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Antibiotic activities of propanolamine containing 1,4-benzoxazin-3-ones against phytopathogenic bacteria†

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Various 1,4-benzoxazin-3-one derivatives containing propanolamine groups have been shown to exhibit good antibacterial activity against *Pseudomonas syringae* pv *actinidiae* (Psa), *X. axonopodis* pv *citri* (Xac) and *Xanthomonas oryzae* pv *oryzae* (Xoo). 1,4-benzoxazin-3-one **4n** showed the best inhibitory effects against Psa, Xac and Xoo, exhibiting *in vitro* EC₅₀ values of 4.95, 4.71 and 8.50 µg mL⁻¹, respectively. These potencies were superior to the corresponding EC₅₀ values of the commercial antibiotics bismethiazol (BT, 89.10, and 116.90 µg mL⁻¹) and thiodiazole copper (TC, 127.30, 82.73 and 87.50 µg mL⁻¹). Treatment on the bacterial leaf blight of rice revealed that compound **4n** displayed better curative (51%) and protective (48%) activities for reducing rice BLB than either BT (41%, 39%) or TC (43%, 41%). Scanning electron microscopy (SEM) imaging of Xoo that had been treated with 1,4-benzoxazin-3-one **4n** (50–100 µg mL⁻¹) revealed that the bacterial cells had experienced extensive cell wall damage, which is the likely cause of its antimicrobial activity and bacterial death.

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1. Introduction

Plant diseases caused by microbial infections have become one of the world's largest agricultural problems, resulting in serious reductions in crop yields and the production of poor quality agricultural products that can cause significant economic losses.^{1,2} Amongst these pathogens, *Pseudomonas syringae* pv *actinidiae* (Psa; which causes kiwi disease), *X. axonopodis* pv (Xac; which causes rice bacterial leaf blight [BLB]) and *Xanthomonas oryzae* pv *oryzae* (Xoo; which causes citrus canker) are particularly problematic.^{3–6} Currently, only a few established commercial bactericides, namely, bismethiazol (BT), etridiazole, benthiazole and thiodiazole copper (TC), are used for the treatment of these plant bacterial diseases. Unfortunately, long-term use of these antimicrobial agents has resulted in the emergence of antimicrobial resistance in many of these pathogenic bacteria,^{7–9} so the discovery of new antibiotics that elicit their antimicrobial activity through novel modes of action is very important.^{10,11}

Recently, 1,4-benzoxazin-3-one derivatives have gained increasing attention because of their broad pharmacological activities, including examples that display antibacterial,^{12,13}

antifungal,^{14,15} herbicidal,^{16,17} anticancer,^{18,19} neurodegenerative diseases²⁰ and platelet aggregation inhibitor activities.²¹ Anti-bacterial 1,4-benzoxazin-3-one derivatives include ethyl-2-(3-oxo-3,4-dihydro-2*H*-benzo[b][1,4]oxazin-2-yl)acetate (Fig. 1, A), 4-cyclo-hexyl-7-fluoro-2*H*-benzo[b][1,4]oxazin-3(4*H*)-one (Fig. 1, B), 2*H*-benzo[b][1,4]oxazin-3(4*H*)-one (Fig. 1, C), and methyl 3-(2-oxo-2*H*-benzo[b][1,4]oxazin-3-yl) propanoate derivatives (Fig. 1, D). Consequently, 2*H*-1,4-benzoxazin-3-ones can be considered as a potential pharmacophores candidate for developing new agrochemical antibiotics for the treatment of plant bacterial diseases.

Propanolamine based compounds are also known to exhibit a wide range of medically useful properties, including anti-fungal,^{22,23} antibacterial,^{24,25} antimalarial,^{26,27} antihyperglycemic²⁸ and anticancer drug activities.^{29,30} For example, Zhao *et al.* have

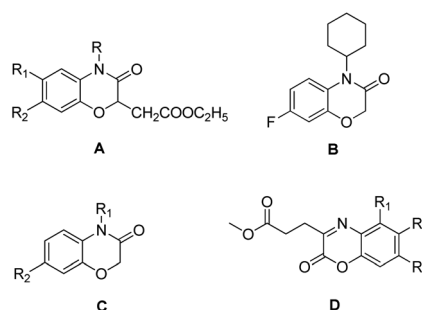


Fig. 1 Selected 1,4-benzoxazin-3-one compounds that exhibit anti-bacterial activities against plant bacterial pathogens.

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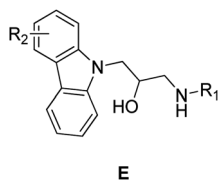


Fig. 2 Propanolamine containing carbazoles **E** that exhibit antibiotic activity against plant pathogens.

recently shown that a propanolamine derived carbazole exhibited good antibacterial activities (Fig. 2). Consequently, we decided to investigate whether incorporating a propanolamine group into a 4-benzoxazin-3-one ring system would afford a hybrid compound with good antibiotic activity for the treatment of citrus canker, rice BLB and kiwi disease. These studies reveal that propanolamine 4-benzoxazin-3-ones display good antimicrobial activity against these plant pathogens *in vitro*, with 1,4-benzoxazin-3-one **4n** used as an effective *in vivo* treatment of BLB in rice. Scanning electron microscopic (SEM) analysis of bacteria treated with 1,4-benzoxazin-3-one **4n** revealed significant damage to their cell walls, which is the likely cause of its antibacterial activity.

2. Experimental

2.1. Methods and materials

XT-4 binocular microscope (Beijing Tech Instrument Co., China) was used for melting point determinations with uncorrected documentation. ^1H , ^{13}C and ^{19}F NMR spectra were measured using a Bruker Biospin AG-400 (400 MHz) spectrometer in the presence of solvent (CDCl_3) solvent and internal standard (TMS). Chemical shifts were recorded in ppm by referring to TMS, while coupling constants were recorded in Hz by referring to the peak multiplicity. SEM images were acquired by Nova NanoSEM 450. TLC with silica gel GF254 as stationary phase was used to monitor the progress of each reaction. All reagents were of analytical grade. All solvents and reagents were in analytically or chemically pure form, and not purified further.

2.2. General procedure for preparing intermediate 1

An equimolar amount of KOH (376.17 mg, 6.70 mmol) was mixed with 1,4-benzoxazin-3-one (1 g, 6.70 mmol) dissolved in 5 mL *N,N*-dimethylformamide. Subsequently, the resultant mixture was stirred at 0°C for 0.5 hours. Epibromohydrin

(1.84 g, 13.41 mmol) was then added portionwise with stirring at 0°C for 8 hours. After the reaction was completed, the reaction solution was then poured into 60 mL ethyl acetate and rinsed with 3×30 mL water, following by drying (Na_2SO_4), filtering and concentrating in a vacuo. The resultant crude product was then purified with column chromatography or preparative TLC consisting of petroleum ether : ethyl acetate (1 : 8, v/v) mixture as solvent.

2.3. General procedure for preparing intermediate 3

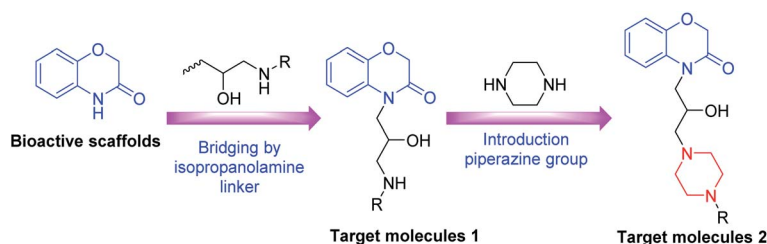
A stirred solution of intermediate **1** (1 g, 4.87 mmol), piperazine (2.1 g, 24.36 mmol), K_2CO_3 (673.47 mg, 4.87 mmol) in isopropyl alcohol (6 mL) was heated at 60°C for 4 hours. After the reaction was completed, the reaction solution was then poured into 50 mL ethyl acetate and rinsed with 3×25 mL water, followed by drying (Na_2SO_4), filtering and concentrating in a vacuo. The resultant crude product was then purified using column chromatography or preparative TLC consisting of methanol : -methylene chloride (1 : 20, v/v) mixture as solvent.

2.4. General procedure for preparing 1,4-benzoxazin-3-one 2a–2d

A stirred solution of intermediate **1** (300 mg, 1.46 mmol), reaction compound (1.46 mmol), and K_2CO_3 (202.04 mg, 1.46 mmol) dissolved in 3 mL isopropyl alcohol was heated at 60°C for 6 hours. After the reaction was completed, the reaction solution was then poured into 25 mL ethyl acetate, rinsed with 3×15 mL water, followed by drying (Na_2SO_4), filtering and concentrating in a vacuo. The resultant crude products were then purified using column chromatography or preparative TLC consisting of petroleum ether : ethyl acetate (1 : 8, v/v) mixture as solvent.

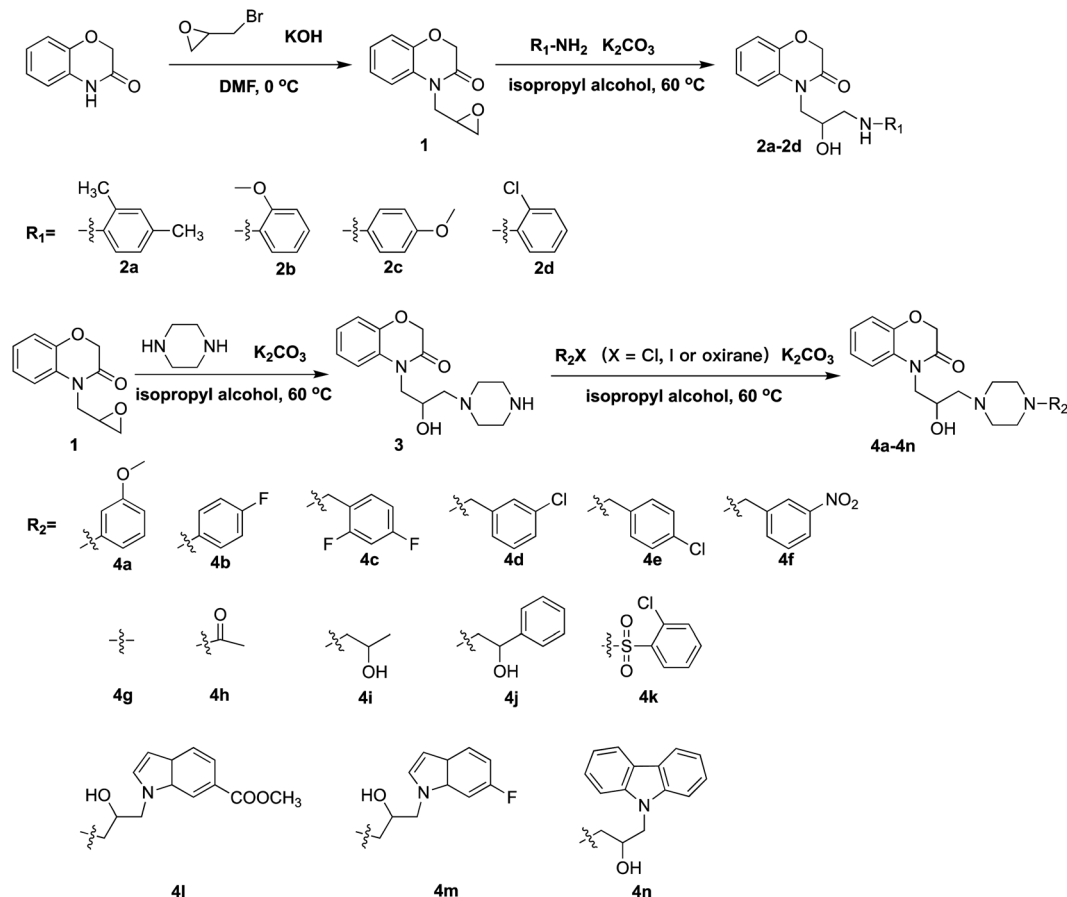
2.5. General procedure for preparing 1,4-benzoxazin-3-ones 4a–4n

A stirred solution of intermediate **3** (300 mg, 1.03 mmol), reaction compound (1.03 mmol), and K_2CO_3 (142.31 mg, 1.03 mmol) dissolved in 3 mL isopropyl alcohol was heated at 60°C for 8 hours. After the reaction was completed, the reaction solution was then poured into 25 mL ethyl acetate, rinsed with 3×15 mL water, followed by drying (Na_2SO_4) filtering and concentrating in a vacuo. The resultant crude products were then purified using column chromatography or preparative TLC



Scheme 1 Design route of the target compounds.





Scheme 2 Synthesis of propanolamine derived 1,4-benzoxazin-3-one derivatives 4a–4n.

consisting of methanol : CH_2Cl_2 (1 : 20, v/v) mixture as solvent (Scheme 1).

3. Results and discussion

3.1. Synthesis of propanolamine derived 1,4-benzoxazin-3-one derivatives 2a–2d and 4a–4n

The synthetic approaches used to prepare two different series of novel 1,4-benzoxazin-3-one derivatives consisting of a propanolamine moiety are illustrated in Scheme 2. Intermediates 1 and 3 were prepared using a method described previously.⁹ Treatment of intermediate 1 (or 3) with different substituted phenylamine nucleophiles resulted in ring-opening reactions to afford the desired 1,4-benzoxazin-3-one derivatives 2a–2d (or 4a–4n). All the new compounds were confirmed by ^1H NMR, ^{13}C NMR, ^{19}F NMR and HRMS spectroscopy (ESI).

Compound 2a has no intramolecular hydrogen bonds, but a crystal of compound 2a was formed by two intermolecular hydrogen bonds. One intermolecular hydrogen bond was formed by hydroxy oxygen of 2a and the imino hydrogen of the second 2a, the other bond was formed by hydroxyl hydrogen of 2a and the carbonyl oxygen of the third 2a with bond length of 3.012 Å and 2.761 Å respectively. The structural composition of compound 2a was determined using single-crystal X-ray analysis. The deposition number is CCDC 1938560 (see Fig. 3).

3.2. Antibacterial bioassay *in vitro*

The *in vitro* bioactivities of the 1,4-benzoxazin-3-ones 2a–2d and 4a–4n against Xoo, Xac and Psa were determined using turbidimetric tests, with their bioassay results reported in Tables 1 and 2. Most of the novel 1,4-benzoxazin-3-one derivatives exhibited moderate to excellent antibacterial activities against Psa, Xac and Xoo (see Table 1). Compounds 4b, 4d, 4e, 4f, 4m and 4n exhibited excellent antibacterial activities of $\geq 62\%$ and $\geq 51\%$ against Xoo at 100 and $50\text{ }\mu\text{g mL}^{-1}$, respectively. Notably,

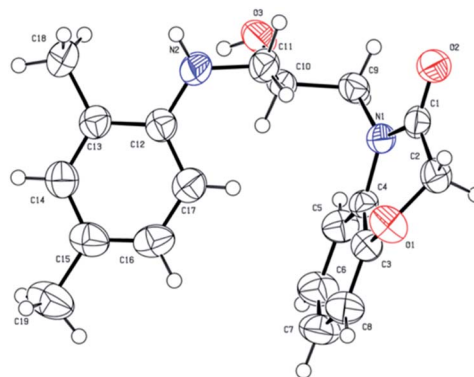


Fig. 3 Crystalline structure of compound 2a.



Table 1 Inhibition effect of 1,4-benzoxazin-3-ones against Xoo, Xac and Psa^a

Compd	Inhibition (%)					
	Xoo		Xac		Psa	
	100 µg mL ⁻¹	50 µg mL ⁻¹	100 µg mL ⁻¹	50 µg mL ⁻¹	100 µg mL ⁻¹	50 µg mL ⁻¹
2a	27.5 ± 2.0	18.1 ± 2.4	27.0 ± 1.4	15.6 ± 2.3	17.7 ± 2.3	11.0 ± 1.5
2b	12.7 ± 1.1	9.1 ± 1.2	19.2 ± 1.2	5.7 ± 2.1	1.4 ± 3.7	— ^c
2c	12.3 ± 1.9	8.7 ± 1.2	17.5 ± 4.8	6.1 ± 1.5	—	—
2d	15.7 ± 4.8	8.7 ± 5.2	12.0 ± 1.2	6.7 ± 1.5	14.1 ± 2.1	—
3	16.6 ± 4.4	14.8 ± 3.5	7.9 ± 0.6	—	—	—
4a	24.4 ± 4.5	14.9 ± 1.0	35.4 ± 0.0	23.4 ± 1.8	—	—
4b	62.4 ± 2.2	59.4 ± 0.6	62.5 ± 1.2	59.1 ± 3.8	40.8 ± 0.7	24.3 ± 0.9
4c	48.6 ± 0.6	30.0 ± 3.5	56.1 ± 1.8	37.2 ± 1.5	30.7 ± 1.3	26.1 ± 2.6
4d	71.7 ± 1.8	63.5 ± 1.1	90.5 ± 0.7	78.7 ± 1.4	54.0 ± 1.6	43.5 ± 1.4
4e	91.6 ± 3.3	71.0 ± 2.6	96.9 ± 0.5	86.0 ± 2.1	78.9 ± 0.4	64.2 ± 1.9
4f	66.9 ± 4.3	50.6 ± 1.2	57.1 ± 2.1	35.5 ± 1.9	46.3 ± 3.2	27.0 ± 2.1
4g	19.4 ± 4.6	16.9 ± 3.4	22.6 ± 1.6	16.2 ± 2.5	—	—
4h	17.8 ± 2.6	14.3 ± 4.5	12.0 ± 4.3	8.6 ± 4.2	3.1 ± 1.4	1.0 ± 4.6
4i	18.9 ± 0.9	15.0 ± 1.5	19.2 ± 3.9	12.7 ± 1.6	15.0 ± 1.1	7.5 ± 1.4
4j	27.9 ± 4.4	15.9 ± 2.1	21.3 ± 0.6	16.0 ± 3.7	21.5 ± 2.0	12.2 ± 2.1
4k	15.2 ± 4.8	5.6 ± 0.9	15.4 ± 3.2	8.1 ± 2.8	14.2 ± 2.7	7.2 ± 1.7
4l	34.3 ± 4.5	27.9 ± 1.0	54.7 ± 1.9	33.8 ± 5.0	28.8 ± 2.6	17.8 ± 3.4
4m	80.0 ± 3.8	64.9 ± 2.7	81.4 ± 2.0	77.7 ± 2.3	71.4 ± 1.8	63.7 ± 2.3
4n	100 ± 0.5	97.4 ± 1.1	100 ± 1.3	97.4 ± 1.9	83.5 ± 2.1	74.0 ± 3.8
BT ^b	58.4 ± 3.7	31.7 ± 1.5	—	—	49.1 ± 1.5	21.4 ± 0.8
TC ^b	48.7 ± 0.5	30.6 ± 1.9	58.8 ± 3.5	27.5 ± 0.9	55.0 ± 2.4	30.1 ± 2.7

^a Average of three replicates; BT: bismethiazol; TC: thiadiazole copper. ^b The antibacterial agents applied to generate comparative antibacterial activities. ^c “—” indicate as not active.

Table 2 EC₅₀ values of 1,4-benzoxazin-3-ones against Psa, Xac and Xoo^a

Compd	Xoo		Xac		Psa	
	Regression equation	EC ₅₀ (µg mL ⁻¹)	Regression equation	EC ₅₀ (µg mL ⁻¹)	Regression equation	EC ₅₀ (µg mL ⁻¹)
4b	y = 0.5434x + 4.1434	37.70 ± 1.63	y = 1.0179x + 3.3998	37.33 ± 1.46	—	—
4d	y = 0.7887x + 3.8552	28.28 ± 2.42	y = 1.2451x + 3.682	11.44 ± 1.73	y = 0.5991x + 3.8205	93.07 ± 1.54
4e	y = 1.2439x + 3.6936	11.23 ± 1.16	y = 1.7851x + 3.0969	11.64 ± 1.27	y = 0.8778x + 3.7733	24.97 ± 2.33
4m	y = 1.1474x + 3.3538	27.21 ± 1.61	y = 2.1693x + 2.57	13.19 ± 1.28	y = 1.0277x + 3.5593	25.23 ± 1.32
4n	y = 1.9139x + 3.6705	4.95 ± 0.73	y = 1.7394x + 3.8296	4.71 ± 0.52	y = 1.1107x + 3.9675	8.50 ± 0.83
BT ^b	y = 1.2472x + 2.5681	89.10 ± 1.96	—	—	y = 1.7949x + 1.2883	116.9 ± 1.71
TC ^b	y = 1.1471x + 2.5854	127.3 ± 1.50	y = 1.6235x + 1.8867	82.73 ± 1.45	y = 1.9802x + 1.1106	87.50 ± 1.42

^a Average of three replicates; BT: bismethiazol; TC: thiadiazole copper. ^b The antibacterial agents applied to compare antibacterial activities.

Table 3 Protective and curative effects of compound 4n (200 µg mL⁻¹) on the BLB of rice

Treatment	Protection activity (14 days following the last spray application)			Curative activity (14 days following the last spray application)		
	Morbidity (%)	Disease index (%)	Control efficiency ^b (%)	Morbidity (%)	Disease index (%)	Control efficiency ^b (%)
4n	100	40.61	48.16	100	39.48	51.08
BT	100	47.92	38.82	100	47.92	40.63
TC	100	45.83	41.49	100	46.15	42.82
CK ^a	100	78.33	—	100	80.71	—

^a Negative control. ^b Data were analyzed using ANOVA method, and *p* value less than 0.05 indicated statistically significant different. The same uppercase letters describe the non-significant difference in protective activities between two groups.



these inhibitory values were superior to those of bismethiazol (BT; 58% and 32%) and thiodiazole copper (TC; 49% and 31%). Similarly, compounds **4b**, **4d**, **4e**, **4m** and **4n** assayed at 100 and 50 $\mu\text{g mL}^{-1}$ showed greater antibacterial activities ($\geq 62\%$ and $\geq 59\%$, respectively) against Xac compared to BT (59% and 28%, respectively). Finally, the antibacterial activities of compounds **4e**, **4m** and **4n** against Psa were $\geq 71\%$ and $\geq 64\%$ at 100 and 50

$\mu\text{g mL}^{-1}$, respectively, which were better than BT (49% and 21%) and TC (55% and 30%), respectively.

The EC_{50} values of these 1,4-benzoxazin-3-one derivatives against Xoo were found to range from 4.58–37.70 $\mu\text{g mL}^{-1}$, which were better than BT (89.10 $\mu\text{g mL}^{-1}$) and TC (127.30 $\mu\text{g mL}^{-1}$) (see Table 2). These compounds also expressed moderate to good bioactivities (EC_{50} = 4.57–37.33 $\mu\text{g mL}^{-1}$) against Xac

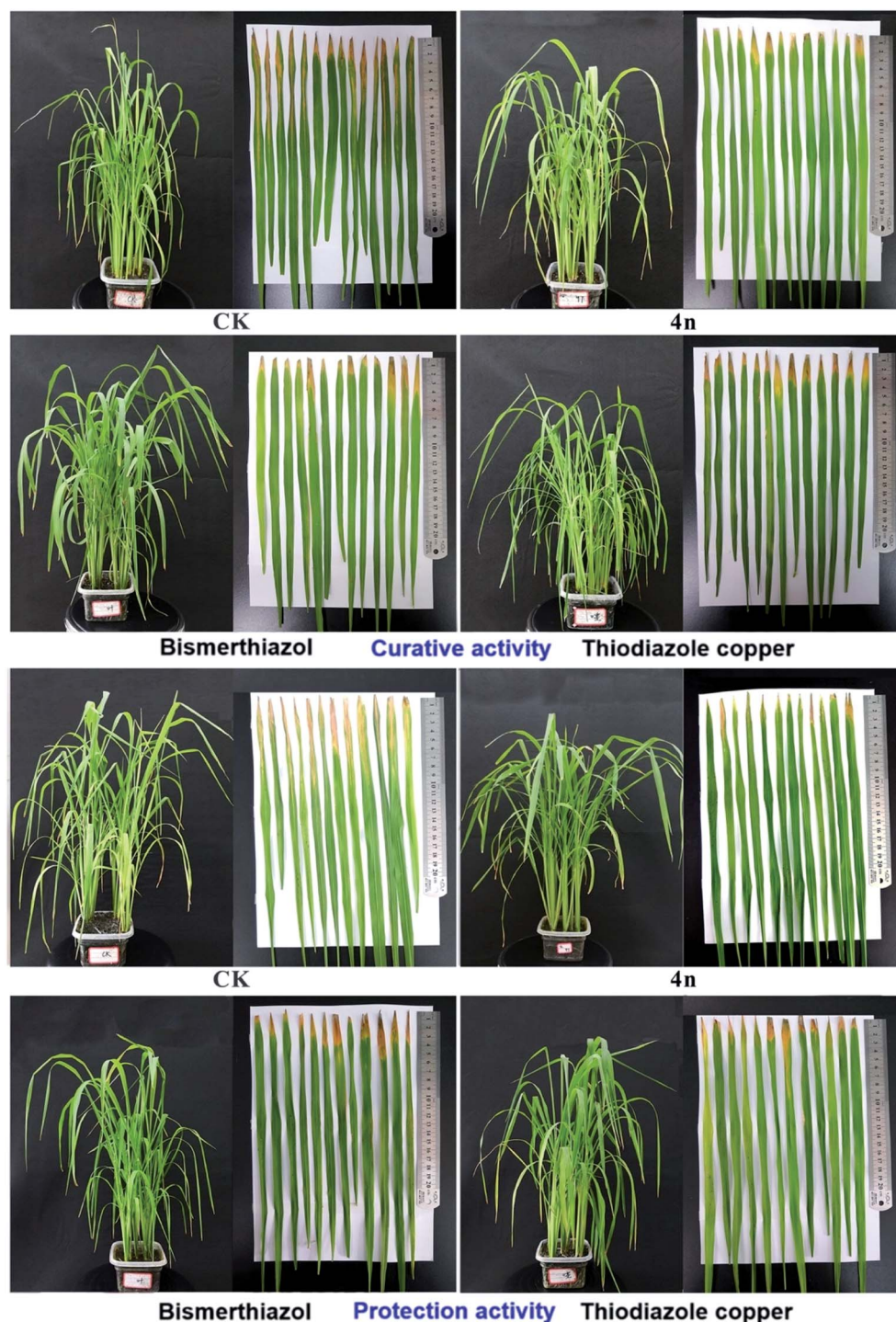


Fig. 4 Protective and curative effects of compound **4n** (200 $\mu\text{g mL}^{-1}$) on the BLB of rice. BT and TC were used as positive controls under similar experimental conditions.



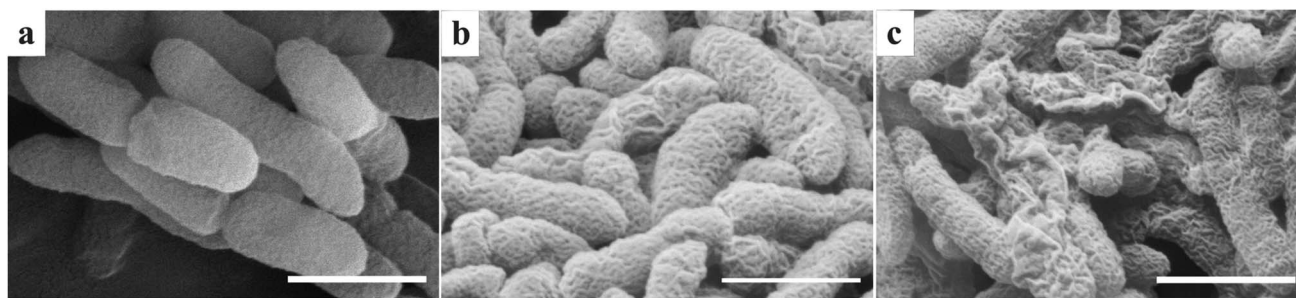


Fig. 5 SEM images of Xoo treated with different amounts of compound **4n**. Images of Xoo treated with 0 (a), 50 (b), and 100 (c) $\mu\text{g mL}^{-1}$ of **4n**. Scale bars are 1 μm .

and excellent antibacterial activities against Psa (EC_{50} values = $8.08\text{--}93.07 \mu\text{g mL}^{-1}$), with these inhibitory values superior to BT ($116.90 \mu\text{g mL}^{-1}$) and TC ($87.50 \mu\text{g mL}^{-1}$), respectively. Compound **4n** exhibited the best overall antibacterial activities against Psa ($\text{EC}_{50} = 8.50 \mu\text{g mL}^{-1}$), Xac ($\text{EC}_{50} = 4.71 \mu\text{g mL}^{-1}$) and Xoo ($\text{EC}_{50} = 4.95 \mu\text{g mL}^{-1}$).

3.3. Structure–activity relationship of antibacterial compounds

A structure–activity relationship (SAR) analysis was carried out according to the antibacterial activities reported in Tables 1 and 2. Analysis of the antibacterial data acquired for compounds **2a–2d** revealed that the aryl groups with both electron-donating and-withdrawing groups were well tolerated, which led us to synthesis the second generation 1,4-benzoxazin-3-one derivatives **4a–4n** that contained a wider range of substituents. Analysis of the data for **4a–n** reveals that the antibacterial activities of 1,4-benzoxazin-3-ones containing electron-withdrawing group on their aryl rings were generally higher than those containing electron-donating groups. Compounds containing a single electron withdrawing group were more active than compounds containing two electron withdrawing groups (*cf.* **4c** > **4b**). The biological activity of 1,4-benzoxazin-3-one derivatives containing aromatic substituents were much greater than those containing alkyl derivatives (*cf.* **4a**, **4b**, **4c**, **4d**, **4e**, **4f**) > (**4g**, **4h**, **4i**). Finally, the best overall antibacterial activity was produced by the carbazole derived 1,4-benzoxazin-3-one derivative **4n**, which gave better antibacterial activity than other heterocyclic derivatives (*e.g.*, **4n** > **4m** > **4j** > **4k**). Therefore, these novel propanolamine appended this class of 1,4-benzoxazin-3-ones template represent a promising class of lead compound for the discovery potential agrochemicals for the treatment of phytochemical diseases.

3.4. *In vivo* bioactivity of 1,4-benzoxazin-3-one **4n** against rice BLB

In vivo experiments were carried to determine the potential of 1,4-benzoxazin-3-one as an agrochemical for treating the BLB of rice. As shown in Table 3 and Fig. 4, compound **4n** exhibited a greater curative activity (51%) against BLB of rice compared to BT (41%) and TC (42%). Furthermore, compound **4n** demonstrated excellent protection activity (48.16%) against rice BLB

when compared to BT (38.8%) and TC (41.49%) (see Table 3 and Fig. 4). These results suggests that propanolamine substituted 1,4-benzoxazin-3-ones may be considered as an effective structural template for the development of antibiotic agents to treat phytopathogenic bacterial infections.

3.5. Scanning electron microscopy (SEM) of Xoo bacteria treated with 1,4-benzoxazin-3-one **4n**

SEM images of Xoo cells were acquired to determine whether any morphological changes could be detected after treatment with different dosages of compound **4n**. Treatment of Xoo with compound **4n** at a dose of $50 \mu\text{g mL}^{-1}$ resulted in the morphologies of the surfaces of the cells being changed from being well-rounded (Fig. 5a) to being partially corrugated with a number of pore-like shapes visible (Fig. 5b). Furthermore, the extent of cell wall damage was increased significantly when bacteria were exposed to a higher $100 \mu\text{g mL}^{-1}$ concentration of **4n**, with numerous corrugated and fractured bacterial cells being observed (Fig. 5c). This suggests that the antibiotic **4n** causes physiological changes within the bacteria that result in cell-wall damage that results in lysis and bacterial cell death.

4. Conclusions

Novel 1,4-benzoxazin-3-one derivatives containing propanolamine groups were prepared, and their antibacterial activities were evaluated against three phytopathogenic bacteria. Bioassay results showed that most of these 1,4-benzoxazin-3-ones possess outstanding antibacterial activities against Psa, Xac and Xoo *in vitro*, with the best compound **4n** exhibiting EC_{50} values for Xoo, Xac and Psa of 4.95, 4.71 and $8.50 \mu\text{g mL}^{-1}$, respectively. 1,4-benzoxazin-3-one **4n** also displayed good *in vivo* antibacterial activities against the BLB of rice (curative activity 51% and protective activity 48%) which was better than the curative and protection activities of BT (41% and 39%) and TC (43%, 41%), respectively. SEM images of Xoo incubated with compound **4n** revealed the presence of corrugated and broken cells, consistent with bacterial cell wall damage having occurred that results in bacterial death. These results demonstrate that 1,4-benzoxazin-3-one derivatives containing propanolamine moieties can effectively control the growth of Xoo, Xac and Psa bacteria, thus providing new structural templates for the

development of promising antibiotic agents against phyto-bacterial pathogens.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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Notes and references

- 1 X. P. Song, P. Li, M. W. Li, A. Yang, L. Yu, L. Z. Luo, D. Y. Hu and B. A. Song, *Pestic. Biochem. Physiol.*, 2018, **147**, 11–19.
- 2 B. Li, B. P. Liu, C. L. Shan, M. Lbrahim, Y. H. Lou, Y. L. Wang, G. L. Xie, H. Y. Li and G. C. Sun, *Pest Manage. Sci.*, 2013, **69**, 312–320.
- 3 P. Dai, K. Luo, X. Yu, W.-C. Yang, L. Wu and W.-H. Zhang, *Adv. Synth. Catal.*, 2018, **360**, 468–473.
- 4 F. D. Lorenzo, A. Palmigiano, A. Silipo, Y. Desaki, D. Garozzo, R. Lanzetta, N. Shibuya and A. Molinaro, *Carbohydr. Res.*, 2016, **427**, 38–43.
- 5 Z. Li, S. Chen, S. Zhu, J. Luo, Y. Zhang and Q. Weng, *Molecules*, 2015, **20**, 13941–13957.
- 6 Z. F. Li, S. L. Wu, X. F. Bai, Y. Liu, J. F. Lu, Y. Liu, B. G. Xiao, X. P. Lu and L. J. Fan, *J. Bacteriol.*, 2011, **193**, 6088–6089.
- 7 H. P. T. Ngo, T. H. Ho, I. Lee, H. T. Tran, B. Sur, S. Kim, J. G. Kim, Y. J. Ahn, S. S. Cha and L. W. Kang, *J. Agric. Food Chem.*, 2016, **64**, 7307–7314.
- 8 A. Piazza, T. Zimaro, B. S. Garavaglia, F. A. Ficarra, L. Thomas, C. Marondedze, R. Feil, J. E. Lunn, C. Gehring, J. Ottado and N. Gottig, *Exp. Bot.*, 2015, **66**, 2795–2811.
- 9 Y. L. Zhao, X. Huang, L. W. Liu, P. Y. Wang, Q. S. Long, Q. Q. Tao, Z. Li and S. Yang, *J. Agric. Food Chem.*, 2019, **67**, 7512–7525.
- 10 Y. Zhang, X. Pan, Y. B. Duan, X. F. Zhu, X. W. Ma, T. T. Huang, T. C. Gao and M. G. Zhou, *Australas. Plant Pathol.*, 2015, **44**, 541–543.
- 11 P. Li, L. Shi, M. N. Gao, X. Yang, W. Xue, L. H. Jin, D. Y. Hu and B. A. Song, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 481–484.
- 12 M. Gleńsk, B. Gajda, R. Franiczek, B. Krzyżanowska, I. Biskup and M. Włodarczyk, *Nat. Prod. Res.*, 2015, **30**, 1–4.
- 13 R. Bollu, S. Banu, R. Bantu, A. G. Reddy, L. Nagarapu, K. Sirisha, C. G. Kumar, S. K. Gunda and K. Shaik, *Bioorg. Med. Chem. Lett.*, 2017, **27**, 5158–5162.
- 14 Y. Q. Jiang, L. F. Mao, L. Zhu, B. Q. Ren, W. Li and G. Q. Xu, *Z. Naturforsch. B Chem. Sci.*, 2014, **69b**, 103–108.
- 15 M. Śmist, H. Kwiecień and M. Krawczyk, *J. Environ. Sci. Health, Part B*, 2016, **51**, 1–9.
- 16 M. E. d. l. Calle, G. Cabrera, D. Cantero, A. Valle and J. Bolivar, *New Biotechnol.*, 2019, **50**, 9–19.
- 17 H. B. Li, L. Li, J. X. Li, T. F. Han, J. L. He and Y. Q. Zhu, *Pest Manage. Sci.*, 2017, **74**, 579–589.
- 18 C. L. Su, C. L. Tseng, C. Ramesh, H. S. Liu, C. Y. F. Huang and C. F. Yao, *Eur. J. Med. Chem.*, 2017, **132**, 90–107.
- 19 P. Wang, F. Liu, Q. Zhong, S. L. Zheng, Y. Chen, G. D. Wang and L. He, *Chin. Chem. Lett.*, 2017, **28**, 1243–1247.
- 20 L. L. Wang, H. Ankati, S. K. Akubathini, M. Balderamos, C. A. Storey, A. V. Patel, V. Price, D. Kretzschmar, E. R. Biehl and S. R. D'Mello, *J. Neurosci. Res.*, 2010, **88**, 1970–1984.
- 21 S. Xia, J. Q. Liu, X. H. Wang, Y. Tian, Y. Wang, J. H. Wang, L. Fang and H. Zuo, *Bioorg. Med. Chem. Lett.*, 2014, **24**, 1479–1483.
- 22 S. P. Zhu, W. Y. Wang, K. Fang, Z. G. Li, G. Q. Dong, Z. Y. Miao, J. Z. Yao, W. N. Zhang and C. Q. Sheng, *Chin. Chem. Lett.*, 2014, **25**, 229–233.
- 23 Y. Zhang, V. K. R. Tangadanchu, R. R. Y. Bheemanaboina, Y. Cheng and C. H. Zhou, *Eur. J. Med. Chem.*, 2018, **155**, 579–589.
- 24 C. Y. Cheng, C. P. Chang, T. L. Y. Lauderdale, G. Y. Yu, J. C. Lee, Y. W. Jhang, C. H. Wu, Y. Y. Ke, A. A. Sadani, C. F. Yeh, I. W. Huang, Y. P. Kuo, D. J. Tsai, T. K. Yeh, C. T. Tseng, J. S. Song, Y. W. Liu, L. K. Tsou and K. S. Shia, *ACS Med. Chem. Lett.*, 2016, **7**, 1191–1196.
- 25 Y. Zhang, V. K. R. Tangadanchu, Y. Cheng, R. G. Yang, J. M. Lin and C. H. Zhou, *ACS Med. Chem. Lett.*, 2018, **9**, 244–249.
- 26 A. Robin, F. Brown, N. Bahamontes-Rosa, B. H. Wu, E. Beitz, J. F. J. Kun and S. L. Flitsch, *J. Med. Chem.*, 2007, **50**, 4243–4249.
- 27 J. Molette, J. Routier, N. Abila, D. Besson, A. Bombrun, R. Brun, H. Burt, K. Georgi, M. Kaiser, S. Nwaka, M. Muzerelle and A. Scheer, *ACS Med. Chem. Lett.*, 2013, **4**, 1037–1041.
- 28 P. S. Humphries, R. Bersot, J. Kincaid, E. Mabery, K. McCluskie, T. Park, T. Renner, E. Riegler, T. Steinfeld, E. D. Turtle, Z. L. Wei and E. Willis, *Bioorg. Med. Chem. Lett.*, 2018, **28**, 293–297.
- 29 J. Y. Jin, C. X. Miao, Z. L. Wang, W. L. Zhang, X. W. Zhang, X. Xie and W. Lu, *Eur. J. Med. Chem.*, 2018, **150**, 757–770.
- 30 Z. Chen, T. Y. Yang, W. S. Wang, J. M. Yao, S. M. Han, Y. Tao, R. Wang and L. P. Duan, *ChemistrySelect*, 2018, **3**, 12630–12638.

