


 Cite this: *RSC Adv.*, 2020, 10, 6223

The development of isoguanosine: from discovery, synthesis, and modification to supramolecular structures and potential applications

 Tingting Ding,[†] Fan Tang,[†] Guangcheng Ni, Jiang Liu,^{*} Hang Zhao ^{*} and Qianming Chen

Isoguanosine (isoG), an isomer of guanosine (G), differs from G by the translocation of the C2 carbonyl and C6 amino groups. This minor translocation of functional groups results in significant differences in the properties of G and isoG. Like G, isoG can self-assemble into various supramolecular structures such as tetramers and decamers in the presence of various cations. And it shows a wide range of applications including in ionophores, genetics, gel formation, and cancer treatment. Although there have been many studies and reviews concerning G in recent years, to the best of our knowledge, there have been no reviews that summarized isoG works. Herein, the discovery, synthesis, modification, supramolecular structures and potential applications of isoG are systematically reviewed in this article, in order to provide a reference for future studies.

Received 12th November 2019

Accepted 17th January 2020

DOI: 10.1039/c9ra09427j

rsc.li/rsc-advances

1 Introduction

Nucleic acids are one of the most basic substances of life, not only comprising genetic material but also playing a crucial role in protein biosynthesis. Nucleic acids are biopolymers composed of nucleotide monomers. Nucleotides in living organisms are composed of a five-carbon sugar, three phosphate molecules and a nitrogenous base (adenine, guanine, cytosine, thymine or uracil, Fig. 1). Compared with the other four bases, guanine has more unique properties because of its unique 2-amino and 6-carbonyl groups. Its 2-amino and 6-carbonyl groups can act as a hydrogen bond donor and receptor, respectively, resulting in its assembly into different structures, such as the cation-templated G-quartet and G-quadruplex.^{1,2} G-quartets have various applications in ionophore and nanostructure formation. And G-quadruplexes are present in telomeres, suppressing the growth of tumors as telomerase inhibitors. They can also take part in gel formation, and recognize proteins and small molecules. In addition, G-quadruplexes are found in promoter regions of genes and in some untranslated regions of messenger RNA (mRNA).

isoG is a natural isomer of G, differing from G because the C2 carbonyl and C6 amino groups are transposed (purine numbering is widely used in this paper) (Fig. 1). This minor

change leads to significant differences between isoG and G. isoG, is also known as crotonoside, 2-hydroxyadenosine (2-OH-Ado), 2-oxoadenosine (2-oxo-Ado), 1,2-dihydro-2-oxoadenosine (1,2-dihydro-2-oxo-Ado), 2-oxy-6-amino-isoguanine- β -D-ribose and 6-amino-9- β -D-ribofuranosyl-9H-purine-2-oxo-isoguanine riboside. Significant research into isoG has been carried out since Fischer³ first synthesized isoguanine in 1897. isoG/isoguanine were firstly separated from natural substances such as croton beans (*Crotonis oleum*),⁴ butterfly wings⁵ and marine mollusks.⁶ Later, isoG and its derivatives were synthesized from three categories of methods, one is from the protected 2,6-diamino-9- β -D-ribofuranosylpurine 1,⁷ adenosine 1-oxygen 2,⁸ the protected 6-chloroxanthosine 3 (Fig. 3)⁹ by the introduction of functional groups to the base. The second is from G by exchanging C2 amino and C6 carbonyl groups,¹⁰ and the third is from 5-amino-1-(β -D-ribofuranosyl)-imidazole-4-carboxamide (AICA riboside, 4, Fig. 3) to construct the nitrogen heterocycle.¹¹ In addition, researchers have modified the sugar or base groups of isoG/isoguanine to explore differences among them and the potential unique properties. In the meantime, the structures, self-assembly properties of isoG with/without metal cations (M^+) have been studied in monomer and multi-molecular layers. isoG can self-assemble into different cation-templated supramolecular structures, such as $(isoG)_4 \cdot M^+$ (ref. 12) and $(isoG)_{10} \cdot M^+$,¹³ in a different manner from G. Finally, its potential applications in fields of ionophore, gel-formation, and anticancer treatments *etc.* are also of great significance and have been systematically reviewed. We hope that this review can not only give a glimpse of the comprehensive

State Key Laboratory of Oral Diseases, National Clinical Research Center for Oral Diseases, Chinese Academy of Medical Sciences Research Unit of Oral Carcinogenesis and Management, West China Hospital of Stomatology, Sichuan University, Chengdu, Sichuan, 610041, P. R. China. E-mail: zhaohanghy@scu.edu.cn; liujiang@scu.edu.cn

[†] These authors contributed equally to this work.



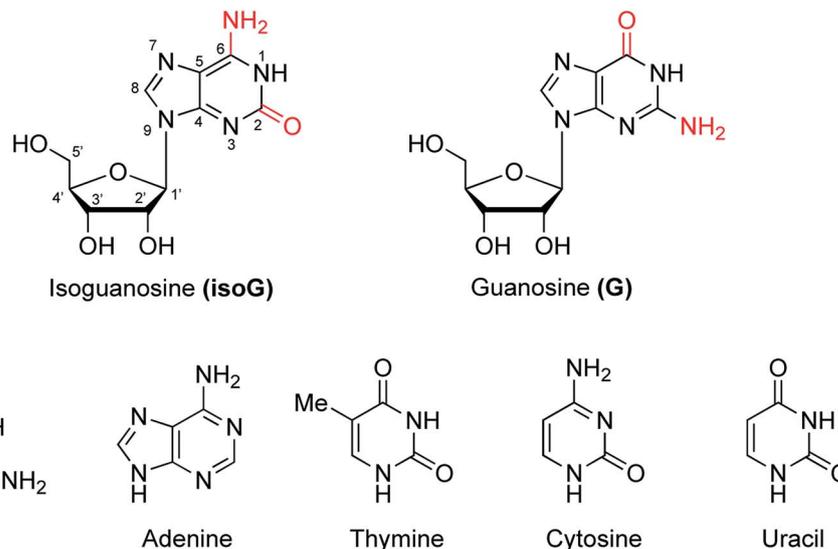


Fig. 1 Structures of isoG and G as well as the five natural bases.

development surrounding isoG, but also inspire further research into this fascinating topic.

2 Discovery process of isoG

In 1897, Fischer³ synthesized many purines and identified a strong similarity between guanine and one particular product 2-oxo-6-aminopurine, which had not been discovered in nature (Fig. 2). Believing that 2-oxo-6-aminopurine was oxidized adenine, he suggested that 2-oxo-6-aminopurine would most likely be discovered in animals. And 2-oxo-6-aminopurine may have already been mistaken for guanine. Then in 1927, Buell¹⁴ *et al.* found that oxyadenine, a purine isolated from pig blood, is consistent with 2-oxo-6-aminopurine. However, the presence of 2-oxo-6-aminopurine in pig blood could not be confirmed. In 1932, during a study of croton seeds, Cherbuliez and Bernhard⁴ accidentally discovered a new glucoside, which had not been

discovered in nature. They identified it as 2-hydroxy-6-amino-purine-D-ribose. However, the yields obtained from the seeds were variable and very low, and glycosyl crystals could not be obtained by hydrolysis. Subsequently, in 1935, Spies¹⁵ *et al.* successfully enriched the new glucoside and obtained the crystals, and identified the structure as the D-ribose form. In 1939, Spies¹⁶ confirmed the observations of Fischer³ and Cherbuliez⁴ *et al.* that isoguanine is converted to xanthine by hydrochloric acid but is resistant to nitric acid, whereas guanine is converted to xanthine in nitric acid. This reactivity was evidence of the 2-oxo-6-aminopurine structure that Bendich had observed, *i.e.*, that 2,6-diaminopurine can only be converted to isoguanine rather than xanthine in the reaction with excess nitrous acid. In 1939, Falconer¹⁷ *et al.* confirmed the ribose was linked to the base at N9 position. Afterwards, isoguanine was separated from the butterfly wings of *Prioneris thestylis* by Purmann⁵ in 1940 and by Pettit¹⁸ *et al.* in 1976. In 1951,

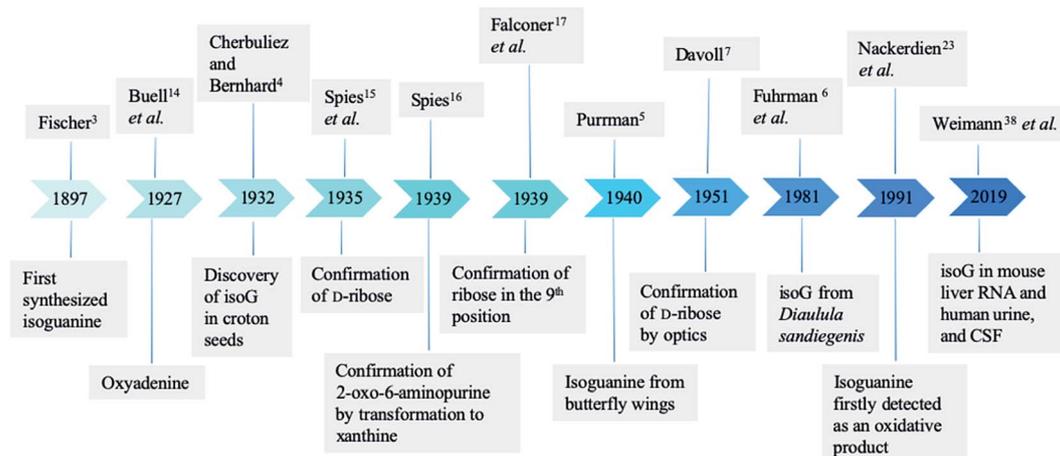


Fig. 2 Natural discovery of isoguanine/isoG and the confirmation of structure.



experiments by Davoll^{7,19} *et al.* into optical properties of these compounds revealed that isoG was consistent with 9-β-D-ribofuranosylisoguanine. Later, in a study of *Diaulula sandiegensis* in 1981, Fuhrman⁶ *et al.* confirmed the presence of isoG in extracts of the digestive glands of the sea slug *Diaulula sandiegensis*. However, the yield was very low, only one milligram of crystals can be obtained from per gram of dry tissue.

Interestingly, although isoG is a nucleoside that has been isolated from a variety of species, it does not exist as a natural nucleic acid. In 1950, Brown²⁰ *et al.* demonstrated that isoguanine is not a precursor of nucleoside biosynthesis in mice. In 1952, they reported that isoguanine could not be used as a pure source of purines in *Lactobacillus casei*, nor could it be integrated into nucleic acid.²¹ Notably, C¹⁴ crotonoside has been used to investigate its role in the metabolism of *Escherichia coli* (*E. coli*). The results indicate that isoguanine and its derivatives play an important role in nucleic acid metabolism.^{21,22}

In 1991, using gas chromatography/mass spectrometry (GC-MS) in isolated human chromatin isoguanine, Nackerdien²³ *et al.* firstly detected 2-OH-Ade within Ni(II)/Co(II)-H₂O₂-treated DNA. And in 1992, Dizdaroglu²⁴ reported that in the presence of Fe(III), Cu(II), Ni(II) and Co(II), isoguanine (2-hydroxyadenine) was a product of H₂O₂-treated mammalian chromatin both *in vivo* and *in vitro*. Oxidation reaction commonly occurs in cells because of the presence of reactive oxygen species (ROS), which plays an important role in DNA damage. In this respect, it is logical that isoguanine is more abundant in human cancerous tissue than in normal tissue, which is reported by Olinski^{25,26} *et al.* in 1992 and 1994. In 1993, Dizdaroglu²⁷ *et al.* identified isoguanine in mice at levels of only a few oxidation sites per 10⁵ DNA bases. They also identified isoguanine as an oxidation product of hepatic chromatin exposed to γ-ray in pregnant rats and fetuses of theirs. In 1996, using GC-MS, Jaruga²⁸ *et al.* reported that isoguanine is produced as a DNA oxidation product of adenine in *E. coli*, as well as in cultured human lymphoblast

cells. These have drawn attention toward isoguanine as a marker for oxidative damage formation, repair, mutagenicity.^{29–34}

In 2002, Cadet³⁵ *et al.* demonstrated that after γ-ray radiolysis and Fenton reaction of free nucleosides, the yield of 2'-deoxyisoguanosine (isoG_d) was low, compared to the yield of 8-oxo-2'-deoxy-adenosine, expect in the presence of ferrous ion (Fe²⁺)/ethylenediaminetetraacetic acid (EDTA) with oxygen or cuprous ion (Cu⁺). However, Fe²⁺/EDTA with oxygen or Cu⁺ are typically at very different conditions from those found in cells. They concluded that isoG_d cannot be detected under a limit of per 10⁷ bases in human DNA and *E. coli*. They also reported that the process of conventional GC-MS, which involves acid hydrolysis and derivation at high temperature, may have caused an overestimation of isoguanine/isoG.^{35–37} Notably, in 2019, using the ultra-performance liquid chromatography (UPLC) and electrospray ionization isotope dilution tandem mass spectrometry, Weimann³⁸ *et al.* firstly identified and quantified the ribonucleoside isoG in mouse liver RNA, human urine, and cerebrospinal fluid (CSF). However, the amount of detected isoG is more than or equal to that of 8-oxo-G, which is most easily oxidized, and isoG_d was not detected. These data are opposed to the common hypothesis that the free nucleoside isoG may originate from oxidation reaction. As a result, the reason why isoG exists in mammals still remains a matter of study.

3 Syntheses of isoG

As a special nucleoside, isoG is expected to present applications in various areas. However, its yield from natural sources is too low. Thus, many synthetic methods have been reported (Table 1), and these synthetic methods can be divided into three categories: (1) the introduction of the functional groups to existing heterocyclic 9-β-D-ribofuranosylpurine nucleosides

Table 1 Syntheses of isoguanine/isoG are divided into three categories: (1) the introduction of functional groups to the existing heterocycle, (2) G as a initial reactant to isoG by exchanging C2 amino and C6 carbonyl groups, (3) construction of a nitrogen heterocycle with AICA

Year	Authors	Initial substrates	Productivity	References
1897	Fischer	6-Oxy-2,8-dichlorpurine	— ^a	3
1951	Davoll	2,6-Diamino-9-β-D-glucopyranosylpurine/ 2,6-diamino-9-β-D-ribofuranosyl purine	55% 9-β-D-Glucopyranosylisoguanine/ 57% 9-β-D ribofuranosylisoguanine	7
1964	Brown <i>et al.</i>	Adenosine 1-oxide	Variable	8
1965	Ravindranathan <i>et al.</i>	2,6-Diaminopurine riboside	— ^a	40
1968	Montgomery <i>et al.</i>	The protected 2,6-diaminopurine nucleoside	By-product	42
1995	Napoli <i>et al.</i>	The protected 6-chloroxanthosine	20% isoG	43
1995	Napoli <i>et al.</i>	2',3',5'-tri-O-acetyl-xanthosine	76% isoG	43
1997	Napoli <i>et al.</i>	The protected 6-chloroxanthosine	80% isoG	48
2012	Cheng <i>et al.</i>	Adenine	— ^a	44
1985	Nair <i>et al.</i>	G	34% isoG	39
1991	Divakar <i>et al.</i>	G	64% isoG	10
1968	Yamazaki <i>et al.</i>	4-Amino-5-imidazole carbonitrile (AICN)	— ^a	45
1976	Yamazaki <i>et al.</i>	AICA (riboside)	By-product	46
1987	Reese <i>et al.</i>	The protected AICA riboside	— ^a	9
1987	Chern <i>et al.</i>	AICA (riboside)	77% isoG/(— ^a) isoguanine	11

^a The yield is not mentioned.



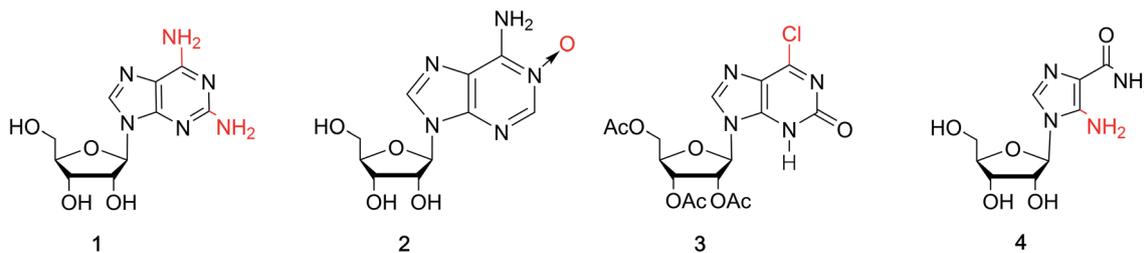


Fig. 3 Common substrates for the synthesis of isoG.

(Fig. 3), such as 2,6-diamino-9- β -D-ribofuranosylpurine **1**,⁷ adenosine 1-oxide **2**,⁸ the protected 6-chloroxanthosine **3**,³⁹ (2) G as an initial reactant to build isoG by exchanging the C2 amino and C6 carbonyl groups, (3) the construction of nitrogen heterocycle with AICA riboside (**4**, Fig. 3) as a precursor for base synthesis to introduce oxo group at C2 position. Notably, the third method is challenging because it is difficult to obtain AICA. These syntheses will be discussed in detail in the following parts.

3.1 Introducing functional groups on the existing heterocycle

Isoguanine was initially synthesized by Fischer³ in 1897. It can resist the deamination by nitrous acid, and the reaction of 2,6-diaminopurine with excess nitrous acid only yields isoguanine not xanthine. In 1951, Davoll^{7,19} *et al.* reported that 9- β -D-glucopyranosyl-isoguanine can be obtained in 55% yield by the reaction of 2,6-diamino-9- β -D-glucopyranosylpurine with nitrous acid, while 9- β -D-ribofuranosyl-isoguanine can be obtained in 57% yield from the reaction of 2,6-diamino-9- β -D-ribofuranosylpurine (**1**, Fig. 3) and nitrous acid (Fig. 4). However, the overall yields are low and harmful heavy metal ions (Hg and Pb) are used. So in 1965, Ravindranathan⁴⁰ *et al.* synthesized isoG using the way of Davoll but using much charcoal instead of a lead salt for purification. On account of the relevance of ultraviolet (UV) light and the mutations in nucleic acids, in 1964, Brown⁸ *et al.* reported a photochemical

preparation of isoG from adenosine 1-oxide (**2**, Fig. 4). Adenosine 1-oxide is more sensitive to UV light than adenosine. However, this method gives variable proportions of two major products. One product is adenosine (**6**, Fig. 4) *via* the direct loss of oxygen. The other product is isoG *via* the rearrangement of oxygen atom to the neighboring carbon atom. Subsequently in 1975, Mantsch⁴¹ *et al.* concluded that if nucleotides of the initial substance **2** is irradiated with UV light under alkaline condition (0.01 M NH_4OH), isoG nucleotides can be the corresponding primary products. This is also the case for adenine 1-oxide (**10**, Fig. 5).⁸

In 1968, in a synthesis of 2-fluoroadenosine by Montgomery⁴² *et al.*, isoG was reported to be a by-product, relying on a protected 2,6-diaminopurine nucleoside (**5**, Fig. 4). Generally, the introduction of N to the assigned activated nucleoside bases of guanine, cytosine and adenine is easy but this is not the case for isoG. A significant intermediate **3** can be obtained from 2'3'5'-tri-*O*-acetylxanthosine **7** (Fig. 4). Nevertheless, in 1995, Napoli⁴³ *et al.* reported a direct route from the protected 6-chloroxanthosine **3** to isoG, but this gives a very low yield of 20%. So, they synthesized isoG through the reagent to nucleophilic attack on the pyridinium α -carbon, and obtained the intermediate (**8**, Fig. 4). Then isoG was isolated by HPLC in 76% overall yield by ring opening (Zincke reaction) of the compound **8**. Then in 1997, they proposed another way by introducing a strong electron-withdrawing group in the heterocyclic to enhance the reactivity of C6 towards aqueous ammonia, in

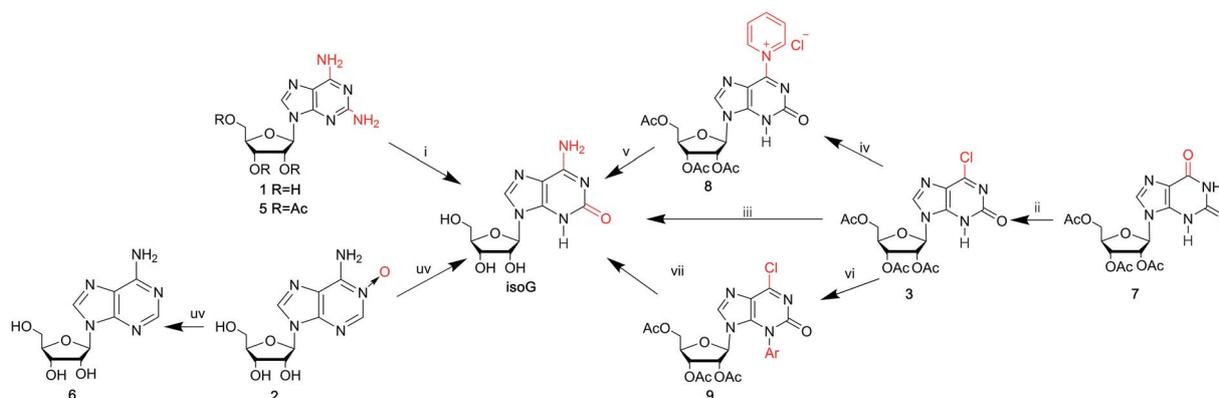


Fig. 4 Syntheses of isoG *via* introducing functional groups. (i) Nitrous acid for compound **1**; hydrogen and palladium-on-charcoal catalyst, 40% fluoroboric acid, sodium nitrite for compound **5**; (ii) PPh_3 (2 mol equiv.), CCl_4 , 3 h, reflux; (iii) aq. NH_3 (32%), 3 days, room temperature; (iv) pyridine–water (1 : 1), 2 h, 50 °C; (v) aq. NH_3 (32%), 5 h, 50 °C; (vi) 2,4-dinitrochlorobenzene in DMF solution, 1.5 h, 80 °C; (vii) NH_4OH .



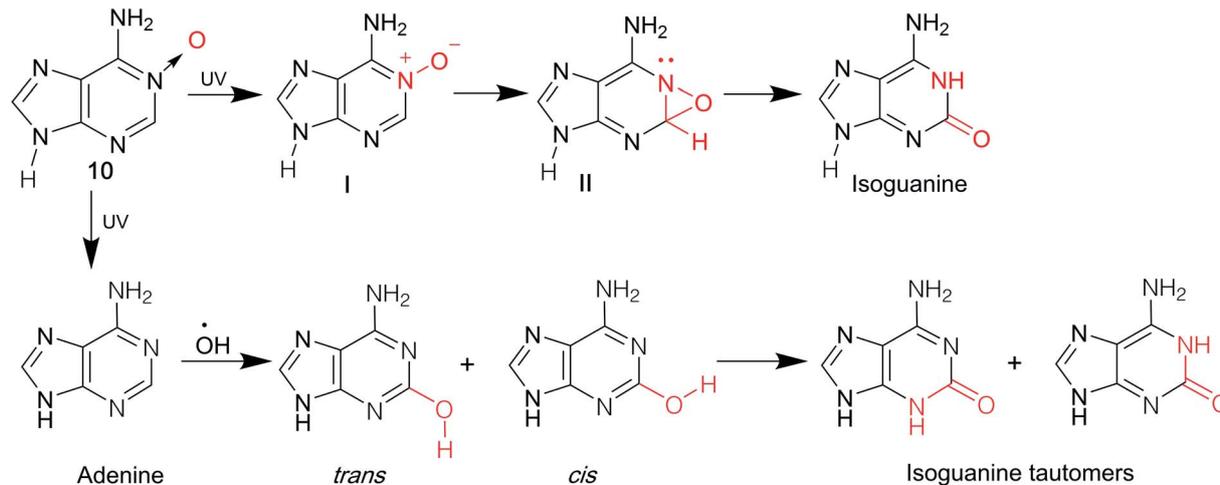


Fig. 5 Syntheses of isoguanine *via* introducing functional groups.

a yield of 80%, with an important intermediate (9, Fig. 4). In 2012, Cheng⁴⁴ *et al.* published a new two-step method based on adenine. A mixture of isoguanine tautomers was obtained (Fig. 5). Reactions had been proved where OH radicals attack adenine on the C2, C4, C5, and C8 positions, and the reactions are spontaneous and exothermic. In the first step, attack by OH radicals at the C2 position of adenine forms a favorable low-energy complex (Fig. 5). In the second step, the enol *cis*-*trans* forms are converted into the corresponding N1H/N3H keto forms.

3.2 G to isoG by exchanging C2 amino and C6 carbonyl groups

As mentioned above, the route from 2,6-diaminopurines to isoG requires harmful heavy metal ions (Hg and Pb) and the overall yields are low. isoG was reported as a by-product in a synthesis of nucleosides from 2,6-diaminopurines, as well as from adenosine-1-oxide, both in variable yields. In 1985, Nair³⁹ *et al.* found a new reproducible and efficient method for the synthesis of isoG starting from G (Fig. 6). This method contains five steps. Firstly, the sugar is protected by selective acetylation. Secondly, reaction with phosphorus oxychloride and *N,N*-dimethylaniline

yields the 6-chloropurine nucleoside (11, Fig. 6). Thirdly, *n*-pentyl nitrite reacts with diiodomethane to obtain protected 2-iodo-6-chloropurine nucleoside (12, Fig. 6). Then, 2-iodoadenine (13, Fig. 6) is obtained by reacting with ethanolic ammonia. And finally, in the last key step in this process, photo-induced hydration occurs. The product isoG was isolated by HPLC. However, this method gives a low yield about 34%. In addition, in the fifth step, the requirement of UV irradiation limits the spread of this synthetic route. Therefore, in 1991, Divakar¹⁰ *et al.* developed an improved five-step route (Fig. 6) starting from G, which achieved isoG with a 64% overall yield involving the same 6-chloropurine nucleoside (11, Fig. 6), with an important intermediate 14. This means that, for the latter reaction, inavailability of direct synthesis to 6-chloroxanthosine is not required. In contrast, a better yield is obtained. Moreover, conditional complex photochemical steps are avoided.

3.3 isoG from AICA by constructing a nitrogen heterocycle

In 1968, Yamazaki⁴⁵ *et al.* found that isoG/isoguanine can be obtained from 4-amino-5-imidazole carbonitrile (AICN, 16, Fig. 7) and its protected riboside (21, Fig. 7), but the yield is too low. For a long time, there have been great efforts to synthesize

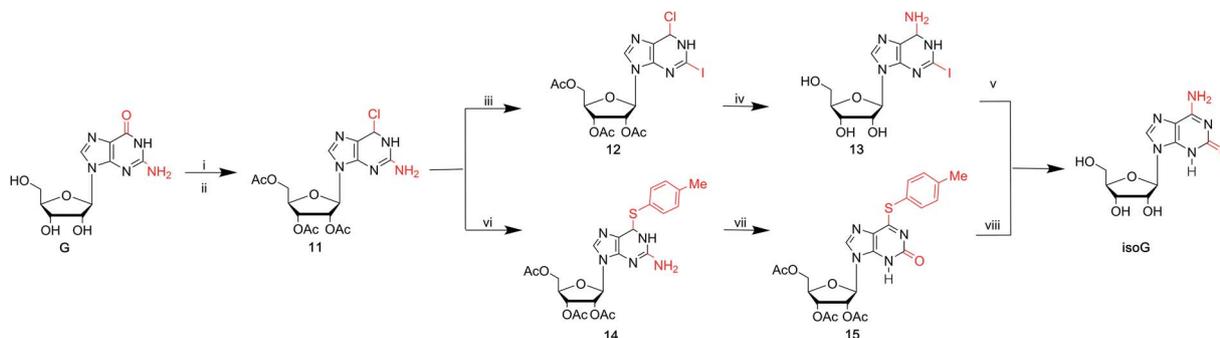


Fig. 6 Syntheses of isoG from G. (i) Ac₂O, pyridine, DMF, 70 °C; (ii) POCl₃, *N,N*-dimethylaniline, CH₃CN, 100 °C; (iii) *n*-C₅H₁₁ONO, CH₂I₂, heating; (iv) NH₃, C₂H₅OH; (v) H₂O, UV; (vi) toluerthiol, TEA, DMF, 100 °C; (vii) 50% AcOH, NaNO₂, 50 °C; (viii) ammonia, ethanol, 70 °C.



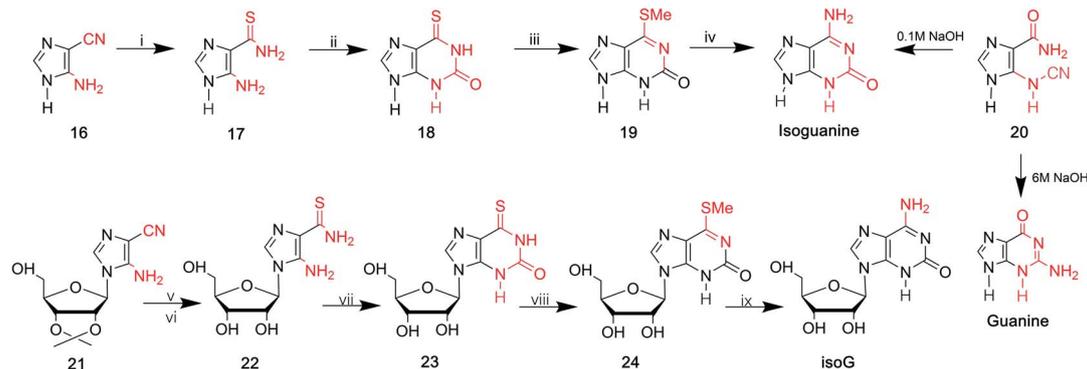


Fig. 7 Syntheses of isoG/isoguanine from AICN. (i) H_2S , KOH; (ii) fusion with urea; (iii) CH_3I , alkaline solution; (iv) ammonia, autoclave at 130°C , 3 h; (v) H_2S ; (vi) acid condition, hydrolyzation; (vii) diethyl carbonate; (viii) CH_3I ; (ix) ammonia.

guanine from AICA. In 1976, isoguanine was discovered as an unexpected product in a synthetic process of guanine by Yamazaki⁴⁶ *et al.* from AICA. Isoguanine was obtained from an intermediate substance 4-cyanamido-imidazole-carboxamide (20, Fig. 7) in weakly alkaline or neutral conditions. Whereas in alkaline conditions, the compound 20 was converted into guanine. They synthesized isoG depending on the protected AICA riboside (25, Fig. 8), in low yield with three plausible and non-isolated intermediates (27, 28, 30, Fig. 8). Subsequently in 1987, Reese⁹ *et al.* reported that the compound 30 can also be obtained from compound 25 *via* the intermediate putative methoxyacetyl thioureido derivative 29 (Fig. 8). Moreover, in 1987, enlightened by a synthesis of 1-methyl-isoG, Chern¹¹ *et al.* proposed that an N,N' -dicyclohexylcarbodiimide (DCC)-mediated cyclodesulfurative method would allow milder reaction conditions and produce a higher yield of isoG. They reported an improved, more efficient route in milder reaction conditions using AICA 32 and its riboside 4 to synthesize isoguanine and isoG, respectively (Fig. 9). When AICA/AICA riboside 32/4 reacts with benzoyl isothiocyanate in

dimethylformamide (DMF) at room temperature, it can be converted to benzoylthiourea intermediates 33/34, respectively. Next, DCC was added to this system and isoguanine/isoG can be obtained *via* stirring, washing and separation. And isoG can be obtained in a high overall yield of 77%. In 1991, they also synthesized several 1-substituted isoG through similar DCC-mediated ring-closure reactions.⁴⁷

In summary, as shown in Table 1, over time, the syntheses of isoG/isoguanine have developed toward milder reaction conditions and higher yield. To date, the most outstanding synthetic route to isoG is that involving 6-chloroxanthosine by Napoli⁴⁸ *et al.*, with a yield of 80%, under mild conditions.

4 The modified isoG

In the above sections, we have discussed the discovery of natural isoG, as well as a variety of synthetic methods. isoG contains a nitrogenous base (a purine ring) and a carbohydrate residue. Minor modification to the base or sugar may lead to great differences in the higher structures and properties compared to the parent compound. Thus, a number of base-

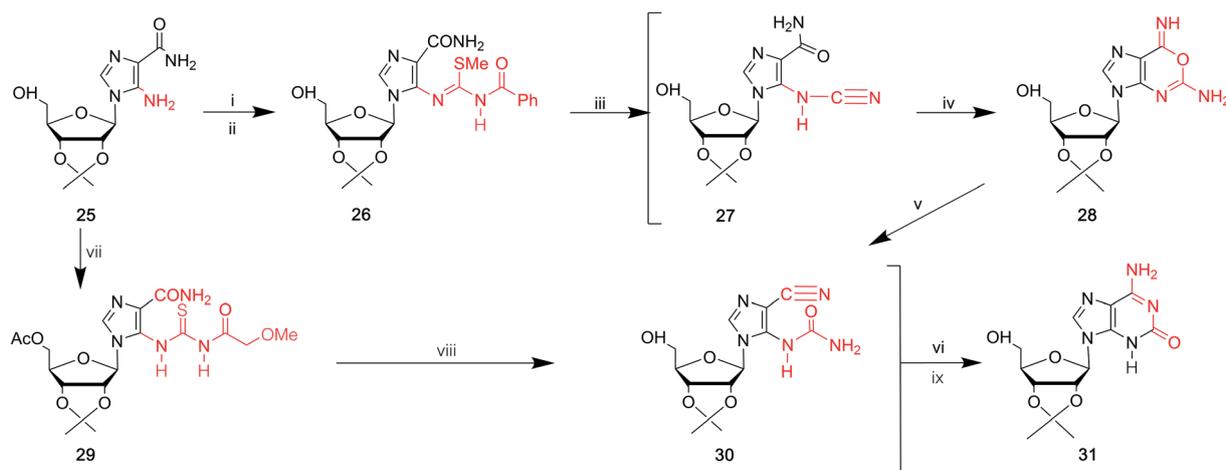


Fig. 8 The syntheses of isoG from the protected AICA riboside. (i) Benzoyl isothiocyanate; (ii) methyl iodide, alkali; (iii)–(vi) 0.1 M NaOH, heating, reflux; (vii) acetic anhydride, in pyridine solution, then methoxyacetyl isothiocyanate; (viii) mercury(II) perchlorate, pyridine, in tetrahydrofuran solution, room temperature, then methanolic ammonia; (ix) N^1,N^1,N^3,N^3 -tetramethylguanidine, water, in tetrahydrofuran, room temperature.



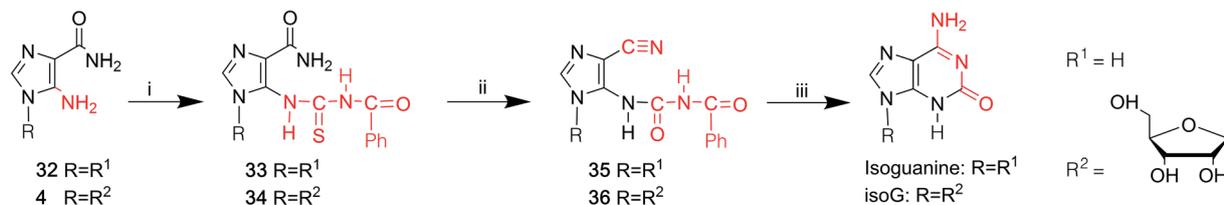


Fig. 9 Syntheses of isoguanine/isoG from AICA and its riboside. (i) Benzoyl isothiocyanate, DMF, room temperature; (ii) dicyclohexylcarbodiimide, DMF, room temperature; *vacuo*, 50 °C, hot toluene; (iii) ethanol, 33% aqueous ammonia.

modified (Fig. 10) or sugar-modified (Fig. 11) analogs of isoG have been discovered in nature or synthesized. Summaries are shown in Table 2. And some of their properties are discussed in the following part.

4.1 Base-modified isoG

4.1.1 1-Methyl-isoG (doridosine). *Dorid nudibranchs* are shell-less sea slugs. Interestingly, they have long lives despite lacking the typical protection given by a shell. Thus, there has been significant conjecture about their defense mechanism. In 1979, a tissue analysis by Fuhrman⁴⁹ *et al.* revealed that only the digestive gland extract is toxic, and this acts as the unique defense mechanism of *Dorid nudibranch*. In 1980, Quinn⁵⁰ *et al.* isolated a new pharmacologically active substance from the Australian sponge *Tedania digitate*, in a yield of 0.71% from the crude extract. They also developed a synthesis of doridosine from AICA riboside (4, Fig. 3) by acetylation in four steps, giving a yield of 39%. Meanwhile, they identified doridosine as identical to 1-methyl-isoG (37, Fig. 10) by spectrum, chemical degradation, synthetic analysis. In 1981, Fuhrman⁵¹ *et al.* concluded that the substances isolated from nudibranch and sponge are indeed the same. Moreover, 1-methyl-isoG 37 has been shown to have several pharmacological effects related to hypotension, hypothermia and anti-inflammatory, and these will be discussed in Section 6.3.

4.1.2 2-Methoxyadenine (spongosine). Nucleosides from marine organism have been found to have various biological activities, and this has aroused great interests among scholars. In a study of Caribbean sponge *Cryptotethya crypta* in Florida,^{52,53} a mixture of two unnatural nucleosides spongothymidine and spongouridine were obtained. And these were identified as 3- β -arabofuranosides of thymidine and uridine, respectively. The development of materials has promoted elucidation of the composition and structures of these purines and the carbohydrate. Then in 1951, Bergmann⁵³ *et al.* firstly isolated spongosine (38a, Fig. 10) from *Cryptotethya crypta*. It was identified as pentosylmethyl-aminooxypurine. In 1956, they determined the structure of β -D-ribose of 2-methoxy-6-aminopurine (2-methoxyadenine) using UV absorption spectra and the synthesis of 2-methyladenine.⁵⁴ They concluded that it is not only the first methoxy purine found in nature, but also the first occurrence of methoxy substance isolated from animals. In addition, in 1994, Searle⁵⁵ *et al.* firstly identified 2'-deoxy-spongosine [9 β -(2'-deoxy-D-ribofuranosyl)-2-methoxyadenine, 38b, Fig. 10] as a natural product from Western Australia.

4.1.3 More analogs. In terms of the great effects of 1-methyl-isoG 37, many analogs of 1-methyl-isoG have been prepared

from the protected imidazole nucleoside to explore the structure–activity correlation. As shown in Fig. 10, some *N1*-substituted isoG 39 have been developed to explore the importance of *N1*-methyl-substituted compounds in marine products. Some of these have been synthesized in good yield, by Bartlett⁵⁶ *et al.* in 1981 and Chern⁴⁷ *et al.* in 1991. In addition, in 1981, using 1-methyl-isoG 37 as a starting material, C8-substituted-1-methyl-isoG 40 was prepared by Bartlett⁵⁶ *et al.* Their effects on cardiovascular response and muscle relaxation, *etc.* will be discussed later in Section 6.3. Moreover, in 2017, Thomas⁵⁷ *et al.* identified 8-oxo-1-methyl-isoG 41a and 3'-*O*-acetyl-8-oxo-1-methyl-isoG 41b as natural derivatives of isoG in a sponge from the Northeastern Atlantic, *Clathria (Microcionia) strepsitoxa*. And these compounds showed no significant cytotoxic or antimicrobial activity. In addition, in 1994, Lee⁵⁸ *et al.* synthesized a series of 6-substituted isoG derivatives 42 and isoG by one-step nucleophilic substitution from the compound 15 (Fig. 6). These were synthesized to explore the antitumor activities, and the results will be discussed later in Section 6.3.

8-Aza-7-deazapurines and their 7-halogenated derivatives have been reported to possess excellent biological and physical properties, such as antiparasitic and antiviral effects, because of their structural similarity to purines in nucleic acids. And earlier studies showed that 7-substituents of 8-aza-7-deazapurine nucleosides and 7-deazapurine nucleosides incorporated oligodeoxynucleosides could increase antiparallel strand (aps) duplexes stability.^{59–66} Thus, in 2003 and 2007, Seela^{67,68} *et al.* synthesized 7-halogenated 8-aza-7-deazaisoguanosine derivatives 43 and 7-halogenated 7-deazaisoguanosine derivatives 44 from 8-aza-7-deazapurine-4,6-diamine riboside derivatives. Their tautomerism will be discussed later in Section 5.1. Moreover, in 2004, 8-aza-3-deazaisoG 45 was synthesized by Jeselnik⁶⁹ *et al.* to study the influence of the lack of *N3* on the hydrogen-bonding modes. 8-Azapurine nucleosides, including 8-azaadenine, 8-azaguanine, and their corresponding nucleosides, have been reported to be fluorescent. And they show various applications as fluorescent probes and in biochemical process.^{70–72} Consequently, in 2010, Seela⁷³ *et al.* synthesized 8-aza-2'-deoxyisoguanosine (z^8 isoG_d, 46) from the protected 8-aza-2'-deoxyguanosine (z^8 G_d) in a yield of 27%. z^8 isoG_d shows great fluorescence that becomes stronger in alkaline conditions than at neutral pH.⁷³ Moreover, 8-azaisoguanine 47 and its *N*-methyl derivatives *N3*-, or *N8*-methyl-8-azaisoguanine and *N3,N8*-dimethyl-8-azaisoguanine 48–50 were synthesized in the method of Fox⁷⁴ *et al.* In 2012, Wierchowski⁷⁵ *et al.* reported these compounds 48–50 also showed



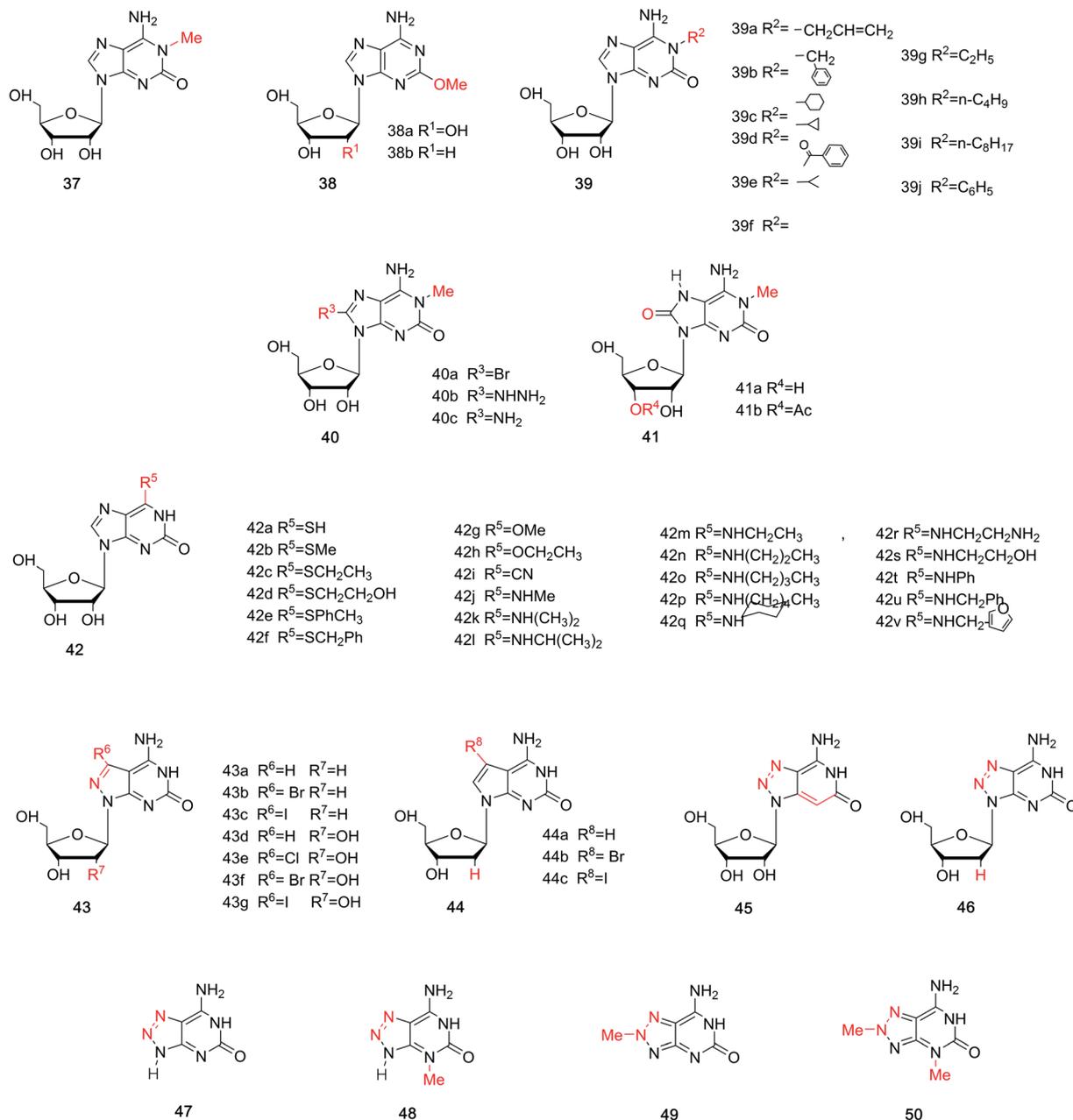


Fig. 10 Base-modified derivatives of isoG/isoguanine.

fluorescence, which is highly sensitive to the microenvironment.

4.2 Sugar-modified isoG. The structural modifications of isoG results in significant changes to its structure and functions. In addition to the previously discussed modification to the heterocycle, the glycosylation of isoG has also been studied. For example, as shown in Fig. 11, isoG_d **51a** was first synthesized by Kazimierzuk⁷⁶ *et al.* in 1973. Then in 1991, Seela⁷⁷ *et al.* improved the synthesis of isoG_d from 2-halogenated-isoG_d, in a yield of 52–53% by photochemical methods. isoG_d has been reported to be an oxidative damage product of DNA. Oxidation shows great impacts on cells, and ROS may affect multiple cellular locations, such as the connection between DNA and

proteins, as well as bases mutations. In 1995, Kamiya²⁹ confirmed 2-hydroxy-2'-deoxyadenosine triphosphate (2-OH-dATP) to be an oxidative product of dATP. But it is a minor product compared to 8-OH-G, which is generally believed as a significant marker of DNA oxidation, after γ -ray irradiation and exposure to the Fenton reaction.

Fluorine-substituted nucleosides have been reported to be efficient antiviral compounds. The fluorine-carbon bond is particularly stable, and its introduction to the sugar moiety can increase the lipophilicity of a drug.⁷⁷ Fluorine is generally incorporated into the 2'-position of the sugar, and it has the near radius as hydrogen and can exist in duplexes. Apart from these, fluorine also has a significant effect on the sugar



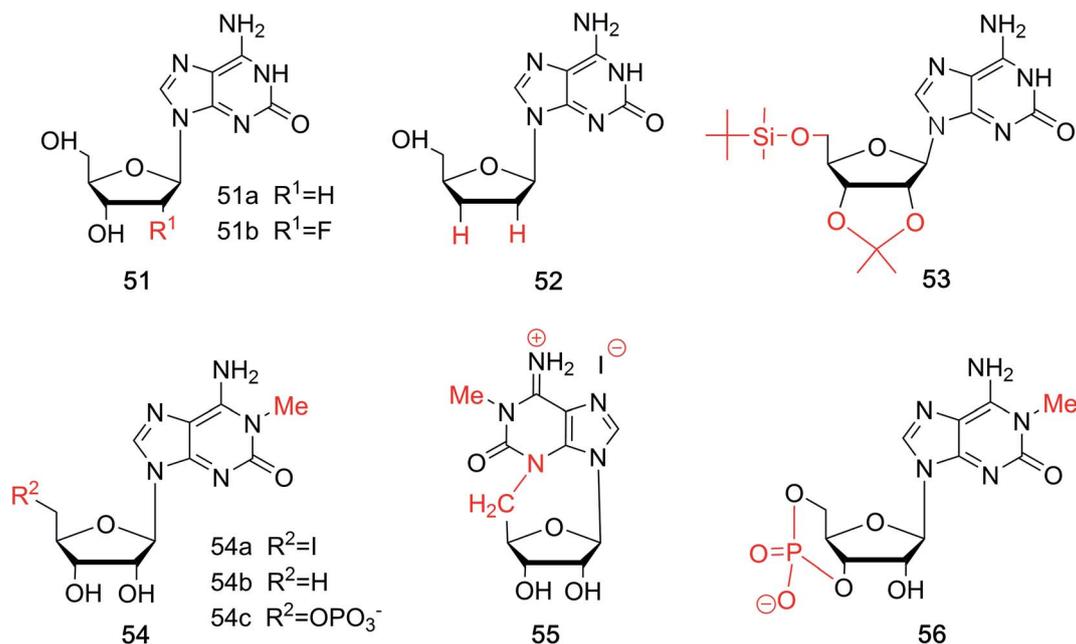


Fig. 11 Sugar-modified derivatives of isoG.

conformation because of its strong electronegativity. Based on the characteristic of fluorine mentioned above, through deamination of 2,6-diamino-2'-deoxy-2'-fluoropurine nucleoside, Seela⁷⁸ *et al.* first synthesized 2'-fluorine-2'-deoxyisoguanosine

(^FisoG_d, **51b**, Fig. 11) in a yield of 79%. Studies concerning the stability of its duplexes will be discussed in Section 5.3.4.

Some 2'3'-dideoxynucleoside derivatives have been reported to show effects such as inhibiting reverse transcription.^{79,80}

Table 2 Derivatives of isoG referred above found in nature or by synthesis

Analogs	Naming	From	First	Authors	References
Base-modified	1-Methyl-isoG 37	Natural	1979/1980	Fuhrman <i>et al.</i> /Quinn <i>et al.</i>	49 and 50
	Spongosine 38a	Natural	1951	Bergmann <i>et al.</i>	52
	2'-Deoxyspongosine 38b	Natural	1994	Searle <i>et al.</i>	55
	N1-Substituted isoG 39	Syn ^a	1991/1981	Chern <i>et al.</i> /Bartlett <i>et al.</i>	48 and 56
	8-Substituted-1-methyl-isoG 40	Syn ^a	1981/2017	Bartlett <i>et al.</i> /Firsava <i>et al.</i>	56 and 57
	8-Oxo-1-methyl-isoG 41a	Natural	2017	Thomas <i>et al.</i>	57
	3'- <i>o</i> -Acetyl-8-oxo-1-methyl-isoG 41b	Natural	2017	Thomas <i>et al.</i>	57
	6-Substituted isoG 42	Syn ^a	1994	Sung Jun Lee <i>et al.</i>	58
	7-Halogenated-8-aza-7-deaza-isoguanosine derivatives 43	Syn ^a	2003/2007	Seela <i>et al.</i>	67 and 68
	7-Halogenated-7-deaza-isoguanosine derivatives 44	Syn ^a	2007	Seela <i>et al.</i>	68
	8-Aza-3-deaza-isoG 45	Syn ^a	2004	Jeselnik <i>et al.</i>	69
	z ⁸ isoG _d 46	Syn ^a	2010	Seela <i>et al.</i>	73
	8-Aza-isoguanine 47	Syn ^a	1970	Fox <i>et al.</i>	74
	N3-Methyl-8-aza-isoguanine 48	Syn ^a	1970	Fox <i>et al.</i>	74
	N8-Methyl-8-aza-isoguanine 49	Syn ^a	1970	Fox <i>et al.</i>	74
	N3,N8-Dimethyl-8-aza-isoguanine 50	Syn ^a	1970	Fox <i>et al.</i>	74
	Sugar-modified	isoG _d 51a	Syn ^a	1973	Kazimierczuk <i>et al.</i>
^F isoG _d 51b		Syn ^a	2015	Seela <i>et al.</i>	78
2'3'-Dideoxy-isoG 52		Syn ^a	1999	Kim <i>et al.</i>	81
5'-(<i>tert</i> -Butyldimethylsilyl)-2',3'- <i>O</i> -isopropylidene-substituted isoG 53		Syn ^a	1995	Davis <i>et al.</i>	82
5'-Substituted-1-methyl-isoG 54		Syn ^a	1981	Bartlett <i>et al.</i>	56
3,5-Anhydro- <i>l</i> -methyl-isoG hydriodide 55		Syn ^a	1981	Bartlett <i>et al.</i>	56
1-Methyl-isoG cyclic 3',5'-phosphate 56		Syn ^a	1981	Bartlett <i>et al.</i>	56

^a Synthesized.



Inspired by these findings, in 1999, Kim⁸¹ *et al.* synthesized 2',3'-dideoxy-isoG (52, Fig. 11) from G in a yield of 10%. And studies on its biological activities are ongoing. Besides, in 1995, the protected isoG derivatives 5'-(*tert*-butyldimethylsilyl)-2',3'-O-isopropylidene-substituted isoG (53, Fig. 11) has been prepared by Davis⁸² *et al.* to study the self-assembly of isoG. So hydrogen bonds of isoG can be researched at different temperature and in different solvents. 1-Methyl-isoG (37, Fig. 10) has been identified as a natural product. And in 1981, using 1-methyl-isoG as a starting material, Bartlett⁵⁶ *et al.* prepared several N9-substituted doridosine compounds (54–56, Fig. 11). The effects of these compounds are shown later in Section 6.3. All the derivatives mentioned above can be found in Table 2 for their original sources.

5 Structure properties of isoG

The discovery of isoG and isoguanine in nature and their structural determination have been discussed above. In addition, we have discussed the low content and low extraction efficiency of these compounds in natural products, which led to the development of a series of synthetic methods. Next, some of the special structural properties of isoG, including its tautomerism at the single molecule level and the resulting unnatural pairing in nucleic acids, its self-assembled structures such as tetramers, the more highly ordered “sandwich-like” decamer structure, the isoG₄ quadruplexes formed in helix, the parallel-stranded (ps) and antiparallel-stranded (aps) chains containing isoG will be discussed as follows.

5.1 Tautomerism enol–keto of isoG

The tautomeric phenomenon of nitrogen heterocycle has been widely investigated, for example, the N1H, N3H and O6H forms of guanine.^{83,84} In a study of poly (isoG) by Golas⁸⁵ *et al.* in 1976, the study of the tautomeric forms of isoG seems particularly important to identify correct base pairings containing isoG. In 1976, Sepiol⁸⁶ *et al.* reported that the tautomerism of isoG is affected by concentration, temperature, and solvent polarity. As the polarity of the solution changes, the enol ratio can vary from 5% in aqueous medium to 80% in polar solvents. They proposed that isoG is often present in solution as a mixture of enol and keto forms. isoG shows a UV absorption 310 nm for the N1H keto form and 270 nm for the O2H enol form. In general, the enol form (O2H) is favored in lower polarity solvents such as dioxane or at higher temperature in chloroform solution.^{67,86} In contrast, the N1H-2-oxo-6-amino form (I, Fig. 12) is mainly present in polar solvents. However, the N3H form of isoG was not found until Eschenmoser and co-workers confirmed its existence in 1993.^{87–89} In 1997, Switzer⁸⁴ *et al.* reported that isoG may possess nine tautomerism forms (I–IX, Fig. 12). The tautomeric forms I–V are the five most stable forms by gas-phase calculation. They also concluded that *cis*-enol configuration of isoguanine is more stable than *trans*-enol configuration. Notably, if isoG is methylated, the enol–keto shift is faster, and the ketone 6-amino-N3H form of 9-methylisoguanine has been confirmed in solid state by Banerjee⁹⁰ *et al.* in 1978. However, researchers have not successfully isolated the enol

form of 9-methylisoguanine.⁹⁰ In 2012, Schaefer⁴⁴ *et al.* reported that the *cis/trans* enol isoguanine can be converted to the corresponding N1H and N3H ketone forms, respectively (Fig. 5).

In a study of pyrazolo[3,4-*d*]pyrimidine by Seela^{68,91} *et al.* in 2007, it is concluded that for isoG and its derivatives, a higher ratio of the ketone form results in a stronger mismatch recognition ability in duplexes. Compared to isoG_d (51a, Fig. 11) and 7-deaza-isoG_d 44a, 7-halogenated-7-deaza-isoG_d (44b, 44c, Fig. 10) prefers the ketone form, thus, the compounds 44b and 44c show a stronger mismatch ability. The proportion of the ketone form of 8-aza-7-deaza-isoG and its 7-halogenated derivatives (43, Fig. 10) are higher than the original isoG, so the mismatch recognition ability is higher. However, the effect of 7-halogenation on the tautomerism of 8-aza-7-deaza-isoG 43d is less than that on 7-deaza-isoG_d 44a. Thus, 7-halogen substitution should not change the base recognition of 8-aza-7-deaza-isoG derivatives 43a/43d. Pyrazolo[3,4-*d*]pyrimidines 43d–43g is more enolized than pyrrolo[2,3-*d*]pyrimidines 44, while isoG itself shows the highest enol content. For compound 43d–43g the equilibrium constants [K_{TAUT} (keto/enol)] range from 400 to 1200, whereas for compound 43a–43c they range from 1000 to 10 000. Therefore, these compounds are expected to exhibit better mismatch recognition than isoG.

5.2 Mispairing

Many researchers have studied the tolerance of base mutation that do not affect the original complementary pairing of oligonucleotides, for the purpose of expanding the genetic alphabet, designing tools for monitoring biological mechanism, and creating new gene systems. A common method is to study the pairing by inserting irregular bases. As mentioned above, isoG is an unnatural nucleoside, so the study of its base pairing is of great significance for subsequent applications. In 1962, Rich⁹² *et al.* hypothesized that isocytosine (isoC) and isoG, the isomers of C and G, respectively, may make up a third Watson–Crick base pair. The complementary isoG–isoC pair would form a similar geometry to that of G–C. But there were questions as to why the isoG–isoC is not a constituent of nucleic acids. Subsequently, in 1990, Benner⁹³ *et al.* confirmed the existence of the isoG–isoC base pair, as well as a second pair, isoG–T, *in vitro* polymerase experiments. Afterwards, evidence for isoG–isoC pair without enzymes was obtained by Horn⁹⁴ *et al.* and Switzer⁹⁵ *et al.* in 1995. The base pairing of isoG in the enol form is stronger than that in the ketone form, which is the first experimental evidence that tautomerism can lead to base pairing differences. The keto–enol tautomerism of isoG and the possible *cis* and *trans* conformers result in various types of mispairing and different hydrogen-bonding modes. It has been reported that in different tautomeric forms, isoG_d can pair with several natural bases, deoxycytidine, deoxythymidine, deoxyuridine, deoxyguanosine, and deoxyadenosine (C_d, T_d, U_d, G_d, A_d, respectively), as well as with some special unnatural bases such as isoC_d and 5-aza-7-deaza-G_d in different templates (Fig. 13).^{84,94,96–98} On the basis of the tautomerism of the bases described above, the possible hydrogen-bonding of isoguanine–isocytosine, isoguanine–uracil, isoguanine–cytosine were put



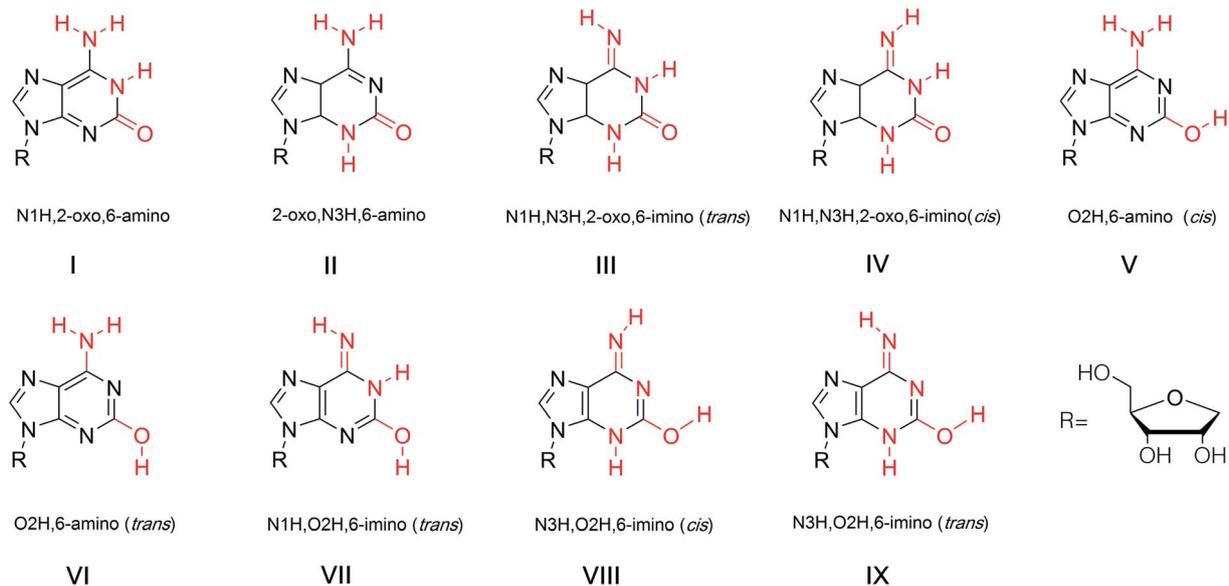


Fig. 12 Tautomerism of isoG.

forward (Fig. 13).⁸⁴ And in 2003, the impact of the temperature and the adjacent base pairs on base pairings of isoguanine was investigated by Maciejewska⁹⁹ *et al.* They concluded both affect base-pairing by changing the polarity of the solution around the

isoguanine to cause tautomerism. The temperature was found to have only a slight influence on the base pairing. The 3'-adjacent base affects the tautomerism, while the 5'-neighbor

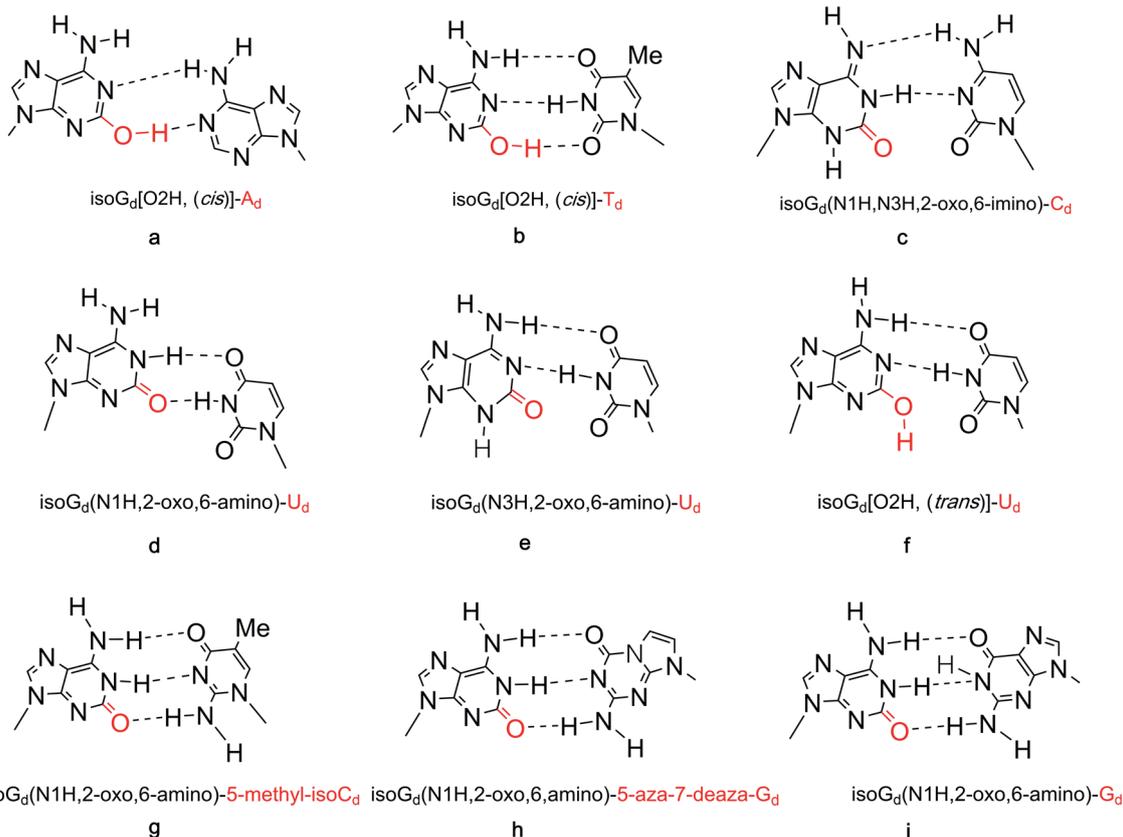


Fig. 13 Base pairs of isoG.



can not, because the 5'-terminal is not at the polymerase active site for the stacking with template.

In 1986, Jaworski⁹⁶ *et al.* carried out experiments to explore the interaction energies of isoguanine–isocytosine and isoguanine–uracil, and compared them with energies of natural base pairs guanine–cytosine, adenine–thymine/adenine–uracil. It was found that isoguanine–uracil interaction energy is smaller than that of guanine–cytosine and isoguanine–isocytosine but stronger than that of adenine–thymine/adenine–uracil pairs. Additionally, isoguanine–isocytosine pairing is slightly stronger than guanine–cytosine pairing. And isoG_d-5-methyl-isoC_d pair was reported to be isoenergetic with the G–C pair, and each base can effectively identify mismatches in synthetic oligonucleotides, by Horn⁹⁴ *et al.* in 1995 and Switzer⁸⁴ *et al.* in 1997. The isoguanine–cytosine and uracil–adenine base pairs have the same energy. In 1997, using *ab initio* data from calculations combined with thermodynamic data, Switzer⁸⁴ *et al.* concluded the reason why isoguanine–isocytosine shows unnatural stability may be that isoguanine adopts a carbonyl-imino form.

Isoguanine is a modified base, so its enzymatic recognition and removal have aroused wide interest among scholars. Various DNA and RNA enzymes can catalyze the recognition of different bases by hydrogen bonding interactions. Thus, the enzymological properties of these varied base pairs have been investigated to explore whether they are incorporated into duplexes by DNA and RNA polymerases, and, if so, under what conditions. It has been demonstrated that various enzymes catalyze the entry of isoG into the nucleotide chains *in vitro* (Table 3). For the first time, Switzer¹⁰⁰ *et al.* reported that d-isoGTP can be incorporated opposite T/isoC by the Klenow fragment (KF) of *E. coli* DNA polymerase (pol) I and T7 RNA polymerase in 1989. Then in 1993, they reported that in different templates, avian myeloblastosis virus (AMV) reverse transcriptase can enhanced the formation of isoG–isoC, and isoG–U base pairs *via* T7 RNA pol.¹⁰¹ And T7 RNA pol has a preference in catalyzing the incorporation of U over isoC. However, T4 DNA pol rejected isoguanine's incorporation under reported experimental conditions. Moreover, in 1993, Dervan¹⁰² *et al.* reported that 5-methyl-isoC_d can direct the incorporation of N6-(6-aminoethyl)-isoG into the transcribed RNA by T7 RNA pol. And this amino group affords site-specifically modified RNA sequences beneficial for future studies after transcription. Until then, the possibility of isoG opposite A/G and the incorporation by mammalian polymerase were not investigated. Subsequently, in 1995, *in vitro* DNA synthesis, Kamiya³¹ *et al.* reported that deoxythymidine monophosphate (dTMP) and deoxyadenosine monophosphate (dAMP) can be incorporated

to pair with isoG by KF of *E. coli* DNA pol I, recombinant rat DNA pol β and calf thymus DNA pol α. And deoxyguanosine monophosphate (dGMP) can pair with isoG by KF of *E. coli* DNA pol I. They also concluded that d-isoGTP can mismatch with T/C (showing a preference for T) using mammalian (calf) DNA pol α.²⁹ Whereas d-isoGTP can only pair with T by KF of *E. coli* DNA pol I. In 1996, using calf thymus DNA pol α and recombinant rat DNA pol β, they confirmed the dTMP and deoxycytidine monophosphate (dCMP) pairs with isoG.¹⁰³ Whereas dTMP and dGMP were incorporated to form base pairs with isoG by KF of DNA pol I. In addition, human immunodeficiency virus 1 reverse transcriptase (HIV-1RT) has a high tolerance for nonstandard bases, indicating that it promotes nonstandard base pairing containing isoG_d, and thus preventing HIV-infected cells from being recognized and dying in the cell's S phase.^{104,105} Then in 1998, Lutz¹⁰⁶ *et al.* reported that compared to eukaryotic DNA polymerases, the incorporation of T or U by HIV-1RT to mismatch with d-isoGTP is more efficient, whether dATP exists or not. In 2000, Kamiya^{107,108} *et al.* reported the 2-OH-dATP can pair with T and G using the α (catalytic) subunit of *E. coli* DNA pol III and KF of DNA pol I. Above all, the incorporation of isoG depends on the types of polymerases, as well as the sequence contexts in various templates.

In 1975, Mantsch⁴¹ *et al.* reported that isoguanine nucleotides are not recognized by the mitochondrial endometrial translocation enzyme system, but they can be well recognized by mitochondrial phosphatases in the inter-membrane space. They are also involved in phosphoryl group transfer reactions. These reactions are catalyzed by pyruvate kinase, phosphofructokinase and hexokinase. Additionally, in 1998, Kamiya¹⁰⁹ *et al.* reported that isoguanine opposite thymine/cytosine cannot be moved by a glycosylase-type mechanism in rat organs. In bacteria such as in *E. coli*, there are some specific enzymes for the removal of unnatural base from damaged nucleic acid. In 2011, Rauschel¹¹⁰ *et al.* reported that cytosine deaminase (CDA) from *E. coli* can catalyze the deamination of isoguanine to xanthine. However, in 2000, Kamiya^{111,112} *et al.* demonstrated that the removal of isoguanine by the MutY and MutM proteins of *E. coli* is slow because of poor affinity. Surprisingly, they reported that among the known akin nucleotide substrates, the nucleotide forms of isoG (2-OH-ATP and 2-OH-dATP, where ATP is adenosine triphosphate and dATP is deoxyadenosine triphosphate) are substrates with the highest affinity for the human MTH1 (hMTH1) protein, which is a hydrolytic enzyme.^{113,114} Moreover, 2-OH-dADP and 8-OH-dGDP (dADP and dGDP are deoxyadenosine diphosphate and deoxyguanosine diphosphate, respectively) can competitively inhibit the effect of hMTH1 on 2-OH-dATP and 8-OH-dGTP (dGTP is

Table 3 Enzymes incorporating isoG opposite different natural/unnatural nucleosides

Enzymes	<i>E. coli</i> DNA pol I	<i>E. coli</i> DNA pol III	DNA pol β	DNA pol α	T7 RNA pol	AMV reverse transcriptase	T4 pol	HIV-1RT
Mispairing with isoG	T/isoC/A/G	T/G	T/A/C	T/A/C	U/T/isoC	isoC	— ^a	T/U
References	29, 31, 100, 103, 107 and 108	107 and 108	31 and 103	29, 31 and 103	100 and 101	101	101	106

^a T4 pol do not catalyze any nucleosides opposite isoG under experimental conditions.



deoxyguanosine triphosphate), which indicates that hMTH1 takes effect on the damaged substances *via* the same receptor.¹¹⁴

5.3 Self-assembly of isoG

We have discussed the tautomeric forms of isoG in different solvents and the various base pairs catalyzed by different DNA and RNA enzymes above. Now, we will describe several types of advanced structures such as tetramers, decamers, oligonucleotides, and ps/aps strands that form based on the self-assembly properties of isoG. Self-assembly is a process where substances combine into a supramolecular system through intermolecular forces, such as electrostatic and hydrophobic interactions. The discoveries of poly(isoguanic acid), the oligonucleotide d(T₄-isoG₄T₄), and G-quartets indicate that isoG may also self-assemble into supramolecular structures. In fact, both G and isoG can self-assemble into supramolecular systems, such as supramolecular gel by hydrogen-bonding, π - π stacking, and hydrophobic interactions. And these structures show a wide range of applications in material science, medical chemistry, and nanotechnology *etc.* As mentioned before, isoG differs from G in that the C2 carbonyl and C6 amino groups are transposed. This minor change causes significant differences between isoG and G. In particular, the variation in hydrogen bond donors and acceptors affects the hydrogen-bonded systems, resulting in great changes in self-assembled supramolecular structures. For G, the angle between the hydrogen bond donor (N1 amide and N2 amino) on its Watson-Crick face and the receptor (N7 and O6) in the Hoogsteen face is 90°, and this is favorable for the formation of planar tetrads (Fig. 14).^{115,116} On the contrary, the isoG donor-receptor angle of 67° prefers cyclic pentad (Fig. 14). In 1994, Davis⁸² *et al.* raised that for isoG, there are two prerequisites for self-association. One is that the hydrogen-bonding receptors of isoG (O2-C2-N3) are close to 180°, whereas the N7-C5-C6-O6 bond of G is nonplanar. And the ribose of isoG, as well as the hydrogen-bond acceptors, are more likely to form hydrogen bonds in the lower position of

purine.^{117,118} The other prerequisite is the formation of hydrogen bond between the N6H proton and adjacent isoG monomer O2'. In contrast, G is unlikely to form such a hydrogen bond because the hydrogen bond acceptor O6/N7 and the glycosyl group are in opposite positions in the heterocycle.⁸² This is also an important reason why isoG self-assembled oligonucleotides are more stable than the corresponding G self-assembled oligonucleotides.¹¹⁹ It has also been proposed that isoG has three variable N protons that are important for supporting the self-assembly.⁸² C6NH_A and C6NH_B (Fig. 14) rotates slowly, allowing the exocyclic amino group to form a strong hydrogen bond, and the third proton N1H is confirmed based on nuclear Overhauser effect (NOE) and UV studies in CDCl₃.⁸²

5.3.1 The tetramer model of isoG. As shown in Fig. 14, without cations, some free G lipophilic derivatives such as G_d can only self-assemble to different ribbons by taking N3 as a hydrogen-bonding receptor.¹²⁰ And in 2002, Gottarelli¹²¹ *et al.* reported that some lipophilic derivatives are potential gelators. However, for isoG, self-assembly into tetramers or decamers without cations is possible. In the presence of cations, G-quartet was identified as the basis of the self-assembly of G.¹²² It is widely accepted that G-quartet takes shape *via* eight Hoogsteen hydrogen-bonds, and a central cavity containing four oxygens that bind to cation through ion-dipole interactions. The stability of G-quartets varies with different radii and charges of the cations. In 1978, Becker¹²³ *et al.* reported that G-quartets are stabilized in the presence of sodium ion (Na⁺) and potassium ion (K⁺). In 1994, using 2D nuclear Overhauser spectroscopy (NOESY) and 1D NOE experiments, the isoG tetramer, a basic unit of isoG self-assembled structures, was confirmed. When two nuclei (A and B) are close to each other in space and have strong mutual relaxation, NOE occurs. This means that when A is saturated by irradiation, its energy is transferred to B, so that the energy absorbed by B increases and the resonance signal also increases. The specific NOEs between the exocyclic C6NH₂

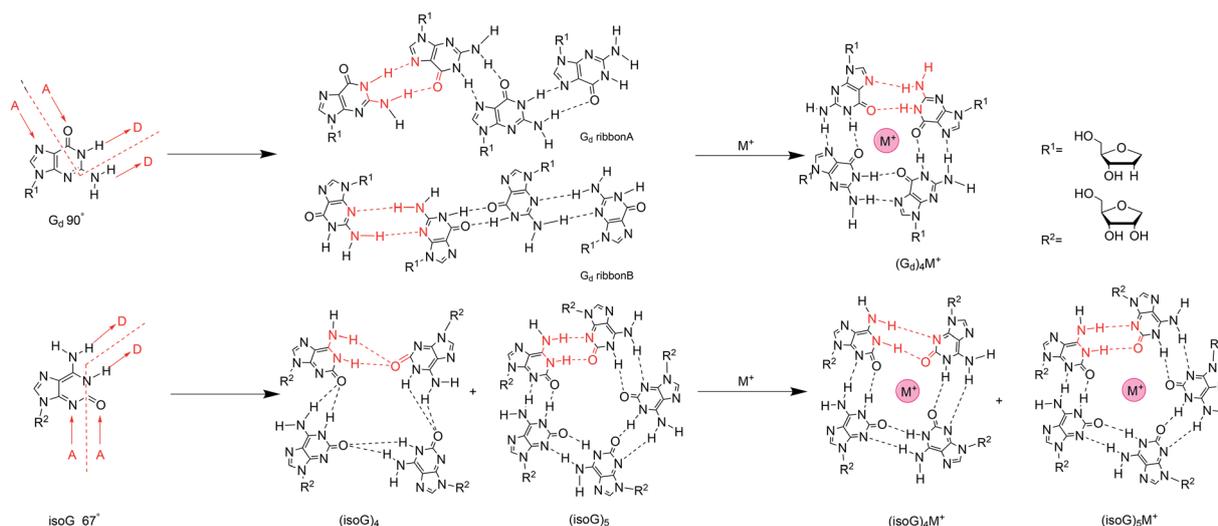


Fig. 14 Self-assembly of isoG and G with/without cations.



amino proton and the adjacent ribose H1' and H2' protons are confirmation of isoG self-association.⁸² Only the isoG tetramer model conforms with the observed NOEs.

Similar to the interaction of the cations with G, alkali ions have a great influence on the self-assembled structures of isoG. Different ions have been titrated into pure isoG mononucleoside in *d*₆-acetone to identify the different structures.⁸² Tetramerization with lithium ion (Li⁺) is unstable because Li⁺ is too small for the central cavity. Li⁺ may be combined with the O2 or N3 hydrogen-bond donors to hinder hydrogen-bond formation. A low concentration of Na⁺ can stabilize the tetramer, probably *via* ionic coupling. But no new NMR peaks have been observed with Na⁺, which indicates that no higher-ordered self-assemblies, such as stacking structures, are formed in the presence of Na⁺. Nevertheless, high concentrations of Na⁺ destroy isoguanine tetramers, probably for the same reason as that for Li⁺. Notably, the addition of K⁺/barium ion (Ba²⁺) form octamers, presenting a new peak with a sandwich structure.^{12,82} And K⁺/Ba²⁺ enhanced the thermal stability compared to the original tetramer. Moreover, the Ba²⁺ octamer is more stable than K⁺ octamer.

The hydrogen-bonding modes are different in the isoG tetramer in the presence and absence of cations. Next, the hydrogen bonds in alkali-metal-free isoG tetrad and pentad structures will be introduced. In 2003, Meyer¹²⁴ *et al.* proposed that the hydrogen-bond modes in tetrad and pentad are different (Fig. 14), one difference is that the N1–H1...O2 hydrogen bond (H bond) in the tetrad is longer than that in the pentad. The other is the tetrad N6–H6...O2 H bond forms a crossing hydrogen bond with the N1–H1...O2 H bond. Whereas two hydrogen bond, N1–H1...O2 and N6–H6...N3 are formed without crossing in the pentad. Besides, the second H bond N6–H6...N3 in pentad is shorter than the second bond N6–H6...O2 in tetrad. When the ions are added to the isoG solution, there are no crossing hydrogen bonds occurring. In other words, alkali ions can affect the crossing hydrogen-bond mode of the protected isoG tetramer (53)₄ to form

independent N6–H6...N3 and N1–H1...O2 hydrogen bonds (Fig. 14). This is not surprising because base-cation linkages are usually stronger than base–base linkages for isoG. That is, the cation interactions are the dominant contribution to the interaction energy. As a result of metal cations present in the cavity, the base–base hydrogen bond in the tetrad is shorter than in the pentad, and the distance between cation and O2 is also shorter. It indicates the hydrogen bond distance is shortened in the presence of alkali ions, and this effect is more pronounced for Li⁺ and decreases with increasing ion size. From Li⁺ to cesium ion (Cs⁺), the distance between O2 and the cation increases. Further, larger ions result in smaller connection energies between the ion and C2 carbonyl group.

The protected isoG tetramer (53)₄·M⁺ is stabilized by the hydrogen-bond system of the outer ring and the inner ring (Fig. 14). The inner ring is composed of four hydrogen bonds formed between the imino N1H proton and the C2 carbonyl oxygen of an adjacent isoG. The outer ring is comprised of hydrogen bonds between N6H and N3H of the adjacent monomer. Four nonbonded C2 oxygen atoms point to the cavity, interacting with metal alkali ions, and selectively bind to monovalent or divalent cations to form a stable complex. The presence of multiple metal ions causes different changes to hydrogen-bond model, and metal cations are often required to stabilize hydrogen-bonded macrocycles. The self-assembled structures of isoguanine with different metal ions varies. For example, the tetramer of isoG is strongly nonplanar except with Li⁺, whereas the pentamer is often planar. In fact, most nonplanar structures converge into planar structures, so most of the known tetrad/pentad structures are planar, which is more stable. However, there is no selectivity over monovalent and divalent ions. That is, there is no difference in affinity for different charge numbers. Moreover, the tetrad has strong selectivity and preference for small ions. However, for large ions, there is no preference between the tetrad and pentad. The energy of (isoG)₄·Li⁺ tetrad is higher than that of its pentad, whereas the energy of the pentad with a larger radius is higher

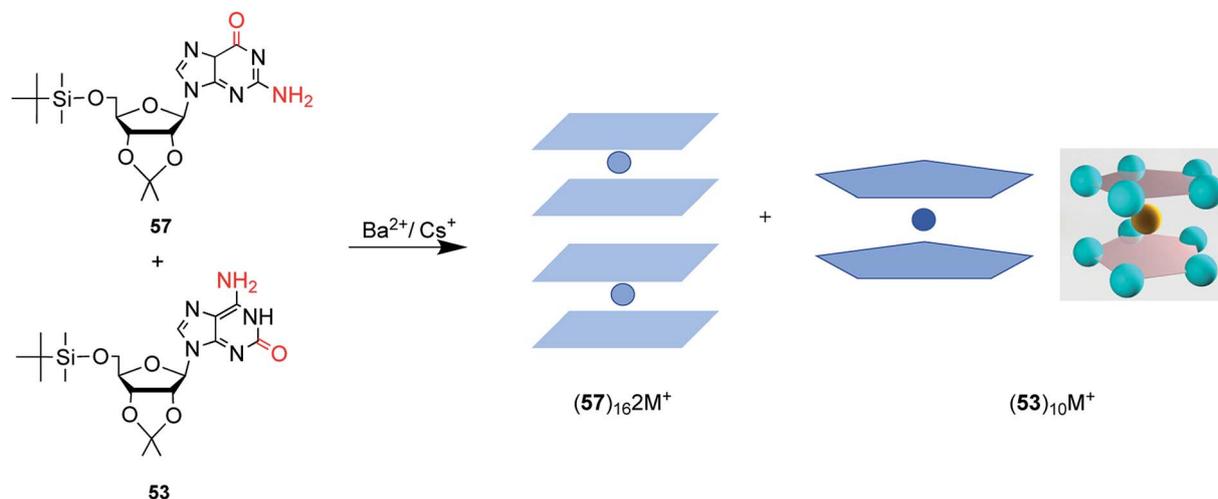


Fig. 15 The protected G and isoG self-assemble and stack into hexadecamer and decamer respectively in the presence of Ba²⁺.



than the corresponding tetrad energy. The energies of the planar tetrad structures are relatively low; *i.e.*, they are more stable. And the metal-free and (isoG)₅·K⁺/rubidium ion (Rb⁺) pentad energies are also relatively low. Li⁺ and Na⁺ ions are too small for the pentameric cavity, whereas Cs⁺ is too large to be in the cavity so it may sit over the isoG₅ pentad plane. The protected isoG (53, Fig. 11) can also self-associate in the gas phase and studies into this fact are ongoing.

5.3.2 Sandwiched structure formed by isoG. In 2000, by X-ray crystallography, Davis¹²⁵ *et al.* showed that the protected G (57, Fig. 15) can stack into an ordered hexadecamer. And solid-state decamers have been obtained with Ba²⁺ and strontium (Sr²⁺) in 2001 by his group.^{120,126–128} They also showed the role of organic anions in the cation-filled quadruplexes, which predominate in solution.¹²⁹ In 2000, they reported isoG, an isomer of G, can also stack into a sandwiched structure (53)₁₀·Cs⁺ (Fig. 15), comprising two hydrogen-bonded isoG pentads.¹³⁰ This structure forms in both solution and solid state. Two-side views of the crystal sandwich structure are shown (Fig. 16): a overlooking view showing the two pentameric tail-tail connections that directly overlap with ten chemically equivalent isoG units having Cs⁺ and ten carbonyl groups in the center; the side view shows a sandwich model with Cs⁺ as the bisector of the plane. In 1999, depending on conventional NMR, an inaccurate isoG pentamer structure was confirmed by Davis¹³¹ *et al.*, and was generally accepted for a long time. But advances in experimental instruments have revealed that the use of conventional NMR is insufficient to probe the distinction

between highly symmetrical supramolecular structures. And it is also difficult to determine the structure of a self-assembly system similar to its monomeric and oligomeric structures. Consequently, in 2007, they used the present diffusion NMR, which uses pulse field gradient techniques, to determine the exact size and molecular shape in solution.¹³² As a result, the wrong pentamer structure (53)₅·M⁺ was overturned, and the correct (53)₁₀·2M⁺ or singly charged decamer (53)₁₀·M⁺ structures were identified. Of the two structures, the latter (53)₁₀·M⁺ form is more commonly seen. If there are more cations, then the (53)₁₀·2M⁺ sandwiched structure is obtained, with one metal ion in the middle and the other one in the pentad. In addition, for (53)₁₀·2M⁺, another structure having two cations in the pentads was rejected because it is impossible on the basis of combined anion limited. (53)₁₀·2M⁺ can more easily bind anions than (53)₁₀·M⁺, but it has been shown that anions are not involved in the self-assembly process.

isoG is selective for the largest alkali cation, Cs⁺ (*r* = 1.67 Å), whereas G-quartets have a preference for K⁺ (*r* = 1.33 Å).¹³² Importantly, in the presence of Cs⁺, isoG only formed the (53)₁₀·Cs⁺ structure. However, for other cations such as Li⁺, (53)₁₀·M⁺ and (53)₁₀·2M⁺ structures are formed, and stay nonstop transition between each other.¹³² Notably, (53)₁₀·2K⁺ directly comes from two pentamers rather than by the addition of more K⁺. Furthermore, in 2014, Rodriguez¹³³ *et al.* reported that small dendritic molecules containing isoG can also self-assemble into a decameric nucleodendrimers, even without cations. And they demonstrated the existence of G analogs can promote the formation of decameric nucleodendrimers, but are not involved in decameric structures. Experiments to explore the abilities of different ions to induce nucleodendrimers identified that the isoG-based nucleodendrimers show a preference for Cs⁺ over K⁺, which reflects the fact that the cavity size is more suitable for the large radius of Cs⁺. In addition to the above, in 2002, from the NMR spectrum, Davis¹³⁴ *et al.* concluded that G and isoG self-assemble independently in the presence of Cs⁺/Ba²⁺, rather than form crosslinked structures (Fig. 15). However, in CD₂Cl₂ where there is no cation, G and isoG are crosslinked.^{13,129,134}

Moreover, in 2000, Davis¹³ *et al.* found that (D,L)-isoG (53, 59, Fig. 17) undergoes enantiomeric self-recognition in solution containing Cs⁺. Then in 2001, they found that the protected (D,L)-G (57, 58, Fig. 17) can self-associate with diastereoselectivity in Ba²⁺-containing solutions, yielding two homochiral G-quadruplexes (57)₁₆·2Ba²⁺·pic⁻ and (58)₁₆·2Ba²⁺·4pic⁻.¹²⁶ But in the presence of K⁺, (D,L)-G forms a diastereomeric mixture. As mentioned above, hydrogen bonds can take shape between neighboring isoG ribose residue to help conquer the entropic demands in relation to enantiomeric self-sorting, whereas this does not occur for G. Such hydrogen bonds can transfer conformational information between pentameric isoG to ensure homochiral conformation. And the achiral Cs⁺ promotes the enantioselectivity of isoG, thus promoting formation of the Cs⁺ sandwich meso decamer, (53)₅·Cs⁺·(59)₅·Ph₄B⁻, as determined by X-ray crystal structure analysis. However, enantioselectivity is not possible with other ions.

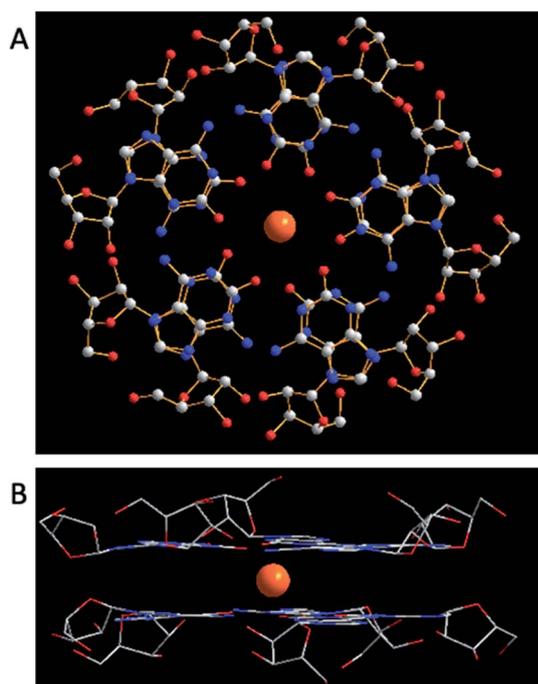


Fig. 16 (A) A top view of X-ray crystal structure of (53)₁₀·Cs⁺. Two pentamers directly overlap with ten chemically equivalent isoG units, having Cs⁺ (yellow) and ten carbonyl groups in the center. C atoms are grey, N atoms are blue, O atoms are red. (B) A sandwich model with Cs⁺ as the bisector of the plane. Anions are omitted for clearness.



5.3.3 Oligonucleotide aggregates $d(T_4\text{isoG}_4T_4)_4$. The tetramer and decamer structures of isoG have been summarized above, next, the isoG-based aggregated structures in duplexes will be talked about. In 1984, Blackburn¹³⁵ *et al.* reported that some $(G_d)_4$ aggregates are natural parts of telomeres. This has aroused great interest in G-quadruplexes. In 1989, Williamson¹³⁶ *et al.* found that oligonucleotides including short runs of G, such as $d(T_4G_4T_4)$, can form four-stranded nucleic acid structure named G-quadruplexes. And the fundamental structure is the G-quartet composed of four G units interacting *via* Hoogsteen hydrogen bonds in planar (Fig. 14). G-quartets stacked in a helical mode *via* π - π interactions are stabilized by cations such as K^+ and Na^+ (preference $K^+ > Na^+$). And with cations linked with O6 of G in the interior channel, G-quartets show a parallel/antiparallel orientation of strands. The oligonucleotides containing short runs of G can self-assemble into higher aggregates, which can be classified depending on their molecularity [*i.e.*, unimolecular (monomer), bimolecular (dimer), and tetraplex (tetramer)] or in terms of their strand orientation (*i.e.*, chair or basket) (Fig. 18a-f).¹³⁷⁻¹³⁹ In 1996, based on ion-exchange HPLC and circular dichroism (CD) spectroscopy, Seela¹⁴⁰ *et al.* demonstrated the identification of an isoG-containing oligonucleotide quartet $d(T_4\text{isoG}_4T_4)$, which is more stable than $d(T_4G_4T_4)$. This conclusion also applies to the comparison of thermal stability of their tetraplexes, *i.e.*, $d(T_4\text{isoG}_4T_4)_4$ is more stable than $d(T_4G_4T_4)_4$. Subsequently, in 1997, they proved that the stability of $d(T_4\text{isoG}_4T_4)$ against exonuclease hydrolysis is much higher than that of $d(T_4G_4T_4)$.¹⁴⁹ The same is true for their tetraplexes. In general, base residues can pair to cause reverse trimerization or tetramerization, but when the increasing energy arising from stacking or hydrogen bonding is greater than the energy lost because of the main degeneration, the double helix structure is generally more stable, and still able to maintain a stable Watson-Crick base pair model.

In 1997, by native polyacrylamide gel electrophoresis (PAGE), Roberts¹⁴¹ *et al.* reported that similar to G, isoG also shows a tendency to form tetraplexes, showing the similar monomer, dimer, tetramer structure (Fig. 18a-f). Notably, they found that isoG can even form mixed quartets with G, which may induce the formation of mixed tetraplexes. Moreover, isoG-tetraplex may show higher stability than G-tetraplex in the presence of K^+ . And isoG-tetraplex shows a preference for K^+ over Na^+ . Afterwards, in 1998, Seela⁹⁸ *et al.* concluded that oligonucleotide tetraplex $d(T_4\text{isoG}_4T_4)_4$ has an ion preference in the following order: $K^+ > Rb^+ > Cs^+ > Na^+ > Li^+$. In contrast, the order for

$d(T_4G_4T_4)_4$ is $K^+ > Rb^+ > Na^+ > Cs^+ > Li^+$. In 1999, Switzer¹¹⁵ *et al.* reported that ionic identity can determine the self-assembly of DNA strands into quadruplexes or pentaplexes. In the presence of Cs^+ , isoG self-assembles into pentaplexes. Whereas, in the presence of Na^+ and K^+ , it forms quadruplexes. However, it was found that G self-assembles into almost the same quadruplexes in Cs^+ and K^+ , which is consistent with the results of Sen¹⁴² *et al.* in 1993. Interestingly, in 2001, Seela¹⁴³ *et al.* found that 8-aza-7-deaza-isoG_d (**43a**, Fig. 10) can self-assemble into $d(T_4(\mathbf{43a})_4T_2)$. And the compound **43a** can form a pentaplex (Fig. 18g) in the presence of Cs^+ , and a tetraplex in the presence of Na^+ , K^+ and Rb^+ . This is different from $d(T_4\text{isoG}_4T_2)$ because the $d(T_4\text{isoG}_4T_2)$ can self-assemble into both four-stranded and five-stranded structure in Rb^+ solution. In addition, in 1997, they reported that 7-deaza-isoG_d (**44a**, Fig. 10) can also form a tetraplex, identical in hydrogen-bonding structure to $d(T_4\text{isoG}_4T_4)_4$, but the former is less stable because of the heterocycle change.¹⁴⁴ This observation is also a confirmation that N7 does not participate in the isoG hydrogen-bonding system. Moreover, in 1997, they proved that isoG_d is sensitive to acid and can be easily hydrolyzed under acidic conditions.¹⁴⁴⁻¹⁴⁶ But 7-deaza and 8-aza-7-deaza purines can add to the stability of the glycosylic bond. Moreover, 7-deaza-isoG_d can form stronger tetraplexes in the presence of Na^+/K^+ comparing to isoG_d, respectively. The 7-halogen substitution is capable of enhancing stabilities of the base pairs (even in mismatches), glycosylic bond, and duplexes of 8-aza-7-deaza-isoG derivatives. However, 7-deaza-isoG_d show the topmost glycosylic bond stability but the lowest duplex stability.¹⁴⁵

5.3.4 Parallel-stranded (ps) and antiparallel stranded (aps) chains containing isoG. A high A_d-T_d content and the special sequences are prerequisites for ps DNA duplexes.¹⁴⁷ In general, ps DNA is more unstable than the aps counterparts, because of weaker stacking interactions and hydrogen bonds by A_d-T_d reversed Watson-Crick (Donohue) base pairing. G and 5-methyl-isoC base pair incorporated with A-T can lead to ps duplexes with lower stability than the corresponding aps duplexes (Fig. 19).¹⁴⁸ Ps DNA can be found in some oligonucleotide duplexes, triplexes and some tetrameric aggregates. In 1993, Seela¹⁴⁸⁻¹⁵⁰ *et al.* initially reported that the oligonucleotides containing isoG-C can form a new type of ps DNA duplexes (Fig. 19). And then, in 1999, they proposed that if a central A_d-T_d is replaced by a isoG_d-C_d , the stability of the original aps duplex will be reduced.¹⁵¹ Further, if isoG_d-C_d replaces two A_d-T_d pairs, it will form the ps chains. However, oligonucleotides containing $\text{isoG}_d\text{-isoC}_d$ still keep aps duplexes (Fig. 19). Notably, in 2013,

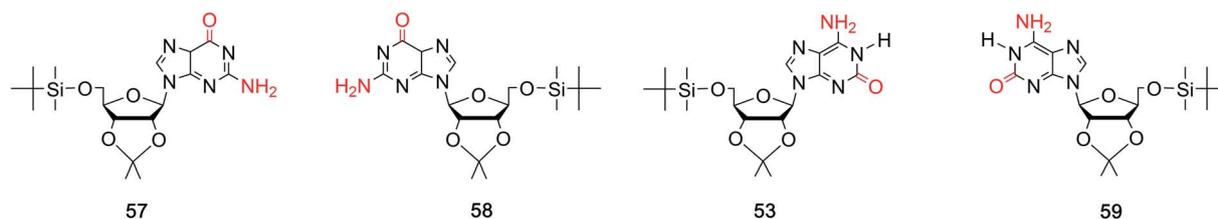


Fig. 17 The D and L structures of the protected G and isoG.



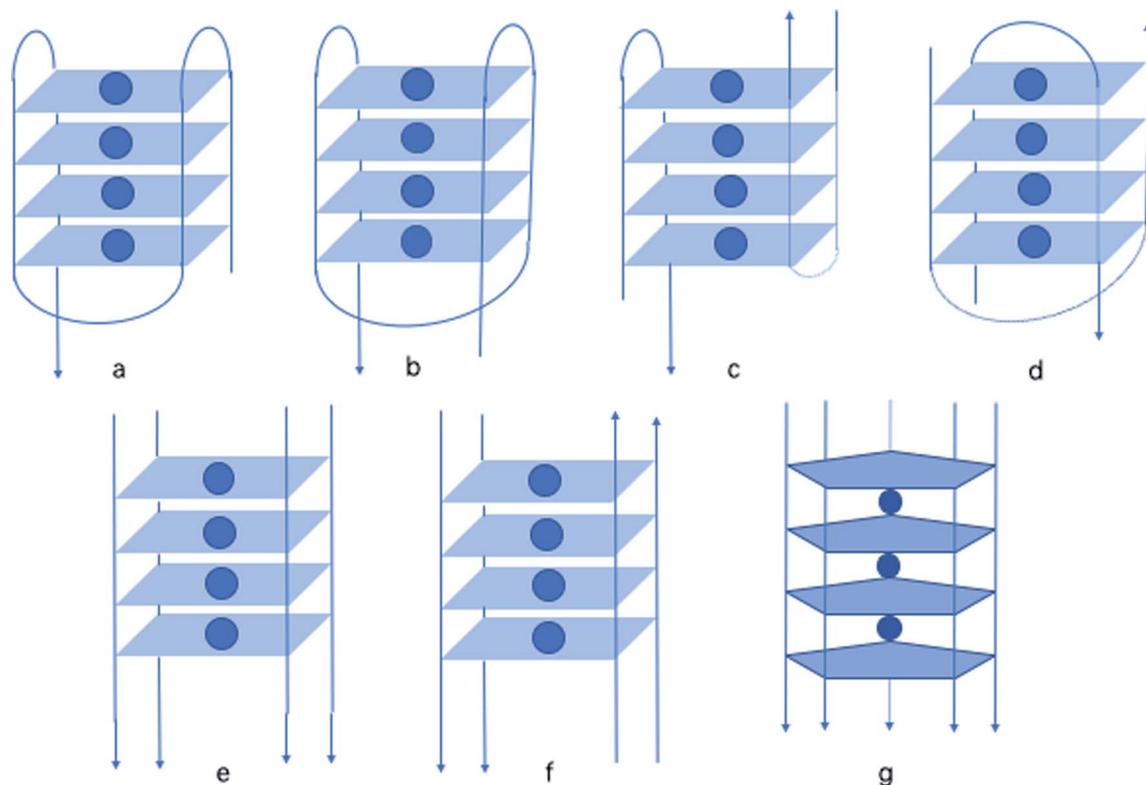


Fig. 18 A variety of oligonucleotide structures: (a) monomer chair, (b) monomer basket, (c) dimer chair, (d) dimer basket, (e) tetramer parallel, (f) tetramer antiparallel, (g) pentaplex.

5'- d(T isoC A T A A isoC T G T A T) -3'

5'- d(A G T A T T G A C A T A) -3'

5'- d(T isoG A T A A isoG T G T A T) -3'

5'- d(A C T A T T C A C A T A) -3'

5'- d(T isoC A T A A isoC T G T A T) -3'

3'- d(A isoG T A T T isoG A C A T A) -5'

Fig. 19 Ps duplexes with isoC–G/isoG–C and aps duplexes with isoC–isoG.

they reported that tridentate isoG_d–C_d base pairs can be used to compensate for the loss and stabilize ps DNA.¹⁵² In addition, in 2003, they proposed that 7-halogenated 8-aza-7-deaza-isoG_d (43b, 43c, Fig. 10) can significantly increase the duplex stability of 8-aza-7-deaza-isoG_d (43a) both in ps and aps DNA.⁶⁷ In 2015, they raised that ^FisoG_d (51b, Fig. 11) can add to the stability of aps duplexes, and this effect correlates to the number of incorporated ^FisoG_d units.⁷⁸ But this is not true for ps duplexes because the sugar conformation of ^FisoG_d does not apply to ps duplexes.

6 Applications

In the above parts, the discovery of isoG, its synthesis, modifications, tautomerism, and different mismatches, as well as the self-assembly properties of isoG, have been introduced. Based on the above characteristics, the applications of isoG have been investigated, such as the formation of supramolecular gels, the role of base-pairing in duplexes, its use as an ionophore, and its use as an antitumor agent. In the following sections, these applications will be discussed in detail.

6.1 isoG-based supramolecular hydrogels

Hydrogels have been an important topic for centuries because of their diverse applications in many fields, including in nanoscience, medicine, and the cosmetics industry. Hydrogels are viscoelastic solid-like materials composed of an elastic crosslinked network, and the major component solvent.¹⁵³ Hydrogels can be divided into two categories. One is polymer hydrogels, whose shape is maintained by strong chemical bonds crosslinked long chains, which cannot be broken easily and are not thermally reversible. The other one is low molecular weight hydrogels (LMWH). LMWH formed by the self-assembly of materials through noncovalent interactions, such as hydrogen-bonding, π – π stacking, hydrophobic forces, and van der Waals interactions. LMWH, also known as supramolecular hydrogels or physical hydrogels, will be discussed in this paper. Nucleosides are desirable candidates for building



supramolecular hydrogels because of their ability to self-assemble *via* noncovalent interactions, as well as their biocompatibility in cells.

The gelation of guanylic acid was firstly identified in 1910 by Bang.¹⁵⁴ G and some of its derivatives can self-assemble into various supramolecular gels based on its unique hydrogen bond donors and receptors.¹⁵⁴ There have been many reports concerning G-based hydrogels and their applications in diagnostics and nanoscience. But these G-based hydrogels all show short lifespans. So finding new nucleosides that can form long-life supramolecular hydrogels is crucial. Like G, isoG can form gels as well. In a synthesis of isoG in 1951, Davoll⁷ noted the formation of a “gelatinous material” during the deamination of 2,6-diaminopurine, but he did not characterize this material further by biochemistry. It was preliminarily described that the aggregation state of nucleosides turns viscous at higher concentrations and when cooled. Then in 1965, Ravindranathan⁴⁰ *et al.* synthesized isoG and gave proof for the phenomenon observed by Davoll.⁷ They reported the formation of asymmetry, ruled, ordered, spiral and temperature-depending structures where the nucleosides were uncharged. And these structures are similar to the gel formed by 5'-monophosphate-guanosine (5'-GMP), as identified by infrared spectroscopy. However, the preference of isoG for alkali ions is different from that of G. They also concluded that the purine ring may be hydrogen-bonded with the amino group of another molecule, and the carbonyl group does not participate in the formation of the ordered structure.

As shown in Table 4 and Fig. 20, remarkably, in 2017, Seela¹⁵⁵ *et al.* found that isoG-based hydrogels formed in various solutions containing Li⁺, Na⁺, K⁺, Rb⁺ and Cs⁺, but not in water. And all hydrogels present significant longer lifespans (two or three months) compared to those of G (several days), as well as higher thermodynamic stability and wider pH range. The rheological results suggested that the stabilities of isoG-based gels are strikingly higher than those of G. And scanning electronic microscope (SEM) images revealed that isoG-based hydrogels form helix-like cylinders, whereas G-based gels form individual flat ribbons. As reported, hydrogels formed by isoG at

physiological Na⁺ concentration showed a good small molecule loading and releasing capacity in phosphate buffered saline (PBS) buffer. This suggested their potential applications in drug delivery and as ionophores carrying different ions. In addition, supramolecular hydrogel systems of isoG_d (51a, Fig. 11) solutions containing Li⁺, K⁺ and Rb⁺ have been established. However, in Na⁺ and Cs⁺ solutions, these hydrogels can not take shape. Moreover, the derivative ^FisoG_d (51b, Fig. 11) can also form supramolecular hydrogels in the presence of Li⁺, Na⁺, K⁺ and Rb⁺. While in solutions containing Cs⁺, precipitates form, indicating the ion dependence of the three nucleosides-based gels is different.

Hydrogels with fluorescence have aroused much attention due to its latent applications in drug delivery, molecule marking, and tissue engineering. As discussed before, z⁸isoG_d, z⁸G_d are virtually fluorescent in alkaline environment and at neutral pH, similar to those reported for 8-aza-G and relevant 8-azapurine nucleosides.⁷⁰⁻⁷³ Notably, isoG_d and G_d are not fluorescent. In addition, z⁸isoG_d and z⁸G_d both show self-assembly properties. So experiments have been carried out by Seela¹⁵⁶ *et al.* in 2017 to prepare smart (stimulus-responsive) gels with fluorescence, which can be regulated by external changes, such as pH, heat and nucleoside mimics. Remarkably, z⁸isoG_d formed the first G-similar hydrogel with fluorescence properties even in solid state. And the hydrogel also shows a longer lifetime and higher thermal stability than isoG and G hydrogels. However, z⁸G_d does not form a gel under experimental conditions. z⁸isoG_d hydrogel has a great selectivity for K⁺, and its pentad and tetrad structures are the basis of its self-assembly into gels. SEM images revealed that z⁸isoG_d gel forms dense fibers and bundles, whereas z⁸G_d forms nanotubes. It makes great significance that the fluorescence hydrogel formed by z⁸isoG_d can take response to external changes such as pH, UV, and heat. Additionally, in terms of the ability to form gels even at a very low KCl concentrations, z⁸isoG_d hydrogel is expected to be applied in various fields of nanobiotechnology, chemistry, biology and medicine. Interestingly, although N7 does not participate in the isoG hydrogen-bonding system, the absence of N7 in 7-deaza-isoG_d results in this system being unable to form a hydrogel. Moreover, the addition of 1.0 equivalent of 7-deaza-isoG_d can break the hydrogels formed by isoG and the fluorescence hydrogels formed by z⁸isoG_d.

Self-healing is an ability to self-repair when the external stimuli, such as pH, external force and temperature, are moved. It is one of the most attractive properties of hydrogels and empowers them with diverse applications in many fields like tissue engineering. For example, so-called injectable hydrogels undergo shear-thinning on injection, but the hydrogels recover soon after injection.¹⁵⁷ There have been many reports on self-healing polymer hydrogels, but only a few studies concerning the self-healing supramolecular nucleoside hydrogels have been published. In 2018, Das¹⁵⁸ *et al.* developed a self-healing arylboronate esters mediated G-quartet hydrogel as promising 3D-bioink. However, the applications of G-based hydrogels are limited for their short lifetime within hours and low stabilities. This has drawn attention to isoG, an isomer of G. However, the injection of isoG hydrogels is challenging.¹⁵⁹ Recently, inspired

Table 4 The property of supramolecular gel forming by isoG, its derivatives and G-isoG co-gel

	H ₂ O	Li ⁺	Na ⁺	K ⁺	Rb ⁺	Cs ⁺
isoG	S ^c	Gel	Gel	Gel	Gel	Gel
G	— ^e	Cry ^b	Cry ^b	Gel	Cry ^b	Cry ^b
isoG _d	Gel	Gel	S ^c	Gel	Gel	S ^c
G _d	Cry ^b	Cry ^b	Cry ^b	Gel	VS ^d	Cry ^b
^F isoG _d	Gel	Gel	Gel	Gel	Gel	S ^c
z ⁸ isoG _d	Pre ^a	Pre ^a	Pre ^a	Gel	Pre ^a	Pre ^a
z ⁸ G _d	Pre ^a	S ^c	S ^c	S ^c	S ^c	S ^c
G : isoG (1 : 1)	— ^e	Pre ^a	Cry ^b	Gel	Gel	Cry ^b
G : isoG (1 : 3)	— ^e	Cry ^b	Cry ^b	Gel	Gel	Cry ^b
G : isoG (3 : 1)	— ^e	Cry ^b	Cry ^b	Gel	Cry ^b	Cry ^b

^a Precipitation. ^b Crystallization. ^c Solution. ^d Viscous solution. ^e Not experimented.



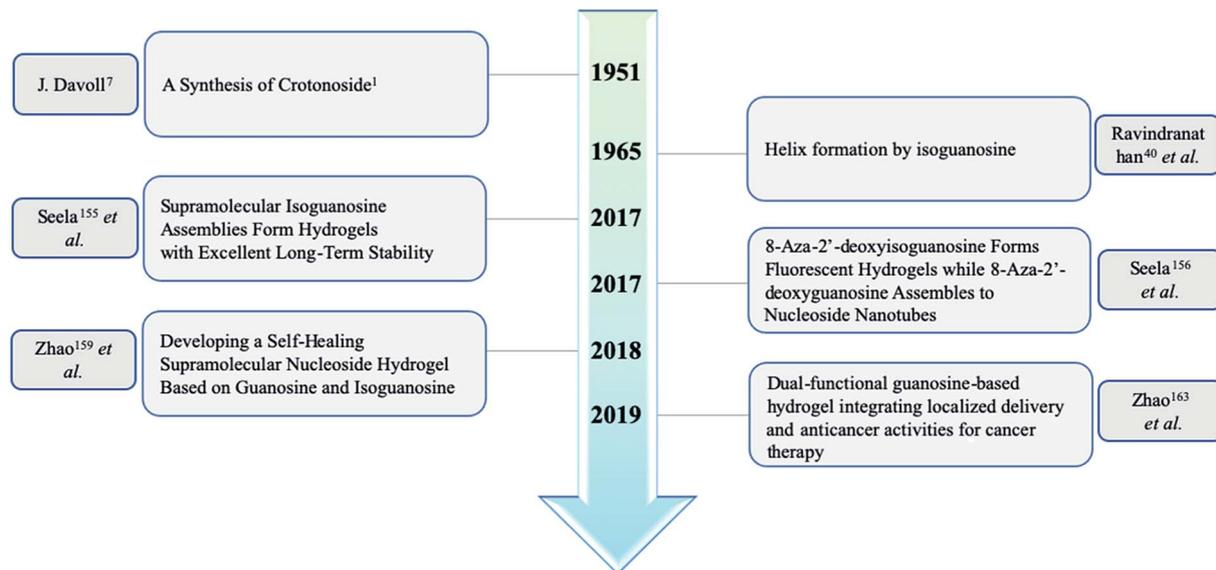


Fig. 20 The development of hydrogels formed by isoG.

by the self-assembled tetramers formed by mixing $d(T_4G_4T_4)$ and $d(T_4isoG_4T_4)$ verified by Seela¹⁴⁰ *et al.* in 1996, the tetraplexes formed by $d(T_8isoG_4T)$ and $d(T_4G_4T)$ in the presence of K^+ proved by Roberts¹⁴¹ *et al.* in 1997, and the self-healing co-gel developed by mixing G and G_d by Adhikari¹⁶⁰ *et al.* in 2014, Zhao¹⁵⁹ *et al.* successfully constructed a self-healing supramolecular co-hydrogel in 2018 by simply mixing G and isoG with 1 : 1 and 1 : 3 molar ratios in the presence of K^+ . In terms of ion dependence, both mixtures can form co-gels in the presence of K^+ and Rb^+ . But the 3 : 1 could only produce a co-gel in the presence of K^+ . Furthermore, the lifespan of the gels varied from hours to months. SEM images show that these co-gels form flower-like structures. Rheological and injection experiments concluded the excellent self-healing properties, short recovery time, and syringeability of the co-gel. Moreover, G_2isoG_2 -quartets are possibly formed inside the self-healing supramolecular nucleoside co-gel.

Furthermore, it is widely known that isoG possesses anti-tumor property toward several cancer lines (details are shown in Section 6.5);^{58,161} and the injectable property could greatly promote the application of these supramolecular gels in anti-tumor treatment. In addition, borate ester is well-known as a dynamic covalent bond to construct self-healing hydrogels. Based on these, in 2017, Sadler¹⁶² *et al.* developed an anticancer hydrogel (dopamine-conjugated platinum IV- G_4K^+ -borate ester, Pt- G_4K^+B hydrogel) by using borate ester linkage. Inspired by above-mentioned findings, recently, Zhao¹⁶³ *et al.* developed another type of dual-functional co-gel called isoguanosine-borate-guanosine (isoGBG) hydrogel (Fig. 21), which possesses delivery and antitumor activities. isoGBG hydrogel displays excellent stability, self-healing property and biocompatibility, as well as highly antitumor activity and excellent inhibition of tumor recurrence *in vivo*. Moreover, the mechanism was preliminarily explored, and it was concluded that the mechanism of inhibiting oral cancer cells might be through the

caspase-dependent signaling pathway to induce apoptosis *via* regulating dephosphorylation of epidermal growth factor receptor (EGFR) (Fig. 21).

6.2 Applications of isoG mispairing

In Section 5.2, different mismatches and the incorporation of isoguanine have been discussed. Although isoG can be integrated into DNA/RNA by multiple enzymes in various templates, isoG-isoC is not accepted as a natural base pair. The three reasons why isoG-isoC cannot be a natural base pair are: (1) tautomerism causes unstable base pairing;^{96,101} (2) deoxygenated isoC is easily degraded to deoxygenated U;¹⁰¹ (3) isoG-isoC cannot form an R-type triple-chain structure.¹⁶⁴ Although the tautomerism of isoG limits its stable transmission as a gene letter, it enables the existence of gene mismatch, suggesting isoG's various alternative applications. In 1996, for the first time, Kamiya¹⁰³ *et al.* suggested that isoG may be mutagenic and lead to A to C, A to G and A to T transversions in cells. Then in 1997, isoG is proved to induce mutagenicity in *E. coli*,³² as well as in living COS-7 cells,¹⁶⁵ but it does not hinder the replication. These indicate the mutation caused by isoG can occur in both eukaryotic and prokaryotic cells. Interestingly, the mutagenicity is affected by the sequence and strands that the base is located in. 8-OH-dGTP is a significant marker of DNA oxidation. In 1998, Kamiya¹⁶⁶ *et al.* demonstrated that the mutagenicity induced by 2-OH-dATP in *E. coli* was more significant than that caused by 8-OH-dGTP, which also gave a confirmation of 2-OH-dATP-G base pair.

Interestingly, the nonstandard isoG-isoC base pair contributes to the stability of duplexes. In 2001, Turner⁹⁷ *et al.* reported this effect has a sequence dependency in duplexes because of the large difference in electron density. In 2004, Prudent^{167,168} *et al.* proved that isoG-isoC stabilizes the G-C double helix and can be applied to polymerase chain reaction (PCR) to detect the



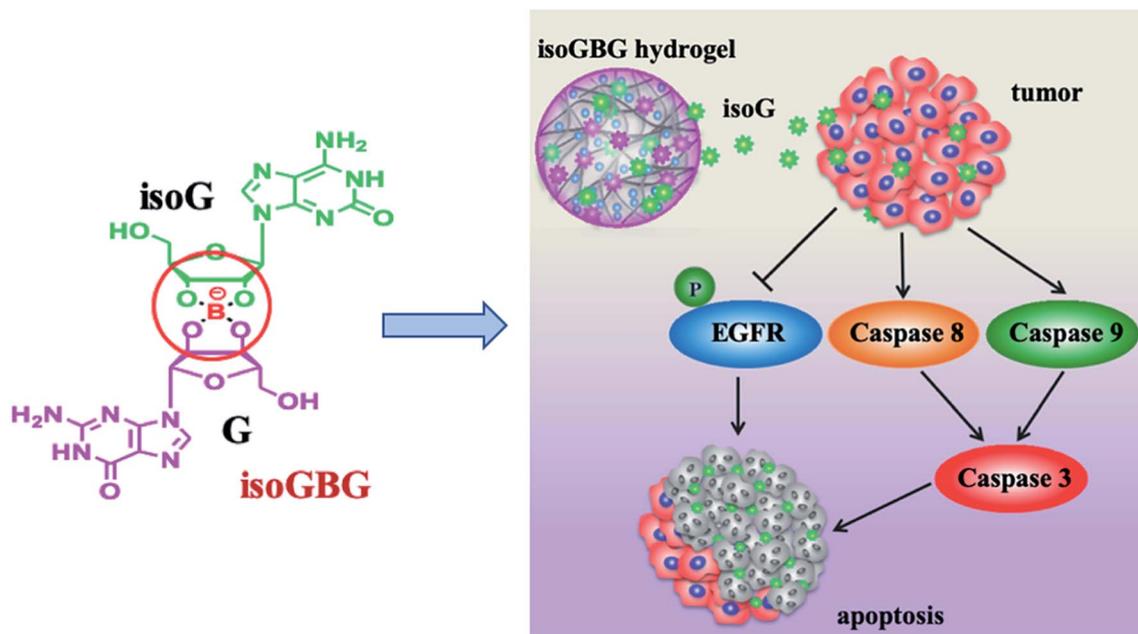


Fig. 21 The dual-functional isoGBG displays antitumor activity, may via the caspase-dependent signaling pathway to induce apoptosis via regulating dephosphorylation of EGFR.

mutant genes. Beyond that, in 1992, Benner¹⁶⁹ *et al.* raised that it can help regulate ribosome translation. 91% of isoG-containing RNA could be read, which is higher than the control, because the frameshift with isoG skips the isoC. However, the transfer RNA (tRNA) containing the meaningless CUA codon is meaningless and results in translation termination. This property can also aid the development of nonstandard base pairs to extend gene dictionaries and provide a deeper understanding of translation termination. Moreover, in 1993, Switzer¹⁰¹ *et al.* reported that the tautomerism of isoG leads to various mispairings, which may enhance the antiviral activity. The stability of base-pairs of z^8 isoG_d (46, Fig. 10) with 5-methyl-isoC, A_d, T_d, C_d and G_d can be monitored by fluorescence,⁷³ in other words, the fluorescence signal is correlated with DNA base pairing stability. Thus, z^8 isoG_d can be applied as a sensor with fluorescence for detecting mismatch DNA duplexes, even more complicated DNA structures,⁷³ such as in triplexes reported by Seela¹⁷⁰ *et al.* in 2010.

6.3 Applications of 1-methyl-isoG

In 1981, Fuhrman¹⁷¹ *et al.* reported that isoG presents negative inotropic and chronotropic effects in the myocardium. And those effects can be antagonized by theophylline. The effects of negative inotropic and chronotropic activity ranked in the order isoG > 1-methyl-isoG > adenosine > 2-methoxyadenosine. And these substances may act on the same receptor. In addition, as reported by Emerson¹⁷² *et al.* in 1949 and Brown¹⁷³ *et al.* in 1952, isoG can produce smooth muscle relaxation, hypotension and bradycardia effects that are more potent and longer-lasting than those of adenosine. Remarkably, 1-methyl-isoG (37, Fig. 10) was also found to have muscle relaxation, hypothermia,

hypotension, anti-inflammatory, activating adenosine cyclase (AC) and anti-allergic effects (Fig. 22).^{30,50,51,56,174–177} In 1981, as shown in Table 5, a comparison of effects of the derivatives of 1-methyl-isoG have been explored by Cook⁵⁶ *et al.* Moreover, the effects of halogenated analog 2-chloroadenosine are greater.³⁰

These effects induced by 1-methyl-isoG are more potent than adenosine because 1-methyl-isoG can resist the action of adenosine deaminase. So 1-methyl-isoG can exist in the extracellular fluid for a long time, whereas adenosine has been degraded into inactive inosine.⁵¹ And there is a positive correlation between the potency of resistance to adenosine deaminase and the muscle relaxation and cardiovascular effects.^{30,51,178} However, in 1985, Radulovacki¹⁷⁹ *et al.* reported 1-methyl-isoG does not have a strong hypnosis effect similar to

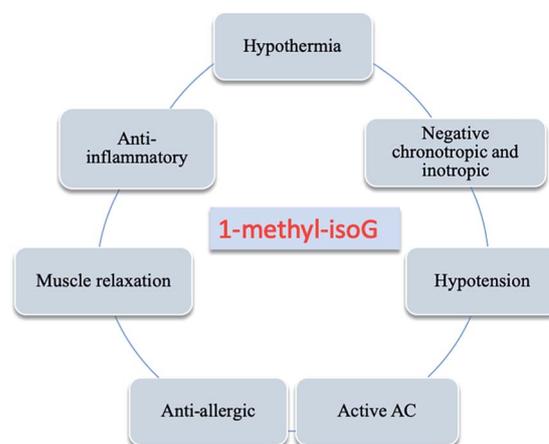


Fig. 22 Effects of 1-methyl-isoG.



Table 5 Comparisons of the effectiveness of 1-methyl-isoG and its derivatives in muscle relaxation, hypothermia, hypotension, anti-inflammatory and antiallergy effects

Number	Muscle relaxation	Hypothermia	Hypotension	Anti-inflammatory	Anti-allergic
37	Y ^a	Y ^a	Y ^a	Y ^a	Y ^a
38a	Y ^a	Y ^a	Y ^a	Weak ^b	Y ^a
39g	N ^c	N ^c	Y ^a	— ^d	Y ^a
39h	N ^c	N ^c	Weak ^b	— ^d	Y ^a
39i	N ^c	N ^c	— ^d	— ^d	Weak ^b
39j	N ^c	N ^c	— ^d	— ^d	Y ^a
40a	N ^c	N ^c	— ^d	— ^d	— ^d
40b	N ^c	N ^c	Weak ^b	— ^d	— ^d
40c	N ^c	N ^c	Weak ^b	— ^d	— ^d
54a	N ^c	N ^c	— ^d	— ^d	— ^d
54b	N ^c	N ^c	— ^d	— ^d	— ^d
54c	Y ^a	Y ^a	Y ^a	Y ^a	Y ^a
55	N ^c	N ^c	— ^d	Weak ^b	Y ^a
56	Y ^a	Y ^a	Y ^a	— ^d	Weak ^b

^a Effective. ^b Limited effect. ^c Invalid. ^d Not experimented.

adenosine. And, in 1981, Spence¹⁸⁰ *et al.* showed that the effects of 1-methyl-isoG and adenosine on neuromuscular transmission in the vas deferens are similar. This is probably because the enzyme activity of adenosine deaminase in the vas deferens is inherently low, and thus the degradation is low. The ability of 1-methyl-isoG to cause the accumulation of cyclic adenosine monophosphate (cAMP) in brain is not effective as that of adenosine,¹⁸¹ but neither of these two nucleosides show inhibition of rat brain cAMP phosphodiesterase (PDE). Besides, in 1980, Taylor³⁰ *et al.* proved that accumulated adenylate cyclase (AC) caused by 1-methyl-isoG may be associated with cardiovascular/relaxation effects.

In 1981, Marwood¹⁷⁷ *et al.* showed that 1-methyl-isoG has equal negative inotropic and chronotropic effects in guinea-pig isolated atria. Nevertheless, adenosine has a greater negative inotropic effect. The active site of 1-methyl-isoG is only on the atria, whereas adenosine can also act on ventricle.³¹ Furthermore, 1-methyl-isoG showed a biphasic response on cardiac output, which means a slow increase followed by a rapid initial decrease.¹⁷⁷ The impact of 1-methyl-isoG in lowering response electrical stimulation in isolated perfused rat tail arteries is similar to that of adenosine. In 1980, Jamieson¹⁷⁸ *et al.* demonstrated that 1-methyl-isoG shows an inhibition of nerve-mediated stimulation, which is induced by electricity or nicotine in isolated guinea-pig ileum. But it is not useful for acetylcholine (ACh) and histamine-mediated contraction. Moreover, the site of action is at the post ganglionic nerve terminals because the effect disappears in isolated mammalian motor axons. Further, in 1981, Spence¹⁷⁶ *et al.* demonstrated that the inhibitory dose for central excitation and monosynaptic or polysynaptic spinal reflex in conscious animals (central pathways) is much lower than that of neuromuscular junction. This means that muscle relaxation may be related to the central pathway (probably the spinal pathway).

In 1980, Taylor¹⁸² *et al.* reported 1-methyl-isoG does not compete with adenosine for uptake sites to activate adenosine

cyclase. This indicates that it does not attenuate the effect of adenosine by competition, but because 1-methyl-isoG is not degraded and remains for a long time in the extracellular fluid. 1-Methyl-isoG was thought to act on a benzodiazepine receptor, but that the hypnotic effect was not the same as benzodiazepine ruled that out by Hall¹⁸³ *et al.* in 1987. Those effects above are dose-dependent.^{30,51,176} And theophylline, a widely-used blocker of adenosine receptors, also acts as a dose-dependent antagonist of the effects of 1-methyl-isoG.^{51,178,179} So 1-methyl-isoG may act on the same receptor as adenosine in the intestinal smooth muscle and pig brain.

6.4 Ionophores

Nuclear waste remnants from the Second World War, nuclear testing, and the nuclear industry have left a large amount of radioactive contamination, mostly Cs⁺ and Ra²⁺. They are challenging to remove, leaving behind huge potential risk for human being and pollution of water and soil. Compounds that are highly selective for Cs⁺ are required because of the large quantity of Na⁺ and K⁺ in nuclear waste. In 1990s, crown ethers have been developed to bind Cs⁺ by McDowell¹⁸⁴ *et al.* and Sachleben^{185–188} *et al.* But the Cs⁺ selectivity are modest resulting from their flexibility. So rigid macrocycles, especially calixarene crowns and their derivatives with different binding groups attached, have been used as recycling ligands. Rigid macrocycles show better complexation and selectivity for Cs⁺ over other ions such as Li⁺, Na⁺, K⁺ and Rb⁺.¹⁸⁹ In addition, in 1995, calixarene carriers were reported to have rattling cell membrane penetration in polymer inclusion membrane (PIMs) by Reinhoudt¹⁹⁰ *et al.* However, these macrocycles are expensive to synthesize and the ionophore recovery, as well as cations release, are difficult. Nowadays, noncovalent ionophores containing isoG are widely used. They show greater affinity and selectivity for Cs⁺ compared to those of calixarene and its derivatives. In 1997, Davis¹⁸⁹ *et al.* reported the self-assembled (isoG)₁₀·M⁺ ionophore shows significant selectivity for Cs⁺ in



competition experiments. And $(\text{isoG})_{10} \cdot \text{M}^+$ can even extract bound Cs^+ from a calixarene crowns ether. Thus, these compounds can be used for purification and metal nuclear waste disposal.

Further, in 2000, Davis⁸⁰ *et al.* then reported that in the $(\text{isoG})_{10} \cdot \text{Cs}^+$ system (Fig. 15 and 16), Cs^+ and the protected isoG 53 are in constant exchange with the outside world. However, the exchange rate of Cs^+ itself is 40 000 times faster than the compound 53, as shown by multinuclear NMR spectroscopy. So the ion carrier does not disintegrate during ion exchange. The ions can be easily dissociated from the noncovalent ionophore when the solvent polarity changes. The same exchange process is true for other metal ions. Specially, Cs^+ possesses an advantage that it can even slow down the isoG exchange rate. In 2001, they proved that the protected isoG 53 assembly can not only release Cs^+ more easily without dissociation, but also show a stronger affinity for larger cations such as Cs^+ and Ba^{2+} than K^+ , Sr^{2+} and Na^+ .¹⁹¹ They also demonstrated that $(53)_{10} \cdot \text{Cs}^+$ can also promote selective transport through organic polymer inclusion membrane (PIMs) and bulk liquid membranes (BIMs), with high transport rates and excellent selectivity for Cs^+ . This cation-binding selectivity is attributed to the geometric hydrogen-bonding mode composed of donors and receptors. As discussed earlier, G forms G-quartets, whereas isoG prefers the formation of pentads. The sizes of these macrocyclic cavities of self-assembled ionophores determine the cation-binding selectivity. Moreover, in 2004, they reported that in the case of G, the selectivity for Ra^{2+} prior to magnesium ion (Mg^{2+}) and calcium ion (Ca^{2+}) requires an existing lipophilic anion.¹⁹² Nevertheless, isoG shows preferential selectivity for Cs^+ over Mg^{2+} , Ca^{2+} , Sr^{2+} and Ba^{2+} without any additional environmental conditions. In addition, a significant radium ion ($^{226}\text{Ra}^{2+}$) preference over other monovalent alkali cations was observed for both 57 and 53, with a “precipitation prevention/dissolving” property over a wide pH range. And, in 2005, Reinhoudt¹⁹³ *et al.* reported that Ra^{2+} can be selectively extracted from gas-field produced water by isoG-based self-assemblies.

6.5 Antitumor activity

In 1994, Kim¹⁶¹ *et al.* found that isoG has strong anti-cancer effects against P338, L5178Y, Sp2/O, HL60, Lymphoma Raji, both *in vitro* and *in vivo*. And isoG shows especial effectiveness against ascites tumor and malignant solid tumor. It is generally believed that isoG enters the cell by simple diffusion or diffusion combined with active transporters. The phosphorylation of isoG in the cytoplasm produces active nucleotides that inhibit tumor growth. Moreover, in 1994, they also reported that most of 6-substituted isoG derivatives (42, Fig. 10) show low cytotoxicity against cancerous Pograss 1, HeLa and Molt-4 cells, except those with amino, sulfur and furfurylamine substituents.⁵⁸ This suggests that high cytotoxicity depends on the N2H or SH perssard at C6 position on the heterocyclic ring. In particular, when these substituents are displaced by nonpolar alkyl groups, the cytotoxicity decreases. Then in 1995, they demonstrated that the combination of berberine with isoG monophosphate greatly increases the antitumor effect.¹⁹⁴

However, in 2004, Nowak¹⁹⁵ *et al.* reported that similar to other telomerase inhibitors, isoG_d showed low potency and specificity in inhibiting tumor cell growth.

6.6 More applications

Apart from applications in gel formation, ionophore, and as antitumor agent, in 1973, isoG was reported to be an inhibitor of inosine monophosphate (IMP) pyrophosphorylase by Hagen.¹⁹⁶ And in 1974, Holy¹⁹⁷ *et al.* reported that isoG can also inhibit inducible *E. coli* binding site. In addition, in 1975, Mantsch⁴¹ *et al.* reported that its 5'-di- and -tri-phosphates can strongly bind and inhibit glutamic acid dehydrogenase. In 1988, Wissler¹⁹⁸ *et al.* reported that isoG also acts as an essential constituent of Cu(II)-containing ribonucleic acid produced by cultured pig macrophages. This unusual Cu(II)-containing extracellular RNA was a formidable angiogenesis factor, and acted as an adenosine A1-receptor agonist. Besides, in 1999, Davis¹³¹ *et al.* raised that the protected isoG (53, Fig. 11) self-assembly can act as phase transfer catalysts. And in 2018, Anna¹⁷⁴ *et al.* proposed that by strongly affecting the highly ordered structure of the G-quartet in TBA, isoG greatly affects the anticoagulant activity of the thrombin binding aptamer (TBA).

7 Summary and outlook

In summary, since Fischer first synthesized isoguanine in 1897, there have been many reports focusing on the comparison of isoG with G. In this review, we have detailed the discovery, and synthesis of base- or sugar-modified derivatives of isoG, its monomer structure and aggregates, and the diverse applications of isoG/isoguanine. Since the yields of isoG/isoguanine in natural sources such as croton beans, butterfly wings and marine mollusk are too low, a series of synthetic methods of isoG/isoguanine by introducing functional groups to the heterocycle of G and AICA have been proposed. And various base- or sugar-modified derivatives have been synthesized or found in nature. Additionally, some unique properties of isoG and its derivatives including enol-keto tautomerism, mispairing, self assembly into tetramer ($(\text{isoG})_4$), decamer ($(\text{isoG})_{10}$) and isoG_4 quadruplex, and fluorescence suggest its potential diverse applications in the formation of supramolecular hydrogels, in ps and aps duplexes and even triplexes, also in waste-collection as an ionophore. In addition, further applications such as antitumor activity, inhibition of IMP-pyrophosphorylase, and mutation-inducing have been talked about. As the structure and properties of isoG become better understood, there will be further studies and more applications will be identified. For example, because of its unique hydrogen bond acceptors and donors, isoG is expected to have a wider range of applications in the field of supramolecular self-assembly, the formation of hydrogel and ionophore. Additionally, isoG is most likely to play a significant role in genetic disease monitoring probes because of the variable nonstandard pairs caused by its tautomerism and its incorporation by different enzymes. Moreover, benefited from its antitumor ability and self-assembly properties, the



preparation of supramolecular gels integrating antitumor activity and drug delivery appears extremely significant. Thus, isoG may play a significant role in the clinical treatment of various cancers and other therapies involving smooth muscle relaxation, hypotension, and bradycardia. In conclusion, the unique properties of isoG, such as its use in supramolecular assemblies, as a nonstandard base and a medicine with anti-tumor activity, will broaden its applications in diverse fields.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This study is supported by the National Natural Science Foundations of China (81922020, 81970950, 81621062).

References

- M. Chen, W. Lin, L. Hong, N. Ji and H. Zhao, *BioMed Res. Int.*, 2019, **2019**, DOI: 10.1155/2019/6258248.
- J. T. Davis, *Angew. Chem., Int. Ed.*, 2004, **43**, 668–698.
- E. Fischer, *Ber. Dtsch. Chem. Ges.*, 1897, **30**, 2226–2254.
- E. Cherbuliez and K. Bernhard, *Helv. Chim. Acta.*, 1932, **15**, 464–471.
- R. Purrmann, *Justus Liebigs Ann. Chem.*, 1940, **544**, 182–190.
- F. A. Fuhrman, G. J. Fuhrman, R. J. Nachman and H. S. Mosher, *Science*, 1981, **212**, 557–558.
- J. Davoll, *J. Am. Chem. Soc.*, 1951, **73**, 3174–3176.
- G. B. Brown, G. Levin and S. Murphy, *Biochemistry*, 1964, **3**, 880–883.
- C. B. Reese, Y. S. Sanghvi and R. Kuroda, *J. Chem. Soc., Perkin Trans. 1*, 1987, 1527–1531.
- K. J. Divakar, M. Mottahedeh, C. B. Reese, Y. S. Sanghvi and K. A. D. Swift, *J. Chem. Soc., Perkin Trans. 1*, 1991, **4**, 771–774.
- C. Ji-Wang, L. Horng-Yuh, H. Min and S. Fang-Jy, *Tetrahedron Lett.*, 1987, **28**, 2151–2154.
- S. Tirumala and J. T. Davis, *J. Am. Chem. Soc.*, 1997, **119**, 2769–2776.
- X. Shi, J. C. Fettinger, M. Cai and J. T. Davis, *Angew. Chem., Int. Ed.*, 2000, **39**, 3124–3127.
- M. V. Buell and M. E. Perkins, *J. Biol. Chem.*, 1927, **72**, 745–749.
- J. R. Spies and N. L. Drake, *J. Am. Chem. Soc.*, 1935, **57**, 774.
- J. R. Spies, *J. Am. Chem. Soc.*, 1939, **61**, 350–351.
- R. Falconer, J. M. Gulland and L. F. Story, *J. Am. Chem. Soc.*, 1939, 1784, DOI: 10.1039/jr9390001784.
- G. R. Pettit, R. H. Ode, R. M. Coomes and S. L. Ode, *Lloydia*, 1976, **39**, 363–367.
- J. Davoll and B. A. Lowy, *J. Am. Chem. Soc.*, 1951, **73**, 1650–1655.
- A. Bendich, G. B. Brown, F. S. Philips and J. B. Thiersch, *J. Biol. Chem.*, 1950, **183**, 267–277.
- B. A. Lowy, J. Davoll and G. B. Brown, *J. Biol. Chem.*, 1952, **197**, 591–600.
- S. Friedman and J. S. Gots, *Arch. Biochem. Biophys.*, 1951, **32**, 227–229.
- Z. Nackerdien, K. S. Kasprzak, G. Rao, B. Halliwell and M. Dizdaroglu, *Cancer Res.*, 1991, 5837–5842.
- M. Dizdaroglu, *Mutat. Res.*, 1992, **275**, 331–342.
- R. Olinski, T. Zastawny, J. Budzbon, J. Skokowski, W. Zegarski and M. Dizdaroglu, *FEBS Lett.*, 1992, **309**, 193–198.
- P. Jaruga, T. H. Zastawny, J. Skokowski, M. Dizdaroglu and R. Olinski, *FEBS Lett.*, 1994, **341**, 59–64.
- T. Mori, Y. Hori and M. Dizdaroglu, *Int. J. Radiat. Biol.*, 1993, **64**, 645–650.
- P. Jaruga and M. Dizdaroglu, *Nucleic Acids Res.*, 1996, **24**, 1389–1394.
- H. Kamiya and H. Kasai, *J. Biol. Chem.*, 1995, **270**, 19446–19450.
- J. Baird-Lambert, J. F. Marwood, L. P. Davies and K. M. Taylor, *Life Sci.*, 1980, **26**, 1069–1077.
- H. Kamiya, T. Ueda, T. Ohgi, A. Matsukage and H. Kasai, *Nucleic Acids Res.*, 1995, **23**, 761–766.
- H. Kamiya and H. Kasai, *Nucleic Acids Res.*, 1997, **25**, 304–311.
- H. Kamiya, *Mutat. Res., Genet. Toxicol. Environ. Mutagen.*, 2010, **703**, 32–36.
- H. Kasai, *Free Radicals Biol. Med.*, 2002, **33**, 450–456.
- S. Frelon, T. Douki and J. Cadet, *Free Radical Res.*, 2002, **36**, 499–508.
- J. Cadet, T. Douki and J. L. Ravanat, *Environ. Health Perspect.*, 1997, **105**, 1034–1039.
- J. Cadet, T. Douki, J.-L. Ravanat and J. Wagner, *Bioanal. Rev.*, 2012, **4**, 55–74.
- A. Weimann, G. McLeod, T. Henriksen, V. Cejvanovic and H. E. Poulsen, *Scand. J. Clin. Lab. Invest.*, 2019, **79**, 1–8.
- V. Nair and D. A. Young, *J. Org. Chem.*, 1985, **50**, 406–408.
- R. V. Ravindranathan and H. T. Miles, *Biochim. Biophys. Acta*, 1965, **94**, 603–606.
- H. H. Mantsch, I. Goia, M. Kezdi, O. Barzu, M. Dansoreanu, G. Jebeleanu and N. G. Ty, *Biochemistry*, 1975, **14**, 5593–5601.
- J. A. Montgomery and K. Hewson, *J. Org. Chem.*, 1968, **33**, 432.
- L. De Napoli, D. Montesarchio, G. Piccialli, C. Santacrose and M. Varra, *J. Chem. Soc., Perkin Trans. 1*, 1995, **15**, 45–47.
- Q. Cheng, J. Gu, K. R. Compaan and H. F. Schaefer, *Chem.–Eur. J.*, 2012, **18**, 4877–4886.
- A. Yamazaki, I. Kumashiro, T. Takenishi and M. Ikehara, *Chem. Pharm. Bull.*, 1968, **16**, 2172–2181.
- A. Yamazaki, M. Okutsu and Y. Yamada, *Nucleic Acids Res.*, 1976, **3**, 251–260.
- J. W. Chern, G. S. Lin, C. S. Chen and L. B. Townsend, *J. Org. Chem.*, 1991, **56**, 4213–4218.
- L. De Napoli, A. Messere, D. Montesarchio, G. Piccialli and M. Varra, *Nucleosides Nucleotides*, 1997, **16**, 183–191.
- F. A. Fuhrman, G. J. Fuhrman and K. Deriemer, *Biol. Bull.*, 1979, **156**, 289–299.
- R. J. Quinn, R. P. Gregson, A. F. Cook and R. T. Bartlett, *Tetrahedron Lett.*, 1980, **21**, 567–568.



- 51 Y. H. Kim, R. J. Nachman, L. Pavelka, H. S. Mosher, F. A. Fuhrman and G. J. Fuhrman, *J. Nat. Prod.*, 1981, **44**, 206–214.
- 52 W. Bergmann and R. J. Feeney, *J. Org. Chem.*, 1951, **16**, 981–987.
- 53 W. Bergmann and D. C. Burke, *J. Org. Chem.*, 1955, **20**, 1501–1506.
- 54 W. Bergmann and D. C. Burke, *J. Org. Chem.*, 1956, **21**, 226–228.
- 55 P. Searle and T. Molinski, *J. Nat. Prod.*, 1994, **57**, 1452–1454.
- 56 R. T. Bartlett, A. F. Cook, M. J. Holman, W. W. McComas, E. F. Nowoswait, M. S. Poonian, J. A. Baird-Lambert, B. A. Baldo and J. F. Marwood, *J. Med. Chem.*, 1981, **24**, 947–954.
- 57 D. Firsova, K. Calabro, P. Lasserre, F. Reyes and O. P. Thomas, *Tetrahedron Lett.*, 2017, **58**, 4652–4654.
- 58 S. Lee, J. Kim, Y. Cho and J. Kim, *Arch. Pharmacol Res.*, 1994, **17**, 170–174.
- 59 C. Buhr, R. Wagner, D. Grant and B. Froehler, *Nucleic Acids Res.*, 1996, **24**, 2974–2980.
- 60 G. Balow, V. Mohan, E. A. Lesnik, J. F. Johnston, B. P. Monia and O. L. Acevedo, *Nucleic Acids Res.*, 1998, **26**, 3350–3357.
- 61 F. Seela and H. Thomas, *Helv. Chim. Acta*, 1995, **78**, 94–108.
- 62 N. Ramzaeva and F. Seela, *Helv. Chim. Acta*, 1996, **79**, 1549–1558.
- 63 F. Seela and Y. Chen, *Chem. Commun.*, 1996, 2263–2264, DOI: 10.1039/CC9960002263.
- 64 G. Becher, J. He and F. Seela, *Helv. Chim. Acta*, 2001, **84**, 1048–1065.
- 65 F. Seela and G. Becher, *Helv. Chim. Acta*, 1999, **82**, 1640–1655.
- 66 F. Seela and G. Becher, *Chem. Commun.*, 1998, 2017–2018.
- 67 F. Seela and R. Kröschel, *Nucleic Acids Res.*, 2003, **31**, 7150–7158.
- 68 F. Seela and K. Xu, *Org. Biomol. Chem.*, 2007, **5**, 3034–3045.
- 69 M. Jeselnik, S. Jakska and J. Kobe, *ChemInform*, 2004, **35**, 153–160.
- 70 J. Wierzchowski, J. Sepioł, D. Sulikowski, B. Kierdaszuk and D. Shugar, *J. Photochem. Photobiol., A*, 2006, **179**, 276–282.
- 71 F. Seela, A. M. Javelakar and I. Munster, *Helv. Chim. Acta*, 2005, **88**, 751–765.
- 72 F. Seela, I. Munster, U. Loechner and H. Rosemeyer, *Helv. Chim. Acta*, 1998, **81**, 1139–1155.
- 73 D. Jiang and F. Seela, *J. Am. Chem. Soc.*, 2010, **132**, 4016–4024.
- 74 H. U. Blank, I. Wempfen and J. J. Fox, *J. Org. Chem.*, 1970, **35**, 1131–1138.
- 75 J. Wierzchowski, G. Mędza, J. Sepioł, M. Szabelski and D. Shugar, *J. Photochem. Photobiol., A*, 2012, **237**, 64–70.
- 76 Z. Kazimierzczuk and D. Shugar, *Acta Biochim. Pol.*, 1973, **20**, 395–402.
- 77 Z. Kazimierzczuk, R. Mertens, W. Kawczynski and F. Seela, *Helv. Chim. Acta*, 1991, **74**, 1742–1748.
- 78 S. A. Ingale, P. Leonard, Q. N. Tran and F. Seela, *J. Org. Chem.*, 2015, **80**, 3124–3138.
- 79 K. Ono, M. Ogasawara, Y. Iwata, H. Nakane, T. Fujii, K. Sawai and M. Saneyoshi, *Biochem. Biophys. Res. Commun.*, 1986, **140**, 498–507.
- 80 M. Cai, V. Sidorov, Y. F. Lam, R. A. Flowers and J. T. Davis, *Org. Lett.*, 2000, **2**, 1665–1668.
- 81 S. Kim, S. Lee, W. Sun, S. Oh and J. Kim, *Arch. Pharmacol Res.*, 1999, **22**, 619–623.
- 82 T. Davis, S. Tirumala, J. R. Jenssen, E. Radler and D. Fabris, *J. Org. Chem.*, 1995, **60**, 4167–4176.
- 83 W. Pfeleiderer, *Justus Liebigs Ann. Chem.*, 1961, **647**, 167–173.
- 84 C. Roberts, R. Bandaru and C. Switzer, *J. Am. Chem. Soc.*, 1997, **119**, 4640–4649.
- 85 T. Golas, M. Fikus, Z. Kazimierzczuk and D. Shugar, *Eur. J. Biochem.*, 1976, **65**, 183–192.
- 86 J. Sepiol, Z. Kazimierzczuk and D. Shugar, *Z. Naturforsch., C: Biosci.*, 1976, **31**, 361–370.
- 87 R. Krishnamurthy, S. Pitsch, M. Minton, C. Miculka, N. Windhab and A. Eschenmoser, *Angew. Chem., Int. Ed.*, 1996, **35**, 1537–1541.
- 88 J. Hunziker, H. J. Roth, M. Böhringer, A. Giger, U. Diederichsen, M. Göbel, R. Krishnan, B. Jaun, C. Leumann and A. Eschenmoser, *Helv. Chim. Acta*, 1993, **76**, 259–352.
- 89 A. Eschenmoser, *Pure Appl. Chem.*, 1993, **65**, 1179–1188.
- 90 A. Banerjee, W. Saenger, B. Lesyng, Z. Kazimierzczuk and D. Shugar, *Acta Crystallogr., Sect. B: Struct. Crystallogr. Cryst. Chem.*, 1978, **34**, 2472–2477.
- 91 F. Seela, X. Peng and K. Xu, *Nucleosides, Nucleotides Nucleic Acids*, 2007, **26**, 1569–1572.
- 92 *Horiz. Biochem*, ed. A. Rich, M. Kasha and B. Pullman, 1962, pp. 103–126.
- 93 A. P. Joseph, A. B. Steven, K. Tilman and E. M. Simon, *Nature*, 1990, **343**, 33–37.
- 94 T. Horn, C.-A. Chang and M. L. Collins, *Tetrahedron Lett.*, 1995, **36**, 2033–2036.
- 95 C. Roberts, R. Bandaru and C. Switzer, *Tetrahedron Lett.*, 1995, **36**, 3601–3604.
- 96 A. Jaworski, J. S. Kwiatkowski and B. Lesyng, *Int. J. Quantum Chem.*, 1985, **28**, 209–216.
- 97 X. Chen, R. Kierzek and D. H. Turner, *J. Am. Chem. Soc.*, 2001, **123**, 1267–1274.
- 98 F. Seela, C. Wei, A. Melenewski and E. Feiling, *Nucleosides Nucleotides*, 1998, **17**, 2045–2052.
- 99 A. Maciejewska, K. Lichota and J. Kusmierek, *Biochem. J.*, 2003, **369**, 611–618.
- 100 C. Switzer, S. Moroney and S. A. Benner, *J. Am. Chem. Soc.*, 1989, **111**, 8322–8323.
- 101 C. Y. Switzer, S. E. Moroney and S. A. Benner, *Biochemistry*, 1993, **32**, 10489–10496.
- 102 Y. Tor and P. Dervan, *J. Am. Chem. Soc.*, 1993, **115**, 4461–4467.
- 103 H. Kamiya and H. Kasai, *FEBS Lett.*, 1996, **391**, 113–116.
- 104 J. Horlacher, M. Hottiger, V. N. Podust, U. Hübscher and S. A. Benner, *Proc. Natl. Acad. Sci. U. S. A.*, 1995, **92**, 6329–6333.
- 105 M. J. Lutz, H. A. Held, M. Hottiger, U. Hübscher and S. A. Benner, *Nucleic Acids Res.*, 1996, **24**, 1308–1313.



- 106 M. J. Lutz, J. Horlacher and S. A. Benner, *Bioorg. Med. Chem. Lett.*, 1998, **8**, 499–504.
- 107 H. Kamiya and H. Kasai, *Nucleic Acids Res.*, 2000, **28**, 1640–1646.
- 108 H. Kamiya, H. Maki and H. Kasai, *Biochemistry*, 2000, **39**, 9508–9513.
- 109 Y. Tsurudome, T. Hirano, H. Kamiya, R. Yamaguchi, S. Asami, H. Itoh and H. Kasai, *Mutat. Res.*, 1998, **408**, 121–127.
- 110 D. S. Hitchcock, A. A. Fedorov, E. V. Fedorov, L. J. Dangott, S. C. Almo and F. M. Raushel, *Biochemistry*, 2011, **50**, 5555–5557.
- 111 T. Ohtsubo, K. Nishioka, Y. Imaiso, S. Iwai, H. Shimokawa, H. Oda, T. Fujiwara and Y. Nakabeppu, *Nucleic Acids Res.*, 2015, **43**, 3870–3871.
- 112 H. Kamiya and H. Kasai, *J. Radiat. Res.*, 2000, **41**, 349–354.
- 113 K. Fujikawa, H. Kamiya, H. Yakushiji, Y. Nakabeppu and H. Kasai, *Nucleic Acids Res.*, 2001, **29**, 449–454.
- 114 K. Fujikawa, H. Kamiya, H. Yakushiji, Y. Fujii, Y. Nakabeppu and H. Kasai, *J. Biol. Chem.*, 1999, **274**, 18201–18205.
- 115 J. C. Chaput and C. Switzer, *Proc. Natl. Acad. Sci. U. S. A.*, 1999, **96**, 10614–10619.
- 116 C. Switzer and J. C. Chaput, *Methods*, 2001, **23**, 141–148.
- 117 P. Schuster, G. Zundel and C. Sandorfy, *The Hydrogen Bond*, North Holland Co., New York, 1976, p. 403.
- 118 G. A. Jeffrey and W. Saenger, *Hydrogen Bonding in Biological Systems*, Springer-Verlag, New York, 1991, pp. 103–110.
- 119 F. Seela, C. Wei and A. Melenewski, *Origins Life Evol. Biospheres*, 1997, **27**, 597–608.
- 120 J. T. Davis and G. P. Spada, *Chem. Soc. Rev.*, 2007, **36**, 296–313.
- 121 T. Giorgi, F. Grepioni, I. Manet, P. Mariani, S. Masiero, E. Mezzina, S. Pieraccini, L. Saturni, G. P. Spada and G. Gottarelli, *Chem.–Eur. J.*, 2002, **8**, 2143–2152.
- 122 J. T. Davis, *Angew. Chem., Int. Ed.*, 2004, **43**, 668–698.
- 123 T. J. Pinnavaia, C. L. Marshall, C. M. Mettler, C. L. Fisk, H. T. Miles and E. D. Becker, *J. Am. Chem. Soc.*, 1978, **100**, 3625–3627.
- 124 M. L. Meyer and J. Suhnel, *J. Phys. Chem. A*, 2003, **107**, 1025–1031.
- 125 S. L. Forman, J. C. Fettinger, S. Pieraccini, G. Gottarelli and J. T. Davis, *J. Am. Chem. Soc.*, 2000, **122**, 4060–4067.
- 126 X. Shi, J. C. Fettinger and J. T. Davis, *J. Am. Chem. Soc.*, 2001, **123**, 6738–6739.
- 127 X. Shi, J. C. Fettinger and J. T. Davis, *Angew. Chem., Int. Ed.*, 2001, **40**, 2827–2831.
- 128 X. Shi, J. C. Fettinger and J. T. Davis, *Angew. Chem.*, 2001, 2909–2913.
- 129 X. Shi, K. M. Mullaugh, J. C. Fettinger, Y. Jiang, S. A. Hofstadler and J. T. Davis, *J. Am. Chem. Soc.*, 2003, **125**, 10830–10841.
- 130 M. Cai, A. L. Marlow, J. C. Fettinger, D. Fabris, T. J. Haverlock, B. A. Moyer and J. T. Davis, *Angew. Chem., Int. Ed.*, 2000, **39**, 1283–1285.
- 131 A. L. Marlow and J. T. Davis, *Tetrahedron Lett.*, 1999, **40**, 3539–3542.
- 132 T. Evan-Salem, L. Frish, F. W. B. Van Leeuwen, D. N. Reinhoudt, W. Verboom, M. S. Kaucher, J. T. Davis and Y. Cohen, *Chem.–Eur. J.*, 2007, **13**, 1969–1977.
- 133 V. Abet, R. Evans, F. Guibbal, S. Caldarelli and R. Rodriguez, *Angew. Chem.*, 2014, **126**, 4962–4966.
- 134 M. Cai, X. Shi, V. Sidorov, D. Fabris, Y.-F. Lam and J. T. Davis, *Tetrahedron*, 2002, **58**, 661–671.
- 135 E. H. Blackburn and J. W. Szostak, *Annu. Rev. Biochem.*, 1984, **53**, 163–194.
- 136 J. R. Williamson, M. K. Raghuraman and T. R. Cech, *Cell*, 1989, **59**, 871–880.
- 137 F. Aboul-Ela, A. I. H. Murchie, D. G. Norman and D. M. J. Lilley, *J. Mol. Biol.*, 1994, **243**, 458–471.
- 138 A.-E. Fareed, I. H. M. Alastair and M. J. L. David, *Nature*, 1992, **360**, 280–282.
- 139 W. Yong and D. J. Patel, *Biochemistry*, 1992, **31**, 8112–8119.
- 140 F. Seela, C. Wei and A. Melenewski, *Nucleic Acids Res.*, 1996, **24**, 4940–4945.
- 141 C. Roberts, J. C. Chaput and C. Switzer, *Chem. Biol.*, 1997, **4**, 899–908.
- 142 E. A. Venczel and D. Sen, *Biochemistry*, 1993, **32**, 6220–6228.
- 143 F. Seela and R. Kröschel, *Bioconjugate Chem.*, 2001, **12**, 1043–1050.
- 144 F. Seela and C. Wei, *Chem. Commun.*, 1997, 1869–1870.
- 145 F. Seela, C. Wei, A. Melenewski, Y. He, R. Kröschel and E. Feiling, *Nucleosides Nucleotides*, 1999, **18**, 1543–1548.
- 146 F. Seela and C. Wei, *Helv. Chim. Acta*, 1999, **82**, 726–745.
- 147 N. B. Ramsing and T. M. Jovin, *Nucleic Acids Res.*, 1988, **16**, 6659–6676.
- 148 F. Seela, B. Gabler and Z. Kazimierczuk, *Collect. Czech. Chem. Commun.*, 1993, **58**, 170–173.
- 149 F. Seela, C. Wei and A. Melenewski, *Nucleosides Nucleotides*, 1997, **16**, 1523–1527.
- 150 X.-L. Yang, H. Sugiyama, S. Ikeda, I. Saito and A. H. J. Wang, *Biophys. J.*, 1998, **75**, 1163–1171.
- 151 F. Seela, Y. He and C. Wei, *Tetrahedron*, 1999, **55**, 9481–9500.
- 152 S. S. Pujari and F. Seela, *J. Org. Chem.*, 2013, **78**, 8545–8561.
- 153 N. M. Sangeetha and U. Maitra, *Chem. Soc. Rev.*, 2005, **34**, 821–836.
- 154 I. Bang, *Biochem. Z.*, 1910, **26**, 293–311.
- 155 H. Zhao, A. H. Schäfer and F. Seela, *ChemPlusChem*, 2017, **82**, 826–833.
- 156 H. Zhao, D. Jiang, A. H. Schäfer and F. Seela, *ChemPlusChem*, 2017, **82**, 778–784.
- 157 M. Guvendiren, H. D. Lu and J. A. Burdick, *Soft Matter*, 2011, **8**, 260–272.
- 158 A. Biswas, S. Malferrari, D. M. Kalaskar and A. K. Das, *Chem. Commun.*, 2018, **54**, 1778–1781.
- 159 F. Tang, H. Feng, Y. Du, Y. Xiao, H. Dan, H. Zhao and Q. Chen, *Chem.–Asian J.*, 2018, **13**, 1962–1971.
- 160 B. Adhikari, A. Shah and H.-B. Kraatz, *J. Mater. Chem. B*, 2014, **2**, 4802–4810.
- 161 J. Kim, S. Lee, Y. Han, J. Moon and J. Kim, *Arch. Pharmacol. Res.*, 1994, **17**, 115–118.
- 162 V. Venkatesh, N. K. Mishra, I. Romero-Canelon, R. R. Vernooij, H. Shi, J. P. C. Coverdale,



- A. Habtemariam, S. Verma and P. J. Sadler, *J. Am. Chem. Soc.*, 2017, **139**, 5656–5659.
- 163 H. Zhao, H. Feng, J. Liu, F. Tang, Y. Du, N. Ji, L. Xie, X. Zhao, Z. Wang and Q. Chen, *Biomaterials*, 2019, **230**, DOI: 10.1016/j.biomaterials.2019.119598.
- 164 M. M. Cox, *Mutat. Res.*, 1997, **384**, 15–22.
- 165 H. Kamiya and H. Kasai, *Biochemistry*, 1997, **36**, 11125–11130.
- 166 M. Inoue, H. Kamiya, K. Fujikawa, Y. Ootsuyama, N. Murata-Kamiya, T. Osaki, K. Yasumoto and H. Kasai, *J. Biol. Chem.*, 1998, **273**, 11069–11074.
- 167 S. C. Johnson, C. B. Sherrill, D. J. Marshall, M. J. Moser and J. R. Prudent, *Nucleic Acids Res.*, 2004, **32**, 1937–1941.
- 168 S. C. Johnson, D. J. Marshall, G. Harms, C. M. Miller, C. B. Sherrill, E. L. Beaty, S. A. Lederer, E. B. Roesch, G. Madsen, G. L. Hoffman, R. H. Laessig, G. J. Kopish, M. W. Baker, S. A. Benner, P. M. Farrell and J. R. Prudent, *Clin. Chem.*, 2004, **50**, 2019–2027.
- 169 J. D. Bain, S. Christopher, C. Richard and A. B. Steven, *Nature*, 1992, **356**, 537–539.
- 170 F. Seela, D. Jiang and S. Budow, *ChemBioChem*, 2010, **11**, 1443–1450.
- 171 F. A. Fuhrman and G. J. Fuhrman, *Comp. Biochem. Physiol., C: Comp. Pharmacol.*, 1982, **72**, 203–210.
- 172 P. L. Ewing, F. Schlenk and G. A. Emerson, *J. Pharmacol. Exp. Ther.*, 1949, **97**, 379–383.
- 173 D. A. Clarke, J. Davoll, F. S. Philips and G. B. Brown, *J. Pharmacol. Exp. Ther.*, 1952, **106**, 291–302.
- 174 K. Weronika, C. Tomasz and P. Anna, *PLoS One*, 2018, **13**, e0197835.
- 175 F. A. Fuhrman, G. J. Fuhrman, Y. H. Kim, L. A. Pavelka and H. S. Mosher, *Science*, 1980, **207**, 193–195.
- 176 P. Buckle and I. Spence, *Naunyn-Schmiedeberg's Arch. Pharmacol.*, 1981, **316**, 64–68.
- 177 J. F. Marwood, *Clin. Exp. Pharmacol. Physiol.*, 1981, **8**, 575–584.
- 178 D. Jamieson and P. Davis, *Eur. J. Pharmacol.*, 1980, **67**, 295–300.
- 179 M. Radulovacki, R. M. Virus, D. Rapoza and R. C. Crane, *Neuropharmacology*, 1985, **24**, 547–549.
- 180 P. Buckle and I. Spence, *Naunyn-Schmiedeberg's Arch. Pharmacol.*, 1982, **319**, 130–135.
- 181 M. Huang, H. Shimizu and J. W. Daly, *J. Med. Chem.*, 1972, **15**, 462–466.
- 182 L. P. Davies, K. M. Taylor, R. P. Gregson and R. J. Quinn, *Life Sci.*, 1980, **26**, 1079–1088.
- 183 L. P. Davies, J. Baird-Lambert and J. G. Hall, *Neuropharmacology*, 1987, **26**, 493–497.
- 184 W. J. McDowell and G. N. Case, *Anal. Chem.*, 1992, **64**, 3013–3017.
- 185 J. C. Bryan, K. Kavallieratos and R. A. Sachleben, *Inorg. Chem.*, 2000, **39**, 1568–1572.
- 186 J. C. Bryan, R. A. Sachleben and B. P. Hay, *Inorg. Chim. Acta*, 1999, **290**, 86–94.
- 187 Y. Deng, R. A. Sachleben and B. A. Moyer, *J. Chem. Soc., Faraday Trans.*, 1995, **91**, 4215–4222.
- 188 R. A. Sachleben, Y. Deng, D. R. Bailey and B. A. Moyer, *Solvent Extr. Ion Exch.*, 1996, **14**, 995–1015.
- 189 J. T. Davis, S. K. Tirumala and A. L. Marlow, *J. Am. Chem. Soc.*, 1997, **119**, 5271–5272.
- 190 A. Casnati, A. Pochini, R. Ungaro, F. Ugozzoli, F. Arnaud, S. Fanni, M.-J. Schwing, R. J. M. Egberink, F. d. Jong and D. N. Reinhoudt, *J. Am. Chem. Soc.*, 1995, **117**, 2767.
- 191 S. Lee, J. Lamb, M. Cai and J. Davis, *J. Inclusion Phenom. Macrocyclic Chem.*, 2001, **40**, 51–57.
- 192 F. W. B. van Leeuwen, W. Verboom, X. Shi, J. T. Davis and D. N. Reinhoudt, *J. Am. Chem. Soc.*, 2004, **126**, 16575–16581.
- 193 F. W. B. van Leeuwen, C. J. H. Miermans, H. Beijleveld, T. Tomasberger, J. T. Davis, J. T. Davis, W. Verboom and D. Reinhoudt, *Environ. Sci. Technol.*, 2005, **39**, 5455–5459.
- 194 S. Lee, J. Kim, S. Lee and J. Kim, *Arch. Pharmacol. Res.*, 1995, **18**, 138–139.
- 195 R. Nowak, M. Sawardo-Rochowska, Z. Kazimierzczuk and R. J. Nowak, *Eksperimental'naia Onkologiya*, 2004, **26**, 20–23.
- 196 C. Hagen, *Biochim. Biophys. Acta*, 1973, **293**, 105–110.
- 197 J. Doskocil and A. Holý, *Nucleic Acids Res.*, 1974, **1**, 645–652.
- 198 J. H. Wissler, S. Kiesewetter, E. Logemann, M. Sprinzl and L. M. G. Hellmeyer Jr, *Biol. Chem.*, 1988, **369**, 948–949.

