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# Measuring nanoparticle-induced resonance energy transfer effect by electrogenerated chemiluminescent reactions†

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Electrogenerated chemiluminescence (ECL) efficiencies, redox potentials, photoluminescent (PL) (quenching and coupling) effects, and AFM images for the [Ru(bpy)<sub>3</sub>]<sup>2+</sup>/Au@tiopronin system were determined in aqueous solutions of the gold nanoparticles (NPs) at pH 7.0. The most remarkable finding was that ECL measurements can display the nanoparticle-induced resonance energy transfer (NP-RET) effect. Its effectiveness was quantified through a coefficient,  $K_{(NP-RET)ECL}$ , which measures how much an ECL reaction has been enhanced. Moreover, the NP-RET effect was also checked using PL measurements, in such a way that a coefficient,  $K_{(NP-RET)PL}$ , was determined; both constants,  $K_{(NP-RET)ECL}$  and  $K_{(NP-RET)PL}$  being in close agreement. It is important to highlight the fact that the NP-RET effect is only displayed in diluted solutions in which there is no NPs self-aggregation. The existence of the NPs self-aggregation behavior is revealed through AFM measurements.

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## 1. Introduction

In a broad sense relevant aspects of non-covalent interactions in supramolecular systems can be studied using the Förster resonance energy transfer (FRET) approach.<sup>1–8</sup> In this type of energy transfer process an excited donor fluorophore (dye) transfers its energy to an acceptor molecule, which normally relaxes by emitting light if it is photoluminescent (PL); if this species is not itself PL, then there is a quenching effect. Certain requirements must be met for the process to take place.<sup>5–7</sup> First, sufficient spectral overlap is required between the emission donor and absorption acceptor species.<sup>5</sup> Second, the distance between the molecules must be less than approximately 10 nm.<sup>6</sup> Finally, the fluorophores should be conveniently oriented with respect to each other.<sup>7</sup> In fact, FRET efficiency depends on these three factors, and is highly sensitive to the donor–acceptor distance ( $R$ ). Therefore, using the concept of FRET coupling, quantitative information can be obtained about several relevant characteristics of supramolecular phenomena.<sup>8</sup>

However, a major limitation of FRET interactions is the relatively small distance ( $\approx 10$  nm) at which the energy can be transferred. There are several ways to resolve this situation. One of these comprises the formation of cascades, in which several

FRET fluorophores with increasingly longer excitation wavelengths are organized in devices to facilitate energy transfer over distances greater than 10 nm or more.<sup>9–11</sup> Efforts made in this regard, outside the regime of interaction between point dipoles, are also important. That is, resonance energy transfer (RET) efficiency has been predicted based on  $R^{-4}$  or  $R^{-2}$  donor–acceptor distance dependence<sup>12–14</sup> and not on  $R^{-6}$  (Förster theory). Thus, the energy can be transferred over a distance of 20 nm from an organic dye to a spherical metal nanoparticle (NP) which acts as a surface better than a point dipole.<sup>15</sup> Currently, FRET is being replaced by plasmon-enhanced or metal-induced resonance energy transfer, which as demonstrated can work at distances of up to 100 nm.<sup>15–18</sup> Therefore, the challenge now is to use metal NPs for the study of non-covalent interactions in supramolecular media using FRET or plasmon-enhanced techniques.

In addition, the study of the plasmon–molecule interaction is gaining increasing attention.<sup>4,19–21</sup> In point of fact, the enhancement of photoluminescence (PL) by a nanoparticle uses in an effective way the coupling between excited dyes and the localized surface plasmon resonance (SPR) in metal nanostructures in an effective way. This interaction effect has contributed to the development of nanotechnology:<sup>22–27</sup> for instance, solar energy storage and conversion<sup>22–24</sup> and optical biosensing,<sup>25–27</sup> rely on this type of the energy transfer.

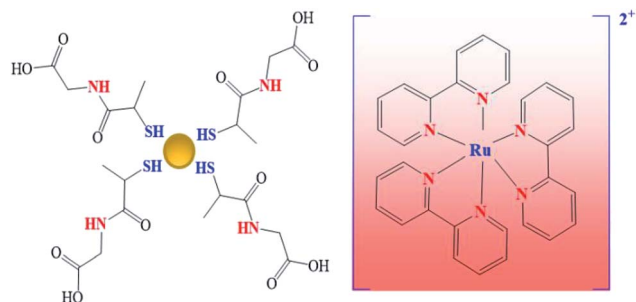
Based on the preceding results, the subject of plasmon–molecule couplings is of interest because the understanding of these interactions can help address important issues related to the potential applications of nanomaterials and supramolecular media. The aim of this study is to obtain a deeper insight

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Scheme 1 Au@tiopronin/[Ru(bpy)<sub>3</sub>]<sup>2+</sup> system.

into the plasmon–molecule interactions using a high-energy electrogenerated chemiluminescent (ECL) reaction in colloidal gold solutions. ECL is a process that follows several fundamental pathways. In the first step, an electrode originates species that in successive paths will give rise to high-energy electron-transfer reactions in solution, which in turn will produce one or two species in their excited state.<sup>28–37</sup>

After about 50–60 years of study, ECL has proven to be a useful research tool in a wide variety of areas.<sup>31–39</sup> Indeed, there are studies in which the increase in ECL due to the coupling between plasmons and dyes is applied to the detection of different species.<sup>38–42</sup> These studies have contributed to the development of the ECL reactions as an analytical technique, given their significant impact. However, to our knowledge, there are few studies in which the magnitude of the coupling effect is quantified.<sup>43</sup>

To study plasmon–molecule interactions was used the ECL reaction, [Ru(bpy)<sub>3</sub>]<sup>2+</sup> + C<sub>2</sub>O<sub>4</sub><sup>2–</sup>, in the presence of several Au@tiopronin<sup>†</sup> NPs (see Scheme 1) aqueous solutions. In addition, photoluminescent (PL) (quenching and coupling) of the [Ru(bpy)<sub>3</sub>]<sup>2+\*</sup> species as well as atomic force microscopy (AFM) measurements were carried out; redox potentials of the [Ru(bpy)<sub>3</sub>]<sup>3+/2+</sup> couple were also determined in several NP solutions. The noteworthy findings were that ECL measurements can display the emission-dye-NPs coupling effect or as we have called it, the nanoparticle-induced resonance energy transfer (NP-RET) effect, and it is also possible to quantify its effectiveness. A coefficient was defined which measures the efficiency that a given system of NPs has for the improvement of ECL in a particular reaction. Specifically, this coefficient measures how much an ECL reaction is improved by the NP-RET effect. The novelty here lies in using an ECL reaction as a technique to measure the NP-RET efficiency of a dye-NP system and confirm

<sup>†</sup> The gold clusters protected with tiopronin are NPs of alkanethiolate in which the presence of a carboxylic and an amino group on the tiopronin can modify the charge of the particles depending on the medium's pH. Consequently, this colloidal system offers the possibility of being positively or negatively charged. In fact, at pK<sub>a</sub> ≈ 5.6, Au@tiopronin nanoparticles are commonly neutral and hydrophilic.<sup>44,45</sup> Thus, at pH 7.0, for example, the Au@tiopronin NPs will be negatively charged. The judicious choice of the medium pH will determine not only how these NPs interact with the surrounding solvent molecules but also the aggregation of the nanoparticles themselves.<sup>46</sup> In order to minimize the self-aggregation, the medium pH chosen was 7.0.

its value *via* photoluminescence measurements. All experiments were carried out at pH 7.0 and 298.15 ± 0.01 K.

## 2. Materials and methods

### 2.1. Materials

All chemicals [Ru(bpy)<sub>3</sub>]<sub>2</sub>Cl<sub>2</sub>, Na<sub>2</sub>C<sub>2</sub>O<sub>4</sub>, CH<sub>3</sub>AsO<sub>2</sub>Na<sub>3</sub>H<sub>2</sub>O (cacodylate), NaCl, HCl, *N*-(2-mercapto-propionyl)glycine (tiopronin), HAuCl<sub>4</sub>·3H<sub>2</sub>O, MeOH, CH<sub>3</sub>COOH and NaBH<sub>4</sub> were analytical grade, purchased from Sigma-Aldrich and used without further purification. Gold nanoparticles were synthesized following the method of Templeton *et al.*<sup>45</sup> Dialysis sacks 25EA from Sigma were utilized for purification of the gold nanoparticles. These were characterized by UV-visible absorption spectra (*vide infra*) (Fig. 1), TEM microscopy (*vide infra*) (Fig. 2) and C, H, N, and S microanalysis (11.80% C; 1.86% H; 2.89% N; 7.37% S). From gold core size distribution (see Fig. 2) the average size of Au colloids was 3.3 ± 0.5 nm. Accordingly, the relation between the number of Au atoms and tiopronin ligands<sup>45</sup> was 314/101. Consequently, the molecular weight calculated for C<sub>505</sub>H<sub>909</sub>O<sub>303</sub>N<sub>101</sub>S<sub>101</sub>Au<sub>314</sub> was 79 680 g mol<sup>–1</sup>. All the solutions were prepared with deionized water obtained from a Millipore Milli-Q system, having a conductivity less than <10<sup>–6</sup> S m<sup>–1</sup>. All experiments were carried out with Au colloid solutions prepared by weight.

### 2.2. Electrogenerated chemiluminescence measurements

ECL measurements (cyclic voltammetry (CV) experiments plus ECL emissions) were carried out as in previous works.<sup>47–49</sup> All ECL measurements were recorded using the CV technique working at a scan rate of 100 mV s<sup>–1</sup>. The uncertainty of the relative measured ECL efficiency (Φ<sub>ECL</sub>) (see Section 3.1) was less than 4% from the average of two or three measurements in

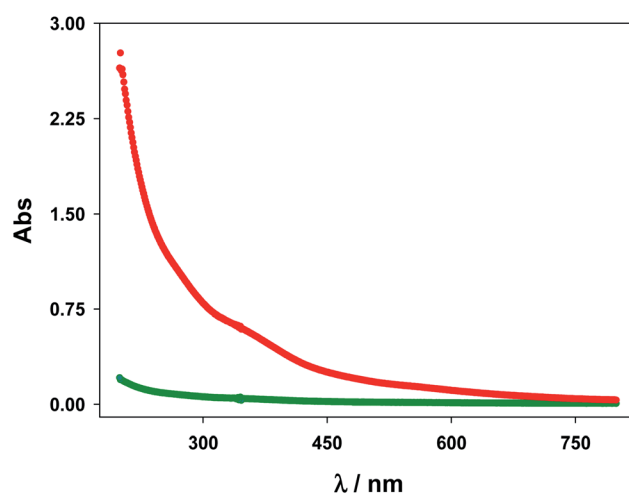


Fig. 1 UV-visible absorption spectra of the Au@tiopronin nanoparticles in the absence of the [Ru(bpy)<sub>3</sub>]<sup>2+</sup> species and presence of the cacodylate buffer, pH 7.0 (*vide infra*). Green and red lines denote 2.0 × 10<sup>–7</sup> M and 2.0 × 10<sup>–6</sup> M [Au@tiopronin]. As can be seen, a slight but detectable surface plasmon band (SPR) appears as a consequence of the small size of nanoclusters when the [Au@tiopronin] increases.



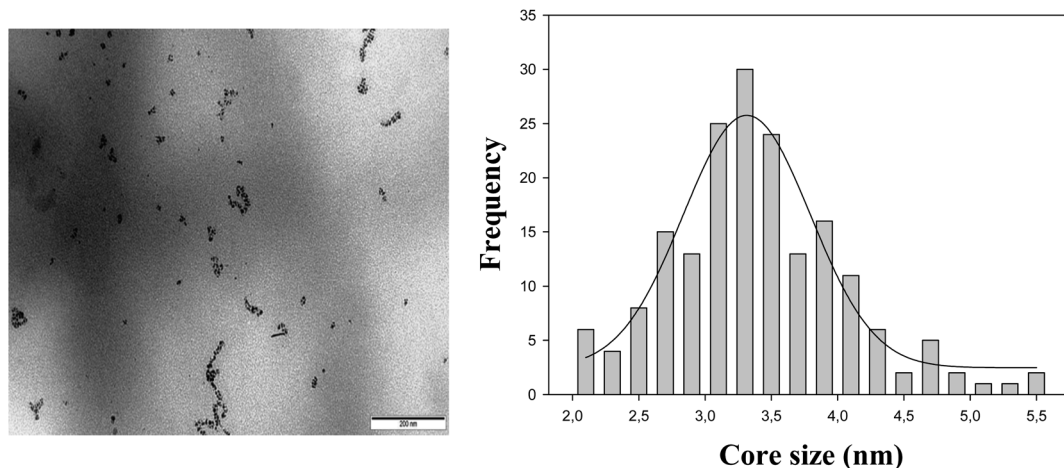


Fig. 2 TEM image (left) and size distribution (right) of the tiopronin gold nanoparticles in the absence of the Ru(II) complex and presence of the cacodylate buffer at pH 7.0.

four scans, after system calibration. ECL measurements were taken for several AuNPs solutions (from  $1.00 \times 10^{-8}$  M to  $1.00 \times 10^{-6}$  M (see Table 1)).  $2.0 \times 10^{-4}$  M and  $2.0 \times 10^{-3}$  M were always the concentrations of  $[\text{Ru}(\text{bpy})_3]^{2+}$  and  $\text{C}_2\text{O}_4^{2-}$ , respectively. All experiments were carried out at pH 7.0 by using a buffer  $3.0 \times 10^{-2}$  M  $\text{NaCH}_3\text{AsO}_2/3.8 \times 10^{-3}$  M HCl plus  $6.6 \times 10^{-3}$  M NaCl. In this way, the ionic strength was also fixed at 0.040 M in each solution.

### 2.3. Electrochemical measurements

Peak potentials ( $E_{\text{peak}}$ ) for the  $[\text{Ru}(\text{bpy})_3]^{3+/2+}$  couple in several AuNPs solutions were determined using the differential pulse voltammetry (DPV) technique, as in previous works.<sup>47–49</sup> The

experimental conditions were identical with those corresponding to ECL measurements (*vide supra*).

### 2.4. Spectroscopic measurements

A Cary 500 Scan UV-vis-NIR spectrophotometer was used to record the absorption spectra of the AuNP solutions. Fig. 1 shows characteristic UV-vis spectra of some Au@tiopronin solutions as an example. As can be seen a slight detectable surface plasmon band (SPR) appears as a consequence of the small size of nanoclusters.

Photoluminescent spectra for the  $[\text{Ru}(\text{bpy})_3]^{2+*}$  excited species were acquired with a Photon Technology International (PTI). Several sets of experiments were carried out in the presence of the cacodylate buffer at pH 7.0. To study the quenching effect (see Section 3.4) in the first set of experiments, the excitation wavelength was that corresponding to the absorption maximum (454 nm), whereas in the second set of experiments it was 500 nm, the one corresponding to an absorbance of 0.2. The concentrations of the  $[\text{Ru}(\text{bpy})_3]^{2+}$  complex were  $2.0 \times 10^{-6}$  M and  $2.0 \times 10^{-4}$  M, respectively. To study the coupling effect (see Section 3.5) the excitation wavelength was 454 nm and  $4.0 \times 10^{-7}$  M  $[\text{Ru}(\text{bpy})_3]^{2+}$ .

### 2.5. Transmission electronic microscopy (TEM) measurements

TEM analysis was carried out using a Philips CM 200 electron microscope working at 200 kV. 10  $\mu\text{L}$  of the AuNPs aqueous solution was placed on a copper grid coated with a carbon film. The grid was left to air dry for several hours at room temperature. The solutions were prepared in the absence of the Ru(II) complex and presence of the cacodylate buffer at pH 7.0 (see Fig. 2).

### 2.6. Atomic force microscopy (AFM) measurements

Images were obtained with a Molecular Imaging PicoPlus 2500 AFM (Agilent Technologies). Silicon cantilevers (Model

Table 1 Relative ECL efficiencies ( $\Phi_{\text{ECL}}$ ), integrated charges (Q) and integrated ECL emission intensities ( $I_{\text{ECL}}$ ) for  $[\text{Ru}(\text{bpy})_3]^{2+} + \text{C}_2\text{O}_4^{2-}$  reaction, and redox potentials ( $E_{\text{peak}}$ ) of the  $[\text{Ru}(\text{bpy})_3]^{3+/2+}$  couple for several Au@tiopronin nanoparticle solutions<sup>a</sup>

$10^8 \times [\text{AuNPs}]/\text{M}$	$10^2 \times \Phi_{\text{ECL}}$	$10^3 \times Q/C$	$10^{-6} \times I_{\text{ECL}}/\text{counts}$	$E_{\text{peak}}/V$
0.00	3.85	0.149	0.304	1.092
1.00	4.55	0.341	0.341	1.097
3.00	5.23	0.483	0.587	1.112
4.00	5.81	0.453	0.609	1.087
5.00	6.22	0.404	0.641	1.082
5.50	5.78	0.339	0.497	1.087
6.00	4.98	0.326	0.414	1.092
7.00	4.46	0.395	0.426	1.095
8.00	3.39	0.341	0.267	1.097
10.0	2.74	0.366	0.257	1.092
13.0	2.95	0.309	0.239	1.107
40.0	2.35	0.360	0.218	1.106
50.0	2.51	0.320	0.194	1.112
100	2.49	0.310	0.199	1.115

<sup>a</sup> pH 7.0, cacodylate buffer ( $[\text{NaCH}_3\text{AsO}_2] = 3.0 \times 10^{-2}$  M,  $[\text{HCl}] = 3.8 \times 10^{-3}$  M),  $6.2 \times 10^{-3}$  M NaCl. Ionic strength 0.040 M.



Pointprobe, Nanoworld) with a resonance frequency of around 240 kHz and nominal force constant of  $42 \text{ N m}^{-1}$  were used. All AFM images were recorded in air and in tapping mode, with scan speeds of about 0.5 Hz and data collection at  $256 \times 256$  pixels. At least two mica specimens were prepared for each solution, and at least 6 distinct areas of each specimen were investigated using the sample probe tip. The solutions were prepared in the absence and presence of  $2.0 \times 10^{-4} \text{ M}$   $[\text{Ru}(\text{bpy})_3]^{2+}$  complex and  $2.0 \times 10^{-3} \text{ M}$   $\text{C}_2\text{O}_4^{2-}$  plus cacodylate buffer at pH 7.0 plus the suitable  $[\text{AuNP}]$ .

100  $\mu\text{L}$  of solutions was dropped onto a mica surface and adsorbed for 30 min in a humidified chamber. Following incubation, samples were washed with ultra-pure water and subsequently air dried for AFM imaging. In order to prevent the formation of salt crystals on the mica surface, the rinse was performed without cacodylate buffer.

### 3. Results

#### 3.1. Electrogenerated chemiluminescence

In order to determine the ECL efficiency values (*vide infra*) the ECL emission-time and current-time curves corresponding to a given CV voltammogram were obtained. Fig. 3 shows a set of experimental measurements generated *via* CV as an example: current-time curves for  $[\text{Ru}(\text{bpy})_3]^{3+/2+}$  couple (top) and their corresponding ECL intensity-time curves for  $[\text{Ru}(\text{bpy})_3]^{2+*}$  excited state (bottom), (see also Fig. S1† (ECL spectrum) and Fig. S2† (CV voltammogram) in ESI†). Of course, background experiments were done for all cases, whereas ECL emissions were irrelevant, the current-time curves of the backgrounds for all the measurements were taken into account when calculating ECL efficiency values.

The ECL emission efficiency or ECL quantum yield ( $\Phi_{\text{ECL}}$ ) (emitted photon per transferred proton) can be approximately defined by the coulometric efficiency by:<sup>50–52</sup>

$$\Phi_{\text{ECL}} = \frac{\int_0^t I_e dt}{\int_0^t i dt \left( \frac{N_A}{F} \right)} \quad (1)$$

where  $I_e$  and  $i$  are the intensity in photons per second and the current in amperes (coulombs per second), respectively, integrated over a finite period of time,  $t$ .  $F$  and  $N_A$  are the Faraday and Avogadro constants, respectively. As an alternative, a standard reaction is often used, simplifying both the experimental measurements and the calculation of  $\Phi_{\text{ECL}}$ .<sup>47–49</sup>

$$\Phi_{\text{ECL}} = \Phi_{\text{ECL}}^{\circ} \frac{Q^{\circ} I_{\text{ECL}}}{Q I_{\text{ECL}}^{\circ}} \quad (2)$$

where  $Q$  and  $Q^{\circ}$  are the integrated charge for the actual and standard reactions, respectively;  $I_{\text{ECL}}$  and  $I_{\text{ECL}}^{\circ}$  are the integrated photon intensities for both cases.  $\Phi_{\text{ECL}}^{\circ}$  is the ECL efficiency in the absence of Au@tiopronin nanoparticles for a standard reaction ( $[\text{Ru}(\text{bpy})_3]^{2+} + \text{C}_2\text{O}_4^{2-}$  in 0.01 M phosphate buffer (pH 6.1) + 0.1 M NaCl)<sup>53</sup> equal to 0.0185 at 298.15 K.<sup>47</sup> Relative ECL efficiency values were determined by means of eqn (1) and (2) for all NP solutions; the results are given in Table 1.

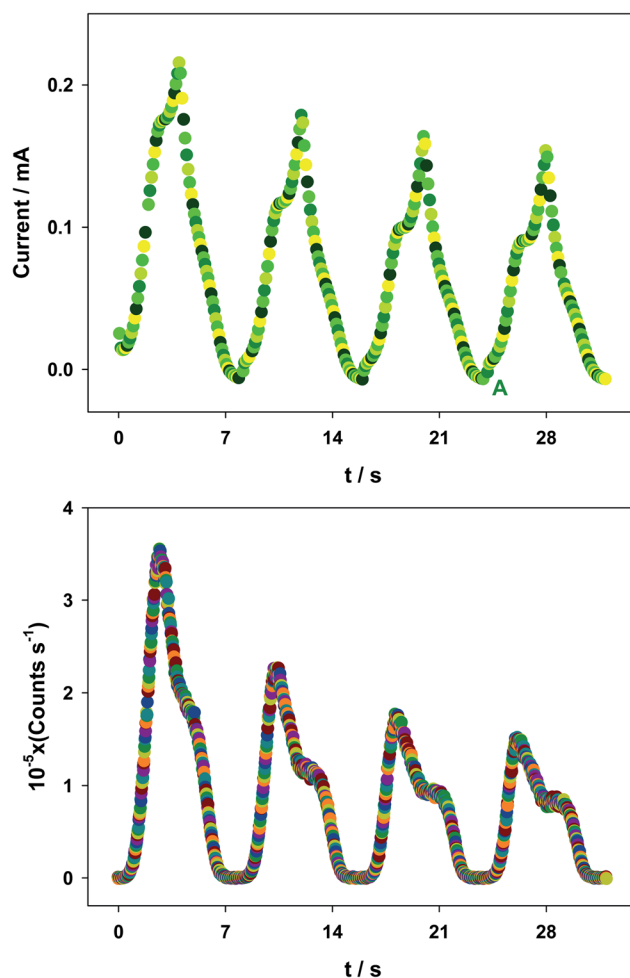


Fig. 3 Current (top) and ECL intensity (bottom) as functions of time in a CV measurement of four scans for the  $[\text{Ru}(\text{bpy})_3]^{2+} + \text{C}_2\text{O}_4^{2-}$  reaction in  $5.0 \times 10^{-8} \text{ M}$  AuNPs. Scan rate:  $100 \text{ mV s}^{-1}$ . Cacodylate buffer (pH 7.0). Ionic strength 0.040 M.

#### 3.2. Redox potentials

In order to obtain some information about the microenvironment around the  $[\text{Ru}(\text{bpy})_3]^{2+}$  complex, redox potentials were evaluated. The DPV technique was used to determine the redox potential values by means of eqn (3), which relates the peak and half-wave potentials for a reversible system:

$$E_{\text{peak}} = E_{1/2} \pm \Delta E/2 \quad (3)$$

where  $\Delta E$  is the pulse voltage amplitude (2.5 mV in our experiments) and the signs  $\pm$  denote anodic or cathodic scans, thus  $E_{\text{peak}} \approx E_{1/2}$ , resulting, their uncertainty in the values measured being about  $\pm 3 \text{ mV}$ . Fig. 4 illustrates a typical DPV voltammogram as an example. The results of redox potentials are collected in Table 1. As can be seen there is no significant variation in the values of the redox potentials. This fact seems to indicate that the addition of increasing amounts of nanoparticles does not affect the environment surrounding the  $[\text{Ru}(\text{bpy})_3]^{3+/2+}$  couple. This feature seems to show that the electric field created by the charged NPs is very weak and,





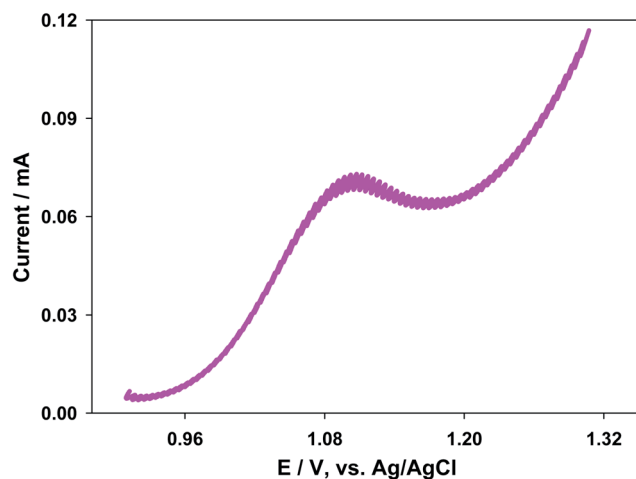


Fig. 4 Differential pulse voltammogram for the  $[\text{Ru}(\text{bpy})_3]^{3+/2+}$  couple in the presence of  $7.0 \times 10^{-8}$  M  $[\text{AuNPs}]$ . Cacodylate buffer (pH 7.0). Ionic strength 0.040 M.

consequently, the variation of the interaction between NPs and the  $\text{Ru}(\text{II})$  and  $\text{Ru}(\text{III})$  species is not significant.

### 3.3. Self-aggregation performance

In order to rationalize the trends of  $\Phi_{\text{ECL}}/[\text{AuNPs}]$  self-aggregation measurements were necessary. To this end, AFM method was carried out. Fig. S3† (bottom) and 5 display AFM images for  $8.0 \times 10^{-8}$  M  $[\text{AuNPs}]$  in the absence and presence of  $2.0 \times 10^{-4}$  M  $[\text{Ru}(\text{bpy})_3]^{2+}$ , respectively. Likewise, Fig. S3† (top) and 6 show the images at  $1.0 \times 10^{-8}$  M  $[\text{AuNPs}]$ .

Comparison between Fig. 5 and 6 allowed us to verify the self-aggregation of particles. As can be seen in Fig. 5 when the concentration is  $8.0 \times 10^{-8}$  M, large aggregates can be distinguished. However, in the case of solutions at  $1.0 \times 10^{-8}$  M  $[\text{AuNPs}]$ , aggregation does not seem to occur (Fig. 6). In addition, the  $\text{Ru}(\text{II})$  complex induced the formation of more aggregates (see Fig. S3† (top and bottom)), but this does not determine their formation in any of the cases. That is to say, at  $1.0 \times 10^{-8}$  M  $[\text{AuNPs}]$ , in the absence or presence of the  $\text{Ru}(\text{II})$  complex there is no aggregation (see Fig. S3† (top) and 6). For both histograms (see Fig. 5 and 6), the average diameters of NPs at  $8.0 \times 10^{-8}$  M and  $1.0 \times 10^{-8}$  M  $[\text{AuNPs}]$  were  $8.6 \pm 0.9$  nm and  $3.7 \pm 0.6$  nm, respectively. Comparison between them allows us to discover which monomers on average form a NP self-aggregate,  $n = 2.3 \pm 0.6$ .

### 3.4. Quenching effect

To verify whether the quenching effect was at work or not under our experimental conditions, the photoluminescent spectra of the  $[\text{Ru}(\text{bpy})_3]^{2+*}$  species were recorded for the whole concentration range of AuNPs (see Table 1). Fig. S4A† displays some spectra as examples (see ESI† for details). Bear in mind that there is no significant quenching effect in diluted solutions. For this reason, the  $[\text{AuNP}]$  was increased up to  $2.0 \times 10^{-6}$  M.

The Stern–Volmer approach can be applied in order to take into account the magnitude of the quenching effect:

$$\left(\frac{I_0}{I}\right)_{\text{PL}} = 1 + (K_D + K_S)[\text{AuNPs}] \quad (4)$$

where  $K_D$  and  $K_S$  are the dynamic and static quenching constants, respectively,  $[\text{AuNPs}]$  is the [quencher], and  $(I_0/I)_{\text{PL}}$  the emission ratio for  $\text{Ru}(\text{II})^*$  species (with  $I_0$  and  $I$  having their usual meaning). Eqn (4), which predicts a linear relationship between  $[\text{AuNPs}]$  and  $(I_0/I)_{\text{PL}}$ , is fulfilled as shown in Fig. 7, from which slope  $K_Q = (K_D + K_S)$  was calculated, being equal to  $4.5 \times 10^5 \text{ M}^{-1}$ . Notice that if the concentration of NPs is further increased up to  $4.0 \times 10^{-6}$  M a Stern–Volmer quadratic equation taking into account static ( $K_S$ ) and dynamic ( $K_D$ ) quenching constants may be operative (see eqn (1S), Fig. S4B and S5†). However, although the fit between experimental data and eqn (1S)† is good the results for  $K_S$  and  $K_D$  are not consistent. This behavior could be due to the formation of self-aggregates, which seems to be more significant when the concentration of the AuNPs increases as demonstrated previously (see Section 3.3).

### 3.5. Coupling effect

Another set of photoluminescent spectra was recorded to ascertain whether the photoluminescent intensity of the  $[\text{Ru}(\text{bpy})_3]^{2+*}$  species increases or not in the presence of the Au@tiopronin particles at a range of diluted concentrations. These spectra were carried out at  $1.0 \times 10^{-8}$  M and  $8.0 \times 10^{-8}$  M  $[\text{AuNPs}]$  under buffer conditions (pH 7.0) at  $4.0 \times 10^{-7}$  M  $[\text{Ru}(\text{bpy})_3]^{2+}$ . Fig. 8 clearly shows that the photoluminescent intensity of the  $[\text{Ru}(\text{bpy})_3]^{2+*}$  species increases at the diluted solutions of the NPs, which is not so for the concentrated solutions (see Fig. S6†). For the latter solutions, a slight decay of light intensity is observed.

## 4. Discussion

As shown in Table 1,  $\Phi_{\text{ECL}}$  values increase up to approximately  $5.00 \times 10^{-8}$  M  $[\text{AuNPs}]$  and then  $\Phi_{\text{ECL}}$  values decrease. In order to explain this particular  $\Phi_{\text{ECL}}/[\text{AuNP}]$  trend, redox potentials, quenching and coupling effects together with AFM photograms were measured for gold nanocluster solutions. However, before continuing, an explanation from a qualitative point of view concerning the trends of  $\Phi_{\text{ECL}}$ ,  $I_{\text{ECL}}$  and  $Q$  among one another seems pertinent.

It is clear from Table 1 that the integrated charge of Au@tiopronin nanoparticles increases about 2.7-fold its value when the  $[\text{AuNPs}]$  increase up to about  $5.0 \times 10^{-8}$  M, in relation to that in the absence of NPs. In this same range of concentrations,  $\Phi_{\text{ECL}}$  values increase 1.6-fold, whereas integrated ECL emission shows an enhancement of about 2.1-fold in comparison with its value in the absence of AuNPs. Hence, the increase in ECL efficiency is due to the augmentation of the ECL emission intensity at the  $0.0\text{--}5.00 \times 10^{-8}$  M NPs concentration range.

Subsequently, from  $[\text{AuNPs}] > 5.00 \times 10^{-8}$  M, both  $\Phi_{\text{ECL}}$  and ECL emission intensity follow the same tendencies; they decrease when NP content increases (see Table 1). Notice that the increase in the amount of NPs from  $5.00 \times 10^{-8}$  M does not produce any variation in the  $Q$  values, which remain nearly



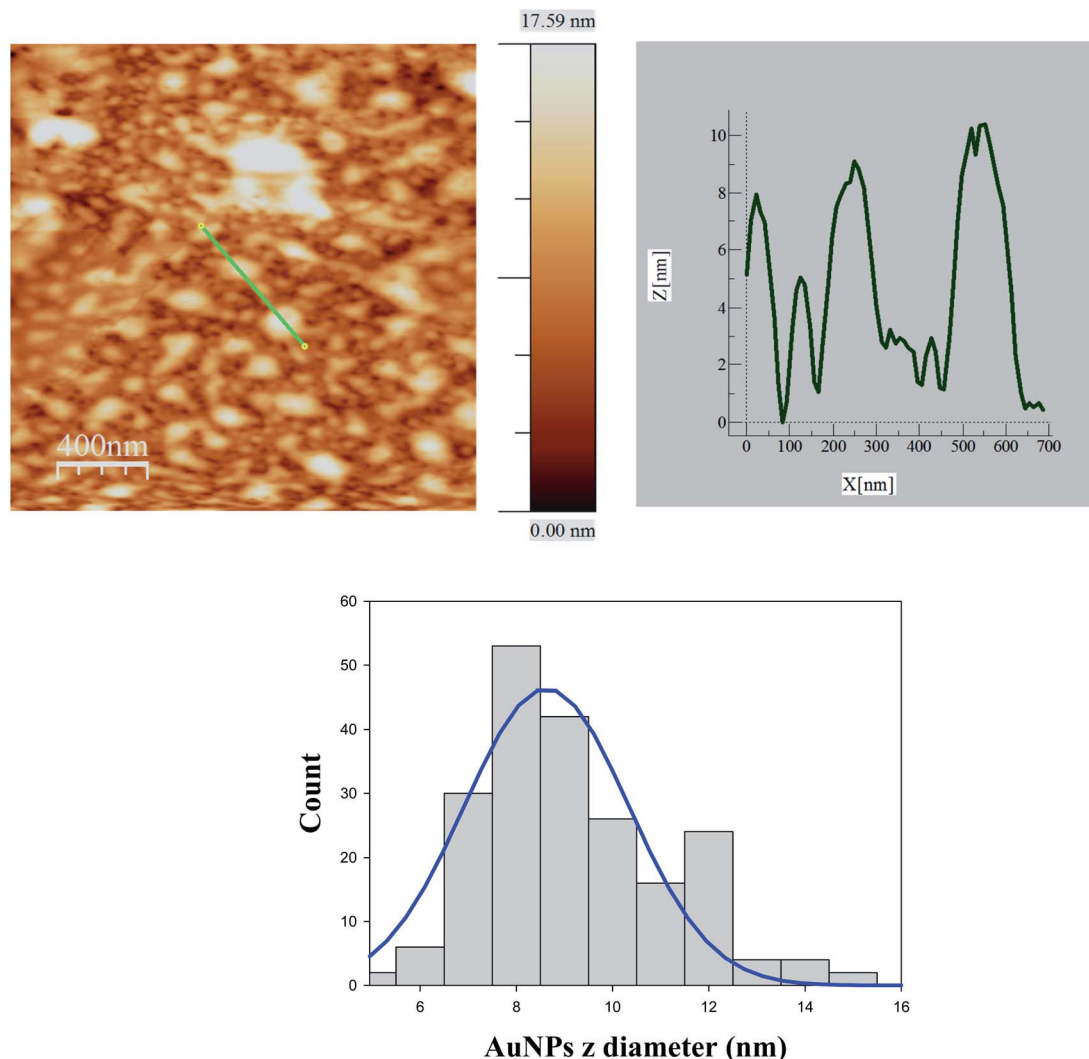


Fig. 5 AFM image for  $8.0 \times 10^{-8}$  M [Au@tiopronin] in the presence of  $[\text{Ru}(\text{bpy})_3]^{3+/2+}$  and corresponding histogram from which the average diameter obtained was  $8.6 \pm 0.9$  nm.

constant within the experimental uncertainty ( $Q = (0.37 \pm 0.06) \times 10^{-3}$  C). This behavior may be a consequence of the formation of self-aggregates when increasing the concentration of NPs. That is, the addition of charged Au@tiopronin species in sufficient quantity would result, on average, in the neutralization of the monomers therein. Therefore, the  $\Phi_{\text{ECL}}[\text{AuNPs}]$  trend is determined to a considerable extent by the  $I_{\text{ECL}}[\text{AuNPs}]$  trend.

In keeping with the preceding discussion,  $\Phi_{\text{ECL}}$  behavior is worthy of consideration, taking into account two ranges of concentrations: one diluted, up to  $\approx 5.00 \times 10^{-8}$  M [AuNPs], and another concentrated from  $\approx$  the previous value of [AuNPs]. These two ranges of NP concentrations will be treated below in this way: diluted and concentrated ranges.

As mentioned above, by means of the redox potential variation as a function of a particular medium content, it is possible to obtain information about the microenvironment that, on average, would have a redox couple around it. In line with this idea, the redox potentials of the  $[\text{Ru}(\text{bpy})_3]^{3+/2+}$  couple were

determined, and the results are summarized in Table 1 (see Section 3.2). These results show that the addition of increasing amounts of nanoparticles does not affect the environment surrounding the couple  $[\text{Ru}(\text{bpy})_3]^{3+/2+}$  (see Section 3.2), which seems to corroborate the previous discussion about the dominance of  $I_{\text{ECL}}$  on the integrated charge.

As aforesaid, the trend of ECL efficiency is governed by that of the ECL emission intensity. In addition, Section 3.3 shows that under our experimental conditions the Au@tiopronin system suffers self-aggregation at the concentrated range of the [AuNPs], whereas at the dilute range of NP concentrations there are no aggregates in either the presence or absence of the Ru(II) complex. Therefore, both light emission and self-aggregation will allow us to rationalize the  $\Phi_{\text{ECL}}[\text{AuNP}]$  trend.

In effect, from a qualitative point of view, it is easy to understand that both the quenching effect and the NP self-aggregation would reduce  $I_{\text{ECL}}$ , and, therefore,  $\Phi_{\text{ECL}}$ , and that these effects would be rendered more efficient by increasing the NPs concentration. Nevertheless, in the diluted range of NPs, an



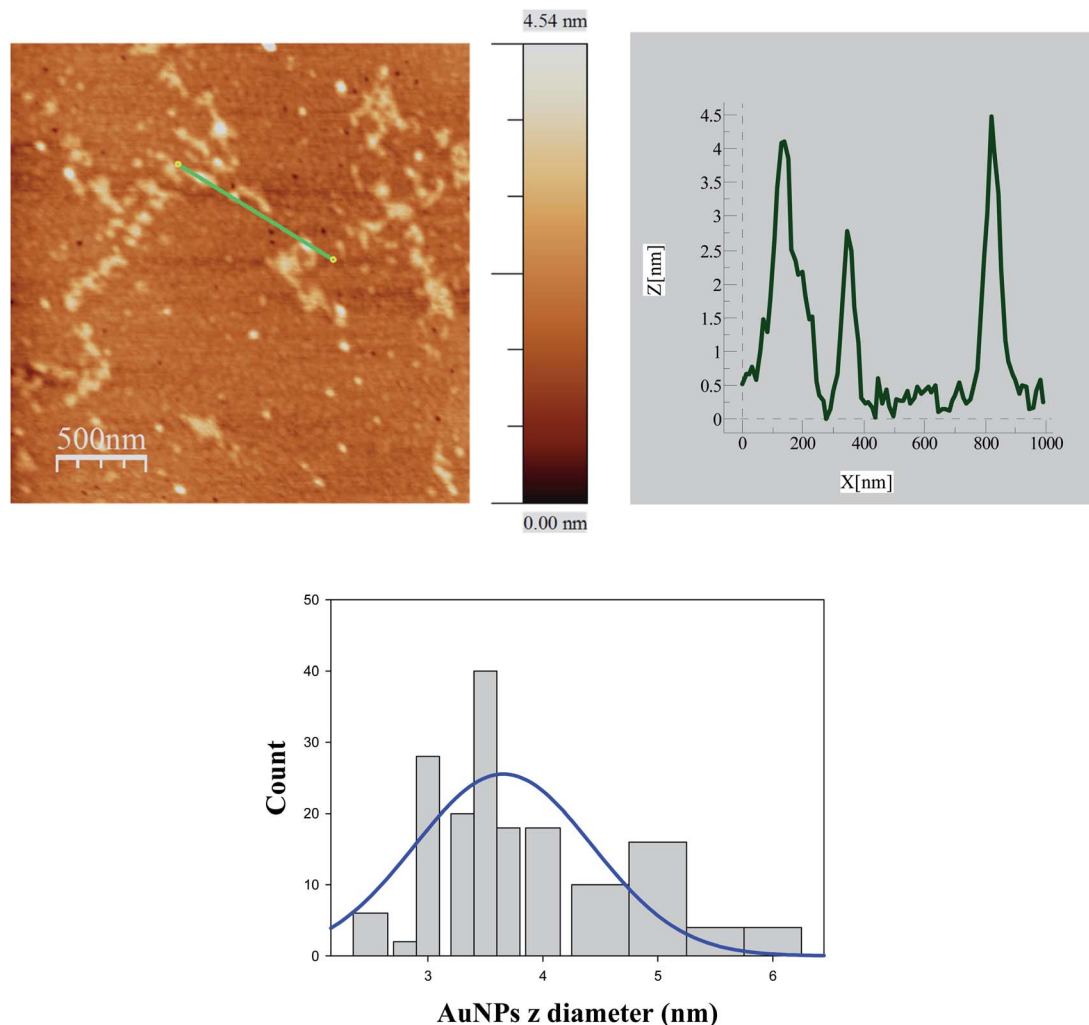


Fig. 6 AFM image for  $1.0 \times 10^{-8}$  M [Au@tiopronin] in the presence of  $[\text{Ru}(\text{bpy})_3]^{2+}$  and corresponding histogram from which the average diameter obtained was  $3.7 \pm 0.6$  nm.

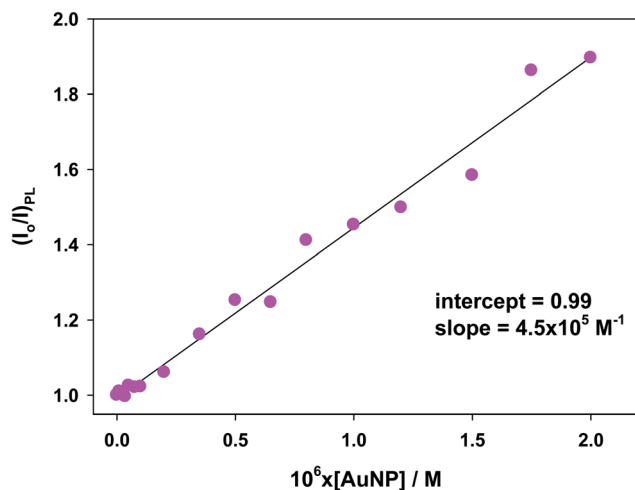
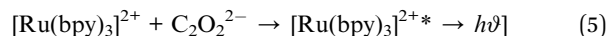


Fig. 7 Stern-Volmer plot for  $[\text{Ru}(\text{bpy})_3]^{2+}$  species in several solutions of gold nanoparticles in cacodylate buffer at pH 7.0.

augmentation of  $I_{\text{ECL}}$  and  $\Phi_{\text{ECL}}$  occurs when the [AuNPs] increases. This behaviour, the ECL-enhancement effect, can be explained by the fact that the photoluminescent coupling effect (see Section 3.5) between the emission bands of the  $[\text{Ru}(\text{bpy})_3]^{2+*}$  and  $\text{AuNP}^*$  species (*vide infra*) is more effective than the quenching effect in this concentration range.

The physical meaning of the ECL-enhancement effect in the  $[\text{Ru}(\text{bpy})_3]^{2+}/\text{Au@tiopronin}$  system will be better understood if several global processes (5)–(9) are taken into account. In simple terms, these give rise to the ECL pathway in the presence of Au@tiopronin nanoparticles. If we consider only that the free metal complex contributes to the ECL reaction (see Fig. S1,† suggesting that the ECL reaction could follow the same mechanism in the absence as well as in the presence of NPs) these processes are:



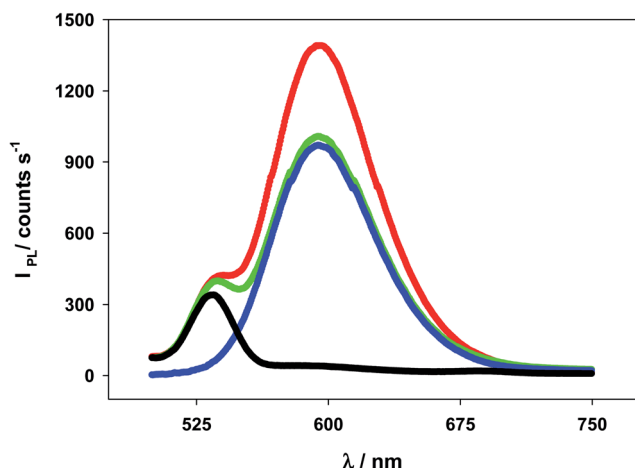
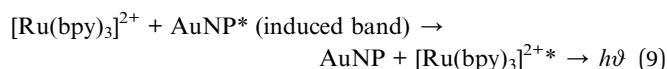
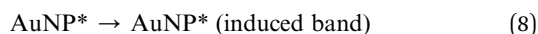


Fig. 8 Photoluminescent spectra showing the coupling effect between the emission spectra of the  $[\text{Ru}(\text{bpy})_3]^{2+*}$  and  $\text{AuNP}^*$  excited species. Red, coupling spectrum. Blue,  $[\text{Ru}(\text{bpy})_3]^{2+*}$  spectrum. Black,  $\text{AuNP}^*$  spectrum. Green, sum of the  $[\text{Ru}(\text{bpy})_3]^{2+*}$  and  $\text{AuNP}^*$  spectra.  $4.0 \times 10^{-7} \text{ M}$   $[\text{Ru}(\text{bpy})_3]^{2+}$ ,  $1.0 \times 10^{-8} \text{ M}$   $[\text{AuNPs}]$ , cacodylate buffer pH 7.0.



where eqn (5) represents the overall ECL reaction, equal to that taking place in the absence of NPs (the mechanism given in ref. 53 is assumed). Eqn (6) and (7) symbolize a non-reactive dynamic quenching process: the first, a non-radiative energy transfer process, involves the sensitization of an  $\text{AuNP}$ ; the second denotes a non-radiative loss of energy from  $\text{AuNP}^*$ . Eqn (8) indicates the generation of an induced band from sensitized  $\text{AuNP}^*$  species. Finally, eqn (9) represents the enhancement of ECL emission: a radiative energy transfer which may increase the emission intensity of the emitting species,  $[\text{Ru}(\text{bpy})_3]^{2+*}$ , because the photons already emitted are recaptured through the  $\text{AuNP}^*$ -induced band.

Certainly, processes (7) and (8) compete with each other; though, the latter process could be favoured when the quenching effect is insignificant, as is the case in dilute solutions (see Section 3.4) and in the absence of the NPs self-aggregation (see Section 3.3); that is the case when the intermolecular interactions are negligible.<sup>5</sup> Furthermore, the resonance requirement for energy transfer from  $\text{Ru}^*$  to NP species must be met, this condition is satisfied in our case as verified *via* the coupling measurements (see Fig. 8). Consequently, the effective coupling between both bands (ECL- $\text{Ru}^*$  emission and induced-NP\*) is an essential requirement for the improvement in ECL to occur. Therefore, the ECL-enhancement in the diluted range of the NPs can be rationalized by the ECL- $\text{Ru}^*$ -induced-NP\* coupling effect or nanoparticle-induced resonance transfer

(NP-RET) effect. In order to quantify the NP-RET effect on the  $[\text{Ru}(\text{bpy})_3]^{2+}/\text{Au@tiopronin}$  system using an ECL reaction, eqn (10) was applied for the diluted range:

$$\frac{\Phi_{\text{ECL}}}{\Phi_{\text{ECL}}^0} = 1 + K_{(\text{NP-RET})_{\text{ECL}}} [\text{AuNP}] \quad (10)$$

where,  $K_{(\text{NP-RET})_{\text{ECL}}}$  denotes the NP-RET effectiveness (defined as the efficacy of a nanoparticle to increase the emission of an ECL reaction by resonance energy transfer), and  $\Phi_{\text{ECL}}$  and  $\Phi_{\text{ECL}}^0$  account for the ECL efficiencies for the actual and standard reactions, respectively (see Section 3.1). Explicitly,  $K_{(\text{NP-RET})_{\text{ECL}}}$  quantifies how much an ECL reaction has been enhanced. Fig. 9 shows a plot for eqn (10): from the slope, a value of  $1.2 \times 10^7 \text{ M}^{-1}$  is found for  $K_{(\text{NP-RET})_{\text{ECL}}}$ , being about 28 times higher than  $K_{\text{Q}}$  for the system  $[\text{Ru}(\text{bpy})_3]^{2+}/\text{Au@tiopronin}$ .

In addition, an estimate of the photoluminescent coupling constant  $K_{(\text{NP-RET})_{\text{PL}}}$  can be made by:

$$\frac{\left(\frac{I}{I_0}\right)_{\text{PL}} - 1}{[\text{AuNP}]} = K_{(\text{NP-RET})_{\text{PL}}} \quad (11)$$

$I$  and  $I_0$  being the intensities at the emission maximum of the photoluminescent spectra for coupled and uncoupled  $\text{Ru}(\text{n})^*$  species, respectively. Through the emission spectra of Fig. 8, the value for  $K_{(\text{NP-RET})_{\text{PL}}}$  was  $4.3 \times 10^7 \text{ M}^{-1}$ , in agreement with the result achieved from ECL measurements.

Therefore, in diluted NP solutions the observed ECL increase is due to an NP-RET effect that surpasses the quenching effect. We must emphasize that the NP-RET effect is only operative in diluted solutions in which the aggregation phenomenon does not occur.

Returning to the ECL-detraction result for the concentrated range of NPs, this is easily understood through the combined action of the quenching and NPs self-aggregation effects. Assuming that only the free metal complex contributes to the ECL reaction as previously mentioned, and bearing in mind eqn

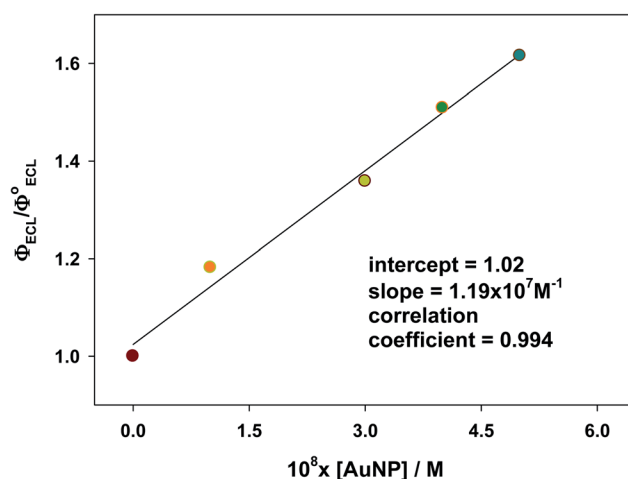


Fig. 9 Relative ECL efficiency plotted vs. AuNPs concentration showing the NP-RET effectiveness of the  $[\text{Ru}(\text{bpy})_3]^{2+}/\text{Au@tiopronin}$  system. Cacodylate buffer, pH 7.0.





(5) and (9) as the only sources of the emission energy, two phenomena can lead to a decrease in the light emission: on one hand, static quenching, which would begin to be significant together with dynamic quenching; and on the other, the self-aggregation that would decrease the number of free NPs. The first would cause a decrease in the available  $[\text{Ru}(\text{bpy})_3]^{2+}$  species for ECL reaction; the second by decreasing the number of free NPs also decreases the probability that reactions (6)–(9) of the previously proposed mechanism can occur. If this last effect were predominant,  $I_{\text{ECL}}$  and  $\Phi_{\text{ECL}}$  should reach a constant value by increasing the concentration of AuNPs, which does not occur. Therefore, in the concentrated range of NPs both  $I_{\text{ECL}}$  and  $\Phi_{\text{ECL}}$  decrease as a consequence of the combined action of the self-aggregation and the static quenching effects, causing a decrease in free NPs and available  $[\text{Ru}(\text{bpy})_3]^{2+}$  species for ECL reaction, respectively.

## 5. Conclusions

ECL efficiencies, redox potentials, PL quenching and coupling effects, and AFM photograms of the  $[\text{Ru}(\text{bpy})_3]^{2+}/\text{Au}@$ tiopronin system were determined in several aqueous solutions of AuNPs. The most remarkable finding was that ECL measurements can display the nanoparticle-induced resonance energy transfer (NP-RET) effect. Its effectiveness was quantified through a coefficient,  $K_{(\text{NP-RET})_{\text{ECL}}}$ , which measures how much an ECL reaction has been enhanced. Moreover, it should also be noted that the NP-RET effect was also verified through PL measurements, in such a way that a coefficient,  $K_{(\text{NP-RET})_{\text{PL}}}$ , was determined, both constants,  $K_{(\text{NP-RET})_{\text{ECL}}}$  and  $K_{(\text{NP-RET})_{\text{PL}}}$  being in close agreement. We must highlight the fact that the NP-RET effect is only displayed in diluted solutions in which there is no NPs self-aggregation. The existence of the NPs self-aggregation behavior is revealed through AFM measurements.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

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