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## Colorectal cancer stem cells: a review of targeted drug delivery by gold nanoparticles

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Colorectal cancer (CRC) is one of the most common cancers with equal probability of affecting both men and women worldwide. Recently, the newly emerged theory of cancer stem cells has associated CRC stem cells (CRCSCs) with the high rates of recurrence and poor prognosis. Thus, targeting CRCSCs instead of CRC may resolve cancer relapse. Among the chemotherapeutic drugs, the antimetabolite 5-fluorouracil (5-FU) has shown high efficiency in CRC treatment. However, due to several limitations, the usefulness of 5-FU has been restricted. The application of gold nanoparticles (AuNPs) in drug delivery systems is reported to enhance the effectiveness of anticancer drugs due to their biostability, non-toxicity and feasibility for surface modification. Furthermore, the overexpression of biomolecular surface markers in CRCSCs may elevate the specific targeting by AuNPs, and hence, reduce the non-specific binding, which could lead to systemic side effects. This review briefly presents the proposed therapeutic potential of AuNPs loaded with 5-FU, conjugated with specific antibodies targeting CRCSCs, which could be valuable to improve some limitations in current CRC management.

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## 1. Introduction

Colorectal cancer (CRC) is among the most common cancers and accounts for nearly 9% of all cancers in the world<sup>1,2</sup> with high mortality rate due to advanced stage diagnosis and high rates of recurrence.<sup>3</sup> Studies have underlined the cause of recurrence and metastases as due to the presence of cancer stem cells (CSCs), a small subpopulation of cancer cells which are able to self-renew, differentiate and sustain tumor growth.<sup>2,4,5</sup> Colorectal cancer stem cells (CRCSCs) are characterized by the overexpression of CD133, a five-transmembrane glycoprotein.<sup>6,7</sup>

Currently, common cytotoxic chemotherapies used for the treatment of CRC are 5-fluorouracil (5-FU), oxaliplatin and cisplatin. Chemotherapeutic drugs kill cancer cells by interfering with their cell growth and division mechanisms.<sup>8</sup> A few studies both on cellular and animal models have proved that treatment with these chemotherapeutic drugs, either alone or

in combination with other chemotherapies, is effective in treating CRC.<sup>9–11</sup> Among these chemotherapeutic agents, 5-FU is one of the most prescribed anti-tumor drugs for CRC therapy.<sup>12,13</sup> Owing to its analogous structure to uracil, 5-FU can be incorporated into RNA and DNA and interferes with nucleoside metabolism. However, 5-FU-based chemotherapy also interferes with rapidly dividing healthy cells due to its lack of site specificity, and hence, causes common side effects such as hair loss, nausea and vomiting.<sup>14,15</sup> In addition, rapid clearance from blood circulation and poor distribution limit the therapeutic action of 5-FU.<sup>16</sup>

Drug delivery systems in cancer therapeutics provide an alternative to minimize the limitations of conventional cancer chemotherapy by improving specific drug targeting, prolonging the circulation time and controlling drug release.<sup>17,18</sup> In addition, targeted drug delivery increases bioavailability to maintain the drug concentration upon arriving at the cancer site.<sup>19</sup> Within the past few decades, nanoparticles have received great interest



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Table 1 The advantages of the proposed 5-FU loaded AuNPs conjugated with CD133 antibody system targeting CRCSCs

System	Advantages	Reference
AuNPs as drug carrier	Facile synthesis Biocompatible Multi-functionalization Tunable surface	27
mPEG-surface coating	Reduce protein adsorption on AuNPs Avoid macrophage uptake	28 and 29
5-FU	Antimetabolite Inhibit DNA synthesis	30
Anti-CD133 monoclonal antibody conjugate	Overexpression of CD133 has been associated with the relapse, metastasis and chemotherapy resistant Interfere on specific proteins overexpressed CD133 involved in tumorigenesis (targeted cancer therapy)	7 and 31
CRCSCs targeting	Targeting CRCSCs rather than tumor bulk could be effective in reducing risk of relapse and metastasis	32

among the CRC research community as one of the effective drug delivery systems. Some nanoparticles targeting CRC have been studied previously, including solid lipid nanoparticles,<sup>20,21</sup> heparin-polyethyleneimine nanoparticles,<sup>22</sup> silica nanoparticles<sup>23</sup> and liposome-based nanoparticles.<sup>24,25</sup> However, according to a literature survey (2005–2015) by Wilhelm *et al.*, only 0.7% of the administered nanoparticle dose reaches the solid tumor, which

eventually hinders the effective use of nanoparticles as drug delivery agents.<sup>26</sup>

Here, we summarize a proposed system of targeted drug delivery consisting of 5-FU loaded gold nanoparticles (AuNPs) conjugated with CD133 antibody to target CRCSCs, supported by previous studies (Table 1).

Previously, the specific targeting of the CSCs in CRC by CD133 antibody has been studied using methoxy poly(ethylene

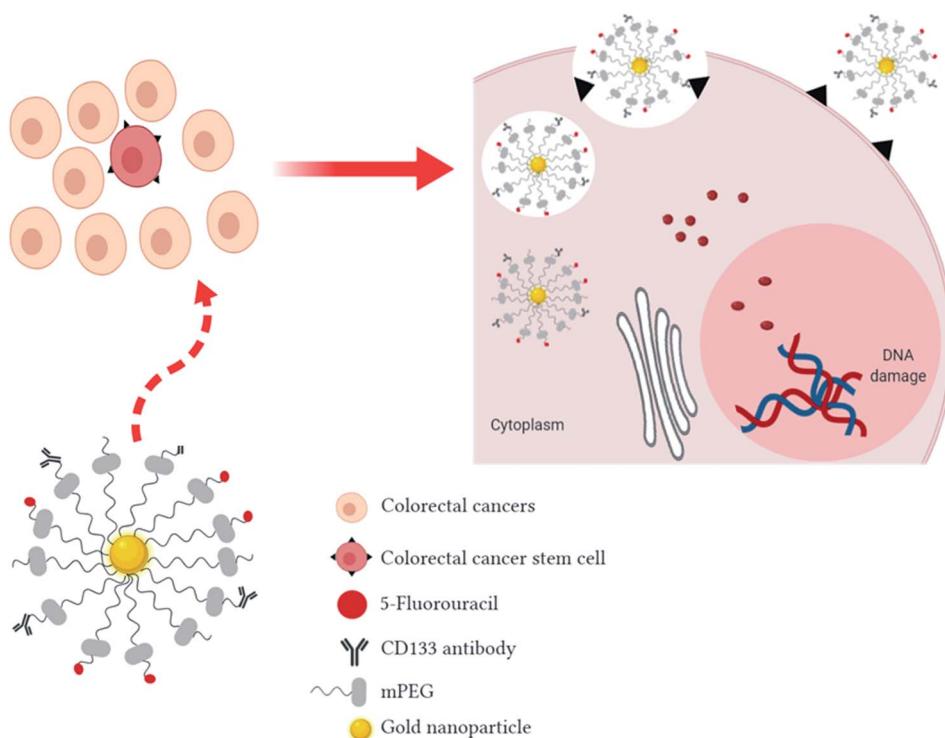


Fig. 1 The proposed schematic mechanism of 5-fluorouracil (5-FU) loaded gold nanoparticles (AuNPs) conjugated with CD133 antibodies to target colorectal cancer stem cells (CRCSCs) in a targeted drug delivery system. Methoxy polyethylene glycol (mPEG)-stabilized AuNPs loaded with 5-FU and conjugated with CD133 antibody will target the CRCSCs instead of the bulk of colorectal cancers due to the overexpression of CD133 antigen on the surface of CRCSCs. High affinity binding of the CD133 antibody ligand with the targeted cells will increase the delivery efficiency and hence protect healthy cells. The AuNPs with loaded 5-FU will be internalized by cells via endocytosis. The acidic environment in the tumor may trigger the cleave of 5-FU from the AuNP complex inside cell endosomes to enhance cell toxicity by interfering with DNA synthesis.



glycol)-poly( $\epsilon$ -caprolactone) (mPEG-PCL) nanoparticles loaded with anticancer drug 7-ethyl-10-hydroxy-camptothecin (SN-38).<sup>33</sup> The *in vitro* and *in vivo* studies showed that CD133-targeting nanoparticles have increased cytotoxicity and inhibited tumor growth in HCT116 cells and mouse xenograft models, respectively, compared with the none antibody-targeted nanoparticles. In this case, the therapeutic effects on CRC have been enhanced by the CD133 antibody conjugates because the chemotherapeutic agent SN-38 is efficiently guided to the overexpressed CD133 markers at the tumor site.

Fig. 1 shows the proposed schematic mechanism of 5-FU loaded AuNPs conjugated with CD133 antibody to target CRCSCs. In drug delivery systems, nonionic polymer-surface modification of AuNPs by methoxy polyethylene glycol (mPEG) thiol is commonly used to improve blood residence time, reduce reticulo-endothelial system (RES) uptake and avoid non-specific targeting.<sup>28</sup> The AuNP complexes will be internalized by CRCSCs through passive (neovasculature) and active targeting (anti-CD133 ligand-receptor docking) followed by fusion with lysosomes. Owing to the changes of some pathological events in tumor cells, such as pH and concentration of intracellular glutathione (GSH), AuNPs can be engineered to be activated by these endogenous stimuli in order to release the payloads at the tumor site and hence enhance the therapeutic effects and avoid systemic side effects to healthy cells. Regularly, the extracellular pH of solid tumor cells is acidic (5.5 to 6.5) compared to the physiological condition<sup>34</sup> due to the anaerobic glycolysis-

produced lactic acid in hypoxia<sup>35</sup> and substantial H<sup>+</sup> generation because of higher metabolic activity.<sup>36</sup> Besides the acidic tumor environment, 5-FU will be released on exposure to lysosomal enzymes followed by interfering with DNA synthesis and RNA processing and functioning to cause tumor cell death.

## 2. Colorectal cancer stem cells

According to the cancer stem cell (CSC) theory, a small subpopulation of cells share embryonic stem cell characteristics,<sup>2</sup> giving rise to metastatic cancer cells and responsible for the high rate of recurrence.<sup>37</sup> Indeed, CSCs share the same major signaling pathways with embryonic stem cells, including the Wnt, Notch, transforming growth factor beta (TGF- $\beta$ ), and Hedgehog signaling pathways.<sup>38</sup> Nevertheless, quiescent CSCs express stemness genes, have the capability to self-renew and differentiate into other non-stem cancer cells and resist traditional chemotherapy and radiotherapy.<sup>39</sup> Another crucial feature of CSCs is possessing tumorigenic potential, which is measured by the ability of the cells to initiate xenograft tumors upon transplantation.<sup>38</sup>

Thus, targeting CSCs instead of the tumor bulk population is a therapeutic goal in CRC management, particularly in preventing cancer relapse. The isolation and identification of CSCs from CRC further elucidates several putative stem cell markers. The identification of CRCSC markers will be beneficial in identifying the disease progression as well as the risk of recurrence among patients.<sup>40</sup> Table 2 shows several putative stem cell

Table 2 Biomarkers of colorectal cancer stem cells with their prognostic values

CRCSC Markers	Molecular weight	Prognostic significance	Tumorigenicity	References
CD133	120 kDa	<ul style="list-style-type: none"> <li>Highly correlated with low survival in CRC patients</li> <li>Higher expression is associated with resistance of CRC to 5-FU based chemotherapy</li> </ul>	<ul style="list-style-type: none"> <li>Subcutaneous injection of a CD133<sup>+</sup>, but not CD133<sup>-</sup> subpopulation of CRC into immunodeficient mice established a tumor</li> </ul>	41–45
CD44	81 kDa	<ul style="list-style-type: none"> <li>Associated with lower patient survival</li> <li>Not associated with prognosis</li> </ul>	<ul style="list-style-type: none"> <li>CD44<sup>+</sup> cells from a patient's tumor initiated a xenograft tumor <i>in vivo</i> and form a sphere <i>in vitro</i></li> </ul>	43, 46 and 47
CD166	100–105 kDa	<ul style="list-style-type: none"> <li>The expression correlated with a shortened period of survival</li> <li>No significant correlation with poor survival</li> </ul>	<ul style="list-style-type: none"> <li>CD166<sup>+</sup>/CD44<sup>+</sup> or CD166<sup>+</sup>/ESA<sup>+</sup> from human CRC cells can induce tumorigenesis in immunodeficient mice</li> </ul>	43, 48 and 49
CD24	35–45 kDa	<ul style="list-style-type: none"> <li>No significant correlation with patient low survival</li> </ul>	<ul style="list-style-type: none"> <li>Tumor spheroid cells capable of inducing tumors upon xenotransplantation</li> </ul>	4 and 50
CD26	110 kDa	<ul style="list-style-type: none"> <li>Associated with enhanced invasiveness and chemoresistance</li> </ul>	<ul style="list-style-type: none"> <li>Isolated CD26<sup>+</sup>, but not CD26<sup>-</sup> cells of CRC, developed distant metastasis when injected into the mouse cecal wall</li> </ul>	51
Epithelial cell adhesion molecule (EpCAM)	30–40 kDa	<ul style="list-style-type: none"> <li>Inversely correlated between EpCAM glycoprotein expression and tumor staging</li> </ul>	<ul style="list-style-type: none"> <li>Injection of 200 to 500 EpCAM<sup>high</sup>/CD44<sup>+</sup> cells in immunodeficient mice was sufficient to give rise to a tumor</li> </ul>	4 and 52



biomarkers of CRC and their prognostic value as previously reported.

It is interesting to note that several discrepancies of prognosis values and tumorigenic potential have been reported among the listed CRCSC biomarkers. Despite the above listed inconsistencies, there is strong evidence from numerous studies suggesting that CD133 is a notable marker to identify CRCSCs.<sup>6,7,53</sup> CD133, also known as prominin-1, is a 97 kDa pentaspan transmembrane glycoprotein<sup>54,55</sup> and has been reported to be mostly localized in membrane protrusions.<sup>56</sup> However, due to glycosylation, CD133 yields a 120 kDa protein.<sup>55</sup> It was first discovered in 1997 as a hematopoietic stem cell (HSC) marker, which is associated with stem cell maintenance.<sup>55</sup>

Recently, CD133 has been widely studied as a putative biological marker for CRCSCs to predict tumor progression, prognosis value and chemoradiotherapy resistance.<sup>7</sup> The enhanced CD133 expression in CRCSCs suggests that CD133 is a suitable biomarker target for the treatment of CRC. Fang *et al.* demonstrated that CD133<sup>+</sup> cells isolated from primary CRC gave rise to spheroid cultures, which have the ability to self-renew and maintain CD133 expression in serum-free media.<sup>57</sup> This finding supports the notion that CRCSCs that are CD133<sup>+</sup> are able to initiate tumors, which may increase CRC occurrence.

It was also suggested that CRCSCs escaped conventional chemotherapy targeting rapidly dividing cells due to their quiescent nature, thereby inhibiting the complete eradication of CRC.<sup>58</sup> In addition, the presence of overexpressed multidrug resistance protein 1 (MDR-1), detoxifying enzymes and DNA repair proteins in CSCs increased chemoresistance in malignant tumors.<sup>59,60</sup> A study by Ong *et al.* reported that CD133<sup>+</sup> CRC cells were more resistant towards 5-FU-based chemotherapy.<sup>44</sup> Therefore, CD133-targeted therapeutic strategies in CRC might be valuable to eliminate CSC, which is associated with a high risk of relapse.

### 3. 5-Fluorouracil

5-FU is an antimetabolite and was discovered in 1957 to display tumor inhibitory activities by Heiderberger *et al.*<sup>61</sup> Currently, it is used as a first line chemotherapy agent and is widely used as an antineoplastic drug.<sup>12,62</sup> 5-FU is a natural pyrimidine uracil analog with a fluorine atom inserted into the C-5 position in place of hydrogen (Fig. 2).<sup>63</sup> Besides CRC, 5-FU is also commonly used to treat other solid malignancies arising from breast, stomach, pancreatic and head and neck cancers.<sup>64,65</sup>

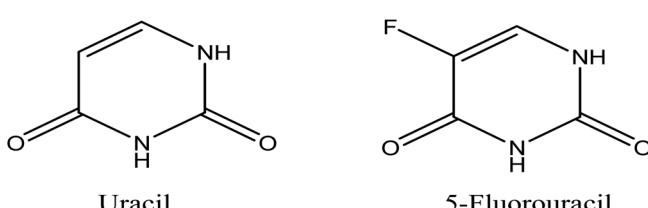


Fig. 2 Chemical structure of uracil and 5-fluorouracil.

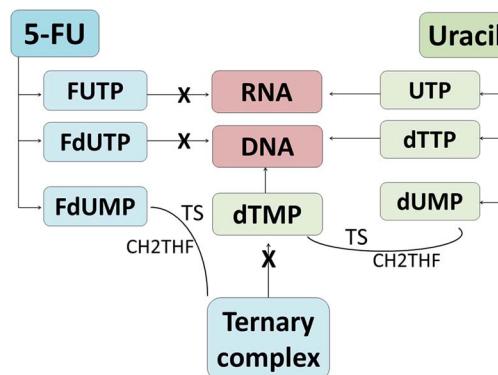


Fig. 3 Metabolism routes of 5-fluorouracil and uracil. 5-FU metabolites inhibit DNA synthesis and RNA processing and function, which eventually results in cell death.

The regulation of 5-FU metabolism involves thymidylate synthase (TS), dihydropyrimidine dehydrogenase (DPD) and orotate phosphoribosyl transferase (OPRT) enzymes.<sup>66</sup> It has been reported that only 1–3% of the original dose of 5-FU contributes to the cytotoxicity in rapidly proliferating tumor and normal cells *via* anabolic actions.<sup>67</sup> Meanwhile, about 80–85% of 5-FU is subjected to biotransformation in the liver and catabolized into inactive metabolites, fluorinated  $\beta$ -alanine, by DPD<sup>67,68</sup> and 15–20% is eliminated in urine.<sup>68</sup> Thus, the development of drug delivery agents for 5-FU will enhance the 5-FU cytotoxicity to the targeted tumor cells. As an analog of uracil, 5-FU readily crosses the cell membrane using the same facilitated transport mechanism as uracil and is incorporated into nucleic acids, which eventually contributes to the cytotoxicity.<sup>30,69</sup> 5-FU and uracil are in continuous competition with each other due to their identical metabolic pathways (Fig. 3). Instead of uridine triphosphate (UTP), the active metabolite of 5-FU, fluorouridine triphosphate (FUTP), is incorporated into ribonucleic acid (RNA) and interferes with its processing and functioning.<sup>67</sup> Fluorodeoxyuridine triphosphate (FdUTP) is another 5-FU metabolite that can be incorporated into deoxyribonucleic acid (DNA) instead of deoxythymidine triphosphate (dTTP). In addition, instead of normal pyrimidines, deoxyuridine monophosphate (dUMP), the metabolite of 5-FU, fluorodeoxyuridine monophosphate (FdUMP), forms a covalent ternary complex with TS and 5,10-methylene tetrahydrofolate (CH<sub>2</sub>THF) and inhibits deoxythymidine monophosphate (dTDP) production, which eventually interferes with DNA synthesis.<sup>62,67</sup> In general, 5-FU causes cancer cell death by interfering with DNA synthesis and RNA processing and function.

Despite the effectiveness of 5-FU as a chemotherapeutic drug, one of its limitations is short plasma half-life (10–20 min) caused by fast degradation.<sup>63</sup> In addition, due to the non-selectivity and poor bioavailability of 5-FU to tumor sites, a higher dose is required in order to reach the effective drug concentration. Nonetheless, high doses of chemotherapeutic agents lead to elevated systemic side effects and increased incidence of drug resistance, thereby limiting the usefulness of 5-FU in CRC chemo-treatment.

## 4. Considerations on using nanoparticles in drug delivery

The application of nanoparticles in drug delivery systems has enhanced the conventional anticancer treatments, particularly in terms of bioavailability, specific targeting, and reducing adverse side effects. However, nanoparticle clearance from the circulation by the reticuloendothelial system (RES), including the mononuclear phagocytic system (MNS), is one of the major concerns in drug delivery systems. The MNS consists of the liver, spleen, lymph nodes, bone marrow, skin and macrophages.<sup>26</sup> Meanwhile, scavenger receptors on Kupffer cells are responsible for recognizing, engulfing and eliminating the opsonized nanoparticles in the liver.<sup>26,70</sup> Thus, several requirements should be incorporated in designing nanoparticles to evade rapid clearance and enhance effective delivery of chemotherapeutic agents. Generally, the mechanisms of nanoparticles in drug delivery are affected by the physicochemical properties of the nanoparticles, such as size, surface chemistry and particle shape. These characterizations may impact nanoparticle-cell interactions, particularly in cellular and tissue uptake.

First and foremost, it is apparent that nanoparticles offer an ideal system for chemotherapeutic drug delivery due to their tunable sizes. In medical applications, nanoparticles are materials with preferential dimensions of 1–100 nm.<sup>71</sup> In addition, to avoid recognition by the RES, nano-sized particles also contribute to passive targeting into the tumor zone. The idea of nanoparticles as promising carriers for anti-tumor drugs was owing to the study by Matsumura and Maeda, which suggested that nanoparticles could accumulate in tumors.<sup>72</sup> The formation of neovasculature in tumors increases the permeability of the vascular endothelial layer, resulting in leaks and susceptibility for the passage of molecules and/or nanosystems with sizes in the range of hundreds of nm.<sup>73,74</sup> It is reported that the range of size of the vascular pore cut-off exhibited by most solid tumors is between 380 nm and 780 nm.<sup>75</sup> Thus, nano-sized particles within this size range can easily pass through the cancer vascular pores. This phenomenon is known as the enhanced permeability and retention (EPR) effect (Fig. 4); it

enables passive targeting of nanoparticles carrying drugs and is reported to enhance drug efficiency by up to 2.3-fold compared with when loaded with the free drugs.<sup>76</sup>

Secondly, hydrophobic and highly surface charged nanoparticles are highly recognized by the RES and removed from the circulation.<sup>77</sup> Thus, various current nanoparticle-based drug delivery systems undergo surface modifications by grafting polymer coatings such as polyethylene glycol (PEG), poly(*N*-vinyl-2-pyrrolidone) (PVP) and dextran in order to avoid agglomeration and systemic clearance by macrophages, thereby increasing the bioavailability of chemotherapeutic agents.<sup>78–80</sup> Hydrophilic PEG shielded the presence of nanoparticles to avoid opsonization by passive diffusion across the lipophilic cell membrane and hence increase the nanoparticle availability in systemic circulation.<sup>79</sup> For instance, doxorubicin-loaded PEGylated liposomes have shown significant efficacy in breast cancer treatment.<sup>81</sup> In addition, the stability of the formulated nanoparticles in the delivery system influences the cellular uptake. Monodispersity of nanoparticles in suspension is required to avoid agglomeration, which increases the rate of clearance. Basically, more positively or negatively charged nanoparticles prevent agglomeration in suspension due to their larger degree of repulsion. However, since many plasma membrane surfaces are negatively charged, nanoparticles with cationic surfactants display high affinity for cellular uptake *via* endocytosis during nanoparticle-cell adhesion.<sup>82</sup>

Lastly, nanoparticle shape is another crucial factor that can affect the cellular uptake efficiency and actions of nanoparticle delivery. In a study conducted by Huang *et al.*, mesoporous silica nanoparticles (MSNs) of long-rod shape showed an increased number of internalized particles compared with short-rod shaped and sphere-shaped MSNs.<sup>83</sup> However, when compared to spherical shape MSNs, the long-rod shaped MSNs showed higher cytotoxicity than the short-rod shaped and sphere-shaped particles. Another study found that the sharpness of the nanoparticles influenced the intracellular translocation and excretion of nanoparticles.<sup>84</sup> In that study, it was reported that the sharp-shaped nanodiamonds translocated from the endosome to the cytoplasm quickly, with difficult cellular excretion compared with the round-shaped nanodiamonds, suggesting that the sharp-shaped nanoparticles ruptured the endosomal membrane, thereby reduced the

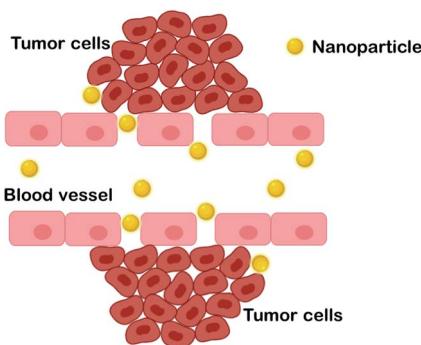


Fig. 4 The enhanced permeability and retention (EPR) effect, which allows passive targeting of nanoparticles. The neovasculature phenomenon in the tumor causes disordered vascular endothelial layers and permits the passage of nanoparticles.

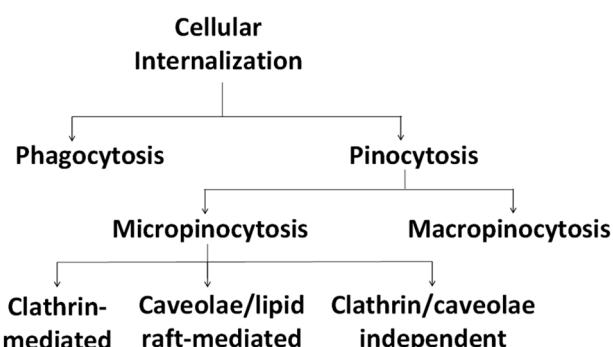


Fig. 5 Different pathways for cellular internalization of nanoparticles.



Table 3 Summary of the physicochemical properties of engineered nanoparticles and their cellular internalization pathways

Physicochemical properties		Cellular internalization
Size	60–80 nm	Caveolae-mediated endocytosis which could avoid lysosomes
	~100 nm	Clathrin-mediated endocytosis
	Up to 5 $\mu$ m	Engulfed by macropinocytosis
	Larger particle	Opsonized and engulfed by phagocytosis
Surface charge	Cationic	Have strong electrostatic interaction with the anionic cell membrane surface leading to rapid entry
	Neutral	May interact with the cells with the aid of hydrophobic and hydrogen bond interactions
	Anionic	May interact with the cationic protein membrane and create repulsive interactions with the negatively charged cell surface leading to endocytosis
Surface hydrophobicity	Hydrophobic	High affinity for the cell membrane
	Hydrophilic	Suppresses interaction between the nanoparticles and lipid bilayer of cells

excretion rate, and remained in the cytoplasm for an extended time. These studies suggest the interplay between cell interactions and nanoparticle design as one of the important aspects in developing drug delivery systems.

In addition, it is important to note that in cellular systems, particles enter the cells *via* the endocytosis route, which consists of phagocytosis or pinocytosis as summarized in Fig. 5. Large particles are taken up by phagocytosis/micropinocytosis, while nanoparticles with sizes lower than 200 nm are engulfed by micropinocytosis, either by a clathrin-dependent, caveolae-dependent or clathrin/caveolae-independent route.<sup>78,85</sup>

The clathrin-mediated pathway transfers the nanoparticles into lysosomes and a portion of the nanoparticles are recycled back to the extracellular space. Meanwhile, nanoparticles that follow the caveolae pathway are transported to caveosomes to be either translocated to the endoplasmic reticulum/Golgi body or enter the endosomal pathway.<sup>86</sup> Engineered physicochemical properties as well as elemental compositions of the nanoparticles are crucial, since they are responsible for the fate of the nanoparticles generally in blood circulation and hence determine the effectiveness of the carried chemotherapeutic agents.<sup>87</sup> Table 3 summarizes the common physicochemical properties of nanoparticles and the pathways for their internalization as described by Kou *et al.*<sup>78</sup>

Apart from the physicochemical properties of the particles, it is also important to note that the initial materials for nanoparticles should be non-toxic materials or biocompatible to avoid toxic effects.<sup>88</sup>

## 5. The applications of gold nanoparticles in drug delivery systems

Gold (Au) has been applied in the biomedical field for many years. It has been used in bioimaging<sup>89</sup> as well as in the treatment of arthritis/inflammation since the beginning of the 20<sup>th</sup> century.<sup>90</sup> Since gold salts show anti-inflammatory activities, they have been incorporated into disease-modifying anti-rheumatic drugs (DMARDs), one of the drug classes to treat rheumatic arthritis.<sup>91</sup> However, gold salts as DMARDs have been discontinued and replaced by others, probably due to the affinity of gold for DNA and hence its interference in cell

function.<sup>80</sup> The toxic effects of AuNPs have been supported by a study conducted by Qiu *et al.* which reported that acute and chronic exposure to AuNPs interrupts gene expression.<sup>92</sup> To date, no gold-based nanomedicines have been approved by the US Food and Drug Administration (FDA) yet.<sup>80</sup>

Besides as a direct therapeutic agent, AuNPs have been intensively studied in other biomedical applications, including in plasmonic photothermal therapy, photodynamic therapy and targeted drug delivery.<sup>93</sup> To date, various studies have proven the overwhelming potential of AuNPs with some in clinical trial phases. A recent phase II clinical study into the photothermal effect (AuroLase Therapy with identification number: NCT02680535) utilized the unique optical tunability of AuNPs that can convert light into heat to thermally kill prostate cancer.<sup>94</sup> A near-infrared laser that did not destroy healthy tissues was specifically designed to excite the accumulated AuNPs in cancerous cells and hence, the removal of the prostate tumor was precise enough without significant damage to surrounding healthy tissues.

One of the advantages of AuNPs compared to other metal nanoparticles is that the gold core is relatively inert, and so it is considered to be biocompatible and non-toxic.<sup>95</sup> Previously, it has been reported that ionic AuNPs show obvious toxicity at 25 mM.<sup>96</sup> Nevertheless, another study found that plain AuNPs did not show any cytotoxicity to human cells up to 250 mM.<sup>97</sup> Thus, it is suggested that AuNPs themselves are non-toxic, but then toxicity could be derived from their functionalization, such as with surface coatings. In spite of this, AuNPs have shown promising potential to deliver chemotherapeutic drugs targeting cancer cells either *via* passive targeting, active targeting or both. As previously described, passive targeting is due to the neovasculature formed by tumor cells. Meanwhile, the active targeting approach is *via* the surface functionalization of AuNPs using immobilized ligands such as proteins, antibodies or small molecules for specific cell targeting on the targeted surface of the membrane.<sup>80</sup> Thus, targeted cancer therapies can be beneficial for the controlled delivery of chemotherapeutic agents stemming from the fact that cancer cells overexpress specific antigens.<sup>98</sup> Owing to the expression of specific antigens, drug carriers are often conjugated with specific antibodies to bind with the particularly expressed antigen, which will be discussed later.



Table 4 The current applications of gold nanoparticle-loaded anticancer drugs in drug delivery systems

Drugs loaded	Surface coatings/ligands	Treatments	Key findings	Reference
5-FU	Thiols thioglycolic acid (TGA) and glutathione (GSH)	Colon cancer tissue samples were obtained from patients and incubated with TGA/GSH-AuNPs-5-FU or 5-FU alone	TGA/GSH-AuNPs-5-FU exhibited 2-fold higher anticancer effect compared with free 5-FU	100
Doxorubicin (DOX)	PEG	Carcinoma cells were intraperitoneally injected into female Balb/c mice and treated with DOX	PEG coatings showed high drug accumulation compared to passive targeting without PEG	101
TGF- $\beta$ 1 antibody and methotrexate	Folic acid	MDA-MB-231 breast cancer cell line	Folate conjugation enhanced the cellular uptake and TGF- $\beta$ 1 antibody on the AuNPs reduced extracellular TGF- $\beta$ 1 of cancer cells by 30%	102
Mitoxantrone (MTX)	Methoxy-polyethylene-glycol (mPEG)	Combination of photodynamic therapy (PDT) and chemotherapy on human melanoma (DFW) and breast cancer (MCF7) cell lines	MTX-mPEG-AuNP complex improved the efficacy of PDT with a light emitting diode	103
Doxorubicin (DOX)	Oligonucleotides (ONT)	DOX-ONT-AuNP complex treated SW480 CRC cell line and xenograft mice which were subcutaneously inoculated by the transfected SW480 cells	DOX-ONT-AuNP complex reduced cell viability and inhibited tumor growth in CRC xenograft mice	104

The incorporation of a biocompatible coating seems to enhance the usefulness of AuNPs in drug delivery systems. A recent clinical trial on an AuNP-based product, Aurimune (CYT-6091), recombinant human tumor necrosis factor (rhTNF) bound to colloidal gold *via* a PEG-linker, showed that AuNP-loaded rhTNF was three times as effective in treating advanced stage cancer patients compared to the native rhTNF without introducing any significant toxic effects.<sup>99</sup> This finding supports the notion that AuNPs with some modifications could be a promising targeted drug carrier in mediating conventional chemotherapeutic agents to eradicate tumors.

Undoubtedly, in order to justify further development, *in vitro* studies using human cell line models are very valuable for initial screening prior to clinical validation and translation. However, preclinical *in vitro* studies are beneficial to study specific tumor biology and treatment responses only. Thus, the *in vitro* effects may not fully represent the effects in medical application, since more complex interactions exist among the diverse organs and physiological systems in the human body. In spite of this, there are some AuNP studies that have reached clinical trials as previously discussed, and this could be a promising area for researchers to further explore their applications, particularly in drug delivery systems for cancer management.

Table 4 shows the application of functionalized-AuNPs in drug delivery systems targeting various types of cancers that have been reported *in vivo* and *in vitro*.

## 6. Design of gold nanoparticles for cellular uptake

As previously discussed, physicochemical properties in terms of size, shape and surface properties play a major role in determining the competency of AuNPs as an optimal drug carrier, particularly in order to overcome certain biological barriers, such

as macrophage clearance. For drug delivery purposes, the interaction between membrane receptors and nanoparticles is one of the most important aspects that regulates the rate of cellular uptake (endocytosis) and hence increases the accumulation of drug-loaded nanoparticles at the tumor site. In this regard, the nanoparticle size plays an important role to avoid early clearance by the MPS organs. Several previous studies have reported the rate of cellular uptake and AuNP accumulation among different sizes of AuNPs (Table 5). In AuNP chemical synthesis using the Turkevish method, the size of the synthesized AuNPs can be easily adjusted by varying the ratio of gold salt and citrate.<sup>105</sup> Thus, tunability of the AuNP size during chemical synthesis can maximize the efficient delivery of therapeutic drugs to targeted cells.

In biomedical applications, it is important to take note that the size of AuNPs should not be too small to avoid the interactions with biological macromolecules leading to cytotoxicity, but not too large to reduce the chance of macrophage clearance. Based on previous studies, it was highly suggested that the size of endocytosis-susceptible guest particles should be around 40–50 nm.<sup>106,112,114</sup> This is in agreement with the 100 nm size limit of the plasma membrane to form endocytic vesicles.<sup>119</sup> In addition, AuNPs with a size around 40 to 60 nm were reported to have optimal values of membrane bending rigidity and ligand-receptor binding interaction for endocytosis.<sup>114</sup> However, it is another important consideration to note that the hydrodynamic diameter of AuNPs will increase upon coating or surface modification and hence affect the rate of cellular uptake. Thus, it is crucial to consistently monitor the increase of AuNP size before and after modifications to avoid clearance due to larger size.

In addition to size, the shape of AuNPs is another crucial physical factor to affect endocytic uptake by cells. The particle shape can enhance the process of cellular membrane wrapping during endocytosis.<sup>120</sup> It was reported that spherical AuNPs increased the cellular uptake when compared to rod-shaped



Table 5 Size of gold nanoparticles (AuNPs) and effects on cellular uptake

Size of AuNPs (nm)	Cell lines or animals	Findings	References
14, 30, 50, 74, 100	HeLa cells	50 nm AuNPs showed the maximum uptake by HeLa cells, which was 1294 nanoparticles per hour, and the maximum time for cellular uptake was 6 hours	106
3, 10, 25, 50	Hep-2 cells	A toxicity study of plain AuNPs on Hep-2 cells revealed that smaller AuNPs could enter the nucleus thereby damaging the cellular and nuclear membranes and it was suggested that the cytotoxicity of AuNPs is highly size-dependent (3 > 10 > 25 > 50 nm)	107
1.4	HeLa cells	AuNPs with the size of 1.4 nm were reported to cause necrosis due to the oxidative stress and disruption of membrane integrity	108
1.4, 18, 80	Pregnant rats	Smaller size of AuNPs was associated with teratogenic effects	109
2, 6, 15	MCF-7 cells and female Balb/c nude mice	Higher uptake of smaller AuNPs; 2 and 6 nm AuNPs distributed evenly throughout the cytoplasm and nucleus, and 15 nm AuNPs were only found in the cytoplasm and formed agglomerates	110
		No significant toxic effects of AuNPs on the liver, spleen, kidneys, lungs, and heart	
2.4, 5.5, 8.2, 16, 38, 89	COS-1 cells	Intracellular uptake and localization of AuNPs: 2.4 nm: primarily in the nucleus 5.5 and 8.2 nm: partially in the cytoplasm 16 nm and above: no cellular uptake and located at the cellular periphery	111
2, 10, 25, 40, 50, 70, 80, 100	SK-BR-3 cells	40 and 50 nm AuNPs showed the greatest effect on membrane receptor internalization and enhanced apoptotic activities	112
4, 12, 17	HeLa cells	Uptake forces <i>via</i> raft-mediated endocytosis increased with the size of AuNPs	113
13, 45, 70, 110	CL1-0 and HeLa cells	45 nm AuNPs showed the highest uptake rate and entered cells <i>via</i> endocytosis	114
30, 50, 90	PC-3 cells	30 and 50 nm plain AuNPs were highly taken up by the cells compared to the larger particles PEGylation on the AuNP surface reduced the cellular uptake by reducing the interactions of the anionic plain AuNPs with the negatively charged domains on the cell membrane surface	115
13, 45	CF-31 cells	The major pathway for 45 nm AuNPs was clathrin-mediated endocytosis while for 13 nm AuNPs it was phagocytosis	116
16, 26, 40, 58	RAW264.7 and HepG2 cells	Negatively charged 40 nm AuNPs showed the highest uptake in both cells while positively charged AuNPs had no certain tendency	117
20, 30, 50, 80	CHO-K1 cells	Less internalization with increasing size; 60 800 AuNPs/cell for 20 nm AuNPs, and 50 AuNPs/cell for 80 nm AuNPs	118

AuNPs<sup>106,115</sup> probably due to the elongated particles requiring longer time for membrane wrapping.<sup>121</sup> Despite the fact there were some previous studies reported that rod-shaped AuNPs enhanced the therapeutic outcomes in cancer drug delivery, including prolonged circulation times<sup>122</sup> and enhanced drug loading,<sup>123</sup> it is also important to note that the synthesis of gold nanorods requires the use of a growth-directing surfactant (cetyltrimethylammonium bromide, CTAB). Free CTAB without nanorods was shown to induce cytotoxicity in the human colon cancer HT-29 cell line.<sup>124</sup> However, it was suggested that a polyacrylic acid (PAA) coating was able to reduce the exposure of the cells to the CTAB and hence attenuated the CTAB-capped gold nanorod toxicity. Thus, instead of focusing on the rate of cellular uptake alone, the toxic effects of drug carriers on the cellular system need to be prioritized as well.

In a separate study by Xie *et al.*, mPEGylation of triangular AuNPs resulted in the highest cellular uptake by RAW264.7 cells, followed by rods and star-like structures.<sup>85</sup> The highest uptake of the triangular-like shape was highly associated with the cytoskeleton arrangement and mediated by the dynamin-dependent

endocytosis pathway. These studies indicate that the geometry of AuNPs is an additional feature that can influence the biological interactions, particularly in cellular uptake. However, more repetitive studies need to be conducted to gain a better understanding of the cellular uptake and toxicity implications of drug carriers with different shapes on the cellular system.

## 7. Effect of surface properties of AuNPs on cellular uptake

In addition to size and shape, the interactions of nanoparticles with lipid bilayer cell membranes mainly rely upon the chemical functionalities coated on the nanoparticle surfaces. Generally, modification of the AuNP surface is an important feature that influences the effective use of the particles for drug delivery systems. According to Pissuwan *et al.*, surface modifications for AuNPs are needed in drug delivery systems to prolong residence of the AuNP conjugates in circulation, avoid RES clearance, ensure effective attachment of the desired targeting or therapeutic molecules, improve AuNP stability by



preventing agglomeration and finally, neutralize the possible cytotoxicity caused by stabilizing surfactants during AuNP synthesis.<sup>125</sup> For instance, the surface of AuNPs is coated by either neutral charged groups such as PEG or zwitterionic ligands, or charged functional groups that are anionic or cationic to avoid rapid clearance *via* non-specific uptake by the RES and provide active nanoparticle interactions with cells, respectively.<sup>126</sup> Moreover, a previous preclinical trial of CYT-6091 showed that PEGylated AuNPs carrying rhTNF can escape immunogenicity by avoiding RES phagocytic clearance, thereby allowing the nanotherapeutic to be circulated longer in the blood circulation.<sup>99</sup> Another study using gold nanorods reported that surface-modified gold nanorods with PEG showed no agglomeration due to the nearly complete neutral charge on the nanorod surface, as measured by zeta-potential, while increasing the circulation of the AuNP conjugates in *in vivo* systems.<sup>127</sup>

An example of drug delivery using charged functional groups is provided by Hauck *et al.*<sup>128</sup> In this study, gold nanorods of 18 × 40 nm were coated with various layer-by-layer polyelectrolyte (PE) coatings to produce positively and negatively charged nanorods by electrostatic interactions. Nanorods coated with negatively charged poly(4-styrenesulfonic acid) (PSS) followed by a layer of positively charged poly(diallyldimethylammonium chloride) (PDADMAC) exhibited a higher cellular uptake *via* passive targeting compared to the negatively charged PSS-coated nanorods. Electrostatic interactions between the negatively charged cellular membrane and positively charged nanoparticles may explain this higher uptake.

## 8. Active targeting or tumors by antibody conjugates

To date, no commercial antibody-conjugated drug-loaded nanoparticles have been applied in cancer therapy. However, antibody-conjugated drugs, such as Mylortag®, are commercially available on the market to treat acute myeloid leukaemia.<sup>75</sup> The application of antibodies in cancer therapy has received great attention owing to the expression of specific antigens by cancerous cells. Thus, the development of antibody conjugates as the active targeting ligands in nanoparticle-targeted drug delivery will improve the specificity of chemotherapeutic drug delivery without inducing significant damage to healthy cells.

Antibody grafting *via* covalent linkages at the surface of AuNPs is preferable compared to physical adsorption to reduce competitive displacement of the antibodies in the blood.<sup>75</sup> To demonstrate the efficacy of functionalized antibodies in drug-loaded nanoparticles targeting tumors, Kou *et al.* synthesized cationic SM5-1 single chain antibody (scFv) and polylysine (SMFv-polylys)-coated poly(lactide-*co*-glycolide) (PLGA) nanoparticles loaded with paclitaxel to induce cytotoxicity in human hepatocellular carcinoma cell lines (Ch-hep-3).<sup>129</sup> It was found that the paclitaxel-loaded PLGA in the presence of scFv antibody showed higher cell death compared to the paclitaxel-loaded PLGA only. In a recent AuNP study, conjugation of EpCAM or

TARP antibodies with paclitaxel-loaded AuNPs *via* an *N*-(3-dimethylaminopropyl)-*N'*-ethylcarbodiimide hydrochloride (EDC) and *N*-hydroxysuccinimide (NHS) (EDC/NHS) coupling reaction showed significant T47D breast cancer cell reduction compared to the paclitaxel-loaded AuNPs without antibody conjugates.<sup>130</sup> Both studies support the notion that antibody conjugation provides specific drug delivery in order to enhance the chemotherapeutic action at the targeted tumor site, and hence could be further developed to hinder interactions with healthy cells.

## 9. Concluding remarks

The treatment of CRC has emerged as a topic of interest, since CRC displays high rates of recurrence as well as poor prognosis. The discovery of subpopulations in CRC known as stem cells had led to advanced studies targeting CRCSCs, owing to the stem cell theory that they are able to self-renew and sustain tumor growth. The traditional CRC chemotherapeutic agent, 5-FU, targeting RNA and DNA synthesis and function causes cytotoxicity by interfering with nucleoside metabolism. However, due to the short half-life, non-selectivity and poor biodistribution, the usefulness of 5-FU has been limited. Thus, there is an urgent need to construct an effective targeted drug delivery system to maximize the efficacy of chemotherapeutic agents. Recently, AuNPs have been shown to resolve the limitations of traditional chemotherapy *via* drug delivery systems. The noble AuNPs have various properties that enable them to be designed and functionalized to act as an effective drug carrier, such as biostability, non-toxicity and feasibility for surface modification. Owing to the overexpression of CD133 in CRCSCs, the specific targeting of AuNPs loaded with 5-FU towards CRCSCs can be enhanced *via* bioconjugation of anti-CD133 to the nanoparticles. Therefore, it will be possible for AuNPs to reach and release 5-FU at the targeted site (CRCSCs), thus reducing the interactions with the healthy cells and hence eliminating the systemic side effects previously reported for naked 5-FU without nanodrug carriers.

## Conflicts of interest

There are no conflicts of interest to declare.

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## References

- 1 M. Ghoncheh, M. Mohammadian, A. Mohammadian-Hafshejani and H. Salehiniya, *Ann. Glob. Health*, 2016, **82**, 726–737.
- 2 M. J. Munro, S. K. Wickremesekera, L. Peng, S. T. Tan and T. Itinteang, *J. Clin. Pathol.*, 2018, **71**, 110–116.



3 J. W. Huh, Y. A. Park, K. Y. Lee, S. A. Kim and S. K. Sohn, *Yonsei Med. J.*, 2009, **50**, 704–708.

4 C. Fanali, D. Lucchetti, M. Farina, M. Corbi, V. Cufino, A. Cittadini and A. Sgambato, *World J. Gastroenterol.*, 2014, **20**, 923–942.

5 C. J. Li, X. Zhang and G. W. Fan, *J. Cancer Res. Ther.*, 2014, **10**, 233–239.

6 S. Chen, X. Song, Z. Chen, X. Li, M. Li, H. Liu and J. Li, *PLoS One*, 2013, **8**, e56380.

7 F. Ren, W. Q. Sheng and X. Du, *World J. Gastroenterol.*, 2013, **19**, 2603–2611.

8 B. Viswanath, S. Kim and K. Lee, *Int. J. Nanomed.*, 2016, **11**, 2491–2504.

9 Y. Seium, R. Stupp, T. Ruhstaller, P. Gervaz, G. Mentha, M. Philippe, A. Allal, C. Trembleau, J. Bauer, R. Morant and A. D. Roth, *Ann. Oncol.*, 2005, **16**, 762–766.

10 R. Shiragami, S. Murata, C. Kosugi, T. Tezuka, M. Yamazaki, A. Hirano, Y. Yoshimura, M. Suzuki, K. Shuto and K. Koda, *Int. J. Oncol.*, 2013, **43**, 431–438.

11 M. M. Saber, A. M. Al-Mahallawi, N. N. Nassar, B. Stork and S. A. Shouman, *BMC Cancer*, 2018, **18**, 822.

12 H. Wei, D. Qing, C. De-Ying, X. Bai and F. Li-Fang, *Int. J. Pharm.*, 2008, **348**, 35–45.

13 O. Abdel-Rahman, *Clin. Colorectal Cancer*, 2019, **18**, 58–63.

14 S. Tummala, M. N. S. Kumar and A. Prakash, *Saudi Pharm. J.*, 2015, **23**, 308–314.

15 B. A. Cisterna, N. Kamay, W. Choi, A. Tavakkoli, O. C. Farokhzad and C. Vilos, *Nanomedicine*, 2016, **11**, 2443–2456.

16 K. Liu, Z. Q. Wang, S. J. Wang, P. Liu, Y. H. Qin, Y. Ma, X. C. Li and Z. J. Huo, *Int. J. Nanomed.*, 2015, **10**, 6445–6454.

17 A. K. Mitra, V. Agrahari, A. Mandal, K. Cholkar, C. Natarajan, S. Shah, J. Joseph, H. M. Trinh, R. Vaishya, X. Yang, Y. Hao, V. Khurana and D. Pal, *J. Controlled Release*, 2015, **219**, 248–268.

18 R. V. Kalaydina, K. Bajwa, B. Qorri, A. Decarlo and M. R. Szewczuk, *Int. J. Nanomed.*, 2018, **13**, 4727–4745.

19 R. R. Rasha, *Am. J. Adv. Drug Delivery*, 2016, **4**, 55–57.

20 A. E. Yassin, M. K. Anwer, H. A. Mowafy, I. M. El-Bagory, M. A. Bayomi and I. A. Alsarra, *Int. J. Med. Sci.*, 2010, **7**, 398–408.

21 S. P. Chandran, K. P. Nachimuthu, S. B. Natarajan, M. G. Inamdar and M. S. B. M. Shahimi, *Curr. Cancer Ther. Rev.*, 2018, **14**, 75–87.

22 L. Zhang, X. Gao, K. Men, B. Wang, S. Zhang, J. Qiu, M. Huang, M. Gou, N. Huang, Z. Qian, X. Zhao and Y. Wei, *Int. J. Nanomed.*, 2011, **6**, 2419–2427.

23 J. P. Tiernan, N. Ingram, G. Marston, S. L. Perry, J. V. Rushworth, P. L. Coletta, P. A. Millner, D. G. Jayne and T. A. Hughes, *Nanomedicine*, 2015, **10**, 1223–1231.

24 K. Ogawara, K. Un, K. Tanaka, K. Higaki and T. Kimura, *J. Controlled Release*, 2009, **133**, 4–10.

25 A. Hardiansyah, L. Y. Huang, M. C. Yang, T. Y. Liu, S. C. Tsai, C. Y. Yang, C. Y. Kuo, T. Y. Chan, H. M. Zou, W. N. Lian and C. H. Lin, *Nanoscale Res. Lett.*, 2014, **9**, 497.

26 S. Wilhelm, A. J. Tavares, Q. Dai, S. Ohta, J. Audet, H. F. Dvorak and W. C. W. Chan, *Nat. Rev. Mater.*, 2016, **1**, 16014.

27 B. Duncan, C. Kim and V. M. Rotello, *J. Controlled Release*, 2010, **148**, 122–127.

28 S. S. Banerjee, N. Aher, R. Patil and J. Khandare, *J. Drug Delivery*, 2012, **2012**, 103973.

29 T. A. Larson, P. P. Joshi and K. Sokolov, *ACS Nano*, 2012, **6**, 9182–9190.

30 D. B. Longley, D. P. Harkin and P. G. Johnston, *Nature*, 2003, **3**, 330–338.

31 T. A. Baudino, *Curr. Drug Discovery Technol.*, 2015, **12**, 3–20.

32 X. Yu, S. S. Kanwar, B. B. Patel, J. Nautiyal, F. H. Sarkar and A. P. N. Majumdar, *Transl. Oncol.*, 2009, **2**, 321–328.

33 S. T. Ning, S. Y. Lee, M. F. Wei, C. L. Peng, S. Y. F. Lin, M. H. Tsai, P. C. Lee, Y. H. Shih, C. Y. Lin, T. Y. Luo and M. J. Shieh, *ACS Appl. Mater. Interfaces*, 2016, **8**, 17793–17804.

34 J. J. Casciari, S. V. Sotirchos and R. M. Sutherland, *J. Cell. Physiol.*, 1992, **151**, 386–394.

35 Y. Kato, S. Ozawa, C. Miyamoto, Y. Maehata, A. Suzuki, T. Maeda and Y. Baba, *Cancer Cell Int.*, 2013, **13**, 1–8.

36 M. Carbone, *J. King Saud Univ., Sci.*, 2017, **29**, 284–290.

37 C. L. Zhang, T. Huang, B. L. Wu, W. X. He and D. Liu, *Oncotarget*, 2017, **8**, 75756–75766.

38 D. Abetov, Z. Mustapova, T. Saliev and D. Bulanin, *Tumor Biol.*, 2015, **36**, 1339–1353.

39 J. Xiao, J. Mu, T. Liu and H. Xu, *J. Med. Discovery*, 2017, **2**, jmd17003.

40 R. C. Langan, J. E. Mullinax, M. T. Rajji, T. Upham, T. Summers, A. Stojadinovic and I. Avital, *J. Cancer*, 2013, **4**, 241–250.

41 C. A. O'Brien, A. Pollett, S. Gallinger and J. E. Dick, *Nature*, 2007, **445**, 106–110.

42 L. Ricci-Vitiani, D. G. Lombardi, E. Pilozzi, M. Biffoni, M. Todaro, C. Peschle and R. De Maria, *Nature*, 2007, **445**, 111–115.

43 D. Horst, L. Kriegel, J. Engel, T. Kirchner and A. Jung, *Cancer Invest.*, 2009, **27**, 844–850.

44 C. W. Ong, L. G. Kim, H. H. Kong, L. Y. Low, B. Iacopetta, R. Soong and M. Salto-Tellez, *Mod. Pathol.*, 2010, **23**, 450–457.

45 A. V. Paschall, D. Yang, C. Lu, P. S. Redd, J. H. Choi, C. M. Heaton, J. R. Lee, A. Nayak-Kapoor and K. Liu, *Oncotarget*, 2016, **7**, 78698–78712.

46 K. M. Ropponen, M. J. Eskelinen, P. K. Lipponen, E. Alhava and V. M. Kosma, *Scand. J. Gastroenterol.*, 1998, **33**, 301–309.

47 L. Du, H. Wang, L. He, J. Zhang, B. Ni, X. Wang, H. Jin, N. Cahuzac, M. Mehrpour, Y. Lu and Q. Chen, *Clin. Cancer Res.*, 2008, **14**, 6751–6760.

48 W. Weichert, T. Knosel, J. Bellach, M. Dietel and G. Kristiansen, *J. Clin. Pathol.*, 2004, **57**, 1160–1164.

49 P. Dalerba, S. J. Dylla, I. K. Park, R. Liu, X. Wang, R. W. Cho, T. Hoey, A. Gurney, E. H. Huang, D. M. Simeone, A. A. Shelton, G. Parmiani, C. Castelli and M. F. Clarke, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 10158–10163.



50 J. Papailiou, K. J. Bramis, M. Gazouli and G. Theodoropoulos, *Int. J. Colorectal Dis.*, 2011, **26**, 1–11.

51 R. Pang, W. L. Law, A. C. Chu, J. T. Poon, C. S. Lam, A. K. Chow, L. Ng, L. W. H. Cheung, X. R. Lan, H. Y. Lan, V. P. Y. Tan, T. C. Yau, R. T. Poon and B. C. Y. Wong, *Cell Stem Cell*, 2010, **6**, 603–615.

52 M. Mokhtari and Z. Zakerzade, *Adv. Biomed. Res.*, 2017, **6**, 56.

53 B. B. Wang, Z. J. Li, F. F. Zhang, H. T. Hou, J. K. Yu and F. Li, *Histol. Histopathol.*, 2016, **31**, 299–306.

54 I. S. Hong, G. B. Jang, H. Y. Lee and J. S. Nam, *Int. J. Nanomed.*, 2015, **10**, 251–260.

55 P. M. Glumac and A. M. LeBeau, *Clin. Transl. Med.*, 2018, **7**, 18.

56 D. Corbeil, K. Roper, C. A. Fargeas, A. Joester and W. B. Huttner, *Traffic*, 2001, **2**, 82–91.

57 D. D. Fang, Y. J. Kim, C. N. Lee, S. Aggarwal, K. McKinnon, D. Mesmer, J. Norton, C. E. Birse, T. He, S. M. Ruben and P. A. Moore, *Br. J. Cancer*, 2010, **102**, 1265–1275.

58 B. M. Boman and E. Huang, *J. Clin. Oncol.*, 2008, **26**, 2828–2838.

59 J. N. Rich and S. Bao, *Cell Stem Cell*, 2007, **1**, 353–355.

60 R. Gangemi, L. Paleari, A. M. Orengo, A. Cesario, L. Chessa, S. Ferrini and P. Russo, *Curr. Med. Chem.*, 2009, **16**, 1688–1703.

61 C. Heidelberger, N. K. Chaudhuri, P. Danneberg, D. Mooren and L. Griesbach, *Nature*, 1957, **179**, 663–666.

62 C. Focaccetti, A. Bruno, E. Magnani, D. Bartolini, E. Principi, K. Dallaglio, E. O. Bucci, G. Finzi, F. Sessa, D. M. Noonan and A. Albini, *PLoS One*, 2015, **10**, e0115686.

63 S. Handali, E. Moghimipour, M. Rezaei, Z. Ramezani, M. Kouchak, M. Amini, K. A. Angali, S. Saremy and F. A. Dorkoosh, *Biomed. Pharmacother.*, 2018, **108**, 1259–1273.

64 N. Zhang, Y. Yin, S. J. Xu and W. S. Chen, *Molecules*, 2008, **13**, 1551–1569.

65 K. Bracht, A. M. Nicholls, Y. Liu and W. F. Bodmer, *Br. J. Cancer*, 2010, **103**, 340–346.

66 M. Ishikawa, T. Miyauchi and Y. Kashiwagi, *BMC Cancer*, 2008, **8**, 188.

67 M. W. Saif, K. N. Syrigos and N. A. Katirtzoglou, *Expert Opin. Invest. Drugs*, 2009, **18**, 335–348.

68 F. Casale, R. Canaparo, L. Serpe, E. Muntoni, C. D. Pepa, M. Costa, L. Mairone, G. P. Zara, G. Fornari and M. Eandi, *Pharmacol. Res.*, 2004, **50**, 173–179.

69 R. M. Wohlhueter, R. S. McIvor and G. W. Peter, *J. Cell. Physiol.*, 1980, **104**, 309–319.

70 K. Sztandera, M. Gorzkiewicz and B. Klajnert-Maculewicz, *Mol. Pharm.*, 2018, **16**, 1–23.

71 A. K. Biswas, M. R. Islam, Z. S. Choudhury, A. Mostafa and M. F. Kadir, *Adv. Nat. Sci.: Nanosci. Nanotechnol.*, 2014, **5**, 043001.

72 Y. Matsumura and H. Maeda, *Cancer Res.*, 1986, **46**, 6387–6392.

73 Y. Nakamura, A. Mochida, P. L. Choyke and H. Kobayashi, *Bioconjugate Chem.*, 2016, **27**, 2225–2238.

74 R. Ngoune, A. Peters, D. V. Elverfeldt, K. Winkler and G. Pütz, *J. Controlled Release*, 2016, **238**, 58–70.

75 M. Arruebo, M. Valladares and Á. González-Fernández, *J. Nanomater.*, 2009, **2009**, 439389.

76 H. Maeda, H. Nakamura and J. Fang, *Adv. Drug Delivery Rev.*, 2013, **65**, 71–79.

77 H. Kobayashi, R. Watanabe and P. L. Choyke, *Theranostics*, 2014, **4**, 81–89.

78 L. Kou, J. Sun, Y. Zhai and Z. He, *Asian J. Pharm. Sci.*, 2013, **8**, 1–10.

79 J. R. Nicol, D. Dixon and J. A. Coulter, *Nanomedicine*, 2015, **10**, 1315–1326.

80 D. Bobo, K. J. Robinson, J. Islam, K. J. Thurecht and S. R. Corrie, *Pharm. Res.*, 2016, **33**, 2373–2387.

81 J. W. Park, *Breast Cancer Res.*, 2002, **4**, 95–99.

82 D. L. Cooper, C. M. Conder and S. Harirforoosh, *Expert Opin. Drug Delivery*, 2014, **11**, 1661–1680.

83 X. Huang, X. Teng, D. Chen, F. Tang and J. He, *Biomaterials*, 2010, **31**, 438–448.

84 Z. Chu, S. Zhang, B. Zhang, C. Zhang, C. Y. Fang, I. Rehor, P. Cigler, H. C. Chang, G. Lin, R. Liu and Q. Li, *Sci. Rep.*, 2014, **4**, 4495.

85 X. Xie, J. Liao, X. Shao, Q. Li and Y. Lin, *Nature*, 2017, **7**, 3827.

86 N. Hao, L. Li, Q. Zhang, X. Huang, X. Meng, Y. Zhang, D. Chen, F. Tang and L. Li, *Microporous Mesoporous Mater.*, 2012, **162**, 14–23.

87 R. Mohamud, S. D. Xiang, C. Selomulya, J. M. Rolland, R. E. O'Hehir, C. L. Hardy and M. Plebanski, *Drug Metab. Rev.*, 2014, **46**, 176–190.

88 J. H. Adair, M. P. Parette, E. I. Altinoğlu and M. Kester, *ACS Nano*, 2010, **4**, 4967–4970.

89 E. Hutter and D. Maysinger, *Microsc. Res. Tech.*, 2011, **74**, 592–604.

90 M. F. H. Carneiro and F. J. Barbosa, *J. Toxicol. Environ. Health*, 2016, **19**, 129–148.

91 S. Ramiro, C. Gaujoux-Viala, J. L. Nam, J. S. Smolen, M. Buch, L. Gossec, D. Van Der Heijde, K. Winthrop and R. Landewé, *Ann. Rheum. Dis.*, 2014, **73**, 529–535.

92 T. A. Qiu, J. S. Bozich, S. E. Lohse, A. M. Vartanian, L. M. Jacob, B. M. Meyer, I. L. Gunsolus, N. J. Niemuth, C. J. Murphy, C. L. Haynes and R. D. Klaper, *Environ. Sci.: Nano*, 2015, **2**, 615–629.

93 L. Dykman and N. Khlebtsov, *Chem. Soc. Rev.*, 2012, **41**, 2256–2282.

94 A. Laukka, *UTHHealth News*, <https://www.uth.edu/news/story.htm>, accessed 27 March 2019.

95 S. Verma, P. Utreja, M. Rahman and L. Kumar, *Curr. Nanomed.*, 2018, **8**, 1–18.

96 C. M. Goodman, C. D. McCusker, T. Yilmaz and V. M. Rotello, *Bioconjugate Chem.*, 2004, **15**, 897–900.

97 E. E. Connor, J. Mwamuka, A. Gole, C. J. Murphy and M. D. Wyatt, *Small*, 2005, **1**, 325–327.

98 J. Wang, X. Hu and D. Xiang, *Drug Delivery*, 2018, **25**, 1319–1327.



99 S. K. Libutti, G. F. Paciotti, A. A. Bymes, H. R. Alexander, W. E. Gannon, M. Walker, G. D. Seidel, N. Yuldasheva and N. Tamarkin, *Clin. Cancer Res.*, 2010, **16**, 6139–6149.

100 M. A. Safwat, G. M. Soliman, D. Sayed and M. A. Attia, *Int. J. Pharm.*, 2016, **513**, 648–658.

101 N. Elbialy, M. M. Fathy and W. M. Khalil, *Int. J. Pharm.*, 2015, **490**, 190–199.

102 N. Rizk, N. Christoforou and S. Lee, *Nanotechnology*, 2016, **27**, 185704.

103 A. Imanparast, M. Bakhshizadeh, R. Salek and A. Sazgarnia, *Photodiagn. Photodyn. Ther.*, 2018, **23**, 295–305.

104 C. S. Lee, H. Kim, J. Yu, S. H. Yu, S. Ban, S. Oh, D. Jeong, J. Im, M. J. Baek and T. H. Kim, *Eur. J. Med. Chem.*, 2017, **142**, 416–423.

105 E. C. Dreaden, L. A. Austin, M. A. Mackey and M. A. El-Sayed, *Ther. Delivery*, 2012, **3**, 457–478.

106 B. D. Chithrani, A. A. Ghazani and W. C. W. Chan, *Nano Lett.*, 2006, **6**, 662–668.

107 C. Boyoglu, Q. He, G. Willing, S. Boyoglu-Barnum, V. A. Dennis, S. Pillai and S. R. Singh, *ISRN Nanotechnol.*, 2013, **2013**, 123838.

108 Y. Pan, A. Leifert, D. Ruau, S. Neuss, J. Bornemann, G. Schmid, W. Brandau, U. Simon and W. Jahn-Dechent, *Small*, 2009, **5**, 2067–2076.

109 M. Semmler-Behnke, J. Lipka, A. Wenk, S. Hirn, M. Schäffler, F. Tian, G. Schmid, G. Oberdörster and W. G. Kreyling, *Part. Fibre Toxicol.*, 2014, **11**, 1–12.

110 K. Huang, H. Ma, J. Liu, S. Huo, A. Kumar, T. Wei, X. Zhang, S. Jin, Y. Gan, P. C. Wang, S. He, X. Zhang and X. J. Liang, *ACS Nano*, 2012, **6**, 4483–4493.

111 E. Oh, J. B. Delehanty, K. E. Sapsford, K. Susumu, R. Goswami, J. B. Blanco-Canosa, P. E. Dawson, J. Granek, M. Shoff, Q. Zhang, P. L. Goering, A. Huston and I. L. Medintz, *ACS Nano*, 2011, **5**, 6434–6448.

112 W. Jiang, B. Y. S. Kim, J. T. Rutka and W. C. W. Chan, *Nat. Nanotechnol.*, 2008, **3**, 145–150.

113 Y. Shan, S. Ma, L. Nie, X. Shang, X. Hao, Z. Tang and H. Wang, *Chem. Commun.*, 2011, **47**, 8091–8093.

114 S. H. Wang, C. W. Lee, A. Chiou and P. K. Wei, *J. Nanobiotechnol.*, 2010, **8**, 33.

115 Arnida, A. Malugin and H. Ghandehari, *J. Appl. Toxicol.*, 2010, **30**, 212–217.

116 T. Mironava, M. Hadjiargyrou, M. Simon, V. Jurukovski and M. H. Rafailovich, *Nanotoxicology*, 2010, **4**, 120–137.

117 X. Liu, N. Huang, H. Li, Q. Jin and J. Ji, *Langmuir*, 2013, **29**, 9138–9148.

118 A. Elbakry, E. C. Wurster, A. Zaky, R. Liebl, E. Schindler, P. Bauer-Kreisel, T. Blunk, R. Rachel, A. Goepfertich and M. Breunig, *Small*, 2012, **8**, 3847–3856.

119 F. Osaki, T. Kanamori, S. Sando, T. Sera and Y. Aoyama, *J. Am. Chem. Soc.*, 2004, **126**, 6520–6521.

120 N. Kamaly, Z. Xiao, P. M. Valencia, A. F. Radovic-Moreno and O. C. Farokhzad, *Chem. Soc. Rev.*, 2013, **41**, 2971–3010.

121 V. Voliani, G. Signore, R. Nifosí, F. Ricci, S. Luin and F. Beltram, *Recent Pat. Nanotechnol.*, 2012, **2**, 34–44.

122 Arnida, M. M. Janát-Amsbury, A. Ray, C. M. Peterson and H. Ghandehari, *Eur. J. Pharm. Biopharm.*, 2011, **77**, 417–423.

123 S. Pandey, R. Shah, A. Mewada, M. Thakur, G. Oza and M. Sharon, *J. Mater. Sci.: Mater. Med.*, 2013, **24**, 1671–1681.

124 A. M. Alkilany, P. K. Nagaria, C. R. Hexel, T. J. Shaw, C. J. Murphy and M. D. Wyatt, *Small*, 2009, **5**, 701–708.

125 D. Pissuwan, T. Niidome and M. B. Cortie, *J. Controlled Release*, 2011, **149**, 65–71.

126 A. Verma and F. Stellacci, *Small*, 2010, **6**, 12–21.

127 T. Niidome, M. Yamagata, Y. Okamoto, Y. Akiyama, H. Takahashi, T. Kawano, Y. Katayama and Y. Niidome, *J. Controlled Release*, 2006, **114**, 343–347.

128 T. S. Hauck, A. A. Ghazani and W. C. W. Chan, *Small*, 2008, **4**, 153–159.

129 G. Kou, J. Gao, H. Wang, H. Chen, B. Li, D. Zhang, S. Wang, S. Hou, W. Qian, J. Dai, Y. Zhong and Y. Guo, *J. Biochem. Mol. Biol.*, 2007, **40**, 731–739.

130 Z. Alhalili, J. Shapter, D. Figueroa and B. Sanderson, *Drug Delivery Lett.*, 2018, **8**, 217–225.

