

REVIEW

View Article Online
View Journal | View Issue

Cite this: *Mater. Chem. Front.*,
2020, 4, 400

Received 29th August 2019,
Accepted 23rd October 2019

DOI: 10.1039/c9qm00552h

rsc.li/frontiers-materials

The formation mechanism and fluorophores of carbon dots synthesized *via* a bottom-up route

Dan Qu  and Zaicheng Sun *

Carbon dots (CDs) with incomparable optical properties have attracted extensive attention. However, some unclear issues remain, which has impeded the basic understanding and practical application of CDs. The formation process and chemical structure of CDs are critical factors for understanding their optical properties. In this review, recent progress in the formation and fluorophores of CDs is summarized and discussed, which draws a clear picture of related research and indicates a promising future for further studies.

Introduction

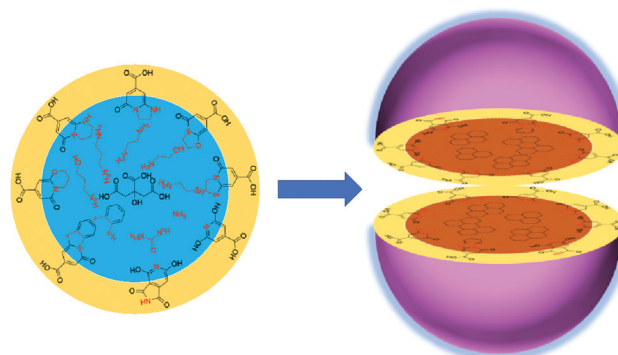
Carbon dots (CDs, encompassing carbon nanodots, C-dots, carbon quantum dots, and graphene quantum dots in this context) have been significantly developed since their discovery in 2004 due to their unique properties and promising potential application.^{1,2} Carbon is generally the main component of CDs; thus, they have very low cytotoxicity and high biocompatibility. They can emit multiple-colored light in the visible light region because they contain both assorted conjugated and abundant surface groups, which are beneficial for functionalization for different applications, such as for use in light-emitting diodes (LEDs), photovoltaic cells, catalysts, fluorescent sensors, bio-imaging agents, and nanomedicines.^{3–8} Recently, great progress has been made in the synthesis of red and near-infrared emissive CDs *via* a bottom-up route, which will further extend the applications of CDs to new fields. It is well known that the typical structure of CDs is treated as a core/shell model that is composed of a carbon core containing graphitic fragments and a shell made of various surface functional groups (Scheme 1). The photoluminescence originated from the conjugated fragments of the core and/or surface chromophore groups. Although many reviews on the synthesis route, properties, and applications of CDs have been published, their formation process and fluorophores remain obscure. For example, it is unknown how to form the conjugated unit from a non-conjugated molecule, *e.g.*, citric acid, as is the fluorophore group on the surface. To address these questions, the formation mechanism must be fully understood, as it will

aid in the understanding of the composition, structure, and photoluminescence origin of CDs. In this review, recent works on the study of the synthesis mechanism of CDs *via* a bottom-up route are summarized. The authors hope that this review will provide the readers with a clear development route that benefits the future design of CDs.

CDs synthesized *via* a bottom-up route are divided into two categories based on the corresponding precursors: those synthesized from non-conjugated molecules and those from conjugated molecules. In the former case, the reactants, such as citric acid, are the non-conjugated molecules, whereas in the latter case, the precursors are usually aromatic molecules.

Non-conjugated molecule-based CDs

In 2008, Bourlinos *et al.*^{9,10} reported the formation of fluorescent carbogenic dots from citric acid (CA) and alkyl amine *via* a pyrolytic process. Since then, CA has become a common and effective carbon source for the preparation of CDs,^{11,12} although other



Scheme 1 Citric acid and various amines for the formation of fluorescent carbon dots.

Center of Excellence for Environmental Safety and Biological Effects, Beijing Key Laboratory of Green Catalysis and Separation, Department of Chemistry and Chemical Engineering, School of Environmental and Energy, Beijing University of Technology, 100 Pingleyuan, Chaoyang District, Beijing 100124, P. R. China.
E-mail: sunzc@bjut.edu.cn

types of carbon sources, such as organic molecules, organic solvents, sugars, and other biomasses, are extensively employed.^{13–19} Until the past 2–3 years, few research groups had devoted themselves to these fields, and a better overview of CDs than what existed before has emerged.

1. Citric acid and ammonia

As per the preceding claim, CA has been extensively used for the synthesis of CDs, and is thus treated as a model non-conjugated molecule. Ammonia is the simplest amine that can be employed in the synthesis of CDs. As early as 1884 and 1894, Behrmann *et al.* and Sell *et al.* investigated the reaction of CA with ammonia or urea, which can release ammonia during the heating process,^{20,21} and indicated the formation of the fluorophore citrazinic acid *via* the reaction. Reckmeier *et al.* synthesized CDs from CA and ammonia *via* hydrothermal reaction and the ammonothermal (supercritical ammonia) method.²² By comparing the optical properties of CDs and citrazinic acid, they confirmed that CDs can be identified as amorphous aggregates of molecular fluorophores based on citrazinic acid derivatives. Schneider *et al.* investigated the reactions of CA with ethylenediamine (EDA), hexamethylenetetramine (HMTA), and triethanolamine (TEOA).²³ The EDA-CDs exhibited strong blue emission with a PL QY of 53% because the fluorophore imidazo[1,2-*a*]pyridine-7-carboxylic acid (IPCA) was generated in the reaction.²⁴ HMTA can release ammonia at elevated temperatures, and citrazinic acid derivatives might be produced from CA and ammonia during the reaction (Scheme 2). However, the fluorophore formation was lower than that in the reaction of CA with EDA because of the slow decomposition and low nucleophilic strength of NH₃. This is the reason why the HMTA-CDs exhibited a weaker fluorescence with a PL QY of 17%. The reaction between CA and TEOA could not produce a similar fluorophore due to the tertiary amines in the reaction. Therefore, citrazinic acid derivatives are the main contributors to the PL of CDs.

2. Citric acid and ethanolamine

In 2011, Krysmann *et al.* investigated the pyrolytic carbonization of CA in ethanolamine (EA).²⁵ They obtained three distinct photoluminescent species that are associated with three different stages of the pyrolytic process. At the low-temperature stage (<180 °C), carbogenic nanoparticles were produced with strong PL, which was assigned to the organic fluorophores. The spectral analyses, including those of FTIR (Fig. 1B), NMR (Fig. 1C), ESI-MS (Fig. 1D), XPS, and UV-vis spectra, revealed an amide functional group in the product formed by a simple

condensation reaction between CA and EA molecules (Fig. 1A). This was further confirmed by the dehydration of tris-(2-hydroxyethyl)ammonium citrate salt at 140 °C under vacuum. The same fluorescent species were obtained with excitation-independent PL at 455 nm with excitation at 375 nm and a high quantum yield (QY = ~50%). Hu *et al.*²⁶ investigated the heating process from room temperature to 170 °C, and the maintenance of 170 °C for different time periods. They found that polymer nanoparticles ~150 nm in diameter were initially formed at a temperature of 130 °C. With the increase of the temperature, the polymer nanoparticles began to shrink to ~60 nm due to dehydration. When the temperature reached 170 °C, many CD seeds with diameters of ~1.5 nm could be found in the shrunken polymer nanoparticles. After 10 minutes, the polymer fragments vanished, and CD particles ~3.5 nm in size could be found (Fig. 1E). Excitation-independent emission was observed after the formation of the nuclei of CDs. The CDs were formed with the extension of the reaction time.

Das *et al.* analyzed the spectra of CDs prepared from CA and EA.²⁷ FTIR spectra confirmed the presence of functional groups including –OH (stretching, 3150–3550 cm^{–1}), sp² C–H (stretching, 3100 cm^{–1}), sp³ C–H (asymmetric stretching, 2940 cm^{–1}, and symmetric stretching, 2882 cm^{–1}), >C=O stretching and –NH bending in amides (1635 and 1540 cm^{–1}), the stretching of sp² C (1422 cm^{–1}), and different vibration modes of –C–N– and –C–O (1355, 1231 and 1055 cm^{–1}). ¹H NMR spectra provided more evidence for the chemical structures. The chemical shifts of ¹H NMR in the region from 8.0307 to 7.68 ppm corresponded to H-bond amide protons, which were similar to that of –CO–NH–CH₂. The peaks at 6.33 and 5.72 ppm were associated with hydroxyl (–OH) protons, and the peaks at 5.87 and 5.63 ppm corresponded to protons attached to sp² carbon. A small peak at 4.7 ppm was attributed to the proton attached to the nitrogen. The triplet peaks ranging from 4.90 to 3.57 ppm, and multiple peaks from 3.28 to 3.08 ppm, corresponded to protons of the methylene group (–CH₂), which attached to O and N atoms in the EA moieties. The AB quartet-type signal from 2.54 to 2.41 corresponded to the methylene group protons of CA (Fig. 1F). The mass spectrum and *m/z* value are presented in Fig. 1G, and the peaks related to the CDs are marked in red. The value of *m/z* = 322 is related to the proton product 1a, which is in agreement with Krysmann's report.²⁵ The peak at *m/z* = 286 indicates the presence of 2-pyridone derivatives, which are proposed as the fluorophores for the CDs. They can further lose water to produce product 1c, the *m/z* values of which are 268.11 and 249.99, corresponding to points 4 and 5 in Fig. 1A. The UV-vis and PL emission spectra of the reaction solution at different stages are presented in Fig. 1H and I. A new absorption band and emission band appeared after the formation of the citrazinic acid derivatives (4 and 5).

Fluorescent CDs were formed when the temperature was above 230 °C for 30 minutes.²⁵ The PL QY of CDs with excitation-independent emission was about 15%, which is much lower than that of the liquid formed at 180 °C. Further pyrolysis at a higher temperature (300 °C) resulted in the formation of CDs with excitation-dependent emission. The PL



Scheme 2 The reaction of citric acid with ammonia.

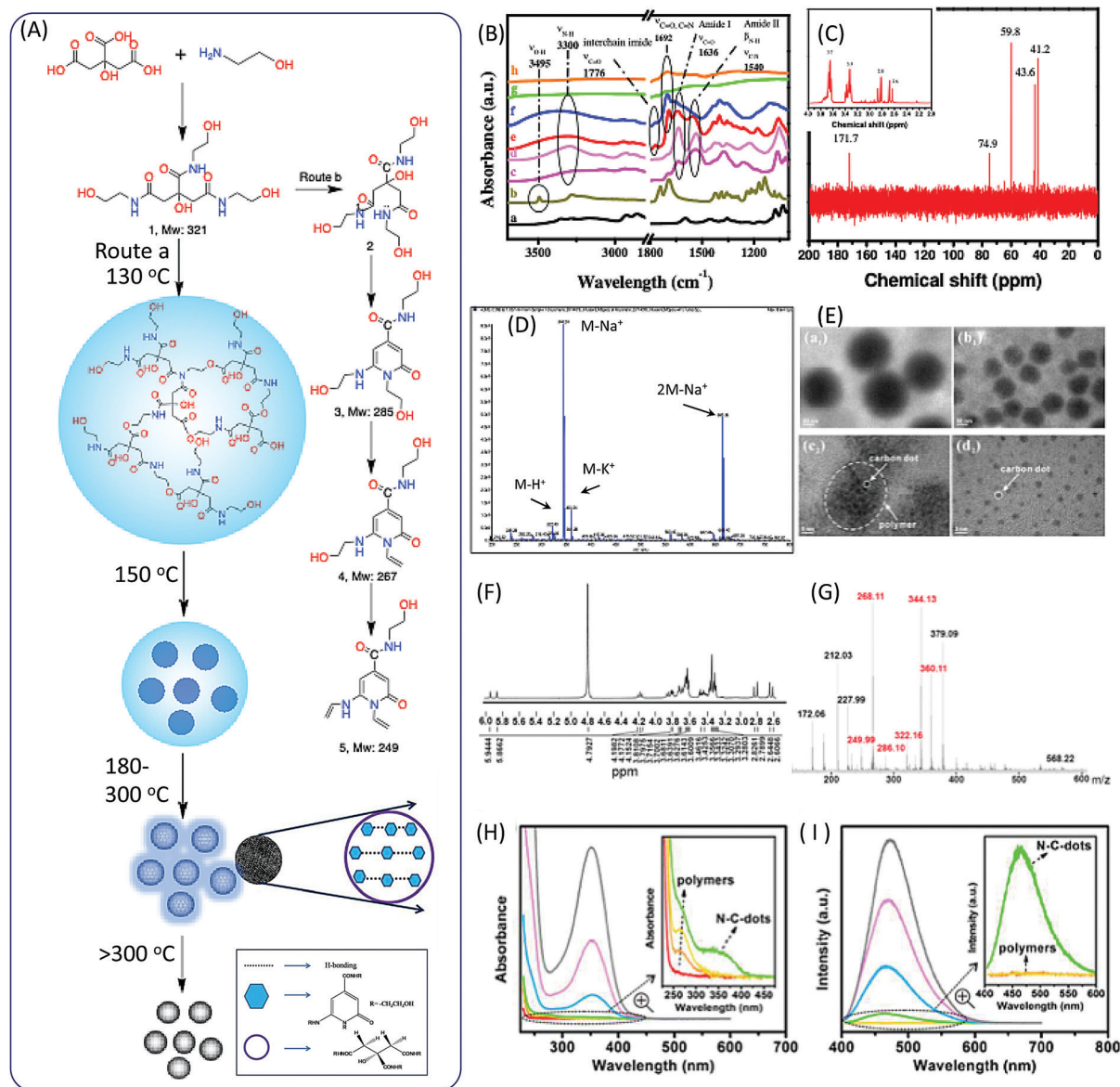


Fig. 1 (A) The scheme of the preparation of CDs from CA and EA. (B) FTIR spectra of (a) CA, (b) EA, (c) CDs prepared at different temperature 180 °C, the PL parts at (d) 180 °C, (e) 230 °C, (f) 300 °C, and (g) 400 °C, and (h) the oxidized part at 400 °C. (C) ¹H-NMR (inset) and ¹³C-NMR spectra of the PL part of CDs prepared at 180 °C. (D) ESI-MS spectrum of the PL part of CDs prepared at 180 °C.²⁵ (E) TEM images of the products at different reaction stages. The temperature reached (a1) 130 °C, (b1) 150 °C, and (c2) 170 °C, and (d2) was maintained at 170 °C for 10 min.²⁶ (F) MALDI MS and (G) ¹H NMR of purified CDs prepared by reflux at 180 °C.²⁷ The (H) adsorption and (I) PL emission of the reaction product of CA and EA at different temperatures (red: room temperature, orange: 130 °C, yellow: 150 °C, green: 170 °C, blue: 170 °C for 2 min, pink: 170 °C for 5 min, and gray: 170 °C for 10 min); the insets show the enlarged UV-vis adsorption and PL spectra.²⁶

intensity and QY were dramatically decreased. The FTIR spectra indicate that imide groups were formed in the CDs. The Raman spectra show a strong G band at 1595 cm⁻¹ in the CDs, indicating the sp² domain embedded in the amorphous carbon (sp³ C) matrix. The authors proposed that the PL behavior of amorphous/disordered graphite containing a mixture of sp² and sp³ carbons could be attributed to the photogeneration of electron-hole pairs; this could have induced the radiative recombination of the trap carriers localized within small sp² carbon clusters that were surrounded by sp³ defects, which is a mechanism that can remain active in the presence of heteroatoms.

The formation of larger and non-uniform particles with diameters of a few hundred nanometers occurred due to aggregation. The PL emission completely vanished when the reaction temperature reached 400 °C.

The overall formation process is illustrated in Fig. 1A. The polymer nanoparticles were first formed between the CA and amine, followed by the formation of citrazinic acid derivatives that exhibited strong fluorescence. CDs with a graphitic core were produced, and contained some aggregates of citrazinic acid derivatives with the extension of carbonization. At the same time, the organic functional groups were left on the

surface due to incomplete carbonization. The citrazinic acid derivatives were assigned as the fluorophores in the CDs.

3. Citric acid and ethylenediamine (EDA)

In 2013, Zhu *et al.* reported highly fluorescent CDs prepared from CA and EDA, the PL QY of which could attain 80% and even higher.^{28,29} Thus, this method has attracted more attention in order to better understand the reaction process. Zhu *et al.*

proposed a reaction process that includes obtaining branched polymers *via* amidation reaction, and then partial carbonization to form graphitic fragments in the amorphous carbonic matrix (Fig. 2A). First, the amide group forms between CA and EDA because CA has three carboxylic acid groups and EDA has two amine groups. Due to the abundant carboxylic acid and amine groups, CA and EDA can react and form either linear³⁰ or cross-linked²⁴ polymers *via* dehydration and condensation depending



Fig. 2 (A) The schematic graph of the formation process of CDs from CA and EDA.²⁸ (B) The reaction of EDA and CA and TA under different conditions to produce three types of CDs. (C) ¹H NMR and (D) ¹³C {¹H} APT NMR of CDs1 (black), CDs2 (blue), and CDs3 (red). (E) Chain isomers for the signal attribution of the top (CDs1, CDs3) and bottom (CDs2).³⁰ (F) Typical TEM (top) and HR-TEM (bottom) images of CDs. (G) ¹H NMR and (H) ¹³C NMR spectra of CDs.²⁸

on the ratio of CA to EDA. Vallan *et al.* investigated CDs prepared from CA and EDA (molar ratio 1:1) in water *via* microwave methods.³⁰ Two types of CDs were synthesized from EDA and either CA or tricarballic acid (TA), and are denoted as CD1 and CD2, respectively. The third type of CD, CD3, was obtained from EDA and CA *via* coupling reaction in the presence of *N,N'*-diisopropylcarbodiimide at room temperature (Fig. 2B). These 3 types of CDs exhibited similar optical properties, as determined by UV-vis and PL spectra. FTIR disclosed that most of the carboxylic groups were involved in H-bonds (peak $I_{1710} > I_{1780}$), and that the amide peaks at 1653 and 1560 cm^{-1} shifted; this was comparable to the peptide, indicating that the amide groups were engaged in H-bonds that strengthened the rigidity of the polymer. Detailed NMR spectroscopy was carried out to reveal the chemical structure. A comparison of the ^1H and ^{13}C $\{^1\text{H}\}$ attached proton test (APT) NMR spectra is presented in Fig. 2C–E. In the ^1H NMR spectra, the sharp line shape of the peaks suggests a compact and static structure. The ^1H – ^{13}C HSQC spectra demonstrate that methylene carbons coupled with a rather condensed set of proton signals. The density of the sharp signals is related to the variety of static chemical environments that surrounded these protons, and can be assigned to the presence of various chain isomers of the repetitive unit that coexisted in the polymer. These three types of CDs exhibited similar NMR spectra, which indicates that the chemical structures of these three CDs can be identified as non-conjugated polymers consisting of the product of the condensation of CA (TA) and EDA. Yang *et al.* thought that the polymer was highly branched, as an excess amount of EDA was employed in the reaction.²⁸ Furthermore, a certain degree of carbonization produced some conjugated fragments in the entangled polymer chains. The carbonization degree is strongly dependent on the reaction conditions. TEM and HR TEM images of the CDs are presented in Fig. 2F, and have a uniform dispersion without apparent aggregation and particle diameters of 2–6 nm. The XRD pattern exhibits a broad diffraction peak at 25 °C corresponding to 0.34 nm, indicating a highly dispersed arrangement of carbon atoms. The Raman spectra also show the presence of a disordered and graphitic band. The chemical shift from 7 to 9 ppm proves the existence of sp^2 C in the ^1H NMR spectrum (Fig. 2G). In the ^{13}C NMR spectrum (Fig. 2H), signals from 30 to 45 ppm, which are assigned to sp^3 carbon atoms, and signals from 100 to 185 ppm, which are indicative of sp^2 carbon, can be observed. Moreover, the X-ray photoelectron spectra also show the presence of sp^2 C, sp^3 C, and oxygenated C. The FTIR spectra show the presence of OH, COOH, and NH_2 groups. However, the defined chemical structure remains unclear, which results in poor reproducibility from batch to batch.

Polymer CDs with blue emission were obtained in the studies by Yang^{16,31,32} and Vallan, and a cross-link-enhanced emission (CEE) effect for the generation of emission without conjugated units was proposed.^{33–37} Vallan *et al.* carried out density function theoretical (DFT) calculations on dimer ($n = 2$) and decamer ($n = 10$) chain configurations (Fig. 3A and B), which exhibited highly intricate structures due to the rigid entanglement of the chain as a result of intramolecular hydrogen bonding (HB). This limited the vibration and/or rotation,

and enhanced the radiative relaxation process. Fig. 3C and D display the HOMO and LUMO of the dimer, respectively. The amide unit ($-\text{CONH}-$) primarily contributed to the HOMO, while the LUMO was attributed to the carboxylic groups ($-\text{COOH}$). The photoinduced charge transfer between these spatially separated groups in the entangled chain can be identified as the origin of fluorescence phenomena in the polymer CDs. This provides an explanation for the fluorescence origin of the polymer dots.

In general, the formation of CDs does not stop in the stage of polymer dots; they are formed in the further carbonization process. CD nanoparticles are treated as having a carbon core with surface groups; thus, they may contain multichromophoric units.³⁸ The core is composed of an sp^2 hybridized carbon domain embedded in an sp^3 hybridized matrix, which is the source of the main absorption feature in the UV region. To mimic the CDs, Fu *et al.* chose three basic polycyclic aromatic hydrocarbons (PAHs), anthracene (3 rings), pyrene (4 rings), and perylene (5 rings), to represent the sp^2 domain.³⁹ Poly(methyl methacrylate) (PMMA) was employed to simulate the sp^3 domain; 0.01 mol% of PMMA monomer units was added into the PMMA film. The absorption and PL spectra, together with those of the CDs, are presented in Fig. 3E and F. The absorbances of pyrene and anthracene are located at the main absorption peak A1, while that of perylene coincides with the absorption shoulder A2. PL from the films demonstrates that the perylene emission overlapped with the maximum emission of the CDs, while the emissions from anthracene and pyrene appeared at a shorter wavelength. The concentration of PAHs in the film also affected the absorption and emission (Fig. 3G). Additionally, PAH molecules could not reproduce the excitation-dependent emission. Based on these results, the CDs cannot be directly mimicked by a single molecule. The film contained a fabricated monomer unit with a molecular molar ratio of 10:10:1:20 anthracene/pyrene/peryene/PMMA. The mimic absorption, PL emission, and excitation-dependent emission are exhibited in Fig. 3I–K. Although the spectrum in Fig. 3I does not fully match the spectrum of the CDs, it confirms that CDs comprise small PAHs embedded in an sp^3 hybridized carbon matrix. The large Stokes shift is due to the exciton self-trapping in the stacked PAH molecules (Fig. 3H). These results successfully demonstrate that the core of the CDs is composed of a few kinds of sp^2 domains embedded in the sp^3 matrix.

Because the CDs were prepared from CA and EDA, the amine took part in the reaction and formed a product that contained heteroatoms in addition to the C, H, and O. Thus, it has been proposed that the blue emission may not be associated with the simple PAH, and is instead associated with citrazinic acid or other 2-pyridone-based molecules, since these molecules can be produced in the common reaction between CA and amine.^{40,41} Song *et al.* analyzed the small fluorescent molecules (1,2,3,5-tetrahydro-5-oxo-imidazo[1,2-*a*]pyridine-7-carboxylic acid, IPCA) that can be produced from CA and EDA at a low temperature (140 °C), and clarified their precise chemical structure (Fig. 4A).²⁴ IPCA showed a strong blue PL emission at 440 nm and two absorption bands at 240 nm and 350 nm, which are similar to

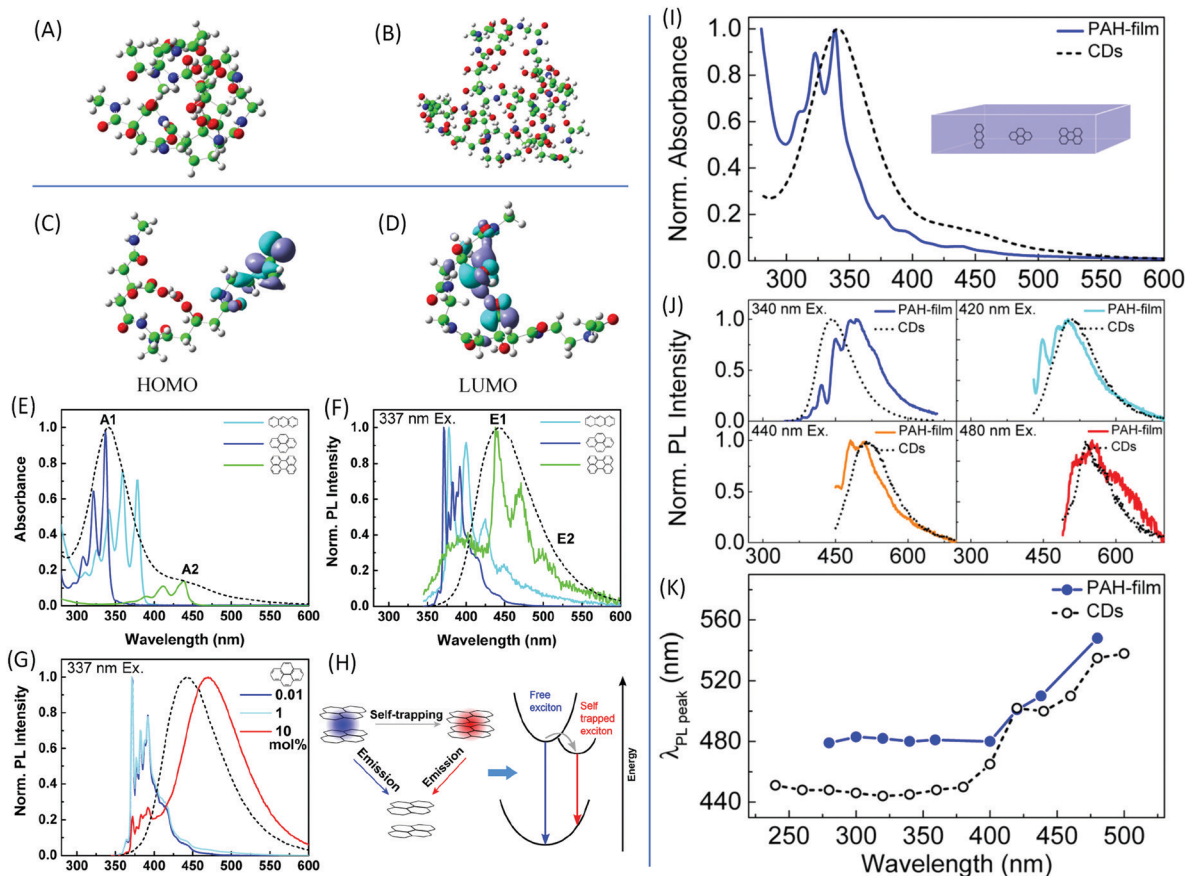


Fig. 3 DFT calculations of the optimized molecular structures of the (A) two-dimer chain and (B) one-decamer chain. (C) HOMO and (D) LUMO involved in the fluorescence phenomenon.³⁰ (E) UV-vis absorption and (F) normalized PL emission spectra upon excitation at 337 nm of anthracene (light blue), pyrene (dark blue), and perylene (green) dispersed in the PMMA matrix with concentrations of 0.01 mol%, as well as of CDs in aqueous solution (black dashed line). (G) Normalized PL spectra of pyrene in PMMA film excited at 337 nm with different pyrene concentrations. (H) Scheme of the exciton self-trapping process in a pyrene molecule pair: a free exciton (blue spot) that may be self-trapped on a molecule pair as a self-trapped exciton (red spot), causing the reduction of energy mobility. (I) Normalized absorption spectra of a PMMA film containing a blend of PAHs (blue) with a molecular ratio of 10 : 10 : 1 : 20 anthracene/pyrene/peryene/PMMA monomer unit, and of the CDs in aqueous solution (black dashed line). (J) Normalized PL spectra of the same samples excited at different wavelengths with the CDs' PL spectra (black dashed line) as references. (K) PL peak wavelength as a function of the excitation wavelength of the same samples as in (I) and (J).³⁹

those of CDs (Fig. 4B). This implies that IPCA can considerably contribute to the absorbance and PL in CDs. The PL lifetime of IPCA was found to be about 14.06 ns, and its single exponential PL intensity decay tendency indicated a simple PL center (Fig. 4C). In addition, -OH, C=O, C-N (CQN), -NH, and -CH groups were also confirmed by the IR spectra (Fig. 4D). The molecular mass of IPCA is 181. In the mass spectrum, the molecular ion (m/z) is 180 (Fig. 4E), and the ^1H and ^{13}C NMR spectra also disclosed the chemical structure (Fig. 4F and G). IPCA was also synthesized by refluxing CA and EDA at ambient pressure, indicating that IPCA is a critical PL center of CDs.

In sum, the entire reaction may include the following steps: (i) polymer aggregate nanoparticles are formed by the condensation of CA and EDA in the hydrothermal reaction; (ii) polymer nanoparticles are further carbonized to form an amorphous matrix; and (iii) at the same time, partial polymer units could form IPCA as a conjugated domain, and the fluorophore is embedded in the matrix (Fig. 4H).

4. CA and EDA derivatives

Song *et al.* also pointed out that EDA derivatives are employed in the synthesis of CDs, and that IPCA derivatives are also produced in the corresponding CDs (Fig. 5A). Alkyl chains or other groups took the place of one hydrogen on the amine, and similar IPCA derivatives were produced similarly. Et-CDs and Ac-CDs with respective PL QY values of 77% and 46% were obtained *via* the same reaction process. The NMR results imply that the products have basic IPCA structures and exhibit similar optical properties. If the propane diamine (PrDA) was employed to take the position of EDA, PPCA would be observed in the reaction; the mass and NMR spectra disclosed the chemical structure, which exhibits strong blue emission with a PL QY of 73% (Fig. 5B and C).

Liu *et al.* investigated the CA and diethylenetriamine (DETA) reaction system to produce CDs at different reaction temperatures (180, 250, and 300 °C).^{42,43} FTIR spectra (Fig. 5D) confirmed the presence of amide groups (1648 cm^{-1} stretching of the amide and 1555 cm^{-1} in-plane bending). XPS spectra indicated the existence

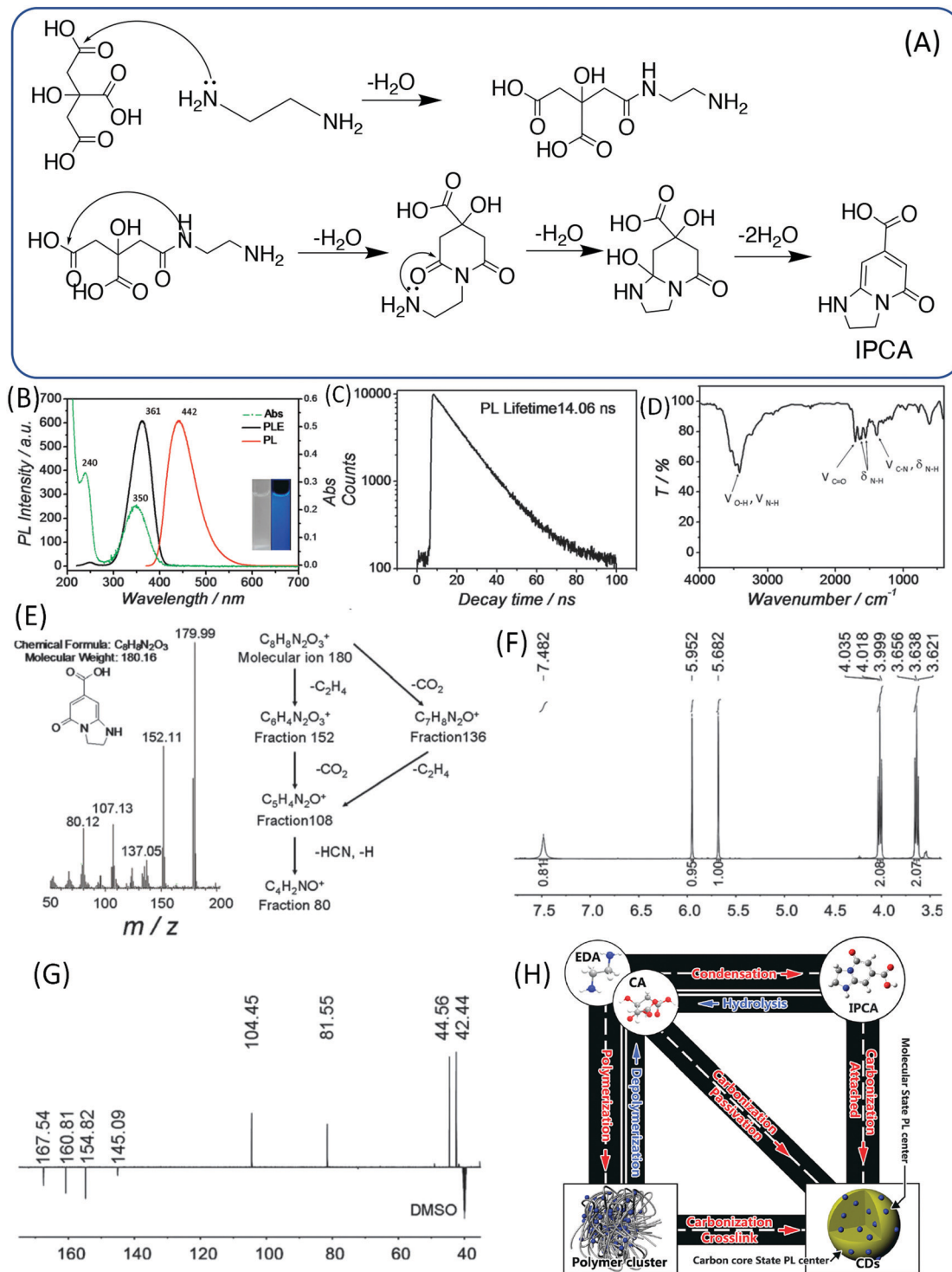


Fig. 4 (A) The assumed process of forming the molecule (IPCA) from CA and EDA. (B) UV-vis absorption, PL emission and excitation spectra of IPCA; inset: photographs of IPCA solution under visible and UV light. (C) Time-resolved PL lifetime decay, (D) FTIR, (E) mass spectrum, (F) ^1H NMR, and (G) DEPTQ ^{13}C NMR spectra of IPCA. (H) A schematic of the relationship between different products in the one-pot hydrothermal system of CA and EDA.²⁴

of amide carbonyl (288.0 eV in C 1s), pyridinic N (398.9 eV), pyrrolic N (399.8 eV), and carbonyl O (531.5 eV). The value of $m/z = 224.1$ was observed in the LC-MS analysis. ^1H NMR spectra showed the signals at 6.0–9.0 ppm for aromatic hydrogen, and 1.5–2.0 ppm for

aliphatic hydrogen. The high reaction temperature resulted in further condensation and carbonization (broadened peaks), and the peaks at 6.0–10.0 ppm indicated the presence of aromatic carboxylic acid and/or phenol (Fig. 5E–G). This spectral evidence proved that



Fig. 5 (A) The reaction of CA with EDA derivatives to produce the corresponding fluorophores. (B) The mass spectrum and (C) PL excitation and emission spectra at different excitation wavelengths of PPCA.²⁴ (D) FTIR and (E–G) ^1H NMR of CDs prepared from CA and DETA at different temperatures. (H) The possible chemical structure of a supramolecular nanocluster in CDs-180, which is composed of AEIOP coupled with DETA@5CA. The dashed lines indicate hydrogen bonds.⁴²

1-(2-aminoethyl)-5-oxo-1,2,3,5-tetrahydroimidazo[1,2-a]-pyridine-7-carboxylic acid (AEIOP) was formed in the reaction (inset of Fig. 5E). The authors also proposed the chemical structure of CDs (Fig. 5H).

5. CA and cysteamine (Cam) or L-cysteine (Cys)

The $-\text{OH}$ group in the EA was replaced by $-\text{NH}_2$ to obtain EDA. Cysteamine is a similar molecule to EDA in that the $-\text{SH}$ takes

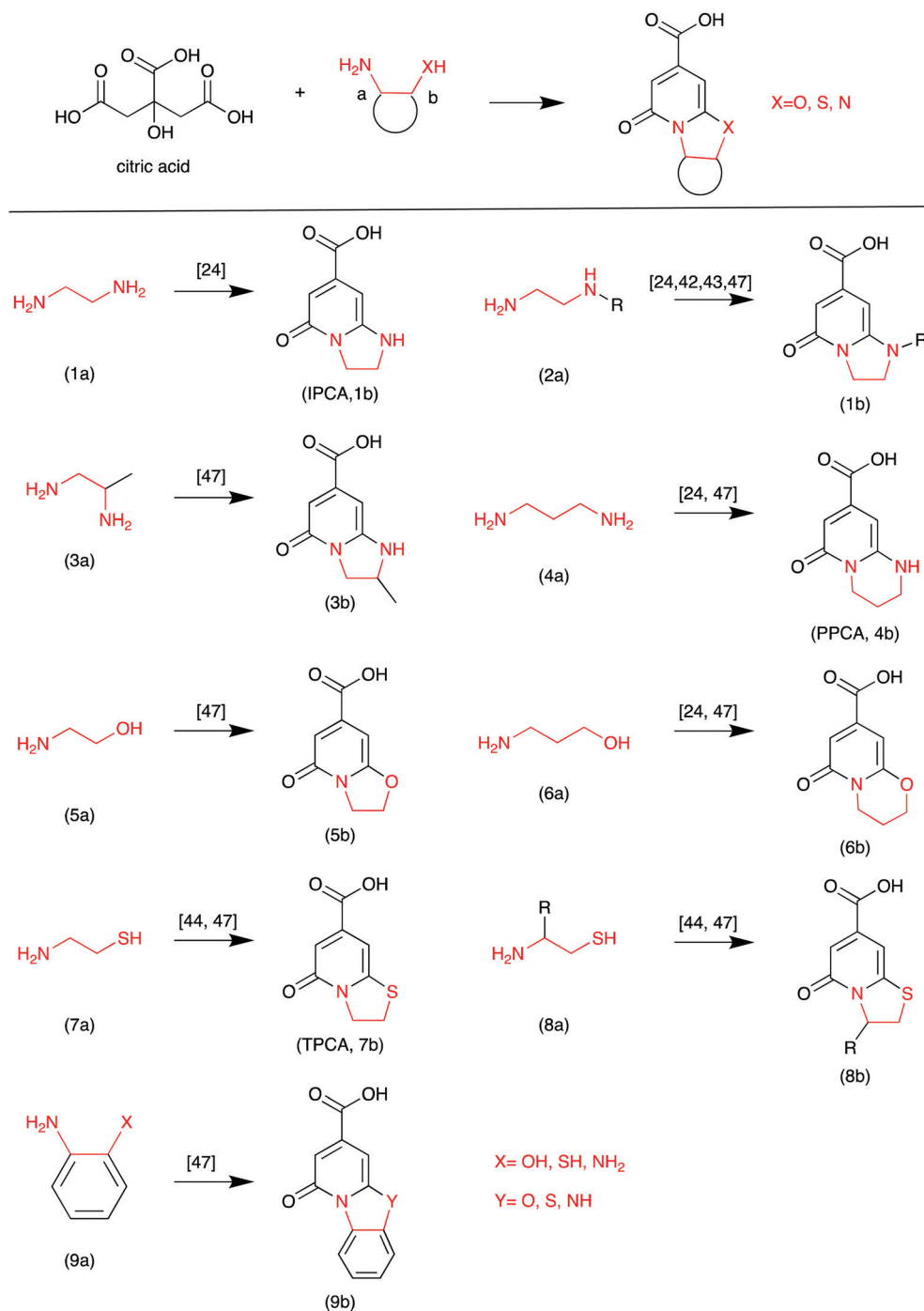
the -OH position. Shi *et al.* investigated N,S-doped CDs prepared from CA and CAM and Cys, which could be treated as a derivative of CAM.^{44,45} They found that N,S-doped CDs also exhibit strong blue fluorescence under UV excitation and excitation-dependent emission. 5-Oxo-3,5-dihydro-2H-thiazolo[3,2-*a*]pyridine-7-carboxylic acid (TPCA) is the main ingredient and actual fluorescence origin of N,S-CDs, which are typical strongly fluorescent citric acid-based CDs with a remarkable QY. TPCA can be directly produced from CA and CAM at relatively low temperatures (Fig. 6A). When CA reacted with Cys, 5-oxo-3,5-dihydro-2H-thiazolo[3,2-*a*]pyridine-3,7-dicarboxylic acid (TPDCA) was formed first, and TPCA was then formed *via* dehydration (Fig. 6A and B). Kasprzyl *et al.* reported the reaction of CA with Cys at 100 °C to synthesize highly fluorescent TPDCA (QY = 62%) (Fig. 6C and D), the chemical structure of which was confirmed by NMR and ESI-MS.⁴⁶ The NMR and MS spectra also showed that the N,S-doped CDs have similar chemical structures to TPCA (Fig. 6E–G). The N,S-doped CDs also exhibit similar optical properties including absorption, fluorescence, and pH-dependent emission, as well as photobleaching phenomena. Based on the

obtained data, Kasprzyl *et al.* believed that the organics might be the main ingredient when the reaction temperature is lower than 200 °C. The PL QY will be reduced with the further increase of the reaction temperature due to the loss of organic fluorophores during carbonization.

Based on the literature, highly fluorescent molecules could be synthesized from CA and α,β -amines, β -amino alcohols, and β -amino thiols. Kasprzyl *et al.* proposed the reaction in Scheme 3.⁴⁷ The fluorescent CDs were synthesized from CA as the carbon source and diamines as the nitrogen source. The authors provided detailed NMR and MS analyses of the small fluorophores from CA and α,β -amines, β -amino alcohols, and β -amino thiols. For example, fluorophore 9b (Y = NH) was synthesized from CA and *o*-phenylenediamine. Its HSQC NMR spectra and ESI-MS/MS are presented in Fig. 8, which further confirms its chemical structure. Recently, Yuan *et al.* confirmed that blue and green emissive CDs can be obtained from CA and diaminonaphthalene (DAN), and have a high PL QY (>70%) due to the extension of the conjugation length.⁴⁸



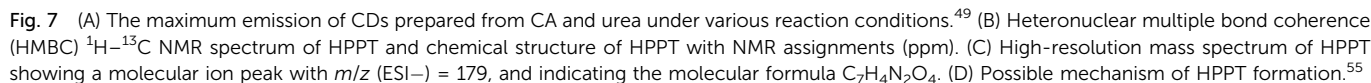
Fig. 6 (A) The synthesis approaches and chemical structures of TPDCA and TPCA. (a) Synthesis of TPDCA by heating a powder mixture of citric acid and L-cysteine, and (b) synthesis of TPCA by heating TPDCA powder to decarboxylate. (B) Synthesis of TPCA by heating a powder mixture of citric acid and cysteamine.⁴⁴ (C) UV-vis spectra and (D) PL excitation and emission spectra of TPDCA, TPCA, and N,S-CDs; inset: photos of TPDCA (left) and TPCA (right) solutions under daylight (top) and under 365 nm UV light (bottom). (E) ^1H -NMR comparison of TPCA and N,S-CDs. (F) ^{13}C -NMR comparison of TPCA and N,S-CDs. (G) High-resolution MS spectrum of N,S-CDs.⁴⁶

Scheme 3 The CA and EDA derivative reaction.⁴⁷

6. CA and urea/thiourea

Beyond the EDA and its derivatives, urea and its derivatives are other common nitrogen sources for CDs. Blue-emissive CDs were obtained from CA and EDA derivatives; however, not only blue-emissive, but also multiple-colored emissive CDs were synthesized from CA and urea by varying the reaction conditions, *e.g.*, the ratio of CA to urea and the reaction temperature (Fig. 7A).^{49–51} Li *et al.* investigated the reaction system of CA

and urea,⁵² and proposed a process similar to that of the CA and EDA system. The CA and urea polymerized to form polymer nanoparticles, and then carbonized to form the sp^2 domain in the amorphous sp^3 matrix. To understand the chemical structure of the fluorophores of CDs, Strauss *et al.* examined the reaction of CA with urea *via* the microwave method.⁵³ They found two kinds of CDs; the closed reaction system produced a blue solution, whereas the open reaction produced a light yellow solution. The former exhibited narrow excitation-independent



blue emission, and the latter displayed excitation-dependent blue-green emission. During the reaction, urea could decompose and release various amine species. While these amines could be released in the open system, they could not in the closed system. The NMR spectra confirmed the presence of carbons stemming from carboxylates and amides, and the aromatic carbon atoms attached to strong electron-withdrawing groups. Zholobak *et al.* thermalized CA and urea in different molar ratios without any solvents, as urea can melt at temperatures greater than 133 °C.⁵⁴ They also found that the blue emission turned into green emission when the molar ratio of CA to urea was greater than 1:3. They hypothesized that the origin of the blue-emissive fluorophore was citrazinic acid,^{23,41} and that the green fluorophore was a result of the formation of citrazinic amide.⁵⁴ However, they did not provide detailed evidence for the chemical structure. Kasprzyk *et al.*⁵⁵ obtained two kinds of CDs with green and blue emission from open and closed systems. Ammonia produced in the reaction could be released from the open system, and the ammonia concentration in the reaction system has a critical influence on the emission of CDs. The authors further carried out ESI-MS and NMR experiments to disclose the chemical structure of the fluorophores for these CDs. They ascertained that the blue fluorophore in the CDs was the citrazinic acid produced from CA and ammonia. The green fluorophore was 4-hydroxy-1*H*-pyrrolo[3,4-*c*]pyridine-1,3,6(2*H*,5*H*)-trione (HPPT). The ¹H-¹³C NMR and ESI-MS/MS spectra are presented in Fig. 7B and C. The authors also proposed the two formation mechanisms presented in Fig. 7D. One possible reaction route is that the citrazinic acid or amide was first formed by the reaction of CA with NH₃. The isocyanic acid, which was produced from the decomposition of urea, was then added at position 3 of the pyridine ring. Finally, the imide ring was closed to form HPPT. Another possible reaction route is the formation of an intermolecular amide between citrazinic acid and urea. Intramolecular condensation and cyclization then occurred with the exclusion of ammonia. Therefore, the CA and urea reaction system is similar to the CD and EDA reaction system, and the formation of CDs contains a few steps such as the formation of polymer nanoparticles and carbonization. The difference is that the fluorophore was HPPT.

To briefly summarize Table 1, which presents the characteristics of the CDs prepared from CA, the general reaction route begins with the polymerization of CA and amines to produce the polymer nanoparticles. Meanwhile, the fluorophores of citrazinic acid derivatives such as IPCA are formed between CA and RNH₂. With the extension of the reaction temperature and time, the aromatization generates conjugated molecules which work as fluorophores in the CDs. These conjugated molecules are further aggregated to enhance the rigidity of the nanoparticles and form the sp² nanodomain *via* intermolecular interaction, such as π - π stacking and H-bonding.^{40,56} Thus, it forms typical CD structures with an sp² nanodomain embedded into the amorphous sp³ matrix, which is composed of non-conjugated molecules, cross-linked polymers, or amorphous carbon materials. The main fluorophores of CDs are the citrazinic acid derivatives (Scheme 1).⁵⁷ Table 1 summarizes all the reactants and the corresponding explanations. Beyond CA,

various non-conjugated molecules such as sugars,⁵⁸ proteins,⁵⁹ amino acids,^{60,61} and other biomasses¹⁴ are extensively used for the synthesis of CDs. However, the systematic investigation of these species in the formation process is rarely reported, and thus will be a potential research hotspot in the future.

Conjugated molecule-based CDs

1. Phenyldiamines and derivatives

In 2015, Lin *et al.*⁶² reported the formation of red-, green-, and blue-emissive CDs from *p*-, *o*-, and *m*-phenylenediamines (PDAs) in ethanol *via* a solvothermal route (Fig. 8A). This promoted the research of CDs to enter another era. Ding *et al.* synthesized and separated a series of color emissive CDs from urea and *p*-PDA *via* a hydrothermal method followed by the column chromatography technique.⁶³ The presence of various surface functional groups (OH, NH₂, C-O, and COOH) was proven by FTIR spectra. The authors emphasized that the surface defects produced by the oxidation are one of the critical factors that result in a narrow bandgap for red emission. Multiple-color emissive CDs can also be prepared from *p*-PDA in different kinds of solvents.^{64,65} The emission peak can be shifted from 540 nm to 614 nm. In this reaction, there is only one reactant, *p*-PDA, which has one type of functional group (-NH₂). The authors proposed that the *p*-PDA was first polymerized in the reaction, and carbon materials were then formed by further coupling (Fig. 8B). Although they did not provide the NMR or MS results, *p*-PDA polymerization reactions have been reported by other scholars, and the findings have matched their explanation.⁶⁶ Based on the investigation of the solvent effect, the red emission of CDs originates from molecular states. Tan *et al.* calculated the formation of tetramers of *o*-PDA, and found that the fully protonated bi-poly(*p*-PDA) tended to be coupled in transverse growth (-1406.07 kJ mol⁻¹) to form a planar structure, rather than longitudinal growth (-616.25 kJ mol⁻¹) (Fig. 8C).⁶⁷ Very recently, Jia *et al.* used *N,N*-dialkyl-*p*-PDA (alkyl = methyl, ethyl, and propyl) as a single reactant in DMF to produce red-emissive CDs with a PL QY of 86.0%.⁶⁸ The reaction process is presented in Fig. 8D. The corresponding Raman spectrum exhibits a sharp graphitic band. Unlike previously reported CDs, there is no indication of -NH₂ or -OH groups in the FTIR spectra of the CDs (Fig. 8E). In the downfield of the ¹H and ¹³C-NMR spectra (Fig. 8F and G), the peaks appearing in the ranges of 6.5–8 and 110–150 ppm are classified as signals of aromatic hydrogen and carbon, respectively, confirming the presence of a π -conjugated structure. In the high field of the ¹H and ¹³C-NMR spectra, peaks appearing in the range of 1–4 ppm and <60 ppm are assigned to aliphatic hydrogen and carbon, respectively. Theoretical calculations were carried out on the optical properties of CD-X with different surface groups (X = 0, -NH₂, -NMe₂, -NEt₂ and -NPr₂). CD-0 exhibited fluorescence emission (λ_{em}) at 559 nm. Pronounced increases in the red-shift in λ_{em} were observed for CD-NH₂ (583 nm), -NMe₂ (614 nm), -NEt₂ (620 nm), and -NPr₂ (623 nm). The -NR₂ passivation led to a

Table 1 The list of reactants, reaction conditions, PL emissions, and explanations of the formation process

Reactants	Reaction conditions	PL emission	Main product	Explanation	Ref.
CA and NH ₃	Reflux at 120–130 °C	Blue	Citrazinic acid	Citrazinic acid	20–22
CA and ethanolamine (EA)	Heat to 180–300 °C	Blue	No solid at 180 °C	FTIR proves the amidation of CA and EA	25 and 47
		Blue	N-doped CDs at 230 °C	Amidation/imidization of CA and EA	
		Excitation-dependent emission	N-doped CDs at 300 °C	Carbonized nanoparticles	
CA and EA	Heat to 130–170 °C	No emission	Polymer nanoparticles at 130 °C	Polymer nanoparticles at 130 °C	26
		Weak blue emission	Shrunken polymer nanoparticles and aromatization at 150 °C	Shrunken polymer nanoparticles and aromatization at 150 °C	
CA and EA	Reflux at 180 °C for 1 hour	Blue	Carbonized nanoparticles formed in the shrunken polymer nanoparticles at 170 °C for 1 minute, CDs formed at 170 °C for 10 minutes	Carbonized nanoparticles formed in the shrunken polymer nanoparticles at 170 °C for 1 minute, CDs formed at 170 °C for 10 minutes	27
CA and ethylenediamine (EDA)	Hydrothermal at 200 °C for 5 hours	Blue	N-doped CDs	Discovered the fluorophore; molecules 3–5 in Fig. 1	28
		Blue	N-doped CDs	The formation of CDs follows the cross-linked polymer and then carbonogenic nanoparticles. A cross-link-enhanced emission (CEE) mechanism was proposed for the highly efficient fluorescence	47
CA and EDA	Microwave at 140 °C for 3 minutes	Blue	N-doped CDs	H-bond-induced amide and carboxylic group assembly contributed to the HOMO and LUMO. The photoinduced charge transfer between these groups thus constitutes the origin of the strong blue fluorescence emission	30
CA and EDA	In the presence of a coupling agent (diisopropyl carbodiimide) at room temperature	Blue	Polymer nanoparticles		
Tricarballic acid and EDA	Microwave at 140 °C for 3 minutes	Blue	N-doped CDs		
CA and EDA	Hydrothermal at 140–300 °C for 10 hours	Blue	N-doped CDs	Provided the detailed formation process, cross-linked polymer, formation of the conjugated molecule IPCA, and further carbonization to form CDs. Assigned the IPCA as the fluorophore of CDs	24 and 47
CA and EDA derivatives such as Et-EDA, Ac-EDA, and PDA	Hydrothermal at 140 °C for 10 hours	Blue	Confirm the IPCA derivatives as fluorophores, such as Et-/Ac-IPCA, PPCA in Scheme 3		24,47
CA and CAm/l-cysteine	Hydrothermal at 180–300 °C for 10 hours	Blue	N-doped CDs	Assigned AEIOP as the fluorophore of CDs (Scheme 3)	42
	Hydrothermal at 200 °C for 3 hours	Blue	S,N-doped CDs	Assigned TPCA and TPDCA as fluorophores of CDs (Scheme 3)	44, 46 and 47
CA and propanol diamine (PoDA)	Hydrothermal at 180 °C for 1 hour	Blue	N-doped CDs	Assigned PPCA as the fluorophore of CDs (Scheme 3)	47
CA and o-aminophenol/o-aminothiophenol/o-phenylenediamine (o-PDA)	Hydrothermal at 180 °C for 1 hour	Blue	N-doped CDs	Assigned molecule 9b as the fluorophore of CDs (Scheme 3)	47
CA and diamionaphthalene (DAN)	Solvothermal at 200 °C for 1–9 hours in ethanol/H ₂ SO ₄	Blue-red	N-doped CDs	DAN could be considered as the smallest sp ² domain with a unique amino-substituted rigid carbon skeleton structure, which concomitantly acts as a building block to form an intact sp ² cluster that is N-doped in the large rigid π -conjugated structure, and highly surface-passivated with the amino at edge sites	48
CA and urea	Hydrothermal at 130–240 °C for 6 hours	Blue-green	N-doped CDs	Excitation-independent emission at 440 nm at a low reaction temperature	52
				Excitation-dependent emission at a high reaction temperature. The surface state decided the emission	
CA and urea	Microwave in closed and open systems	Blue and green	N-doped CDs	Citrazinic acid and derivatives for fluorophores of blue-emissive CDs	55
p-PDA	Solvothermal at 180 °C for 12 hours in ethanol	Red	N-doped CDs	HPPT for green-emissive CDs	62
m-PDA		Blue			

Table 1 (continued)

Reactants	Reaction conditions	PL emission	Main product	Explanation	Ref.
<i>o</i> -PDA	Hydrothermal in organic solution at 200 °C for 5 hours	Green	N-doped CDs	Formed oligomers of PDA in the longitudinal direction, and polymerization in the transverse direction. The red emission of CDs originated from molecular states	64
<i>p</i> -PDA		Red			
<i>p</i> -PDA	Hydrothermal in acidic aqueous solution at 200 °C for 2 hours	Red	N-doped CDs	First, the formation of the dimer of PDA, and further polymerization in the longitudinal and transverse directions. DFT calculation indicates that polymerization in the transverse direction is preferred	67
<i>p</i> -PDA-NR ₂	Solvothermal in DMF at 200 °C for 12 hours	Red	N-doped CDs	Deamination reaction in the solvothermal process	68
DAP	Hydrothermal at 250–380 °C for 16–48 hours	Tuneable emission from blue to NIR	C ₃ N	DAP polymerized in the longitudinal and transverse directions to form 2D C ₃ N	76
<i>o</i> -DHB	Solvothermal in hydrazine ethanol solution at 160 °C for 12 hours	Blue-yellow	N-doped CDs	Benzoquinone was formed by the oxidation of DHB. The cross-linking and carbonization happened between DHB and benzoquinone to obtain the CDs	79 and 81
<i>m</i> -DHB		Green-yellow			
<i>p</i> -DHB	Hydrothermal in acidic solution at 200 °C for 8 hours	Blue-red	N-doped CDs	Dopamine cross-linked with <i>o</i> -PDA; oxidative acid may promote the oxidation of dopamine and/or DPA	82 and 83
Dopamine and <i>o</i> -PDA		NIR			
<i>m</i> -DHB	Solvothermal in ethanol at 200 °C	Green and red	CDs	CDs in the triangular shape. Green and red emissions for 4 and 7 hours of reaction time, respectively	84
Naphthalenediol (DHN) or DHB and oxidant (K ₂ S ₂ O ₈ or anthraquinone, etc.)	Solvothermal in ethanol at 180 °C for 4 hours	Blue to red	CDs	The emission of CDs can be adjusted by tuning the ratio of phenol to the oxidant. Part of the phenol can be oxidized into quinone	80
1,3-DHN and KIO ₃	Solvothermal in ethanol at 180 °C for 4 hours	Red	CDs	Cross-link and dehydration	85
Phloroglucinol	Solvothermal in ethanol at 180 °C for 2–24 hours	Blue to red	Triangular CDs	Blue emission for a reaction time of 9 h; green emission for a reaction time of 24 h; yellow emission for a reaction time of 2 h in H ₂ SO ₄ -ethanol; red emission for a reaction time of 5 h in H ₂ SO ₄ -ethanol	86



Fig. 8 (A) Preparation of RGB PL CDs from three different phenylenediamine isomers (*o*-PDA, *m*-PDA, and *p*-PDA).⁶² (B) Illustration of the formation process of CDs based on the polymerization of PPD. (a) Polymerization of PPD, (b) formation of nitrogen-containing fluorophores, and (c) possible structure of CDs.⁶⁴ (C) Formation energies of longitudinal and transverse growths of a dimer of *p*-PDA.⁶⁷ (D) Synthesis of CD-NMe₂, -NEt₂, and -NPr₂ via solvothermal treatment of *N,N*-dimethyl-, *N,N*-diethyl-, and *N,N*-dipropyl-*p*-PD, respectively. (E) FTIR, (F) ¹H NMR, and (G) ¹³C NMR spectra of CD-NMe₂ (black), -NEt₂ (red), and -NPr₂ (blue).⁶⁸

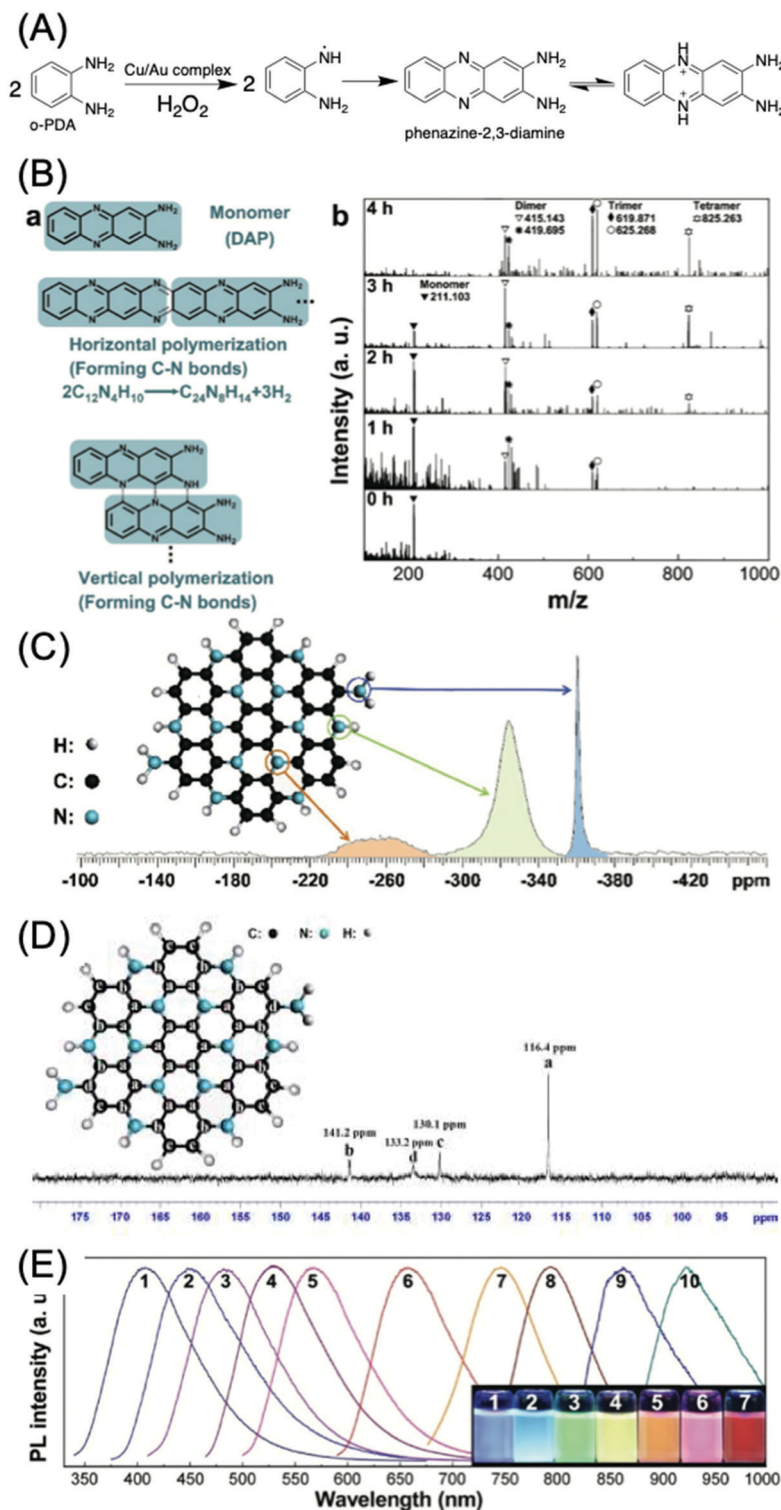


Fig. 9 (A) Mechanism of the catalytic reaction of an o-PDA molecule by the Cu/Au complex.⁷⁵ (B) Polymerization of DAP. (a) Scheme for horizontal and vertical polymerization of DAP forming C–N bonds. The polymerization advances via the abstraction of H from C–H and N–H bonds, followed by the formation of C–N bonds. (b) MALDI-TOFMS spectra of products with different polymerization times (from 0 to 4 h, 623 K, 2.0×10^{-3} M DAP). The peaks in the MALDI-TOFMS spectrum at $m/z = 211.103$, 415.143 (or 419.695), 625.268 (or 625.268), and 825.263 are attributed to the protonated monomer, dimer, trimer, and tetramer, respectively. The lighter fragments are polymerized to heavier fragments as time advances. (C) ¹⁵N-NMR spectrum of a mixture of single-layer and multilayer C₃N sheets. The assignment of the peaks to specific N atoms in the C₃N crystal is indicated. (D) ¹³C-NMR spectrum of a mixture of single-layer and multilayer C₃N sheets. (E) PL spectra of different-sized C₃N QDs (sizes: 1.8, 2.0, 3.0, 3.3, 4.0, and 5.5 nm; for curves 1–6, larger QDs yield IR PL (curves 7–10)); inset: digital photographs of C₃N QD aqueous solutions irradiated using a 100 W mercury lamp (samples 1–7).⁷⁶

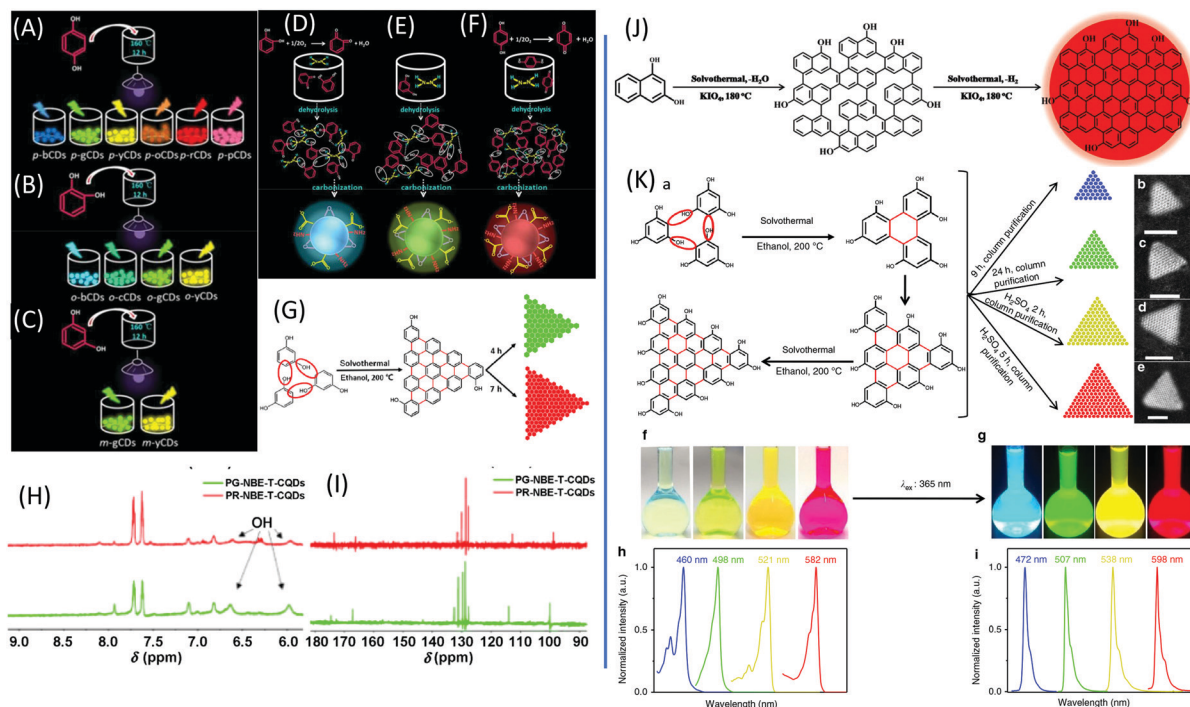


Fig. 10 (A–C) Schematic routes of multiple-color emission CDs from *p*-, *o*-, and *m*-DHB. (D–F) Schematics of the growth processes and structural models of *o*-CDs, *m*-CDs, and *p*-CDs.⁷⁹ (G) The green- and red-emissive CDs prepared from resorcinol via solvothermal treatment. (H) ¹H NMR and (I) ¹³C NMR spectra of green- and red-emissive CDs prepared via route G.⁸⁴ (J) The red emissive CDs prepared from 1,3-dihydroxynaphthalene and KIO₄ via the solvothermal route.⁸⁵ (K) Design and synthesis of narrow bandwidth emission triangular CQDs. (a) Synthesis route of triangular CDs via the solvothermal treatment of PG triangulogen. The typical aberration-corrected HAADF-STEM images of (b) B-, (c) G-, (d) Y-, and (e) R-CDs. Scale bars = 2 nm. (f) Photographs of the CD ethanol solution under daylight and (g) fluorescence images under UV light (excited at 365 nm). The normalized (h) UV-vis absorption and (i) PL spectra of B-, G-, Y-, and R-CDs.⁸⁶

longer λ_{em} than that of $-NH_2$, which is in good agreement with the experimental λ_{em} . These results indicate that the red bandgap emissions of CD-NMe₂, -NEt₂, and -NPr₂ originated from the rigid π -conjugated skeleton structure.

o-Phenylenediamine (*o*-PDA) is another aromatic amine that acts as a reactant for CDs with green, yellow, and red emission.^{69–73} Generally, *o*-PDA is polymerized to form the dimer, trimer, and oligomers (Fig. 9A).^{70,74,75} These oligomers are carbonized and produce emissive CDs. Yang *et al.* began with the dimer of *o*-PDA (2,3-diaminophenazine, DAP) and treated it with a simple hydrothermal method to form C₃N.⁷⁶ The DAP molecules can be polymerized both horizontally and vertically (Fig. 9B(a)). The peaks in the MALDI-TOFMS spectra (Fig. 9B(b)) located at m/z = 211.103, 415.143 (or 419.695), 625.268 (or 625.268), and 825.263 are attributed to the protonated monomer, dimer, trimer, and tetramer, respectively. The lighter fragments were polymerized to heavier fragments with the increase of reaction time, while the DAP was completely consumed after 5 h. Theoretical analysis suggests that the horizontal and vertical polymerization that form C–N bonds are thermodynamically more favorable than other polymerization types. The ¹⁵N NMR spectrum (Fig. 9C) of the C₃N sheet exhibits three singlet peaks, indicating a relative chemical shift. The peaks located at δ = –360.03 and –323.87 ppm can be attributed to the N in the primary amine group and the parahelium group at the edges of the C₃N sheets. The peak

at –261.62 ppm is assigned to aromatic nitrogen (C₃N). The ¹³C NMR spectrum of C₃N is displayed in Fig. 9D, and exhibits four singlet peaks at 141.2, 133.2, 130.1, and 116.4 ppm. The peak located at 116.4 ppm may be attributed to the inner carbon atoms. The peaks located at 141.2 and 130.1 ppm can be attributed to the zigzag-edge carbon atoms, as shown in the schematic diagram. The peak located at 133.2 may be assigned to the zigzag-edge carbon atoms with the amino groups. The size dependence of the bandgap was utilized to tune the PL of C₃N QDs over the entire visible range (400–660 nm) (Fig. 9E, curves 1–6) up to the IR region (curves 7–10). C₃N QDs have a large quantum yield (QY > 0.8) and a relatively long (12.8 ns) lifetime.

m-Phenylenediamine (*m*-PDA), another isomer of PDA, can also be used to generate emissive CDs.^{62,77,78} Zhu *et al.* proposed the presence of a quinoid structure (N=Q=N) and benzenoid (N–B–N) in the emissive CDs, which requires further evidence for confirmation.

2. Phenol and derivatives

Dihydroxybenzene (DHB) and its derivatives were also employed as the reactants for multiple-color emissive CDs. Three DHB isomers (*o*-, *m*-, and *p*-DHB) and hydrazine were used as reactants for the synthesis of N-doped CDs.^{79–81} The reaction was considered to produce benzoquinone, and further dehydration, cross-linking, and carbonization occurred in the reaction to form CDs with

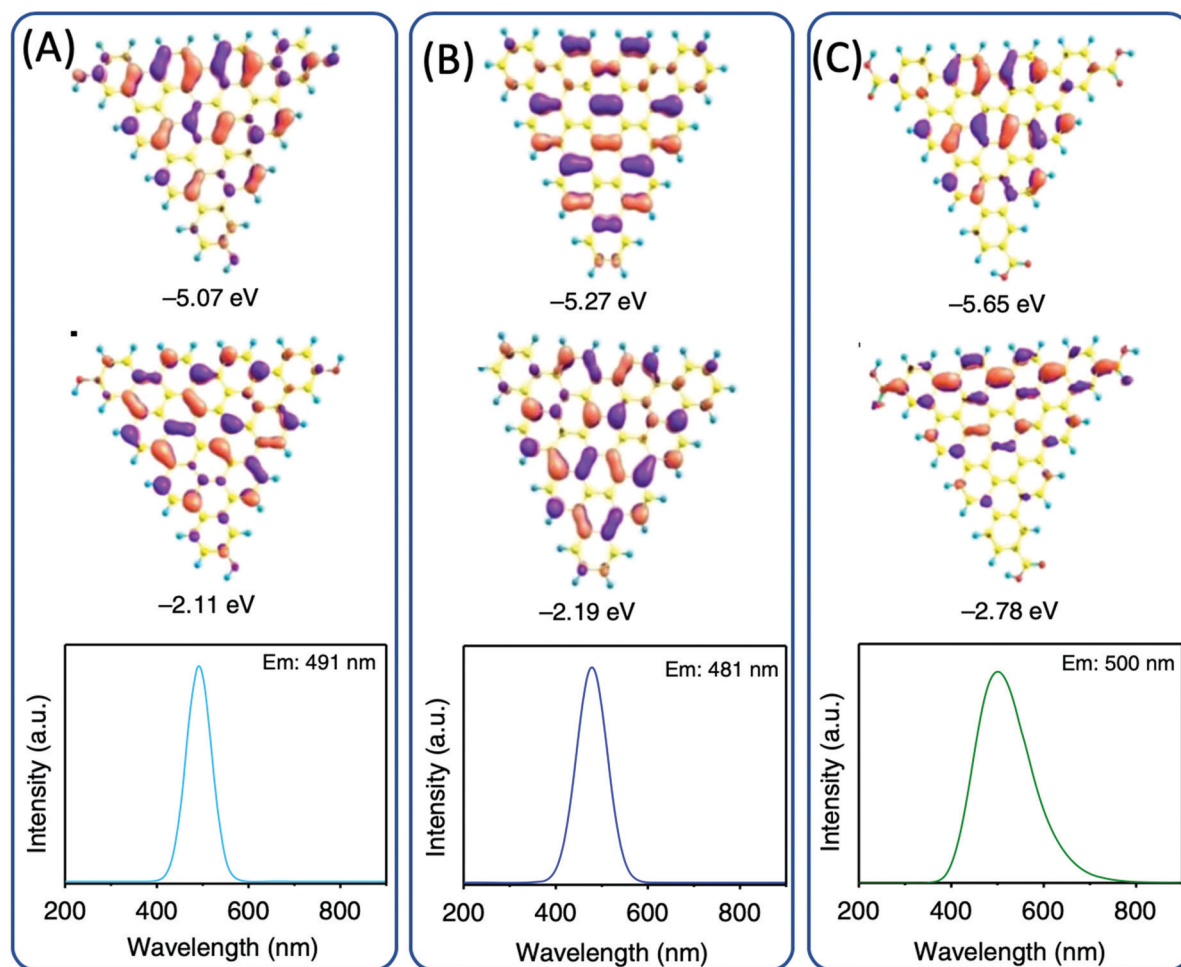


Fig. 11 DFT calculations of the triangular structure model CDs consisting of 19 fused benzene rings (A) functionalized with electron-donating $-OH$ groups and (B) without functionalization and (C) electron-withdrawing $-COOH$ groups. The calculated HOMO (top), LUMO (middle), and PL spectra (bottom).⁸⁶

different color emissions (Fig. 10A–F). Dopamine, as a catechol derivative, was used to synthesize near-IR emissive CDs together with *o*-phenylenediamine.^{82,83} Catechol and *o*-PDA can generate white-emissive CDs that contain three emissive CDs (blue, green, and red light). Pure catechol, pure *o*-PDA, and a mixture of the two produce blue, green, and red emission, respectively.⁶⁹ Yuan *et al.* reported that triangular-shaped, red-emissive CDs can be synthesized from *m*-DHB *via* a simple solvothermal route (Fig. 10G).⁸⁴ In the 1H NMR spectra (Fig. 10H), except for the obvious aromatic hydrogen signals observed in the range of 7–8 ppm, active $-H$ signals from the $-OH$ groups with broad peaks were detected, as indicated by a black arrow. Moreover, ^{13}C NMR spectra (methanol- d_4 , ppm) of the CDs (Fig. 10I) further confirm the functionalization of electron-donating $-OH$ groups at the edge sites. The resonance signals in the range of 160–180 ppm are indicative of the aromatic carbon atoms bonded with $-OH$ groups at the edge sites. The numerous signals observed in the range of 120–140 ppm in the ^{13}C NMR spectra are attributed to sp^2 carbon atoms, further demonstrating the formation of intact sp^2 domains. Similarly, 1,3-dihydroxynaphthalene can also be used as a source for red-emissive CDs with the aid of KIO_3 (Fig. 10J).⁸⁵ Beyond *m*-DHB, phloroglucinol is also a promising reactant for CDs.

Yuan *et al.* reported CDs that were synthesized from phloroglucinol *via* a solvothermal route (Fig. 10K).⁸⁶ The TEM images showed that the CDs were in a typical triangular shape. The triangle size of CDs can be tuned by the reaction conditions. The most promising emission exhibited a relatively narrow full width at half maximum (FWHM) of 29–30 nm, indicating the high color purity of the CDs. Due to the conjugated structure of these CDs, DFT theoretical calculation was carried out to obtain the HOMO, LUMO, and bandgap information of a triangular conjugated structure consisting of 4, 10, and 19 fused benzene rings with electron-donating $-OH$ and electron-withdrawing $-COOH$ groups. Fig. 11 presents the LUMO, HOMO, and calculated PL spectra of triangular CDs containing 19 fused rings and different functional groups. Their corresponding electron cloud density distributions were around the entire molecular structure, indicating a higher degree of delocalization and uniform distribution across the whole molecular structure. The surface group played a critical role in the color purity and emission position. The electron-donating hydroxyl groups at the edge sites exhibited highly delocalized charges and outstanding structural stability, and thus dramatically reduced electron-phonon coupling, which was responsible for the high color-purity excitonic

emission. The electron-withdrawing carboxyl groups on sp^2 -hybridized carbons can induce significant local distortions, and simultaneously act as surface defects that can trap carriers, which ultimately results in the dramatically increased FWHM of the PL spectra of CQDs.

CDs are prepared with an aromatic precursor mainly *via* coupling, condensation polymerization, and carbonization to form fused rings or an sp^2 domain. The fluorophore contributes to the π electron delocalization of the conjugated domain. Thus, an effective conjugation size and electron density distribution play critical roles in the emission position and FWHM.

Conclusions and outlook

With the development of CDs, the understanding of the formation and PL emission of CDs has become increasingly more detailed and clearer. As one of the common carbon sources, citric acid can react with a variety of amines to form citrazinic acid, and with further cyclization and aromatization can produce a conjugated sp^2 domain that enhances the chemical stability and photostability of CDs. Additionally, citrazinic acid and its derivatives are the main contributors to fluorescence in CDs. They are treated as the fluorophores of blue-emissive CDs. Furthermore, red-emissive CDs may be generated from aromatic molecules such as phenylenediamines and dihydroxybenzene. Intermolecular coupling promotes the formation of a planar aromatic structure. These conjugated structures exhibit superior chemical stability and photostability. The delocalized π electron results in an increasingly smaller bandgap. Thus, the optical properties can be tuned by the conjugation size of the CDs.

Although a series of studies has been carried out for the investigation of the formation of CDs, the formation mechanism is still not fully clear. In the citric acid system, citrazinic acid and its derivatives are the main contributors to blue emission; green-, yellow-, and red-emissive CDs can also be produced from citric acid and urea. However, full understanding remains a great challenge because of the many kinds of intermediates produced from urea decomposition. The aromatic system was developed in the past 3–5 years, and thus more research into the formation process is required. In sum, the following questions remain to be clarified.

(1) As is known, CDs have been treated as having a carbon core with an sp^2 domain and surface functional groups. What are the carbonization and aromatization of these non-conjugated molecules?

(2) Citrazinic acid and its derivatives are thought to be the fluorophores of CDs. What are fluorophores from other non-conjugated molecules, such as carbohydrates?

(3) Does the solvent take part in the reaction? What is the solvent effect on the formation of CDs?

(4) The aromatic molecules are treated with a coupling reaction between the molecules. What is the detailed reaction mechanism? How can the coupling reaction between aromatic molecules be tuned and regulated?

(5) Can we finely regulate the reaction process to realize quantum confinement for the carbon materials?

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was financially supported by the Beijing Municipal High Level Innovative Team Building Program (No. IDHT20180504), the Beijing Outstanding Young Scientists Program (No. BJJWZYJH 01201910005017), the National Natural Science Foundation of China (No. 21805004, 21671011, 21872001, and 51801006), the Beijing Natural Science Foundation (No. KZ201710005002 and 2192005), the China Postdoctoral Science Foundation (No. 2018M641133), the Beijing Postdoctoral Research Foundation (No. 2018-ZZ-021), and the Chaoyang District Postdoctoral Research Foundation (No. 2018-ZZ-026).

Notes and references

- 1 X. Xu, R. Ray, Y. Gu, H. J. Ploehn, L. Gearheart, K. Raker and W. A. Scrivens, *J. Am. Chem. Soc.*, 2004, **126**, 12736–12737.
- 2 B. Yao, H. Huang, Y. Liu and Z. Kang, *Trends Chem.*, 2019, **1**, 235–246.
- 3 J. Shen, Y. Zhu, X. Yang and C. Li, *Chem. Commun.*, 2012, **48**, 3686–3699.
- 4 H. Li, Z. Kang, Y. Liu and S.-T. Lee, *J. Mater. Chem.*, 2012, **22**, 24230.
- 5 K. Hola, Y. Zhang, Y. Wang, E. P. Giannelis, R. Zboril and A. L. Rogach, *Nano Today*, 2014, **9**, 590–603.
- 6 X. M. Li, M. C. Rui, J. Z. Song, Z. H. Shen and H. B. Zeng, *Adv. Funct. Mater.*, 2015, **25**, 4929–4947.
- 7 X. T. Zheng, A. Ananthanarayanan, K. Q. Luo and P. Chen, *Small*, 2015, **11**, 1620–1636.
- 8 G. E. LeCroy, S.-T. Yang, F. Yang, Y. Liu, K. A. S. Fernando, C. E. Bunker, Y. Hu, P. G. Luo and Y.-P. Sun, *Coord. Chem. Rev.*, 2016, **320–321**, 66–81.
- 9 A. B. Bourlinos, A. Stassinopoulos, D. Anglos, R. Zboril, M. Karakassides and E. P. Giannelis, *Small*, 2008, **4**, 455–458.
- 10 A. B. Bourlinos, A. Stassinopoulos, D. Anglos, R. Zboril, V. Georgakilas and E. P. Giannelis, *Chem. Mater.*, 2008, **20**, 4539–4541.
- 11 D. Shan, J. T. Hsieh, X. Bai and J. Yang, *Adv. Healthcare Mater.*, 2018, **7**, 1800532.
- 12 S. Zhu, X. Zhao, Y. Song, S. Lu and B. Yang, *Nano Today*, 2016, **11**, 128–132.
- 13 S. N. Baker and G. A. Baker, *Angew. Chem., Int. Ed.*, 2010, **49**, 6726–6744.
- 14 X. Zhang, M. Jiang, N. Niu, Z. Chen, S. Li, S. Liu and J. Li, *ChemSusChem*, 2018, **11**, 11–24.
- 15 M. K. Barman and A. Patra, *J. Photochem. Photobiol., C*, 2018, **37**, 1–22.
- 16 S. Zhu, Y. Song, X. Zhao, J. Shao, J. Zhang and B. Yang, *Nano Res.*, 2015, **8**, 355–381.
- 17 F. Yuan, S. Li, Z. Fan, X. Meng, L. Fan and S. Yang, *Nano Today*, 2016, **11**, 565–586.

- 18 J. Zhang and S.-H. Yu, *Mater. Today*, 2016, **19**, 382–393.
- 19 P. Tian, L. Tang, K. S. Teng and S. P. Lau, *Mater. Today Chem.*, 2018, **10**, 221–258.
- 20 A. Behrmann and A. W. Hofmann, *Ber. Dtsch. Chem. Ges.*, 1884, **17**, 2681–2699.
- 21 W. J. Sell and T. H. Easterfield, *J. Chem. Soc., Trans.*, 1893, **63**, 1035–1051.
- 22 C. J. Reckmeier, J. Schneider, Y. Xiong, J. Häusler, P. Kasák, W. Schnick and A. L. Rogach, *Chem. Mater.*, 2017, **29**, 10352–10361.
- 23 J. Schneider, C. J. Reckmeier, Y. Xiong, M. von Seckendorff, A. S. Susha, P. Kasák and A. L. Rogach, *J. Phys. Chem. C*, 2017, **121**, 2014–2022.
- 24 Y. Song, S. Zhu, S. Zhang, Y. Fu, L. Wang, X. Zhao and B. Yang, *J. Mater. Chem. C*, 2015, **3**, 5976–5984.
- 25 M. J. Krysmann, A. Kellarakis, P. Dallas and E. P. Giannelis, *J. Am. Chem. Soc.*, 2012, **134**, 747–750.
- 26 Y. Hu, J. Yang, J. Tian and J.-S. Yu, *J. Mater. Chem. B*, 2015, **3**, 5608–5614.
- 27 A. Das, V. Gude, D. Roy, T. Chatterjee, C. K. De and P. K. Mandal, *J. Phys. Chem. C*, 2017, **121**, 9634–9641.
- 28 S. Zhu, Q. Meng, L. Wang, J. Zhang, Y. Song, H. Jin, K. Zhang, H. Sun, H. Wang and B. Yang, *Angew. Chem., Int. Ed.*, 2013, **52**, 3953–3957.
- 29 D. Qu, M. Zheng, L. Zhang, H. Zhao, Z. Xie, X. Jing, R. E. Haddad, H. Fan and Z. Sun, *Sci. Rep.*, 2014, **4**, 5294.
- 30 L. Vallan, E. P. Urriolabeitia, F. Ruiperez, J. M. Matxain, R. Canton-Vitoria, N. Tagmatarchis, A. M. Benito and W. K. Maser, *J. Am. Chem. Soc.*, 2018, **140**, 12862–12869.
- 31 S. Zhu, J. Zhang, L. Wang, Y. Song, G. Zhang, H. Wang and B. Yang, *Chem. Commun.*, 2012, **48**, 10889–10891.
- 32 S. Tao, Y. Song, S. Zhu, J. Shao and B. Yang, *Polymer*, 2017, **116**, 472–478.
- 33 E. Zhao, J. W. Y. Lam, L. Meng, Y. Hong, H. Deng, G. Bai, X. Huang, J. Hao and B. Z. Tang, *Macromolecules*, 2014, **48**, 64–71.
- 34 S. Zhu, Y. Song, J. Shao, X. Zhao and B. Yang, *Angew. Chem., Int. Ed.*, 2015, **54**, 14626–14637.
- 35 R. Ye, Y. Liu, H. Zhang, H. Su, Y. Zhang, L. Xu, R. Hu, R. T. K. Kwok, K. S. Wong, J. W. Y. Lam, W. A. Goddard and B. Z. Tang, *Polym. Chem.*, 2017, **8**, 1722–1727.
- 36 T. Han, H. Deng, Z. Qiu, Z. Zhao, H. Zhang, H. Zou, N. L. C. Leung, G. Shan, M. R. J. Elsegood, J. W. Y. Lam and B. Z. Tang, *J. Am. Chem. Soc.*, 2018, **140**, 5588–5598.
- 37 S. Zhu, L. Wang, N. Zhou, X. Zhao, Y. Song, S. Maharjan, J. Zhang, L. Lu, H. Wang and B. Yang, *Chem. Commun.*, 2014, **50**, 13845–13848.
- 38 S. K. Das, Y. Liu, S. Yeom, D. Y. Kim and C. I. Richards, *Nano Lett.*, 2014, **14**, 620–625.
- 39 M. Fu, F. Ehrat, Y. Wang, K. Z. Milowska, C. Reckmeier, A. L. Rogach, J. K. Stolarczyk, A. S. Urban and J. Feldmann, *Nano Lett.*, 2015, **15**, 6030–6035.
- 40 F. Ehrat, S. Bhattacharyya, J. Schneider, A. Lof, R. Wyrwich, A. L. Rogach, J. K. Stolarczyk, A. S. Urban and J. Feldmann, *Nano Lett.*, 2017, **17**, 7710–7716.
- 41 A. Sharma, T. Gadly, S. Neogy, S. K. Ghosh and M. Kumbhakar, *J. Phys. Chem. Lett.*, 2017, **8**, 1044–1052.
- 42 X. Liu, H.-B. Li, L. Shi, X. Meng, Y. Wang, X. Chen, H. Xu, W. Zhang, X. Fang and T. Ding, *J. Mater. Chem. C*, 2017, **5**, 10302–10312.
- 43 Q. Fang, Y. Dong, Y. Chen, C.-H. Lu, Y. Chi, H.-H. Yang and T. Yu, *Carbon*, 2017, **118**, 319–326.
- 44 L. Shi, J. H. Yang, H. B. Zeng, Y. M. Chen, S. C. Yang, C. Wu, H. Zeng, O. Yoshihito and Q. Zhang, *Nanoscale*, 2016, **8**, 14374–14378.
- 45 J. P. Kim, Z. Xie, M. Creer, Z. Liu and J. Yang, *Chem. Sci.*, 2017, **8**, 550–558.
- 46 W. Kasprzyk, S. Bednarz and D. Bogdal, *Chem. Commun.*, 2013, **49**, 6445–6447.
- 47 W. Kasprzyk, S. Bednarz, P. Żmudzki, M. Galica and D. Bogdał, *RSC Adv.*, 2015, **5**, 34795–34799.
- 48 F. Yuan, Z. Wang, X. Li, Y. Li, Z. Tan, L. Fan and S. Yang, *Adv. Mater.*, 2017, **29**, 1604436.
- 49 X. Miao, D. Qu, D. Yang, B. Nie, Y. Zhao, H. Fan and Z. Sun, *Adv. Mater.*, 2018, **30**, 1704740.
- 50 K. Hola, M. Sudolska, S. Kalytchuk, D. Nachtigallova, A. L. Rogach, M. Otyepka and R. Zbořil, *ACS Nano*, 2017, **11**, 12402–12410.
- 51 T. Hu, Z. Wen, C. Wang, T. Thomas, C. Wang, Q. Song and M. Yang, *Nanoscale Adv.*, 2019, **1**, 1413–1420.
- 52 X. Li, S. Zhang, S. A. Kulinich, Y. Liu and H. Zeng, *Sci. Rep.*, 2014, **4**, 04976.
- 53 V. Strauss, J. T. Margraf, C. Dolle, B. Butz, T. J. Nacken, J. Walter, W. Bauer, W. Peukert, E. Spiecker, T. Clark and D. M. Guldi, *J. Am. Chem. Soc.*, 2014, **136**, 17308–17316.
- 54 N. M. Zholobak, A. L. Popov, A. B. Shcherbakov, N. R. Popova, M. M. Guzyk, V. P. Antonovich, A. V. Yegorova, Y. V. Scrypnets, I. I. Leonenko, A. Y. Baranchikov and V. K. Ivanov, *Beilstein J. Nanotechnol.*, 2016, **7**, 1905–1917.
- 55 W. Kasprzyk, T. Swiergosz, S. Bednarz, K. Walas, N. V. Bashmakova and D. Bogdal, *Nanoscale*, 2018, **10**, 13889–13894.
- 56 A. P. Demchenko and M. O. Dekaliuk, *Nanoscale*, 2016, **8**, 14057–14069.
- 57 M. Shamsipur, A. Barati, A. A. Taherpour and M. Jamshidi, *J. Phys. Chem. Lett.*, 2018, **9**, 4189–4198.
- 58 H. Peng and J. Travas-Sejdic, *Chem. Mater.*, 2009, **21**, 5563–5565.
- 59 Z. Zhang, J. Hao, J. Zhang, B. Zhang and J. Tang, *RSC Adv.*, 2012, **2**, 8599.
- 60 J. Jiang, Y. He, S. Li and H. Cui, *Chem. Commun.*, 2012, **48**, 9634–9636.
- 61 F. Arcudi, L. Dordevic and M. Prato, *Angew. Chem., Int. Ed.*, 2016, **55**, 2107–2112.
- 62 K. Jiang, S. Sun, L. Zhang, Y. Lu, A. Wu, C. Cai and H. Lin, *Angew. Chem., Int. Ed.*, 2015, **54**, 5360–5363.
- 63 H. Ding, S. B. Yu, J. S. Wei and H. M. Xiong, *ACS Nano*, 2016, **10**, 484–491.
- 64 T. Zhang, J. Zhu, Y. Zhai, H. Wang, X. Bai, B. Dong, H. Wang and H. Song, *Nanoscale*, 2017, **9**, 13042–13051.
- 65 H. Wang, C. Sun, X. Chen, Y. Zhang, V. L. Colvin, Q. Rice, J. Seo, S. Feng, S. Wang and W. W. Yu, *Nanoscale*, 2017, **9**, 1909–1915.
- 66 T. Plachy, M. Sedlacik, V. Pavlinek, Z. Morávková, M. Hajná and J. Stejskal, *Carbon*, 2013, **63**, 187–195.

- 67 C. Tan, C. Zhou, X. Peng, H. Zhi, D. Wang, Q. Zhan and S. He, *Nanoscale Res. Lett.*, 2018, **13**, 272.
- 68 H. Jia, Z. Wang, T. Yuan, F. Yuan, X. Li, Y. Li, Z. Tan, L. Fan and S. Yang, *Adv. Sci.*, 2019, **6**, 1900397.
- 69 D. Qu, D. Yang, Y. Sun, X. Wang and Z. Sun, *J. Phys. Chem. Lett.*, 2019, **10**, 3849–3857.
- 70 L. Song, Y. Cui, C. Zhang, Z. Hu and X. Liu, *RSC Adv.*, 2016, **6**, 17704–17712.
- 71 D. Bhattacharya, M. K. Mishra and G. De, *J. Phys. Chem. C*, 2017, **121**, 28106–28116.
- 72 K. Jiang, X. Feng, X. Gao, Y. Wang, C. Cai, Z. Li and H. Lin, *Nanomaterials*, 2019, **9**, 529.
- 73 H. Ding, J. S. Wei, P. Zhang, Z. Y. Zhou, Q. Y. Gao and H. M. Xiong, *Small*, 2018, **14**, e1800612.
- 74 M. Vedamalai, A. P. Periasamy, C. W. Wang, Y. T. Tseng, L. C. Ho, C. C. Shih and H. T. Chang, *Nanoscale*, 2014, **6**, 13119–13125.
- 75 Z. Yu, Y. Park, L. Chen, B. Zhao, Y. M. Jung and Q. Cong, *ACS Appl. Mater. Interfaces*, 2015, **7**, 23472–23480.
- 76 S. Yang, W. Li, C. Ye, G. Wang, H. Tian, C. Zhu, P. He, G. Ding, X. Xie, Y. Liu, Y. Lifshitz, S. T. Lee, Z. Kang and M. Jiang, *Adv. Mater.*, 2017, **29**, 1605625.
- 77 P. Zhu, K. Tan, Q. Chen, J. Xiong and L. Gao, *Chem. Mater.*, 2019, **31**, 4732–4742.
- 78 W. Zhou, J. Zhuang, W. Li, C. Hu, B. Lei and Y. Liu, *J. Mater. Chem. C*, 2017, **5**, 8014–8021.
- 79 Y. Wang, Q. Su and X. Yang, *Chem. Commun.*, 2018, **54**, 11312–11315.
- 80 W. Zhu, X. Meng, H. Li, F. He, L. Wang, H. Xu, Y. Huang, W. Zhang, X. Fang and T. Ding, *Opt. Mater.*, 2019, **88**, 412–416.
- 81 J. Wang, C. Cheng, Y. Huang, B. Zheng, H. Yuan, L. Bo, M.-W. Zheng, S.-Y. Yang, Y. Guo and D. Xiao, *J. Mater. Chem. C*, 2014, **2**, 5028–5035.
- 82 S. Lu, L. Sui, J. Liu, S. Zhu, A. Chen, M. Jin and B. Yang, *Adv. Mater.*, 2017, **29**, 1603443.
- 83 B. Wang, J. Li, Z. Tang, B. Yang and S. Lu, *Sci. Bull.*, 2019, **64**, 1285–1292.
- 84 F. Yuan, P. He, Z. Xi, X. Li, Y. Li, H. Zhong, L. Fan and S. Yang, *Nano Res.*, 2019, **12**, 1669–1674.
- 85 Z. Wang, F. Yuan, X. Li, Y. Li, H. Zhong, L. Fan and S. Yang, *Adv. Mater.*, 2017, **29**, 1702910.
- 86 F. Yuan, T. Yuan, L. Sui, Z. Wang, Z. Xi, Y. Li, X. Li, L. Fan, Z. Tan, A. Chen, M. Jin and S. Yang, *Nat. Commun.*, 2018, **9**, 2249.