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# Synthesis and intramolecular ring transformation of *N,N'*-dialkylated 2,6,9-triazabicyclo[3.3.1]nonadienes†

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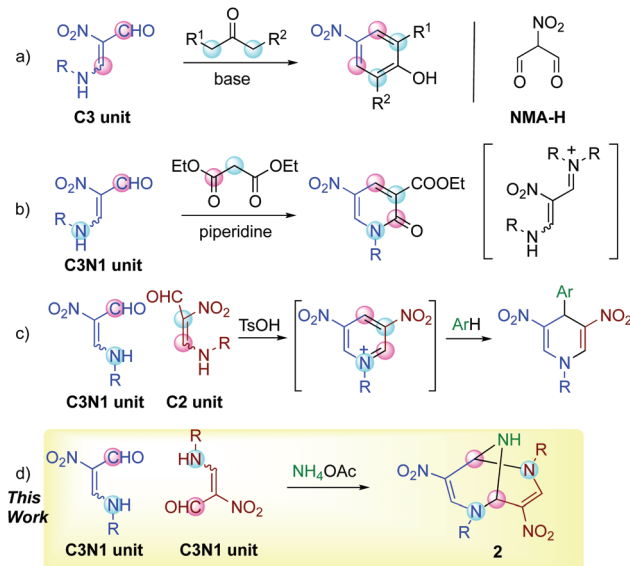
The first and facile synthesis of *N,N'*-dialkylated 2,6,9-triazabicyclo[3.3.1]nonadienes was achieved by the [4 + 4] self-condensation of  $\beta$ -formyl- $\beta$ -nitroenamine in the presence of ammonium acetate. The 2,6- and 2,9-dialkylated products were found to be interconvertible when dissolved in a solvent. This isomerization proceeds through intramolecular ring transformation *via* a common intermediate under equilibrium.

## Introduction

$\beta$ -Formyl- $\beta$ -nitroenamine **1** possesses multiple functionalities such as a formyl, nitro, and amino group, and a carbon-carbon double bond with biased electron density, which bring about diverse reactivities.<sup>1</sup> Two electrophilic sites of **1** react with dinucleophiles such as hydrazines, 1,2-diamines, and ketones to afford nitrated pyrazoles,<sup>2</sup> diazepines<sup>2</sup> and phenols (Scheme 1, a),<sup>3</sup> respectively, in which nitroenamine **1** serves as a synthetic equivalent of nitromalonalddehyde (NMA-H). Its sodium salt (NMA-Na) component has been used for this purpose for a long time. However, NMA-Na should be handled carefully because of the explosive impurity, and aqueous or alcoholic media should be used.<sup>4</sup> In contrast, nitroenamine **1** is safely handleable, and its high solubility in common organic solvents allows its reaction with versatile reagents in organic media.

The dipolar properties of nitroenamine **1** facilitate its reactions with 1,3-dicarbonyl compounds possessing both nucleophilic and electrophilic sites, and polyfunctionalized pyri-

done are synthesized through this process.<sup>5</sup> In these reactions, a secondary amine such as piperidine is found to be a suitable base for condensation, while a tertiary amine such as triethylamine causes no reaction, indicating that the intermediate formation of iminium species is crucial (Scheme 1, b). Furthermore, the [4 + 2] self-condensation of **1** proceeds in the presence of sulfonic acid to form a dinitropyridinium salt *in situ*, which can be trapped by electron-rich benzenes leading to the formation of 4-arylated 1,4-dihydropyridines (Scheme 1, c).<sup>6</sup>



**Scheme 1** Reactions using nitroenamine **1**, in which the pink circles and blue circles indicate electrophilic and nucleophilic sites, respectively.

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†Electronic supplementary information (ESI) available: Spectral data and copies of NMR spectra of compounds 2–6, crystallographic data of **2a** and **3d**, and temperature-dependent NMR spectra. CCDC 2027731 and 2027735. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/d0ob01950j

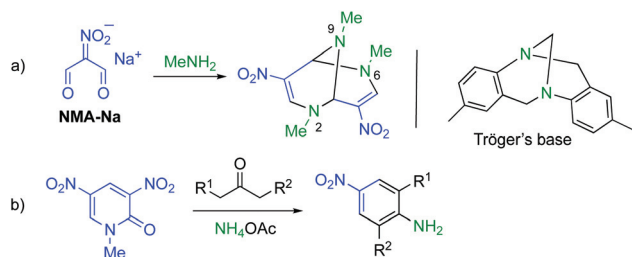


These results prompted us to design a new synthetic method for the triazabicyclic framework, including the [4 + 4] self-condensation of nitroenamine **1**. Tröger's base<sup>7</sup> (3,7-dibenzo-2,6-diazabicyclo[3.3.1]nonane) with a V-shaped structure has attracted much attention because of its chirality,<sup>8</sup> biological activity,<sup>9</sup> molecular recognition ability,<sup>10</sup> and optoelectronic properties.<sup>11</sup> However, the bridgehead nitrogen atoms cannot be modified anymore. Hence, researchers' attention has recently turned towards its aza-analogue, 2,6,9-triazabicyclo[3.3.1]nonane (TABN), because of the availability of further interactions and modifiability of nitrogen atoms in the scaffold.<sup>12</sup> However, it is still difficult to introduce a functional group into the bicyclic framework,<sup>13</sup> except for two examples.<sup>14,15</sup> We focused on one of the examples, Ostercamp's work. The functionalized TABN derivative was first synthesized by the treatment of **NMA-Na** with methylamine (Scheme 2, a).<sup>15</sup> The use of safely handleable nitroenamine **1** was considered to represent a general synthetic method for diverse TABNs because the *N*-substituent of **1** can be easily modified. In Ostercamp's

work, methylamine also serves as a nitrogen source to form a nitrogen bridge, which indicates that the addition of a nitrogen source to our reaction is also necessary. In our study of three-component ring transformation using dinitropyridone, ammonium acetate was found to serve as an excellent nitrogen source and an activator of the substrate (Scheme 2, b),<sup>16</sup> which prompted us to study the synthesis of TABNs using nitroenamine **1** and ammonium acetate (Scheme 1, d).

## Results and discussion

When a solution of nitroenamine **1a** in ethanol was heated at 80 °C for 1 d in the presence of ammonium acetate, four kinds of TABNs **2a–5a** were formed, among which **2a** and **3a** could not be separated by column chromatography (Table 1, entry 1). In the <sup>1</sup>H NMR spectrum of one major product **2a**, a singlet signal assigned to two equivalent olefinic protons and an NH signal showing coupling with the adjacent bridgehead protons were observed. Hence, this isomer was determined to be a 2,6-dipropyl derivative. The other major product **3a** possesses an unsymmetrical structure; one of the olefinic protons was observed as a singlet, and the other was observed as a doublet coupled with the adjacent NH proton, which was also supported by the <sup>1</sup>H–<sup>1</sup>H COSY 2D NMR spectrum. The recrystallization of the **2a** and **3a** mixture afforded a single crystal of **2a**, which was subjected to X-ray crystallography to confirm the structure (Fig. 1, left). Fortunately, a single crystal of dibenzyl derivative **3d** was obtained, and the structure was confirmed to be a 2,9-disubstituted isomer (Fig. 1, right). The use of lower reaction temperature increased the total yield of products **2a–5a**, presumably because the competitive decomposition of



**Scheme 2** Synthetic equivalents of **NMA-H**: (a) sodium salt **NMA-Na** and (b) 1-methyl-3,5-dinitro-2-pyridone.

**Table 1** Formation of four kinds of triazabicyclic compounds **2–5**

					Yield <sup>a</sup> (%)					Recovery of <b>1</b> <sup>a</sup> (%)
Entry	R	NH <sub>4</sub> OAc (equiv.)	Temp. (°C)		2	3	4	5	Total	
1	Pr	<b>a</b>	80		25	12	7	2	46	20
2	Pr	<b>a</b>	50		51	20	19	1	90	10
3	Pr	<b>a</b>	rt		45	12	18	0	75	18
4	Pr	<b>a</b>	50		47	21	23	2	93	4
5	Pr	<b>a</b>	50		39	18	39	1	97	1
6	<i>i</i> -Pr	<b>b</b>	50		27	4	20	0	51	29
7 <sup>b</sup>	<i>t</i> -Bu	<b>c</b>	50		0	0	0	0	0	30
8	–CH <sub>2</sub> Ph	<b>d</b>	50		26	53	11	3	93	4
9	–CH <sub>2</sub> CH=CH <sub>2</sub>	<b>e</b>	50		21	41	15	3	80	6
10	–CH <sub>2</sub> CH <sub>2</sub> OH	<b>f</b>	50		37	24	8	12	81	4

<sup>a</sup> Determined by <sup>1</sup>H NMR. <sup>b</sup> Non-alkylated bicyclic compound **6** was obtained in 26% yield.



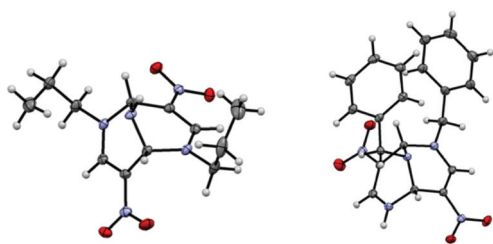


Fig. 1 ORTEP views of **2a** (left, 2,6-di-Pr) and **3d** (right, 2,9-di-Bn). Thermal ellipsoids are represented at probably the 50% level. CCDC: 2027731 (**2a**) and 2027735 (**3d**).†

ammonium acetate was suppressed (entry 2). Although this reaction proceeded even at room temperature, the total yield decreased to an extent (entry 3). The yields of dealkylated products **4a** and **5a** increased when larger amounts of ammonium acetate were used (entries 4 and 5).

The optimized conditions were applied to other nitroenamines **1b–f**. Bicyclic products **2b–5b** possessing bulkier isopropyl groups could be prepared by altering nitroenamine to **1b** (entry 6). When *tert*-butyl-substituted nitroenamine **1c** was employed, the corresponding bicyclic products **2c–5c** were not formed, and non-alkylated bicyclic product **6** was obtained with 26% yield, which was due to the easy elimination of the stable *tert*-butyl cation under acidic conditions (entry 7). It was also possible to introduce functional groups to the ring nitrogen atoms, such as benzyl, allyl, and 2-hydroxyethyl groups, efficiently (entries 8–10).

All attempts to control the selectivity between **2** and **3** failed. During this study, we noticed that the 2/3 ratio changed each time we performed  $^1\text{H}$  NMR. This observation suggests that there is a relationship of equilibrium between **2** and **3**. To confirm this, the single crystals of **2a** and **3d** used for X-ray crystallography were dissolved in deuterated solvents, and the change in the mole fraction ( $[\text{2}]/[\text{2}] + [\text{3}]$ ) was monitored by  $^1\text{H}$  NMR at intervals of several minutes/hours (Fig. 2). Conversion from **2a** to **3a** was observed by only dissolving them in the solvents at room temperature, irrespective of the solvent, without

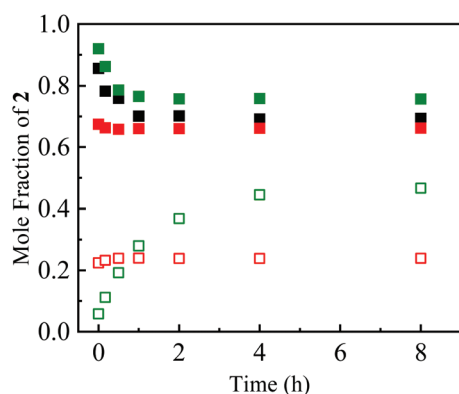


Fig. 2 Time courses of the mole fraction of **2a** (closed squares) and **3d** (open squares) in  $\text{C}_6\text{D}_6$  (black),  $\text{CDCl}_3$  (green) and  $\text{CD}_3\text{CN}$  (red).

the formation of any detectable byproduct. A different behavior was observed when **2a** was dissolved in chloroform-*d*, which might be due to a trace amount of hydrochloric acid in the solvent. Conversely, the formation of **2d** was also observed when **3d** was dissolved in deuterated solvents. The equilibrium constants ( $K = [\text{3}]/[\text{2}]$ ) were temperature-dependent, as shown in Fig. 3. The  $K$  value for **3a/2a** increased upon increasing the temperature, whereas that for **3d/2d** decreased. The Gibbs energy differences between **2** and **3** determined by the Arrhenius-type fittings ( $\ln K = -\Delta G/RT$ ) reveal that **3d** is more stable than **2d** ( $\Delta G < 0$ ) in contrast to the **2a/3a** system (Table 2). The difference originated from the existence of  $\pi$ -stacking between the benzyl groups in **3d**. In addition, the  $K$  values in  $\text{CD}_3\text{CN}$  were smaller than the relevant values in  $\text{CDCl}_3$ , presumably due to the polarity and/or Lewis acidity/basicity of the solvent. These tendencies were qualitatively reproduced by theoretical calculations. Furthermore, when a mixture of **2a** and **3a** was heated in the presence of ammonium acetate, nitroenamine **1a** was formed in 10% yield in the reaction mixture, indicating the existence of equilibrium between **1a** and TABNs **2a** and **3a**.

Based on these results, bicyclic products **2** and **3** are considered to be formed as shown in Scheme 3. The reaction is initiated by the conversion of a formyl group of **1** to iminium

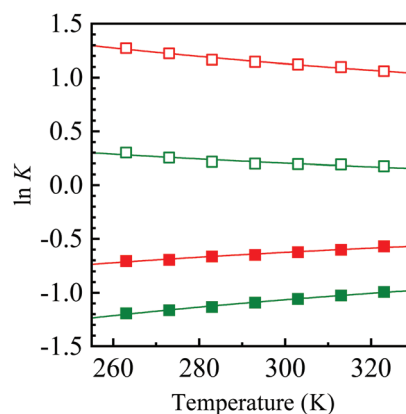


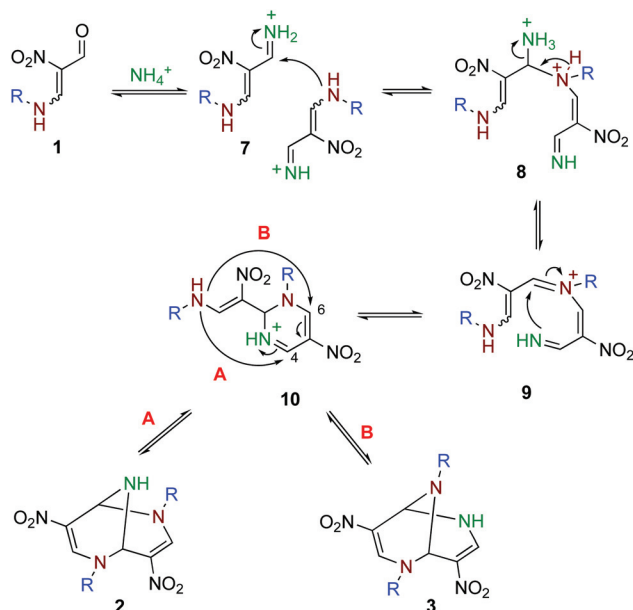
Fig. 3 Temperature dependences of equilibrium constants  $K$  for **3a/2a** (closed squares) and **3d/2d** (open squares) in  $\text{CDCl}_3$  (green) and  $\text{CD}_3\text{CN}$  (red).

Table 2 Observed and calculated Gibbs energy differences in chloroform and acetonitrile

R	Solvent	$\Delta G$ (kJ mol $^{-1}$ )	
		Obsd <sup>a</sup>	Calcd <sup>b</sup>
–Pr	Chloroform	$+2.40 \pm 0.09$	+5.22
	Acetonitrile	$+1.58 \pm 0.12$	+5.29
–CH <sub>2</sub> Ph	Chloroform	$-1.36 \pm 0.22$	+1.87
	Acetonitrile	$-2.38 \pm 0.14$	+0.69

<sup>a</sup> Determined using deuterated solvents. <sup>b</sup> Calculated at the B3LYP/6-311+G(d,p) level.





**Scheme 3** A plausible mechanism for the formation of bicyclic compounds **2** and **3**.

ion **7**, which facilitates the attack of the less nucleophilic amino group of another molecule of **1** forming **8**. After the elimination of ammonia, the formed iminium ion is attacked by the imino group intramolecularly to form a six-membered ring. Dihydropyrimidine **10** is a common intermediate of **2** and **3**. When the amino group in the side chain attacks the 4-position (Path A), 2,6-dialkyl product **2** is formed. On the other hand, 2,9-dialkyl product **3** is formed when the amino group attacks the 6-position of **10** (Path B). A direct rearrangement between **2** and **3** is also possible if a stable cation such as a benzyl and allyl cation is released. However, ring transformation *via* intermediate **10** is considered to be more plausible based on our previous findings; 4,6-dinitro-2,8-diazabicyclo[3.3.1]nona-2-ene and 3-nitropyridine are interconvertible under equilibrium in the presence of ammonium acetate.<sup>17</sup> The use of larger amounts of ammonium acetate is subject to

the conversion of nitroenamine **1** or iminium ion **7** to nitroenamine **11** (Scheme 4), which affords dealkylated products **4** and **5** in a manner similar to that shown in Scheme 3. Products **4** and **5** are also inseparable because there is a relationship between these compounds.

## Conclusions

TABN frameworks were efficiently constructed by the [4 + 4] self-condensation of nitroenamine **1** in the presence of ammonium acetate. Since the *N*-alkyl group of **1** is easily modified, versatile TABNs were available by this protocol. The major products, 2,6- and 2,9-dialkylated TABNs **2** and **3**, were found to be interconvertible when dissolved in a solvent and this isomerization proceeds through intramolecular ring transformation under equilibrium *via* the common intermediate **10**. Further studies on the physical properties of TABNs and chemical conversion using the functionalities are in progress, and the results will be presented in due course.

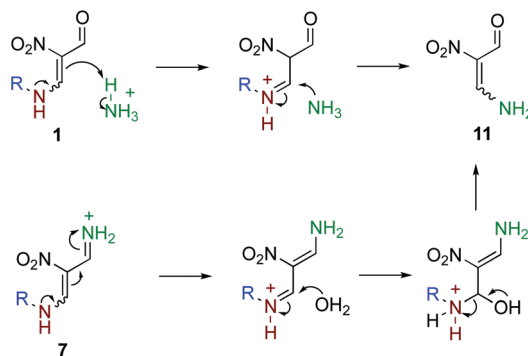
## Experimental section

### General

The melting points were determined using an SRS-Optimelt automated melting point system. All the reagents and solvents were commercially available and used as received. The <sup>1</sup>H NMR spectra were measured on a Bruker Ascend-400 at 400 MHz with tetramethylsilane as an internal standard. The <sup>13</sup>C NMR spectra were measured on a Bruker Ascend-400 at 100 MHz, and the assignments of the <sup>13</sup>C NMR spectra were performed by DEPT experiments. The high-resolution mass spectra were measured on an AB SCIEX Triple TOF 4600. The IR spectra were recorded on a JASCO FT/IR-4200 spectrometer.

### Preparation of nitroenamines 1a–f

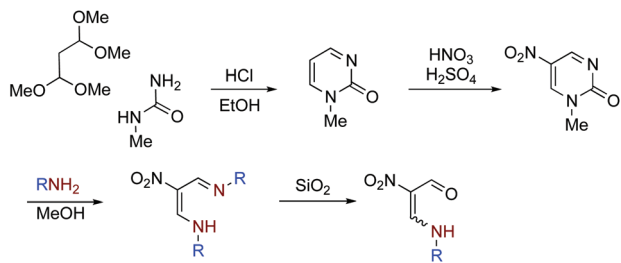
Nitroenamines **1a–f** were prepared according to our established method.<sup>18</sup> Nitropyrimidinone was prepared in 70% overall yield by the condensation of commercially available 1,1,3,3-tetramethoxypropane and *N*-methylurea in 12 M hydrochloric acid followed by nitration with nitric acid in sulfuric acid. To a solution of pyrimidinone (310 mg, 2 mmol) in methanol (40 mL), propylamine (410 μL, 5 mmol) was added, and the mixture was heated under reflux for 3 h. After evaporation, the residue was extracted with hexane (3 × 30 mL), and the removal of hexane afforded NMR pure azadienamine (390 mg, 1.96 mmol, 98%) as a pale yellow oil. A solution of azadienamine (200 mg, 1 mmol) in chloroform (5 mL) was charged on silica gel (20 g) in a column and allowed to stand at room temperature for 1 day, and then it was eluted with chloroform. The solvent was removed under reduced pressure to give nitroenamine **1a** (150 mg, 93%). Other nitroenamines **1b–f** were prepared in the same way by using the corresponding amines instead of propylamine (Scheme 5).



**Scheme 4** Conversion of an alkylamino group to an amino group in the nitroenamine moiety.







Scheme 5 Outline of the preparative method for nitroenamines **1a–f**.

#### 4-Aza-2-nitro-2-heptenal (**1a**)<sup>18</sup>

Brown solid (*E/Z* = 76/24). (*E*)-Isomer; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.01 (t, *J* = 7.4 Hz, 3H), 1.74 (tq, *J* = 7.3, 7.4 Hz, 2H), 3.45 (dt, *J* = 6.8, 7.3 Hz, 2H), 8.48 (dd, *J* = 3.6, 14.5 Hz, 1H), 10.14 (d, *J* = 3.6 Hz, 1H), 10.3–10.8 (br, 1H). (*Z*)-Isomer; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.01 (t, *J* = 7.4 Hz, 3H), 1.76 (tq, *J* = 7.3, 7.4 Hz, 2H), 3.53 (dt, *J* = 6.8, 7.3 Hz, 2H), 8.48 (d, *J* = 15.3 Hz, 1H), 9.5–9.9 (br, 1H), 10.05 (s, 1H).

#### 4-Aza-5-methyl-2-nitro-2-hexenal (**1b**)<sup>19</sup>

Brown solid (*E/Z* = 76/24). (*E*)-Isomer; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.40 (d, *J* = 6.6 Hz, 6H), 3.75 (sep, *J* = 6.6 Hz, 1H), 8.52 (ddd, *J* = 0.6, 3.6, 14.6 Hz, 1H), 10.15 (d, *J* = 3.6 Hz, 1H), 10.3–10.8 (br, 1H). (*Z*)-Isomer; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.42 (d, *J* = 6.6 Hz, 6H), 3.84 (sep, *J* = 6.6 Hz, 2H), 7.95 (d, *J* = 15.4 Hz, 1H), 9.4–9.9 (br, 1H), 10.56 (s, 1H).

#### 4-Aza-5,5-dimethyl-2-nitro-2-hexenal (**1c**)<sup>18</sup>

White solid (*E/Z* = 79/21). (*E*)-Isomer; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.45 (s, 9H), 8.57 (dd, *J* = 3.6, 14.9 Hz, 1H), 10.15 (d, *J* = 3.6 Hz, 1H), 10.3–10.8 (br, 1H). (*Z*)-Isomer; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.47 (s, 9H), 7.99 (d, *J* = 15.7 Hz, 1H), 9.7–10.1 (br, 1H), 9.94 (s, 1H).

#### 4-Aza-2-nitro-5-phenyl-2-pentenal (**1d**)

White solid; mp 119.2–120.0 °C (*E/Z* = 77/23). (*E*)-Isomer; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 4.64 (d, *J* = 5.9 Hz, 2H), 7.2–7.5 (m, 5H), 8.54 (dd, *J* = 3.6, 14.3 Hz, 1H), 10.16 (d, *J* = 3.6 Hz, 1H), 10.6–11.0 (br, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 54.5 (CH<sub>2</sub>), 126.3 (C), 127.8 (CH), 129.1 (CH), 129.5 (CH), 133.9 (CH), 155.0 (C), 186.4 (CH). (*Z*)-Isomer; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 4.71 (d, *J* = 6.0 Hz, 2H), 7.2–7.5 (m, 5H), 7.98 (d, *J* = 15.1 Hz, 1H), 9.7–10.0 (br, 1H), 10.07 (s, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 54.7 (CH<sub>2</sub>), 125.1 (C), 127.9 (CH), 129.2 (CH), 129.5 (CH), 133.9 (CH), 150.3 (C), 183.4 (CH). HRMS (ESI/TOF) calcd for C<sub>10</sub>H<sub>10</sub>N<sub>2</sub>O<sub>3</sub>(M – H<sup>+</sup>): 205.0619, found: 205.0624; IR (KBr, cm<sup>−1</sup>) 3212, 1656, 1606, 1505, 1317.

#### 4-Aza-2-nitro-2,6-pentadienal (**1e**)<sup>5</sup>

Yellow oil (*E/Z* = 78/22). (*E*)-Isomer; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 4.09 (br dd, *J* = 5.7, 5.7 Hz, 2H), 5.3–5.5 (m, 2H), 5.90 (ddt, *J* = 5.7, 11.2, 17.0 Hz, 1H), 8.48 (dd, *J* = 3.6, 14.4 Hz, 1H), 10.15 (d, *J* = 3.6 Hz, 1H), 10.3–10.8 (br, 1H). (*Z*)-Isomer; <sup>1</sup>H NMR

(400 MHz, CDCl<sub>3</sub>) δ 4.16 (br dd, *J* = 5.8, 5.8 Hz, 2H), 5.3–5.5 (m, 2H), 5.8–6.0 (m, 1H, overlapped with signals of *E*-isomer), 7.91 (d, *J* = 15.2 Hz, 1H), 9.5–9.9 (br, 1H), 10.06 (s, 1H).

#### 4-Aza-6-hydroxy-2-nitro-2-hexenal (**1f**)

Brown solid; mp 125.4–126.5 °C (*E/Z* = 71/29). (*E*)-Isomer; <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN) δ 3.1–3.4 (br, 1H), 3.5–3.6 (m, 2H), 3.70 (t, *J* = 5.2 Hz, 2H), 8.58 (br d, *J* = 8.8 Hz, 1H), 10.10 (d, *J* = 3.6 Hz, 1H), 10.3–10.7 (br, 1H); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>CN) δ 53.3 (CH<sub>2</sub>), 60.8 (CH<sub>2</sub>), 126.8 (C), 157.4 (CH), 186.3 (CH). (*Z*)-Isomer; <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN) δ 3.2–3.4 (br, 1H), 3.62 (t, *J* = 4.7 Hz, 2H), 3.6–3.8 (m, 2H), 7.96 (d, *J* = 15.2 Hz, 1H), 9.5–9.9 (br, 1H), 9.98 (s, 1H); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>CN) δ 53.4 (CH<sub>2</sub>), 60.8 (CH<sub>2</sub>), 125.6 (C), 152.7 (CH), 183.8 (CH). HRMS (ESI/TOF) calcd for C<sub>5</sub>H<sub>8</sub>N<sub>2</sub>O<sub>4</sub>(M – H<sup>+</sup>): 159.0411, found: 159.0416; IR (KBr, cm<sup>−1</sup>) 3494, 3216, 1674, 1589, 1491, 1314.

#### Synthesis of triazabicyclic compounds

To a solution of formylnitroenamine **1a** (158.2 mg, 1.0 mmol) in ethanol (4 mL), ammonium acetate (78.5 mg, 1.0 mmol) was added, and the resultant mixture was heated at 50 °C for 1 day. After the removal of the solvent under reduced pressure, the residual brown solid was dissolved in dichloromethane (30 mL) and washed with water (30 mL × 1). The aqueous layer was extracted with dichloromethane (30 mL × 3). The combined organic layer was dried over magnesium sulfate and evaporated to afford a brown solid as a residue. The solid was subjected to column chromatography on silica gel using ethyl acetate/dichloromethane (1/9) as an eluent to afford a mixture of **2a** and **3a** as a yellow solid (81.0 mg, 0.27 mmol, 55%) and a mixture of **3a** and **4a** as a brown solid (17.9 mg, 0.07 mmol, 14%). However, further separation could not be achieved. When other nitroenamines **1b–f** were used, the experiments were performed in the same way. In the cases of **1b** and **1f**, ethyl acetate was used as an extraction solvent instead of dichloromethane. In the case of **1f**, ethyl acetate was used as an eluent for column chromatography.

The NMR spectra were analyzed using a mixture of **2** and **3** because these products are interconvertible under equilibrium when dissolved in solvents. Hence, the content ratio varied in each measurement. With regard to bicyclic products **3** and **4**, the NMR spectra were measured using a mixture because they were inseparable despite several attempts. The yields of **3** and **4** were determined on the basis of the integral ratio.

#### 2,6,9-Triaza-4,8-dinitro-2,6-dipropylbicyclo[3.3.1]nona-3,7-diene (**2a**) and 2,6,9-triaza-4,8-dinitro-2,9-dipropylbicyclo[3.3.1]nona-3,7-diene (**3a**)

Yellow solid, mp 152.5–153.3 °C, *R*<sub>f</sub> = 0.43 (SiO<sub>2</sub>, dichloromethane/methanol = 20/1). **2a**; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 0.98 (t, *J* = 7.4 Hz, 6H), 1.6–1.9 (m, 4H), 2.2–2.3 (br, 1H), 3.43 (ddd, *J* = 7.7, 7.7, 13.9 Hz, 2H), 3.98 (ddd, *J* = 5.8, 7.8, 13.9 Hz, 2H), 5.68 (d, *J* = 2.8 Hz, 2H), 8.20 (s, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 11.1 (CH<sub>3</sub>), 23.3 (CH<sub>2</sub>), 56.9 (CH<sub>2</sub>), 61.9 (CH), 122.4 (C), 144.6 (CH). **3a**; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 0.93 (t, *J* =



7.4 Hz, 3H), 1.00 (t,  $J = 7.4$  Hz, 3H), 1.56 (ddq,  $J = 7.2, 7.2, 7.4$  Hz, 2H), 1.6–1.9 (m, 2H), 2.37 (dt,  $J = 7.2, 12.6$  Hz, 1H), 2.41 (dt,  $J = 7.4, 12.6$  Hz, 1H), 3.39 (ddd,  $J = 7.8, 7.8, 14.0$  Hz, 1H), 3.95 (ddd,  $J = 5.4, 8.1, 14.0$  Hz, 1H), 5.34 (d,  $J = 1.4$  Hz, 1H), 5.32 (s, 1H), 7.0–7.2 (br, 1H), 8.16 (s, 1H), 8.29 (d,  $J = 4.7, 1H$ );  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  11.1 ( $\text{CH}_3$ ), 11.6 ( $\text{CH}_3$ ), 20.8 ( $\text{CH}_2$ ), 23.2 ( $\text{CH}_2$ ), 51.4 ( $\text{CH}_2$ ), 56.8 ( $\text{CH}_2$ ), 61.3 (CH), 66.9 (CH), 122.2 (C), 122.7 (C), 142.2 (CH), 144.4 (CH). HRMS (ESI/TOF) calcd for  $\text{C}_{12}\text{H}_{19}\text{N}_5\text{O}_4$  ( $\text{M} + \text{H}^+$ ): 298.1510, found: 298.1515; IR (KBr,  $\text{cm}^{-1}$ ) 3291, 2965, 1613, 1304, 1226. A single crystal of **2a** was obtained as yellow needles by recrystallization from dichloromethane/hexane.

**2,6,9-Triaza-4,8-dinitro-2-propylbicyclo[3.3.1]nona-3,7-diene (4a) and 2,6,9-triaza-4,8-dinitro-9-propylbicyclo[3.3.1]nona-3,7-diene (5a)**

Yellow solid, mp 91.5–93.6 °C (**4a/5a** = 89/11). **4a**;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  0.92 (t,  $J = 7.4$  Hz, 3H), 1.6–1.9 (m, 2H), 2.7–2.9 (br, 1H), 3.45 (ddd,  $J = 7.7, 7.7, 13.8$  Hz, 1H), 3.87 (ddd,  $J = 5.5, 7.8, 13.8$  Hz, 1H), 5.57 (d,  $J = 1.6$  Hz, 1H), 5.68 (d,  $J = 2.0$  Hz, 1H), 7.6–8.1 (br, 1H), 8.20 (s, 1H), 8.22 (s, 1H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  11.1 ( $\text{CH}_3$ ), 23.6 ( $\text{CH}_2$ ), 55.6 ( $\text{CH}_2$ ), 55.7 (CH), 61.5 (CH), 124.0 (C), 124.3 (C), 143.5 (CH), 145.35 (CH). **5a**;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  0.87 (t,  $J = 7.4$  Hz, 3H), 1.51 (ddq,  $J = 6.7, 6.8, 7.4$  Hz, 2H), 2.34 (dt,  $J = 6.7, 14.7$  Hz, 1H), 2.36 (dt,  $J = 6.8, 14.7$  Hz, 1H), 5.33 (s, 2H), 7.6–8.1 (br, 2H), 8.22 (s, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  11.7 ( $\text{CH}_3$ ), 21.2 ( $\text{CH}_2$ ), 51.3 ( $\text{CH}_2$ ), 62.0 (CH), 124.0 (C), 142.8 (CH). HRMS (ESI/TOF) calcd for  $\text{C}_9\text{H}_{13}\text{N}_5\text{O}_4$  ( $\text{M} - \text{H}^+$ ): 254.0895, found: 254.0890; IR (KBr,  $\text{cm}^{-1}$ ) 3283, 2965, 1611, 1340, 1225, 1139, 926.

**2,6,9-Triaza-2,6-di(2-methylethyl)-4,8-dinitrobicyclo[3.3.1]nona-3,7-diene (2b) and 2,6,9-triaza-2,9-di(2-methylethyl)-4,8-dinitrobicyclo[3.3.1]nona-3,7-diene (3b)**

Yellow solid, mp 160.6–162.1 °C. Signals of **3b** were too small to be assigned. **2b**;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  1.36 (d,  $J = 6.6$  Hz, 12H), 2.1–2.3 (br, 1H), 4.36 (sep,  $J = 6.6$  Hz, 2H), 5.69 (d,  $J = 2.7$  Hz, 2H), 8.27 (s, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  21.3 ( $\text{CH}_3$ ), 24.2 ( $\text{CH}_3$ ), 55.5 (CH), 63.4 (CH), 122.7 (C), 140.9 (CH); HRMS (ESI/TOF) calcd for  $\text{C}_{12}\text{H}_{18}\text{N}_5\text{O}_4$  ( $\text{M} - \text{H}^+$ ): 296.1364, found: 296.1375; IR (KBr,  $\text{cm}^{-1}$ ) 3291, 2973, 1609, 1338, 1259, 1230, 1181.

**2,6,9-Triaza-2-(2-methylethyl)-4,8-dinitrobicyclo[3.3.1]nona-3,7-diene (4b)**

Brown solid, mp 126.2–127.9 °C. Signals of **5b** were too small to be assigned.  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  1.30 (d,  $J = 6.6$  Hz, 3H), 1.33 (d,  $J = 6.6$  Hz, 3H), 2.7–2.9 (br, 1H), 4.24 (qq,  $J = 6.6, 6.6$  Hz, 1H), 5.57 (d,  $J = 1.1$  Hz, 1H), 5.70 (d,  $J = 1.2$  Hz, 1H), 7.7–8.0 (br, 1H), 8.21 (s, 1H), 8.26 (s, 1H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  21.2 ( $\text{CH}_3$ ), 23.9 ( $\text{CH}_3$ ), 56.2 (CH), 57.4 (CH), 64.4 (CH), 124.1 (C), 124.9 (C), 141.4 (CH), 143.4 (CH); HRMS (ESI/TOF) calcd for  $\text{C}_9\text{H}_{12}\text{N}_5\text{O}_4$  ( $\text{M} - \text{H}^+$ ): 254.0895, found: 254.0898; IR (KBr,  $\text{cm}^{-1}$ ) 3283, 1609, 1342, 1232.

**2,6,9-Triaza-4,8-dinitro[3.3.1]nona-3,7-diene (6)**

Yellow solid, mp 207.0–208.1 °C (dec.).  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  3.72 (t,  $J = 2.7$  Hz, 1H), 5.51 (d,  $J = 2.7$  Hz, 2H), 8.24 (s, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{DMSO}-d_6$ )  $\delta$  56.2 (CH), 123.1 (C), 142.8 (CH); HRMS (ESI/TOF) calcd for  $\text{C}_6\text{H}_7\text{N}_5\text{O}_4$  ( $\text{M} - \text{H}^+$ ): 212.0425, found: 212.0427; IR (KBr,  $\text{cm}^{-1}$ ) 3166, 3012, 1607, 1353, 1223.

**2,6,9-Triaza-2,6-dibenzyl-4,8-dinitrobicyclo[3.3.1]nona-3,7-diene (2d) and 2,6,9-triaza-2,9-dibenzyl-4,8-dinitrobicyclo[3.3.1]nona-3,7-diene (3d)**

Brown solid, mp 155.8–157.5 °C,  $R_f = 0.29$  ( $\text{SiO}_2$ , hexane/ethyl acetate/triethylamine = 6/4/0.5). **2d**;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  1.9–2.1 (br, 1H), 4.68 (d,  $J = 14.8$  Hz, 2H), 5.08 (d,  $J = 14.8$  Hz, 2H), 5.65 (d,  $J = 2.6$  Hz, 2H), 6.9–7.5 (m, 10H), 8.26 (s, 2H). **3d**;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  3.29 (d,  $J = 13.4$  Hz, 1H), 3.35 (d,  $J = 13.4$  Hz, 1H), 4.63 (d,  $J = 14.7$  Hz, 1H), 4.99 (d,  $J = 14.7$  Hz, 1H), 5.24 (d,  $J = 1.3$  Hz, 1H), 5.35 (d,  $J = 1.3$  Hz, 1H), 6.9–7.5 (m, 10H), 8.30 (s, 1H), 8.38 (s, 1H). A signal of N–H was absent, which was presumably due to overlapping. The  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ) measurement was performed using a mixture of two isomers **2d** and **3d**.  $\delta$  53.7 ( $\text{CH}_2$ ), 59.1 ( $\text{CH}_2$ ), 59.1 ( $\text{CH}_2$ ), 60.7 (CH), 61.1 (CH), 65.8 (CH), 121.8 (C), 122.8 (C), 123.0 (C), 128.2 (CH), 128.4 (CH), 128.6 (CH), 128.7 (CH), 128.9 (CH), 129.0 (CH), 129.0 (CH), 129.4 (CH), 129.4 (CH), 134.5 (C), 134.7 (C), 135.0 (C), 142.8 (CH), 144.6 (CH), 144.8 (CH); HRMS (ESI/TOF) calcd for  $\text{C}_{20}\text{H}_{19}\text{N}_5\text{O}_4$  ( $\text{M} + \text{H}^+$ ): 394.1510, found: 394.1525; IR (KBr,  $\text{cm}^{-1}$ ) 3279, 3057, 1614, 1318, 1209. A single crystal of **3d** was obtained as yellow blocks by recrystallization from ethanol/diethyl ether.

**2,6,9-Triaza-2-benzyl-4,8-dinitrobicyclo[3.3.1]nona-3,7-diene (4d) and 2,6,9-triaza-9-benzyl-4,8-dinitrobicyclo[3.3.1]nona-3,7-diene (5d)**

Yellow solid, mp 155.0–155.8 °C. **4d**;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  2.6–2.8 (br, 1H), 4.78 (d,  $J = 15.0$  Hz, 1H), 4.99 (d,  $J = 15.0$  Hz, 1H), 5.56 (dd,  $J = 1.0, 2.7$  Hz, 1H), 5.63 (d,  $J = 2.7$  Hz, 1H), 7.1–7.5 (m, 5H), 8.24 (s, 1H), 8.26 (s, 1H). **5d**;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  3.54 (d,  $J = 13.3$  Hz, 1H), 3.59 (d,  $J = 13.3$  Hz, 1H), 5.25 (s, 2H), 7.1–7.5 (m, 5H), 8.27 (s, 2H). A signal of N–H was absent, which was presumably due to overlapping. The  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{CN}$ ) measurement was performed using a mixture of two isomers **4d** and **5d**.  $\delta$  53.7 ( $\text{CH}_2$ ), 57.1 (CH), 58.9 ( $\text{CH}_2$ ), 61.5 (CH), 62.5 (CH), 123.8 (C), 124.1 (C), 124.9 (C), 128.8 (CH), 129.2 (CH), 129.3 (CH), 129.6 (CH), 129.9 (CH), 130.0 (CH), 136.9 (C), 137.4 (C), 142.9 (CH), 143.5 (CH), 145.3 (CH); HRMS (ESI/TOF) calcd for  $\text{C}_{13}\text{H}_{13}\text{N}_5\text{O}_4$  ( $\text{M} - \text{H}^+$ ): 302.0895, found: 302.0895; IR (KBr,  $\text{cm}^{-1}$ ) 3276, 1705, 1610, 1320, 1204, 928.

**2,6,9-Triaza-4,8-dinitro-2,6-di(3-propen-1-yl)bicyclo[3.3.1]nona-3,7-diene (2e) and 2,6,9-triaza-4,8-dinitro-2,9-di(3-propen-1-yl)bicyclo[3.3.1]nona-3,7-diene (3e)**

Brown solid, mp 60.5–62.0 °C. **2e**;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.20 (br, 1H), 4.17 (dddd,  $J = 1.5, 1.5, 5.0, 15.4$  Hz, 2H), 3.60



(br dd,  $J = 7.1, 15.4$  Hz, 2H), 5.82 (dddd,  $J = 1.2, 1.2, 1.2, 17.2$  Hz, 2H), 5.3–5.5 (m, 2H), 5.69 (d,  $J = 2.8$  Hz, 2H), 5.87 (dddd,  $J = 5.0, 7.1, 10.2, 17.2$  Hz, 2H), 8.18 (s, 2H). **3e**;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  3.01 (dddd,  $J = 1.2, 1.2, 6.4, 13.5$  Hz, 1H), 3.08 (dddd,  $J = 1.2, 1.2, 6.3, 13.5$  Hz, 1H), 4.13 (dddd,  $J = 1.3, 1.3, 5.1, 15.3$  Hz, 1H), 4.44 (br dd,  $J = 7.2, 15.3$  Hz, 1H), 5.26 (dddd,  $J = 1.2, 1.2, 1.2, 17.1$  Hz, 1H), 5.29 (dddd,  $J = 1.3, 1.3, 1.3, 10.2$  Hz, 1H), 5.3–5.5 (m, 2H), 5.47 (dddd,  $J = 1.3, 1.3, 1.3, 17.0$  Hz, 1H), 5.34 (d,  $J = 1.5$  Hz, 1H), 5.38 (d,  $J = 1.5$  Hz, 1H), 5.82 (dddd,  $J = 6.3, 6.4, 10.2, 17.1$  Hz, 1H), 5.86 (dddd,  $J = 5.1, 7.2, 10.2, 17.0$  Hz, 1H), 6.2–6.8 (br, 1H), 8.18 (s, 1H), 8.30 (s, 1H). The  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ) measurement was performed using a mixture of two isomers **2e** and **3e**.  $\delta$  52.6 ( $\text{CH}_2$ ), 57.5 ( $\text{CH}_2$ ), 57.5 ( $\text{CH}_2$ ), 60.7 (CH), 61.7 (CH), 66.2 (CH), 120.6 ( $\text{CH}_2$ ), 121.1 ( $\text{CH}_2$ ), 121.4 ( $\text{CH}_2$ ), 121.8 (C), 122.8 (C), 122.9 (C), 131.5 (CH), 131.7 (CH), 132.4 (CH), 142.8 (CH), 144.3 (CH), 144.6 (CH); HRMS (ESI/TOF) calcd for  $\text{C}_{12}\text{H}_{15}\text{N}_5\text{O}_4$  ( $\text{M} - \text{H}^+$ ): 292.1051, found: 292.1054; IR (KBr,  $\text{cm}^{-1}$ ) 3283, 1613, 1482, 1416, 1309, 920, 760.

**2,6,9-Triaza-4,8-dinitro-2-(3-propen-1-yl)bicyclo[3.3.1]nona-3,7-diene (4e) and 2,6,9-triaza-4,8-dinitro-9-(3-propen-1-yl)bicyclo[3.3.1]nona-3,7-diene (5e)**

Yellow solid, mp 135.4–137.5 °C. **4e**;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  2.7–2.9 (br, 1H), 4.19 (dddd,  $J = 1.4, 1.4, 5.0, 15.6$  Hz, 1H), 4.43 (dddd,  $J = 1.4, 1.4, 6.8, 15.6$  Hz, 1H), 5.30 (dddd,  $J = 1.4, 1.4, 1.4, 10.2$  Hz, 1H), 5.39 (dddd,  $J = 1.4, 1.4, 1.4, 17.1$  Hz, 1H), 5.57 (dd,  $J = 1.1, 2.7$  Hz, 1H), 5.65 (br d,  $J = 2.8$  Hz, 1H), 5.94 (dddd,  $J = 5.0, 6.9, 10.2, 17.1$  Hz, 1H), 7.5–8.1 (br, 1H), 8.18 (s, 1H), 8.22 (s, 1H). **5e**;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  3.00 (dddd,  $J = 1.3, 1.3, 6.4, 13.7$  Hz, 1H), 3.05 (dddd,  $J = 1.3, 1.3, 6.4, 13.7$  Hz, 1H), 5.30 (dddd,  $J = 1.3, 1.3, 1.3, 10.2$  Hz, 1H), 5.39 (dddd,  $J = 1.3, 1.3, 1.3, 17.1$  Hz, 1H), 5.33 (s, 2H), 5.85 (dddd,  $J = 6.4, 6.4, 10.2, 17.1$  Hz, 1H), 7.5–8.1 (br, 2H), 8.23 (s, 2H). The  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{CN}$ ) measurement was performed using a mixture of two isomers **4e** and **5e**.  $\delta$  52.6 ( $\text{CH}_2$ ), 57.1 (CH), 57.7 ( $\text{CH}_2$ ), 61.5 (CH), 62.8 (CH), 119.9 ( $\text{CH}_2$ ), 119.9 ( $\text{CH}_2$ ), 123.8 (C), 124.1 (C), 124.9 (C), 134.0 (CH), 134.4 (CH), 142.8 (CH), 143.4 (CH), 145.1 (CH); HRMS (ESI/TOF) calcd for  $\text{C}_9\text{H}_{11}\text{N}_5\text{O}_4$  ( $\text{M} - \text{H}^+$ ): 252.0738, found: 252.0740; IR (KBr,  $\text{cm}^{-1}$ ) 3278, 1610, 1423, 1334, 1215, 927.

**2,6,9-Triaza-2,6-di(2-hydroxyethyl)-4,8-dinitro[3.3.1]nona-3,7-diene (2f) and 2,6,9-triaza-2,9-di(2-hydroxyethyl)-4,8-dinitro[3.3.1]nona-3,7-diene (3f)**

Brown solid, mp 68.1–70.7 °C. **2f**;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  2.8–3.0 (br, 1H), 3.0–3.3 (br, 2H), 3.44 (ddd,  $J = 3.8, 7.0, 14.2$  Hz, 2H), 3.69 (ddd,  $J = 3.8, 6.0, 11.8$  Hz, 2H), 3.77 (ddd,  $J = 3.4, 7.0, 11.8$  Hz, 2H), 4.01 (ddd,  $J = 3.4, 6.0, 14.2$  Hz, 2H), 5.71 (d,  $J = 1.0$  Hz, 2H), 8.24 (s, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  57.4 ( $\text{CH}_2$ ), 61.5 ( $\text{CH}_2$ ), 63.3 (CH), 123.5 (C), 144.6 (CH). **3f**;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  2.49 (ddd,  $J = 5.2, 5.2, 13.7$  Hz, 1H), 2.64 (ddd,  $J = 5.0, 5.0, 13.7$  Hz, 1H), 2.9–3.1 (br, 1H), 3.0–3.3 (br, 1H), 3.5–3.8 (m, 5H, overlapped with signals of **3f**), 3.97 (ddd,  $J = 3.3, 6.0, 14.7$  Hz, 1H), 5.42 (d,  $J = 1.5$  Hz, 1H), 5.61 (d,  $J = 1.5$  Hz, 1H), 7.5–8.2 (br, 1H), 8.26 (s, 1H), 8.27 (s, 1H);  $^{13}\text{C}$

NMR (100 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  51.9 ( $\text{CH}_2$ ), 57.3 ( $\text{CH}_2$ ), 60.7 ( $\text{CH}_2$ ), 61.4 ( $\text{CH}_2$ ), 62.1 (CH), 68.4 (CH), 122.9 (C), 123.1 (C), 143.4 (CH), 145.9 (CH). HRMS (ESI/TOF) calcd for  $\text{C}_{10}\text{H}_{15}\text{N}_5\text{O}_6$  ( $\text{M} - \text{H}^+$ ): 300.0950, found: 300.0954; IR (KBr,  $\text{cm}^{-1}$ ) 1209, 1307, 1612, 2359, 3307.

**2,6,9-Triaza-2-(2-hydroxyethyl)-4,8-dinitro[3.3.1]nona-3,7-diene (4f) and 2,6,9-triaza-9-(2-hydroxyethyl)-4,8-dinitro[3.3.1]nona-3,7-diene (5f)**

Brown solid, mp 95.6–97.2 °C. **4f**;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  2.8–3.0 (br, 1H), 2.9–3.4 (br, 1H), 3.57 (ddd,  $J = 3.8, 6.9, 14.3$  Hz, 1H), 3.67 (ddd,  $J = 3.8, 6.2, 11.6$  Hz, 1H), 3.77 (ddd,  $J = 3.5, 6.9, 11.6$  Hz, 1H), 3.99 (ddd,  $J = 3.5, 6.2, 14.3$  Hz, 1H), 5.57 (s, 1H), 5.73 (s, 1H), 7.4–8.2 (br, 1H), 8.21 (s, 1H), 8.23 (s, 1H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  57.0 ( $\text{CH}_2$ ), 57.2 (CH), 61.5 ( $\text{CH}_2$ ), 63.8 (CH), 124.0 (C), 124.7 (C), 143.5 (CH), 145.6 (CH). **5f**;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  2.49 (ddd,  $J = 5.5, 5.5, 13.4$  Hz, 1H), 2.55 (ddd,  $J = 5.1, 5.1, 13.4$  Hz, 1H), 2.9–3.4 (br, 1H), 3.5–3.7 (m, 2H, overlapped with signals of **4f**), 5.44 (s, 2H), 7.4–8.2 (br, 2H), 8.23 (s, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  51.9 ( $\text{CH}_2$ ), 60.7 ( $\text{CH}_2$ ), 62.5 (CH), 124.0 (C), 142.7 (CH). HRMS (ESI/TOF) calcd for  $\text{C}_8\text{H}_{11}\text{N}_5\text{O}_5$  ( $\text{M} - \text{H}^+$ ): 256.0687, found: 256.0685; IR (KBr,  $\text{cm}^{-1}$ ) 1208, 1226, 1317, 1609, 2359, 3251.

**X-Ray crystallographic analysis**

Diffraction data were collected at 93 or 123 K under a cold  $\text{N}_2$ -gas stream using a Rigaku XtaLAB Synergy-S/Mo system ( $\lambda = 0.71073$  Å (Mo-K $\alpha$ )). The integrated data were analyzed using the Olex2 crystallographic software package.<sup>20</sup> The structures were solved with the ShelXT structure solution program<sup>21</sup> using Intrinsic Phasing and refined with the ShelXL refinement package<sup>22</sup> using least-squares minimization. Anisotropic refinement was performed for all non-hydrogen atoms, and all the hydrogen atoms were placed at the calculated positions.

**Temperature dependent NMR spectra**

$^1\text{H}$  NMR spectra were obtained by using the variable-temperature (VT) NMR technique. The sample solutions were stored at –10–50 °C for over 12 h prior to the measurements.

**Theoretical calculations**

Density functional theory (DFT) calculations for **2a**, **2d**, **3a** and **3d** were conducted using Gaussian 16 W software (revision A.03)<sup>23</sup> using the restricted Becke three-parameter hybrid functionals with the Lee–Yang–Parr correlation (RB3LYP)<sup>24,25</sup> and 6-311+G(d,p) basis set.<sup>26,27</sup> Ground-state geometries were optimized and, then, the Gibbs energies of the molecules were calculated. Solvation was considered using the conductor-like polarizable continuum model.<sup>28,29</sup>

**Conflicts of interest**

There are no conflicts to declare.





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