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Solid-phase synthesis and evaluation of tumour-targeting phenylboronate-based MRI contrast agents†

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Paramagnetic macrocycles functionalized with phenylboronic moieties have proven to be interesting for MRI applications based on their ability to recognize cancer cells and generate local contrast. However, full use of the potential of this class of compounds is hampered by laborious and inefficient synthetic and, especially, purification procedures. The amphiphilic character of water-soluble phenylboronates renders them difficult compounds to be prepared through conventional solution synthesis due to the tendency to aggregate and form adducts with other nucleophiles. The new strategy described herein exploits the advantage of solid-phase synthesis with the application of DEAM-PS resin for anchorage and the subsequent simplified derivatization of boronates. GdDOTA-EN-PBA and its fluorinated analogue GdDOTA-EN-F2PBA were synthesized in a much easier, faster and economically convenient way to achieve good yields and purity. Furthermore, the effect of electron-withdrawing fluorine atoms on the aromatic ring of the latter compound was investigated by comparing the physico-chemical properties of both compounds as well as their binding affinity towards melanoma cancer cells.

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Introduction

Boronic acids are a class of compounds that keep attracting increasing interest in many fields. Their applications range from biomedical research to materials science, chemical synthesis and electronics.¹ For example, boronates have been successfully employed as enzymatic inhibitors,² in the synthesis of organogels,³ as chemosensors,⁴ and as binding sites in the preparation of electroconductive molecules,⁵ not to mention their use as synthetic partners in the Suzuki–Miyaura coupling reaction.⁶ Boron-10 is also the nuclide of choice for neutron capture therapy, an emerging anti-cancer treatment based on the use of thermal neutrons to destroy tumour tissues.⁷

Another interesting biomedical application of boron-based compounds lies in their capability for molecular recognition in

the design of targeting contrast agents. In particular, phenylboronic acid (PBA) and its derivatives have shown the ability to strongly bind sialic acid, a nine-carbon monosaccharide unit that is overexpressed as the terminal group of glycolipids and glycoproteins on the surface of tumour cells.⁸ The mechanism of such interaction is based on the reversible formation of five- and six-membered cyclic boronate esters between the boronic group of PBA and the exocyclic polyol chain of sialic acid.⁹ This property of PBAs has been exploited in the preparation of several diagnostic and/or therapeutic targeting agents,^{10–12} the most successful of them being GdDOTA-EN-PBA (DOTA = 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid, EN = ethylenediamine). Such a Gd(III)-complex bears a PBA moiety conjugated to a ligand (DOTA) through an aminoethylamide linker (EN) (Scheme 1). GdDOTA-EN-PBA has proved to be successful as a magnetic resonance imaging (MRI) contrast agent for *in vivo* tumour targeting based on the recognition of overexpressed sialic acid.¹³ In this case, a two-site cooperative binding mechanism is involved: in addition to the previously mentioned formation of cyclic boronates between PBA and the diol of the substrate, the recognition is enhanced by an electrostatic interaction involving the negatively charged carboxylate on the sialic acid residue and the positively charged amino group of the linker between the macrocycle and PBA. The same strategy was applied later with the radioactive Ga-68 complex of DOTA-EN-PBA enabling visualization of tumours by positron emission tomo-

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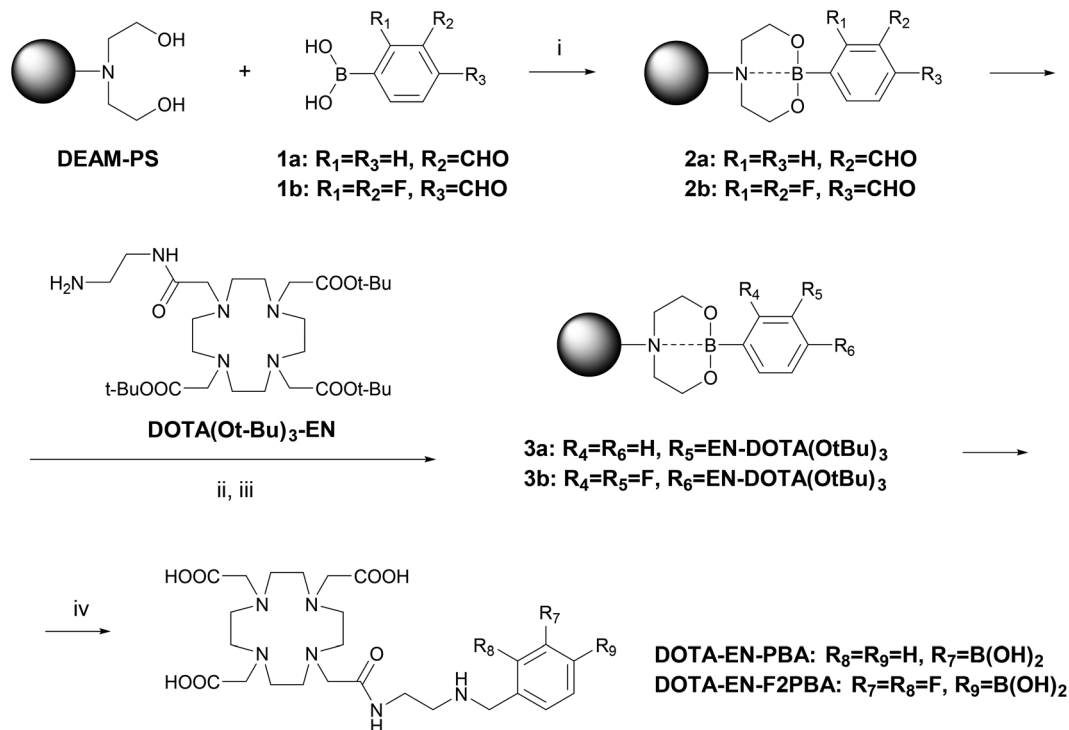
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Scheme 1 Solid-phase synthesis of DOTA-EN-PBA and DOTA-EN-F₂PBA. (i) THF, rt, 2 h; (ii) THF, rt, 3 h; (iii) NaBH₄, THF, rt, 4 h; and (iv) 1 : 1 DCM/TFA, rt, 15 h.

graphy (PET).^{11,14} Additionally, Yamamoto *et al.* have demonstrated the potential of a PBA-containing GdDTPA-based MRI contrast agent (DTPA = diethylenetriaminepentaacetic acid) for simultaneous neutron capture therapy, thus widening the medical applications of such systems.¹⁵

Unfortunately, the amphiphilic character of boronates translates into severe difficulties in their preparation and purification, including solubility problems and the inclination to form oligomers or react with alcohols, amines, carboxylates and other nucleophilic species. Also, the pH-dependent equilibrium between a boronic acid and its boronate form represents an additional source of synthetic obstacles.¹⁶ For instance, the conventional synthesis of DOTA-EN-PBA was heavily affected in terms of the final yield and the time and effort required to purify the ligand.¹⁰ As an example of tedious procedures needed for the purification of PBA derivatives, time-consuming ion-exchange chromatography was also responsible for partial loss of the product.

In order to prevent the boronic acid moiety from affecting the efficacy of the various synthetic steps and the purification of the intermediate products, the boronate can be “protected” by binding it to a solid support. This would also allow the exploitation of the well-known advantages of solid-phase synthesis, such as the easy removal of excess reagents and by-products through simple filtration and washing. A few cases have been already described where conventional solid-phase peptide synthesis was exploited to build molecular parts of contrast agents.^{17–20} Several solid supports are reported in the literature to specifically bind boronic acids, including citronellic acid

attached to Rink's amide resin, polystyrene derivatised with 2-methyl-2-(hydroxymethyl)-1,3-propanediol,²¹ catechol pendant polystyrene polymer,²² and more recently, 1-glycerol Merrifield resin.²³ Among these, polystyrene functionalized with diethanolamine groups (DEAM-PS resin) seems to be endowed with more effective binding properties, and requires mild conditions to release the boronic acids after their derivatization.^{24–26}

Herein, we report a new way to prepare DOTA-EN-PBA derivatives based on solid-phase synthesis. We were also interested in investigating the effect of the increased acidity of the boronate moiety on the binding to sialic acid. Therefore, an additional fluorinated ligand (DOTA-EN-F₂PBA) was synthesised *via* the same successful procedure. Such a strategy allowed both compounds to be obtained in a more straightforward way and with higher yields compared to those reported previously for DOTA-EN-PBA. Gd(III)-complexes of both species were then prepared and their relaxometric properties were determined through standard procedures. Finally, the targeting ability of novel GdDOTA-EN-F₂PBA towards sialic acid was studied on suitable cell cultures and compared to that of GdDOTA-EN-PBA.

Results and discussion

Synthetic procedures

DEAM-PS resin was synthesized according to the procedure described by Gravel *et al.*²⁵ starting from chloromethylated polystyrene (Merrifield) resin and diethanolamine. Generally, a



functionalized boronic acid can be anchored to the resin by simply dissolving it in a suspension of DEAM-PS and shaking the reaction vessel for a short time at room temperature.^{25,26} The synthesis can then be continued in a divergent way analogously to common solid-phase peptide synthesis until the final cleavage is performed. The novel preparation of DOTA-EN-PBA in a few simple steps was therefore carried out accordingly, as shown in Scheme 1.

After swelling the DEAM-PS resin in dry tetrahydrofuran for 30 min, 3-formylphenylboronic acid (**1a**) dissolved in anhydrous THF was first immobilized onto the resin (**2a**) by agitation in a filter syringe fixed onto an orbital shaker (50 rpm) for 2 hours at room temperature. This step was followed by reductive amination under analogous conditions (anhydrous THF, ambient temperature) between the aldehyde moiety and the primary amine of DOTA(OtBu)₃-EN to form the anchored and protected precursor (**3a**) of the aimed ligand. This was obtained in one step by shaking the functionalized resin in dichloromethane and trifluoroacetic acid to achieve deprotection of the carboxylates and cleavage from the resin at the same time.

The yield calculated (as the trifluoroacetate form) with respect to DOTA(OtBu)₃-EN was 62%, which definitely represents an improvement when compared to that reported in the literature for the synthesis in solution (33%).¹¹ Following this protocol, all the solubility problems typical of PBA derivatives were avoided. The identity of DOTA-EN-PBA was confirmed by ESI mass spectrometry and full NMR characterization (see the ESI†) by comparison with the data previously reported.¹¹ It is noteworthy that the main species observable in the mass spectrum of the ligand were the mono- and bis-dehydrated forms (Fig. S3 and S4†). By recording mass spectra at different values of cone voltage, these have been proved to be the consequence of fragmentation happening in the spectrometer and not of degradation prior to analysis; by decreasing the ionization voltage, the bis-dehydrated form progressively disappears while the mono-hydrated species increases (Fig. S4†).

Prompted by the positive outcome, the novel procedure was exploited to prepare a derivative of DOTA-EN-PBA bearing two electron-withdrawing fluorine atoms on the aromatic ring. Such a modification is expected to increase the acidity of the boronic acid,¹⁶ thus strengthening the binding to the sialic acid residues on the targeted tumour cells. Preliminarily, the effect of the F-atoms in different positions on the acidity of the boronic acid was tested by ¹¹B NMR titration measurements (see the ESI†) on four commercial model compounds: one resembling the position of DOTA-EN-PBA (namely, 3-methylphenylboronic acid) and three bearing the halogens and the B(OH)₂ groups in various sites of the aromatic ring [2,3-difluoro-4-methylphenylboronic acid [2,3-F-4-Me-PBA], 2,4-difluoro-5-methylphenylboronic acid [2,4-F-5-Me-PBA], 2,6-difluoro-5-methylphenylboronic acid [2,6-F-5-Me-PBA]]. The measurements (Fig. S1†) revealed a pK_a value of 7.2 for 2,3-F-4-Me-PBA, compared to 8.8 for 3-Me-PBA, 8.6 for 2,6-F-5-Me-PBA and 8.0 for 2,4-F-5-Me-PBA. Expecting an analogous two-

orders-of-magnitude increase in pK_a, a novel ligand (DOTA-EN-F2PBA) was designed with two fluorine atoms in the *ortho*- and *meta*-positions with respect to the attachment site of the pendant arm, while the boronic moiety was switched to the *para*-position. This would also allow the testing of the importance of the formation of the cyclic boronate in the recognition mechanism regardless of the additional contribution of the amine-carboxylate electrostatic interaction mentioned previously.

An analogous synthetic pathway was followed starting from 2,3-difluoro-4-formylphenylboronic acid (**1b**), which was first immobilized onto the DEAM-PS resin (**2b**), then conjugated to DOTA(OtBu)₃-EN (**3b**) and finally deprotected/cleaved. The newly formed DOTA-EN-F2PBA ligand (trifluoroacetate form) was obtained in 52% yield and fully characterized by ESI MS and NMR spectroscopy (Fig. S2 and S8–S11†). The water solubility of DOTA-EN-PBA and DOTA-EN-F2PBA represents an important extension with respect to the compounds prepared by Gravel *et al.* via solid-phase synthesis on the DEAM-PS resin, especially when the amphiphilic behaviour of boronates is considered.

In order to prove the effectiveness of such a protocol, the literature procedure for the preparation of DOTA-EN-PBA was applied to prepare DOTA-EN-F2PBA without a solid-phase synthetic strategy (see the Synthetic procedures section). In this case, the product was achieved in 24% yield, less than half compared to the new procedure.

Besides the final yield, another advantage of the resin-binding approach is clear when the purity of the compound after the deprotection step is considered: the ¹⁹F NMR spectrum of DOTA-EN-F2PBA prepared in solution (Fig. 1 and S12†) revealed the coexistence of two species bearing a fluorinated arene, which could not be observed by ¹H, ¹³C and ¹¹B NMR spectroscopy. In particular, in addition to the two doublets at −132 and −142 ppm from the expected product, two multiplets were also detected at −138 and −141 ppm, which furthermore gave cross-peaks in the ¹⁹F NMR COSY spectrum of the liquid-synthesis product (Fig. S13†). In con-

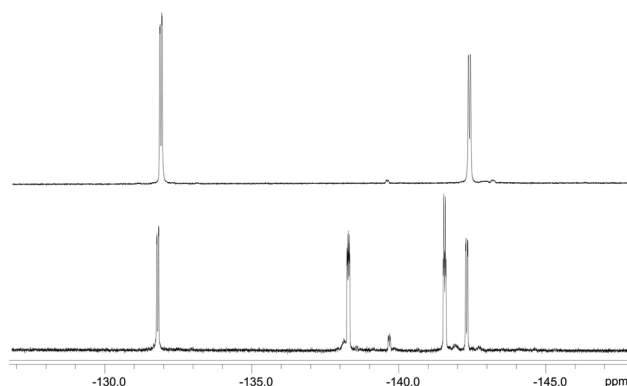


Fig. 1 Comparison between the ¹⁹F NMR spectra of DOTA-EN-F2PBA obtained after cleavage from the solid support (top) and recovered after the final deprotection step of the conventional preparation in solution (bottom).



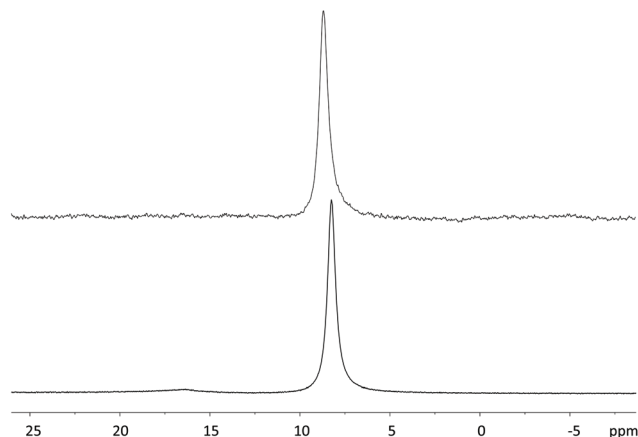


Fig. 2 Comparison between the ^{11}B NMR spectra of DOTA-EN-F2PBA prepared on the solid support (top) and in solution (bottom).

trast, this was not the case for DOTA-EN-F2PBA prepared through the novel procedure, whose ^{19}F NMR spectrum shows only one set of signals (Fig. 1 and S11†). The presence of contaminants in the commercial reactant (2,3-difluoro-4-formylphenylboronic acid) was excluded by checking its NMR spectra.

Since the ^{11}B NMR spectrum of the ligand prepared in solution only reports one resonance from the boronic moiety of the ligand (as compared to the solid-phase synthesis product) without additional signals from side-products (Fig. 2), it was concluded that the extra emerged species is the result of protodeboronation typical of phenylboronic acids with a fluorine in the *ortho*-position, as reported by Cox *et al.*²⁷

The presence of such a by-product was confirmed by both positive and negative ESI MS (Fig. S3†). The cleaner output further emphasizes the benefits of our novel solid-phase synthesis for the preparation of this kind of ligand: should such a side-reaction take place, the by-product would be released from the solid support and eliminated through successive washings.

Relaxometric characterization

Although GdDOTA-EN-PBA is reported in the literature, its relaxometric parameters have never been evaluated. Therefore, full relaxometric characterization of both GdDOTA-EN-PBA and GdDOTA-EN-F2PBA chelates was carried out. The complexes were prepared according to a standard procedure and their formation was preliminarily confirmed by mass spectrometry, including the distinctive Gd isotopic pattern (Fig. S5 and S6†). Then the ^1H nuclear magnetic relaxation dispersion (NMRD) profiles, in the proton Larmor frequency range of 0.01 to 70 MHz, were recorded at 25 and 37 °C (Fig. 3). The exact Gd(III) concentrations were determined according to the Evans method.²⁸

The NMRD profiles measured for the two complexes at 25 °C and 37 °C were analysed using the conventional Solomon–Bloembergen–Morgan theory,²⁹ leading to the determination of a set of characteristic parameters, such as the

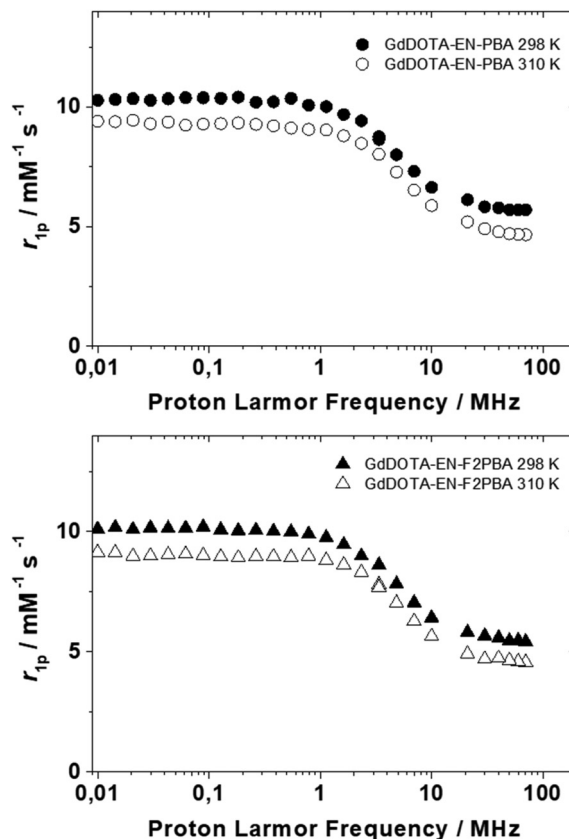


Fig. 3 ^1H NMRD profiles acquired at pH 7 and 25 or 37 °C for aqueous solutions of GdDOTA-EN-PBA (top) and GdDOTA-EN-F2PBA (bottom).

rotational correlation time (τ_R) and the exchange time (τ_M) of water coordinated to the metal centre with the bulk (Table 1). The longitudinal relaxivity values (r_{1p}), representing the proton relaxation rate enhancement per 1 mM Gd(III) ion, were determined at 25 °C and 20 MHz to be 6.24 and 5.80 $\text{mM}^{-1} \text{s}^{-1}$ for GdDOTA-EN-PBA and GdDOTA-EN-F2PBA, respectively. These values, slightly higher than those of typical monomeric GdDOTA complexes, can be attributed to the bigger molecular size due to the additional PBA pendant arm, which results in slower tumbling times as compared to GdDOTA ($\tau_R = 90 \text{ ps}$).³⁰ The increased relaxivity values are expected to translate into an enhanced MRI contrast effect.

As expected, the values for the two complexes are similar, as there is no particular difference of relaxometric interest in

Table 1 Selected parameters obtained from the analysis of the $1/T_1$ NMRD profiles at 298 K for GdDOTA-EN-PBA and GdDOTA-EN-F2PBA^a

Parameter	GdDOTA-EN-PBA	GdDOTA-EN-F2PBA
$^{20} \text{ MHz } r_{1p} [\text{mM}^{-1} \text{s}^{-1}]$	6.24 ± 0.21	5.80 ± 0.35
$\tau_M [\text{ns}]$	69 ± 19	100 ± 23
$\tau_R [\text{ps}]$	115 ± 5	112 ± 6
$\tau_v [\text{ps}]$	36 ± 5	40 ± 7
$\Delta^2 [10^{19} \text{s}^{-2}]$	1.48 ± 0.35	1.14 ± 0.36

^a The following parameters were fixed to common values during the fitting procedure: $r_{\text{Gd-H}} = 3.0 \text{ \AA}$, $D = 2.24 \times 10^{-5} \text{ cm}^2 \text{s}^{-1}$, $q = 1$, $a = 4.0 \text{ \AA}$.



their molecular structures. The slightly lower relaxivity of the fluorinated analogue suggests the influence of the rather remote aromatic group on the first coordination sphere of the complex, whereby a relative decrease in hydrophilicity induced by the fluorine atoms affects the water exchange rate at the Gd(III) ion (100 vs. 69 ns) and consequently the r_{1p} value.

Determination of the effect of pK_a on binding efficacy

As previously mentioned, one of the objectives of this work was to assess the importance of the acidity of the boronic group towards the binding strength regardless of the additional contribution arising from the electrostatic interaction between the amine of the linker and the carboxylate of sialic acid residues overexpressed by cancerous cells. In this perspective, switching the boronate from the *meta*- to *para*-position of the aromatic ring allows the effects of the lower pK_a to be highlighted (see above and the ESI†) in the recognition mechanism while at the same time reducing or removing the electrostatic influence. Since the expression of sialic acid is directly correlated to melanogenesis, the recognition ability of GdDOTA-EN-PBA and GdDOTA-EN-F2PBA was evaluated on murine melanoma B16-F10 cells. In fact, there is a direct correlation between sialic acid expression and the melanogenesis process occurring in these melanoma cells. Upon incubation of the cells in the presence of both Gd(III)-complexes, when the amount of the Gd(III) ion per cell was determined, comparable values were obtained (Fig. 4).

This seems to suggest that the advantageous effect of the electronegative substituents on increasing the acidity of the boronic group was counterbalanced by the shift of the latter from the *meta*-position in GdDOTA-EN-PBA to the *para*-position in the fluorinated derivative. As previously mentioned, because the amino group of DOTA-EN-PBA at physiological pH facilitates the recognition and binding to sialic acid residues by electrostatic interaction with the carboxylate of the glycan chain, when moving from a *meta*- to a *para*-substituted boronic acid, this interaction can be much less effective or even missed completely, thus weakening the targeting ability of the system. The

similar results obtained for GdDOTA-EN-PBA and GdDOTA-EN-F2PBA indicate that the contribution of the electrostatic interaction is basically comparable to the enhanced binding strength achieved by increasing the PBA acidity. This confirms therefore that, as we aimed to prove, playing with the acidity of the vector, *e.g.* by introducing electron-withdrawing groups on the aromatic ring, is a good strategy to improve the targeting abilities of this type of ligand.

Conclusions

The advantages of solid-phase synthesis were successfully exploited to prepare DOTA-EN-PBA and the novel DOTA-EN-F2PBA ligands, the latter bearing two fluorine substituents on the aromatic ring to increase the acidity of the boronic moiety. The positions of the electron-withdrawing groups and the B(OH)₂ groups were chosen upon investigation of the pK_a values of several model compounds by ¹¹B NMR titration, where the strongest acidity was assessed for a PBA derivative with two fluorines in the 2,3-positions and the boronate in the 4-position relatively to the linker. With work-up procedures mostly consisting of washing only, the yields of both compounds were doubled compared to the respective syntheses in solution. Moreover, the products appeared to be cleaner without the final by-products to separate. The relaxometric characterization of the GdDOTA-EN-PBA and GdDOTA-EN-F2PBA complexes allowed the determination of the longitudinal relaxivity values to be 6.24 and 5.80 mM⁻¹ s⁻¹, respectively; such r_{1p} values are higher than those commonly obtained for simple monomeric GdDOTA chelates and are the consequence of a bigger size (due to the PBA functionality) and fast water exchange rates ($\tau_M < 100$ ns). These relatively high relaxivities render GdDOTA-EN-PBA and GdDOTA-EN-F2PBA very promising for applications as targeting MRI contrast agents, as already reported.¹³ When the targeting abilities of the Gd(III)-complexes of the two ligands were tested towards sialic acid residues overexpressed by melanoma cells, a satisfactory response was obtained. However, GdDOTA-EN-F2PBA appeared to bind cancer cells similarly to GdDOTA-EN-PBA; this is due to the different position of the PBA group in the fluorinated agent (*para*) with respect to the non-fluorinated system (*meta*), which affects the secondary interaction between the amine of the ligand and the carboxylate of sialic acid, thus reducing the binding strength. This aspect, being one of the objectives of this work, has to be taken into consideration in the future design of this promising kind of MRI targeting contrast agent. The solid-state method proved to be suitable for the preparation of the targeted species with good yield and purity for applications as theranostic (therapeutic and diagnostic) agents.

Experimental

Materials and methods

DO3ArBu-N-(2-aminoethyl)ethanamide (DOTA(OtBu)₃-EN) was purchased from CheMatech. All chemicals were used as sup-

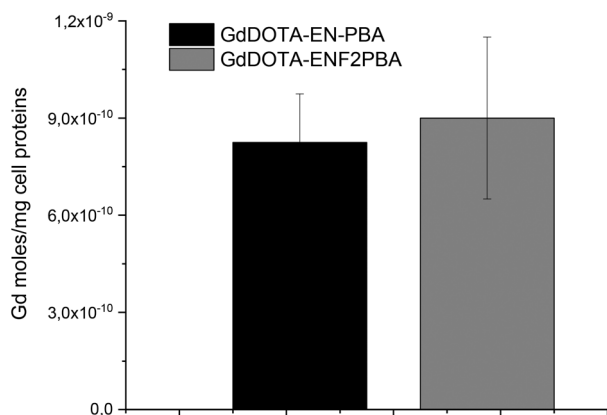


Fig. 4 Evaluation of the bound Gd(III)-complexes upon incubation with melanoma cells: B16-F10 cells, 4 h of incubation in EBSS buffer w/o glucose at a Gd concentration of 0.6 mM.



plied from commercial sources. "H₂O" refers to high-purity water with a conductivity of 0.04 $\mu\text{S cm}^{-1}$, obtained from a Milli-Q purification system. Solid-phase reactions were performed by orbital shaking (50 rpm) in polypropylene vessels (filter syringes) purchased from MultiSynTech GmbH. Resin-washing operations were carried out on a vortexer. Anhydrous THF was obtained by distillation of a commercial solvent over sodium/benzophenone. ¹H, ¹³C, ¹¹B and ¹⁹F NMR was performed on an Agilent MR400DD2 spectrometer operating at 9.4 T. Samples were prepared in 5 mm NMR tubes by dissolving the compounds in D₂O. Chemical shifts are reported in ppm relative to *tert*-BuOH (1.2 and 31.2 ppm for ¹H and ¹³C NMR, respectively), H₃BO₃ (0 ppm for ¹¹B NMR) and CF₃COOH (−74.4 ppm for ¹⁹F NMR) as the internal standards. For ¹¹B NMR experiments, a quartz tube was used. Coupling constants are reported in Hz. Splitting patterns are described as singlet (s), broad singlet (bs), double doublet (dd), multiplet (m) or broad multiplet (bm). Electrospray mass spectra were obtained on an ESI Waters SQD 3100 spectrometer for the low-resolution mass spectra and on a QSTAR Pulsar (AB/MDS Sciex) spectrometer for the high-resolution mass spectra (HR MS).

Synthetic procedures

Immobilization of boronic acids onto DEAM-PS (2a, 2b). DEAM-PS (0.54 mmol, theoretical loading 1.67 mmol g^{−1}, previously swollen by shaking its suspension in 5 mL of dry THF at room temperature for 30 min) and the appropriate PBA (1a or 1b, 0.81 mmol) were suspended in dry THF (6 mL) in a filter syringe and shaken at room temperature for 2 h. The vessel was drained and the functionalized resin was washed with THF (3 × 5 mL).

Reductive amination on DEAM-PS-supported formylphenylboronic acids (3a or 3b). Functionalized resin 2a or 2b (0.54 mmol, theoretical loading 1.67 mmol g^{−1}) was suspended in dry THF (6 mL), DOTA(OtBu)₃-EN (332 mg, 0.54 mmol) was added and the suspension was shaken at room temperature for 3 h. NaBH₄ (20 mg, 2.16 mmol) was added portion-wise to the reaction mixture and the resulting suspension was further agitated at room temperature for 4 h. After draining, the resin was washed with dry DMF (3 × 6 mL), dry DCM (5 × 6 mL) and dry THF (3 × 6 mL).

Deprotection of "DOTA(OtBu)₃-EN"-PBAs and cleavage from DEAM-PS. DEAM-PS-supported DOTA(OtBu)₃-EN-PBA (0.54 mmol, theoretical loading 1.67 mmol g^{−1}) was suspended in 1 : 1 DCM/TFA (6 mL) and shaken at room temperature for 15 h. The suspension was filtered and the resin was washed with 1 : 1 DCM/TFA (3 × 3 mL, 1 min). The combined filtrates were evaporated under reduced pressure and the residue was dissolved again in TFA (1 mL) and added to Et₂O (10 mL). The resulting suspension was centrifuged (4000 rpm, 30 min, 10 °C) and the solid residue was washed/centrifuged with Et₂O (3 × 10 mL) and dried under vacuum.

DOTA-EN-PBA: 62% yield (calculated as the trifluoroacetate form with respect to DOTA(OtBu)₃-EN). The NMR characterization is in agreement with that reported in the literature (Fig. S3†).¹¹

¹H NMR (400 MHz, 25 °C, D₂O): 7.75 (m, 2H), 7.48 (m, 2H), 4.24 (s, 2H), 3.7–3.3 (bm, 8H), 3.3–2.5 (bm, 20H). ¹³C NMR (100 MHz, 25 °C, D₂O): 178.4, 173.0, 169.7, 151.5, 146.9, 129.7, 128.9, 126.8, 123.9, 61.3, 57.1, 56.4, 56.0, 52.0, 50.4, 48.7, 48.3, 46.7, 43.9, 35.6, 20.6. ¹¹B NMR (128 MHz, 25 °C, D₂O): −19.2 (bs). HR MS (ESI): [M − H₂O + H]⁺ *m/z* calculated for C₂₅H₄₀BN₆O₈ 563.2995, observed 563.2989.

DOTA-EN-F2PBA: 52% yield (calculated as the trifluoroacetate form with respect to DOTA(OtBu)₃-EN). ¹H NMR (400 MHz, 25 °C, D₂O[CD₃CN]): 7.28 (m), 7.17 (m), 4.37 (bs), 3.81 (bm), 3.57 (m), 3.6–3.3 (bm), 3.39 (bm), 3.33 (bm), 3.23 (m), 3.2–2.8 (bm). ¹³C NMR (100 MHz, 25 °C, D₂O[CD₃CN]): 173–169 (CO), 155.3, 153.5, 151.3, 149.3, 131.3, 128.5, 61.3, 56–50, 34.9, 29.4, 20.7. ¹¹B NMR (128 MHz, 25 °C, D₂O[CD₃CN]): 8.22 (bs). ¹⁹F NMR (376 MHz, 25 °C, D₂O[CD₃CN]): −131.79 (dd, ³J_{FF} = 23.7 Hz, ⁴J_{FH} = 4.5 Hz, C(F)CCH₂), −141.54 (dd, ³J_{FF} = 23.7 Hz, ⁴J_{FH} = 3.0 Hz, C(F)C(B)). HR MS (ESI): [M − H₂O + H]⁺ *m/z* calculated for C₂₅H₃₈BF₂N₆O₈ 599.2807, observed 599.2803.

Solution-phase synthesis of DOTA(OtBu)₃-EN-F2PBA. DOTA(OtBu)₃-EN (0.50 g, 0.81 mmol) was dissolved in MeOH (5 mL), and 2,3-difluoroformylphenylboronic acid (0.17 g, 0.89 mmol) was added at 0 °C followed by Et₃N (2 mL). The reaction mixture was stirred at room temperature for 2 h. NaBH₄ (0.37 g, 9.72 mmol) was then added portion-wise at 0 °C and the resulting mixture was stirred at room temperature for 12 h. The solvent was evaporated under reduced pressure, and the residue was suspended in H₂O (10 mL) and extracted with DCM (3 × 5 mL). The organic phases were dried over anhydrous Na₂SO₄, filtered and evaporated under vacuum. The *tert*-butyl-protected DOTA-EN-F2PBA was obtained as a pale-yellow solid that was used in the next reaction step without further purification.

Solution-phase synthesis of DOTA-EN-F2PBA. DOTA(OtBu)₃-EN-F2PBA (0.58 g, 0.74 mmol) was dissolved in DCM (3 mL), TFA (1 mL) was added and the solution was stirred at room temperature overnight. The solvents were evaporated under reduced pressure and the residue was taken up in methanol and evaporated again (3 times). Finally, the residue was dissolved in H₂O, the pH was adjusted to 12 with 1 M NaOH and the solution was passed through an ion exchange resin, rinsing first with H₂O until neutral pH was reached. The aimed compound was eluted with a mixture of pyridine/H₂O/EtOH (5 : 47.5 : 47.5). After evaporation of the solvents under reduced pressure, the residue was dissolved again in H₂O and lyophilized (0.12 g, 24% yield calculated from the initial step). The NMR spectra are discussed in the text.

Synthesis of Gd(III)-complexes of DOTA-EN-PBAs. The ligand (0.10 mmol) was dissolved in H₂O (10 mL). GdCl₃·6H₂O (0.11 mmol) was added, the pH of the solution was adjusted to 6.5 by dropwise addition of 0.1 M aqueous NaOH, and the solution was stirred at 50 °C overnight. After adjusting the pH to 10, the resulting precipitate was filtered off, the filtrate was corrected back to neutral pH by adding 0.1 M HCl and freeze-dried, leading to the target complex as a white solid.



GdDOTA-EN-PBA: 96% yield. ESI⁺ MS: [M + H]⁺ *m/z* calculated for C₂₅H₃₉BGdN₆O₉⁺ 736.2, observed 735.8.

GdDOTA-EN-F2PBA: 93% yield. ESI⁺ MS: [M + H]⁺ *m/z* calculated for C₂₅H₃₇BF₂GdN₆O₉⁺ 772.2, observed 771.8.

Relaxometric measurements

The water-proton longitudinal relaxation rates as a function of the magnetic field strength were measured in non-deuterated aqueous solutions on a fast field-cycling Stellar SmarTracer relaxometer (Stellar s.r.l., Mede (PV), Italy) over a continuum of magnetic-field strengths from 0.00024 to 0.25 T (corresponding to 0.01–10 MHz proton Larmor frequencies). The relaxometer operates under computer control with an absolute uncertainty in 1/*T*₁ of ±1%. Additional longitudinal and transverse relaxation data in the range of 15–70 MHz were obtained on a Stellar relaxometer connected to a Bruker WP80 NMR electromagnet adapted to variable-field measurements (15–80 MHz proton Larmor frequency). The exact concentration of Gd(III) ions was determined by measuring the bulk magnetic susceptibility shifts of a *t*BuOH signal. The ¹H *T*₁ relaxation times were acquired by the standard inversion recovery method with a typical pulse width (90°) of 3.5 ms and 16 experiments of 4 scans. The temperature was controlled with a Stellar VTC-91 airflow heater equipped with a calibrated copper-constantan thermocouple (uncertainty of ±0.1 °C).

Cell lines

Mouse melanoma (B16-F10) cell lines were purchased from American Type Culture Collection. Melanogenic B16-F10m cells were obtained by growing cells in standard DMEM (Lonza) medium supplemented with sodium bicarbonate, and glutamine. The cells were incubated at 37 °C under a humidified atmosphere of 5% CO₂. The medium was supplemented with 10% (v/v) FBS, 2 mM glutamine, 100 U ml^{−1} penicillin, and 100 U ml^{−1} streptomycin. Melanogenesis was evaluated by measuring the absorbance of the cell lysate and medium (dissolved in 1 M NaOH) at 490 nm observing its linear dependency on the number of cell passages. The sialic acid expression on B16-F10 cells was determined using a commercially available AbCAM assay.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- W. L. A. Brooks and B. S. Sumerlin, *Chem. Rev.*, 2016, **116**, 1375–1397.
- R. Smoum, A. Rubinstein, V. M. Dembitsky and M. Srebnik, *Chem. Rev.*, 2012, **112**, 4156–4220.
- L. Xu, Y. Hu, M. Liu, J. Chen, X. Huang, W. Gao and H. Wu, *Tetrahedron*, 2015, **71**, 2079–2088.
- S. D. Bull, M. G. Davidson, J. M. H. Van den Elsen, J. S. Fossey, A. T. A. Jenkins, Y. B. Jiang, Y. Kubo, F. Marken, K. Sakurai, J. Z. Zhao and T. D. James, *Acc. Chem. Res.*, 2013, **46**, 312–326.
- T. Miyahara and K. Kurihara, *J. Am. Chem. Soc.*, 2004, **126**, 5684–5685.
- I. Maluenda and O. Navarro, *Molecules*, 2015, **20**, 7528–7557.
- A. H. Soloway, W. Tjarks, B. A. Barnum, F. G. Rong, R. F. Barth, I. M. Codogni and J. G. Wilson, *Chem. Rev.*, 1998, **98**, 1515–1562.
- R. Schauer, *Glycoconjugate J.*, 2000, **17**, 485–499.
- K. Djanashvili, L. Frullano and J. A. Peters, *Chem. – Eur. J.*, 2005, **11**, 4010–4018.
- K. Djanashvili, G. A. Koning, A. van der Meer, H. T. Wolterbeek and J. A. Peters, *Contrast Media Mol. Imaging*, 2007, **2**, 35–41.
- K. Djanashvili, T. L. M. ten Hagen, R. Blange, D. Schipper, J. A. Peters and G. A. Koning, *Bioorg. Med. Chem.*, 2011, **19**, 1123–1130.
- L. Frullano, J. Rohovec, S. Aime, T. Maschmeyer, M. I. Prata, J. J. P. de Lima, C. Geraldes and J. A. Peters, *Chem. – Eur. J.*, 2004, **10**, 5205–5217.
- S. G. Crich, D. Alberti, I. Szabo, S. Aime and K. Djanashvili, *Angew. Chem., Int. Ed.*, 2013, **52**, 1161–1164.
- C. Tsoukalas, S. Geninatti-Crich, A. Gaitanis, T. Tsotakos, M. Paravatou-Petsotas, S. Aime, R. Jimenez-Juarez, C. D. Anagnostopoulos, K. Djanashvili and P. Bouziotis, *Mol. Imaging Biol.*, 2018, **20**, 798–807.
- K. Takahashi, H. Nakamura, S. Furumoto, K. Yamamoto, H. Fukuda, A. Matsumura and Y. Yamamoto, *Bioorg. Med. Chem.*, 2005, **13**, 735–743.
- J. Yan, G. Springsteen, S. Deeter and B. H. Wang, *Tetrahedron*, 2004, **60**, 11205–11209.
- L. Connah, R. Joshi, S. Vibhute, G. Gambino, J. D. G. Correia and G. Angelovski, *Org. Lett.*, 2019, **21**, 5378–5382.
- J. Singh, V. Rustagi, S. R. Zhang, A. D. Sherry and D. G. Udugamasooriya, *Magn. Reson. Chem.*, 2017, **55**, 747–753.
- M. Tripepi, F. Capuana, E. Gianolio, F. V. C. Kock, A. Pagoto, R. Stefania, G. Digilio and S. Aime, *Bioconjugate Chem.*, 2018, **29**, 1428–1437.
- W. R. Li and Y. C. Yo, *Tetrahedron Lett.*, 1999, **40**, 9085–9089.
- B. Carboni, C. Pourbaix, F. Carreaux, H. Deleuze and B. Maillard, *Tetrahedron Lett.*, 1999, **40**, 7979–7983.



- 22 W. Q. Yang, X. M. Gao, G. Springsteen and B. H. Wang, *Tetrahedron Lett.*, 2002, **43**, 6339–6342.
- 23 M. A. M. Behnam, T. R. Sundermann and C. D. Klein, *Org. Lett.*, 2016, **18**, 2016–2019.
- 24 S. Arimori, J. H. Hartley, M. L. Bell, C. S. Oh and T. D. James, *Tetrahedron Lett.*, 2000, **41**, 10291–10294.
- 25 M. Gravel, K. A. Thompson, M. Zak, C. Berube and D. G. Hall, *J. Org. Chem.*, 2002, **67**, 3–15.
- 26 D. G. Hall, J. Tailor and M. Gravel, *Angew. Chem., Int. Ed.*, 1999, **38**, 3064–3067.
- 27 P. A. Cox, M. Reid, A. G. Leach, A. D. Campbell, E. J. King and G. C. Lloyd-Jones, *J. Am. Chem. Soc.*, 2017, **139**, 13156–13165.
- 28 D. M. Corsi, C. Platas-Iglesias, H. Van Bakkum and J. A. Peters, *Magn. Reson. Chem.*, 2001, **39**, 723–726.
- 29 *The Chemistry of Contrast Agents in Medical Magnetic Resonance Imaging*, ed. A. Merbach, L. Helm and E. Toth, John Wiley & Sons Ltd., 2013.
- 30 D. H. Powell, O. M. N. Dhubhghaill, D. Pubanz, L. Helm, Y. S. Lebedev, W. Schlaepfer and A. E. Merbach, *J. Am. Chem. Soc.*, 1996, **118**, 9333–9346.

