

PAPER

View Article Online
View Journal | View Issue



Cite this: *Org. Biomol. Chem.*, 2020, **18**, 8735

Functionalized fluorescent terephthalate monomers and their attempted polyester formation†

Yvonne S. L. Choo,^a Marta Giamberini,^b José Antonio,^c Paul G. Waddell^d and Andrew C. Benniston^{*,e}

The reaction of diethyl 2,5-bis(*tert*-butyl)phenoxy-3,6-dihydroxyterephthalate (**1**) with tetraethylene glycol di(*p*-toluenesulfonate) under high-dilution conditions afforded several isolated products. Two products were identified as macrocycles with one being the 1 + 1 crown ether derivative **3** (10% yield), and the second being the 2 + 2 crown ether compound **D3** (19% yield). The X-ray structure for **3** was determined with the asymmetric unit observed to comprise half of the molecule. The small crown ether ring of **3** interacts with K⁺ or H⁺ ions in MeOH, but binding is weak and the macrocyclic cavity is too small to fully encapsulate the K⁺ ion. Transesterification of compounds **1**, its methylated version **2** and **3** with diols such as ethylene glycol or 1,4-butanediol produced monomers (**M1–M3**) which were reacted with terephthaloyl chloride. Short oligomers were produced (**PolyM1–PolyM3**) rather than extensive polymeric materials and all displayed solid state fluorescence. The absorption and fluorescence properties of **M1–M2** and their polymers can be related to subtle structural changes. The Stokes shift for **M2** of 15 627 cm^{−1} in DCM is one of the largest observed for a simple organic chromophore in fluid solution.

Received 24th July 2020,
Accepted 13th October 2020

DOI: 10.1039/d0ob01533d

rsc.li/obc

Introduction

Terephthalate derivatives are the cornerstone of the manufacture of the widely used polyester plastics,¹ which have found numerous commercial applications in areas such as bottles, packaging, cosmetics, thermal insulating materials and car accessories, to name but a few. The preparation of, for example, poly(ethylene terephthalate) (PET) is relatively straightforward, requiring only high temperatures to perform in the first instance, the transesterification of dimethyl-terephthalate with ethylene glycol.² The final stage is the

driving of polymer formation by vacuum distillation of ethylene glycol under high temperatures. The properties of PET stand it in good stead because of its suitable mechanical features, chemical resistance, thermal stability, high transparency/flexibility and low permittivity to gases.³ Considering all of these positive features of PET it is not surprising that non-commercial applications of the polymer have also been sought in research-biased areas including force-sensing resistor materials,⁴ nanofiber phototransistors,⁵ rechargeable polymer batteries,⁶ solar cells⁷ and OLED devices.⁸ One feature from these studies is the consistency in structure of the basic terephthalate core with any additional functionality arising from blending of additional agents. There is clearly no reason, except based on a steric argument, that the actual terephthalate could not support other substituents, which could themselves have additional properties that could be imparted to the final polymer. Features could include, for example, pH sensitivity,⁹ thermochromic response,¹⁰ electrical conductivity,¹¹ ion transport¹² and fluorescence-based remote sensing.¹³ The final two categories are of special interest for potential applications in proton transport membranes¹⁴ and explosives detection.¹⁵

Recently we developed the preparation of highly substituted terephthalate derivatives based on diethyl 2,5-bis(aryl)phenoxy-3,6-dihydroxyterephthalate, where the aryl groups are several disparate aromatic subunits.¹⁶ The compounds dis-

^aSchool of Energy and Chemical Engineering, Xiamen University Malaysia, Jalan Sunsuria, Bandar Sunsuria, Sepang 43900, Selangor Darul Ehsan, Malaysia

^bDepartament d'Enginyeria Química, Universitat Rovira i Virgili, Av. Països Catalans 26, 43007 Tarragona, Spain

^cDepartament de Química Analítica i Química Orgànica, Universitat Rovira i Virgili, Carrer Marcel·lí Domingo s/n, 43007 Tarragona, Spain

^dCrystallography Laboratory, Chemistry-School of Natural & Environmental Sciences, Newcastle University, Newcastle upon Tyne, NE1 7RU, UK

^eMolecular Photonics Laboratory, Chemistry-School of Natural & Environmental Sciences, Newcastle University, Newcastle upon Tyne, NE1 7RU, UK.

E-mail: andrew.benniston@ncl.ac.uk

†Electronic supplementary information (ESI) available: NMR spectra of compounds, DFT calculation results, binding data and absorption spectra. CCDC 2018846. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/d0ob01533d



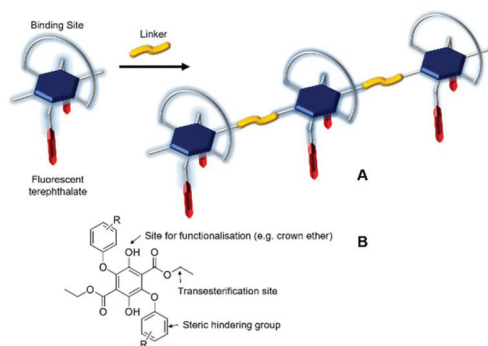


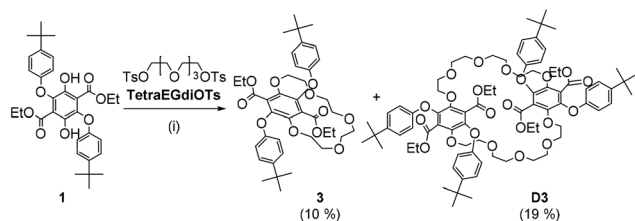
Fig. 1 (A) Cartoon representation of polymer formation by linking of a functionalized fluorescent terephthalate subunit. (B) The basic building block for preparation of a terephthalate monomer.

played moderately intense fluorescence both in fluidic solution as well as the solid state; the latter probably a result of the bulky groups which restricted close packing. In addition, the Stokes shift for the compounds was relatively large, supporting major internal structural change upon excitation. Noting the potential for such compounds to be further functionalized at the hydroxyl (*e.g.*, crown ether) or ester groups (*e.g.*, transesterification) we envisaged that new terephthalate-based polymeric materials could also be feasible (Fig. 1). The additional functionality of a crown ether offering a cation-binding site to manipulate the polymer structure and impose alteration to its fluorescence properties. Despite success in preparation of the crown-based monomer, its polymerisation proved challenging, but simpler monomer versions could be produced. The fluorescence properties of these monomers and their polyesters revealed subtle differences, highlighting larger Stokes shifts than reported previously. In addition, it was shown that the internal polymer structure restricted structural motions within the terephthalate core upon photoexcitation; the flexibility of the spacer group between each monomer being an important parameter in this observation.

Results and discussion

Synthesis and structure

The synthetic procedure used in the production of the crown ether is illustrated in Scheme 1 starting from the precursor **1** prepared previously using published methods.¹⁶ In an attempt



Scheme 1 Reagents and conditions (i) MeCN, K₂CO₃, KPF₆, reflux, 9 days.

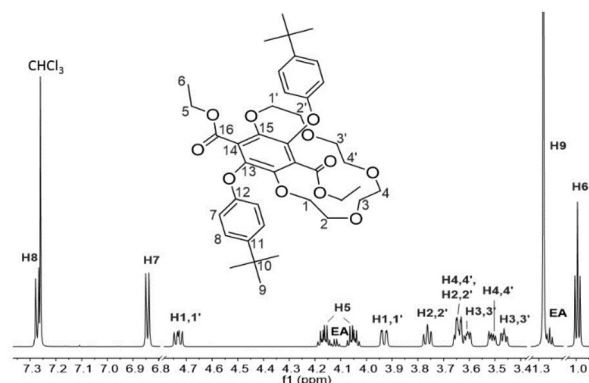


Fig. 2 ¹H NMR spectrum (700 MHz) of **3** in CDCl₃ showing the proton signals assignment. EA = trace of ethyl acetate solvent.

to minimise polymerisation reactions, the high dilution method was used in which the diol **1** and the tetraethylene glycol di(*p*-toluenesulfonate) were added slowly to a refluxing MeCN solution. After 9 days the reaction was worked up and the crude product purified by careful column chromatography on silica gel by using ethyl acetate/petroleum ether (1 : 5 then 1 : 4) as the eluent to afford two main products, **3** and **D3**. The unequivocal identification of the products was performed using a combination of ¹H NMR spectroscopy, mass spectrometry and where possible, X-ray crystallography.

The ¹H NMR spectrum for **3** in CDCl₃ is shown in Fig. 2. The prominent multiplet signals in the δ = 3.94–3.45 and 4.73 ppm are readily assigned to the OCH₂ protons for the crown ether. By careful inspection of the COSY spectrum a partial assignment of the signals to the methylene units was possible. The first point to note is that the ¹H NMR spectrum for **1** is not straightforward because of slowly interconverting conformers which impart inequivalence to the CH₂ protons of the ethyl groups. Very noticeable are the downfield signals at δ = 4.73 corresponding to the H1,1' protons which appear as a doublet of doublet of doublets, confirming that each separate proton of a CH₂ unit are diastereotopic. Variable temperature ¹H NMR spectra, coupled to COSY assignments, were highly informative of changes to the conformation of **3** in *d*₈-toluene (see ESI S1†). A significant downfield shift for the H2,2' proton resonances is observed with increasing temperature whereas upfield shifts are witnessed for the remaining CH₂ protons. Apart from the shift in peaks, no coalescence of the two methylene multiplets (H5) or the diastereotopic CH₂ protons are observed. However, there is a loss in resolution when the temperature is lowered to 183 K.

Further corroboration that the solid was in fact the 1 + 1 product was obtained by positive nanoESI (DCM/MeOH + KOAc) which displayed a prominent peak at m/z = 747.3126 corresponding interestingly to the [M + K]⁺ ion and a peak at m/z = 726.3848 corresponding to the [M + NH₄]⁺ ion (DCM/MeOH + NH₄OAc). However, the unequivocal identification of **3** was obtained by the use of single-crystal X-ray crystallography, and the molecular structure is illustrated in Fig. 3.



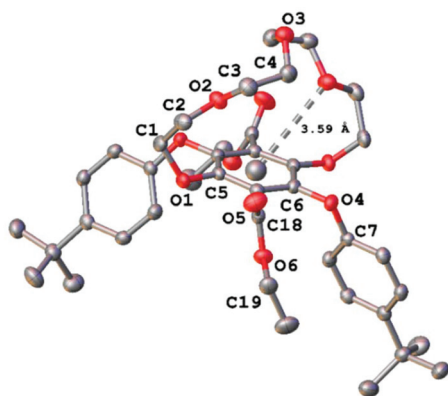


Fig. 3 X-ray crystal structure of **3** with the centroid...O2 distance highlighted. Ellipsoids are drawn at the 50% probability level and hydrogen atoms have been omitted for clarity.

Table 1 Selected bond length and angles for the structure of compound **3**

Atoms	Bond length/Å	Atoms	Angle/°
O1–C1	1.4459(18)	C5–O1–C1	116.84(10)
O1–C5	1.3760(16)	C3–O2–C2	113.70(11)
O2–C2	1.4211(18)	C4–O3–C4	115.93(17)
O2–C3	1.421(2)	C6–O4–C7	117.64(10)
O3–C4	1.4240(18)	C18–O6–C19	117.35(11)
O4–C6	1.3892(17)	O1–C1–C2	112.25(13)
O4–C7	1.3932(17)	O2–C2–C1	107.81(12)
O5–C18	1.1982(18)	O2–C3–C4	109.09(12)
O6–C18	1.3349(18)	O3–C4–C3	113.32(13)
O6–C19	1.4553(17)	O5–C18–O6	124.87(13)

Selected bond lengths and angles are collected in Table 1. The two bulky *tert*-butyl aryl groups are *syn* to each other and point away from the crown ether moiety which encloses the central benzene ring. The effective size of the cavity can be assessed by considering the atom of the crown ether moiety closest to the benzene ring centroid (Fig. 3). This centroid...O2 distance was observed to be 3.59(1) Å, a relatively short contact. It is conceivable however that the conformational flexibility of the ring may allow it to expand further.

Noting the conformational flexibility of the compound in solution, and that the jellyfish-like shape of the molecule in the crystal structure was conceivably not the lowest energy conformation, complementary molecular modelling calculations were performed. Using the molecular geometry of the crystal structure as the starting point an energy minimisation calculation was performed using DFT (B3LYP) and a 6-311G basis set. There was only a minor structural alteration which suggested the solid state structure actually represented the lowest energy conformation. Basic molecular dynamics simulations (MM⁺) were far more informative (see ESI S2†) and several different energy conformations were observed. In full agreement with the solid-state structure and the DFT calculation, the *syn tert*-butyl aryl arrangement would appear to be the lowest energy conformation; the *anti* structure is slightly

higher in energy by *ca.* 5 kcal mol^{−1}. Two other higher energy conformations sampled are associated with variations in the crown ether geometry.

Cation binding studies of **3**

Based on the crystal structure of **3** the crown ether moiety appears unobstructed by the two bulky *tert*-butyl aryl groups as they are orientated away from the cavity and are arranged *syn* to one another. However, the relatively short centroid...O2 distance and the potential steric hindrance of the carbonyl groups for the *tert*-butyl aryl groups either side of the ring raises the question of whether the cavity would be large enough for any cation to fit in it. Upon performing a structural database search on ConQuest,¹⁷ no X-ray crystal structures of compounds with similar features to **3** could be found, let alone examples with a cation bound within the cavity.

To our surprise, the ¹H NMR spectra of **3** recorded in CD₃OD in the absence and presence of K⁺ ions (KPF₆) did show subtle differences as shown in Fig. 4. Small downfield shifts and changes in splitting patterns were observed from the OCH₂ protons on the alkoxy chain but the aromatic protons (H7 and H8) and the CH₃ protons of the ethyl ester groups (H6) remained unchanged. This may potentially be attributed to some cation-crown interaction whereby the K⁺ ion does not necessarily bind within the cavity but may reside at the periphery of the crown and interact with the oxygen atoms on the alkoxy chain. This would explain the positive nanoESI mass spectrometry results of **3** for the presence of [M + K]⁺ species. The unique interaction of a K⁺ ion with **3** in CD₃OD was found to be selective as all the other cation salts (LiPF₆, NaPF₆ and NH₄PF₆) revealed no changes or shifts in the ¹H NMR signals. Since all the salts used were hexafluorophosphates, any anion interaction with the crown ether can be ruled out.

Absorption and emission spectra of **3** in MeOH were also recorded in the absence and presence of cation salts. Unfortunately, no significant change was observed. As for the

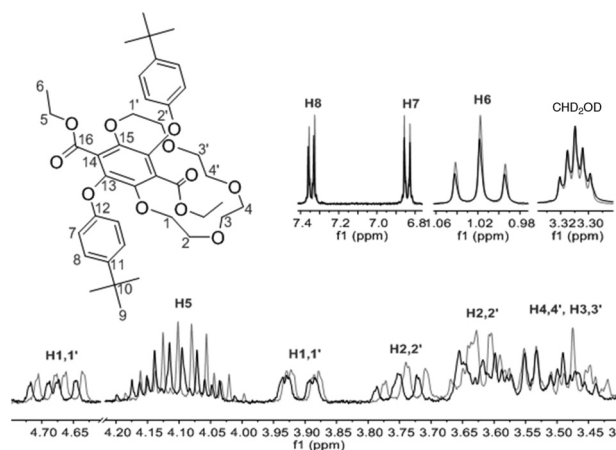


Fig. 4 ¹H NMR spectra (300 MHz, CD₃OD) of **3** before (grey) and after (bold) addition of K⁺ ions.



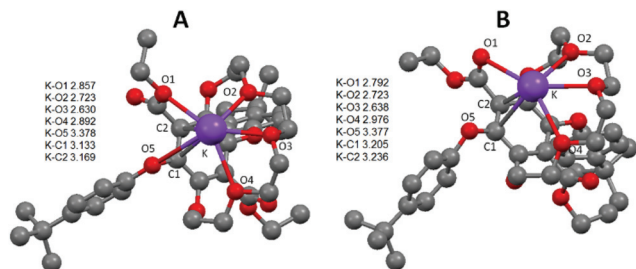


Fig. 5 Gaussian calculated energy-minimised gas phase structures for a K^+ adduct of **3** using B3LYP and a 6-311G basis set. Distances shown as the insert are in Å and hydrogens are omitted for clarity.

crown-ether-cation complex crystallisations attempted with sodium, potassium and ammonium hexafluorophosphate salts, only free **3** (uncomplexed) was crystallised in all of the cases. Since all attempts to grow crystals of the K^+ adduct with **3** failed we turned our attention to DFT calculations to try and ascertain a probable structure. Starting from the solid state structure a K^+ ion was placed in several positions within bonding distances to disparate oxygen atoms of the crown, and the structures were energy minimised in Gaussian 09 using B3LYP and a 6-311G basis set. From several calculations two structures, as shown in Fig. 5, emerged as conceivable adducts of **3** with a K^+ ion; differing only slightly in energy because of small structural variations. Several key aspects are apparent from examination of the structures. The first to note is that only three oxygen atoms of the crown ether bind to the potassium ion and that the two crown-based phenoxyl atoms do not. The latter observation is perhaps not too surprising considering the severe structural distortion that would be required to fully accommodate the potassium ion. The K–O bond distances relating to the oxygen atoms of the crown ether are in good agreement with those found in several X-ray structures of relevant macrocyclic complexes.¹⁸ Additional oxygen donor interactions to the K^+ appear to be *via* the ethyl ester, using the ethyl oxygen in structure A and the carbonyl oxygen for structure B; the latter structure being *ca.* 4 kcal mol^{−1} more stable. For both structures there is the possibility of a K–O5 interaction although the actual distances are rather long. The K–C1 and K–C2 distances are potentially just within the crystallographically determined range (2.990–3.117 Å) to be considered an η^2 interaction to the aryl ring.¹⁹ It is worth noting that the analogues structures calculated for **3** with the smaller Li^+ ion (see ESI S3†) showed that only two oxygen atoms of the crown bind to the metal ion, but with the ester still able to participate in ion binding. The Li–O5 distance is also far too long to be considered significant. The combination of weaker interactions is likely one reason for the observed lack of binding of **3** with Li^+ . The structure of **3** with Na^+ (see ESI S4†) is more akin to the K^+ structure shown.

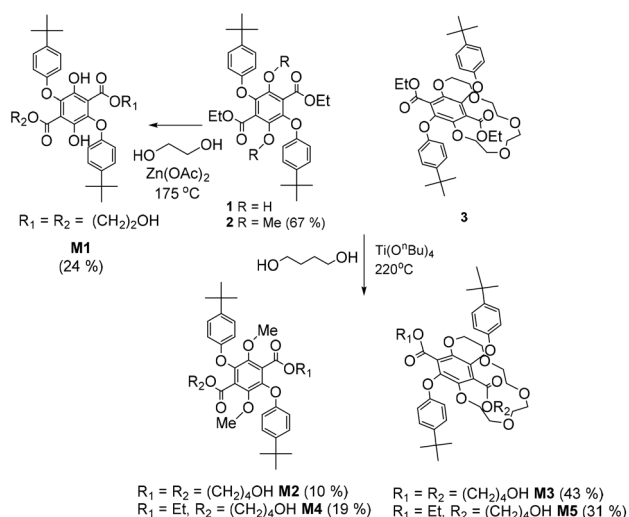
Proton binding studies of **3**

Since **3** was designed with the intention of its being polymerised to create a polymer containing proton binding sites, it was

deemed reasonable to first test its ability to coordinate with protons. To this end, several NMR titration experiments were conducted in $CDCl_3$ using trifluoroacetic acid as the proton source. Interestingly, no chemical shift changes were observed throughout the titration experiment for most of the proton peaks of **3** aside from H3,3' and H4,4' (protons from the crown ether chain) and H5 (protons from the ester side groups). By focusing on the region between 3.90–3.40 ppm in the stacked 1H NMR spectra (see ESI S74–76(a)†) the chemical shift data obtained from peaks corresponding to H3,3', H3,3' and H4,4' were non-linearly fitted to a binding equation (see ESI S74–76(b)†). The average binding constant $\log K_a$ was calculated to be 2.2 M^{−1} which is a relatively small value. There are two potential binding sites: the ester group and the crown cavity. Based on literature, the association constant for the protonation of an ester is found to be very small with $\log K_a$ values between −15 and −1 M^{−1}.²⁰ The binding constants of cation complexes fall within the $\log K_a$ range of 1.0 and 2.1 M^{−1} while proton complexes have $\log K_a$ value of around 2.9 M^{−1} and ΔG° (300 K) of −15.9 kJ mol^{−1} (15-crown-5).²⁰ Therefore, to a certain degree of certainty, the values obtained were indeed the binding constants of **3** with H^+ ($\log K_a$ of 2.2 M^{−1} and ΔG° of −13 kJ mol^{−1}). Several attempts to crystallise the protonated complex of **3** with TFA were to no avail. Likewise, no $[M + H]^+$ ion could be detected in the positive nanoESI spectrum of **3**. Given the results it is speculated that the proton interacts weakly with oxygen atoms of the crown moiety and is not buried deep within the cavity.

Polymerisation results

Polymerization of terephthalate derivatives is often performed using high temperature transesterification in the presence of an appropriate diol and a Lewis acid catalyst.²¹ To aid in the polymerization studies control compounds were employed to ascertain appropriate conditions and identify any problems (Scheme 2). The attempted polymerization of **1** with stoichio-



Scheme 2 Procedures used in the preparation of the diol monomers **M1–M3** and the side products **M4–M5**.



metric ethylene glycol at high temperature immediately identified a drawback in that the *O*-aryl group tended to be cleaved off. The design protocol was altered slightly to introduce two terminal alcohol groups so that simple esterification with a suitable diacid chloride would form a polyester. The reaction of **1** with excess ethylene glycol containing zinc(II) acetate at 175 °C produced the monomer diol **M1** in 24% yield, but still showed signs of some degradation in the purification products. Given this problem a different catalyst system was identified and the two OH groups were protected as their methyl ether (**2**). To also extend the spacer length the reaction of **2** in excess butanol but using $\text{Ti}(\text{O}^i\text{Bn})_4$ as the catalyst was employed and afforded both **M2** and **M4** after column chromatography.

Given the information gleaned by the test procedures the reaction of **3** in excess butanol using $\text{Ti}(\text{O}^i\text{Bn})_4$ as the catalyst worked well and afforded **M3** and the side product **M5** after purification. In the ^1H NMR spectrum of **M3** (see ESI S5†), peaks attributed to the CH_2 protons on the newly attached butyl chains are seen as two sets of mirroring multiplets (at 4.20 and 4.07 ppm – H5), an overlapped multiplet at 3.48 ppm (H18) and additional multiplets at 1.50 ppm (H6) and 1.35 ppm (H17). The two protons from the terminal OH groups (H19) could also be seen as a broad peak at 1.59 ppm. The diastereotropic nature of the OCH_2 protons of the crown ether characteristic of **3** was unaltered by transesterification, and was observed as prominent multiplet signals around 3.93–3.42 ppm and 4.69 ppm. The ^1H NMR spectrum of asymmetrical **M5** (see ESI S56†) appeared to be much more complicated that its symmetrical counterpart with two sets of peaks each in addition to overlapping ones.

Polyester formation

The attempted polyester formation reactions are illustrated in Scheme 3 and for simplicity terephthaloyl chloride was used as the linker unit. In all the reactions a slight excess of the monomer was used to ensure that any polymer would be end capped with a hydroxyl group. An excess of the base triethylamine was used to mop up the HCl generated by the ester

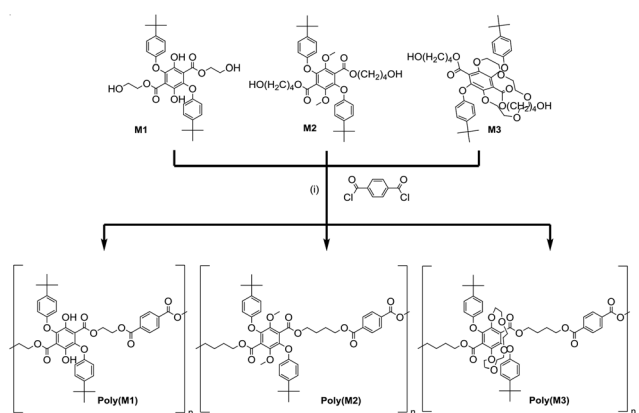
formation reaction. The ^1H NMR spectrum of **PolyM1** (see ESI S10†) showed several broad peaks which are generally observed in the spectrum of a polymeric material. In addition, by comparing the ^{13}C NMR spectrum of **PolyM1** with that of its monomer (see ESI S13†), the stronger intensity carbon peaks could be attributed to the repeating unit of the polymer and the weaker intensity carbon peaks to the end groups.

DOSY NMR spectroscopy was employed for the determination of the diffusion coefficient ($D = 9.06 \times 10^{-11} \text{ m}^2 \text{ s}^{-1}$), and the identification of proton peaks of a polymer from a mixture of compounds. The diffusion coefficient was used to estimate the molecular weight (M_w) of **PolyM1** ($23\,360 \text{ g mol}^{-1}$), which is relatively close to the M_w value obtained from a GPC experiment ($25\,047 \text{ g mol}^{-1}$). In contrast, the calculated M_n from the ^1H NMR spectrum (8423 g mol^{-1}) is much lower reflecting the difference in the methods. It is important to note that the difference may be attributed to the fact that GPC operates with the assumption that the polymer introduced in the system behaves like that of PS in THF/toluene. The copolymer in this case could have had very different hydrodynamic volume and behaviour in THF/toluene, resulting in an under/overestimation of the M_w and M_n . The glass transition temperature, T_g of the polymer, as determined from the second heating curve in the DSC thermogram, was found to be 91 °C. This value is consistent with its brittle nature of the polymer as observed at room temperature.

The copolymer, **PolyM2** was synthesised in a similar manner to **PolyM1** and characterised using a similar method (see ESI S14–18†). Collected in Table 2 are data comparing the two polymers. The T_g for **PolyM2** of 48 °C is significantly lower than that of **PolyM1** (91 °C). This difference is likely a result of the longer aliphatic chain (butylene spacer instead of ethylene spacer) and a smaller number of repeating units ($n = 8$). Given the test conditions the attempted polymerisation of **M3** was carried out and afforded an opaque off-white material. The ^1H NMR spectrum of the material (see ESI S19†) appeared only slightly broadened with respect to the starting material. Most of the peaks observed in the ^1H NMR spectrum could be readily assigned to the main parts of the chemical structure for **PolyM3**, and suggested that the compound only contained a few repeating units. The lack of polymerisation of **M3** was disappointing and seemed to be related to the presence of the crown group, but the reason is not immediately obvious. Molecular dynamics simulations for the oligomers ($n = 2, 4, 8$) show that all the structures tend to fold in on themselves the greater the number of repeat units, and hence start to enclose and hinder the terminal alcohol groups. The crown ether of **M3** unlike the other two monomers provides multiple hydrogen bonding sites which may interact with the terminal hydroxyl groups to retard the esterification reaction.

Absorption and emission

All three polymers exhibited solid-state fluorescence as shown in Fig. 6, but with slightly different colours as observed by eye. The subtle colour difference between **PolyM1** and **PolyM2** appears to arise from the more structured emission in the



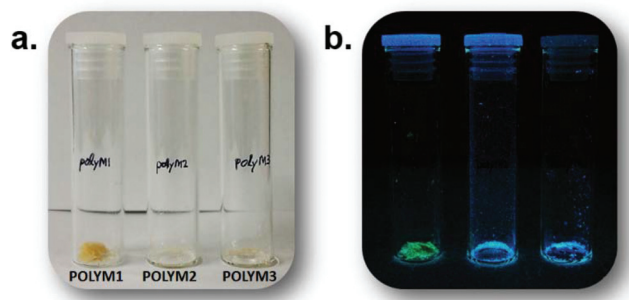
Scheme 3 Reagents and conditions: (i) Triethylamine, THF, 0 °C then room temperature, 3 days.



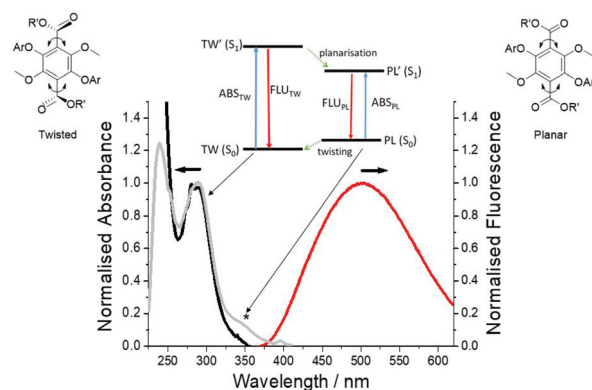
Table 2 Number average molecular weight (M_n), weight average molecular weight (M_w), and polydispersity index (PDI) of **PolyM1** and **PolyM2**

Polymer	^1H and DOSY NMR				GPC ^d		
	M_n^a (g mol ⁻¹)	M_w^b (g mol ⁻¹)	PDI ^c	D (m ² s ⁻¹)	M_n (g mol ⁻¹)	M_w (g mol ⁻¹)	PDI
PolyM1	8423	23 360	2.77	9.06×10^{-11}	12 806	25 047	1.96
PolyM2	7839	10 670	1.36	1.38×10^{-10}	1809	3471	1.92

^a Number average molecular weight (M_n) was calculated from the ^1H NMR spectrum. ^b Weight average molecular weight (M_w) was calculated from diffusion coefficient, D obtained from the 2D DOSY NMR spectrum together with the equation of PS calibration curve by Grubbs *et al.* $\log D = -0.537 \log M_w - 7.697$ ($R^2 = 0.9991$). ^c $\text{PDI} = M_w/M_n$. ^d Monodispersed PS standard as the calibrant, THF/toluene as the eluent.

**Fig. 6** Side-by-side comparison of **PolyM1**, **PolyM2** and **PolyM3** in (a) normal light conditions and (b) under 312 nm UV irradiation.

blue-end of the spectrum (see ESI S20†). Although there are several polymers that emit in the blue region,²² the unique features of the two materials are revealed by careful inspection of the absorption and emission/excitation profiles for **M1**, **M2** and the starting material **1**. The X-ray crystal structure and molecular modelling calculations (DFT, B3LYP, 6-311G) for **1** show that the ground-state molecule is planar with the hydroxyl group forming an intramolecular hydrogen bond to the ester. The structure also remains planar in the first-excited singlet state, but there is a rearrangement manifest in changes to the bond lengths within the terephthalate core (see ESI S21†). The absorption maximum (λ_{ABS}) in DCM is located at 392 nm ($25\,510\text{ cm}^{-1}$) and the broad emission profile has a maximum (λ_{EM}) at 488 nm ($20\,492\text{ cm}^{-1}$). The fluorescence excitation spectrum is a good match to the absorption profile (see ESI S23†). The large Stokes shift (SS) of 5018 cm^{-1} is a prominent feature of this type of structure. It is noted that an absorption spectrum calculated using time-dependent DFT (TD-DFT) and using a planar structure as the starting geometry is consistent with the experimental spectrum, but it is considerably blue-shifted if both ester groups are twisted out of the plane (see ESI S22†). This observation is important when comparing the absorption and emission properties of **M1**, **M2** and their polymers. The ethylene glycol transesterification of **1** has little effect on the absorption and emission properties; the λ_{ABS} and λ_{EM} for **M1** are located at 393 nm and 488 nm, respectively (see ESI S25†). The absorption and fluorescence spectra for **M2** (Fig. 7) are significantly different and must be related to the methylation of the two hydroxyl groups. The absorption profile is slightly structured and considerably blue-

**Fig. 7** Absorption (black), excitation (grey) and fluorescence (red) spectra of **M2** in dilute DCM. $\lambda_{\text{ex}} = 320\text{ nm}$. The asterisk notes the additional feature seen in the excitation spectrum to longer wavelength. Insert shows a simple picture of the absorption and emission from the two different conformers. PL = planar, TW = twisted and that the potential energy surfaces $\text{TW}^* \neq \text{TW}$ and $\text{PL} \neq \text{PL}^*$.

shifted compared to **M1** ($\lambda_{\text{ABS}} = 281\text{ nm}$). Again, the fluorescence profile is broad with λ_{EM} located at 501 nm, and red-shifted with respect to **M1**. Even though the fluorescence excitation spectrum matches well to the absorption profile in most parts, there is an additional feature (marked) which is significant and is related to the structure of the compound. The high-energy absorption profile is consistent with a ground-state structure in which the ester groups are twisted out of the aromatic plane (*cf.* **1**). Further evidence for this can be observed in the calculated structure (DFT, B3LYP, 6-311G) of the ground-state for **2** (simplified version of **M2**) (see ESI S26†). The two ester groups adopt a more co-planar arrangement with the central aromatic ring as illustrated in the TD-DFT calculated structure of the first-excited singlet state (see ESI S26†). Such a large-scale structural alteration accompanying the ground to relaxed excited-state transition is the reason for the large SS. It should be noted that the apparent SS of $15\,627\text{ cm}^{-1}$ for **M2** is one of the largest observed for a simple organic chromophore in fluid solution.²³ This large SS is certainly conducive for the application of **M2** (or a derivative) as a bio-fluorescent tag, but with the drawback of requiring excitation in the UV region.

Given that the absorption profile for a planar structure is more red-shifted, the additional feature seen in the excitation



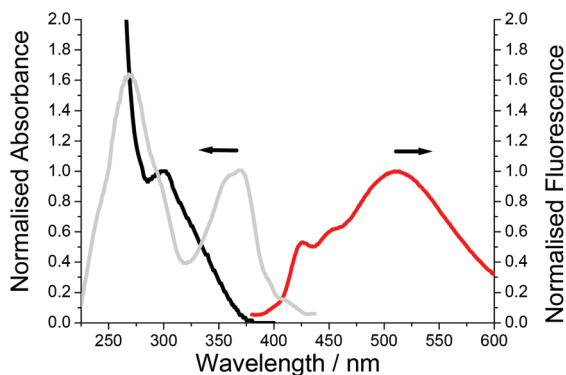


Fig. 8 Absorption (black), excitation (grey) and fluorescence (red) spectra of **PolyM1** in dilute DCM. $\lambda_{\text{ex}} = 320$ nm.

spectrum is assigned to a planar conformation, which is formed following deactivation of the planar S_1 state. This planar conformation must relax readily to the lower energy twisted conformation under normal conditions. The low-energy absorption feature is much more prominent ($\lambda_{\text{abs}} = 365$ nm) in the spectrum for **PolyM2** (see ESI S20†). In addition, the emission profile contains a slight shoulder at *ca.* 428 nm, which is not observed for the monomer (Fig. 7). As the absorption spectrum for **M2** and **PolyM2** are remarkably similar, the ground-state structures of the aromatic core are likely comparable and hence exhibit the twisted ester subunits. The vertical Franck–Condon transition in both cases produces the twisted S_1 conformation. However, in **PolyM2** the twisting of the ester groups to generate the planar conformation is more hindered and emission is also observed from this state, explaining the high-energy shoulder in the emission profile. This type of dual emission is not uncommon in molecular systems that can twist and is observed in porphyrin dimers.²⁴

Major perturbation of the ground-state structure is observed for **PolyM1** by inspection of its absorption spectrum; λ_{abs} is located at 300 nm but the profile is broad stretching to around 375 nm (Fig. 8). By comparison to the absorption profile of Fig. 7, the major conformer comprises ester groups twisted out of the plane, implying that the intramolecular hydrogen bonding is disrupted when the monomer **M1** is polymerised. The broadness of the absorption profile does suggest that there is still a small distribution of the hydrogen bonded planar conformer in the ground state. The excitation spectrum however clearly indicates that this conformer is populated after deactivation of the first-excited state using the model discussed for **PolyM2**. That the excitation profile is more intense and the high-energy structured emission band is far more prominent suggests that the short ethylene glycol spacer severely restricts molecular rearrangement after excitation.

Conclusion

The two hydroxyl groups of diethyl 2,5-bis(*tert*-butyl)phenoxy-3,6-dihydroxyterephthalate are readily functionalised, and are

ideal sites for crown ether formation. The ring size for the first example produced (**3**) is too small for complete cation encapsulation, but the methodology could be readily adapted to increase the number of ethylene glycol spacer units. However, as observed in the macrocycle forming reaction the 2 + 2 adduct may also be formed, and dominate the mixture of products. The 2 + 2 adduct (**D3**) is interesting as its core resembles the well-known 34-Crown-10 macrocycle.²⁵ This compound has found wide application in the binding of electron deficient moieties such as paraquat (*N,N'*-dimethyl-4,4-bipyridinium chloride).²⁶ Preliminary results support the same type of self-assembly with **D3** despite the steric demands imposed by the additional ester and aromatic groups.

The highly substituted phthalate unit is also a prime candidate for the production of fluorescent polymeric materials, but direct transesterification is not appropriate due to problems caused by degradation. A better approach appears to be the attachment of short spacer groups under milder transesterification conditions, which contain an end group that facilitates polymerization. Even so, using the esterification method detailed in this work only short oligomers are produced. A RAFT approach may be more conducive to control the growth of the polymer.

Experimental

Materials

Reagents and solvents were purchased from Sigma Aldrich at the highest purity possible and were used without further purification unless otherwise stated. Standard solvents were dried by literature methods before being distilled and stored under nitrogen over 4 Å molecular sieves. Thin layer chromatography (TLC) was performed with silica gel TLC plates. The spots were visualized under UV light at 312 nm and 365 nm to monitor progress of the reaction.

^1H and ^{13}C NMR spectra were recorded on either the Bruker Avance III 300 MHz, Bruker Avance III HD 500 MHz or Bruker Avance III 700 MHz. 2D NMR such as ^1H – ^1H correlation (COSY), ^1H – ^{13}C heteronuclear single quantum correlation (HSQC), ^1H – ^{13}C heteronuclear multiple bond correlation (HMBC) were used to substantiate ^1H and ^{13}C NMR assignments of the compounds. Deuterated chloroform (CDCl_3), methanol (d_4 -MeOH) and dimethylsulphoxide (DMSO-d_6) were used as the solvents and TMS as the internal standard. FTIR spectra were obtained using Varian 800 FTIR instrument (Varian Inc.) fitted with a diamond crystal plate ATR unit (Pike Technologies). Melting points of intermediates were measured using a StuartTM melting point apparatus SMP3. Absorption spectra were recorded using UV-1800 UV-Vis Spectrophotometer. Matrix-assisted laser desorption ionization (MALDI) mass spectrometry was carried out using the Applied BioSystems Voyager DE-STR and Bruker ultrafleXtreme mass spectrometers at the EPSRC UK National Mass Spectrometry Facility at Swansea. Thermogravimetric analysis experiments (TGA) were carried out in a Mettler TGA SDTA 851e thermoba-



lance. Approximately 8 mg of samples were heated from 30 to 650 °C at 10 °C min⁻¹ in a nitrogen atmosphere (100 mL min⁻¹). Calorimetric studies were carried out in a Mettler DSC822e Differential Scanning Calorimeter. Approximately 5 mg of samples were weighed into 40 µL aluminium crucibles covered with a pierced lid. The analyses were performed in dynamic mode at a heating and cooling rate of 10 °C min⁻¹ using nitrogen as a purge gas (100 mL min⁻¹). The number average (M_n) and weight average (M_w) molecular weight of the polymers were determined by the use of an Agilent 1200 Series GPC-SEC system consisting of three columns in series (PLgel 20 µm MIXED-A, PLgel 5 µm MIXED-D and PLgel 3 µm MIXED-E) and a refractive index detector. Monodispersed polystyrene (PS) standards were used for calibration and the mobile phase (THF with toluene as internal standard) was eluted at a flow rate of 1.0 mL min⁻¹. The sample concentrations used were 5–10 mg mL⁻¹.

Crystal structure data for **3** were collected at 150 K on an Xcalibur, Atlas, Gemini ultra diffractometer equipped with a fine-focus sealed X-ray tube ($\lambda_{\text{CuK}\alpha} = 1.54184 \text{ \AA}$) and an Oxford Cryosystems CryoStreamPlus open-flow N₂ cooling device. Cell refinement, data collection and data reduction were undertaken *via* the software CrysAlisPro.²⁷ Intensities were corrected for absorption empirically using spherical harmonics. All structures were solved using XT²⁸ and refined using XL.²⁹ All non-hydrogen atoms were refined as anisotropic. Hydrogen atoms were positioned with idealised geometry with the exception of those bound to heteroatoms the locations of which were identified using peaks in the Fourier difference map. The atomic displacement parameters of the hydrogen atoms were set to be an appropriate multiple of those of the parent atom.

Computational calculations were performed using a 32-bit version of Gaussian09³⁰ on a quadruple-core Intel Xeon system with 4GB RAM. The calculations were run in parallel, fully utilising they multi-core processor. To reduce computational time low-level calculations were carried out to minimise structures using Hartree–Fock and a low basis set. Energy-minimised structures were then used to feed high-level DFT calculations starting firstly with B3LYP and the 3-21G basis set. The complexity of the basis set was then increased and the results compared to the previous calculations. The 6-311G basis set was deemed sufficient for comparison purposes. Simple MM⁺ calculations were performed using Chem3D and conformations were sampled using the dihedral angle driver and minimization function. Dynamic simulations used a step interval of 2 fs and a frame interval of 10 fs. The heating/cooling rate was set at 1 kcal/atom/ps and a target temperature of 300 K.

Synthesis of compounds **3** and **D3**

Anhydrous DCM (150 mL) was added to a flask (F1) containing **1** (6.2 g, 11.3 mmol) and **TetraEGdiOTs** (5.7 g, 11.3 mmol) under N₂. In a separate flask (F2), anhydrous MeCN (50 mL) was added to K₂CO₃ (4.7 g, 34.0 mmol) and KPF₆ (3.1 g, 16.8 mmol). F2 was heated to reflux and was kept under N₂. The solution in F1 was then added dropwise to F2 *via* a syringe

pump over 3 days. The reaction mixture was vigorously stirred for another 6 days under N₂ and reflux conditions. The flasks were both covered in aluminium foil throughout the 9-days-long reaction with TLC monitoring. At day 9, the reaction mixture was cooled to RT and the suspension was filtered and washed with DCM. The filtrate was dried under reduced pressure. The resulting residue was re-dissolved with DCM and washed with H₂O. The combined organic layer was rapidly flushed through a silica plug with DCM as eluent and gradually switched to ethyl acetate (EA). All of the flushes were combined and dried under reduce pressure to yield a viscous oil. The crude material was then dry packed into a silica column with DCM and was eluted with petroleum ether:EA (5:1), slowly increasing the ratio to 4:1. From the viscous oil, several products (**3** and **D3**) were isolated and identified. The isolated **3** was recrystallized from hot MeOH (1 mg **3** in 0.4 mL hot MeOH) to yield pure **3** (0.80 g, 1.1 mmol, 10%) in the form of sugar-like rectangular crystals while **D3** (1.60 g, 1.1 mmol, 19%) was isolated as a viscous oil. Crystals of **3** suitable for X-ray crystallography were grown from hot MeOH upon slow cooling.

Analytical data 3. ¹H NMR (700 MHz, CDCl₃) δ 7.29–7.26 (m, 4H, H8), 6.86–6.83 (m, 4H, H7), 4.73 (ddd, $J = 13.0, 9.0, 1.3 \text{ Hz}$, 2H, H1, H1'), 4.17 (dq, $J = 10.9, 7.1 \text{ Hz}$, 2H, H5), 4.05 (dq, $J = 10.9, 7.1 \text{ Hz}$, 2H, H5), 3.93 (ddd, $J = 13.1, 3.5, 1.5 \text{ Hz}$, 2H, H1, H1'), 3.76 (ddd, $J = 10.7, 9.1, 1.5 \text{ Hz}$, 2H, H2, H2'), 3.67–3.63 (m, 4H, H4, H4', H2, H2'), 3.61 (ddd, $J = 10.0, 5.7, 2.2 \text{ Hz}$, 2H, H3, H3'), 3.51 (ddd, $J = 11.8, 5.8, 2.2 \text{ Hz}$, 2H, H4, H4'), 3.47 (ddd, $J = 9.3, 6.7, 2.2 \text{ Hz}$, 2H, H3, H3'), 1.28 (s, 18H, H9), 0.99 (t, $J = 7.1 \text{ Hz}$, 6H, H6). ¹³C NMR (176 MHz, CDCl₃) δ 164.50 (C16), 155.33 (C12), 145.08 (C11, C15), 140.79 (C13), 126.28 (C8), 125.93 (C14), 115.13 (C7), 72.13 (C2, C2'), 71.98 (C3, C3'), 71.60 (C1, C1'), 70.90 (C4, C4'), 61.58 (C5), 34.32 (C10), 31.66 (C9), 13.88 (C6). FT-IR (ν_{max} , cm⁻¹) 2956 (asymmetric C–H stretching vibration from –CH₂–, –CH₃), 1737 (aromatic ester C=O stretching vibration), 1507 (aromatic C=C stretching vibration), 1212, 1179 (aromatic ester C–O stretching vibrations), 838, 811 (aromatic C–H out of plane bending vibrations). (+)nanoESI-FTMS (m/z): found $[\text{M} + \text{K}]^+$ 747.3126, calcd for C₄₀H₅₂O₁₁K: 221.0608 and $[\text{M} + \text{NH}_4]^+$ 726.3834, calcd for C₄₀H₅₆O₁₁N: 726.3848.

Analytical data D3. ¹H NMR (700 MHz, CDCl₃) δ 7.28–7.24 (m, 8H, H8), 6.85–6.81 (m, 8H, H7), 4.19 (t, $J = 5.4 \text{ Hz}$, 8H, H1), 4.12 (q, $J = 7.1 \text{ Hz}$, 8H, H5), 3.57 (t, $J = 5.3 \text{ Hz}$, 8H, H2), 3.51–3.46 (m, 16H, H3, H4), 1.28 (s, 36H, H9), 1.03 (t, $J = 7.1 \text{ Hz}$, 12H, H6). ¹³C NMR (176 MHz, CDCl₃) δ 163.85 (C16), 155.51 (C12), 145.41 (C15), 145.23 (C11), 141.69 (C13), 126.66 (C14), 126.31 (C8), 115.22 (C7), 73.07 (C1), 70.73, 70.61 (C3, C4), 69.99 (C2), 61.78 (C5), 34.33 (C10), 31.66 (C9), 13.92 (C6). FT-IR (ν_{max} , cm⁻¹) 2960 (asymmetric C–H stretching vibration from –CH₂–, –CH₃), 1733 (aromatic ester C=O stretching vibration), 1507 (aromatic C=C stretching vibration), 1212, 1177 (aromatic ester C–O stretching vibrations), 836, 815 (aromatic C–H out of plane bending vibrations). (+)nanoESI-FTMS (m/z): found $[\text{M} + \text{NH}_4]^+$ 1434.7349, calcd for C₈₀H₁₀₈O₂₂N: 1434.7358.



Synthesis of M1

Ethylene glycol (26 mL, 0.5 mol, excess) was added to a flask containing **PI** (1.0 g, 1.8 mmol) and $\text{Zn}(\text{OAc})_2 \cdot 2\text{H}_2\text{O}$ (50.0 mg, 0.2 mmol). The distillation set-up was covered in aluminium foil and was heated to 175 °C with a sand bath for 17 hours with TLC monitoring. The reaction mixture was cooled to RT. The suspension was suction filtered, the resulting solid was rinsed with H_2O and left to dry. The solid was dissolved in DCM and quickly flushed through a silica plug with DCM/EA. The flushes were collected and dried under reduced pressure. The resulting solid was dry packed into a silica column with DCM:petroleum ether (1:1) and was eluted with the same solvent gradually increasing to DCM and finally DCM with 5% MeOH. Note that this column chromatography had to be done quickly as the compound seemed to decompose on silica. Alternative purification attempts with alumina column and recrystallisation were both unsuccessful. The purification yielded greenish yellow crystals (0.25 g, 0.4 mmol, 24%). ^1H NMR (400 MHz, CDCl_3) δ 10.27 (s, 2H, H6), 7.41–7.34 (m, 4H, H4), 6.88–6.81 (m, 4H, H3), 4.30–4.25 (m, 4H, H1), 3.55–3.49 (m, 4H, H2), 1.31 (s, 18H, H5). ^{13}C NMR (101 MHz, CDCl_3) δ 168.59 (C8), 155.96 (C12), 147.41 (C11), 145.46 (C13), 138.69 (C10), 126.88 (C4), 114.79 (C9), 113.68 (C3), 68.53 (C1), 60.45 (C2), 34.40 (C14), 31.59 (C5). FT-IR (ν_{max} , cm^{-1}) 3574 (alcohol O–H stretching vibration), 2960 (asymmetric C–H stretching vibration from $-\text{CH}_2-$, $-\text{CH}_3$), 1676 (aromatic ester C=O stretching vibration), 1507 (aromatic C=C stretching vibration), 1227, 1174 (aromatic ester C–O stretching vibrations), 826, 721 (aromatic C–H out of plane bending vibrations). (+)nanoESI-FTMS (m/z): found $[\text{M} + \text{H}]^+$ 583.2538, calcd for $\text{C}_{32}\text{H}_{39}\text{O}_{10}$: 583.2538 and $[\text{M} + \text{NH}_4]^+$ 600.2798, calcd for $\text{C}_{32}\text{H}_{42}\text{O}_{10}\text{N}$: 600.2803. Crystals suitable for X-ray crystallography were grown in the fridge *via* vapour diffusion from chloroform/hexane. X-ray crystal structure and refinement data are available in Table 1 of ESI.†

Synthesis of 2

CH_3I (0.8 mL, 12.9 mmol) was added dropwise into a flask containing **1** (1.5 g, 2.7 mmol) and K_2CO_3 (1.9 g, 13.7 mmol) pre-dissolved in acetone (106 mL) at reflux. The reaction mixture was left to stir for 1 hour with TLC monitoring. It was then cooled to RT. The suspension was filtered and the filtrate was dried under reduced pressure. The crude was recrystallized with hot hexane to form colourless sugar-like tiny cube crystals (1.03 g, 1.8 mmol, 67%). ^1H NMR (300 MHz, CDCl_3) δ 7.34–7.22 (m, 4H, H4), 6.86–6.80 (m, 4H, H3), 4.16 (q, J = 7.1 Hz, 4H, H1), 3.81 (s, 6H, H6), 1.29 (s, 18H, H5), 1.08 (t, J = 7.1 Hz, 6H, H2). ^{13}C NMR (75 MHz, CDCl_3) δ 164.00 (C10), 155.58 (C11), 146.48 (C7), 145.40 (C12), 142.04 (C8), 126.84 (C9), 126.34 (C4), 115.11 (C3), 62.08 (C6), 61.89 (C1), 34.34 (C13), 31.63 (C5), 13.96 (C2). FT-IR (ν_{max} , cm^{-1}) 2959 (asymmetric C–H stretching vibration from $-\text{CH}_2-$, $-\text{CH}_3$), 1736 (aromatic ester C=O stretching vibration), 1507 (aromatic C=C stretching vibration), 1205, 1170 (aromatic ester C–O stretching vibrations), 821 (aromatic C–H out of plane bending vibration).

(+)nanoESI-FTMS (m/z): found $[\text{M} + \text{H}]^+$ 579.2946, calcd for $\text{C}_{34}\text{H}_{43}\text{O}_8$: 579.2952. Crystals suitable for X-ray crystallography were grown from acetone at RT. X-ray crystal structure and refinement data are available in Table 2 of ESI.†

Synthesis of M2

1,4-Butandiol (35 mL, 0.4 mol, excess) was added to a flask containing **2** (1.8 g, 3.1 mmol) and $\text{Ti}(\text{O}^n\text{Bu})_4$ (0.3 mL, 0.9 mmol). The distillation set-up was covered in aluminium foil and was heated to 220 °C with a sand bath for 76 hours with TLC monitoring. The reaction mixture was cooled to RT. H_2O was added to the reaction mixture until a suspension formed before it was suction filtered. The residue was allowed to dry and was dissolved in DCM. The solution was quickly flushed through silica plug with DCM/EA. The first flush containing unreacted **2** was discarded while the subsequent flushes were combined and dried under reduced pressure. The resulting viscous oil was wet packed into a silica column with minimum amount of DCM and was eluted with DCM:EA (19:1) gradually increasing the polarity to 4:1. The fractions were collected in vials and were combined based on TLC. From the crude, several products were isolated and identified as follows: **M2** (0.21 g, 0.3 mmol, 10%) and its asymmetric form **M4** (0.35 g, 0.6 mmol, 19%). Both products were isolated in the form of a brown oil.

Analytical data M2. ^1H NMR (400 MHz, CDCl_3) δ 7.31–7.25 (m, 4H, H4), 6.85–6.80 (m, 4H, H3), 4.15 (t, J = 6.4 Hz, 4H, H1), 3.78 (s, 6H, H6), 3.46 (t, J = 6.3 Hz, 4H, H15), 2.61 (s, 2H, H16), 1.59–1.50 (m, 4H, H2), 1.44–1.35 (m, 4H, H14), 1.28 (s, 18H, H5). ^{13}C NMR (101 MHz, CDCl_3) δ 164.02 (C10), 155.46 (C11), 146.29 (C7), 145.45 (C12), 141.82 (C8), 126.79 (C9), 126.34 (C4), 114.96 (C3), 65.65 (C1), 62.19 (C15), 62.01 (C6), 34.29 (C13), 31.59 (C5), 28.84 (C14), 24.89 (C2). FT-IR (ν_{max} , cm^{-1}) 3596 (alcohol O–H stretching vibration), 2959 (asymmetric C–H stretching vibration from $-\text{CH}_2-$, $-\text{CH}_3$), 1732 (aromatic ester C=O stretching vibration), 1507 (aromatic C=C stretching vibration), 1213, 1170 (aromatic ester C–O stretching vibrations), 830 (aromatic C–H out of plane bending vibration), 733 (C–C skeletal rocking vibration of $-(\text{CH}_2)_2-$ and $-(\text{CH}_2)_3-$). (+)nanoESI-FTMS (m/z): found $[\text{M} + \text{Na}]^+$ 689.3285, calcd for $\text{C}_{38}\text{H}_{50}\text{O}_{10}\text{Na}$: 689.3296 and $[\text{M} + \text{NH}_4]^+$ 684.3737, calcd for $\text{C}_{38}\text{H}_{54}\text{O}_{10}\text{N}$: 684.3742.

Analytical data M4. ^1H NMR (400 MHz, CDCl_3) δ 7.31–7.27 (m, 4H, H4, H4'), 6.86–6.81 (m, 4H, H3', H3), 4.21–4.12 (m, 4H, H1, H1'), 3.81 (s, 3H, H6'), 3.78 (s, 3H, H6), 3.49 (t, J = 6.3 Hz, 2H, H15), 1.62–1.52 (m, 2H, H2), 1.47–1.37 (m, 2H, H14), 1.30–1.28 (m, 18H, H5', H5), 1.07 (t, J = 7.1 Hz, 3H, H2'). ^{13}C NMR (101 MHz, CDCl_3) δ 164.10 (C10), 163.93 (C10'), 155.56 (C11, C11'), 146.47 (C7'), 146.37 (C7), 145.47, 145.44 (C12, C12'), 142.07 (C8'), 141.85 (C8), 126.93 (C9'), 126.77 (C9), 126.37, 126.35 (C4', C4), 115.13 (C3'), 115.01 (C3), 65.67 (C1), 62.34 (C15), 62.12 (C6'), 62.02 (C6), 61.89 (C1'), 34.35, 34.34 (C13', C13), 31.64 (C5, C5'), 29.03 (C14), 24.99 (C2), 13.94 (C2'). FT-IR (ν_{max} , cm^{-1}) 3596 (alcohol O–H stretching vibration), 2959 (asymmetric C–H stretching vibration from $-\text{CH}_2-$, $-\text{CH}_3$), 1736 (aromatic ester C=O stretching vibration), 1507



(aromatic C=C stretching vibration), 1213, 1170 (aromatic ester C=O stretching vibrations), 830 (aromatic C-H out of plane bending vibration), 733 (C-C skeletal rocking vibration of $-(CH_2)_2-$ and $-(CH_2)_3-$). (+)nanoESI-FTMS (m/z): found $[M + H]^+$ 623.3206, calcd for $C_{36}H_{47}O_9$: 623.3215.

Synthesis of M3

1,4-Butandiol (7.3 mL, 0.1 mol, excess) was added to a flask containing **3** (0.4 g, 0.6 mmol) and $Ti(O^iBu)_4$ (80 μ L, 0.2 mmol). The distillation set-up was covered in aluminium foil and was heated to 220 °C with a sand bath for 18 hours with TLC monitoring. The reaction mixture was cooled to RT. H_2O was added to the reaction mixture until a suspension formed before it was suction filtered. The residue was allowed to dry and was wet packed into a silica column with a minimum amount of DCM and was eluted with DCM with 0.5% MeOH gradually increasing polarity to DCM with 3.5% MeOH. The fractions were collected in vials and were combined based on TLC. From the crude, several products were isolated and identified as follows: **M3** (0.20 g, 0.3 mmol, 43%) and its asymmetric form **M5** (0.13 g, 0.2 mmol, 31%).

Analytical data M3. 1H NMR (500 MHz, $CDCl_3$) δ 7.33–7.26 (m, 4H, H8), 6.86–6.82 (m, 4H, H7), 4.69 (ddd, $J = 13.0, 8.9, 1.3$ Hz, 2H, H1, H1'), 4.20 (ddd, $J = 10.9, 6.8, 5.9$ Hz, 2H, H5), 4.07 (ddd, $J = 10.9, 6.7, 5.9$ Hz, 2H, H5), 3.91 (ddd, $J = 13.0, 3.6, 1.5$ Hz, 2H, H1, H1'), 3.80–3.71 (m, 2H, H2, H2'), 3.66–3.55 (m, 6H, H2, H2', H4, H4', H3, H3'), 3.54–3.41 (m, 8H, H4, H4', H3, H3', H18), 1.59 (s, 2H, H19), 1.55–1.43 (m, 4H, H6), 1.41–1.31 (m, 4H, H17), 1.29 (s, 18H, H9). ^{13}C NMR (126 MHz, $CDCl_3$) δ 164.47 (C16), 155.29 (C12), 145.24 (C11), 144.95 (C15), 140.59 (C13), 126.37 (C8), 125.98 (C14), 115.01 (C7), 72.00 (C2, C2'), 71.94 (C3, C3'), 71.58 (C1, C1'), 70.85 (C4, C4'), 65.43 (C5), 62.35 (C18), 34.35 (C10), 31.67 (C9), 29.09 (C17), 24.90 (C6). FT-IR (ν_{max} , cm^{-1}) 3337 (alcohol O-H stretching vibration), 2953 (asymmetric C-H stretching vibration from $-CH_2-$, $-CH_3$), 1733 (aromatic ester C=O stretching vibration), 1507 (aromatic C=C stretching vibration), 1212, 1179 (aromatic ester C-O stretching vibrations), 837, 811 (aromatic C-H out of plane bending vibrations). (+)nanoESI-FTMS (m/z): found $[M + NH_4]^+$ 814.4367, calcd for $C_{44}H_{64}O_{13}N$: 814.4372.

Analytical data M5. 1H NMR (500 MHz, $CDCl_3$) δ 7.30–7.26 (m, 4H, H8, H8'), 6.87–6.81 (m, 4H, H7, H7'), 4.71 (dddd, $J = 18.8, 13.0, 9.0, 1.3$ Hz, 2H, H1, H1'), 4.24–4.12 (m, 2H, H5, H5'), 4.11–4.00 (m, 2H, H5, H5'), 3.92 (dddd, $J = 17.5, 13.0, 3.5, 1.5$ Hz, 2H, H1, H1'), 3.76 (dddd, $J = 10.5, 8.8, 4.8, 1.4$ Hz, 2H, H2, H2'), 3.68–3.57 (m, 6H, H2, H2', H4, H4', H3, H3'), 3.55–3.41 (m, 6H, H4, H4', H3, H3', H18), 1.54–1.44 (m, 2H, H6), 1.42–1.32 (m, 2H, H17), 1.29 (s, 18H, H9, H9'), 1.11 (s, 1H, H19), 0.99 (t, $J = 7.1$ Hz, 3H, H6'). ^{13}C NMR (126 MHz, $CDCl_3$) δ 164.55 (C16'), 164.41 (C16), 155.32 (C12, C12'), 145.18, 145.13 (C11, C11'), 144.92 (C15, C15'), 140.87 (C13'), 140.53 (C13), 126.35, 126.30 (C8, C8'), 126.02 (C14), 125.91 (C14'), 115.16 (C7'), 115.01 (C7), 72.06 (C2, C2'), 72.01 (C3, C3'), 71.61, 71.59 (C1, C1'), 70.93, 70.85 (C4, C4'), 65.39 (C5), 62.36 (C18), 61.59 (C5'), 34.34 (C10, C10'), 31.67 (C9, C9'), 29.10 (C17), 24.91 (C6), 13.88 (C6'). FT-IR (ν_{max} , cm^{-1}) 3337

(alcohol O-H stretching vibration), 2956 (asymmetric C-H stretching vibration from $-CH_2-$, $-CH_3$), 1737 (aromatic ester C=O stretching vibration), 1507 (aromatic C=C stretching vibration), 1212, 1179 (aromatic ester C-O stretching vibrations), 837, 811 (aromatic C-H out of plane bending vibrations). (+)nanoESI-FTMS (m/z): found $[M + NH_4]^+$ 770.4103, calcd for $C_{42}H_{60}O_{12}N$: 770.4110.

Synthesis of PolyM1

M1 (250.0 mg, 0.4 mmol) was dissolved in anhydrous THF (5 mL) at RT. Upon complete solubilisation, the solution was cooled to 0 °C in an ice bath. TEA (180 μ L, 1.3 mmol) was added. In a separate flask, terephthaloyl chloride (87.0 mg, 0.4 mmol) was dissolved in anhydrous THF (5 mL). The solution was then added dropwise into the reaction mixture *via* a syringe pump over the course of 1 hour. The reaction mixture was allowed to react at RT over 3 days covered in aluminium foil. The reaction mixture was then suction filtered and the filtrate was dried under reduced pressure. The residue was purified by re-dissolving in $CHCl_3$ and added to Et_2O . The precipitate was dissolved in small amounts of $CHCl_3$ and allowed to dry to yield a semi-opaque pale yellow film (61.0 mg, 7.2 μ mol, 2%).

Synthesis of PolyM2

M2 (250.0 mg, 0.4 mmol) was dissolved in anhydrous THF (5 mL) at RT. Upon complete solubilisation, the solution was cooled to 0 °C in an ice bath. TEA (160 μ L, 1.1 mmol) was added. In a separate flask, terephthaloyl chloride (76.0 mg, 0.4 mmol) was dissolved in anhydrous THF (4 mL). The solution was then added dropwise into the reaction mixture *via* a syringe pump over the course of 1 hour. The reaction mixture was allowed to react at RT over 4 days covered in aluminium foil. The reaction mixture was then suction filtered and the filtrate was dried under reduced pressure. The residue was purified by re-dissolving in $CHCl_3$ and added to Et_2O . The precipitate was dissolved in small amounts of $CHCl_3$ and allowed to dry to yield an off-white opaque brittle film (28.00 mg) which was found to be a mixture. The filtrate from the purification was dried under reduced pressure, re-dissolved in $CHCl_3$ and re-precipitated in hexane twice to yield **PolyM2**, similarly being off-white, opaque and brittle (263.00 mg, 33.6 μ mol, 8%).

Synthesis of PolyM3

M3 (115.0 mg, 0.1 mmol) was dissolved in anhydrous THF (2 mL) at RT. Upon complete solubilisation, the solution was cooled to 0 °C in an ice bath. TEA (60 μ L, 0.4 mmol) pre-dissolved in anhydrous THF (0.8 mL) was added. In a separate flask, terephthaloyl chloride (29.0 mg, 0.1 mmol) was dissolved in anhydrous THF (1 mL). The solution was then added dropwise into the reaction mixture *via* a syringe over the course of 1 hour. The reaction mixture was allowed to react at RT over 6 days covered in aluminium foil. The reaction mixture was then filtered through a pipette cotton plug and the filtrate was dried under reduced pressure. The residue was purified by dissolving in THF and added to $CHCl_3$. The filtrate was dried



under reduced pressure to yield **PolyM3** as an off-white opaque polymer film (95.8 mg).

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We thank the mass spectrometry service at Swansea University for collecting mass spectra and Newcastle University for financial support. Dr C. Wills is thanked for collecting the DOSY spectra. Financial support from the Spanish Ministerio de Ciencia, Innovación y Universidades (ENE2017-86711-C3-3-R) is gratefully acknowledged.

Notes and references

- 1 B. Lepoittevin and P. Roger, *Handbook of Engineering and Speciality Thermoplastics: Polyethers and Polyesters*, ed. S. Thomas and P. M. Visakh, Wiley, Hoboken, 2011, p. 97.
- 2 J. Pang, M. Zheng, R. Sun, A. Wang, X. Wang and T. Zhang, *Green Chem.*, 2016, **18**, 342.
- 3 K. Ravindranath and R. A. Mashelka, *Chem. Eng. Sci.*, 1986, **41**, 2197.
- 4 D. Giovanelli and E. Farella, *J. Sens.*, 2016, 9391850.
- 5 M. Y. Lee, J. Hong, E. K. Lee, H. Yu, H. Kim, J. U. Lee, W. Lee and J. H. Oh, *Adv. Chem. Mater.*, 2016, **26**, 1445.
- 6 K. Peng, B. Wang and C. Ji, *J. Appl. Polym. Sci.*, 2017, **134**, 44907.
- 7 T. Kuwabara, T. Nakashima, T. Yamaguchi and K. Takahashi, *Org. Electron.*, 2012, **13**, 1136.
- 8 K. Hong and J.-L. Lee, *J. Electrochem. Soc.*, 2007, **154**, H782.
- 9 G. Kocak, C. Tuncera and V. Bütün, *J. Polym. Chem.*, 2017, **8**, 144.
- 10 A. Seeboth, D. Löttsch, R. Ruhmann and O. Muehling, *Chem. Rev.*, 2014, **114**, 3037.
- 11 (a) G. Wegner, *Angew. Chem., Int. Ed.*, 1981, **20**, 361; (b) G. Inzelt, *J. Electrochem. Sci. Eng.*, 2018, **8**, 3.
- 12 P. Wang, X. Wang, Y. Ling, M. Wang, S. Ding, W. Shen, Z. Wang, Y. Wang and F. Liu, *Radiat. Meas.*, 2018, **119**, 80.
- 13 D. Gopalakrishnan and W. R. Dichtel, *J. Am. Chem. Soc.*, 2013, **135**, 14853.
- 14 S. J. Peighambaroust, S. Rowshanzamir and M. Amjadi, *Int. J. Hydrogen Energy*, 2010, **35**, 9349.
- 15 Y. Salinas, R. Martínez-Máñez, M. D. Marcos, F. Sancenón, A. M. Costero, M. Parraad and S. Gilad, *Chem. Soc. Rev.*, 2012, **41**, 1261.
- 16 A. C. Benniston, T. P. L. Winstanley, H. Lemmetyinen, N. V. Tkachenko, R. W. Harrington and C. Wills, *Org. Lett.*, 2012, **14**, 1374.
- 17 Software used in the Cambridge Crystallographic Data Centre.
- 18 P. Liebing, A. Zaeni, F. Olbrich and F. T. Edelmann, *Acta Crystallogr., Sect. E: Crystallogr. Commun.*, 2016, **72**, 1757.
- 19 D. L. Clark, J. C. Gordon, J. C. Huffman, R. L. Vincent-Hollis, J. G. Watkin and B. D. Zwick, *Inorg. Chem.*, 1994, **33**, 5903.
- 20 (a) Y. Chiang, J. Kresge and J. Jones, *J. Am. Chem. Soc.*, 1994, **116**, 8358–8359; (b) J. M. A. Spiteri, C. J. Mallia, G. J. Scerri and D. C. Magri, *Org. Biomol. Chem.*, 2017, **15**, 10116–10121; (c) T. Terashima, M. Kawabe, Y. Miyabara, H. Yoda and M. Sawamoto, *Nat. Commun.*, 2013, **4**, 2321; (d) J. B. Smith, S. H. Kerr, P. S. White and A. J. M. Miller, *Organometallics*, 2017, **36**, 3094–3103; (e) F. Arnaud-Neu, R. Delgado and S. Chaves, *Pure Appl. Chem.*, 2003, **75**, 71–102; (f) R. B. Sharma, P. Kebarle and A. T. Blades, *J. Am. Chem. Soc.*, 1984, **106**, 510–516; (g) J. C. Chambron and M. Meyer, *Chem. Soc. Rev.*, 2009, **38**, 1663–1673.
- 21 J. Lui, S.-G. Bian, M. Xiao, S.-J. Wang and Y.-Z. Meng, *J. Appl. Polym. Sci.*, 2010, **115**, 3401.
- 22 X. Yang, X. Xu and G. Zhou, *J. Mater. Chem. C*, 2015, **3**, 913.
- 23 (a) A. Marchesi, S. Brenna and G. A. Ardizzoia, *Dyes Pigm.*, 2019, **161**, 457; (b) A. K. Gupta, A. Kumar, R. Singh, M. Devi, A. Dhir and C. P. Pradeep, *ACS Omega*, 2018, **3**, 14341.
- 24 F. Hammerer, G. Garcia, P. Charles, A. Sourdon, S. Achelle, M.-P. Teulade-Fichou and P. Maillard, *Chem. Commun.*, 2014, **50**, 9529.
- 25 B. L. Allwood, N. Spencer, H. Shahriari-Zavareh, J. F. Stoddart and D. J. Williams, *J. Chem. Soc., Chem. Commun.*, 1987, 1061.
- 26 B. L. Allwood, N. Spencer, H. Shahriari-Zavareh, J. F. Stoddart and D. J. Williams, *J. Chem. Soc., Chem. Commun.*, 1987, 1064.
- 27 O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard and H. Puschmann, *J. Appl. Crystallogr.*, 2009, **42**, 339.
- 28 G. M. Sheldrick, *Acta Crystallogr., Sect. A: Found. Adv.*, 2015, **71**, 3.
- 29 G. M. Sheldrick, *Acta Crystallogr., Sect. A: Found. Crystallogr.*, 2008, **64**, 112.
- 30 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, *Gaussian 09, Revision D.01*, Gaussian, Inc., Wallingford CT, 2013.

