



Cite this: *Org. Biomol. Chem.*, 2020, **18**, 1957

Chiral 1,5-disubstituted 1,2,3-triazoles – versatile tools for foldamers and peptidomimetic applications†

Anna Said Stålsmeden, ^a Andrew J. Paterson, ^a Imola Cs. Szigyártó, ^b Linda Thunberg, ^c Johan R. Johansson, ^{*d} Tamás Beke-Somfai ^{*b} and Nina Kann ^{*a}

1,4- and 1,5-Disubstituted triazole amino acid monomers have gained increasing interest among peptidic foldamers, as they are easily prepared *via* Cu- and Ru-catalyzed click reactions, with the potential for side chain variation. While the latter is key to their applicability, the synthesis and structural properties of the chiral mono- or disubstituted triazole amino acids have only been partially addressed. We here present the synthesis of all eight possible chiral derivatives of a triazole monomer prepared *via* a ruthenium-catalyzed azide alkyne cycloaddition (RuAAC). To evaluate the conformational properties of the individual building units, a systematic quantum chemical study was performed on all monomers, indicating their capacity to form several low energy conformers. This feature may be used to effect structural diversity when the monomers are inserted into various peptide sequences. We envisage that these results will facilitate new applications for these artificial oligomeric compounds in diverse areas, ranging from pharmaceuticals to biotechnology.

Received 24th January 2020,
Accepted 18th February 2020

DOI: 10.1039/d0ob00168f

rsc.li/obc

Introduction

The structural and functional properties of natural biomolecules can be mimicked by synthetic compounds. Such man-made molecules can retain the beneficial properties of their natural counterparts while also benefiting from additional useful attributes. They can provide increased stability against enzymatic degradation,¹ better designability, and diverse side-chain chemistry,² characteristics that are often desired for novel bioactive molecules. Consequently, peptidomimetics and protein engineering often employ synthetic, non-natural, amino acid-derived insertions to improve characteristics of natural compounds.³

In the last two decades, it has become apparent that peptidic oligomers containing, or composed almost entirely of, non-natural amino acids can fold into well-defined secondary structures, leading to the emerging field of peptidic foldamers, or foldamers.^{3c,4} Since the initial studies demonstrating helical oligomers containing homologated derivatives of natural amino acids,^{2,5} a wide variety of compounds have been investigated in this direction.^{3c} Numerous reports have also pointed out that these compounds will have a propensity to fold into secondary structures. Furthermore they can provide antibacterial,⁶ antiviral,⁷ and cell-penetrating activity,⁸ as well as having self-assembling properties.⁹ This may lead to applications in pharmaceuticals or bionanotechnology.

One advantage of constructing foldamers from non-natural amino acids is that standard techniques for peptide coupling can be applied for assembling these monomers into a sequence.^{3c,10} Such monomers may include synthetic amino acids based on a cycloalkyl, aromatic or heteroaromatic scaffold.¹¹ 1,4-Disubstituted 1,2,3-triazoles, easily accessible *via* the copper-catalyzed azide alkyne cycloaddition (CuAAC) reaction,¹² have also been applied in this context. Examples include oligo(phenyl-amide-triazole)-type foldamers that function as halide ion receptors,¹³ oxazolidinone-triazole units incorporated into dipeptide mimics for use as foldamer building blocks.¹⁴ as well as other examples of peptide backbone modifications using 1,4-disubstituted 1,2,3-triazoles.¹⁵ In con-

^aDepartment of Chemistry and Chemical Engineering, Chalmers University of Technology, SE-41296 Göteborg, Sweden. E-mail: kann@chalmers.se

^bResearch Centre for Natural Sciences, Hungarian Academy of Sciences, Magyar tudósok krt. 2, H-1117 Budapest, Hungary. E-mail: beke-somfai.tamas@ttk.mta.hu

^cEarly Chemical Development, Pharmaceutical Sciences, BioPharmaceuticals R&D, AstraZeneca, Gothenburg, Sweden

^dMedicinal Chemistry, Research and Early Development Cardiovascular, Renal and Metabolism, BioPharmaceuticals R&D, AstraZeneca, Gothenburg, Sweden.

E-mail: johan.x.johansson@astrazeneca.com

†Electronic supplementary information (ESI) available: Computational data, determination of enantiomeric excess for monomers, ¹H and ¹³C NMR spectra. See DOI: 10.1039/d0ob00168f



trast to CuAAC, the ruthenium-catalyzed cycloaddition reaction (RuAAC) instead provides access to the 1,5-disubstituted 1,2,3-triazole with a different spatial arrangement of the carboxylic acid and amino functionality used for linking the monomeric units.¹⁶ Homo- and heterochiral peptidomimetics have been developed from enantiomerically pure propargylamines and α -azido acids using the RuAAC reaction.¹⁷ NMR and MD simulations have shown the presence of a β -turn-like structure in homochiral peptido-triazolamers, while a heterochiral triazolamer shows a polyproline-like helix. Moreover, a well defined protocol for the synthesis of chiral propargylamines, with a side chain in the α -position, is feasible using Ellman's *tert*-butylsulfonamide as a chiral auxiliary.¹⁸ The RuAAC reaction has been frequently utilized in medicinal chemistry applications,¹⁹ including the synthesis of peptidomimetics,²⁰ but its use in the area of foldamers is as yet very limited. In earlier reports from our group, we have developed microwave-assisted methodology for the synthesis of 1,5-disubstituted 1,2,3-triazole monomers *via* RuAAC and also investigated the structures and conformations of δ -peptides prepared from these monomers.²¹ In addition, we have compared the conformational properties of triazole monomers prepared *via* CuAAC and RuAAC.²² In this study, we instead focus on the conformational effects of introducing chiral substituents, on either side or both sides of the triazole. During the preparation of this manuscript, a related report, concerning the synthesis of some 1,5-disubstituted peptidotriazolomers, was published by Sewald and co-workers.¹⁷ While there are some touching points between the two studies, the present study describes the complete set of eight possible combinations for mono- and disubstituted chiral derivatives. Thus, these studies are different both in terms of the triazoles prepared, as well as with respect to the computational studies performed. Our results from these investigations are described herein.

Results and discussion

Monomer synthesis

The chiral alkyne **1** and azide **2** (Fig. 1) used in this study originate from the amino acid alanine. Employing both enantiomers, in combination with the corresponding non-chiral non-methylated *N*-Boc propargylamine **3** and methyl 2-azidoacetate

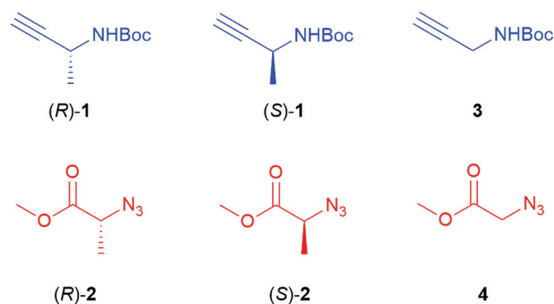


Fig. 1 Chiral and non-chiral alkynes and azides used in this study.

4, allowed the synthesis of eight different stereoisomers of the triazole derivatives (Fig. 2). The chiral alkynes (*R*)-**1** and (*S*)-**1** were initially prepared *via* a Bestmann–Ohira reaction²³ from Boc-protected (*R*)- and (*S*)-alaninal, where the (*R*)-enantiomer was obtained *via* Dess–Martin oxidation²⁴ of Boc-(*R*)-alaninol, and the (*S*)-form was commercially available. While both reactions are documented to proceed without causing racemisation,^{23,25} we found that erosion of the stereochemistry had taken place for both enantiomers of **1** in our case. Raines has reported the preparation of (*S*)-**1** *via* reduction of the corresponding Weinreb amide of alanine, followed by a Bestmann–Ohira reaction of the formed aldehyde *in situ*, and this may be a better option to avoid prolonged handling of the sensitive aldehyde.^{20b} We instead opted for using commercially available (*R*)-**1** and (*S*)-**1** for the continued studies. For compound **2**, both enantiomers were prepared *via* diazo transfer from the corresponding amine by using 2-azido-1,3-dimethylimidazolium hexafluoro-phosphate (ADMP), following a report by Kitamura *et al.*²⁶ The non-chiral derivatives **3** and **4** are both commercially available. With all starting materials in place, reaction conditions for the RuAAC coupling were then investigated. This involved screening various catalysts, including the two most commonly used for RuAAC, *i.e.* Cp*RuCl(PPh₃)₂ and Cp*RuCl(COD), as well as [Cp*RuCl₂]_n and [Cp*RuCl]₄. A solution of (*S*)-**1** and azido ester **4** in THF was heated to 60 or 80 °C for 20 minutes under microwave conditions, and products were purified by HPLC or flash chrom-

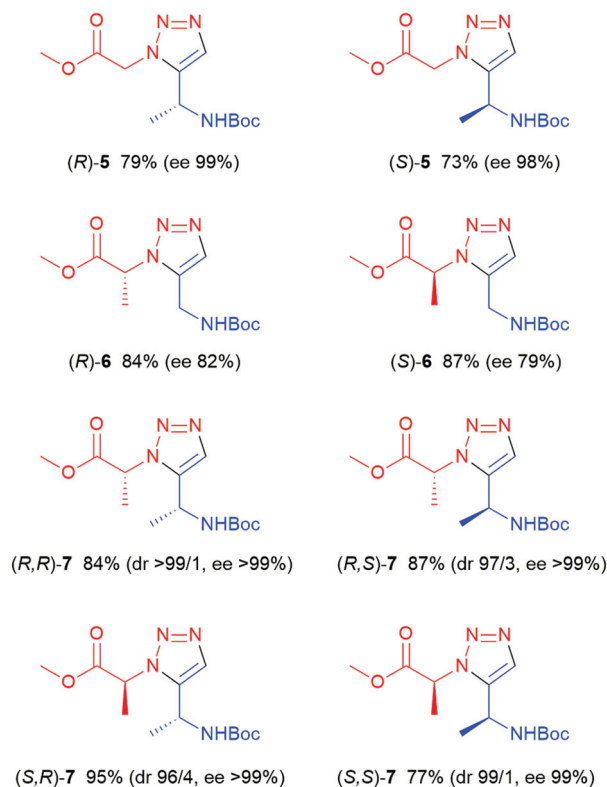
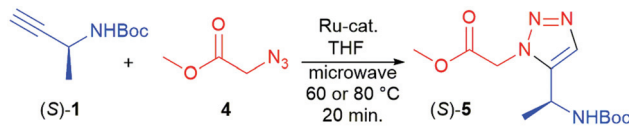


Fig. 2 Chiral 1,5-disubstituted triazoles prepared *via* RuAAC. See Experimental section for method used.



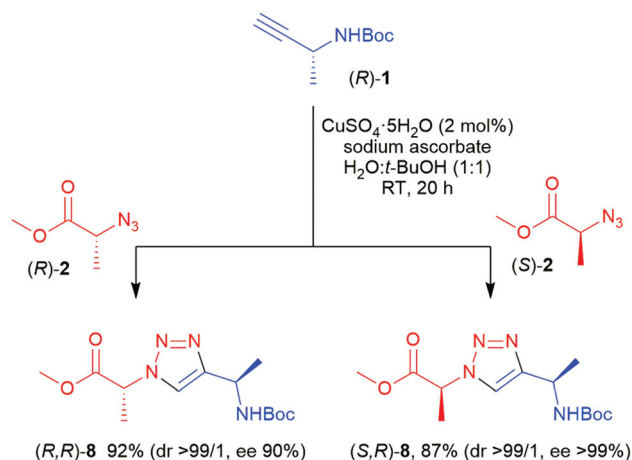
Table 1 Catalyst evaluation in the RuAAC reaction between (S)-1 and 4


Entry	Catalyst	Temp. (°C)	Yield ^a (%)	ee ^b (%)
1	Cp*RuCl(PPh ₃) ₃	80	42	97.7
2	Cp*RuCl(COD)	60	73	98.4
3	[Cp*RuCl ₂] _n	80	57	97.3
4	[Cp*RuCl] ₄	80	5	97.5

^a Isolated yield. ^b Determined by chiral SFC (see ESI† for details).

atography. The results are shown in Table 1. All the catalysts screened, with the exception of [Cp*RuCl]₄, provided triazole (S)-5 in moderate to good yields, and showed no significant differences in enantiomeric excess. The preparation of the full set of chiral triazoles was then pursued using either Cp*RuCl(COD) or [Cp*RuCl₂]_n as the catalyst.

Alkyne (R)-1 was reacted with 4 using 4 mol% [Cp*RuCl₂]_n as the catalyst, affording (R)-5, *i.e.* the enantiomer of the earlier prepared triazole (S)-5, in a somewhat lower yield (52%). Conventional heating at 100 °C, using Cp*RuCl(COD) as the catalyst, was found to be more successful in this case, affording (R)-5 in 79% yield (Fig. 2) and high enantiomeric excess (>99%). Subsequently, the (R)- and (S)-enantiomers of chiral azide 2 were applied and combined with *N*-Boc-propargylamine 3, yielding the two enantiomers of triazole 6, with the methyl substituent in the α-position relative to the ester, in 84 and 87% yield respectively. 2D NOESY NMR confirmed the 1,5-regioselectivity of the 1,2,3-triazoles (see ESI†). However, some loss of stereochemistry was found for compounds (R)-6 and (S)-6, with approximately 10% of the other enantiomer formed alongside the desired product. As the chiral centre next to the alkyne 1 remains intact during the reaction, as seen in Table 1, the cause of this partial racemisation was pinpointed to the methyl-substituted carbon of azides 2. To determine if this racemisation occurs during the synthesis of the chiral precursor azides 2, or during the RuAAC reaction itself, azides 2 were subjected to a CuAAC reaction at room temperature, using CuSO₄ as the catalyst (Scheme 1). Both 1,4-triazoles (R,R)-8 and (S,R)-8 were formed in good yields (92% and 87% respectively) and good to excellent stereoselectivity, indicating that the minor loss of stereochemistry is most likely occurring during the triazole formation itself, rather than in the preparation of 2. With these results in hand, all four stereoisomers of the dimethylated triazole 7 (Fig. 2) were prepared in the same fashion. Azide (R)-2 was first combined with both enantiomers of alkyne 1 to produce the diastereomers (R,R)- and (R,S)-7 in similar yields as for compounds 6. The same reaction starting instead from (S)-2 afforded the second set of diastereomers (S,R)- and (S,S)-7 in 95% and 77% yield, respectively. Interestingly, all these compounds were obtained with excellent enantioselectivity and good to excellent diastereo-

**Scheme 1** Chiral 1,4-substituted triazoles prepared *via* CuAAC.

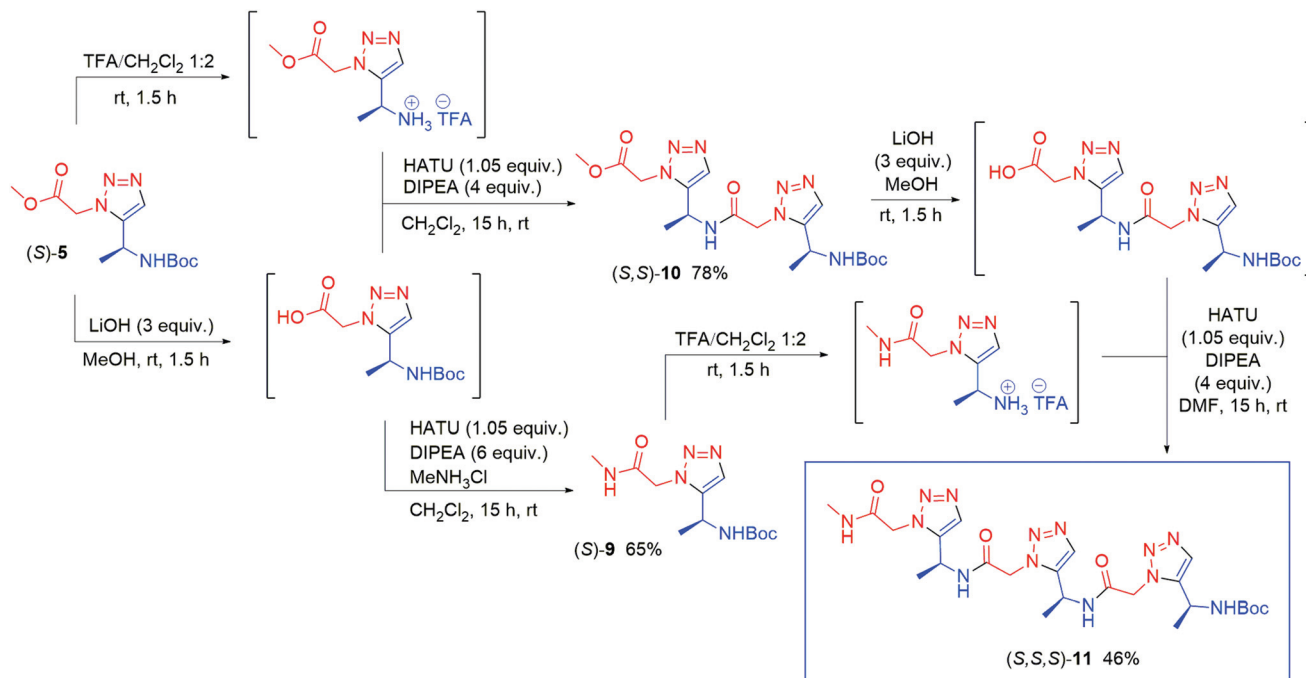
selectivity. The loss of stereochemical integrity for compounds 6 may thus be a result of the lower degree of sterical hindrance in these compounds, allowing racemisation *via* enolisation to take place, while the extra methyl group in compounds 7 may in some manner block this process. More detailed studies are needed, however, to elucidate the mechanism for this racemisation.

Trimer synthesis

To verify that the chiral triazoles prepared could be used for foldamer synthesis, a trimer was prepared from triazole (S)-5 (Scheme 2). *N*-Methyl amide (S)-9 was first synthesized *via* hydrolysis of (S)-5 with LiOH to the corresponding carboxylic acid, followed by HATU-mediated amide formation, affording (S)-9 in 65% yield. A dimer of (S)-5 was then prepared. In one experiment, (S)-5 was Boc-protected with trifluoroacetic acid, and in a second experiment, (S)-5 was again hydrolyzed with LiOH. The two parts were then coupled directly without further purification, using HATU as the coupling agent and DIPEA as the base, affording dimer (S,S)-10 in 78% yield. (S)-9 was then deprotected and subsequently coupled with hydrolysed (S,S)-10, affording trimer (S,S,S)-11 in 46% yield as a white solid. 2D NOESY NMR was performed on (S,S,S)-11 to study the conformational properties of the trimer. The ¹H-¹H NOESY was recorded at 24 mM concentration in DMSO-*d*₆, using a mixing time of 500 ms. In comparison to a corresponding non-chiral trimer reported by us earlier,²¹ with the *N*-methyl amide replaced by a methyl ester, the results are much less conclusive. Where the non-chiral structure displays a long range coupling between the Boc *tert*-butyl group and the terminal methyl group, *i.e.* between the two ends of the oligomer, indicating a turn-like conformer, no such correlation can be seen for (S,S,S)-11. This is perhaps not surprising, as a greater backbone flexibility would be expected for the corresponding non-chiral trimer.

Of more interest is a weak short-range coupling between the triazole CH signals and the methyl substituents on the chiral carbons in (S,S,S)-11, indicating a twisted conformer that allows these groups to come into proximity, rather than a





Scheme 2 Synthesis of trimer (S,S,S)-11.

structure with a straighter elongated backbone chain. Excluding expected correlations, no other interactions of interest could be discerned.

Molecular modelling

With the full set of chiral triazoles at hand, our attention was then directed towards the conformational properties of these compounds. To explore the conformational properties of the eight chiral derivatives prepared, quantum chemical (QM) calculations were performed employing three different levels of theory. QM methods are regularly employed in understanding structural and energetic properties of non-natural amino acid compounds. In contrast to natural amino acids and proteins, proper parametrization of these exotic residues is often missing, making extensive use of molecular dynamics (MD) simulations less reliable.²⁷ Previously, the potential energy hypersurface of an achiral 1,5-disubstituted 1,2,3-triazole amino acid was explored in an exhaustive scan.²² In the present study, 20 of these previously found achiral conformers were employed to construct the eight chiral models, which resulted in the general structure of Ac-5Tzl-NHMe (for more details see Computational methods, and ESI, Tables S1–S8†). Since the *R* and *S* conformers could be considered enantiomer pairs, these were tested against the same initial torsional angles from results of the achiral triazole calculations, and conformational enantiomers were not considered. This way we obtained the relative energies for only one side of the asymmetric potential energy hypersurface (PEHS) for each model, where those of the enantiomer pairs can be converted into each other by their inversion center. Therefore the obtained results are discussed for the enantiomer pairs together.

Considering the (*R*)-5 and (*S*)-5 models, respectively, 9 and 7 out of the located 18 and 17 conformers are within 3 kcal mol⁻¹ in relative energy, counted from the most stable conformer 2' and 2 (see ESI†). This indicates that in principle both derivatives have over ten conformers which have low relative energies and thus could be the structural building units of several low energy secondary structures. For (*R*)-6 and (*S*)-6, there are 6 and 11, respectively, low energy conformers, *i.e.* below 3 kcal mol⁻¹ relative energy, with the most stable conformers being 2 and 1 (see ESI†), respectively. These results indicate that the single substituted models closely resemble to energetic properties of the achiral triazole derivative where numerous low-energy secondary structures can be found. In contrast, for the (*R,R*)-7 and (*S,S*)-7 enantiomer pair, there are only 3 and 5 low energy conformers, with 6 and 1 (see ESI†), respectively, being the most stable. The (*R,S*)-7 and (*S,R*)-7 have 3 and 7 low energy conformers, respectively, with conformers 2 and 2' as most stable ones (see ESI†). The lower number of stable conformers for the disubstituted models most likely originate from steric interactions.

Overall, all chiral compounds have several low energy conformers, which may result in conformational diversity when homo-oligomers are synthesized from these monomers.

Structure and energetics of heptamers

The eight sets of secondary structures were investigated in order to predict the potentially most preferred structure for each substitution pattern. To allow formation of defined secondary structures, longer peptide sequences, heptamers, were employed. All these sets were investigated in an aqueous environment, as detailed in the Experimental section under



Computational methods. These conformations have intramolecular hydrogen bonds formed between relevant parts of the amide groups. Our nomenclature for the conformations is based on the pseudo-cycles formed by hydrogen bonds, using the number of atoms in the peptide backbone involved in forming the H-bond. Thus, conformers H8, H10, H14, and H16 were investigated for both right (P) and left handed (M) structures.

When using water as an environment, for (R)-6, (R)-5, and (R,R)-7, the most preferred secondary structure is the right-handed H16 conformation, H16_P (Fig. 3 and Table S9[†]) while the second most stable conformation is H14_P. Similarly, due to being conformational enantiomers, for (S)-6, (S)-5, and (S,S)-7, the most preferred structure is also H16, but its left handed form, H16_M. For these compounds, the above H16 conformations have a high relative stability compared to the other investigated secondary structures, as their energy is nearly 9 kcal mol⁻¹ lower than that of the second most stable conformer, H14. Note, that for (R)-5 and (R,R)-7 the H10_P is quite close in relative free energy to H14_P. Thus, for these compounds it is very likely that an oligomer with the above chiral substitution patterns could fold into a helical structure, such as H16, H14 or H10. In general, when investigating handedness there is a strong preference towards either P or M, as for (R)-6, and (S)-6, respectively, where the least stable conformer is H14 for both of them, but with the inverse handedness. For

the (R)-5 and (S)-5 substitution patterns we see a very similar relative energy distribution, and preference of handedness is also pronounced because all conformers belonging to the preferred P, or M subsets have higher or lower relative energies than its inverse conformer, respectively. For (R,R)-7 and (S,S)-7, H16 is again the most stable conformer with H14 being the second most stable one, however, here these conformers have a somewhat higher stability relative to the other conformers investigated (Fig. 3).

In contrast to the above six substitution patterns, from a structural point of view the oligomers built from (S,R)-7 and (R,S)-7 monomers seem to be the most interesting. In the case of these two oligomers, the relative energy of the investigated conformers have values much closer to each other. For (R,S)-7, the most stable conformer is H10_M, however its relative energy is only ~3 kcal mol⁻¹ from the second most stable conformer, H14_M (Fig. 4). Furthermore, the third most stable conformer is H16_P, which has the opposite handedness as the above two conformers, yet its energy is still within 7 kcal mol⁻¹ to that of H10_M. Similarly, in case of (S,R)-7, the same energy distribution can be observed. Based on these results for (S,R)-7, (R,S)-7 systems, there is probably a steric factor introduced which renders various secondary structures to comparable stability. This will probably give rise to a conformational diversity that is comparable to that observed for natural peptidic compounds in living systems. Note, that for homooligomers, the

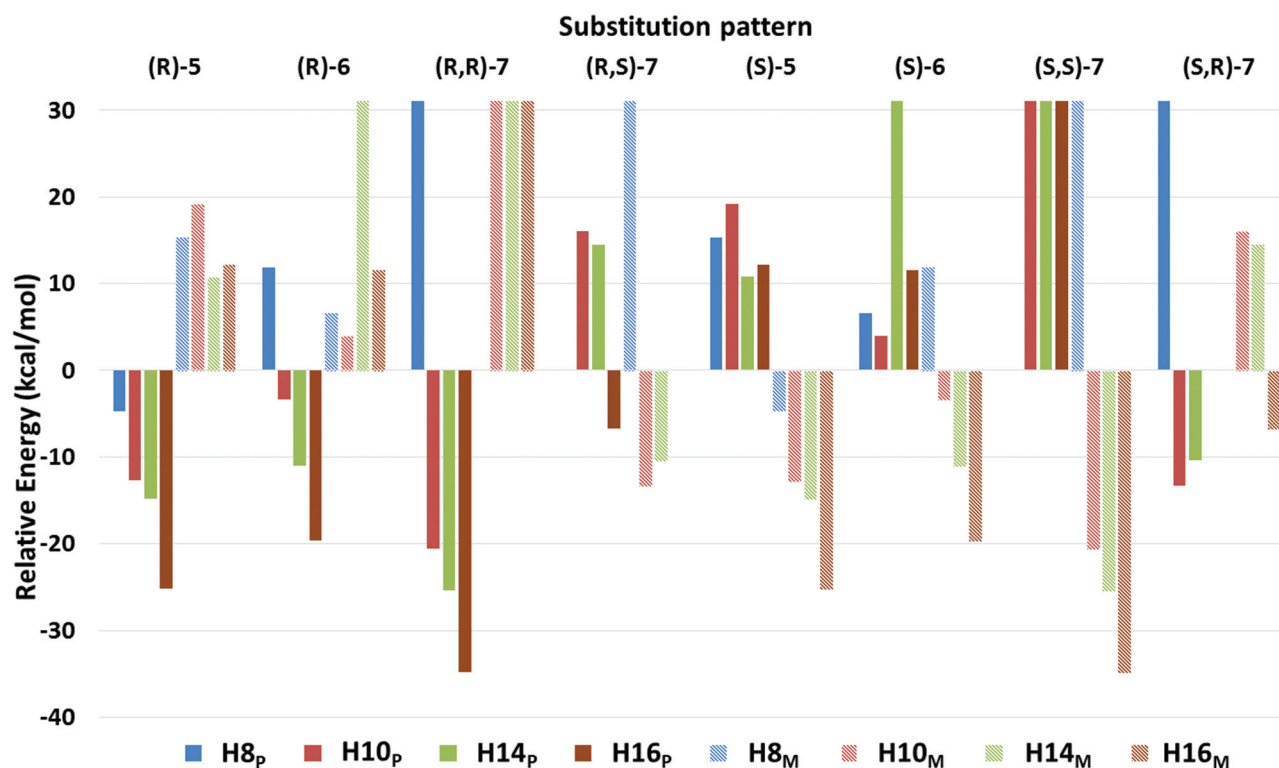


Fig. 3 The relative free energies of the investigated secondary structures for hepta-homooligomers using all 8 possible substitution patterns. Values were obtained at the ω B97X-D/6-31+G(d,p)// ω B97X-D/6-31G(d) level of theory using water as solvent and involving entropy contributions as well as zero point and thermal energy corrections. Relative values were determined for each peptide using the average energy value of all secondary structures as reference point. For more details, see section Computational methods.



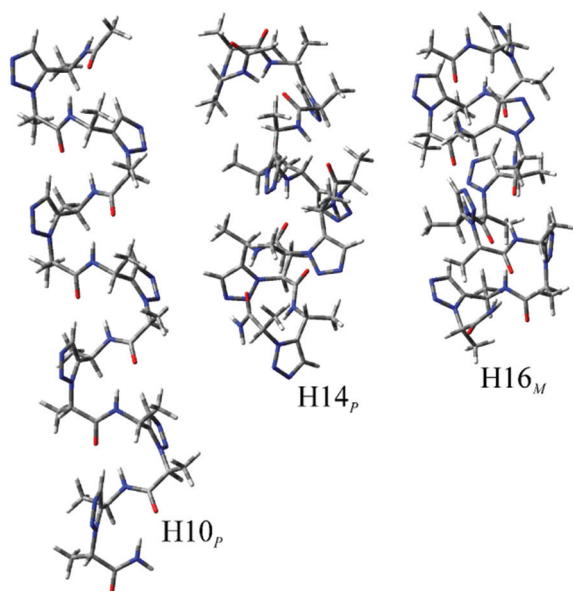


Fig. 4 The three most stable conformation of (S,R) -7 heptahomooligomer. Calculations were performed at the ω B97X-D/6-31+G(d,p)// ω B97X-D/6-31G(d) level of theory.

above results are in contrast to the conformational diversity observed for monomeric compounds. Once the monosubstituted Tzl amino acids are oligomerized, there is a clear preference for one secondary structure among the ones investigated. This observation is also valid for disubstituted compounds with both substitutions having the same chirality, (S,S) -7 and (R,R) -7. In contrast to the above, the mixed substitution patterns, (S,R) -7 and (R,S) -7, may result in conformational diversity for homooligomeric foldamers as several secondary structure elements have close relative energies, irrespective of handedness.

Conclusions

The complete set of possible chiral combinations for 1,5-disubstituted 1,2,3-triazole units have been prepared *via* a ruthenium-catalyzed azide alkyne cycloaddition (RuAAC), in moderate to good yields. These chiral monomeric triazole building blocks are intended for applications in the area of foldamer synthesis. The precursor alkynes and azides used for the RuAAC reaction all originate from the amino acids glycine and alanine. To demonstrate the applicability of the 1,5-triazole monomer building blocks in the synthesis of foldamers, a trimer structure was also prepared. A conformational and structural investigation of these monomers was then carried out to predict the effect of introducing these units into a foldamer. Computational studies on the monomers indicate that several low energy conformers are present for all eight compounds, where several secondary structures may be built with low relative energies for peptidic molecules containing these compounds as single insertions. This is especially indicated

for the monosubstituted compounds, where the number of lower energy conformers exceeds ten. However, when considering homooligomer foldamers built from these compounds, relative energies show a single most stable conformer for 6 out of the 8 substitution patterns. The only two exceptions are (R,S) and (S,R) , where several secondary structures result in relative energies very close to each other, indicating that structural diversity of the backbone, as a key feature for fine-tuning structural properties with various side chain patterns, is present. Overall, we anticipate that the outcome of these studies can be of use in the design and synthesis of new classes of foldamers from non-natural amino acids based on a 1,5-disubstituted 1,2,3-triazole scaffold.

Experimental

General experimental methods

Starting materials were purchased from commercial sources unless otherwise stated. All reactions were performed under an inert atmosphere (argon or nitrogen). Chiral alkynes **1** were initially synthesized from commercial Boc- (R) -alaninol and Boc- (S) -alaninol in one or two steps, but were subsequently purchased from Ark Pharm Inc. (*tert*-butyl (S) -but-3-yn-2-yl-carbamate, 99.5% ee) and J&W PharmLab LLC (*tert*-butyl (R) -but-3-yn-2-yl-carbamate, 98.8% ee). THF was purchased as anhydrous grade. Automated flash chromatography was performed on a Biotage Isolera One. ^1H , ^{13}C and 2D NOESY NMR were acquired on a Varian MR 400 MHz instrument or on a Bruker Avance III 500 MHz spectrometer fitted with a cryoprobe in CDCl_3 or DMSO, using residual solvent peaks for reference. FTIR was performed on a PerkinElmer Frontier FTIR Spectrometer with an ATR device using Spectrum software. The enantiomeric excess was determined by chiral SFC, using a Chiralpak IC column (25% EtOH in CO_2) or a Lux Cellulose-5 column (25% EtOH in CO_2) or a Lux Amylose-1 column (10% EtOH in CO_2) at 120 bar, 40 °C, flow rate 3.5 mL min^{-1} , 220 nm detection (see ESI† for chromatographic data for each compound).

Experimental procedures

Synthesis of chiral propargylic amines

***tert*-Butyl (R) -but-3-yn-2-ylcarbamate ((R)-1).** Alkyne (R)-1 was prepared *via* oxidation of *N*-Boc-D-alaninol ((R)-2-(Boc-amino)-1-propanol) to *N*-Boc-D-alaninal, followed by a Bestmann–Ohira reaction to afford (R)-1. **Oxidation:** To a solution of *N*-Boc-D-alaninol (146 mg, 0.833 mmol) in CH_2Cl_2 (15 mL) was added Dess–Martin periodinane (429 mg, 1.01 mmol) in one portion. The reaction mixture was stirred at ambient temperature for 2.5 h and then extracted with EtOAc (20 mL). After phase separation, the organic phase was washed with 2 M NaHSO_3 (5 mL) and the aqueous phase was extracted with EtOAc (3×10 mL). The combined organic phases were washed with 1 M NaOH (15 mL) and brine (10 mL), then dried



(Na₂SO₄) and concentrated to afford *N*-Boc-D-alaninal (138 mg, 91%) as a white solid of approximately 95% purity. ¹H and ¹³C NMR data were in accordance with published data for this compound.²⁸ This aldehyde was used directly in the next step without further purification. *Bestmann–Ohira reaction*: To a solution of *N*-Boc-D-alaninal (253 mg, 1.46 mmol) in MeOH (25 mL) was added K₂CO₃ (404 mg, 2.92 mmol) in one portion. The reaction was stirred at ambient temperature for 4 h. The solution was concentrated by rotary evaporation and water (25 mL) and EtOAc (25 mL) were added. After phase separation, the aqueous phase was further extracted with EtOAc (3 × 25 mL) and the combined organic phases washed with brine (25 mL), dried (Na₂SO₄) and concentrated. Flash chromatography (silica gel, eluent: CH₂Cl₂) afforded 143 mg of (*R*)-**1** as a white solid. ¹H and ¹³C NMR data were in accordance with published data for this compound.²⁹ However, the ee of (*R*)-**1** was found to be 13.4%. For further studies, compound (*R*)-**1** was instead purchased from J&W PharmLab (ee 98.8%).

tert-Butyl (S)-but-3-yn-2-ylcarbamate ((S)-1). Alkyne (*S*)-**1** was prepared *via* a Bestmann–Ohira reaction of *N*-Boc-D-alaninal using the same procedure as for (*R*)-**1**. ¹H and ¹³C NMR data were in accordance with published data for this compound.³⁰ However, the ee of (*S*)-**1** was found to be 25.8%. For further studies, compound (*S*)-**1** was thus instead purchased from Ark Pharm Inc. (ee 98.5%).

Synthesis of chiral azides

Methyl (R)-2-azidopropanoate ((R)-2). *NOTE*: This is a small-molecule azide and should be handled with care. This reaction should not be scaled up. Do not apply heat to the neat azide product and any reaction with it should be diluted at least 10 times. (*R*)-Methyl 2-aminopropanoate-HCl (855 mg, 6.0 mmol) was suspended in anhydrous acetonitrile (10 mL) and diethylamine (3.0 mL, 28.7 mmol) was added. A suspension of 2-azido-1,3-dimethyl-4,5-dihydro-1*H*-imidazol-3-ium hexafluorophosphate (ADMP) (2.07 g, 7.26 mmol) in anhydrous acetonitrile (10 mL) was added dropwise at 23 °C over 10 min (during the addition, the temperature increased to 32 °C). The reaction mixture was stirred ambient temperature for 1 h, then quenched with saturated NaHCO₃ (15 mL) and stirred for 10 min. CH₂Cl₂ (20 mL) and water (5 mL) was added. After phase separation, the aqueous layer was extracted with CH₂Cl₂ (3 × 5 mL). The combined organic layers were filtered through a phase separator (Telos) and concentrated under reduced pressure. The crude product was purified by gel filtration through silica gel (25 g), eluting with CH₂Cl₂. Concentration of this solution under reduced pressure afforded the product as a colourless oil (351 mg, 45%). ¹H NMR (500 MHz, DMSO): δ 4.33 (q, *J* = 7.1 Hz, 1H), 3.72 (s, 3H), 1.34 (d, *J* = 7.1 Hz, 3H); ¹³C NMR (126 MHz, DMSO) δ 171.20, 56.64, 52.52, 16.48. This compound has been reported earlier,³¹ using a different preparative procedure. Due to the sensitivity of this azide, the ee was not determined at this stage.

Methyl (S)-2-azidopropanoate ((S)-2). Procedure as for (*R*)-**2** (340 mg, 43%). ¹H NMR (500 MHz, DMSO) δ 4.33 (q, *J* = 7.1 Hz, 1H), 3.72 (s, 3H), 1.34 (d, *J* = 7.1 Hz, 3H); ¹³C NMR

(126 MHz, DMSO) δ 171.20, 56.64, 52.52, 16.48. This compound has been reported earlier, using a different preparative procedure.³² Due to the sensitivity of this azide, the ee was not determined at this stage.

General procedure for the RuAAC reaction

RuAAC reactions were performed using microwave heating (*Method A*) or conventional oil bath heating (*Method B*).

Method A: The corresponding alkyne (0.44 mmol) and azide (0.44 mmol) were charged in a Biotage microwave vial with a stirrer bar and dissolved in anhydrous THF (3 mL). Cp*RuCl (COD) (8.4 mg, 0.02 mmol) was added and the vial was sealed, evacuated and refilled with nitrogen three times. The reaction mixture was then heated to 60 °C for 20 min in a microwave reactor or heated in an oil bath at 100 °C for 15 h. The resulting solution was filtered through a syringe filter and concentrated. The crude product was purified by preparative HPLC on a Kromasil C8 column (10 μm 250 × 20 ID mm), using a gradient of 5–45% acetonitrile in a H₂O/acetonitrile/formic acid (95/5/0.2) buffer, over 20 minutes with a flow rate of 19 mL min⁻¹. The compounds were detected by UV at 225 nm. Pure fractions were combined and lyophilized to obtain the product.

Method B, exemplified by the formation of (*S*)-**6**: To a microwave vial equipped with a stirrer bar was added methyl (*S*)-2-azidopropanoate (26 mg, 0.2 mmol) and *N*-Boc-propargylamine (34 mg, 0.2 mmol) along with Cp*RuCl(COD) (3.5 mg, 0.009 mmol) and THF (1 mL). The vessel was sealed in an argon atmosphere and stirred at 100 °C for 15 h or until crude TLC analysis indicated no more starting materials were being consumed (visualized by staining with ninhydrin solution and heating). The crude product was purified by automated flash chromatography on a Biotage Isolera One system, using a gradient of 30–60% EtOAc in petroleum ether. Pure fractions were combined and concentrated by rotary evaporation to afford the product.

Methyl (R)-2-(5-(1-((*tert*-butoxycarbonyl)amino)ethyl)-1*H*-1,2,3-triazol-1-yl)acetate ((R)-5). Prepared from (*R*)-**1** and **4**, affording (*R*)-**5** as a brown solid (method A: 66 mg, 52%, ee 99.1%; method B: 45 mg, 79%; ee 99.1%); [α]_D²⁵ +28.1 (*c* 1.1 in CH₂Cl₂); ν_{max} (neat) 2979, 1753, 1698, 1517, 1453, 1367, 1304, 1243, 1167, 1058, 986, 859, 706; ¹H NMR (400 MHz, CDCl₃): δ 7.53 (s, 1H, triazole-CH), 5.32–5.19 (m, 2H, CH₂), 4.92–4.80 (m, 1H, CHCH₃), 4.88 (br s, 1H, NH), 3.74 (s, 3H, OCH₃), 1.53 (d, *J* = 7.0 Hz, 3H, CHCH₃), 1.37 (s, 9H, C(CH₃)₃); ¹³C NMR (100 MHz, CDCl₃): δ 167.20, 154.91, 140.09, 131.53, 80.33, 52.84, 49.06, 40.07, 28.19, 20.27; HRMS (ESI⁺): found 285.1578 [M + H]⁺; C₁₂H₂₁N₄O₄ requires 285.1563.

Methyl (S)-2-(5-(1-((*tert*-butoxycarbonyl)amino)ethyl)-1*H*-1,2,3-triazol-1-yl)acetate ((S)-5). Prepared from (*S*)-**1** and **4**, affording (*S*)-**5** as a brown solid (method A: 92 mg, 73%, ee 98.4%; method B: 42 mg, 74%, ee 98.4%); [α]_D²⁵ –28.6 (*c* 0.925 in CH₂Cl₂); ν_{max} (neat) 2980, 1752, 1698, 1513, 1453, 1392, 1304, 1239, 1160, 1058, 986, 859, 704; ¹H NMR (400 MHz, CDCl₃): δ 7.58 (s, 1H, triazole-CH), 5.24–5.37 (m, 2H, CH₂), 4.87–4.95 (m, 1H, CHCH₃), 4.76 (br s, 1H, NH), 3.78 (s, 3H, OCH₃), 1.58 (d, *J* = 6.9 Hz, 3H, CHCH₃), 1.41 (s, 9H, C(CH₃)₃);



^{13}C NMR (100 MHz, CDCl_3): δ 167.22, 154.91, 140.08, 131.56, 80.42, 52.87, 49.10, 40.09, 28.22, 20.33; HRMS (ESI+): found 285.1581 $[\text{M} + \text{H}]^+$; $\text{C}_{12}\text{H}_{21}\text{N}_4\text{O}_4$ requires 285.1563.

Methyl (R)-2-(5-(((tert-butoxycarbonyl)amino)methyl)-1H-1,2,3-triazol-1-yl)propanoate ((R)-6). Prepared from (R)-2 and 3, using method A, affording (R)-6 as a pale yellow oil (186 mg; 84%; ee 81.8%); $[\alpha]_{\text{D}} +12.2$ (c 0.975 in CH_2Cl_2); ν_{max} (neat) 2978, 1749, 1749, 1703, 1514, 1449, 1392, 1366, 1247, 1165, 1110, 1079, 1019, 978, 930, 858, 757, 658; ^1H NMR (500 MHz, DMSO-*d*): δ 7.54 (s, 1H, triazole-CH), 7.44 (br s, 1H, triazole-CH), 5.61 (q, $J = 7.2$ Hz, 1H, CHCH₃), 4.18–4.28 (m, 2H, CH₂), 3.66 (s, 3H, OCH₃), 1.76 (d, $J = 7.2$ Hz, 3H, CHCH₃), 1.37 (s, 9H, C(CH₃)₃); ^{13}C NMR (126 MHz, DMSO-*d*): δ 169.45, 155.55, 136.37, 132.39, 78.44, 55.10, 52.77, 32.55, 28.09, 16.90; HRMS (ESI+): found 285.1557 $[\text{M} + \text{H}]^+$; $\text{C}_{12}\text{H}_{21}\text{N}_4\text{O}_4$ requires 285.1563.

Methyl (S)-2-(5-(((tert-butoxycarbonyl)amino)methyl)-1H-1,2,3-triazol-1-yl)propanoate ((S)-6). Prepared from (S)-2 and 3, using method A, affording (S)-6 as a pale yellow oil (173 mg; 87%; ee 79.2%); $[\alpha]_{\text{D}} = -14$ (c 1.1 in CH_2Cl_2); ν_{max} (neat) 2978, 1748, 1698, 1506, 1449, 1367, 1246, 1165, 1079, 1020; ^1H NMR (400 MHz, DMSO-*d*): δ 7.57 (s, 1H, triazole-CH), 5.54–5.35 (m, 1H, CHCH₃), 5.11 (s, 1H, NH), 4.54–4.27 (m, 2H, CH₂), 3.72 (s, 3H, OCH₃), 1.91 (d, $J = 7.3$ Hz, 3H, CHCH₃), 1.42 (s, 9H, C(CH₃)₃); ^{13}C NMR (100 MHz, CDCl_3): δ 169.66, 155.70, 135.28, 133.51, 80.57, 56.31, 53.21, 33.17, 28.36, 28.36, 17.07; HRMS (ESI+): found 285.1556 $[\text{M} + \text{H}]^+$; $\text{C}_{12}\text{H}_{21}\text{N}_4\text{O}_4$ requires 285.1563.

Methyl (R)-2-(5-((R)-1-(((tert-butoxycarbonyl)amino)ethyl)-1H-1,2,3-triazol-1-yl)propanoate ((R,R)-7). Prepared from (R)-1 and (R)-2, using method B, affording (R,R)-7 as a brown solid (50 mg; 84% yield; dr >99/1; ee >99%); $[\alpha]_{\text{D}} +0.59$ (c 1.3 in CH_2Cl_2); ν_{max} (neat) 3206, 2982, 1753, 1698, 1500, 1448, 1436, 1383, 1367, 1344, 1307, 1265, 1249, 1160, 1135, 1117, 1081, 1060, 1021, 976, 856, 834, 732, 703, 663, 636, 556, 513, 471, 418; ^1H NMR (400 MHz, CDCl_3): 7.57 (s, 1H, triazole CH), 5.31 (q, $J = 5.7$ Hz, 1H, COCHCH₃), 5.02–4.90 (m, 1H, NHCH), 4.69 (d, $J = 8.2$ Hz, 1H, NH), 3.72 (s, 3H, OCH₃), 1.95 (d, $J = 7.3$ Hz, 3H) and 1.58 (d, $J = 6.8$ Hz, 3H, COCHCH₃), 1.41 (s, 9H, C(CH₃)₃); ^{13}C NMR (100 MHz, CDCl_3): δ 169.97, 154.89, 138.99, 131.69, 80.54, 56.19, 53.15, 40.20, 28.40, 20.56, 17.60; HRMS (ESI+): found 299.1732 $[\text{M} + \text{H}]^+$; $\text{C}_{13}\text{H}_{23}\text{N}_4\text{O}_4$ requires 299.1719.

Methyl (R)-2-(5-((S)-1-(((tert-butoxycarbonyl)amino)ethyl)-1H-1,2,3-triazol-1-yl)propanoate ((R,S)-7). Prepared from (S)-1 and (R)-2, using method B, affording (R,S)-7 as a pale brown solid (52 mg; 87%; dr >93/7, ee >99%); $[\alpha]_{\text{D}} -0.22$ (c 0.8 in CH_2Cl_2); ν_{max} (neat) 3322, 2979, 1750, 1696, 1514, 1451, 1367, 1305, 1246, 1213, 1162, 1118, 1079, 1057, 1024, 976, 859, 761, 705, 659; ^1H NMR (400 MHz, CDCl_3): 7.57 (s, 1H, triazole-CH), 5.55 (q, $J = 5.3$ Hz, 1H, COCHCH₃), 5.00–4.89 (m, 1H, NHCH), 4.78 (d, $J = 7.8$ Hz, 1H, NH), 3.72 (s, 3H, -OCH₃), 1.91 (d, $J = 7.1$ Hz, 3H) and 1.57 (d, $J = 6.9$ Hz, 3H, COCHCH₃), 1.41 (s, 9H, C(CH₃)₃); ^{13}C NMR (100 MHz, CDCl_3): 169.60, 155.00, 140.22, 131.29, 80.65, 56.04, 53.17, 40.08, 28.39, 20.53, 16.83; HRMS (ESI+): found 299.1735 $[\text{M} + \text{H}]^+$; $\text{C}_{13}\text{H}_{23}\text{N}_4\text{O}_4$ requires 299.1719.

Methyl (S)-2-(5-((R)-1-(((tert-butoxycarbonyl)amino)ethyl)-1H-1,2,3-triazol-1-yl)propanoate ((S,R)-7). Prepared from (R)-1 and

(S)-2, using method B, affording (S,R)-7 as a pale brown solid (56 mg; 95%; dr 96/4, ee >99%); $[\alpha]_{\text{D}} +0.23$ (c 1.05 in CH_2Cl_2); ν_{max} (neat) 3329, 2979, 1750, 1699, 1517, 1452, 1367, 1306, 1247, 1166, 1118, 1079, 1058, 1025, 976, 859, 761, 660; ^1H NMR (400 MHz, CDCl_3): δ 7.55 (s, 1H, triazole-CH), 5.69–5.40 (m, 1H, COCHCH₃), 4.92 (bra app s, 2H, NHCH and NH), 3.70 (s, 3H, OCH₃), 1.89 (d, $J = 7.2$ Hz, 3H) and 1.62–1.50 (m, 3H, COCHCH₃), 1.40 (s, 9H, C(CH₃)₃); ^{13}C NMR (100 MHz, CDCl_3): 169.57, 155.01, 140.21, 131.24, 80.53, 55.98, 53.12, 40.03, 28.33, 20.46, 16.79; HRMS (ESI+): found 299.1728 $[\text{M} + \text{H}]^+$; $\text{C}_{13}\text{H}_{23}\text{N}_4\text{O}_4$ requires 299.1719.

Methyl (S)-2-(5-((S)-1-(((tert-butoxycarbonyl)amino)ethyl)-1H-1,2,3-triazol-1-yl)propanoate ((S,S)-7). Prepared from (S)-1 and (S)-2, using method B, affording (S,S)-7 as a light brown solid (46 mg; 77%; dr >99/1; ee 98.8%); $[\alpha]_{\text{D}} -0.71$ (c 0.925 in CH_2Cl_2); ν_{max} (neat) 3055, 2983, 1751, 1702, 1499, 1451, 1382, 1368, 1306, 1265, 1160, 1118, 1079, 1058, 1021, 975, 896, 858, 835, 731, 702, 555; ^1H NMR (400 MHz, CDCl_3): δ 7.57 (s, 1H, triazole-CH), 5.31 (q, $J = 7.0$ Hz, 1H, COCHCH₃), 4.97 (dt, $J = 13.3, 6.2$ Hz, 1H, NHCH), 4.67 (br s, 1H, NH), 3.72 (s, 3H, OCH₃), 1.95 (d, $J = 7.3$ Hz, 3H) and 1.58 (d, $J = 6.8$ Hz, 3H, COCHCH₃), 1.42 (s, 9H, C(CH₃)₃); ^{13}C NMR (100 MHz, CDCl_3): δ 169.97, 154.87, 138.98, 131.69, 80.54, 56.19, 53.16, 40.22, 28.40, 20.56, 17.60; HRMS (ESI+): found 299.1725 $[\text{M} + \text{H}]^+$; $\text{C}_{13}\text{H}_{23}\text{N}_4\text{O}_4$ requires 299.1719.

General procedure for the CuAAC reaction

To a Biotage microwave vial equipped with a stirrer bar was added the corresponding azide (26 mg, 0.2 mmol) and alkyne (34 mg, 0.2 mmol) along with $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ (1 mg, 0.004 mmol), sodium ascorbate (2.5 mg, 0.012 mmol) and a 1 : 1 mixture of $\text{H}_2\text{O} : t\text{BuOH}$ (1 mL). The vessel was sealed in an air atmosphere and stirred at room temperature for 20 h at which point TLC analysis indicated the reaction was complete. The crude reaction mixture was then diluted in CH_2Cl_2 (10 mL) and water (10 mL) and the aqueous layer was extracted a further two times with CH_2Cl_2 (2 \times 10 mL). The combined organic fractions were then washed with brine (10 mL) and dried using MgSO_4 . The MgSO_4 was then removed from the organic fractions by filtration, concentrated, and purified by flash column chromatography (40–60% EtOAc/hexane) to afford the pure products.

Methyl (R)-2-(4-((R)-1-(((tert-butoxycarbonyl)amino)ethyl)-1H-1,2,3-triazol-1-yl)propanoate ((R,R)-8). Prepared from (R)-1 and (R)-2, affording (R,R)-8 as an oil (55 mg; 92%; dr >99/1; ee 89.8%); $[\alpha]_{\text{D}} +20.2$ (c 1.0 in CH_2Cl_2); ν_{max} (neat) 2978, 2155, 1749, 1699, 1515, 1450, 1366, 1244, 1166, 1095, 1047, 860, 780; ^1H NMR (400 MHz, CDCl_3) δ 7.59 (s, 1H), 5.42 (q, $J = 7.4$ Hz, 1H), 5.13 (s, 1H), 4.93 (dq, $J = 7.5, 6.2$ Hz, 1H), 3.74 (s, 3H), 1.79 (d, $J = 7.3$ Hz, 3H), 1.54 (d, $J = 6.9$ Hz, 3H), 1.41 (s, 9H); ^{13}C NMR (101 MHz, cdCl_3) δ 169.80, 155.20, 150.10, 119.93, 79.58, 58.11, 53.14, 43.01, 28.43, 21.35, 18.25. HRMS (ESI+): found 299.1725 $[\text{M} + \text{H}]^+$; $\text{C}_{13}\text{H}_{23}\text{N}_4\text{O}_4$ requires 299.1719.

Methyl (S)-2-(4-((R)-1-(((tert-butoxycarbonyl)amino)ethyl)-1H-1,2,3-triazol-1-yl)propanoate ((S,R)-8). Prepared from (R)-1 and (S)-2, affording (S,R)-8 as an oil (52 mg; 87%; dr >99/1; ee



99.4%); $[\alpha]_D +50.4$ (c 0.5 in CH_2Cl_2); ν_{max} (neat) 2978, 1749, 1702, 1516, 1454, 1366, 1246, 1169, 1048; ^1H NMR (400 MHz, CDCl_3) δ 7.58 (s, 1H), 5.42 (q, $J = 7.4$ Hz, 1H), 5.13 (br s, 1H), 4.92 (dq, $J = 8.1, 7.6$ Hz, 1H), 3.73 (s, 3H), 1.79 (d, $J = 7.3$ Hz, 3H), 1.54 (d, $J = 6.9$ Hz, 3H), 1.41 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 169.79, 155.22, 150.22, 119.93, 119.90, 79.60, 58.12, 53.14, 43.01, 28.44, 21.32, 18.26. HRMS (ESI+): found 299.1713 $[\text{M} + \text{H}]^+$; $\text{C}_{13}\text{H}_{23}\text{N}_4\text{O}_4$ requires 299.1719.

tert-Butyl (S)-(1-(1-(2-(methylamino)-2-oxoethyl)-1H-1,2,3-triazol-5-yl)ethyl)carbamate ((S)-9). To a microwave vial equipped with a stirrer bar was added (S)-5 (56 mg, 0.2 mmol) and MeOH (2 mL). 1 M LiOH (0.6 mL, 0.6 mmol) was then added dropwise and the solution was stirred at rt for 1.5 h. Water (5 mL) was then added and 1 M HCl (0.6 mL, 0.6 mmol) was added dropwise with stirring. The mixture was transferred to a separating funnel and extracted with CH_2Cl_2 (4×5 mL). The combined organic phases were passed through a TELOS phase separator. The solvent was then removed under a reduced pressure and the resulting carboxylic acid was transferred to a microwave vial equipped with stirrer bar and dissolved in CH_2Cl_2 (2 mL). Methylammonium chloride (54 mg, 0.8 mmol) was then added followed by DIPEA (209 μL , 1.2 mmol). HATU (80 mg, 0.21 mmol) was then added, and the vessel was sealed and allowed to react over night at rt. The reaction mixture was then transferred to a separating funnel along with aq. sat. NH_4Cl (5 mL) and was extracted with EtOAc (4×5 mL). The combined organic phases were dried using MgSO_4 , concentrated, and purified by flash column chromatography eluting with 80–100% EtOAc/hexane to afford (S)-9 as a white solid (37 mg, 65% yield). A second chromatography step was sometimes necessary to remove impurities deriving from the coupling reagent. $[\alpha]_D -2.1$ (c 0.75 in CH_2Cl_2); ν_{max} (neat) 3340, 3318, 2986, 2937, 1676, 1643; ^1H NMR (400 MHz, CDCl_3) δ 7.62 (s, 1H), 6.15 (s, 1H), 5.21 (d, $J = 16.7$ Hz, 1H), 5.03 (d, $J = 16.5$ Hz, 1H), 4.97–4.81 (m, 2H), 2.81 (d, $J = 4.8$ Hz, 3H), 1.55 (d, $J = 6.7$ Hz, 3H), 1.41 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 165.74, 154.95, 140.70, 131.60, 80.65, 51.02, 40.66, 28.24, 26.46, 20.65. HRMS (ESI+): found 284.1729 $[\text{M} + \text{H}]^+$; $\text{C}_{12}\text{H}_{22}\text{N}_5\text{O}_3$ requires 284.1722.

Synthesis of dimer (S,S)-10

Boc-deprotection of (S)-5. To a microwave vial equipped with a stirrer bar was added (S)-5 (56 mg, 0.2 mmol) and CH_2Cl_2 (0.2 mL). Trifluoroacetic acid (TFA) (0.1 mL) was then added and the reaction was stirred at rt for 1.5 h. The reaction mixture was then concentrated under a stream of nitrogen and the salt was used directly for the amide coupling reaction.

Hydrolysis of (S)-5. To a microwave vial equipped with a stirrer bar was added (S)-5 (56 mg, 0.2 mmol) and MeOH (2 mL). 1 M LiOH (0.6 mL, 0.6 mmol) was then added dropwise and the solution was stirred at rt for 1.5 h. Deionised water (5 mL) was then added and 1 M HCl (0.6 mL, 0.6 mmol) was added dropwise with stirring. The mixture was transferred to a separating funnel and extracted with CH_2Cl_2 (4×5 mL). The combined organic phases were passed through a TELOS phase separator. The solvent was then removed under reduced

pressure and the resulting carboxylic acid was used directly in the amide coupling reaction.

Amide coupling reaction. To a microwave vial equipped with a stirrer bar was added the Boc-deprotected (S)-5 and the carboxylic acid formed by hydrolysis of (S)-5, together with CH_2Cl_2 (4 mL). DIPEA (140 μL , 0.8 mmol) was then added followed by HATU (80 mg, 0.21 mmol). The vessel was sealed and allowed to react overnight at rt. The reaction mixture was then transferred to a separating funnel along with aqueous saturated NH_4Cl (5 mL) and extracted with EtOAc (4×5 mL). The combined organic phases were passed through a TELOS phase separator, concentrated, and purified by flash column chromatography eluting with 80–100% EtOAc/hexane. The crude white solid was then further purified by trituration using hexane and dried to yield dimer (S,S)-10 as a white solid (68 mg, 78% yield). $[\alpha]_D -33.0$ (c 0.1 in CH_2Cl_2); ν_{max} (neat) 3311 (br), 2986, 1749, 1672, 1535, 1523; ^1H NMR (400 MHz, CDCl_3) δ 7.60 (s, 1H), 7.56 (s, 1H), 7.41 (d, $J = 8.2$ Hz, 1H), 5.43 (d, $J = 17.7$ Hz, 1H), 5.33 (d, $J = 17.6$ Hz, 1H), 5.20–5.07 (m, 2H), 5.07 (d, $J = 7.2$ Hz, 1H), 4.90 (d, $J = 16.6$ Hz, 1H), 4.73 (p, $J = 7.0$ Hz, 1H), 3.77 (s, 3H), 1.57 (d, $J = 7.0$ Hz, 3H), 1.47 (d, $J = 6.9$ Hz, 3H), 1.41 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 167.66, 165.27, 155.49, 141.10, 140.05, 132.04, 131.47, 81.02, 53.13, 50.79, 49.41, 41.13, 39.20, 28.43, 20.76, 19.78. HRMS (ESI+): found 437.2248 $[\text{M} + \text{H}]^+$; $\text{C}_{18}\text{H}_{29}\text{N}_8\text{O}_5$ requires 437.2261.

Synthesis of trimer (S,S,S)-11

Boc-deprotection of (S)-9. To a microwave vial equipped with a stirrer bar was added (S)-9 (28 mg, 0.1 mmol) and CH_2Cl_2 (2 mL). TFA (1 mL) was then added and the reaction mixture was stirred at rt for 1.5 h. The reaction mixture was then condensed under a stream of nitrogen and the salt was used directly in the amide coupling reaction.

Hydrolysis of (S,S)-10. To a microwave vial equipped with stirrer bar was added (S,S)-10 (44 mg, 0.1 mmol) and MeOH (1 mL). 1 M LiOH (0.3 mL, 0.3 mmol) was then added dropwise and the solution was stirred at rt for 1.5 h. Deionised water (5 mL) was then added and 1 M HCl (0.3 mL, 0.3 mmol) was added dropwise with stirring. The mixture was transferred to a separating funnel and extracted with EtOAc (4×10 mL). The combined organic layers were passed through a TELOS phase separator. The solvent was then removed under a reduced pressure and the resulting carboxylic acid was used directly.

Amide coupling reaction. To a microwave vial equipped with stirrer bar was added the Boc-deprotected (S)-9 and the carboxylic acid formed by hydrolysis of (S,S)-10, together with DMF (1 mL). DIPEA (70 μL , 0.4 mmol) was then added, followed by HATU (39 mg, 0.105 mmol). The vessel was sealed and the reaction was allowed to react overnight at rt. The DMF was removed and the crude reaction mixture was purified directly by flash column chromatography eluting with a gradient of 5–15% MeOH/ CH_2Cl_2 . The crude white solid was then further purified by trituration using CHCl_3 , followed by 1 M NaOH and H_2O , before being dried, affording trimer (S,S,S)-11 as a white solid (27 mg, 46% yield). $[\alpha]_D -9.8$ (c 0.22 in CH_2Cl_2); ν_{max} (neat) 3303 (br), 2983, 1749, 1669 (br), 1537;



^1H NMR (400 MHz, DMSO- d_6) δ 8.93 (d, J = 7.6 Hz, 2H), 8.20 (q, J = 4.6 Hz, 1H), 7.73 (s, 1H), 7.72 (s, 1H), 7.57 (s, 1H), 7.45 (d, J = 8.0 Hz, 1H), 5.20–4.96 (m, 7H), 4.92 (p, J = 7.0 Hz, 1H), 4.62 (dt, J = 15.3, 6.6 Hz, 1H), 2.60 (d, J = 4.6 Hz, 3H), 1.45 (d, J = 6.9 Hz, 3H), 1.41 (d, J = 7.0 Hz, 3H), 1.32 (s, 9H), 1.27 (d, J = 7.0 Hz, 3H); ^{13}C NMR (101 MHz, DMSO- d_6) δ 165.71, 165.13, 165.05, 154.98, 141.60, 140.52, 140.29, 131.8, 131.05, 130.70, 78.36, 49.83, 49.65, 49.53, 40.30 (overlaps with DMSO), 38.82, 38.75, 28.10, 25.59, 20.17, 19.77, 19.70; HRMS (ESI+): found 588.3096 $[\text{M} + \text{H}]^+$; $\text{C}_{24}\text{H}_{38}\text{N}_{13}\text{O}_5$ requires 588.3119.

Computational methods

All computations were carried out using the Gaussian 09 software package.³³ In contrast to experiments, for the calculations the monomer amino acids had *N*-methyl and acetamide protecting groups at their C- and N-terminals, respectively, abbreviated as Ac-5Tzl-NHMe. In case of the longer models the C-terminal protecting group was NH_2 , resulting the general Ac-(5Tzl)₇- NH_2 form. Nevertheless, the same nomenclature was applied for the substitution patterns as for the experiments (Fig. 2). For the monomers, the 27 minima obtained previously in an exhaustive conformational search for the achiral models at the RHF/3-21G level of theory,²² were employed in the current study as initial conformations. Out of these 7 starting conformers were completely planar, with relative energies of their achiral version higher than 20 kcal mol⁻¹. Consequently, these were omitted for the current chiral models, leaving 20 starting test conformers for all eight models. The total of 160 optimisations were primarily performed at the RHF/3-21G level of theory. Further on, the optimised structures were submitted to calculations using Becke's three parameter functional with the Lee–Yang–Parr exchange functional (B3LYP),³⁴ the ω B97X-D functional by Chai and Head-Gordon,³⁵ which included dispersion correction and long-range electron correlation corrections, as well as M06-2X functional developed by Zhao and Truhlar.³⁶ For all functionals, the 6-311++G(2d,2p) basis set was employed. The obtained results for all the conformers are found in the ESI.†

To estimate secondary structure preference of longer oligomers composed of the investigated 8 monomers, previously defined secondary structures were employed.²¹ Initially, five secondary structures, four helical, and one extended, were modified with the 8 chiral substitution patterns and optimised at the ω B97x-D/6-31G(d) level of theory. To predict preference of helicity handedness, both P and M helices were optimised resulting a total of 9 secondary structures. Considering the large size and the unfavorable nature of the extended conformation, for many substitution patterns this conformer experienced convergence problems during the calculations, therefore this conformer was excluded from further investigation. To validate the critical points, *i.e.* to confirm that they are local minima, as well as to obtain zero point and thermal corrections to the energy, and entropy contributions to provide Gibbs free energy values, normal mode analysis was performed on the obtained conformers at the ω B97x-D/6-31G(d) level of theory. These optimised conformers were also submitted to point energy calculations employing the same functional and

the 6-31+G(d,p) basis set. Further on, environmental effects of water were considered also by single point energy calculations at the same level of theory, using the implicit solvent model integral equation formalism for polarizable continuum model (IEFPCM) (Tables S1–S9†).

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

The following sources of funding are gratefully acknowledged: the Swedish Research Council (N. K., 2015-05360), the Swedish Research Council Formas (N. K., 2015-1106), the Carl Trygger Foundation (A. P., CTS 16:235), the Hungarian Scientific Research Fund (T. B.-S., OTKA-109588), the Momentum program of the Hungarian Academy of Sciences (T. B.-S., LP2016-2), the National Competitiveness and Excellence Program (NVKP_16-1-2016-0007), the Excellence program BIONANO_GINOP-2.3.2-15-2016-00017, the Nemzeti Versenyképességi és Kiválósági Program, Hungary (NVKP_16-1-2016-0007), the Gazdaságfejlesztési és Innovációs Operatív Program, Hungary (GINOP-2.3.2-15-2016-00017), and a Marie Curie fellowship (T. B.-S., MSCA-IF BARREL 660030).

References

- (a) J. V. Schreiber, J. Frackenhohl, F. Moser, T. Fleischmann, H. P. E. Kohler and D. Seebach, *ChemBioChem*, 2002, **3**, 424–432; (b) D. F. Hook, F. Gessier, C. Noti, P. Kast and D. Seebach, *ChemBioChem*, 2004, **5**, 691–706.
- D. Seebach, A. K. Beck and D. J. Bierbaum, *Chem. Biodivers.*, 2004, **1**, 1111–1239.
- (a) T. Kimmerlin, D. Seebach and D. Hilvert, *Helv. Chim. Acta*, 2002, **85**, 1812–1826; (b) G. V. M. Sharma, B. S. Babu, K. V. S. Ramakrishna, P. Nagendar, A. C. Kunwar, P. Schramm, C. Baldauf and H. J. Hofmann, *Chem. – Eur. J.*, 2009, **15**, 5552–5566; (c) T. A. Martinek and F. Fulop, *Chem. Soc. Rev.*, 2012, **41**, 687–702.
- K. Gademann, T. Hintermann and J. V. Schreiber, *Curr. Med. Chem.*, 1999, **6**, 905–925.
- W. S. Horne and S. H. Gellman, *Acc. Chem. Res.*, 2008, **41**, 1399–1408.
- G. N. Tew, R. W. Scott, M. L. Klein and W. F. Degrado, *Acc. Chem. Res.*, 2010, **43**, 30–39.
- W. S. Horne, L. M. Johnson, T. J. Ketas, P. J. Klasse, M. Lu, J. P. Moore and S. H. Gellman, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 14751–14756.
- N. Umezawa, M. A. Gelman, M. C. Haigis, R. T. Raines and S. H. Gellman, *J. Am. Chem. Soc.*, 2002, **124**, 368–369.
- J. Kim, S. Kwon, S. H. Kim, C. K. Lee, J. H. Lee, S. J. Cho, H. S. Lee and H. Ihee, *J. Am. Chem. Soc.*, 2012, **134**, 20573–20576.



- 10 (a) R. V. Nair, K. N. Vijayadas, A. Roy and G. J. Sanjayan, *Eur. J. Org. Chem.*, 2014, 7763–7780; (b) J. S. Laursen, J. Engel-Andreasen and C. A. Olsen, *Acc. Chem. Res.*, 2015, **48**, 2696–2704.
- 11 R. Gopalakrishnan, A. I. Frolov, L. Knerr, W. J. Drury and E. Valeur, *J. Med. Chem.*, 2016, **59**, 9599–9621.
- 12 (a) C. W. Tornoe, C. Christensen and M. Meldal, *J. Org. Chem.*, 2002, **67**, 3057–3064; (b) V. V. Rostovtsev, L. G. Green, V. V. Fokin and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2002, **41**, 2596–2599; (c) E. Haldon, M. C. Nicasio and P. J. Perez, *Org. Biomol. Chem.*, 2015, **13**, 9528–9550; (d) C. L. Wang, D. Ikhlef, S. Kahlal, J. Y. Saillard and D. Astruc, *Coord. Chem. Rev.*, 2016, **316**, 1–20.
- 13 Y. Wang, J. F. Xiang and H. Jiang, *Chem. – Eur. J.*, 2011, **17**, 613–619.
- 14 N. Castellucci and C. Tomasini, *Eur. J. Org. Chem.*, 2013, 3567–3573.
- 15 (a) W. S. Horne, M. K. Yadav, C. D. Stout and M. R. Ghadiri, *J. Am. Chem. Soc.*, 2004, **126**, 15366–15367; (b) T. Boddaert, J. Sola, M. Helliwell and J. Clayden, *Chem. Commun.*, 2012, **48**, 3397–3399; (c) K. B. Salah, S. Das, N. Ruiz, V. Andreu, J. Martinez, E. Wenger, M. Amblard, C. Didierjean, B. Legrand and N. Inguibert, *Org. Biomol. Chem.*, 2018, **16**, 3576–3583.
- 16 (a) L. Zhang, X. G. Chen, P. Xue, H. H. Y. Sun, I. D. Williams, K. B. Sharpless, V. V. Fokin and G. C. Jia, *J. Am. Chem. Soc.*, 2005, **127**, 15998–15999; (b) B. C. Boren, S. Narayan, L. K. Rasmussen, L. Zhang, H. T. Zhao, Z. Y. Lin, G. C. Jia and V. V. Fokin, *J. Am. Chem. Soc.*, 2008, **130**, 8923–8930.
- 17 O. Kracker, J. Gora, J. Krzciuk-Gula, A. Marion, B. Neumann, H. G. Stammler, A. Niess, I. Antes, R. Latajka and N. Sewald, *Chem. – Eur. J.*, 2018, **24**, 953–961.
- 18 M. Wunsch, D. Schroder, T. Frohr, L. Teichmann, S. Hedwig, N. Janson, C. Belu, J. Simon, S. Heidemeyer, P. Holtkamp, J. Rudlof, L. Klemme, A. Hinzmann, B. Neumann, H. G. Stammler and N. Sewald, *Beilstein J. Org. Chem.*, 2017, **13**, 2428–2441.
- 19 J. R. Johansson, T. Beke-Somfai, A. S. Stålsmeden and N. Kann, *Chem. Rev.*, 2016, **116**, 14726–14768.
- 20 (a) D. S. Pedersen and A. Abell, *Eur. J. Org. Chem.*, 2011, 2399–2411; (b) A. Tam, U. Arnold, M. B. Soellner and R. T. Raines, *J. Am. Chem. Soc.*, 2007, **129**, 12670–12671; (c) J. K. Pokorski, L. M. M. Jenkins, H. Q. Feng, S. R. Durell, Y. W. Bai and D. H. Appella, *Org. Lett.*, 2007, **9**, 2381–2383; (d) D. Tietze, M. Tischler, S. Voigt, D. Imhof, O. Ohlenschlager, M. Gorchach and G. Buntkowsky, *Chem. – Eur. J.*, 2010, **16**, 7572–7578; (e) M. Empting, O. Avrutina, R. Meusinger, S. Fabritz, M. Reinwarth, M. Biesalski, S. Voigt, G. Buntkowsky and H. Kolmar, *Angew. Chem., Int. Ed.*, 2011, **50**, 5207–5211; (f) J. Q. Zhang, J. Kemmink, D. T. S. Rijkers and R. M. J. Liskamp, *Chem. Commun.*, 2013, **49**, 4498–4500; (g) S. Ferrini, J. Z. Chandanshive, S. Lena, M. C. Franchini, G. Giannini, A. Tafi and M. Taddei, *J. Org. Chem.*, 2015, **80**, 2562–2572; (h) X. Yang, L. P. Beroske, J. Kemmink, D. T. S. Rijkers and R. M. J. Liskamp, *Tetrahedron Lett.*, 2017, **58**, 4542–4546; (i) S. Pacifico, A. Kerckhoffs, A. J. Fallow, R. E. Foreman, R. Guerrini, J. McDonald, D. G. Lambert and A. G. Jamieson, *Org. Biomol. Chem.*, 2017, **15**, 4704–4710; (j) E. Engholm, N. Stühr-Hansen and O. Blixt, *Tetrahedron Lett.*, 2017, **58**, 2272–2275.
- 21 J. R. Johansson, E. Hermansson, B. Norden, N. Kann and T. Beke-Somfai, *Eur. J. Org. Chem.*, 2014, 2703–2713.
- 22 N. Kann, J. R. Johansson and T. Beke-Somfai, *Org. Biomol. Chem.*, 2015, **13**, 2776–2785.
- 23 D. Habrant, V. Rauhala and A. M. P. Koskinen, *Chem. Soc. Rev.*, 2010, **39**, 2007–2017.
- 24 (a) D. B. Dess and J. C. Martin, *J. Org. Chem.*, 1983, **48**, 4155–4156; (b) A. Yoshimura and V. V. Zhdankin, *Chem. Rev.*, 2016, **116**, 3328–3435.
- 25 H. Tohma and Y. Kita, *Adv. Synth. Catal.*, 2004, **346**, 111–124.
- 26 M. Kitamura, S. Kato, M. Yano, N. Tashiro, Y. Shiratake, M. Sando and T. Okauchi, *Org. Biomol. Chem.*, 2014, **12**, 4397–4406.
- 27 C. Baldauf and H. J. Hofmann, *Helv. Chim. Acta*, 2012, **95**, 2348–2383.
- 28 J. Ryan, M. Siauciulis, A. Gomm, B. Macia, E. O'Reilly and V. Caprio, *J. Am. Chem. Soc.*, 2016, **138**, 15798–15800.
- 29 B. Rajagopal, Y. Y. Chen, C. C. Chen, X. Y. Liu, H. R. Wang and P. C. Lin, *J. Org. Chem.*, 2014, **79**, 1254–1264.
- 30 J. A. Fehrentz and B. Castro, *Synthesis*, 1983, 676–678.
- 31 R. V. Hoffman and H. O. Kim, *Tetrahedron*, 1992, **48**, 3007–3020.
- 32 A. Paul, H. Bittermann and P. Gmeiner, *Tetrahedron*, 2006, **62**, 8919–8927.
- 33 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, Ö. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski, and D. J. Fox, *Gaussian 09, Revision A.1*, Gaussian, Inc., Wallingford CT, 2009.
- 34 (a) C. T. Lee, W. T. Yang and R. G. Parr, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 1988, **37**, 785–789; (b) A. D. Becke, *J. Chem. Phys.*, 1993, **98**, 1372–1377.
- 35 J. D. Chai and M. Head-Gordon, *Phys. Chem. Chem. Phys.*, 2008, **10**, 6615–6620.
- 36 Y. Zhao and D. G. Truhlar, *Theor. Chem. Acc.*, 2006, **120**, 215–241.

