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## Fabrication of bismuth oxide-modified pencil graphite sensors for monitoring the hazardous herbicide diuron†

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In this work, a bismuth oxide/pencil graphite electrode ( $\text{Bi}_2\text{O}_3/\text{PGE}$ )-based ultrasensitive voltammetric sensor is reported for the quantification of the herbicide diuron (DU) in vegetable samples. A HB pencil lead of diameter 2.0 mm was applied to prepare the electrode. Bismuth oxide nanopowder of the size range 1–10  $\mu\text{m}$  was used. The prepared  $\text{Bi}_2\text{O}_3/\text{PGE}$  was characterized by different techniques such as energy-dispersive X-ray (EDX) spectroscopy, scanning electron microscopy (SEM), electrochemical impedance spectroscopy (EIS) and cyclic voltammetry (CV). A linear calibration curve was plotted in the range of 5.0–160.0  $\text{ng mL}^{-1}$  in a BR buffer (pH 2.5) solution. The sensitivity of the fabricated sensor was calculated to be 2.80  $\text{ng mL}^{-1}$ . The suggested sensor indicated higher sensitivity towards the detection of DU in a small-sized whole-tomato sample with 99.9% recovery.

## 1. Introduction

Diuron (3-(3,4-dichlorophenyl)-1,1-dimethylurea) (DU) is a substituted phenyl urea herbicide applied for the pre-emergence control of a wide range of wild plants in the cultivation of a vast variety of crops such as citrus fruits, rice, cotton, soybeans, sugarcane, potato, wheat, tea and coffee as well as along airport runways, railways and pipelines.<sup>1–3</sup> Undegraded deposits of the herbicide stay in surface and ground water beyond their permissible limit. In humans, methemoglobin in the blood, liver dysfunction, spleen deformity and hormonal imbalance are the most common health diseases that were identified to be caused by the presence of diuron in food and water. Due to its harmful effects on human health and ecology, it is a key concern to develop an easy, selective, rapid and reliable method for the quantification of DU.<sup>4–7</sup> A variety of electrochemical techniques have been applied for the quantification of DU, but due to passivation of the electrode, surface deposition of polymeric products has been detected.<sup>8–13</sup>

Glassy carbon, carbon paste, carbon nanofibers, carbon strips, etc. are the generally used carbon-based electrodes for the voltammetric determination of pesticides and other environmental pollutants.<sup>14–17</sup> Other than carbon electrodes, a number of metal electrodes such as platinum and gold have also been fabricated with a variety of polymer nanocomposites for the electrochemical sensing of environmental pollutants,

but the fouling of their surface due to adherence of intermediate products during electrochemical measurement is the major drawback as it requires cleaning of the electrode surface prior to each measurement.<sup>18–24</sup> To overcome the above-mentioned major limitation, a novel disposable pencil graphite electrode was prepared indigenously and applied for the quantification of the herbicide DU. Nowadays, for on-site measurement, demands for cost-effective, field functional voltammetric sensors and easy-to-fabricate-type sensors have been ever increasing. A wide range of nanomaterials with unique chemical, physical and electronic properties with the significance of the small size are gaining significant interest as electrochemical sensors.<sup>25–32</sup> Along with these electrochemical sensors, metal oxide-based sensors show superior electrocatalytic activity, stability and reproducibility for the determination of various pesticides.<sup>33–37</sup> The advantages of  $\text{Bi}_2\text{O}_3$  nanoparticles over other metals are revealed in their significant performances such as adequate surface area, electrochemical stability, efficient catalytic activity, ease of functionalization, cost-effectiveness, chemical inertness, and less-toxicity. Due to less toxicity and non-reactive heavy metals, this is appropriate for *in vivo* applications compared to other metals. The simplicity in controlling their particle size and shape throughout the preparation is an additional benefit of bismuth oxide nanoparticles.<sup>38,39</sup> On surveying the literature, we found that few polymer-based sensors such as reduced grapheme oxide, MWCNT, and gold nanoparticles have been developed for detecting diuron, but all previously reported sensors were based on the commercial expensive glassy carbon electrode and the modifier applied was of high cost and more toxicity than bismuth oxide. In the current work, a  $\text{Bi}_2\text{O}_3/\text{PGE}$  was applied to monitor the concentration of DU in a variety of real samples.

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The developed sensor has the advantage of lower cost, ease of availability and disposability.

## 2. Experimental

### 2.1. Instruments and chemicals

All voltammetric measurements were carried out using an AUTOLAB 112 Potentiostat galvanostat 302 N (the Netherlands) with software NOVA 1.1. A three-electrode system was applied for all measurements, and in this work, a PGE was used as the working electrode, a calomel electrode as the reference electrode and a platinum wire as the counter electrode. All pH measurements were recorded using a decible DB-1011 digital pH meter. Diuron (DU) ( $\geq 98\%$ ) standard was acquired from TCI Chemicals. Bismuth oxide along with other used chemicals was bought from Sigma Aldrich, of AR grade of  $1\text{ }\mu\text{m}$  with 99.9% purity and employed without further purification. An appropriate amount of methanol was taken as the solvent to prepare the stock solution of  $0.004\text{ mM}$  DU.

### 2.2. Development of $\text{Bi}_2\text{O}_3$ /PGE working sensors

The method of construction of a pencil graphite electrode is the same as reported briefly in earlier papers. A HB pencil lead of diameter  $2.0\text{ mm}$  was hooked in an insulin syringe used as the holder and fixed with bontite used as the fixing material (Fig. 1). A working electrode was prepared by the drop-casting method *via* applying a uniform thin coating of a  $\text{Bi}_2\text{O}_3$  slurry on the PGE followed by drying at room temperature. The concentration of  $\text{Bi}_2\text{O}_3$  was optimized; for this purpose, various concentrations of  $\text{Bi}_2\text{O}_3$ , namely,  $0.500\text{ mg mL}^{-1}$ ,  $1.0\text{ mg mL}^{-1}$ ,  $2.0\text{ mg mL}^{-1}$ ,  $3.0\text{ mg mL}^{-1}$ ,  $4.0\text{ mg mL}^{-1}$ , and  $5.0\text{ mg mL}^{-1}$  were dissolved in DMF and the peak intensity was evaluated. The result indicated that the peak intensity (during oxidation of DU) increased up to  $3.0\text{ mg mL}^{-1}$  and decreased thereafter. The maximum peak current was observed at  $3.0\text{ mg mL}^{-1}$ , and hence, it was selected for further experiments. The amount of the sensing material  $\text{Bi}_2\text{O}_3$  was also optimized (Fig. 1, ESI<sup>†</sup>). By varying the casting volume from  $1.0$  to  $5.0\text{ }\mu\text{L}$ , the peak intensity amplifies up to  $4.0\text{ }\mu\text{L}$  and diminishes thereafter.

### 2.3. Electrochemical measurements

All electrochemical measurements were recorded in  $16.0\text{ mL}$  of buffer solution (BR, pH 2.5), with  $4.0\text{ mL}$  of aqueous KCl solution as the supporting electrolyte poured into the electrochemical cell. Then, the DU solution of different concentrations



Fig. 1 Image of a pencil graphite electrode and its surface prepared from insulin syringe.

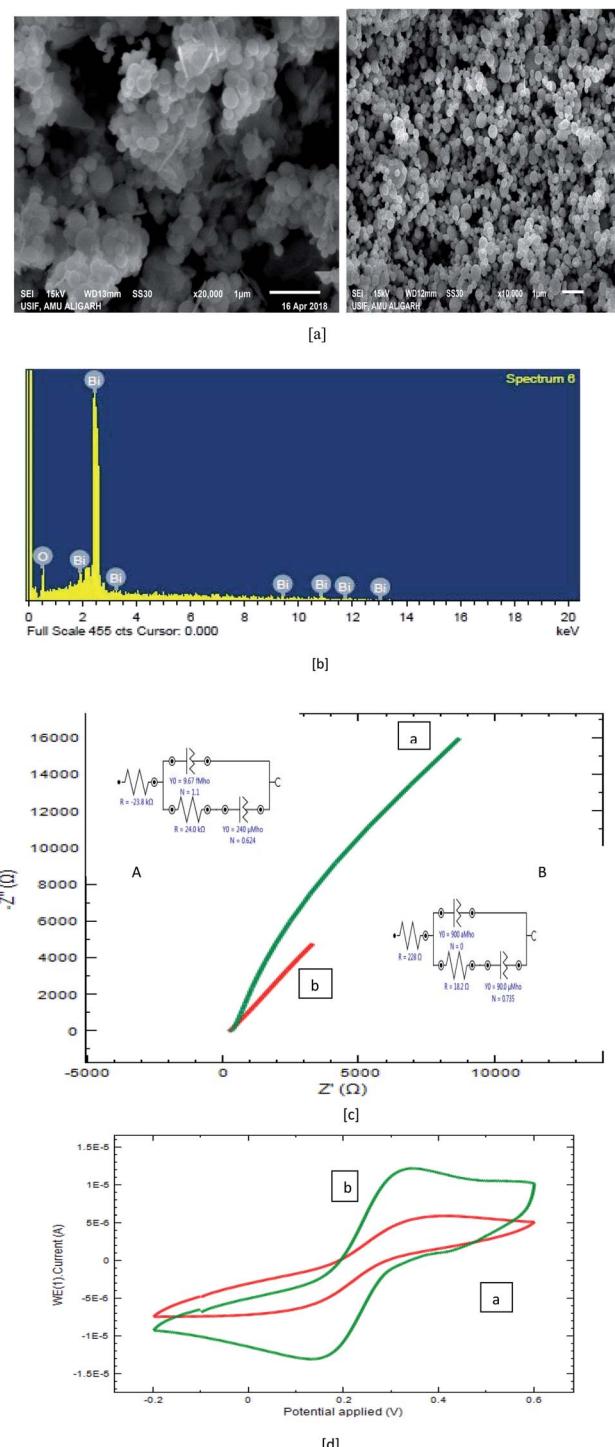


Fig. 2 (a) SEM image of  $\text{Bi}_2\text{O}_3$ , (b) EDX image of  $\text{Bi}_2\text{O}_3$ , (c) Nyquist plot for electrodes in  $5.0\text{ mM K}_3\text{[Fe(CN)}_6\text{]}$  (red: bare PGE; green:  $\text{Bi}_2\text{O}_3$ /PGE). (d) Surface area studies of the bare PGE (red) and  $\text{Bi}_2\text{O}_3$ /PGE (green) in  $1.0\text{ mM K}_3\text{[Fe(CN)}_6\text{]}$  in  $0.1\text{ M KCl}$  by cyclic voltammetry.

diluted from  $0.004\text{ mM}$  stock solution was added subsequently. Voltammograms were recorded by cyclic and differential pulse voltammetry *via* sweeping likely towards the positive direction in the range of  $0.2\text{ V}$  to  $1.2\text{ V}$ . Electrochemical impedance spectroscopy was used for the electrochemical



characterization PGE/Bi<sub>2</sub>O<sub>3</sub> sensors in 1.0 mM K<sub>3</sub>[Fe(CN)<sub>6</sub>] solution (phosphate buffer of pH 7.0).

#### 2.4. Real sample preparation

For the quantification of DU, a small-sized whole tomato was purchased from a local vegetable market and completely ground in a mixer grinder followed by filtration using a filter paper. An appropriate amount of the filtrate was spiked with different concentrations of DU and quantified by cyclic voltammetry.

### 3. Results and discussion

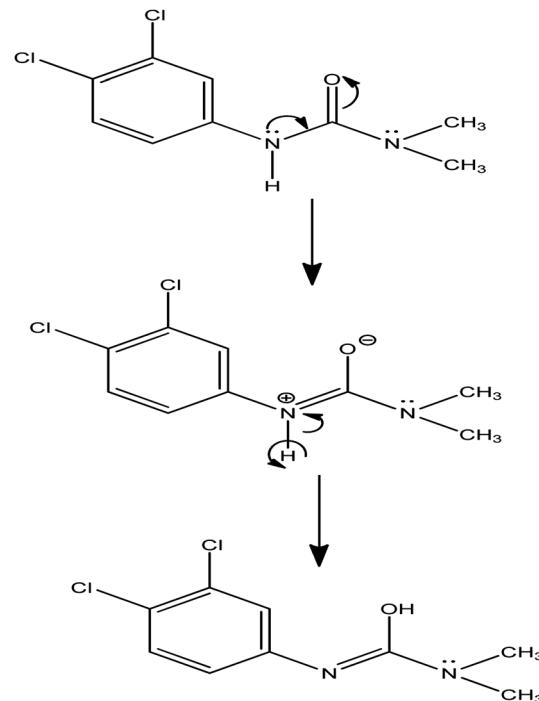
#### 3.1. Bi<sub>2</sub>O<sub>3</sub> sensor characterization

The morphology of the prepared composites was studied by scanning electron microscopy (SEM). The SEM image of Bi<sub>2</sub>O<sub>3</sub> is demonstrated in Fig. 2a, and a pea-like structure of size 1.0  $\mu$ m can be observed in the image at two different magnifications. The elemental composition of Bi<sub>2</sub>O<sub>3</sub> was verified by EDX, and the spectrum is shown in Fig. 2b. The peaks observed for Bi and O confirmed the presence of all elements in the composite.

The surface area of the developed sensor was estimated by cyclic voltammetry *via* a reversible redox reaction of a potassium ferricyanide probe, (K<sub>3</sub>[Fe(CN)<sub>6</sub>]) (Fig. 2c) (prepared in 1 M KCl) and computed using Randles–Ševčík equations (under non-standard conditions), as given in eqn (i)<sup>28–31,41,42</sup>

$$I_{\text{revp,f}} = +0.446nFA_{\text{real}}C\sqrt{nFD\nu/RT} \quad (\text{i})$$

here,  $I$  is the peak current,  $n$  the number of electrons transferred,  $A$  the electroactive area ( $\text{cm}^{-2}$ ),  $D$  the diffusion



Scheme 1 Reaction mechanism.

coefficient of the electroactive species ( $\text{cm}^2 \text{ s}^{-1}$ ),  $C$  the bulk concentration of the same species ( $\text{mol cm}^{-3}$ ),  $\nu$  the scan rate ( $\text{V s}^{-1}$ ), and  $D$  the diffusion coefficient ( $7.6 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$  for ferricyanide). The calculated electroactive surface was  $0.024 \text{ cm}^2$  for bare PGE and  $0.034 \text{ cm}^2$  for Bi<sub>2</sub>O<sub>3</sub>/PGE. The calculated  $A_{\text{real}}$  value and its percentage were also compared to the  $A_{\text{geo}}$  value using the following formula: %Real =  $(A_{\text{real}}/A_{\text{geo}}/100)$ , and % Real was found to be 77.7% for PGE and 109.6% for Bi<sub>2</sub>O<sub>3</sub>/PGE.

The potential of electron transfer at various electrodes was further examined by electrochemical impedance spectroscopy (EIS), and the obtained results are presented in Fig. 2c.

The thickness of the uniform layer was also calculated using the formula:  $d = \epsilon_0 \epsilon_r A/C$ , where  $\epsilon_0$  is the electrical permittivity of vacuum ( $\epsilon_0 = 8.85 \times 10^{-14} \text{ F cm}^{-1}$ ),  $\epsilon_r$  the relative permittivity of the bismuth oxide film and  $A$  its geometric surface area, and  $C$  the capacitance that was found to be 22.0 nm.

In general, the enhancement in the diameter of the semi-circle reveals the increase in the interfacial charge-transfer resistance ( $R_{\text{ct}}$ ). The charge-transfer resistance ( $R_{\text{ct}}$ ) for the two fabricated sensors were estimated to be  $24.8 \text{ k}\Omega$  for the bare PGE and  $18.3 \Omega$  for the Bi<sub>2</sub>O<sub>3</sub>/PGE. The equivalent circuit to EIS for the bare PGE is given in the inset of Fig. 2c. The rule obtained for the bare component was  $[R(Q[RQ])]$  for the bare PGE and  $[R(Q[RW])]$  for the Bi<sub>2</sub>O<sub>3</sub>/PGE. The obtained data were in conformity with the good conductivity and electrochemical properties of the Bi<sub>2</sub>O<sub>3</sub>/PGE.

#### 3.2. Electro-oxidation of DU at a developed sensor

The oxidation of DU at the fabricated sensor was studied by cyclic voltammetry. The study was conducted in a BR buffer (pH 2.5) at

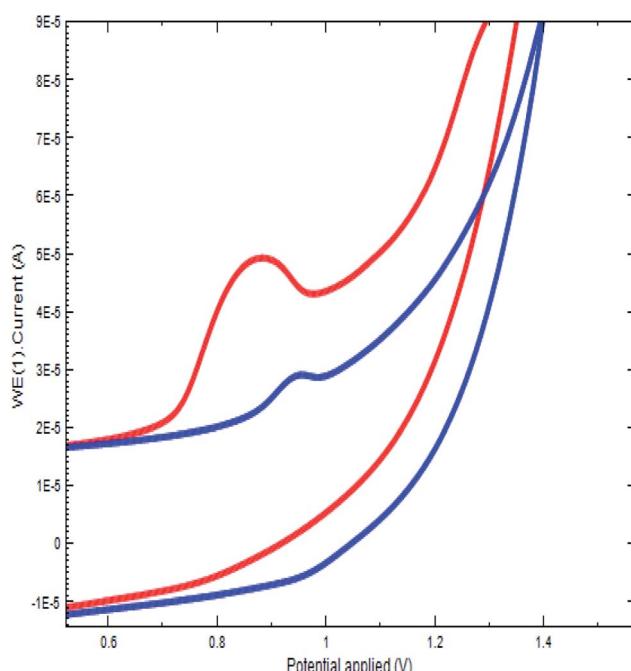


Fig. 3 Electrocatalytic oxidation of DU at bare PGE (blue) and Bi<sub>2</sub>O<sub>3</sub>/PGE (red).



PGE. Upon adding 400.0 ng of DU into the electrochemical cell containing the buffer, an irreversible oxidation peak at 900 mV appeared for the bare PGE due to the oxidation of the amine group. At the same time, this experiment was replicated at  $\text{Bi}_2\text{O}_3$ /PGE and the observed oxidation potential shifted towards the less positive direction (Fig. 3). The raised peak intensity and shifting

of the peak potential at the  $\text{Bi}_2\text{O}_3$ /PGE confirms the fast electrode kinetics of  $\text{Bi}_2\text{O}_3$  at the PGE surface.

### 3.3. Scan rate study

Reversibility of the electrode process was established on the basis of cyclic voltammetry experiments conducted at the

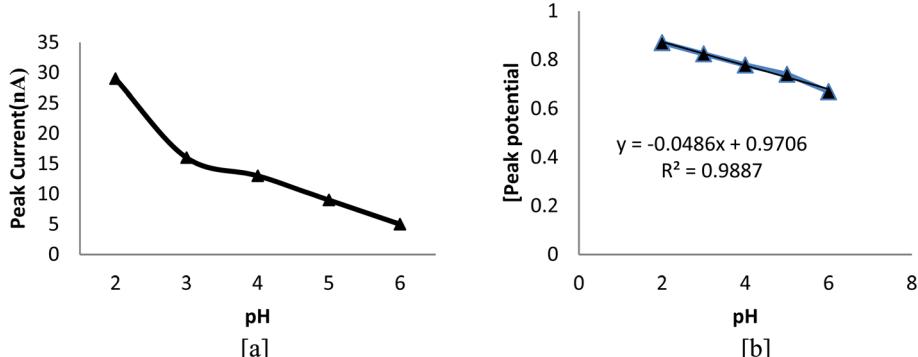


Fig. 4 Plots of (a) pH vs. peak current and (b) pH vs. peak potential.

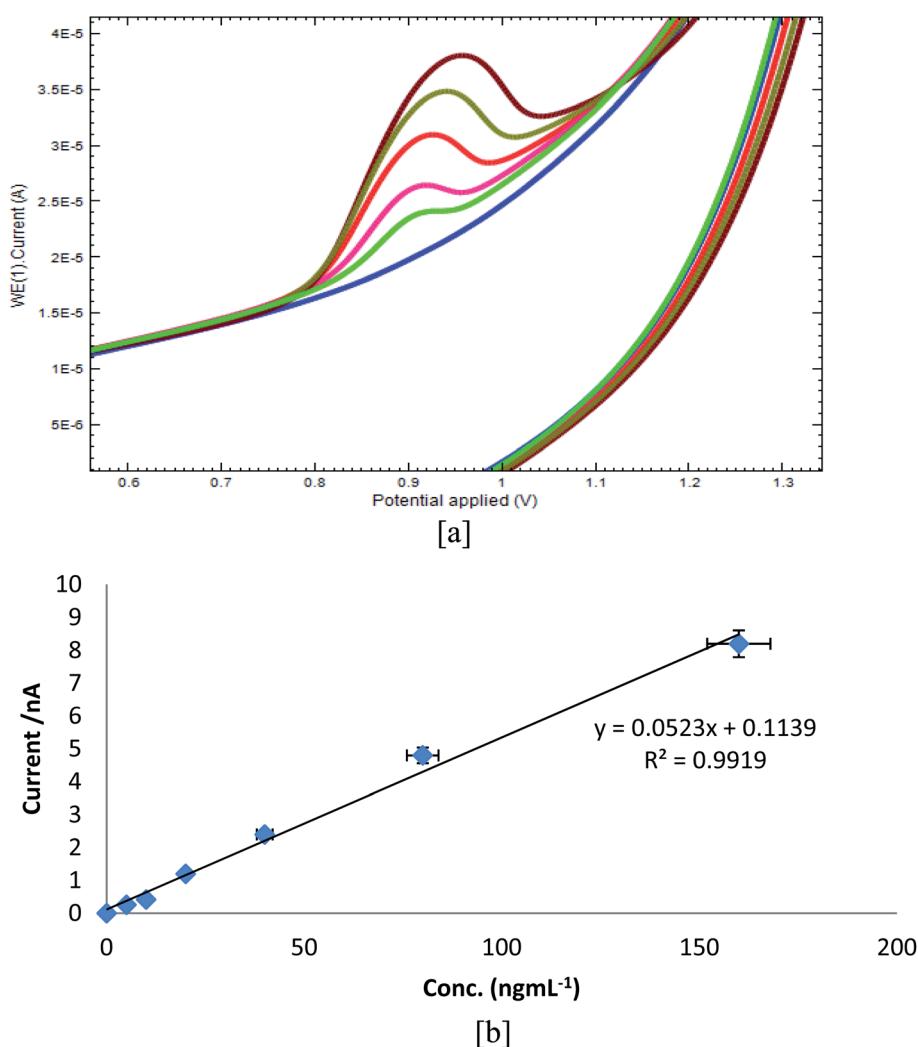


Fig. 5 (a) Voltammograms of different concentrations of DU at  $\text{Bi}_2\text{O}_3$ /PGE (5.0–160.0 ng mL<sup>-1</sup>). (b) Calibration plot concentration vs.  $I_p$ ,  $\mu\text{A}$ .



fabricated  $\text{Bi}_2\text{O}_3/\text{PG}$  sensor. In reverse scan, the absence of reduction peaks proved the irreversible behavior of the electrode phenomena (Fig. 2, ESI†). The effect of varying scan rates in the range of  $10\text{--}100\text{ mV s}^{-1}$  on the peak intensity at different concentrations (800.0 ng) of DU in a BR buffer of pH 2.5 was investigated. In every case, a minute shift in the peak potential with increase in the scan rate and followed by an increase in the peak intensity was examined. The calibration curve plotted between peak currents ( $I_p$ ) and the square root of scan rate ( $\nu^{1/2}$ ) (Fig. 2b, ESI†) was found to be linear passing through the origin (eqn (ii)). The slope of the plot of  $\log \nu$  vs.  $\log I_p$  was 0.478, which validated the diffusion-controlled behaviour of the electrode phenomena (Fig. 2c, ESI†; eqn (iii)):

$$\text{DU}, I_p = 0.575x + 0.213\nu^{1/2}, (R^2 = 0.998) \quad (\text{ii})$$

$$\text{DU}, I_p = 0.478x - 0.167\nu, (R^2 = 0.996) \quad (\text{iii})$$

The calibration curve plotted between the peak potential and logarithm of scan rate was observed to be linear, as shown in Fig. 2d (eqn (iv)):

$$E_p = E^0 + (RT/\alpha_{\text{na}}F)[0.78 + \ln(D_0^{1/2}/K_0) + \ln(\alpha_{\text{na}}F\nu/RT)^{1/2}] \quad (\text{iv})$$

here,  $E^0$  indicates the formal potential,  $\alpha$  the transfer coefficient,  $K$  the standard rate constant and  $n$  the number of electrons transported at the electrode surface. Oxidation of DU occurs at the fabricated sensor, and the peak potential depends on  $\log \nu$ , which can be calculated using eqn (v):

$$E_p = 0.034x + 0.743, (R^2 = 0.998) \quad (\text{v})$$

On equating the above two equations, we calculated:

$$(RT/\alpha nF) = 0.034 \quad (\text{vi})$$

The number of electrons transferred during the electrode fabrication is calculated to be 1. The reaction mechanism is given in Scheme 1.

#### 3.4. Effect of experimental parameters

The variation in peak intensity amid various buffer systems such as, BR, phosphate and acetate was observed, and the highest peak intensity was acquired for BR buffer of pH 2.5. Fig. 4a signifies the effect of different pH values on peak current. From the figure, it can be concluded that on going towards less acidic medium, peak intensity decreases and peak potential also slightly shifts to a less positive value (Fig. 4b), implying the involvement of protons in the rate influential step. The finest result was observed at pH 2.5, and hence, it was opted for the further study. The effects of various solvent systems such as methanol, ethanol, acetone, SLS and CTAB were investigated, and the result indicated that the highest peak current was obtained in methanol. The effects of various electrolytes other than the buffer, namely, 0.01 mM hypochloric acid, 0.01 mM sulphuric acid and 0.01 M potassium chloride were also examined, and the maximum peak current was obtained in 2.0 mL of 0.01 mM sulphuric acid.

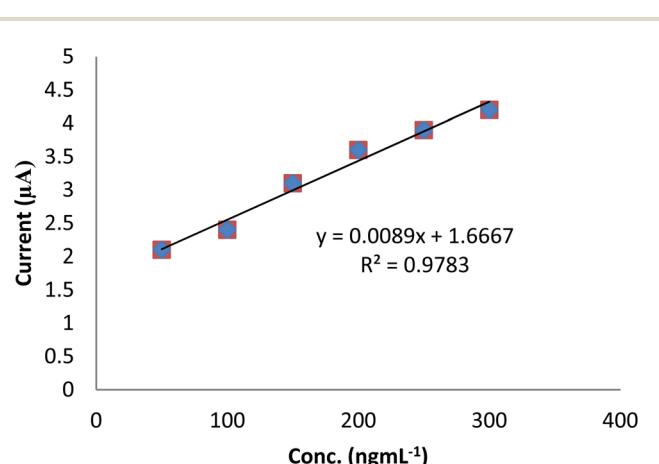
**Table 1** Reproducibility and repeatability of the developed sensor using CV

Sensor	Sensor reproducibility		Single sensor repeatability	
	Mean current ( $I/\text{nA}$ )	RSD	Mean current	RSD
Sensor 1	4.4 <sup>a</sup>	0.12	4.4	0.15
Sensor 2	4.8 <sup>a</sup>	0.17		
Sensor 3	5.2 <sup>a</sup>	0.16		
Average	4.3 <sup>b</sup>	0.15		

<sup>a</sup> Mean of five replicates. <sup>b</sup> Mean of three sensors.

#### 3.5. Calibration curve

Cyclic voltammograms of DU at the  $\text{Bi}_2\text{O}_3/\text{PG}$  sensor under optimized conditions are given in Fig. 5a. It can be concluded that the peak current enhanced upon increasing the concentration of DU. A good linear calibration curve (Fig. 5b) was obtained in the range of 5.0 ng to 160.0 ng with a regression equation,  $I_p = 0.052x + 0.113$ , ( $R^2 = 0.991$ ). The developed sensor was found to be exceedingly sensitive above further reported electrochemical methods, which is evident by the obtained LOD (limit of detection) of  $2.83\text{ ng mL}^{-1}$  as calculated from the formula  $S/N = 3$ , and the LOQ (limit of quantification) of  $8.5\text{ ng mL}^{-1}$ .



**Fig. 6** Plot of  $i_p$  vs. conc. for the analysis of DU in a spiked tomato sample.

**Table 2** Detection of a spiked tomato extract having DU at the developed sensor

S. No	Amount added ( $\text{ng mL}^{-1}$ )	Amount found ( $\text{ng mL}^{-1}$ )	%R	%RSD
1	100.0	99.4	99.9	0.2
2	150.0	148.2	98.8	0.4
3	200.0	199.6	99.8	0.1
4	250.0	249.8	99.9	0.5
5	300.0	298.9	99.6	0.3



Table 3 Comparison of fabricated sensor and other electrodes for the determination of diuron<sup>a</sup>

Sensor	Detection limit	Real Sample	Reference
MWCNT-COOH MIP/CPE	9.0 nM	River water samples	12
rGO–AuNPs/Nafion/GCE	0.3 nM	Tea sample	13
MIP( <i>p</i> -Phe)-PGE	43.43 $\mu$ M	Water samples	40
<b>Bi<sub>2</sub>O<sub>3</sub>/PGE</b>	<b>0.6 nM</b>	<b>Tomatoes</b>	<b>Present work</b>

<sup>a</sup> Abbreviation: rGO/AuNPs/GCE, reduced graphene oxide/gold nanoparticles/glassy carbon electrode; MIP(*p*-Phe)-PGE, molecularly imprinted polymer-pencil graphite electrode; MWCNT-COOH MIP/CPE, molecularly imprinted polymer and carboxyl-functionalized multi-walled carbon nanotubes/carbon paste electrode.

### 3.6. Reproducibility, repeatability, stability and selectivity

The repeatability of the sensor was investigated by calculating the RSD of peak current attained during oxidation of DU from five replicate measurements. The calculated RSD for the modified sensor was found to be 0.15 (Table 1), which indicates the good repeatability. Furthermore, the reproducibility of the developed sensor was studied from the RSD of the current acquired at three comparable developed sensors in the known concentration of 800.0 ng. The obtained RSD of three similar modified sensors was found to be 0.16% (Table 1), which recommends the best reproducibility. Additionally, the fabricated sensor was tested for its stability by examining the reduction in the peak current subsequent to keeping the sensor for 15 days at room temperature. The results indicated that the sensor demonstrates minor decrements in the peak intensity in the earlier first week and maintains approximately 92.0% of its sensitivity, thus signifying the comprehensively consistent stability of the developed sensor. However, selectivity of the modified Bi<sub>2</sub>O<sub>3</sub>/PGE was investigated in the presence of few ions such Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Al<sup>3+</sup>, NO<sub>3</sub><sup>-</sup>, SO<sub>4</sub><sup>2-</sup> and Cl<sup>-</sup>, and the results indicated that the presence of inorganic ions did not affect the quantification.

### 3.7. Recovery analysis

To examine the potential application of the developed electrode in the recovery of real sample analysis, voltammetric determination of DU in spiked tomato was carried out using a standard internal addition method. The recovery analysis of DU was explored using CV technique (Fig. 6). Five different concentrations of DU were studied. A percent recovery of 99.0 real samples was obtained, which demonstrated the appropriateness of the modified sensor and proposed technique (Table 2). For this purpose, previously prepared real samples (Section 2.4) were equally distributed in aliquots in five different volumetric flasks of 5.0 mL, and 2.0 mL of the prepared sample was taken in each flask. Then, the initial flask was subsequently diluted with the solvent up to the mark. The standard solution of DU (200.0 ng mL<sup>-1</sup>) was added in increasing volume (100.0–300.0  $\mu$ L) in succeeding flasks followed by dilution with the solvent up to the mark. The peak intensity was examined for each sample and calibration graph plotted between the analyte concentration and the peak current. To estimate the target analyte quantity in the real sample, linear regression investigated was taken and the slope (*m*) and intercept (*b*) were estimated. The unidentified

concentration of the analyte was quantified using the following eqn (vii):

$$C_x = b \times C_s / m V_x \quad (\text{vii})$$

*C<sub>x</sub>* is the concentration of the sample, *C<sub>s</sub>* the concentration of the standard, *V<sub>x</sub>* the volume of the sample aliquot, *m* the slope of linear regression, and *b* the intercept of linear regression.

From the above equation by substituting the value of “*b*” and “*m*” from the regression equation (*y* = 0.008 + 1.713), *C<sub>s</sub>* = 200.0 ng mL<sup>-1</sup>, and *V<sub>x</sub>* = 2.0 mL, the concentration of DU in the real sample was found to be 215.0 ng mL<sup>-1</sup>.

### 3.8. Comparison of the proposed method with previous reported methods

The analytical performance of the fabricated sensor for the DU quantification at the sensor was compared with different previously reported techniques and given in table (Table 3). The sensitivity of the fabricated sensor was found to be excellent as compared to other reported methods, which suggests that the fabricated Bi<sub>2</sub>O<sub>3</sub>/PGE sensor is highly sensitive for the determination of DU.

## 4. Conclusion

An efficient and novel electrochemical sensor for voltammetric quantification of diuron has been fabricated based on a Bi<sub>2</sub>O<sub>3</sub>-modified pencil graphite electrode. The proposed sensor revealed admirable selectivity, wide linear range (5.0 ng to 160.0 ng), a higher sensitivity of 2.83 ng, fine selectivity, reproducibility and stability. Additionally, the fabricated sensor has advantages of easy preparation procedure, cost-effectiveness and better electrocatalytic performance for the pesticide.

## Conflicts of interest

Authors declare no conflict of interest.

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## References

1 A. Wong, M. V. R. Lanza and M. D. P. T. Sotomayor, *J. Electroanal. Chem.*, 2013, **690**, 83–88, DOI: 10.1016/j.jelechem.2012.11.007.

2 Environmental Monitoring Branch, *Pesticide Chemistry Database*, Department of Pesticide Regulation (DPR) California, 2003.

3 L. Guzzella, E. Capri, A. D. Corcia, A. B. Carracciolo and G. Giuliano, *J. Environ. Qual.*, 2006, **35**, 312–323, DOI: 10.2134/jeq2004.0025.

4 K. Abass, P. Reponen, M. Turpeinen, J. Jalonen and O. Pelkonen, *Drug Metab. Dispos.*, 2007, **35**, 1634–1641, DOI: 10.1124/dmd.107.016295.

5 A. M. Polcaro, M. Mascis, S. Palmas and A. Vacca, *Electrochim. Acta*, 2004, **49**, 649–656, DOI: 10.1016/j.electacta.2003.09.021.

6 C. Tixier, M. Sancelme, F. Bonnemoy, A. Cuer and H. Veschambre, *Environ. Toxicol. Chem.*, 2001, **20**, 1381–1389, DOI: 10.1002/etc.5620200701.

7 C. Lourencetti, M. M. R. Marchi and M. L. Ribeiro, *Talanta*, 2008, **77**, 701–709, DOI: 10.1016/j.talanta.2008.07.013.

8 V. Mani, R. Devasenathipathy, S. M. Chen, T. Y. Wu and K. Kohilarani, *Ionics*, 2015, **21**, 2675–2683, DOI: 10.1007/s11581-015-1459-2.

9 T. Mugadza and T. Nyokong, *Electrochim. Acta*, 2016, **55**, 2606–2613, DOI: 10.1016/j.electacta.2009.12.051.

10 T. Mugadza and T. Nyokong, *Talanta*, 2010, **81**, 1373–1379, DOI: 10.1016/j.talanta.2010.02.037.

11 P. Sharma, K. Sablok, V. Bhalla and C. R. Suri, *Biosens. Bioelectron.*, 2011, **26**, 4209–4212, DOI: 10.1016/j.bios.2011.03.019.

12 A. Wonga, M. V. Foguela, S. Khana, F. M. de Oliveirab, C. R. T. Tarley and M. D. P. T. Sotomayora, *Electrochim. Acta*, 2015, **182**, 122–130, DOI: 10.1016/j.electacta.2015.09.054.

13 K. Zarei and A. Khodadadi, *Ecotoxicol. Environ. Saf.*, 2017, **144**, 171–177, DOI: 10.1016/j.ecoenv.2017.06.030.

14 H. Karimi-Maleh, F. Karimi, M. Alizadeh and A. L. Sanati, *Chem. Rec.*, 2020, **20**, 1–12, DOI: 10.1002/tcr.201900092.

15 F. Tahernejad-Javazmi, M. Shabani-Nooshabadi and H. Karimi-Maleh, *Composites, Part B*, 2019, **172**, 666–670, DOI: 10.1016/j.compositesb.2019.05.065.

16 H. Karimi-Maleh, M. S. Mohammad, A. Taherd, F. Opokuc, E. Muriithi, K. Poomani, P. Govender, S. Ranjbari, M. Rezapoure and Y. Orooji, *J. Mol. Liq.*, 2020, **298**, 112040, DOI: 10.1016/j.molliq.2019.112040.

17 H. Karimi-Maleh, C. T. Fakude, N. Mabuba and G. M. Peleyeju, *J. Colloid Interface Sci.*, 2019, **554**, 603–610, DOI: 10.1016/j.jcis.2019.07.047.

18 H. Karimi-Maleh and O. A. Arotiba, *J. Colloid Interface Sci.*, 2020, **560**, 208–212, DOI: 10.1016/j.jcis.2019.10.007.

19 Z. Shamsadin-Azad, M. A. Tahe, S. Cheraghi and H. Karimi-Maleh, *J. Food Meas. Charact.*, 2019, **13**, 1781–1787, DOI: 10.1007/s11694-019-00096-6.

20 F. E. Gorla, H. Duarte, E. R. Sartori and C. R. T. Tarley, *Microchem. J.*, 2015, **124**, 65–75, DOI: 10.1016/j.microc.2015.07.021.

21 H. Rodrigues, S. Lima, S. J. Silva, E. A. O. Farias, P. R. S. Teixeira, C. Eiras and L. C. C. Nunes, *Biosens. Bioelectron.*, 2018, **108**, 27–37, DOI: 10.1016/j.bios.2018.02.034.

22 M. Ghanei-Motlagh and M. Baghayeri, *J. Electrochem. Soc.*, 2020, **167**, 066508, DOI: 10.1149/1945-7111/ab823c.

23 M. Nodehi, M. Baghayeri, R. Ansari and H. Veisi, *Mater. Chem. Phys.*, 2020, **244**, 122687, DOI: 10.1016/j.matchemphys.2020.122687.

24 M. Baghayeri, M. Ghanei-Motlagh, R. Tayebee, M. Fayazi and F. Narenji, *Anal. Chim. Acta*, 2020, **1099**, 60–67, DOI: 10.1016/j.aca.2019.11.045.

25 M. Rouhi, M. M. Lakouraj and M. Baghayeri, *Polymer Composites*, 2019, DOI: 10.1002/pc.24790.

26 M. Ghanei-Motlagh, M. A. Taher, M. Fayazi, M. Baghayeri and A. R. Hosseinfar, *J. Electrochem. Soc.*, 2019, **166**, B367–B372, DOI: 10.1149/2.0521906jes.

27 M. Baghayeri, R. Ansari, M. Nodehi, I. Razavipanah and H. Veisi, *Electroanalysis*, 2018, **30**, 1–8, DOI: 10.1002/elan.201800158.

28 M. Baghayeri, R. Ansari, M. Nodehi and H. Veisi, *Int. J. Environ. Anal. Chem.*, 2018, **98**, 874–888, DOI: 10.1080/03067319.2018.1512595.

29 M. Baghayeri, R. Ansari, M. Nodehi, I. Razavipanah and H. Veisi, *Microchim. Acta*, **185**, 320, DOI: 10.1007/s00604-018-2838-y.

30 Y. Jing, B. Yu, P. Li, B. Xiong, Y. Cheng, Y. Li, C. Li, X. Xiao, M. Chen, L. Chen, Y. Zhang, M. Zhao and C. Cheng, *Sci. Rep.*, 2017, **7**, 14332, DOI: 10.1038/s41598-017-13716-2.

31 A. H. Oghli, E. Alipour and M. Asadzadeh, *RSC Adv.*, 2015, **13**, DOI: 10.1039/c4ra11399c.

32 A. Rana and A. N. Kawde, *J. Chin. Chem. Soc.*, 2016, **63**, 668–676, DOI: 10.1002/jccs.201600119.

33 M. Taheri, F. Ahour and S. Keshipour, *J. Phys. Chem. Solids*, 2018, **117**, 180–187, DOI: 10.1016/j.jpcs.2018.02.035.

34 S. Pourbeyram and K. Mehdizadeh, *J. Food Drug Anal.*, 2017, **24**, 894–902, DOI: 10.1016/j.jfda.2016.02.010.

35 I. G. David, D. E. Popa and M. Buleandra, *J. Anal. Methods Chem.*, 2017, **2017**, 1–22, DOI: 10.1155/2017/19059.

36 S. B. Hocoevar, B. Ogorevc, J. Wang and B. Pihlarc, *Electroanalysis*, 2017, **14**, 1707–1712, DOI: 10.1002/elan.200290014.

37 E. S. Sá, P. S. Silva, C. L. Jost and Spinelli, *Sens. Actuators, B*, 2015, **209**, 423–430, DOI: 10.1016/j.snb.2014.11.136.

38 H. W. Wang, Z. A. Hu, Y. Q. Chang, Y. L. Chen, Z. Q. Lei, Z. Y. Zhang and Y. Y. Yang, *Electrochim. Acta*, 2010, **55**, 8974–8980, DOI: 10.1016/j.electacta.2010.08.048.

39 M. Shahbazi, L. Faghfouri, M. P. A. Ferreira, P. Figueiredo, H. Maleki, F. S. de, J. Hirvonen and H. A. Santos, *Chem. Soc. Rev.*, 2020, **4**, DOI: 10.1039/c9cs00283a.

40 B. Önde and M. Soysal, *J. Electrochem. Soc.*, 2019, **166**, B395–B401, DOI: 10.1149/2.0631906jes.

41 A. García-Miranda Ferrari, C. W. Foster, P. J. Kelly, D. A. C. Brownson and C. E. Banks, *Biosensors*, 2018, **8**, 1–10, DOI: 10.3390/bios8020053.

42 T. N. Huana, T. Ganesha, K. S. Kimb, S. Kima, S. Hana and H. Chunga, *Biosens. Bioelectron.*, 2011, **8**, 53, DOI: 10.1016/j.bios.2011.06.011 2018.

