



Cite this: *Nanoscale Adv.*, 2020, **2**, 166

Received 23rd October 2019  
Accepted 20th November 2019

DOI: 10.1039/c9na00671k  
[rsc.li/nanoscale-advances](http://rsc.li/nanoscale-advances)

## Regioselective thiocyanation of corroles and the synthesis of gold nanoparticle–corrole assemblies<sup>†‡</sup>

Kasturi Sahu,<sup>ab</sup> Sruti Mondal,<sup>ab</sup> Bratati Patra,<sup>ab</sup> Tanmoy Pain,<sup>ab</sup> Sajal Kumar Patra,<sup>ab</sup> Carsten Dosche <sup>\*c</sup> and Sanjib Kar <sup>\*ab</sup>

Herein we demonstrate a synthetic protocol for the regioselective thiocyanation of corroles. To the best of our knowledge, thiocyanato appended corrole has never been reported earlier. The resulting thiocyanato appended corrole turned out to be a good corrole based precursor for the facile synthesis of thiol protected gold nanoparticles (Au NPs). The ligand system acts as a good bidentate framework and passivates the gold surface. A strong electronic interaction between the corrole and the gold nanoparticles is manifested by their unique photo physical properties and it also confirms that the binding through  $\beta$ -substitutions has a more pronounced effect even though the corrole rings are face-off to the gold surface.

Gold nanoparticles (Au NPs) are extensively used in various areas like catalysis,<sup>1</sup> medicine,<sup>2</sup> sensors,<sup>3</sup> fuel cells,<sup>4</sup> surface enhanced Raman spectroscopy,<sup>5</sup> probes for transmission electron microscopy,<sup>6</sup> and antibiotics.<sup>7</sup> One can easily coat the surface of Au NPs with various molecules, either small or big, and thus can fine tune their physical and chemical properties.<sup>8</sup> In this context, Au NPs bearing  $\pi$ -conjugated molecules are ideal platforms for various nanoelectronic devices.<sup>9</sup> Thus there is widespread research interest in studying porphyrinoid coated gold nanoparticles.<sup>10</sup> Feringa *et al.* demonstrated that porphyrins having  $\pi$ -conjugation can coordinate with the Au (111) surface *via* intramolecular interactions (*via* linker groups) and also *via* interfacial interactions (*via* inner nitrogens).<sup>11</sup> Corrole, a ring contracted version of porphyrin, has interesting and different photo physical properties in comparison to

porphyrin.<sup>12</sup> Thus, it is expected that corrole coated Au NPs will show great promise in designing newer varieties of functional materials having diverse applications. The importance of the multi-dentate ligand in binding the surface of the Au NPs in a specific orientation was demonstrated independently by Ohyama *et al.*<sup>13</sup> and Teranishi *et al.*<sup>14</sup> Very few research papers indeed highlight the importance of polydentate ligands as binders of gold nanoparticles.<sup>15</sup> Tour *et al.* reported that organic thiocyanates can easily form assemblies on gold surfaces without the help of any external agent and have several advantages over free thiols or other thiol based precursors.<sup>16</sup> Their observations have prompted us to choose the thiocyanato appended corrole as a possible precursor for the synthesis of gold nanoparticle–corrole assemblies. The existing literature reports that *meso*-substituted porphyrinoids are used exclusively as binders for gold nanoparticles, and the related  $\beta$ -substituted derivatives are rarely used for this purpose. As  $\beta$ -substitution has a more pronounced effect on the electronic structure, we have chosen  $\beta$ -substitution at the corrole periphery. In order to obtain the thiocyanato appended corrole derivative, we have developed a simple and novel protocol for the regio-selective thiocyanation of corrole. To the best of our knowledge, the thiocyanato group has never been inserted at the corrole periphery. We have synthesized corrole coated gold nanoparticles by using a one-phase strategy.<sup>13</sup> Corrole and  $\text{HAuCl}_4$  solutions were mixed together, and then a  $\text{NaBH}_4$  solution was added and stirred for  $\sim 3$  hours. No co-stabilizers (citrate, TOAB, *etc.*) were used for this purpose. The thiocyanato appended corrole, 3,17-bis(thiocyanato)-5,10,15-triphenylcorrole (**1**), was prepared by following a strategy developed by our group (Scheme 1).

5,10,15-triphenylcorrole (TPC) in a carbon disulfide solution was treated with an excess amount of  $\text{NH}_4\text{SCN}$  and excess pyridine. Refluxing for 3 hours resulted in the formation of **1** in good yields (see the ESI<sup>‡</sup> and Scheme 1). The newly synthesized thiocyanato appended corrole (**1**) has been fully characterized using several spectroscopic techniques like CHN analysis, FT-IR analysis,  $^1\text{H}$  NMR, ESI-mass spectrometry, XPS and single

<sup>a</sup>School of Chemical Sciences, National Institute of Science Education and Research (NISER), Bhubaneswar, Khordha, 752050, India

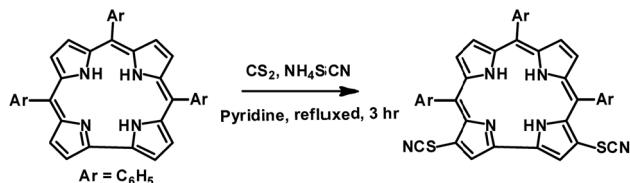
<sup>b</sup>Homi Bhabha National Institute, Training School Complex, Anushakti Nagar, Mumbai, 400094, India. E-mail: [sanjib@niser.ac.in](mailto:sanjib@niser.ac.in)

<sup>c</sup>University of Oldenburg, Institute of Chemistry, Carl-von-Ossietzky-Str. 9-11, 26129 Oldenburg, Germany. E-mail: [carsten.dosche@uol.de](mailto:carsten.dosche@uol.de)

† This contribution is dedicated to the occasion of the 60<sup>th</sup> birthday of Professor Goutam Kumar Lahiri.

‡ Electronic supplementary information (ESI) available: Experimental section, NMR, mass spectrometry, FT-IR, UV-Vis, emission and X-ray data. See DOI: [10.1039/c9na00671k](https://doi.org/10.1039/c9na00671k)





Scheme 1 Synthetic route to 3,17-bis(thiocyanato)-5,10,15-triphenylcorrole, **1**.

crystal XRD (see the ESI; Fig. S1–S6 and Table S1, S2‡). Considering thiocyanate to be a potent nucleophile, this reaction should be considered a unique class of the nucleophilic aromatic substitution reaction in corrole.<sup>17</sup> Paolesse *et al.* described this class of reactions as nucleophilic aromatic substitution of hydrogens, occurring *via* two steps. The first step is the formation of a  $\sigma$ -adduct by simple addition of the nucleophile to the corrole ring and the second step is the elimination of hydride. Pyridine is suspected to be a hydride ion abstractor here. The  $^1\text{H}$  NMR spectrum of **1** exhibits normal chemical shifts in accordance with the ‘twenty one’ aromatic protons in the region  $\delta$ ,  $\sim$ 8.91–7.78 ppm. Negligible shielding is observed in **1** in comparison with 5,10,15-triphenylcorrole (see ESI Fig. S4‡). The FT-IR spectrum of **1** as a KBr pellet shows a peak at  $2154\text{ cm}^{-1}$  due to  $\text{C}\equiv\text{N}$  group stretching (see ESI Fig. S3‡). The crystal structure of **1** is shown in Fig. 1.

The crystal system is triclinic, and the unit cell contains two molecules of **1**. Important crystallographic parameters for compound **1** are shown in Table S1.‡ Bond distances and angles of **1** are in agreement with those of other FB corrole derivatives.<sup>12</sup> Pyrrolic nitrogen atoms have shown deviations from the mean corrole plane (considering the 19-carbon atom plane to be the mean corrole plane) by distances varying from 0.488 to  $-0.052\text{ \AA}$  in **1**. The corresponding deviations in 5,10,15-triphenylcorrole (TPC) are 0.128 to  $-0.195\text{ \AA}$ .<sup>12v</sup> The dihedral angles between the planes of the phenyl rings (*meso*-substituted phenyl group) and the 19-carbon mean corrole plane vary significantly and are 43.74–78.37°. The corresponding dihedral angles in TPC are 41.74–50.33°.<sup>12v</sup> Thus, it appears that the insertion of two thiocyanate groups at the  $\beta$ -pyrrolic position induced

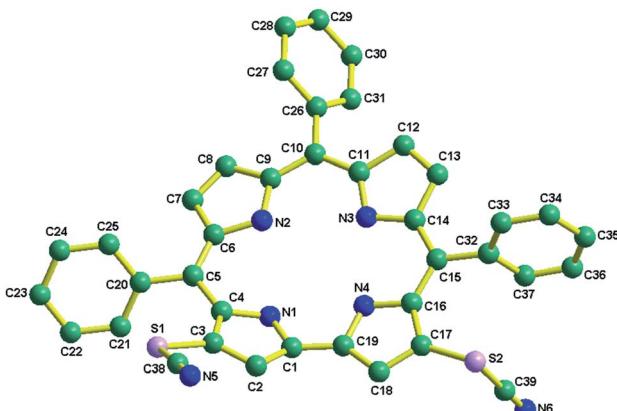


Fig. 1 Perspective view of **1**. Hydrogen atoms are omitted for clarity.

significant saddling of the corrole framework. The two  $\text{C}\equiv\text{N}$  (for N (5)–C (38) and N (6)–C (39)) bond distances of **1** are  $1.13\text{ \AA}$  and  $1.14\text{ \AA}$  and the two  $\text{S}-\text{C}\equiv\text{N}$  bond angles (for S (1)–C (38)–N (5) and S (2)–C (39)–N (6)) are  $177.66^\circ$  and  $178.85^\circ$ .

Gold nanoparticles coated with **1** were prepared by addition of  $\text{NaBH}_4$  to a premixed mixture of **1** and  $\text{HAuCl}_4$  (see the ESI‡). The newly synthesized **1**-Au NPs were characterized by CHN, EDS and FT-IR analysis (see the ESI, Fig. S3 and S7‡). The EDS spectrum of **1**-Au NPs shows the presence of all the constituent elements (Au, N, and S). From CHN analysis (see ESI‡), we can roughly estimate that there are 200 corrole ligands on a single Au NP surface. This implies that there are many physisorbed corrole molecules at the surface which do not have direct contact with the gold surface. The UV-Vis spectra of the solid samples of **1** (very broad and red shifted) clearly indicate extensive aggregate formation (Fig. S1‡). The FT-IR spectrum of Au NP–corrole assemblies obtained using a KBr pellet shows the absence of  $\text{C}\equiv\text{N}$  group stretching at  $2154\text{ cm}^{-1}$  (see the ESI, Fig. S3‡). This confirms that the ligand moiety loses the  $\text{C}\equiv\text{N}$  group while binding with the Au surface. Fig. 2 shows the time evolution of the UV-Vis spectra for mixtures of **1** ( $\sim 10^{-6}\text{ M}$ ),  $\text{NaBH}_4$  ( $\sim 10^{-4}\text{ M}$ ), and  $\text{HAuCl}_4$  ( $\sim 10^{-5}\text{ M}$ ) in DMF. The initial spectrum (black colour, colour online) was for pure **1** in the DMF solution. It was observed that the positions of the Soret and Q bands in **1** are negligibly changed upon addition of  $\text{NaBH}_4$ . However, both bands are drastically different in intensity upon gradual addition of the  $\text{HAuCl}_4$  solution. The red coloured final spectrum (colour online) was obtained after the addition of optimal concentration of the  $\text{HAuCl}_4$  ( $\sim 2 \times 10^{-5}\text{ M}$ ) solution. The final spectrum obtained after 2.5 hours shows a clear signature of the combination of the characteristic Soret band of corrole **1** (as a weak shoulder) and the strong plasmon band of gold nanoparticles. The surface plasmon resonance (SPR) band appears at  $525\text{ nm}$ .<sup>13,14</sup> The intensity of the Q bands of **1** is drastically reduced but it still remains as a shoulder at around  $655\text{ nm}$ . However, the Soret bands are broadened but the shifts of the bands are negligible. A direct comparison of these UV-Vis spectra with that of the FB corrole **1** (prior to Au-thiol contact formation) is not possible because after the

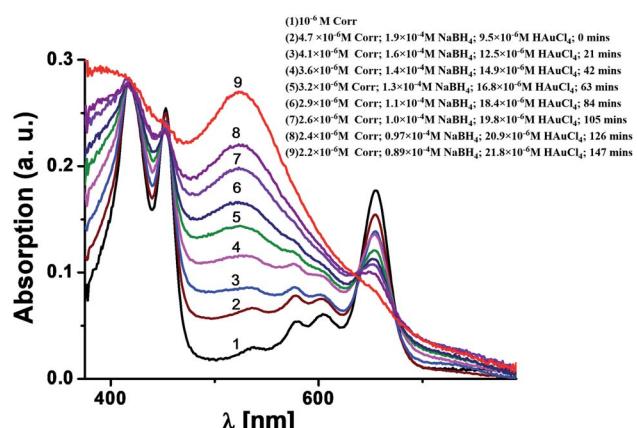


Fig. 2 Time evolution UV-vis spectra of a mixture of **1** ( $\sim 10^{-6}\text{ M}$ ),  $\text{NaBH}_4$  ( $\sim 10^{-4}\text{ M}$ ), and  $\text{HAuCl}_4$  ( $\sim 10^{-5}\text{ M}$ ) in DMF.

interaction of FB corrole **1** with the gold nanoparticles, the thiocyanate group loses the cyanide fragment (after Au–thiol contact formation) and thus the ligand electronic structure is perturbed.

These results are in complete agreement with those for previously reported porphyrin thioester protected gold nanoparticles.<sup>14</sup> In line with the previous report by Teranishi *et al.*, our absorption spectra also suggest that a strong electronic interaction exists between the corrole and the gold surface (Fig. 3).<sup>14</sup> However, in contrast to them, we believe that in our case the parallel binding mode of corrole to the gold surface is not feasible due to the steric hindrance of the *meso*-aryl groups. Thus a face-off binding mode of corrole (due to the lack of one *meso*-carbon) is proposed here. Fig. 4 shows the time evolution fluorescence spectra of a mixture of **1** ( $\sim 10^{-6}$  M),  $\text{NaBH}_4$  ( $\sim 10^{-4}$  M), and  $\text{HAuCl}_4$  ( $\sim 10^{-5}$  M) in DMF.

The initial spectrum (black colour, colour online) was for pure **1** in the DMF solution. The position of the emission bands is negligibly changed upon addition of  $\text{NaBH}_4$ . However, a drastic change of the corrole fluorescence intensity is observed upon gradual addition of the  $\text{HAuCl}_4$  solution. The fluorescence quenching is obvious as the corrole molecules transfer their excitation energy to the gold nanoparticles.<sup>18</sup> The excited state lifetimes of **1** and **1**-Au NPs were found to be 3.69 ns and 1.19 ns, respectively (Fig. 5). Considering that only static quenching is viable,<sup>18a</sup> the change in the lifetime of corrole **1** on binding with the Au surface is attributed to the change of the

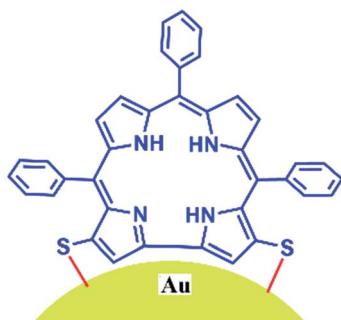


Fig. 3 Plausible structure of gold nano-corrole assemblies.

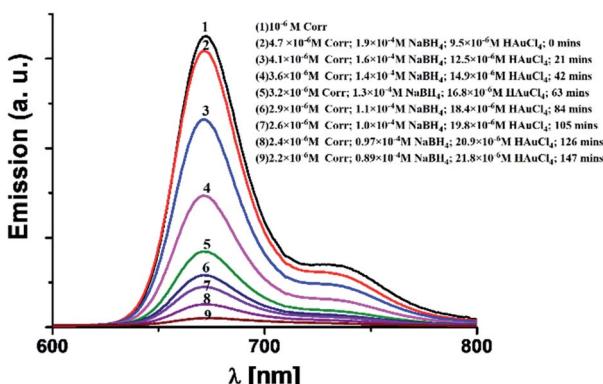


Fig. 4 Time evolution emission spectra of a mixture of **1** ( $\sim 10^{-6}$  M),  $\text{NaBH}_4$  ( $\sim 10^{-4}$  M), and  $\text{HAuCl}_4$  ( $\sim 10^{-5}$  M) in DMF.

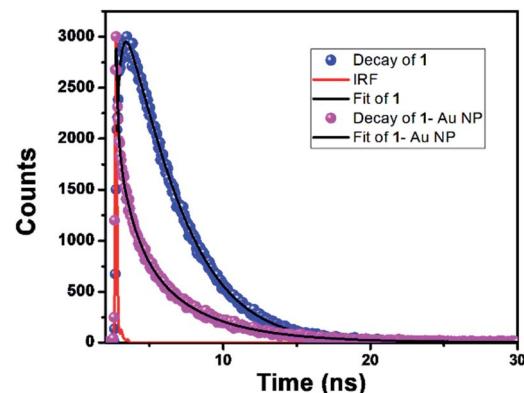


Fig. 5 Fluorescence decay profiles of (a) **1** (~10<sup>-6</sup> M);  $\lambda_{\text{em}} = 672$  nm and (b) **1**-Au NPs (~10<sup>-6</sup> M);  $\lambda_{\text{em}} = 672$  nm. The red line represents the IRF.

ligand structure in **1**-Au NPs (the two thiocyanate groups are converted to thiol groups on binding with Au).

A representative TEM image is shown in Fig. 6. One can easily visualize the diffraction contrast fringes and thus it can confirm the crystalline nature of the **1** coated gold nanoparticles.<sup>13</sup> Particle size analysis indicates that the mean diameter of the **1** coated gold nanoparticles is  $\sim 6.5$  nm.

Gold nanoparticles coated with **1** are stable under ambient conditions for a few weeks as concluded from the characteristic surface plasmon resonance (SPR) band at 525 nm. This is in contrast with the stability of gold nanoparticles alone obtained through  $\text{NaBH}_4$  reduction without the use of the stabilizing corrole molecule, **1**.<sup>13</sup>

**1**-Au NPs were also studied by X-ray photoelectron spectroscopy (XPS). The composition of the organic contributions is 85 at% carbon, 11 at% nitrogen and 4 at% sulphur, approximately matching the expected ratio of 87 : 9 : 4 for **1** with cleaved CN groups. The N 1s signal (Fig. 7) consists of three components which can be assigned to imine nitrogen (398.2 eV, 1.3 at%), amine nitrogen (399.9 eV, 9.6 at%) and amide nitrogen (400.3 eV).

The latter is most probably due to the residual DMF which is left after synthesis and was therefore removed from the calculation of at% for the organic components. The imine and amine nitrogen contributions can be assigned to the four nitrogen atoms in the corrole framework.<sup>19</sup> However, the ratio between

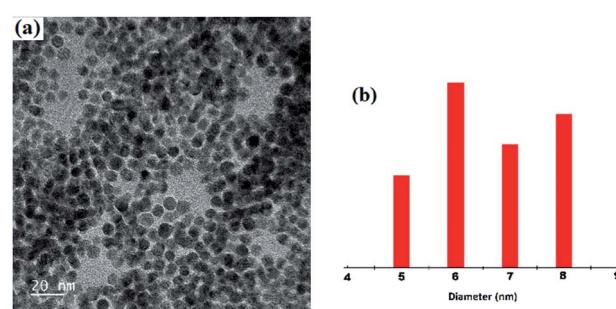


Fig. 6 (a) TEM image of the nanoparticles (the inset shows the diffraction contrast fringes) and (b) histograms of the particle size.



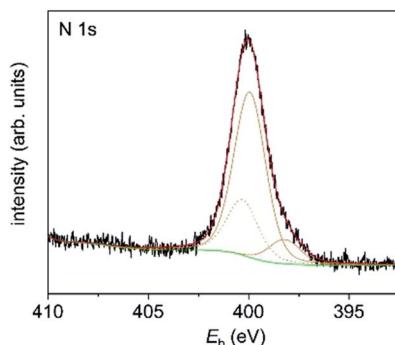


Fig. 7 N 1s XP spectrum of **1**-Au NPs on a Si substrate. Measured data are in black, fitted components are in orange, the background is in green and the sum is in red.

amine and imine nitrogen is much higher than expected for pure corrole and might be an indication of extensive  $\pi$ - $\pi$  stacking. The S 2p signal (Fig. 8) is also composed of three signals, which can be assigned to thiol bound to the gold surface (162.0 eV, 0.8 at%), free thiol (164.0 eV, 1.7 at%) and oxidized thiol (168.1 eV, 1.2 at%).<sup>20</sup> As the ratio of corrole ligands per Au NP has been estimated from CHN analysis to be 200 : 1, it is obvious that only a minor number of corrole ligands can bind directly to the gold surface. Thus, many ligands are only adsorbed by  $\pi$ - $\pi$  stacking and thus have free thiol groups which can easily be oxidized.

In conclusion, we have developed a simple and novel protocol for the regio-selective thiocyanation of corrole. The thiocyanato appended corrole **1** acts as a stabilizer for gold nanoparticles. A unique face-off binding mode in corrole (due to the lack of one *meso*-carbon) is proposed here which is not at all feasible in porphyrins. The corrole **1** and the Au nanoparticle hybrid exhibits interesting photo physical properties. The changes in absorption and emission spectra are remarkable and these also clearly indicate that a strong electronic interaction exists between the corrole and the Au nanoparticles. These studies will certainly be helpful in designing various multi-

dentate corrole based ligands in order to stabilize Au nanoparticles with a specific size and functions depending on the choice of corrole framework.

## Conflicts of interest

There are no conflicts of interest to declare.

## Acknowledgements

Financial support received from the Department of Atomic Energy, (India) is gratefully acknowledged. Authors acknowledge NISER-Bhubaneswar for providing infrastructure and instrumental supports. The authors acknowledge the financial support provided by SERB (Science & Engineering Research Board), India (Grant No: EMR/2016/005484) to carry out this research work. Funding from the DFG, Germany under grant INST 184/106-1 FUGG is acknowledged. The XPS spectrometer is co-financed by the German Science Foundation (INST 184/106-1 FUGG). We would like to acknowledge Dr M. Sadhukhan for TEM Measurement.

## Notes and references

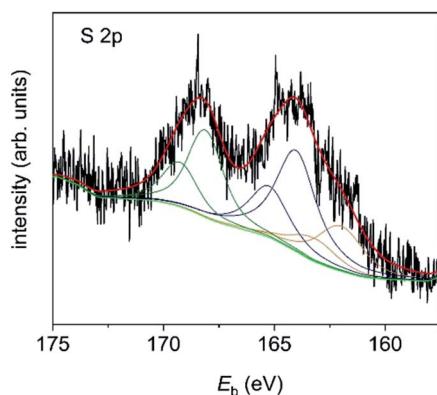


Fig. 8 S 2p spectrum of **1**-Au NPs on a Si substrate. Measured data are in black, the fit for bound thiol is in orange, the fit for free thiol is in blue, the fit for oxidized thiol is in dark green, the background is in green and the sum is in red.

- (a) M.-C. Daniel and D. Astruc, *Chem. Rev.*, 2004, **104**, 293–346; (b) A. Corma and H. Garcia, *Chem. Soc. Rev.*, 2008, **37**, 2096–2126; (c) C. T. Campbell, *Science*, 2004, **306**, 234–235.
- (a) D. A. Giljohann, D. S. Seferos, W. L. Daniel, M. D. Massich, P. C. Patel and C. A. Mirkin, *Angew. Chem., Int. Ed.*, 2010, **49**, 3280–3294; (b) E. C. Dreaden, A. M. Alkilany, X. Huang, C. J. Murphy and M. A. El-Sayed, *Chem. Soc. Rev.*, 2012, **41**, 2740–2779; (c) J. J. Li, L. Zou, D. Hartono, C. N. Ong, B. H. Bay and L. Y. Lanry Yung, *Adv. Mater.*, 2008, **20**, 138–142.
- (a) S. Chah, M. R. Hammond and R. N. Zare, *Chem. Biol.*, 2005, **12**, 323–328; (b) G. Peng, U. Tisch, O. Adams, M. Hakim, N. Shehada, Y. Y. Broza, S. Billan, R. Abdah-Bortnyak, A. Kuten and H. Haick, *Nat. Nanotechnol.*, 2009, **4**, 669.
- (a) Y.-C. Lu, Z. Xu, H. A. Gasteiger, S. Chen, K. Hamad-Schifferli and Y. Shao-Horn, *J. Am. Chem. Soc.*, 2010, **132**, 12170–12171; (b) R. Muszynski, B. Seger and P. V. Kamat, *J. Phys. Chem. C*, 2008, **112**, 5263–5266; (c) J. Luo, L. Wang, D. Mott, P. N. Njoki, Y. Lin, T. He, Z. Xu, B. N. Wanjana, I. I. S. Lim and C. J. Zhong, *Adv. Mater.*, 2008, **20**, 4342–4347.
- (a) C. E. Talley, J. B. Jackson, C. Oubre, N. K. Grady, C. W. Hollars, S. M. Lane, T. R. Huser, P. Nordlander and N. J. Halas, *Nano Lett.*, 2005, **5**, 1569–1574; (b) K. L. Wustholz, A.-I. Henry, J. M. McMahon, R. G. Freeman, N. Valley, M. E. Piotti, M. J. Natan, G. C. Schatz and R. P. Van Duyne, *J. Am. Chem. Soc.*, 2010, **132**, 10903–10910; (c) N. Féridj, J. Aubard, G. Lévi, J. R. Krenn, A. Hohenau, G. Schider, A. Leitner and F. R. Aussenegg, *Appl. Phys. Lett.*, 2003, **82**, 3095–3097.
- (a) T. A. Taton, C. A. Mirkin and R. L. Letsinger, *Science*, 2000, **289**, 1757–1760; (b) J. J. Storhoff, R. Elghanian, R. C. Mucic, C. A. Mirkin and R. L. Letsinger, *J. Am. Chem. Soc.*, 1998,



120, 1959–1964; (c) S. G. Penn, L. He and M. J. Natan, *Curr. Opin. Chem. Biol.*, 2003, **7**, 609–615.

7 I. Yacoby and I. Benhar, *Infect. Disord.: Drug Targets*, 2007, **7**, 221–229.

8 (a) J. S. Lee, M. S. Han and C. A. Mirkin, *Angew. Chem., Int. Ed.*, 2007, **46**, 4093–4096; (b) G. H. Woehrle, L. O. Brown and J. E. Hutchison, *J. Am. Chem. Soc.*, 2005, **127**, 2172–2183; (c) S.-Y. Lin, S.-W. Liu, C.-M. Lin and C.-h. Chen, *Anal. Chem.*, 2002, **74**, 330–335; (d) K.-S. Kim, D. Demberelnyamba and H. Lee, *Langmuir*, 2004, **20**, 556–560.

9 (a) B. C. Sih and M. O. Wolf, *Chem. Commun.*, 2005, 3375–3384; (b) M. Ikeda, N. Tanifuji, H. Yamaguchi, M. Irie and K. Matsuda, *Chem. Commun.*, 2007, 1355–1357; (c) J. van Herrikhuyzen, S. J. George, M. R. Vos, N. A. Sommerdijk, A. Ajayaghosh, S. C. Meskers and A. P. Schenning, *Angew. Chem., Int. Ed.*, 2007, **119**, 1857–1860; (d) K. Matsuda, H. Yamaguchi, T. Sakano, M. Ikeda, N. Tanifuji and M. Irie, *J. Phys. Chem. C*, 2008, **112**, 17005–17010.

10 (a) T. Hasobe, H. Imahori, P. V. Kamat, T. K. Ahn, S. K. Kim, D. Kim, A. Fujimoto, T. Hirakawa and S. Fukuzumi, *J. Am. Chem. Soc.*, 2005, **127**, 1216–1228; (b) T. Akiyama, M. Nakada, N. Terasaki and S. Yamada, *Chem. Commun.*, 2006, 395–397; (c) D. Conklin, S. Nanayakkara, T.-H. Park, M. F. Lagadec, J. T. Stecher, M. J. Therien and D. A. Bonnell, *Nano Lett.*, 2012, **12**, 2414–2419; (d) A. Satake, M. Fujita, Y. Kurimoto and Y. Kobuke, *Chem. Commun.*, 2009, 1231–1233.

11 N. Katsonis, J. Vicario, T. Kudernac, J. Visser, M. M. Pollard and B. L. Feringa, *J. Am. Chem. Soc.*, 2006, **128**, 15537–15541.

12 For selected examples see: (a) E. Vogel, S. Will, A. S. Tilling, L. Neumann, J. Lex, E. Bill, A. X. Trautwein and K. Wieghardt, *Angew. Chem., Int. Ed.*, 1994, **33**, 731–735; (b) Z. Gross, *J. Biol. Inorg. Chem.*, 2001, **6**, 733–738; (c) R. Paolesse, S. Nardis, M. Stefanelli, F. R. Fronczek and M. G. H. Vicente, *Angew. Chem., Int. Ed.*, 2005, **44**, 3047–3050; (d) R. S. Czernuszewicz, V. Mody, A. Czader, M. Galezowski and D. T. Gryko, *J. Am. Chem. Soc.*, 2009, **131**, 14214–14215; (e) M. Autret, S. Will, E. V. Caemelbecke, J. Lex, J.-P. Gisselbrecht, M. Gross, E. Vogel and K. M. Kadish, *J. Am. Chem. Soc.*, 1994, **116**, 9141–9149; (f) J. H. Palmer, A. C. Durrell, Z. Gross, J. R. Winkler and H. B. Gray, *J. Am. Chem. Soc.*, 2010, **132**, 9230–9231; (g) H.-Y. Liu, Y. Fei, Y.-T. Xie, X.-Y. Li and C. K. Chang, *J. Am. Chem. Soc.*, 2009, **131**, 12890–12891; (h) B. Brizet, N. Desbois, A. Bonnot, A. Langlois, A. Dubois, J.-M. Barbe, C. P. Gros, C. Goze, F. Denat and P. D. Harvey, *Inorg. Chem.*, 2014, **53**, 3392–3403; (i) S. Kuck, G. Hoffmann, M. Broering, M. Fechtel, M. Funk and R. Wiesendanger, *J. Am. Chem. Soc.*, 2008, **130**, 14072–14073; (j) K. E. Thomas, H. Vazquez-Lima, Y. Fang, Y. Song, K. J. Gajnon, C. M. Beavers, K. M. Kadish and A. Ghosh, *Chem.-Eur. J.*, 2015, **21**, 16839–16847; (k) M. Bröring, F. Bregier, E. Cónsul Tejero, C. Hell and M. C. Holthausen, *Angew. Chem., Int. Ed.*, 2007, **46**, 445–448; (l) A. Ghosh, *Chem. Rev.*, 2017, **117**, 3798–3881; (m) R. Orłowski, D. Gryko and D. T. Gryko, *Chem. Rev.*, 2017, **117**, 3102–3137; (n) Y. Fang, Z. Ou and K. M. Kadish, *Chem. Rev.*, 2017, **117**, 3377–3419; (o) K. Fujino, Y. Hirata, Y. Kawabe, T. Morimoto, A. Srinivasan, M. Togano, Y. Miseki, A. Kudo and H. Furuta, *Angew. Chem., Int. Ed.*, 2011, **50**, 6855–6859; (p) S. Hiroto, K. Furukawa, H. Shinokubo and A. Osuka, *J. Am. Chem. Soc.*, 2006, **128**, 12380–12381; (q) A. Mahammed, I. Giladi, I. Goldberg and Z. Gross, *Chem.-Eur. J.*, 2001, **7**, 4259–4265; (r) S. Hirabayashi, M. Omote, N. Aratani and A. Osuka, *Bull. Chem. Soc. Jpn.*, 2012, **85**, 558–562; (s) B. Koszarna and D. T. Gryko, *Chem. Commun.*, 2007, 2994–2996; (t) B. Koszarna and D. T. Gryko, *J. Org. Chem.*, 2006, **71**, 3707–3717; (u) H. L. Buckley, W. A. Chomitz, B. Koszarna, M. Tasior, D. T. Gryko, P. J. Brothers and J. Arnold, *Chem. Commun.*, 2012, **48**, 10766–10768; (v) C. F. Zipp, J. P. Michael, M. A. Fernandes and H. M. Marques, *S. Afr. J. Chem.*, 2013, **66**, 158–166; (w) W. Sinha, M. G. Sommer, N. Deibel, F. Ehret, M. Bauer, B. Sarkar and S. Kar, *Angew. Chem., Int. Ed.*, 2015, **54**, 13769–13774; (x) W. Sinha, M. G. Sommer, N. Deibel, F. Ehret, B. Sarkar and S. Kar, *Chem.-Eur. J.*, 2014, **20**, 15920–15932; (y) W. Sinha, M. G. Sommer, L. Hettmanczyk, B. Patra, V. Filippou, B. Sarkar and S. Kar, *Chem.-Eur. J.*, 2017, **23**, 2396–2404; (z) B. Patra, S. Sobottka, W. Sinha, B. Sarkar and S. Kar, *Chem.-Eur. J.*, 2017, **23**, 13858–13863; (aa) A. Garai, S. Sobottka, R. Schepper, W. Sinha, M. Bauer, B. Sarkar and S. Kar, *Chem.-Eur. J.*, 2018, **24**, 12613–12622; (ab) X.-S. Ke, Y. Hong, V. M. Lynch, D. Kim and J. L. Sessler, *J. Am. Chem. Soc.*, 2018, **140**, 7579–7586.

13 J. Ohyama, Y. Hitomi, Y. Higuchi, M. Shinagawa, H. Mukai, M. Kodera, K. Teramura, T. Shishido and T. Tanaka, *Chem. Commun.*, 2008, 6300–6302.

14 M. Kanehara, H. Takahashi and T. Teranishi, *Angew. Chem., Int. Ed.*, 2008, **47**, 307–310.

15 B. C. Mei, K. Susumu, I. L. Medintz and H. Mattossi, *Nat. Protoc.*, 2009, **4**, 412.

16 J. W. Ciszek, M. P. Stewart and J. M. Tour, *J. Am. Chem. Soc.*, 2004, **126**, 13172–13173.

17 (a) M. Stefanelli, F. Mandoj, S. Nardis, M. Raggio, F. Fronczek, G. T. McCandless, K. Smith and R. Paolesse, *Org. Biomol. Chem.*, 2015, **13**, 6611–6618; (b) S. Hiroto, N. Aratani, N. Shibata, Y. Higuchi, T. Sasamori, N. Tokitoh, H. Shinokubo and A. Osuka, *Angew. Chem.*, 2009, **121**, 2424–2426.

18 (a) A. J. Shaikh, F. Rabbani, T. A. Sherazi, Z. Iqbal, S. Mir and S. A. Shahzad, *J. Phys. Chem. A*, 2015, **119**, 1108–1116; (b) A. Ishida and Y. Sakata, *Chem. Commun.*, 1998, 57–58.

19 (a) H. Aldahhak, M. Paszkiewicz, F. Allegretti, D. A. Duncan, S. Tebi, P. S. Deimel, P. Casado Aguilar, Y.-Q. Zhang, A. C. Papageorgiou, R. Koch, J. V. Barth, W. G. Schmidt, S. Müllegger, W. Schöfberger, F. Klappenberger, E. Rauls and U. Gerstmann, *J. Phys. Chem. C*, 2017, **121**, 2192–2200; (b) E. Steene, A. Dey and A. Ghosh, *J. Am. Chem. Soc.*, 2003, **125**, 16300–16309.

20 M. S. Boeckl, A. L. Bramblett, K. D. Hauch, T. Sasaki, B. D. Ratner and J. W. Rogers Jr, *Langmuir*, 2000, **16**, 5644–5653.

