



Cite this: *Green Chem.*, 2020, **22**, 2712

Spherical lignin particles: a review on their sustainability and applications

Monika Österberg,^{ID}*^a Mika H. Sipponen,^{ID}^{a,b} Bruno D. Mattos^{ID}^a and Orlando J. Rojas^{ID}^{a,c}

There is an increased interest in renewable carbon as a source of materials, where lignin is expected to play a prominent role. This stems, partially, from new regulations aiming to achieve a cleaner and safer environment. Lignin, as a polyaromatic plant-derived biomolecule, is not only abundant but widely accessible in industrial streams. Due to recent developments in production scalability as well as promising application prospects, nanoscaled lignin particles have recently generated interest in the research and industrial communities. This review describes the main routes to prepare spherical lignin particles, highlighting aspects associated to their shape and topology as well as performance. We discuss the use of spherical lignin particles as dispersants and in the formulation of coatings, adhesives and composites, focusing on the advantages of the spherical shape and nanoscaled size. The state of the particles is furthermore compared in terms of their applicability in dry and wet forms. Finally, we discuss the sustainability, stability and degradation of lignin particles, which are issues that are critically important for any prospective use.

Received 8th January 2020,
Accepted 3rd April 2020

DOI: 10.1039/d0gc00096e

rsc.li/greenchem

^aAalto University, School of Chemical Engineering, Department of Bioproducts and Biosystems, P.O. Box 16300, Aalto, Finland. E-mail: monika.osterberg@aalto.fi

^bDepartment of Materials and Environmental Chemistry, Stockholm University, SE-106 91 Stockholm, Sweden

^cDepartments of Chemical and Biological Engineering, Chemistry and Wood Science, University of British Columbia, 2360 East Mall, Vancouver, BC, Canada

Introduction

Lignin is a polyphenolic biomolecule found in woody parts of trees and other vascular plants. The monomeric lignin precursors, corresponding structural units and the most common covalent linkages in polymeric lignin are shown in Fig. 1.



Monika Österberg

Monika Österberg is Associate Professor in bioproducts chemistry at the Department of Bioproducts and Biosystems at the School of Chemical Engineering, Aalto University. She received her PhD in surface chemistry in 2000 from the Royal Institute of Technology (KTH), Sweden. In 2012 she joined the faculty of Aalto University and was tenured in 2016. The aim of her research is to enhance the sustainable use of natural

resources. Her research interests are fundamental interfacial phenomena of forest biomaterials, like lignin, cellulose and hemicelluloses, and the development of new materials from these polymers. Lignin nanoparticles being a focus area since 2014.



Mika H. Sipponen

Mika H. Sipponen received his D.Sc. (Tech.) degree in the field of chemical technology from Aalto University, Finland in 2015 (Prof. Simo Laakso). He has worked on lignin-related topics since 2008. As Postdoctoral researcher, he worked during 2016–2019 on lignin-based functional materials with Prof. Monika Österberg and made research stays in University of Rome Tor Vergata, Italy (Prof. Claudia Crestini) and

Shinshu University, Japan (Prof. Yoshihiko Amano). He is presently an Assistant Professor in Materials chemistry in MMK at Stockholm University. His Sustainable Materials Chemistry (SUSMATCHEM) group develops novel lignin-based processes and materials following the principles of green chemistry.



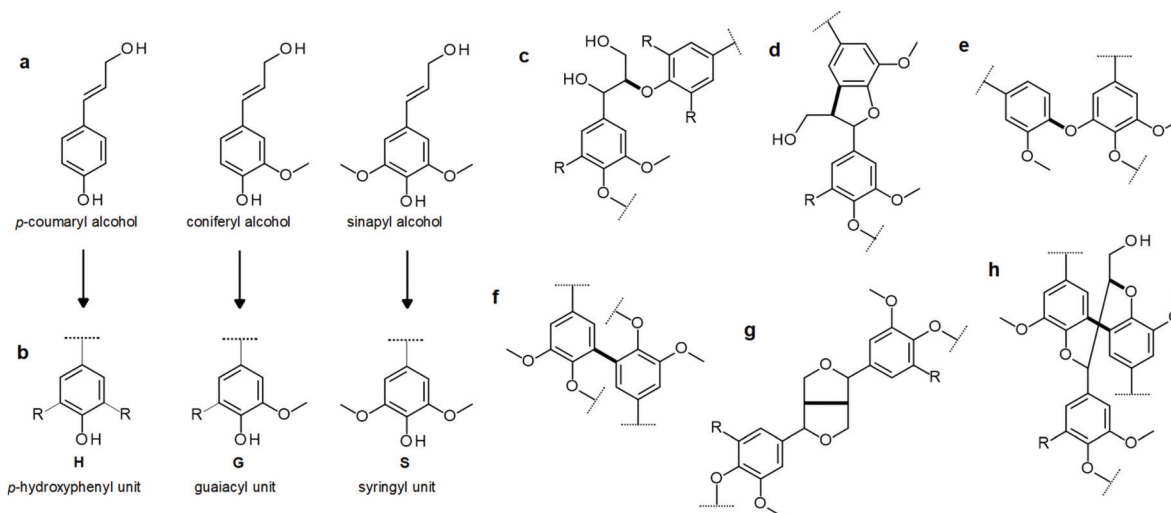


Fig. 1 (a) Monomeric lignin precursors (monolignols) and (b) the corresponding general structural units in lignin. The most common covalent linkages depicted in G-type lignin: (c) β -aryl ether (β -O-4') (d) phenylcoumaran (β -5' (α -O-4')), (e) biphenyl ether (5-O-4'), (f) biphenyl (5-5'), (g) resinol (β - β' (γ -O-4')), (h) dibenzodioxocin (5'-5''/ β -O-4' (α -O-4'')).

During both pulping and biofuel production, lignin is removed as an unwanted side stream. The efficient valorization of lignin has been hampered by its complex and varying chemical structure partly arising from chemical reactions during these pulping^{1–3} and biomass pretreatment processes.⁴ In effect, cleavage of the inter-unit linkages of native lignin (Fig. 1c–h) and associated formation of new covalent bonds leads to technical lignins with complex and incompletely elucidated structures. Lignin nanoparticles (LNPs) or colloidal lignin particles (CLPs), as they are also referred to, have attracted recent increased attention. Some reports even envision that lignin nanoparticles will have a strong influence on promoting lignin valorization, akin to the development of syn-

thetic nanoparticles in the polymer industry.⁵ Indeed, as will be discussed in this review, the formation of spherical, well-defined particles removes many of the obstacles that have hindered efficient and broad utilization of lignin.

There are two main drivers in the application of lignin particles. One is the need to valorize lignin as an industrial stream. The second one is to replace synthetic particles with a biobased alternative. Some of the possibilities for lignin particles involve large volume applications, like adhesives, composites and emulsion stabilizers, *i.e.* applications already proven for other forms of lignin, but where spherical particles could give an advantage. Focusing on the size and shape of the particles introduce totally novel lignin applications, including



Bruno D. Mattos

Aalto University, Finland (since 2018). Dr Mattos' research is devoted to wood nanotechnologies, nano-enabled agriculture, and multifunctional materials from biocolloids.

Bruno D. Mattos is a Wood Engineer (2012; UFPel, Brazil) by training and holds a Master (2014; UFPel, Brazil) and PhD (2018; UFPR, Brazil) in Materials Science and Engineering. Dr Mattos has been with the University of Basque Country, Spain (2013) and Embrapa Florestas, Brazil (2014–2018). Currently, he holds a postdoctoral researcher position in the Department of Bioproducts and Biosystems at



Orlando J. Rojas

an European Research Commission Advanced Grant (ERC-Advanced). He has published over 380 peer-reviewed papers related to the core research of his group, "Bio-based Colloids and Materials", which mainly deals with nanostructures from renewable materials and their utilization in multiphase systems.

Professor Orlando Rojas is a Canada Excellence Research Chair and Director of the Bioproducts Institute in UBC. He is the recipient of the 2018 Anselme Payen Award, one of the highest recognitions in the area of cellulose and renewable materials. He is fellow of the American Chemical Society, the Finnish Academy of Science and Letters and received the 2015 Tappi Nanotechnology Award. His most recent project includes



drug delivery^{6–11} and enzyme immobilization^{12,13} for biocatalysis.

Usually, the approach to use LNPs is described as sustainable or “green” solely based on the fact that a synthetic material can be replaced by a renewable alternative. However, in this review we intend to address the sustainability and greenness of LNP production and applications in more depth from the viewpoint of the most central ones of the 12 principles of green chemistry¹⁴ and by assessing some environmental prospects. Waste prevention is without doubt one of the most important of the green chemistry principles. Lignin valorization can be considered “green” as it utilizes a renewable feedstock that is currently available in excess, hence reducing the amount of waste. Furthermore, isolating lignin for applications instead of burning it for energy, during the recovery of chemicals in the kraft pulping process, would contribute to carbon fixation in materials, and conform to the principles of the circular economy. One of the breakthroughs in the production of LNPs was the finding that no chemical derivatization of lignin is required, provided that there is a suitable organic solvent system. The solvent selection is a compelling engineering challenge that encompasses for instance safety considerations and economical recycling process design. Nevertheless, one needs to be critical as to when the energy input and solvents used for the production of LNPs is compensated by the added value of the products. Other issues that are not always addressed are (1) to what extent the processes are preventing the formation of waste, especially wastewater, (2) what is the atom efficiency of the process, (3) are all materials incorporated in the product, and (4) to what extent could LNPs be designed for degradation?

Lignin particles have in general been found non-toxic (in reasonable concentrations) by *in vitro*^{6,11,15,16} or *in vivo*¹¹ toxicological assessment, but this has to be re-assessed every time the particles are modified or compounded with other materials.¹⁷ In contrast, the eco-toxicity and degradability of the particles have received no attention unlike that of non-particulate lignins.¹⁸ Chemical products should be designed so they do not pollute the environment, hence, when their function is complete, they should break down to non-harmful products. In this scenario, a more comprehensive discussion on the environmental fate, in terms of stability and degradability, of lignin and lignin particles is required to amplify successful applications of such particles as greener solutions for current challenges or to improve the sustainability of already consolidated processes or products.

In this review, we describe the importance of the morphology of particles in general, describe the main methods for producing spherical lignin particles and review the applications in which they have been used. The sustainability of lignin particles, their production and uses are briefly assessed with respect to carbon balance and life cycle assessment. Additionally, we discuss the fate and stability of lignin particles during storage and application, comparing their degradation rates and destabilization mechanisms with those from crude lignins. We will close with our perspectives for future

research needed to advance fundamental understanding and usability of these low-cost particulate materials in applications that support implementation of sustainable circular bio-economy.

Importance of particle morphology on performance

Within materials sciences, particles with controlled morphological features allow gaining control on properties that are otherwise difficult to achieve with irregular, heterogeneous geometries. Relevant to particulate systems, properties such as packing density, color, strength and transport (mass, heat) can be cited. Improved or new functions can be achieved from multiple components integrated in the form of particles.¹⁹ Typically, nano- and micro-particle isolation is less energy demanding compared to the workup of chemical synthesis. This is a direct result of surface energy, especially if the given production method involves high reaction enthalpies (*e.g.* dissolution, covalent bond formation, reduction, *etc.*).¹⁹ Thus, small, chemically engineerable particles can be used prominently, for example as pigments for coatings, as polymer fillers, or for reinforcement, in surface finishing or as substrate in reaction media.

Two morphological considerations are implicit in this review, namely, shape and surface topology. Contrary to the irregular crude lignin that is typically available from a variety of sources, as a dried powder of particles with irregular size and shape, we here focus on recent developments to produce regular spherical lignin particles with uniform size and bearing smooth surfaces. This is critically important since shape regularity is shown to be a determining factor in colloidal behavior, for example, from the point of view of flow, agglomeration and packing.²⁰ Moreover, the effect of shape and its relation with functions have been a standing issue that is recognized for its importance in physics, materials science and other fields, including biology.²¹ Nevertheless, related aspects are far from being fully understood. For example, the relationship between shape and function, especially at the nanoscale, still requires attention.

From the engineering perspective, an advantage of spherical particles over other shapes is the fact that there are no sharp edges, which may break or wear off during the processing. This is particularly true in operations where, for example, finely divided catalyst particles are employed.²² On the other hand, size and size regularity matter for the efficient ordering of objects, which can be in closely packed states, where necks and pores exist. Thus, a two-component system can be assumed for any prediction, *e.g.* the solid and void phases, allowing theoretical and experimental approximations. For monodisperse perfect spheres, the highest packing density comprises approximately 74% of the volume. A random packing of equal spheres, however, generally has a density around 64%. Numerical simulation results show that the upper bound order of random packing densities of basic



monodisperse 3D objects is cube (0.78) > ellipsoid (0.74) > cylinder (0.72) > spherocylinder (0.69) > tetrahedron (0.68) > cone (0.67) > sphere (0.64).²⁰

Similarly as the morphologies of molecules affect the binding forces that hold them together and their ability to form crystals, also the forces between particles are affected by their size and shape, which again will affect the properties of particle assemblies. Another aspect, especially relevant to those working in the area of colloids and surfaces, is the recognized need to control interfacial interactions, which is well described for spherical particles.²³ However, this aspect becomes more complex in the presence of rough surfaces.^{24–26} Roughness features create a distribution of interaction energies, where local inter-particle interaction potentials may be shifted from those predicted under the assumption of smooth surfaces. In general, the double-layer potential is, on average, reduced by surface roughness because the interfacial separation is effectively larger. Hence, particles with rough surfaces are more prone to aggregate due to attractive van der Waals interactions and the particles may be in a primary minimum preventing dispersion upon decrease in ionic strength. However, if colloids are attached on the asperities of rough surfaces, both theoretical and experimental results show that they may be released upon lowering of the ionic strength, demonstrating the importance of surface heterogeneity on colloidal stability.²⁷ Likewise, the idea of matching the shapes of colloid particles to maximize the strength of depletion interactions stems from considerations of entropy effects. For example, small particles or polymers can be excluded from the spaces between larger colloid particles. This exclusion increases the osmotic pressure

outside—and so provides an effective attraction between—the larger particles.

The effect of shape of particles is best illustrated if one considers it as a parameter affecting the toxicity *in vitro* and *in vivo*.^{28–30} High aspect ratio particles can more easily penetrate a biological cell, increasing both cyto- and eco-toxicity. Hence, spherical particles are in general considered to have the lowest toxicity level. Relevant to this review is to highlight the advantages inherent to spherical particles: they facilitate the most efficient utilization if used as a substrate for reactions or as a catalyst support. A practical challenge is therefore to afford particles with a smallest suitable diameter. In operations wherein the particles are employed in a fixed bed, spherical shapes provide a means of obtaining uniform packing, thus preventing a variation in pressure drop through the column. If variations in pressure drop occur, channeling results and a large portion of the bed is by-passed, thereby decreasing the effectiveness of the system. These aspects have been relevant in some of the recent reports on the use of lignin particles in multifunctional membranes, anti-oxidative microfiltration, patterning and 3D structuring.³¹ The following section describes the processes most typically used to produce spherical lignin particles.

Processes to produce spherical lignin particles

There is a multitude of methods reported on preparation of lignin particles and the main ones are summarized in Table 1. Unfortunately, the nomenclature can be confusing. Just as the

Table 1 Summary of the main lignin nanoparticle production methods and the demonstrated applications

Method	Lignin type	Solvent	Antisolvent	Dry/wet	Spherical particles	Demonstration of application	Ref.
Solvent shifting	AL, KL, others	THF	H ₂ O	Wet	Yes	Drug delivery and antiproliferation of cancer cells, UV blocking	7, 8, 35, 42 and 43
	KL	THF/H ₂ O	H ₂ O	Wet/dry	Yes	Glue, composite with PLA, Pickering emulsion	37 and 44
	KL	Acetone/H ₂ O	H ₂ O	Wet	Yes	Pickering emulsion, enzyme immobilization, drug delivery, composite films with functionality (anti UV, antioxidant)	12 and 45–47
Acidification	AL	Ethanol	H ₂ O	Wet	Yes	Drug delivery	9
	KL	EG	HCl	Wet	—	Drug delivery	48
	KL	NaOH (aq)	HNO ₃	Wet	—		48
	KL	EG	HNO ₃			Antimicrobial (silver infused particles)	49 and 50
	AL	EG	HCl	Wet		Composites, adhesives	51 and 52
Reverse micelle formation		Dioxane	Cyclohexane	Wet	Yes	Composites	53
Aerosol flow	KL/HTT, OSL	DMF, acetone	None	Dry	Yes	Pickering emulsions, particulate membranes and anti-oxidative microfiltration	38, 40, 54 and 55
Aerosol + ice segregation		DMSO	H ₂ O	Dry	Yes	Coatings	56
Mechanical treatment	AL	H ₂ O		Dry	—	—	57
	KL	H ₂ O		Dry	—	Polymer blends	58
CO ₂ precipitation	KL	DMF	CO ₂	Wet	—	UV absorption	59

THF, tetrahydrofuran, DMF = *N,N*-dimethylformamide, EG = ethyleneglycol, KL = kraft lignin, AL = alkalilignin, OSL = organosolv lignin.



produced particles are called lignin nanoparticles, colloidal lignin particles, lignin nanospheres *etc.*, the same method has been described differently in the literature. One reason for this confusion is that there is no consensus about the mechanism behind the particle formation. In this review we attempt to clarify discrepancies in nomenclature and instead of using the specific nomenclatures adopted so far, we refer to the given work depending on the underlying particle formation method. As described above, compared to amorphous and irregular powders, spherical lignin particles can often add advantages in applications. Whereas irregular lignin powders may be sufficient for certain uses *e.g.* as fillers,^{32–34} a better control over the particle morphology is preferable for high-performance applications, such as in Pickering stabilization. As a consequence, we will focus on methods leading to spherical particles but also correlate to results obtained using as-produced, irregular powders. The methods to prepare spherical lignin particles can roughly be divided into dry and wet methods (Fig. 2). These are reviewed in the following section.

Dry particles

Dry lignin particles are formed simply from a well-controlled drying of dilute lignin solutions. Uncontrolled drying, *e.g.* oven-drying of a lignin solution, leads to the formation of aggregated, irregular, and heterogeneous micron-sized

powder. A few strategies have been presented for the preparation of dry lignin particles with a variety of controlled surface topologies and particle size. Briefly, such methods involve the controlled evaporation of atomized droplets of lignin solution,^{36,38} or the post-drying of dispersed particles formed by emulsion or solvent exchange techniques.^{37,39} The latter are described in more detail in the next section related to wet particles.

Ago *et al.*³⁸ introduced the use of an aerosol flow reactor for the production of dried lignin particles, in a wide range of size fractions and from different sources. They showed that spherical particles could be produced from a broad variety of lignin types, like alkali, kraft or organosolv lignins, provided that a suitable solvent is used. Unless the lignin stream is already sourced as a solution, the process involves dissolution followed by atomization of the solution into droplets, which are suspended and carried by a carrier gas (for example, air or nitrogen) through a heated laminar flow system.

Common solvents include water, alcohols, acetone and DMF, depending on the lignin type, thus allowing the synthesis to occur, in a temperature range that also depends on the solvent, from 25 to 150 °C.⁴⁰ After formation, the particles are cooled in an air stream and subsequently fractionated in a Berner-type low-pressure impactor with multiple collection stages. Fractions with narrow particle distribution from 230 nm to 1.9 μm can be directly collected. The lignin type affected both particle charge and wettability, with reported contact angles of water ranging from 57° for particles formed from kraft lignin to 69° for those from organosolv lignin (Fig. 3). The cost to produce dried lignin particles can be

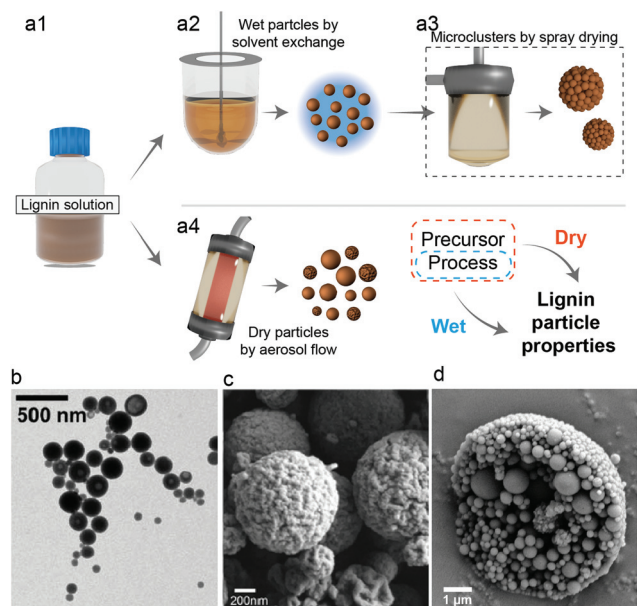


Fig. 2 The most common processes involved in the formation of spherical lignin particles. (a1) Lignin solutions can be converted into spherical particles by (a2) controlled solvent exchange resulting in aqueous colloidal lignin particles dispersions that can be used as such or, if needed, be further spray dried to obtain (a3) lignin particle micro-clusters. (a4) Lignin solutions can be atomized into micro/nano scaled droplets and dried under, fluidized, controlled atmospheres. These processes yield (b) spherical smooth³⁵ (c) wrinkled³⁶ or (d) superstructured³⁷ particles. Reproduced with permission from ref. 34, 35 and 36. Copyright © 2016 and 2018, Royal Society of Chemistry.

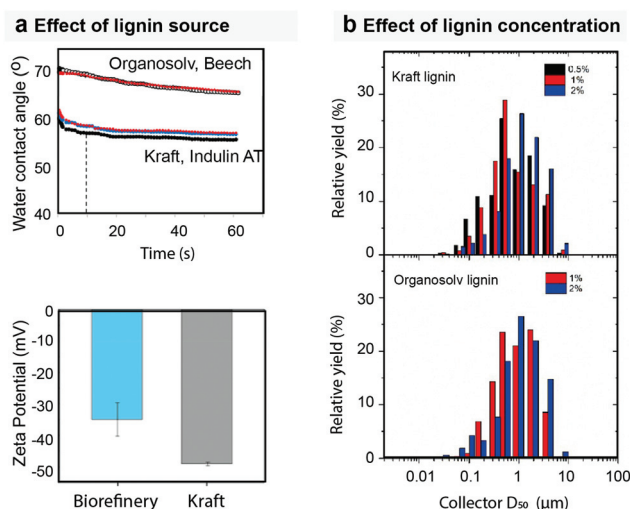


Fig. 3 Factors affecting the formation of dry lignin particles. Process parameters (solvent, reactor conditions, flow, etc.) and source (botanical and extraction process) affect the final properties of the lignin particles. (a) The source of lignin typically affects the surface energy³⁸ and charge⁴⁰ of the obtained particles, whereas (b) the concentration of the starting solution affects the size and size distribution over a wide range of sizes.³⁸ Reproduced with permission from ref. 37 and 41. Copyright © 2016 and 2020, American Chemical Society.



reduced by the utilization of selected solvents, *i.e.* acetone, that favor the aerosol formation and droplet consolidation under conditions of low temperature and short time. This would make such process scalable and cost-effective, with an estimated selling price of 1015 US\$ per metric ton.⁴¹ Recently, acetone-extracted lignin from hydrothermally treated wood was converted into lignin nanoparticles *via* the aerosol flow reactor. The authors experimentally demonstrated the possibility of achieving lower time and temperature for particle formation. Additionally to that, the surface charge of the particles from biorefinery lignin differed from those obtained from kraft lignin (Fig. 3a).⁴⁰

A variation of the aerosol flow process was discussed by Kämäräinen *et al.*³⁶ for the preparation of wrinkled kraft lignin particles. From a different selection of solvents and with the use of a blowing agent, it was possible to engineer the drying phase of the atomized droplets to induce drying stresses exceeding the critical buckling stresses of the partially dried lignin droplets. Ammonium carbonate was used as blowing agent, and ammonium hydroxide as solvent. An early-formed rigid shell around the droplets buckles toward the concave side, thus leading to particles with high roughness. Smooth particles were obtained without the use of the blowing agent, which indicate the need of a blowing agent for the formation of wrinkled lignin particles with a higher surface area.

Dried particles can also be obtained by freeze-drying³⁹ or spray-drying³⁷ lignin particle dispersions previously prepared by either emulsion templating or solvent exchange. Sameni *et al.*³⁹ prepared non-aggregated dried lignin microspheres after freeze-drying a diluted suspension, at 0.2 mg mL⁻¹. Although such process was efficient in obtaining dried particles, freeze-drying is a time-demanding and relatively expensive process, causing freezing gradients, and thus hindering the large-scale production of dried particles and therefore their wide application.

On the other hand, Lintinen *et al.*³⁷ demonstrated an efficient and scalable method to obtain dried particles from colloidal lignin suspensions. After the formation of the lignin particles by solvent exchange, a regular spray-drier can be used to process the concentrated aqueous LNP dispersion. The process led to the formation of superstructured particle microclusters (from 5 to 80 μm), which are partially redispersible or may as well be used in their higher hierarchical architecture in *e.g.* delivery platforms where nanoparticles may be too mobile.⁶⁰ Such microclusters can be redispersed by either tip or bath ultrasound or high-shear homogenization (Ultra-Turrax). Dry lignin nanoparticles have also been formed by evaporating solvents from a reverse micelle dispersion of lignin,⁵³ and by using a so-called ice segregation method, which involves freezing lignin solutions on cold surfaces.^{56,61}

Wet particles

Various precipitation methods exist for the fabrication of nano- and microscaled lignin particles,^{9,10,35,42,48,57,62–66} but not all of them produce spherical lignin particles in colloidally stable aqueous dispersions. The process of dissolving lignin in

(aqueous) organic solvent and precipitating in a non-solvent (typically water) is commonly referred to as solvent shifting, solvent exchange or nanoprecipitation. Regardless of the precipitation scheme, lignin has an intrinsic tendency for assembly into spherical particles that minimize the surface area in contact with the non-solvent phase. However, there are some prerequisites to obtaining stable dispersions of individual particles. For example, although acidification of an alkaline lignin solution leads to precipitation, the precipitation continues to an irregular network structure and inevitable sedimentation, since acidification protonates the charged groups on lignin, preventing electrostatic stabilization of the particles. It is important to note that this differs from the nanoprecipitation methods for preparation of LNP dispersions that can remain colloidally stable for months.

The morphology and colloidal stability of the lignin particles also depend on the type of lignin raw material, properties of solvent, and process parameters, such as the rate of addition of the non-solvent, or if the lignin solution is added to the water, the end-point solvent concentration, and pH. The molecules associate *via* non-covalent interactions including hydrogen bonding, and pi-stacking as well as hydrophobic interactions and others. Some papers discuss pi-pi interactions as the main driving force for the aggregation process that leads to particle formation.^{35,67} Combining molecular dynamics simulations and atomic force microscopy it was recently shown that the hydrophilic groups of enzymatic hydrolysis lignin interact with water and the hydrophobic skeleton with the organic solvent during lignin dissolution in organic solvent-water binary mixtures. When the ratio of water to organic solvent becomes high enough, lignin self-assembles into spherical particles. Interestingly, the authors found that spherical particles were also formed when the water concentration in acetone was decreased below 0.2%.⁶⁸

Except for this study, we can only speculate on the impacts of different structural motifs on particle morphology. One viewpoint is the morphology that lignin molecules adopt when precipitated from solution. Motifs with different bonding segments such as β -aryl ether *versus* biphenyl ether probably behave differently because of the different distances between the interconnected aromatic units.

Another point of interest is the molecular weight, which has been found to determine the pattern of formation of spherical lignin particles.⁹ In general, it appears that precipitation of molecules with large molecular weight (and low water-solubility) initiate the process and act as nuclei for the growing particles. The last lignin molecules to precipitate and coat the particle surfaces are the low molecular weight fragments that are often enriched with hydrophilic functionalities such as carboxylic acid or sulfonic acid groups. Increasing the molecular weight by enzymatic crosslinking of the lignin molecules has been shown to lead to smaller particle size.⁴⁴ In another study, ethanol and sulfuric acid were used for lignin dissolution and the authors found that this enabled cleavage of β -O-4 and β - β linkages resulting in condensed lignin with higher molecular weight.⁶⁹ In line with Mattinen *et al.*⁴⁴ they



also found that a smaller particle size was obtained. This phenomenon was explained to be due to increased hydrophobic interactions. It is presently unclear whether differences in molecular weight distribution and structural motifs of lignin influence particle properties such as colloidal stability or porosity that was recently determined for the first time.⁷⁰ However, many of the above points would benefit from more detailed experimental evidence, and ideally be supported by molecular dynamics simulations.

The effect of pH on the formation of lignin particles has received surprisingly little attention despite the fact that many technical lignins contain carboxylic acid groups in addition to the acidic phenolic hydroxyl groups. Leskinen *et al.* adjusted the pH of the water used as non-solvent in the preparation of colloidal lignin particles from aqueous THF solution of softwood kraft lignin. They found that the particle diameter decreases almost linearly when pH of the resulting lignin particle dispersion increases from 3 to 6.⁷¹ A similar trend was confirmed to take place when the pH of aqueous acetone solution of softwood kraft lignin was adjusted prior to the nanoprecipitation step with deionized water. This trend is expected, since deprotonation of the carboxylic groups leads to increased surface charge and electrostatic stabilization of the particles (Fig. 4).

Another important aspect to consider when comparing different solvent systems is the concentration of lignin before precipitation and also the concentration of lignin particles in the final dispersion (Fig. 4b). Aggregation of particles or formation of interlinked particle networks occur if the concentration of lignin is too high. For instance, the concentration threshold for wheat straw soda lignin prior to the precipitation step was 6 g L⁻¹ when using aqueous ethanol as solvent,⁹ while typically concentrations exceeding 40 g L⁻¹ still enable the formation of freely dispersed particles from softwood kraft lignin in aqueous THF solvent system.^{47,71} Addition of ethanol as a co-solvent in aqueous THF has been shown to increase the applicable concentration range, giving particles with hydrodynamic diameter below 300 nm in final dispersion concentrations of up to 3 wt%.³⁷

The wet particles are typically formed in aqueous medium, have a hydrophilic surface, and are electrostatically stabilized by the weak carboxylic groups present in the lignin. Thus, they are sensitive to the electrolyte concentration and pH of the aqueous media. The surface charge furthermore enables easy surface modification by adsorption of oppositely charged polymers or proteins to the particles. The carboxylic groups additionally offer sites for covalent modification of the surface chemistry. These various surface modifications will be discussed more with respect to applications in the following sections. However, one aspect that deserves more studies is the intra-particle porosity of the wet particles.⁷⁰ Thus far, the particles have been used for entrapment and adsorption of many different active substances, as reviewed,¹⁷ and their release properties have provided indirect information of the packing and porosity of the particles.^{9,10,72,73} In general, the accumulated results indicate that LNPs from the solvent-exchange

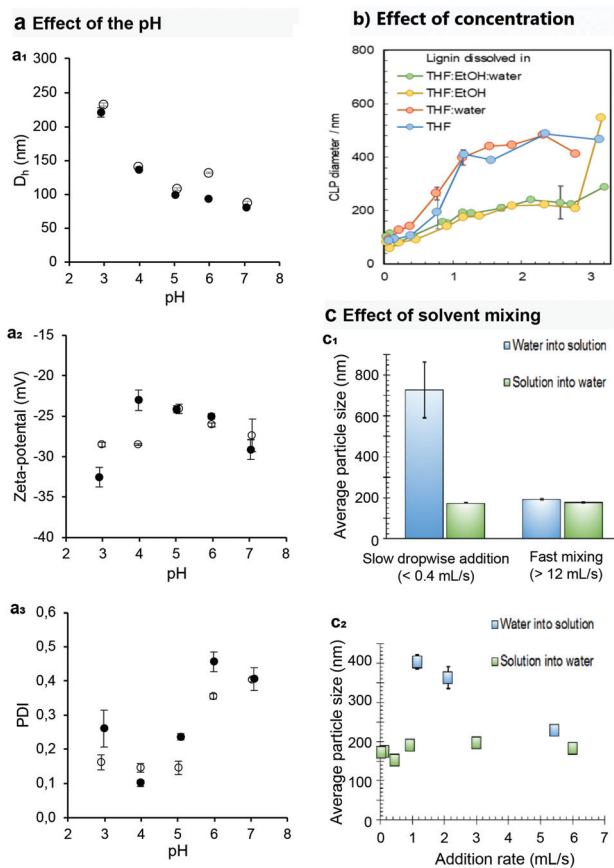


Fig. 4 Typically the process parameters affect the formation and properties of the wet lignin particles. (a) pH of the final solution has a clear effect on (a1) the size, (a2) surface charge and (a3) dispersivity of the particle population. (b) The lignin concentration and solvent composition during the solvent exchange have a decisive effect on the particle size.³⁷ (c) The (c1) order and (c2) rate of solvent mixing during the formation process affect the particle size, being tools for engineering the size of the colloids.⁷¹ The order (solvent into water or water into solvent) has a more pronounced effect at lower addition rates. Panels in "a" are unpublished results from the authors using softwood kraft lignin dissolved in acetone water (3:1 in wt%) mixture, adjusted to target pH using 1 M hydrochloric acid or 1 M sodium hydroxide, and precipitated by rapidly adding deionized water into the vigorously stirred lignin solution. The final pH of the dispersion (CLP concentration 0.2 wt%) was measured and reported. Reproduced with permission from ref. 36 and 67. Copyright © 2018, Royal Society of Chemistry; and Copyright © 2016, Walter de Gruyter GmbH.

method are penetrable to water, which explains a rather rapid release of water-soluble small molecules,^{9,10} while bulkier compounds are released at lower rates.^{72,73}

Applications

In the following section the most common applications that have been reported for spherical lignin particles are reviewed and whenever possible, the achieved properties and functionalities are compared to the results obtained using crude, irregular lignin.



Dispersants

The most obvious application for spherical lignin particles and the one that was first reported is the use of the particles as dispersants. Dispersants are very practical engineering additives that play a key role across a wide range of economically important industrial segments. "Dispersant" is a term that can be generically used to describe surfactants, plasticizers or emulsifiers, depending on the field of application. Healthcare, food, civil construction and agriculture greatly benefit from dispersants that enable the mixing of immiscible liquid phases, and enhance the stability of particle suspensions. Lowering the interfacial tension between immiscible liquids, as well as increasing the repulsive forces between suspended particles are physical phenomena expected from dispersants. Such dispersive forces prevent settling and aggregation of phases, thus improving technical properties of the multiphase system such as rheology, lifetime and function. Amphiphilic molecules are the most common industrial dispersants; however, more recently, solid nanoparticles have been used to stabilize immiscible liquid phases by the formation of so-called Pickering-emulsions.⁷⁴ Pickering emulsions are in general more stable against coalescence than emulsions stabilized using soluble surfactants. The majority of the industrially used dispersants are synthetic, raising concerns over their shortcomings in sustainability. There is a dual drawback in their use, namely their synthesis from non-renewable precursors and the lack of biodegradability. Therefore, there has been a wave of efforts toward the development of biobased, or more sustainable dispersants.

Biomolecules, especially lignin, have a powerful dispersive capacity. Within its complex molecular structure, lignin possesses both hydrophobic and hydrophilic moieties that results in an amphiphilic character that can be engineered based, mostly, on the plant source and extraction process. Lignosulfonates (LS) are successful examples of lignin derivatives used as dispersants mostly in cement,⁷⁵ but also in other minerals,⁷⁶ coal,⁷⁷ pesticides and dyes.⁷⁸ The addition of low loadings of LS in cement mixtures greatly improves the rheological properties of the system, thus allowing the utilization of more concentrated dispersions with less water in their preparation. The major drawback of LS is that it is obtained as a side product from the sulfite pulping process that accounts only for *ca.* 2% of the total worldwide production of chemical pulp. Hence, there has been many attempts to use readily available kraft lignin, which accounts for 90% of the production capacity of technical lignins,⁷⁹ as dispersants. However, in contrast to lignosulfonates, kraft lignin has a poor solubility in neutral water and several chemical modification efforts have been presented to increase the water solubility of kraft lignin. Some of the recent examples being sulfomethylation,⁸⁰ sulfonation,⁸¹ carboxymethylation⁸² and a few other such as oxidations.⁸³ Applying chemical routes to derivatize lignin, however, stands against the principles of green chemistry, generating chemical waste, sometimes using high-energy intensive processes, and decreasing the biodegradability of the final materials.

Spherical lignin particles are envisioned to overcome current challenges in the use of lignin derivatives from non-sulfite pulping process. Lignin from virtually any source can be assembled into particles, as discussed earlier, either by controlled solvent shifting or atomization. Up to date, from the best of our knowledge, there is no report on the utilization of spherical lignin particles as dispersants for cement or other mineral compositions. One reason for this may be their relatively higher production cost when compared to the direct utilization of lignosulfonates, or the fact that lignin particles have not yet been available in large quantities. However, recently, there has been significant advances in process engineering that is leading to large scale production of lignin particles at lower cost,^{41,71} which will open opportunities for the utilization of technical lignin-based particles in large scale applications. A great advantage of lignin particles over their macromolecular counterpart is that they are readily dispersible in water, thus not being limited by solubility. Lignin particles, if not modified, possess negative surface charge (ζ potential *ca.* -30 mV depending on pH, lignin source and particle preparation method). Cationic lignin particles (ζ potential *ca.* +30 mV) can be easily prepared by adsorption of synthetic (PDADMAC)^{35,37} or natural polycations (chitosan)⁴⁵ or cationized lignin.⁴⁷ Such versatility enables the use of spherical lignin particles as dispersants for mineral particles with either negative or positive charges. It has been noted that the size of the macromolecular lignin dispersant (discussed as molecular weight) has a strong effect on its dispersive action, in which bigger molecules are more efficient due to steric effects. We envision that the charged lignin nanoparticles could lead to intensified dispersive forces driven by a combination of electrostatic and steric repulsion. Even if considered big in relation to their molecular counterpart, lignin nanoparticles are at least four orders of magnitude smaller than the mineral particles that are commonly used (*e.g.* cement), thus avoiding aggregation and particle clustering resulting from their adsorption. This may warrant their increased and efficient utilization as dispersants.

Whereas spherical lignin particles have not yet been applied as a dispersant for mineral suspensions, there has been many demonstrations of their applicability as Pickering emulsifiers to stabilize oil-in-water (O/W) emulsions.⁸⁴ Such multiphase systems have potential for applications in many technical fields, but mostly in cosmetics, paints, food, and agriculture. Pickering emulsions usually display longer lifetime when compared to surfactant-based emulsions, which is due to the higher resistance against droplet coalescence induced by the particles adsorbed at the O/W interface.^{85,86} Indeed, by tuning the amphiphilicity and charge of the lignin nanoparticles a large variety of O/W emulsions can be stabilized. While O/W emulsions based on aliphatic or aromatic hydrocarbons have been easy to stabilize using LNPs,^{38,47,87-90} vegetable oils have been more challenging, even if using chemically modified lignin such as the graft copolymer of lignin and *N*-isopropylacrylamide as a source of nanoparticles.⁹¹ Nevertheless, food, cosmetics and biomedical



materials require non-toxic oils and emulsifiers. Recently, Zou *et al.* showed that vegetable oil–water emulsions could be stabilized using LNPs coated with chitosan.⁴⁵ The chitosan-coated LNPs were further cross-linked with sodium triphosphate forming emulsion capsules that resisted drying and re-wetting.⁴⁵ In Fig. 5 examples of the use of LNPs in Pickering emulsions are shown.

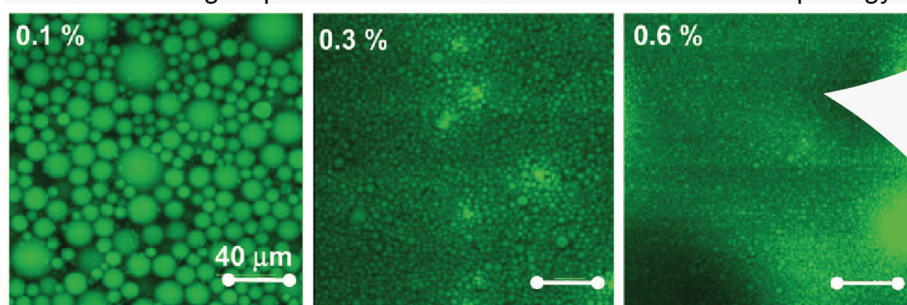
The use of lignin particles to stabilize Pickering emulsions additionally brings, technical and environmental benefits when compared to *e.g.* silica^{93–95} or polystyrene⁹⁶ particles that are currently used in Pickering emulsions. Lignin particles are, if not biodegradable, at least compostable,^{97,98} which is not the case of the common synthetic counterparts. Additionally, lignin possesses high antioxidant and UV-shielding properties.^{46,99,100} Such features enhance the long-term stability

of the emulsified oil by reducing the access of light and air, therefore reducing photodegradation and oxidation of the active ingredient. Lignin-based Pickering emulsions are attractive for outdoor uses, especially for pesticide applications in agriculture where the losses of the efficiency driven by photodegradation and oxidation leads to overuse of pesticides and contamination of non-target organisms.¹⁰¹

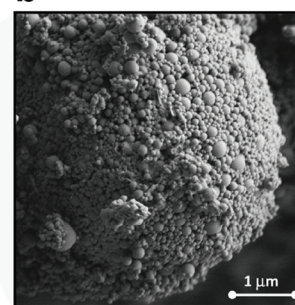
Emulsions and colloids for sunscreens

Due to their UV-light absorbance, finely ground wood powder,¹⁰² various technical lignins,^{103–106} chemically modified lignins,^{107,108} and more recently spherical lignin particles^{43,109–111} have been evaluated in sunscreen formulations. Tan *et al.* showed that LNPs prepared from pine wood organosolv lignin (OL) exhibited better sunscreen performance

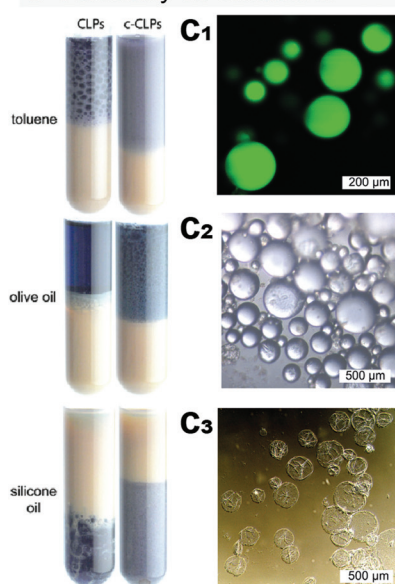
a Effect of the lignin particles concentration on the emulsion morphology



b



c Versatility as emulsifier



d Effect of the particle surface modification

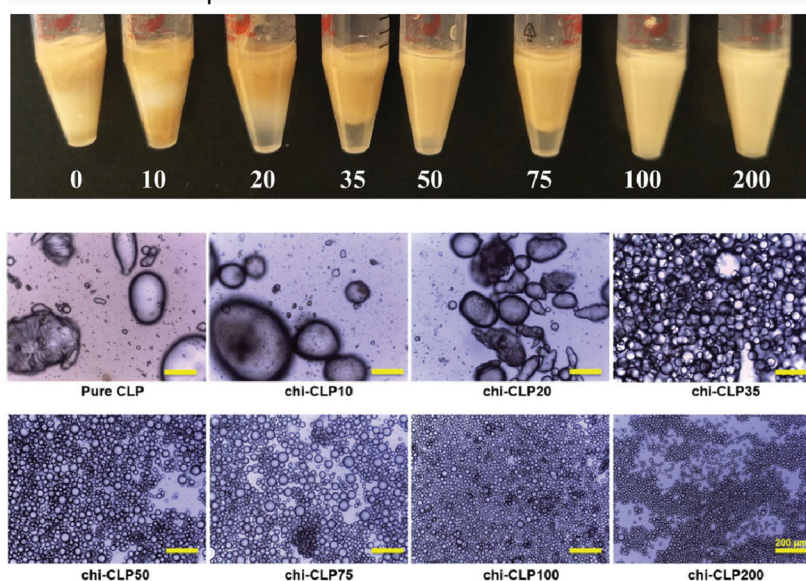


Fig. 5 Use of lignin particles as dispersants for oil toward the formation of Pickering emulsions. (a) The concentration of lignin particles changes the morphology of the emulsion, commonly decreasing the oil droplet size with the addition of more particles.³⁸ (b) Scanning electron microscope image showing lignin particles assembling around the oil droplet at the oil/water interface.³⁸ (c) Lignin particles can be used to effectively disperse a wide range of oils, such as toluene, olive and silicone oil. Confocal (c1) and optical (c2) microscopy images of toluene-in-water emulsions before and after (c3) evaporation of toluene.⁴⁷ (d) Surface chemical modification of the lignin particles can be further used to control their stabilizing action as demonstrated by the small droplet size obtained using chitosan coated LNPs.⁹² Reproduced with permission of ref. 37, 47 and 88. Copyright © 2016, American Chemical Society; Copyright © 2017, Royal Society of Chemistry; and Copyright © 2019, Zou, Sipponen, Österberg.



than the non-spherical OL particles.¹¹¹ Another benefit of using LNPs is that the sunscreen formulations are less brownish than those containing non-spherical lignins. Zhu *et al.* prepared pH-responsive sprayable hydrogels based on chitosan and nanoparticles consisting of lignin reacted with amine and carboxylic acid capped poly(ethylene glycol) (PEG).¹⁰⁷ In this process, peptide formation catalyzed by 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC) is expected to result in a mixture of lignin-grafted PEG and PEG-based peptides. Kai *et al.* synthesized lignin-PEG methacrylate (PEGMA) copolymers *via* atom transfer radical polymerization (ATRP) to enhance the dispersion efficiency of lignin in the commercial creams.¹⁰⁸ The modified lignins had only a minor increment on the SPF (sun protection factor) levels of the moisturizing cream and were less effective than the corresponding lignosulfonate supplements. Blending of the lignin copolymers at 10 wt% concentration in commercial sunscreen formulations increased the SPF of the blend. However, comparative values for lignosulfonate were not shown in the tested sunscreens.

Recent investigations have specifically focused on the development of light-colored lignin-based sunscreens.¹¹² One way to control the whiteness is to use lignin isolated under mild thermochemical conditions.¹¹² Another approach assessed recently is to fractionate lignin based on solubility in organic solvents, and acetylation of the fractions.¹¹³ Compared to these solvent-intensive processes, morphology control through particle size and shape modifications seems more environmentally benign. Zhang *et al.* spray-dried lignin from aqueous dispersion to obtain spherical microparticles that were then formulated in sunscreens.¹⁰⁶ Despite the pursuit of developing white sunscreens, the consumer preference for white *versus* brownish cream may differ. Gutiérrez-Hernández *et al.* suggested that LNPs could be matched to the skin tone of the person using the sunscreen formulation.¹¹⁴

Lignin has been combined with photoactive metal oxide nanoparticles to quench photocatalytic activity of titanium dioxide NPs^{115–117} and to broaden the UV-blocking spectrum of zinc oxide NPs in sunscreen formulations.¹¹⁴ The excellent ability of LNPs to stabilize emulsions is an additional advantage of their use in sunscreens.^{38,45,47,89} A challenge for further work is to develop all-natural sunscreen formulations in which lignin provides multi-functionality such as emulsion stability, sun protection, anti-oxidant, and antimicrobial activity.

Adhesives

Technical adhesives. One of the purposes of lignin in the plant cell wall is to bind the fibrils and fibers together. Hence, the most natural application for lignin is as wood adhesive and lignin has been applied to replace the phenol formaldehyde resins for decades.^{118,119} Phenol formaldehyde resins are known to have excellent moisture resistance, thermal resistance, chemical durability and bonding strength¹²⁰ and are hence still today extensively used in products like plywood, oriented strand board or other engineered wood products. Both the toxicity of formaldehyde and phenol and the fact that they are produced from petroleum have motivated the search

for less toxic, bio-based alternatives like lignin. Nevertheless, these attempts are hampered by the low water resistance of most lignin based adhesives and in most cases the lignin needs to be chemically modified or then only up to 50% of the phenol has been replaced.¹²¹ Kalami *et al.* succeeded in replacing 100% of the phenol in the phenol formaldehyde resin with lignin and demonstrated acceptable wet strength of the bonds in plywood samples¹²⁰ but some excessive purification steps of the lignin were needed and the presence of free formaldehyde could still be an issue in the final product. Recently another approach using epoxy chemistry was reported.¹²² In this study, kraft lignin and glycerol diglycidyl ether were combined to prepare an epoxy resin, and the viability for scale up was assessed. Interestingly, Li *et al.*¹²² tested the effect of lignin particle size on resin properties and found that by decreasing the particle size they were able to increase the lignin content in the formulation. Although not directly focusing on lignin particles, these findings illustrate the advantages of using lignin in the sub-micrometer range.

There are very few reports on the use of LNPs in technical adhesives. Recently Yang *et al.*⁵¹ added 5–10 wt% lignin micro- and nanoparticles to a phenol-formaldehyde resin and found that low concentrations of lignin particles could increase the shear strength of wood lap joints when using the mixtures under dry conditions, but moisture resistance was not studied. However, already 10 wt% of lignin particles decreased the joint strength. They used LNPs prepared by acidification, and the irregularity of the particles may be one reason that higher amounts of lignin could not be used. Unfortunately, there are no systematic studies on the use of well-defined spherical LNPs in technical adhesives. The spherical shape may enable higher lignin concentrations without detrimental effect on viscosity of the resins. Chemical modification, increasing the reactivity of the LNPs, could also enable higher lignin contents without loss in adhesive properties.

Biomedical adhesives. An interesting application of spherical LNPs is to glue soft tissue and hydrogels in biomedical applications. Hydrogels have received attention as ideal materials for repairing soft tissue like skin,^{123,124} cartilage¹²⁵ and muscle,¹²⁶ but most of the developed hydrogels still possess poor adhesion to human tissues.¹²⁷ The curing methods used for technical adhesives, like ultraviolet irradiation or heating are not applicable for human tissues, and more gentle approaches are needed. Rose *et al.* showed that silica nanoparticles can glue together soft gels simply by spreading the NP sol on one of the hydrogels and pressing the gels together.¹²⁸ The polymers from the hydrogels adsorb onto the particle surface and the particles act as connectors between the two hydrogels. The polymer chains reorganize upon adsorption and dissipate energy under stress. Inspired by this, Mattinen *et al.* applied spherical LNPs modified with proteins to glue chamois leather specimens together.⁴⁴ Gan *et al.* used lignosulphonate-stabilized silver nanoparticles to prepare an adhesive and tough hydrogel.¹²⁷ They demonstrated adhesion to a large range of synthetic and biological surfaces in addition to a good cell affinity and antibacterial



activity of the hydrogel. The fact that the NPs not only induce adhesion, but also toughen the hydrogel itself will most probably inspire further work. In Gan *et al.*'s study the bactericidal ability arose from the incorporated silver,¹²⁷ but lignin has also demonstrated antibacterial properties.^{129,130} In future work, focus should be placed on decreasing the use of silver and developing purely natural tissue adhesives.

Coatings and paints

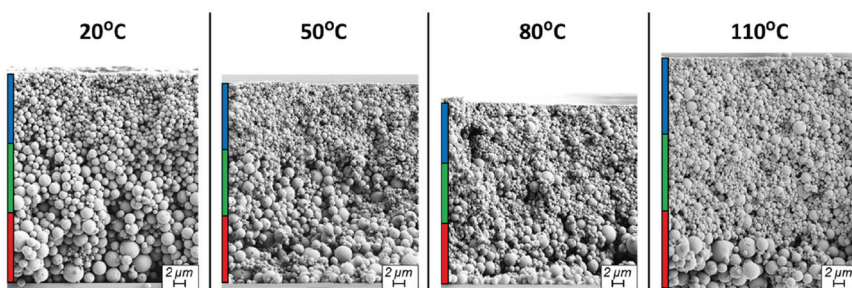
The purpose of applying a coating is to protect the substrate, introduce or enhance functional properties, or to ameliorate decorative or technical properties. According to Mathiazhagan and Joseph, technical requirements for functional coatings include: (i) durability (abrasion, pH, ions and other solutes, temperature, moisture, *etc.*) (ii) reproducibility (iii) easy application and cost-effectiveness (iv) tailored surface morphology and (v) environmental friendliness.¹³¹ Spherical lignin particles seem beneficial regarding many of the above requirements, since lignin can provide surfaces with UV-protection,¹³²

corrosion inhibition,^{133,134} antimicrobial activity,^{129,130} removable/self-healable,¹³⁵ as well as fire-retardant¹³⁶ and drug release properties,¹³⁷ making lignin attractive for functional coating purposes.

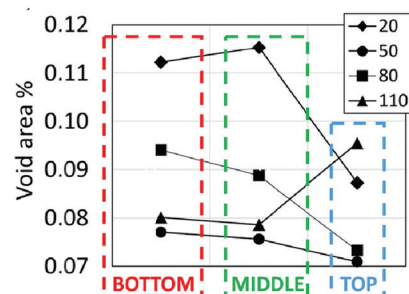
Biocompatible and biodegradable coating materials are gaining importance in response to increasing consumer awareness to sustainable materials that help tackle environmental pollution. Particulate coatings have in general some intrinsic benefits over solution coatings, such as the ability to achieve water-resistant and breathable coating layer that is important in *e.g.* technical textile applications.¹³⁸ The possibility to apply particulate coating from aqueous dispersion of spherical lignin particles cuts off the emission of volatile organic compounds (VOCs), contributing to the waste prevention goal.

There are only a few works involving spherical lignin particles in coatings or films. Yi *et al.* synthesized multilayer composite microcapsules by using Pickering emulsion templates and demonstrated their application in a self-healing coating for steel (Fig. 6b).¹³⁹ Cusola *et al.* experimented and simulated

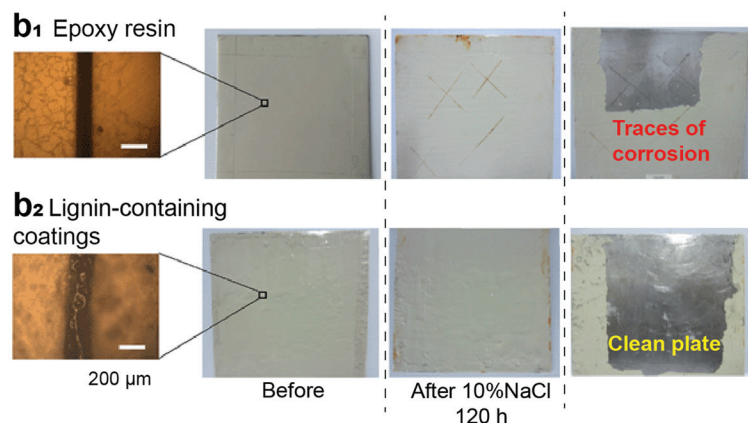
a₁ Formation of lignin particulated coatings with controlled morphology



a₂



b Anticorrosion coatings using lignin particle-based self-healing capsules



c Epoxy resin added of lignin particles

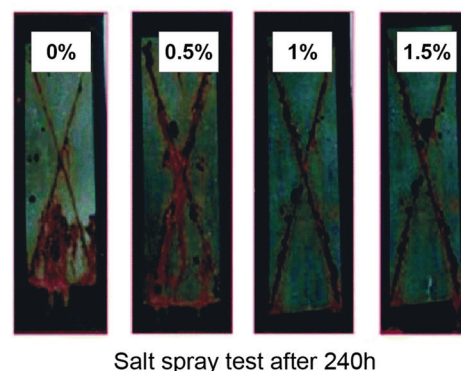


Fig. 6 Utilization of lignin particles in coatings of paints toward given functionalities. (a1) The morphology of the lignin particle coatings can be manipulated by controlling the drying step, in which (a2) the packing of the spherical lignin over a given substrate is influenced by the dispersivity of the system and drying rate.⁵⁵ (b) Anticorrosion films were obtained using microcapsules prepared from a lignin particle stabilized emulsion. The coating displays self-healing properties and advanced corrosion protection when compared to their neat epoxy counterparts.¹³⁹ Brine accelerating and (c) salt spray test methods have showed the high performance of lignin particles against corrosion. (c) A simple addition of lignin particles to epoxy films already warrants anticorrosion features to the coating.¹³³ Reproduced with permission of ref. 55, 135, and 129. Copyright © 2018, American Chemical Society; Copyright © 2015, Royal Society of Chemistry; and Copyright © 2018, Royal Society of Chemistry.



evaporation-induced assembly of spherical lignin particles into 30–40 μm thick coatings on silica wafers (Fig. 6a).⁵⁵ They observed stratification upon drying of the polydisperse particles. Microscaled particles sedimented into the bottom layer, while <300 nm colloidal particles formed the top layer upon evaporation of water from the dispersion. Drying at higher temperatures increased heterogeneity of particle size distribution across the film thickness – a phenomenon that the authors explained by kinetic constraint for arrangement of the particles at higher evaporation rates. Zikeli *et al.* isolated lignin from sawdust for the preparation of LNPs that were successively used for coating of beech wood.¹⁴⁰ Spherical LNPs that originated from spruce sawdust and contained aromatic extractive compounds exhibited better protection against overall color change during accelerated weathering compared to the LNPs from hardwood sawdust. The authors observed disintegration of the spherical particles after one to five days exposure to UV irradiation.

Rahman *et al.* studied synthesis, characterization and corrosion protection performance of acid-precipitated lignin nanoparticles (Fig. 6c).¹³³ These non-spherical lignin particles were blended with epoxy resin and the resulting nanocomposite was used to coat carbon steel. Potentiodynamic polarization and electrical impedance spectroscopy measurements showed that the nanocomposite coating inhibited corrosion of carbon steel better than the sole epoxy resin. There are a few other studies on composite coatings that contain non-spherical lignin.^{141–143} The combination of cellulose nanocrystals (CNC) and lignin produced smooth and homogeneous nanocomposite coatings on quartz surfaces.¹⁴¹ In another composite approach, incorporation of lignin in siloxane–poly(methyl methacrylate) coating increased hydrophobicity, scratch resistance, and thermal stability as well as provided corrosion resistance to carbon steel in saline.¹⁴³

Overall, lignin-based particulate coatings are relatively rare in the current literature, and future work should demonstrate their properties compared to technically and commercially relevant benchmarks. For example, multilayer coatings are typical in industrial products.¹³¹ Furthermore, strong adhesion of the coating on the substrate is in most cases essential to ensure stability under application conditions.¹⁴⁴ Commercial formulations contain curing agents that “harden” the coating and strengthen its adhesion on the substrate. Stabilization of coatings based on lignin particles is very likely needed as well. Cusola *et al.* demonstrated this by using wet strength agents to stabilize spherical lignin particle membranes against disintegration in water.³¹

Fundamental understanding of adhesion, adsorption and other relevant interfacial phenomena rely on studies using well-defined thin films and surface sensitive techniques like atomic force microscopy and quartz crystal microbalance. These methods have been used to understand, for example, direct surface forces between lignin surfaces,¹⁴⁵ or enzyme¹⁴⁶ or protein¹⁴⁷ adsorption to lignin. However, during the solvent shifting method for the production of stable lignin particle dispersions, lignin will assemble with hydrophilic moieties

concentrated at the surface, leading to different surface chemistry compared to lignin model films used so far. Hence, more research is needed also to develop methodology to control packing density and uniformity of model films made of spherical lignin particles in order to investigate how particulate films differ from films prepared from dissolved lignin.

Composites

With the societal demand for eco-friendly and renewable materials there has been an increasing interest to use lignin in various composite materials. As summarized in a recent review on lignin valorization,¹⁴⁸ lignin can enhance the mechanical and thermal stability of polymer blends or composite, act as adsorbing agent, or as reactive component in various resins, act as UV blocker, antimicrobial agent or flame retardant.¹⁴⁸ However, it has been observed that the use of raw, irregular lignin powders hinders high loads in composites, mostly because of the natural poor compatibility but also due to the uneven distribution of particles with varying size and morphology. Spherical lignin particles with controlled surface chemistry could overcome these issues, thus allowing their broader use in composite applications and being very promising although the reported results are rather contradictory. The main reason is that the geometry, morphology and surface chemistry display a crucial effect on miscibility with the matrix, and to date there are very few reports on spherical well-defined particles.

In a study comparing mechanical and thermal properties of particulate composites from polylactic acid (PLA) and lignin with PLA and tannin, Anwer *et al.*¹⁴⁹ found that said properties decreased with increasing amount of either tannin or lignin particles. Although the authors did not use spherical lignin particles, it is interesting to notice that they found that the droplet size of lignin within the PLA matrix had an effect on the mechanical properties, with smaller droplets having less pronounced detrimental effect. This would suggest that the easily dispersible spherical lignin particles with diameters of a few hundred nanometers or less would probably enhance further the mechanical and thermal performance of lignin-containing PLA composites. Indeed Yang *et al.*¹⁵⁰ used LNPs prepared using the acidolysis method⁴⁸ to reinforce PLA composites. They found that the lignin particles induced nucleation when added at 1 wt% in melt-extruded samples, leading to enhanced mechanical properties, but already an addition of 3 wt% led to aggregation and to a decrease in mechanical properties. The acidolysis method leads to irregular particles, which may be the reason for the aggregation and negative impact already at very low addition levels. Del Saz-Orozco *et al.*¹⁵¹ reported reinforcement of phenolic foams using lignin particles and found over 100% increase in both compressive strength and modulus. However, they used calcium lignosulfonate microparticles.

Several authors have reported increased antioxidant and UV shielding properties by the addition of lignin particles to various polymer matrixes (Fig. 7c and d).^{46,152–155} Most probably due to the spherical structure of the LNPs used in their



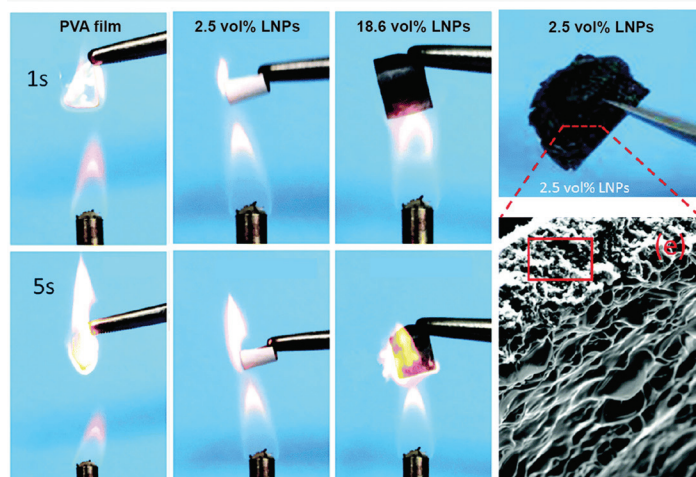
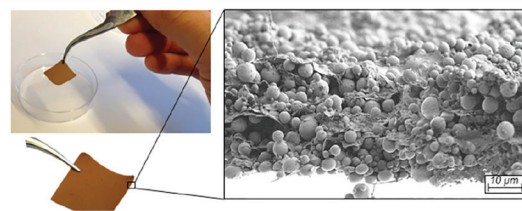
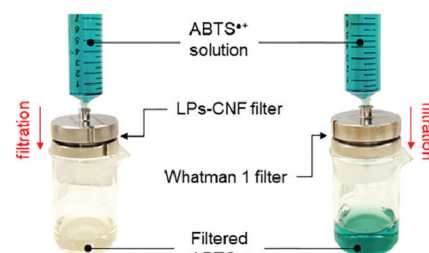
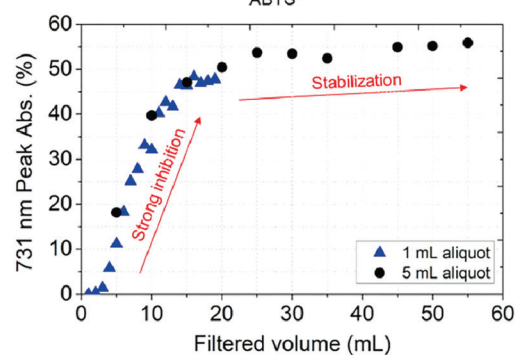
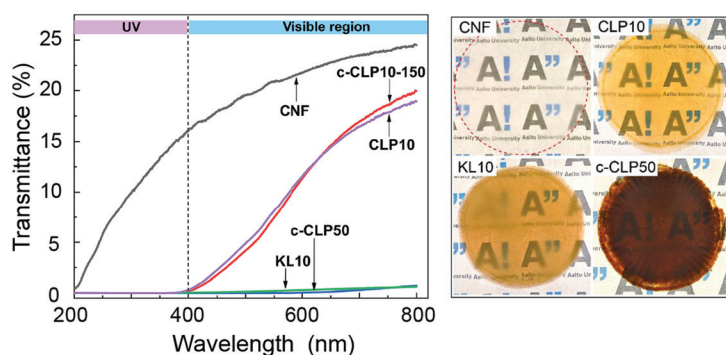
a Fire resistant composites enabled by lignin particles**b** Lignin particle-WSA-CNFs composite**c** Membranes with antioxidant properties**d** Lignin particle composites with UV-shield properties

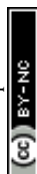
Fig. 7 Composites containing lignin particles displaying a series of important technical properties. (a) PVA films with added lignin particles display anti-fire resistance.¹⁵⁶ Composites with cellulose nanofibrils, lignin particles and a small fractions of wet-strength agent (polyamidoamine-epichlorohydrin resin) formed to (b) water resistant and (c) antioxidant membranes.³¹ (d) Lignin particles were also shown to form very strong UV-shielding films together with cellulose nanofibrils.⁴⁶ Reproduced with permission of ref. 152, 30, and 46. Copyright © 2019, Royal Society of Chemistry; and Copyright © 2018 and 2019, American Chemical Society.

studies, both Farooq *et al.*⁴⁶ and Tian *et al.*¹⁵⁴ were able to add higher amounts of lignin particles without loss in the mechanical properties when compared to previous reports. The hydrogen bonding ability between lignin and the matrices (PVA or cellulose nanofibrils) was an additional feature inducing high mechanical strength. In a recent study LNPs were also shown to reduce thermal conductivity, stability and flame retardancy to boron nitride PVA composites (Fig. 7a).¹⁵⁶

The particle-matrix interactions are crucial to achieve an even distribution of the particles. Frequently, chemical modification of the particles is required for such endeavor. This was demonstrated by He *et al.* who showed that esterification and etherification of lignin nanoparticles using citric acid enables an even distribution of up to 10 wt% of the particles in a PLA matrix to form transparent and active films.¹⁵⁵ Lignin nanoparticles have also been chemically modified to achieve specific properties. Setälä *et al.* prepared LNPs from tall oil

fatty acid ester of kraft lignin and bound these to cellulose fibers achieving composites with antimicrobial properties.¹⁵⁷

UV shielding, antioxidant and antimicrobial properties gained by the use of mostly nano but also submicron lignin particles argue for their use in composite materials, especially for packaging applications. However, the enhanced mechanical performance reported justifies the use of purely spherical particles instead of their irregular counterpart. The next steps should be focused on the formulation optimization as far as the choice of particle production method and interfacial compatibilization. The hydrophilic particles obtained by solvent shifting seem promising to combine with hydrophilic matrixes like cellulose or PVA, while dry particles, or surface modified ones may be a better choice to combine with PLA or other melt extruded matrices. Since chemical modification of the particles adds both cost and environmental burden, the need for modification should be carefully evaluated.



Other applications

The application of spherical lignin particles in biomedical applications has been recently reviewed¹⁷ and will hence not be further discussed here. Some other interesting applications of LNPs are as templates for carriers and for enzyme immobilization^{12,13,158,159} polymer composites and blends,¹⁶⁰ precursors for carbonaceous materials^{161–163} and metal-organic particles,¹⁶⁴ agglomeration agents to facilitate virus removal in water purification,¹⁶⁵ and hybrid lignin–fatty acid capsules as phase-change materials for thermal energy storage.⁷⁰ These examples exemplify that spherical lignin par-

ticles may find use in a plethora of versatile and even unexpected applications.

The stability in applications and rate of degradation of lignin particles

A series of factors affect the stability of lignin during application and its rate of degradation upon use, storage and disposal. Destabilization or degradation can derive from, but are not limited to, variations in salinity or pH conditions of the environment, light, microorganisms (mostly through enzymatic activity) and exposure to solvents and heat (Fig. 8a).

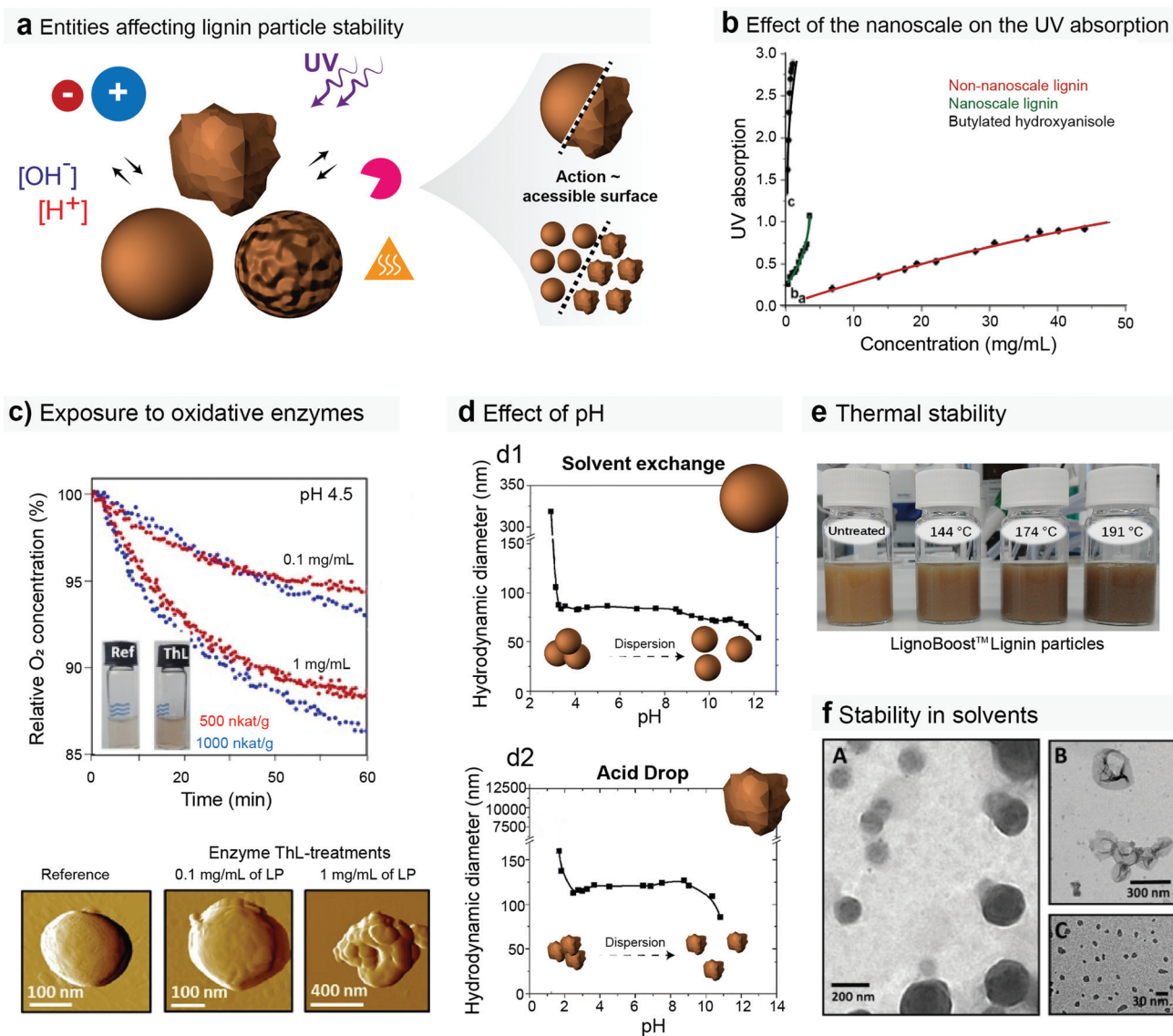


Fig. 8 (a) Possible entities (salts, pH conditions, light, enzymes, heat and solvents) affecting the degradation rate and stability of lignin particles either in storage conditions or applications. The degradation rate of surface-associated degradation may be proportional to the accessible surface area of the lignin particulates. (b) Effect of the nanoscale on the light absorption of the lignin particles.¹⁶⁸ (c) Oxidation of lignin particles induced by enzymes.¹⁶⁹ (d) Effect of pH on the stability, as far as hydrodynamic diameter, of irregular and spherical-shaped lignin nanoparticles.⁴⁹ (e) Effect of thermal treatments on the lignin particles dispersions properties.⁷¹ (f) Stability of unmodified and cross-linked lignin particles upon contact with THF.¹⁶⁹ Reproduced with permission of ref. 163, 164, 49, and 67. Copyright © 2012 and 2018, Elsevier Ltd; Copyright © 2016, American Chemical Society; and Copyright © 2016, Walter de Gruyter GmbH.



Unless any modification is done during the lignin particle preparation (e.g. bulk or surface functionalization),^{47,147,166} the physicochemical features (e.g. molecular mass, surface energy, solvation, thermal stability) of the lignin particles tend to mirror the ones from the starting lignin precursor.^{38,64,65,167} Therefore it is reasonable to suggest that the morphological distinction between the lignin nanoparticles and their macrostructured counterparts is the only parameter modifying their rate of degradation and stability. For instance, the rate of surface-associated degradation mechanisms (e.g. photodegradation and enzymatic depolymerization) should be proportional to the accessible surface area of the lignin particulates (colloids or irregular powder) (Fig. 8a).

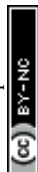
Regardless of the type of molecular structures (as affected by plant source and processing conditions), lignin is composed by chromophore species (phenols) in abundance. Such characteristic is, in fact, what puts lignin among the best bio-based light-absorbing compounds. This has been harnessed in many applications where light needs to be filtered or blocked, such as the applications in sunscreens discussed in earlier sections of this review. However, such intense light absorption triggers photodegradation of lignin¹⁷⁰ and lignin-containing materials.¹⁷¹ When comparing the absorbance at 700 nm of nano-scaled and macroscopic lignin powders it has been shown that the upper boundaries for light absorption are similar; however, this maximum value is achieved at remarkably lower concentration (12-fold lower) when lignin is used at the nano-scale (Fig. 8b).^{46,154} It is likely, although not yet scientifically proven, that such behavior does also intensify the photodegradation mechanisms of lignin, especially as light can easily penetrate micrometers into particulated, macroscopic materials⁶⁰ thus exposing the whole core of the nanoparticles to light. However, even with the potentially higher degradation rate, lignin nanoparticles are highly efficient to act as light barriers even at low fraction, creating a positive relationship between function and degradation over time.

Fungi is the most intense group of living organisms degrading lignin. Several bacteria can also decompose lignin, but to a lesser extent. Fungi excrete enzymes (e.g. peroxidases and laccases) that are able to modify and partially depolymerize the lignin macromolecule to further decompose its primary units. Although not yet fully understood, this subject has been discussed in different contexts.^{172–174} Surprisingly, although this has been a discipline of study for biomass-based materials (e.g. wood),¹⁷⁵ biodegradation of lignin particles by fungi has not been addressed in depth in the available literature. Fungal enzymes have been, however, exploited toward the controlled surface modification of the lignin particles (Fig. 8c). Mattinen *et al.* observed a 13% decrease in the dissolved oxygen concentration when lignin particles were treated with laccases, thus indicating evident oxidation of the lignin particles by the enzyme.¹⁶⁹ This suggests that enzymes could severely damage the lignin colloids if they interact for a longer time. In fact, enzymatic oxidation is the first stage for degradation of lignin in soil and therefore is key for composting efforts.¹⁷⁶ Also relevant for the fate of lignin particles in soil is the fact that they

may possess the same strong chelating properties commonly observed in polyphenolic molecules.^{177,178} This may warrant their low mobility in soil due to ionic binding with alkali metals and especially alkaline earth metals (e.g. Na⁺, K⁺, Mg²⁺, Ca²⁺) that are abundant in several types of soil, leading to aggregation and retention of the lignin nanoparticles in the organic and the surface layers of the soil.

The colloidal stability of lignin particles is heavily influenced by the conditions of the surrounding media, such as pH and ionic strength. A few recent studies have investigated the effects of pH and salt concentration on the hydrodynamic diameter and surface charge of the lignin particles when suspended in aqueous media. The formation of particle aggregates at pH below 3 is a consensus, regardless of particle source, shape or size. The protonation of the carboxyl groups present in the chemical structure of lignin takes place predominantly at pH 3–6, while phenolic hydroxyls in many lignin model compounds have pK_a values in the range of 7–10,¹⁷⁹ both thus providing surface charge on the particles that promotes the formation of repulsive electrical double layers. Such phenomenon has been investigated by tracking the zeta potential profiles of the lignin particles as a function of pH. The surface charge of the lignin particles gets closer to the isoelectric point under acidic conditions.^{35,47,49} Therefore, only above pH 3 the lignin particles are stabilized through electrical double layer repulsion. Interestingly, lignin particles from the same source (Indulin AT) but with different morphologies (irregular and spherical) presented different patterns in particle destabilization – particle aggregation – over pH changes (Fig. 8d). Spherical particles showed an extended region for particle stability with a sharper aggregation at pH ~3. Irregular lignin nanoparticles showed stability, with respect to size, from pH 3 to 9, with a remarkable decrease in size at pH 10 (Fig. 8d2). This is likely to be arising from fragmentation of the particles or even their partial dissolution.⁴⁹ Sipponen *et al.* showed that spherical particles prepared from soda lignin display a sharp onset of aggregation below pH 3, and that particle size decreases slightly during 4 h incubation at pH 7–10.⁹ In another study, Lievonen *et al.* showed that lignin particles are extremely stable in slightly acidic to neutral pH as, as their size and monodispersity in such conditions remained constant for over 60 days.³⁵ Carboxylic acid groups appear to be central also for the formation and dissolution of the particles. Figueiredo *et al.* synthesized carboxylated kraft lignin using succinic anhydride and found that excessive succinylation rendered lignin water-soluble, while a lower extent of succinylation allowed for peptide grafting on LNPs in water.⁸

Like for pH, the ionic strength of the dispersing media interferes with the stability of the kraft (LignoBoost) lignin particles. Lievonen *et al.* showed a significant shift to less negative zeta potential of the lignin particles upon addition of NaCl (from –60 to –10 mV); however, the dispersion was still stable at 500 mM NaCl and the particle size remained unchanged after seven days.³⁵ Particle aggregation took place at 1 M NaCl with average particle size increasing from 300 nm to 1.3 μm seven days after the preparation. The addition of salts changes



the interparticle interactions due to the accumulation of counterions around the particles, thus reducing the thickness of the electrical double layer (predicted by DLVO theory) and favoring the attractive van der Waals forces that may eventually lead to particle aggregation. Similar observations were made from the aggregation behavior of Indulin AT lignin nanoparticles upon addition of NaCl.⁴⁹ Such nanoparticles retained their colloidal stability up to 300 mM NaCl added, after which an exponential increase was observed reaching diameters of ~2 μm . The slightly distinct thresholds for destabilization may arise either from differences in surface charge or size of the lignin particles.

A few cross-linking or coating strategies have been carried out toward higher stability of the lignin particles for specific applications, *e.g.* soft tissue adhesion, emulsion stabilization, and for use in physiological conditions. Enzymes have been used to cross-link protein coated lignin particles.¹⁸⁰ With a combination of β -casein coating followed by crosslinking with transglutaminase, Mattinen *et al.* have shown remarkably high long-term stability (25 days) at pH below 4, conditions under which lignin particles usually start to aggregate.⁴⁴ On another instance, Leskinen *et al.* have taken advantage of the protein corona formation on lignin particles to tailor their surface properties to meet requirements of the potential applications in advanced biomaterials.¹⁴⁷ Relevant for stability purposes, the authors have shown that the addition of poly-L-lysine onto lignin particles reverses their zeta potential from negative (−40 mV) to positive (+40 mV); however, at a concentration range from 0.04 to 0.2 g protein by g of particles intense particle aggregation occurs due to neutralization of the surface charges. The size of the lignin particles starts to increase significantly after the coating with gelatin. The threshold for destabilization is at 0.1 g of gelatin by g of particles, at which point the zeta potential of the particles drops from −40 to −10 eV, thus being close to the isoelectric point of the system.

Coating of lignin particles was also carried out with synthetic polymeric polyelectrolytes, such as poly(diallyldimethylammonium chloride) (PDAC),^{47,49} aiming at extending their properties to broaden the applications. Charge reversal is observed for PDAC-coated lignin particles. Additionally, the PDAC coating increased the stability of the lignin particles against dissolution at pH over 10 due to the formation of a stable shell of polyelectrolyte–lignin complex around the particles.⁴⁹ PDAC-coated particles heavily aggregate at such high pH, going from 100 nm to 10 μm of diameter. The PDAC-coated nanoparticles aggregate at lower molarity of NaCl (at least one order of magnitude lower) when compared to native lignin particles. Sipponen *et al.* have taken a different, greener, approach to tune the surface properties of lignin nanoparticles and therefore to manipulate their performance and stability in applications.⁴⁷ Colloidally stable cationic dispersions were obtained with adsorption of only 40 mg g^{−1} of cationic lignin (Catlig) onto the lignin particles. The zeta potential was reversed from −30 to +30 mV using such strategy, and the size remained *ca.* 300 nm after the destabilization region – Catlig:LNPs ratio 10 to 40 mg g^{−1} – is overcome. With posi-

tively-charged LNPs, the authors could prepare efficiently stabilized toluene-in-water emulsions, with an additional feature related to emulsion breakage stimulated by pH (*ca.* pH 6) that is not possible when using anionic lignin particles.

Exposure of the lignin particles to heat and solvents can affect their stability or performance in applications. Nair *et al.* have demonstrated that the conversion of lignin macromolecules into nanoparticles does not significantly change their thermogravimetric profiles, with a maximum of thermal degradation taking place at 350 °C.⁵⁸ This is a key observation for the utilization of lignin particles in polymeric composites prepared using any heat-centered molding process (*e.g.* extrusion, hot press, injection). In another effort, thermal treatments (up to ~200 °C) of aqueous lignin particle dispersions were performed in order to increase the stability of the particles in organic solvents.⁷¹ The treated particles showed reduced solubility in lignin solvent systems, such as THF and methyl ethyl ketone (MEK)–water. However, under harsher treatment conditions the particles were heavily modified displaying darker color and sulfuric malodorous odors.

Higher solvent stability of the lignin particles can also be achieved through enzymatic cross-linking of the particle surfaces. Mattinen *et al.* performed intraparticle cross-linking reaction on the lignin particle surfaces using laccases from *Trametes hirsuta* and *Melanocarpus albomyces* fungi. With particle size and surface charges roughly unchanged, the cross-linked particles presented significantly higher resistance to organic solvents after dispersion and incubation. It is important to note that for some applications, especially as template for advanced materials,¹⁵⁹ the control over solubility of lignin particles in organic solvents is a key feature for their utilization. Additionally, it is still not known what the effects of cross-linking on biodegradability and recyclability of lignin particles are. These are paramount features for creating a robust platform of sustainable materials.

Sustainability aspects

In its simplest form, valorization of lignin has been shown to produce environmental benefits.¹⁸¹ For example, a cradle-to-gate analysis of the production of bleached fibers indicates that introducing lignin extraction in the kraft process, the most common process available in industry to produce wood pulp for papermaking, resulted in global warming potential of 602 kg CO₂eq per air dry metric ton (ADmt) pulp as compared to 722 kg CO₂eq per ADmt pulp for the reference scenario of the kraft mill with no lignin extraction. Moreover, if the so-called “system expansion” is considered, for example, considering the displacement of petroleum derived phenolic resins, the global warming potential is further reduced to 234 kg CO₂eq.¹⁸¹ More specifically, the system expansion calculations included credits by displacing grid electricity and petroleum derived phenolic resins. Thereby, significant greenhouse gas emission reductions were determined. However, lignin particles possess even further advantages, and it is possible to



achieve even better environmental metrics and sustainability scores, for example, in such applications as emulsion stabilizers, UV protection products, chelating agents, carbon nano-fillers (other than carbon black), and for composite reinforcement, all of which are economically attractive.⁴¹ Finally, these possibilities for substitution are of relatively limited market size but, as introduced in this review, other potential applications of larger scales can be considered for lignin particles, such as in thermoplastics, advanced resin composites, foams, bactericides, and others. It is apparent that, while LCA studies are available to assess the environmental impacts of lignocellulosic biorefineries, around the cellulosic and hemicellulose fractions, LCA studies on lignin, and especially lignin nanoparticles, are far less common. A first attempt in this regards was recently made by Koch *et al.*¹⁸² They reported life cycle assessment calculations on a lignin nanoparticle biorefinery using hot ethanol:water mixture to dissolve lignin and precipitating the lignin particles in water at pH 2, thus combining solvent and pH shifting. They found that one of the hot spots was the thermal energy needed for ethanol recovery, which is in accordance to the techno economical assessment of another solvent shifting process that also found that solvent recovery was the most energy demanding step during lignin particle production.¹⁸³ That calculation was made for THF:ethanol:water system. However, recently Lourençon *et al.* found that using acetone:water as solvent in the atomization process the reactor did not need to be heated for solvent evaporation and lignin concentration up to 5% could be used.⁴⁰ Although the authors concentrated on the economic savings, it can be concluded that the right choice of solvent will have a decisive effect on energy consumption and an impact on the overall sustainability of the processes. With regards to eutrophication, ethanol leakage into fresh water was found to be a hot spot.¹⁸² There is clearly a need for more LCA regarding lignin nanoparticle production. Various particle production methods should be compared and, most importantly, the applications should be included in the cradle-to-gate investigations.

Conclusions and prospects

From the literature reviewed above it is clear that spherical lignin particles with well-defined surface chemistry and morphology have a great potential in many applications. They can improve the properties of renewable and biodegradable composites, be part of greener adhesives and decrease the need for synthetic emulsifiers, to name a few applications. Even reports on experiments using crude lignin often conclude that the size of particles and dispersability are crucial for good performance, suggesting that spherical lignin particles with narrow size distributions in nanometer range and controlled surface chemistry would have an advantage over irregular lignin. Good spreading on a surface, even distribution in media, or high affinity to *e.g.* biological cells are important factors that can be achieved by controlling the surface chemistry of the particles.

With dry particles the surface energy can be controlled by the right choice of lignin source and the surface topography of the particles using a blowing agent. The solvent shifting method, on the other hand, results in hydrophilic particles with excellent dispersability in aqueous media. They can be modified by adsorption or grafting of molecules on the surface to tune the interactions with composite matrix, surfaces, cells *etc.*

In many of the applications the lignin particle production method does not matter, both dry and wet particles work well. However, to minimize the consumption of solvents and energy it would be beneficial to assess what type of particles would be more beneficial. In applications that require large volumes of particles the production process needs to be scalable. The techno-economical feasibility of production processes has been evaluated for both dry⁴¹ and wet approaches¹⁸³ and both are scalable. For the wet approach, the most energy consuming steps are the drying of particles and recycling of the solvents. However, demonstrations of producing large quantities (kg) of particles have so far been made only using the wet approach and a continuous process has also been developed.¹⁸⁴

Most reported methods rely on dissolving lignin that has been precipitated from the pulping process, sometimes further purified, and dried. These intermediate steps add substantial energy and chemical consumption and it would be ideal if the black liquor could be used as such as raw material or alternatively if the lignin particles could be produced directly from biorefinery residues. It has been shown that sequentially isolated lignin fractions from the same feedstock differ in their structural features,^{185,186} and that these fractions lead to different properties of the spherical lignin particles.¹⁸⁶ However, there is a lack of correlation of particle properties with lignin properties from various sources and after different extent of purification. When precipitated from spent pulping liquors, lignin often contains impurities such as polysaccharides, rosin acids, and other extractives from wood. These impurities may affect the properties of the lignin nanoparticles, but more studies are needed to elucidate such relationship. Different solvents have been used with the solvent shifting process but not much knowledge is yet available related to how the choice of solvent system affects the properties. There would be a demand for fundamental research into these issues, utilizing *e.g.* molecular modelling or surface force measurements to gain a better understanding of the interactions and process–property relationship for the particles.

In applications where dry particles are needed, like in the production of carbonized materials or composites together with nonpolar matrixes, it is probably better to apply the dry approach of particle production to avoid the energy needed to remove the excess of water from wet particles. Also for producing highly hydrophobic particles, this may be the approach of choice. Although hydrophobic molecules may be adsorbed onto the surface of lignin nanoparticles in aqueous media, successful hydrophobization will inevitably lead to phase separation. However, for applications in aqueous media, like emulsion stabilization, drug delivery or composites with cellulose fibers or fibrils or other hydrophilic polymers, the easy surface



modification by adsorption and low energy consumption of the process speaks for the choice of the wet nanoprecipitation method for these applications.

To guide the decision making during process development and application of spherical lignin particles, one aspect that needs more focus is the LCA and sustainability metrics. Especially considering both the effect of chemical modification during the production as well as the impact of using lignin particles instead of current, non-renewable components, in the applications.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was a part of the Academy of Finland's Flagship Programme under Projects No. 318890 and 318891 (Competence Center for Materials Bioeconomy, FinnCERES). O. J. R. also acknowledges support by the European Research Council under the European Union's Horizon 2020 research and innovation program (ERC Advanced Grant Agreement No. 788489, "BioElCell"), the Canada Excellence Research Chair initiative and the Canada Foundation for Innovation (CFI).

References

- 1 F. S. Chakar and A. J. Ragauskas, *Ind. Crops Prod.*, 2004, **20**, 131–141.
- 2 J. H. Lora and W. G. Glasser, *J. Polym. Environ.*, 2002, **10**, 39–48.
- 3 J. Gierer, *Wood Sci. Technol.*, 1980, **14**, 241–266.
- 4 M. H. Sipponen, J. Rahikainen, T. Leskinen, V. Pihlajaniemi, M.-L. Mattinen, H. Lange, C. Crestini and M. M. O. Österberg, *Nord. Pulp Pap. Res. J.*, 2017, **32**, 550–571.
- 5 W. Zhao, B. Simmons, S. Singh, A. Ragauskas and G. Cheng, *Green Chem.*, 2016, **18**, 5693–5700.
- 6 P. Figueiredo, K. Lintinen, A. Kiriazis, V. Hynninen, Z. Liu, T. Bauleth-Ramos, A. Rahikkala, A. Correia, T. Kohout, B. Sarmiento, J. Yli-Kauhaluoma, J. Hirvonen, O. Ikkala, M. A. Kostiaainen and H. A. Santos, *Biomaterials*, 2017, **121**, 97–108.
- 7 P. Figueiredo, C. Ferro, M. Kemell, Z. Liu, A. Kiriazis, K. Lintinen, H. F. Florindo, J. Yli-Kauhaluoma, J. Hirvonen, M. A. Kostiaainen and H. A. Santos, *Nanomedicine*, 2017, **12**, 2581.
- 8 P. Figueiredo, M. H. Sipponen, K. Lintinen, A. Correia, A. Kiriazis, J. Yli-Kauhaluoma, M. Österberg, A. George, J. Hirvonen, M. A. Kostiaainen and H. A. Santos, *Small*, 2019, **15**, 1901427.
- 9 M. H. Sipponen, H. Lange, M. Ago and C. Crestini, *ACS Sustainable Chem. Eng.*, 2018, **6**, 9342–9351.
- 10 Y. Li, X. Qiu, Y. Qian, W. Xiong and D. Yang, *Chem. Eng. J.*, 2017, **327**, 1176–1183.
- 11 L. Dai, R. Liu, L. Q. Hu, Z. F. Zou and C. L. Si, *ACS Sustainable Chem. Eng.*, 2017, **5**, 8241–8249.
- 12 M. H. Sipponen, M. Farooq, J. Koivisto, A. Pellis, J. Seitsonen and M. Österberg, *Nat. Commun.*, 2018, **9**, 2300.
- 13 E. Capecchi, D. Piccinino, I. Delfino, P. Bollella, R. Antiochia and R. Saladino, *Nanomaterials*, 2018, **8**, 438.
- 14 P. Anastas and N. Eghbali, *Green Chem.*, 2010, **39**, 301–312.
- 15 M. Tortora, F. Cavaliere, P. Mosesso, F. Ciaffardini, F. Melone and C. Crestini, *Biomacromolecules*, 2014, **15**, 1634–1643.
- 16 S. Rai, B. K. Singh, P. Bhartiya, A. Singh, H. Kumar, P. K. Dutta and G. K. Mehrotra, *J. Lumin.*, 2017, **190**, 492–503.
- 17 M. H. Sipponen, H. Lange, C. Crestini, A. Henn and M. Österberg, *ChemSusChem*, 2019, **12**, 2039–2054.
- 18 F. J. Ruiz-Dueñas and Á. T. Martínez, *Microb. Biotechnol.*, 2009, **2**, 164–177.
- 19 W. J. Stark, P. R. Stoessel, W. Wohlleben and A. Hafner, *Chem. Soc. Rev.*, 2015, **44**, 5793–5805.
- 20 S. Li, J. Zhao, P. Lu and Y. Xie, *Chin. Sci. Bull.*, 2010, **55**, 114–119.
- 21 A. Albanese, P. S. Tang and W. C. W. Chan, *Annu. Rev. Biomed. Eng.*, 2012, **14**, 1–16.
- 22 A. T. Bell, *Science*, 2003, **299**, 1688–1691.
- 23 J. C. Fröberg, O. J. Rojas and P. M. Claesson, *Int. J. Miner. Process.*, 1999, **56**, 1–30.
- 24 L. Suresh and J. Y. Walz, *J. Colloid Interface Sci.*, 1996, **183**, 199–213.
- 25 L. Suresh and J. Y. Walz, *J. Colloid Interface Sci.*, 1997, **196**, 177–190.
- 26 S. Y. Shulepov and G. Frens, *J. Colloid Interface Sci.*, 1996, **182**, 388–394.
- 27 C. Shen, L.-P. Wang, B. Li, Y. Huang and Y. Jin, *Vadose Zone J.*, 2012, **11**(1), DOI: 10.2136/vzj2011.0057.
- 28 S. Sharifi, S. Behzadi, S. Laurent, M. L. Forrest, P. Stroeve and M. Mahmoudi, *Chem. Soc. Rev.*, 2012, **41**, 2323–2343.
- 29 A. E. Nel, L. Mädler, D. Velegol, T. Xia, E. M. V. Hoek, P. Somasundaran, F. Klaessig, V. Castranova and M. Thompson, *Nat. Mater.*, 2009, **8**, 543–557.
- 30 X. Zhao, S. Ng, B. C. Heng, J. Guo, L. Ma, T. T. Y. Tan, K. W. Ng and S. C. J. Loo, *Arch. Toxicol.*, 2013, **87**, 1037–1052.
- 31 O. Cusola, O. J. Rojas and M. B. Roncero, *ACS Appl. Mater. Interfaces*, 2019, **11**, 45226–45236.
- 32 S. Baumberger, C. Lapierre, B. Monties and G. Della-Valle, *Polym. Degrad. Stab.*, 1998, **59**, 273–277.
- 33 B. Košíková, A. Gregorová, A. Osvald and J. Krajčovičová, *J. Appl. Polym. Sci.*, 2007, **103**, 1226–1231.
- 34 O. Gordobil, R. Delucis, I. Egués and J. Labidi, *Ind. Crops Prod.*, 2015, **72**, 46–53.



- 35 M. Lievonen, J. J. Valle-Delgado, M.-L. Mattinen, E.-L. Hult, K. Lintinen, M. A. Kostianen, A. Paananen, G. R. Szilvay, H. Setälä and M. Österberg, *Green Chem.*, 2016, **18**, 1416–1422.
- 36 T. Kämäräinen, M. Ago, J. Seitsonen, J. Raula, E. I. Kauppinen, J. Ruokolainen and O. J. Rojas, *Soft Matter*, 2018, **14**, 3387–3396.
- 37 K. Lintinen, Y. Xiao, R. Bangalore Ashok, T. Leskinen, E. Sakarinen, M. Sipponen, F. Muhammad, P. Oinas, M. Österberg and M. Kostianen, *Green Chem.*, 2018, **20**, 843–850.
- 38 M. Ago, S. Huan, M. Borghei, J. Raula, E. I. Kauppinen and O. J. Rojas, *ACS Appl. Mater. Interfaces*, 2016, **8**, 23302–23310.
- 39 J. Sameni, S. Krigstin, S. A. Jaffer and M. Sain, *Ind. Crops Prod.*, 2018, **117**, 58–65.
- 40 T. V. Lourençon, L. G. Greca, D. Tarasov, M. Borrega, T. Tamminen, O. J. Rojas and M. Y. Balakshin, *ACS Sustainable Chem. Eng.*, 2020, **8**, 1230–1239.
- 41 C. Abbati de Assis, L. G. Greca, M. Ago, M. Y. Balakshin, H. Jameel, R. Gonzalez and O. J. Rojas, *ACS Sustainable Chem. Eng.*, 2018, **6**, 11853–11868.
- 42 Y. Qian, Y. Deng, X. Qiu, H. Li and D. Yang, *Green Chem.*, 2014, **16**, 2156–2163.
- 43 S. X. Li, M. F. Li, J. Bian, X. F. Wu, F. Peng and M. G. Ma, *Int. J. Biol. Macromol.*, 2019, **132**, 836–843.
- 44 M.-L. Mattinen, G. Riviere, A. Henn, R. Nugroho, T. Leskinen, O. Nivala, J. Valle-Delgado, M. Kostianen and M. Österberg, *Nanomaterials*, 2018, **8**, 1001.
- 45 T. Zou, M. H. Sipponen and M. Österberg, *Front. Chem.*, 2019, **7**, 2300.
- 46 M. Farooq, T. Zou, G. Riviere, M. H. Sipponen and M. Österberg, *Biomacromolecules*, 2019, **20**, 693–704.
- 47 M. H. Sipponen, M. Smyth, T. Leskinen, L.-S. Johansson and M. Österberg, *Green Chem.*, 2017, **19**, 5831–5840.
- 48 C. Frangville, M. Rutkevičius, A. P. Richter, O. D. Velev, S. D. Stoyanov and V. N. Paunov, *ChemPhysChem*, 2012, **13**, 4235–4243.
- 49 A. P. Richter, B. Bharti, H. B. Armstrong, J. S. Brown, D. Plemmons, V. N. Paunov, S. D. Stoyanov and O. D. Velev, *Langmuir*, 2016, **32**, 6468–6477.
- 50 A. P. Richter, J. S. Brown, B. Bharti, A. Wang, S. Gangwal, K. Houck, E. A. Cohen Hubal, V. N. Paunov, S. D. Stoyanov and O. D. Velev, *Nat. Nanotechnol.*, 2015, **10**, 817–823.
- 51 W. Yang, M. Rallini, M. Natali, J. Kenny, P. Ma, W. Dong, L. Torre and D. Puglia, *Mater. Des.*, 2019, **161**, 55–63.
- 52 W. Yang, F. Dominici, E. Fortunati, J. M. Kenny and D. Puglia, *Ind. Crops Prod.*, 2015, **77**, 833–844.
- 53 Y. Zhou, Y. Qian, S. Wu, X. Zhong, J. Huang and X. Qiu, *Holzforchung*, DOI: 10.1515/hf-2019-0091.
- 54 M. Ago, B. L. Tardy, L. Wang, J. Guo, A. Khakalo and O. J. Rojas, *MRS Bull.*, 2017, **42**, 371–378.
- 55 O. Cusola, S. Kivistö, S. Vierros, P. Batys, M. Ago, B. L. Tardy, L. G. Greca, M. B. Roncero, M. Sammalkorpi and O. J. Rojas, *Langmuir*, 2018, **34**, 5759–5771.
- 56 P. K. Mishra and R. Wimmer, *Ultrason. Sonochem.*, 2017, **35**, 45–50.
- 57 I. A. Gilca, V. I. Popa and C. Crestini, *Ultrason. Sonochem.*, 2015, **23**, 369–375.
- 58 S. S. Nair, S. Sharma, Y. Pu, Q. Sun, S. Pan, J. Y. Zhu, Y. Deng and A. J. Ragauskas, *ChemSusChem*, 2014, **7**, 3513–3520.
- 59 A. A. Myint, H. W. Lee, B. Seo, W. S. Son, J. Yoon, T. J. Yoon, H. J. Park, J. Yu, J. Yoon and Y. W. Lee, *Green Chem.*, 2016, **18**, 2129–2146.
- 60 B. D. Mattos, L. G. Greca, B. L. Tardy, W. L. E. Magalhães and O. J. Rojas, *Small*, 2018, **14**, 1801256.
- 61 P. K. Mishra and A. Ekielski, *Colloids Interfaces*, 2019, **3**, 52.
- 62 M. B. Agustin, P. A. Penttilä, M. Lahtinen and K. S. Mikkonen, *ACS Sustainable Chem. Eng.*, 2019, **7**, 19925–19934.
- 63 Y. Pang, S. Wang, X. Qiu, Y. Luo, H. Lou and J. Huang, *J. Agric. Food Chem.*, 2017, **65**, 11011–11019.
- 64 H. Li, Y. Deng, H. Wu, Y. Ren, X. Qiu, D. Zheng and C. Li, *Holzforchung*, 2016, **70**, 725–731.
- 65 H. Li, Y. Deng, B. Liu, Y. Ren, J. Liang, Y. Qian, X. Qiu, C. Li and D. Zheng, *ACS Sustainable Chem. Eng.*, 2016, **4**, 1946–1953.
- 66 M. Yang, W. Zhao, S. Singh, B. Simmons and G. Cheng, *Nanoscale Adv.*, 2019, **1**, 299–304.
- 67 F. Xiong, Y. Han, S. Wang, G. Li, T. Qin, Y. Chen and F. Chu, *Ind. Crops Prod.*, 2017, **100**, 146–152.
- 68 J. Wang, Y. Qian, L. Li and X. Qiu, *ChemSusChem*, 2020, DOI: 10.1002/cssc.201903132.
- 69 Z.-H. Liu, N. Hao, S. Shinde, M. L. Olson, S. Bhagia, J. R. Dunlap, K. C. Kao, X. Kang, A. J. Ragauskas and J. S. Yuan, *ACS Sustainable Chem. Eng.*, 2019, **7**, 2634–2647.
- 70 M. H. Sipponen, A. Henn, P. Penttilä and M. Österberg, *Chem. Eng. J.*, 2020, **393**, 124711.
- 71 T. Leskinen, M. Smyth, Y. Xiao, K. Lintinen, M.-L. Mattinen, M. A. Kostianen, P. Oinas and M. Österberg, *Nord. Pulp Pap. Res. J.*, 2017, **32**, 586–596.
- 72 Z. Liu, R. Qie, W. Li, N. Hong, Y. Li, C. Li, R. Wang, Y. Shi, X. Guo and X. Jia, *New J. Chem.*, 2017, **41**, 3190–3195.
- 73 Y. Deng, H. Zhao, Y. Qian, L. Lü, B. Wang and X. Qiu, *Ind. Crops Prod.*, 2016, **87**, 191–197.
- 74 Y. Chevalier and M. A. Bolzinger, *Colloids Surf., A*, 2013, **439**, 23–34.
- 75 F. M. Ernsberger and W. G. France, *Ind. Eng. Chem.*, 1945, **37**, 598–600.
- 76 D. Liu and Y. Peng, *Fuel*, 2015, **142**, 235–242.
- 77 D. Yang, X. Qiu, M. Zhou and H. Lou, *Energy Convers. Manage.*, 2007, **48**, 2433–2438.
- 78 D. Yang, H. Li, Y. Qin, R. Zhong, M. Bai and X. Q. Qiu, *J. Dispersion Sci. Technol.*, 2015, **36**, 532–539.
- 79 T. Li and S. Takkellapati, *Biofuels, Bioprod. Biorefin.*, 2018, **12**, 756–787.
- 80 M. K. R. Konduri and P. Fatehi, *ACS Sustainable Chem. Eng.*, 2015, **3**, 1172–1182.



- 81 X. Ouyang, L. Ke, X. Qiu, Y. Guo and Y. Pang, *J. Dispersion Sci. Technol.*, 2009, **30**, 1–6.
- 82 M. K. Konduri, F. Kong and P. Fatehi, *Eur. Polym. J.*, 2015, **70**, 371–383.
- 83 J. Chen, A. Eraghi Kazzaz, N. AlipoorMazandarani, Z. Hosseinpour Feizi and P. Fatehi, *Molecules*, 2018, **23**, 868.
- 84 L. Bai, L. G. Greca, W. Xiang, J. Lehtonen, S. Huan, R. W. N. Nugroho, B. L. Tardy and O. J. Rojas, *Langmuir*, 2019, **35**, 571–588.
- 85 H. Fan and A. Striolo, *Soft Matter*, 2012, **8**, 9533–9538.
- 86 A. B. Pawar, M. Caggioni, R. Ergun, R. W. Hartel and P. T. Spicer, *Soft Matter*, 2011, **7**, 7710–7716.
- 87 Z. Wei, Y. Yang, R. Yang and C. Wang, *Green Chem.*, 2012, **14**, 3230–3236.
- 88 Y. Yang, Z. Wei, C. Wang and Z. Tong, *Chem. Commun.*, 2013, **49**, 7144–7146.
- 89 T. E. Nypelö, C. A. Carrillo and O. J. Rojas, *Soft Matter*, 2015, **11**, 2046–2054.
- 90 M. R. V. Bertolo, L. B. Brenelli de Paiva, V. M. Nascimento, C. A. Gandin, M. O. Neto, C. E. Driemeier and S. C. Rabelo, *Ind. Crops Prod.*, 2019, **140**, 111591.
- 91 L. Dai, Y. Li, F. Kong, K. Liu, C. Si and Y. Ni, *ACS Sustainable Chem. Eng.*, 2019, **7**, 13497–13504.
- 92 T. Zou, M. H. Sipponen and M. Österberg, *Front. Chem.*, 2019, **7**, 2300.
- 93 A. Sadeghpour, F. Pirolt and O. Glatter, *Langmuir*, 2013, **29**, 6004–6012.
- 94 K. Lebdioua, A. Aimable, M. Cerbelaud, A. Videcoq and C. Peyratout, *J. Colloid Interface Sci.*, 2018, **520**, 127–133.
- 95 V. O. Ikem, A. Menner and A. Bismarck, *Angew. Chem., Int. Ed.*, 2009, **48**, 632.
- 96 B. P. Binks and S. O. Lumsdon, *Langmuir*, 2001, **17**, 4540–4547.
- 97 T. Iwata, *Angew. Chem., Int. Ed.*, 2015, **54**, 3210–3215.
- 98 M. Tuomela, M. Vikman, A. Hatakka and M. Itävaara, *Bioresour. Technol.*, 2000, **72**, 169–183.
- 99 J. D. Nguyen, B. S. Matsuura and C. R. J. Stephenson, *J. Am. Chem. Soc.*, 2014, **136**, 1218–1221.
- 100 D. Ye, S. Li, X. Lu, X. Zhang and O. J. Rojas, *ACS Sustainable Chem. Eng.*, 2016, **4**, 5248–5257.
- 101 B. D. Mattos, B. L. Tardy, W. L. E. Magalhães and O. J. Rojas, *J. Controlled Release*, 2017, **262**, 139–150.
- 102 S. C. Lee, S. H. Lee and K. Won, *Biotechnol. Bioprocess Eng.*, 2019, **24**, 258–263.
- 103 Y. Qian, X. Qiu and S. Zhu, *Green Chem.*, 2015, **17**, 320–324.
- 104 Y. Qian, X. Qiu and S. Zhu, *ACS Sustainable Chem. Eng.*, 2016, **4**, 4029–4035.
- 105 X. Qiu, Y. Li, Y. Qian, J. Wang and S. Zhu, *ACS Appl. Bio Mater.*, 2018, **1**, 1276–1285.
- 106 H. Zhang, F. Chen, X. Liu and S. Fu, *ACS Sustainable Chem. Eng.*, 2018, **6**, 12532–12540.
- 107 W. Zhu, J. Lu and L. Dai, *Part. Part. Syst. Charact.*, 2018, **35**, 1–8.
- 108 D. Kai, Y. K. Chua, L. Jiang, C. Owh, S. Y. Chan and X. J. Loh, *RSC Adv.*, 2016, **6**, 86420–86427.
- 109 Y. Qian, X. Zhong, Y. Li and X. Qiu, *Ind. Crops Prod.*, 2017, **101**, 54–60.
- 110 B. Wang, D. Sun, H. M. Wang, T. Q. Yuan and R. C. Sun, *ACS Sustainable Chem. Eng.*, 2019, **7**, 2658–2666.
- 111 S. Tan, D. Liu, Y. Qian, J. Wang, J. Huang, C. Yi, X. Qiu and Y. Qin, *Holzforchung*, 2019, **73**, 485–491.
- 112 S. C. Lee, T. M. T. Tran, J. W. Choi and K. Won, *Int. J. Biol. Macromol.*, 2019, **122**, 549–554.
- 113 H. Zhang, X. Liu, S. Fu and Y. Chen, *Int. J. Biol. Macromol.*, 2019, **133**, 86–92.
- 114 J. M. Gutiérrez-Hernández, A. Escalante, R. N. Murillo-Vázquez, E. Delgado, F. J. González and G. Toriz, *J. Photochem. Photobiol., B*, 2016, **163**, 156–161.
- 115 Y. Li, D. Yang, S. Lu, X. Qiu, Y. Qian and P. Li, *ACS Sustainable Chem. Eng.*, 2019, **7**, 6234–6242.
- 116 M. Morsella, M. Giammatteo, L. Arrizza, L. Tonucci, M. Bressan and N. D'Alessandro, *RSC Adv.*, 2015, **5**, 57453–57461.
- 117 J. Yu, L. Li, Y. Qian, H. Lou, D. Yang and X. Qiu, *Ind. Eng. Chem. Res.*, 2018, **57**, 15740–15748.
- 118 B. Danielson and R. Simonson, *J. Adhes. Sci. Technol.*, 1998, **12**, 923–939.
- 119 M. Olivares, J. A. Guzmán, A. Natho and A. Saavedra, *Wood Sci. Technol.*, 1988, **22**, 157–165.
- 120 S. Kalami, M. Arefmanesh, E. Master and M. Nejad, *J. Appl. Polym. Sci.*, 2017, **134**, 1–9.
- 121 T. V. Lourençon, S. Alakurtti, T. Virtanen, A. S. Jääskeläinen, T. Liitiä, M. Hughes, W. L. E. Magalhães, G. I. B. Muniz and T. Tamminen, *Holzforchung*, 2020, **74**, 175–183.
- 122 R. J. Li, J. Gutierrez, Y. L. Chung, C. W. Frank, S. L. Billington and E. S. Sattely, *Green Chem.*, 2018, **20**, 1459–1466.
- 123 X. Sun, Q. Lang, H. Zhang, L. Cheng, Y. Zhang, G. Pan, X. Zhao, H. Yang, Y. Zhang, H. A. Santos and W. Cui, *Adv. Funct. Mater.*, 2017, **27**, 1604617.
- 124 X. Zhao, Q. Lang, L. Yildirimer, Z. Y. Lin, W. Cui, N. Annabi, K. W. Ng, M. R. Dokmeci, A. M. Ghaemmaghami and A. Khademhosseini, *Adv. Healthcare Mater.*, 2016, **5**, 108–118.
- 125 K. L. Spiller, S. A. Maher and A. M. Lowman, *Tissue Eng., Part B*, 2011, **17**, 281–299.
- 126 R. Lev and D. Seliktar, *J. R. Soc., Interface*, 2018, **15**, 20170380.
- 127 D. Gan, W. Xing, L. Jiang, J. Fang, C. Zhao, F. Ren, L. Fang, K. Wang and X. Lu, *Nat. Commun.*, 2019, **10**, 1–10.
- 128 S. Rose, A. Prevotau, P. Elzière, D. Hourdet, A. Marcellan and L. Leibler, *Nature*, 2014, **505**, 382–385.
- 129 S. E. Klein, A. Alzagameem, J. Rumpf, I. Korte, J. Kreyenschmidt and M. Schulze, *Coatings*, 2019, **9**, 494.
- 130 S. Erakovic, A. Jankovic, G. C. P. Tsui, C. Y. Tang, V. Miskovic-Stankovic and T. Stevanovic, *Int. J. Mol. Sci.*, 2014, **15**, 12294–12322.



- 131 A. Mathiazhagan and R. Joseph, *Int. J. Chem. Eng. Appl.*, 2011, **2**, 225–237.
- 132 H. Wang, X. Qiu, W. Liu, F. Fu and D. Yang, *Ind. Eng. Chem. Res.*, 2017, **56**, 11133–11141.
- 133 O. U. Rahman, S. Shi, J. Ding, D. Wang, S. Ahmad and H. Yu, *New J. Chem.*, 2018, **42**, 3415–3425.
- 134 A. Dastpak, K. Yliniemi, M. Cecilio, D. O. Monteiro, S. Höhn, S. Virtanen, M. Lundström and B. P. Wilson, *Coatings*, 2018, **8**, 454.
- 135 C. Hao, T. Liu, S. Zhang, L. Brown, R. Li, J. Xin, T. Zhong, L. Jiang and J. Zhang, *ChemSusChem*, 2019, **12**, 1049–1058.
- 136 C. Zheng, D. Li and M. Ek, *Nord. Pulp Pap. Res. J.*, 2019, **34**, 96–106.
- 137 E. Larrañeta, M. Imízcoz, J. X. Toh, N. J. Irwin, A. Ripolin, A. Perminova, J. Domínguez-Robles, A. Rodríguez and R. F. Donnelly, *ACS Sustainable Chem. Eng.*, 2018, **6**, 9037–9046.
- 138 N. Forsman, A. Lozhechnikova, A. Khakalo, L.-S. Johansson, J. Vartiainen and M. Österberg, *Carbohydr. Polym.*, 2017, **173**, 392–402.
- 139 H. Yi, Y. Yang, X. Gu, J. Huang and C. Wang, *J. Mater. Chem. A*, 2015, **3**, 13749–13757.
- 140 F. Zikeli, V. Vinciguerra, A. D. Annibale, D. Capitani, M. Romagnoli and G. S. Mugnozza, *Nanomaterials*, 2019, **9**, 1–18.
- 141 A. Hambardzumyan, L. Foulon, B. Chabbert and V. Aguié-Béghin, *Biomacromolecules*, 2012, **13**, 4081–4088.
- 142 L. Dumitrescu, D. Perniu and I. Manciulea, *Solid State Phenom.*, 2009, **151**, 139–144.
- 143 S. V. Harb, B. M. Cerrutti, S. H. Pulcinelli, C. V. Santilli and P. Hammer, *Surf. Coat. Technol.*, 2015, **275**, 9–16.
- 144 D. S. Rickerby, *Surf. Coat. Technol.*, 1988, **36**, 541–557.
- 145 S. M. Notley and M. Norgren, *Langmuir*, 2006, **22**, 11199–11204.
- 146 J. L. Rahikainen, R. Martin-Sampedro, H. Heikkinen, S. Rovio, K. Marjamaa, T. Tamminen, O. J. Rojas and K. Kruus, *Bioresour. Technol.*, 2013, **133**, 270–278.
- 147 T. Leskinen, J. Witos, J. J. Valle-Delgado, K. Lintinen, M. Kostiaainen, S. K. Wiedmer, M. Österberg and M.-L. Mattinen, *Biomacromolecules*, 2017, **18**, 2767–2776.
- 148 M. N. Collins, M. Nechifor, F. Tanasă, M. Zănoagă, A. McLoughlin, M. A. Stróżyk, M. Culebras and C.-A. Teacă, *Int. J. Biol. Macromol.*, 2019, **131**, 828–849.
- 149 M. A. S. Anwer, H. E. Naguib, A. Celzard and V. Fierro, *Composites, Part B*, 2015, **82**, 92–99.
- 150 W. Yang, E. Fortunati, F. Dominici, J. M. Kenny and D. Puglia, *Eur. Polym. J.*, 2015, **71**, 126–139.
- 151 B. Del Saz-Orozco, M. Oliet, M. V. Alonso, E. Rojo and F. Rodríguez, *Compos. Sci. Technol.*, 2012, **72**, 667–674.
- 152 T. Ju, Z. Zhang, Y. Li, X. Miao and J. Ji, *RSC Adv.*, 2019, **9**, 24915–24921.
- 153 Y. Liu, *ACS Sustainable Chem. Eng.*, 2018, **6**, 5524–5532.
- 154 D. Tian, J. Hu, J. Bao, R. P. Chandra, J. N. Saddler and C. Lu, *Biotechnol. Biofuels*, 2017, **10**, 192.
- 155 X. He, F. Luzi, X. Hao, W. Yang, L. Torre, Z. Xiao, Y. Xie and D. Puglia, *Int. J. Biol. Macromol.*, 2019, **127**, 665–676.
- 156 X. Wang, S. Ji, X. Wang, H. Bian, L. Lin, H. Dai and H. Xiao, *J. Mater. Chem. C*, 2019, **7**, 14159–14169.
- 157 H. Setälä, H.-L. Alakomi, A. Paananen, G. R. Szilvay, M. Kellock, M. Lievonen, V. Liljeström, E.-L. Hult, K. Lintinen, M. Österberg and M. Kostiaainen, *Cellulose*, 2020, **27**, 273–284.
- 158 D. Piccinino, E. Capecci, L. Botta, P. Bollella, R. Antiochia, M. Crucianelli and R. Saladino, *Catal. Sci. Technol.*, 2019, **9**, 4125–4134.
- 159 B. L. Tardy, J. J. Richardson, J. Guo, J. Lehtonen, M. Ago and O. J. Rojas, *Green Chem.*, 2018, **20**, 1335–1344.
- 160 F. Xiong, Y. Wu, G. Li, Y. Han and F. Chu, *Ind. Eng. Chem. Res.*, 2018, **57**, 1207–1212.
- 161 D. Yiamsawas, S. Beckers, H. Lu, K. Landfester and F. R. Wurm, *ACS Biomater. Sci. Eng.*, 2017, **3**, 2375–2383.
- 162 J. Köhnke, H. Rennhofer, C. Unterweger, N. Gierlinger, J. Keckes, C. Zollfrank, O. J. Rojas and W. Gindl-Altmatter, *Nanomaterials*, 2018, **8**(12), 1055.
- 163 W. Gindl-Altmatter, J. Köhnke, C. Unterweger, N. Gierlinger, J. Keckes, J. Zalesak and O. J. Rojas, *Composites, Part A*, 2019, **121**, 175–179.
- 164 K. Lintinen, M. Latikka, M. H. Sipponen, R. H. A. Ras, M. Österberg and M. A. Kostiaainen, *RSC Adv.*, 2016, **6**, 31790–31796.
- 165 G. N. Rivière, A. Korpi, M. H. Sipponen, T. Zou, M. A. Kostiaainen and M. Österberg, *ACS Sustainable Chem. Eng.*, 2020, **8**, 4167–4177.
- 166 P. Figueiredo, K. Lintinen, J. T. Hirvonen, M. A. Kostiaainen and H. A. Santos, *Prog. Mater. Sci.*, 2018, **93**, 233–269.
- 167 W. Gao and P. Fatehi, *Can. J. Chem. Eng.*, 2019, **97**, 2827.
- 168 Q. Lu, M. Zhu, Y. Zu, W. Liu, L. Yang, Y. Zhang, X. Zhao, X. Zhang, X. Zhang and W. Li, *Food Chem.*, 2012, **135**, 63–67.
- 169 M.-L. Mattinen, J. J. Valle-Delgado, T. Leskinen, T. Anttila, G. Riviere, M. Sipponen, A. Paananen, K. Lintinen, M. Kostiaainen and M. Österberg, *Enzyme Microb. Technol.*, 2018, **111**, 48–56.
- 170 A. M. McNally, E. C. Moody and K. McNeill, *Photochem. Photobiol. Sci.*, 2005, **4**, 268.
- 171 K. Chaochanchaikul, K. Jayaraman, V. Rosarpitak and N. Sombatsompop, *BioResources*, 2012, **7**, 38–55.
- 172 G. Janusz, A. Pawlik, J. Sulej, U. Świdarska-Burek, A. Jarosz-Wilkolażka and A. Paszczyński, *FEMS Microbiol. Rev.*, 2017, **41**, 941–962.
- 173 M. A. Lara, A. J. Rodríguez-Malaver, O. J. Rojas, O. Holmquist, A. M. González, J. Bullón, N. Peñalosa and E. Araujo, *Int. Biodeterior. Biodegrad.*, 2003, **52**, 167–173.
- 174 R. Martin-Sampedro, E. A. Capanema, I. Hoeger, J. C. Villar and O. J. Rojas, *J. Agric. Food Chem.*, 2011, **59**, 8761–8769.
- 175 C. Sánchez, *Biotechnol. Adv.*, 2009, **27**, 185–194.
- 176 R. Datta, A. Kelkar, D. Baraniya, A. Molaei, A. Moulick, S. R. Meena and P. Formanek, *Sustainability*, 2017, **9**.



- 177 M. H. Sipponen, O. J. O. J. Rojas, V. Pihlajaniemi, K. Lintinen and M. Österberg, *ACS Sustainable Chem. Eng.*, 2017, **5**, 1054–1061.
- 178 J. Guo, B. L. Tardy, A. J. Christofferson, Y. Dai, J. J. Richardson, W. Zhu, M. Hu, Y. Ju, J. Cui, R. R. Dagastine, I. Yarovsky and F. Caruso, *Nat. Nanotechnol.*, 2016, **11**, 1105.
- 179 M. Ragnar, C. T. Lindgren and N.-O. Nilvebrant, *J. Wood Chem. Technol.*, 2000, **20**, 277–305.
- 180 A. Henn and M.-L. Mattinen, *World J. Microbiol. Biotechnol.*, 2019, **35**, 125.
- 181 C. Culbertson, T. Treasure, R. Venditti, H. Jameel and R. Gonzalez, *Nord. Pulp Pap. Res. J.*, 2016, **31**, 30–40.
- 182 D. Koch, M. Paul, S. Beisl, A. Friedl and B. Mihalyi, *J. Cleaner Prod.*, 2020, **245**, 118760.
- 183 R. P. Bangalore Ashok, P. Oinas, K. Lintinen, G. Sarwar, M. A. Kostianen and M. Österberg, *Green Chem.*, 2018, **20**, 4911–4919.
- 184 R. P. Bangalore Ashok, Y. Xiao, K. Lintinen, P. Oinas, M. A. Kostianen and M. Österberg, *Colloids Surf., A*, 2020, **587**, 124228.
- 185 M. H. Sipponen, C. Lapierre, V. Méchin and S. Baumberger, *Bioresour. Technol.*, 2013, **133**, 522–528.
- 186 Z. H. Liu, N. Hao, S. Shinde, Y. Pu, X. Kang, A. J. Ragauskas and J. S. Yuan, *Green Chem.*, 2019, **21**, 245–260.

