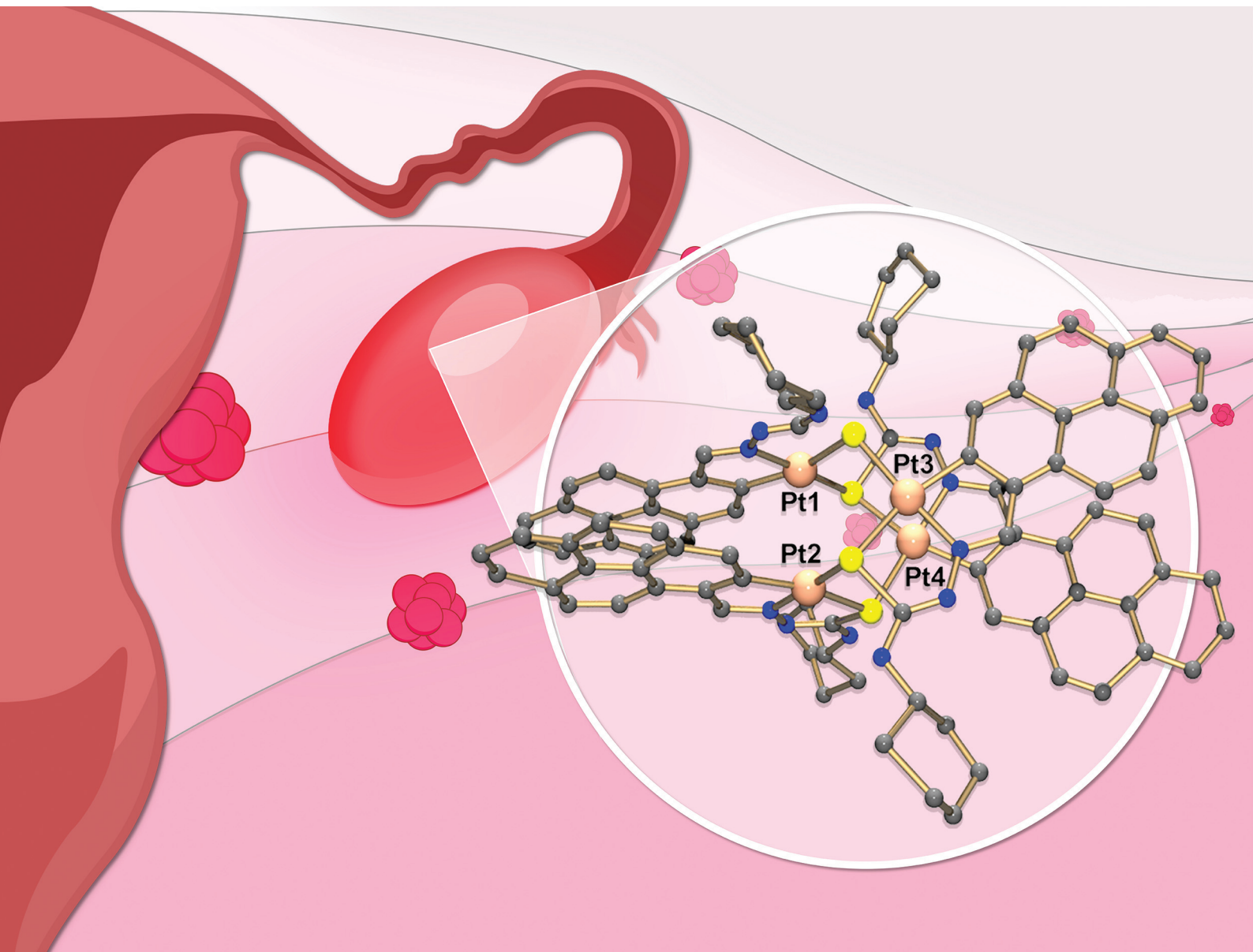


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PAPER

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Novel tetranuclear Pd^{II} and Pt^{II} anticancer complexes derived
from pyrene thiosemicarbazones

PAPER

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[View Journal](#) | [View Issue](#)Cite this: *Dalton Trans.*, 2020, **49**, 9595Novel tetranuclear Pd^{II} and Pt^{II} anticancer complexes derived from pyrene thiosemicarbazones†Carolina G. Oliveira,^{a,b,c} Isolda Romero-Canelón,^d James P. C. Coverdale,^c Pedro Ivo S. Maia,^e Guy J. Clarkson,^c Victor M. Deflon^{*a} and Peter J. Sadler^{*c}

Cyclometallated palladium(II) and platinum(II) pyrenyl-derived thiosemicarbazone (H₂PrR) complexes of the type [M₄(μ-S-PrR-κ³-C,N,S)₄] (M = Pd^{II}, Pt^{II}; R = ethyl, cyclohexyl) have been synthesised in good yields and fully characterised. X-ray crystallography showed that the tetranuclear complex [Pt₄(μ-S-PrCh-κ³-C,N,S)₄](CH₃)₂COCHCl₃ contains an eight-membered ring of alternating M–S atoms. The ethyl derivatives [M₄(μ-S-PrEt-κ³-C,N,S)₄] exhibit potent antiproliferative activity towards A2780 human ovarian cancer cells, with IC₅₀ values of 1.27 μM (for M = Pd^{II}) and 0.37 μM (for M = Pt^{II}), the latter being an order of magnitude more potent than the anticancer drug cisplatin (IC₅₀ 1.20 μM). These promising complexes had low toxicity towards non-cancerous human MRC5 cells, which points towards an early indication of differential toxicity between cancer and normal cells. Experiments that investigated the effects of these tetranuclear complexes on the cell cycle, integrity of the cell membrane, and induction of apoptosis, suggested that their mechanism of action of does not involve DNA targeting, unlike cisplatin, and therefore could be promising for combatting cisplatin resistance.

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Introduction

Thiosemicarbazones (TSCs) are well known as a promising class of compounds with a rich coordination chemistry^{1–3} and a range of pharmacological activities,^{4–6} including antiparasitic, antibacterial and antitumour activities. Some thiosemicarbazones such as the di-2-pyridylketone 4,4-dimethyl-3-thiosemicarbazone Dp44mT (Chart 1) have been extensively studied because of their selective antitumour and antimetastatic properties, causing lysosomal membrane permeabilisation and cell death.⁷ The thiosemicarbazone derivative Triapine (Chart 1), which inhibits cancer cell growth at nanomolar concentrations, has undergone Phase 1 and Phase 2 clinical trials for the treatment of cervical cancer.⁸ It is well established that triapine has the ability to suppress tumour growth by inhibiting ribonucleotide reductase (RR), a key enzyme required for

DNA synthesis.⁹ Following the successful anticancer agent cisplatin (CDDP) and its relatives carboplatin and oxaliplatin, thousands of other metal complexes have been screened in order to study the effects of the metal and their structures on the biological activity.^{10,11} However, the major challenge of this field, is still to eliminate strong side effects, off-target toxicity and achieve cancer-cell specificity. Accordingly, current antitumour metallo-drug research is focused on the development of compounds with new and unusual features, including metal–carbon bonds (organometallic compounds).

Organometallic complexes of palladium and platinum containing thiosemicarbazones as ligands have attracted attention. First because of their diverse range of structures, including oligomers, and second because organometallic bonding can increase the stability of the complex as a whole.¹² Some cyclometallated complexes have potential antitumour

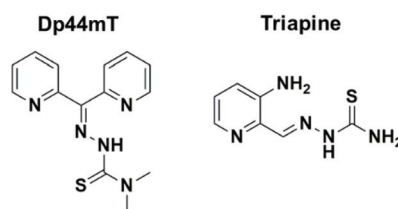


Chart 1 Molecular structures of thiosemicarbazones Dp44mT (topoisomerase IIa inhibitor) and triapine (compound in clinical trials).

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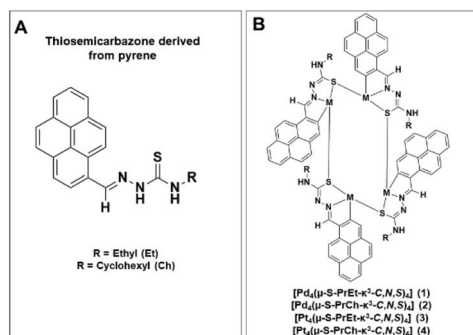


Fig. 1 (A) Thiosemicarbazone ligands derived from pyrene and (B) the general chemical structure of tetranuclear complexes studied in this work.

activity.¹³ Consequently, orthometallated thiosemicarbazone complexes of palladium and platinum have been reported.^{14,15} For example, Quiroga *et al.* have described platinum organometallic complexes derived from thiosemicarbazone ligands which have potent cytotoxic activity against three different cell lines.¹⁶

Thiosemicarbazone ligands are formed by condensation reactions of a thiosemicarbazide and an aldehyde. The ligand features depend on the aldehyde. Both thiosemicarbazide and aldehyde may provide coordination sites. In this work, we have used 1-pyrenecarboxaldehyde to incorporate a fluorescent probe to track complexes inside cells.¹⁷ Furthermore, the well-known intercalating properties of pyrene might enhance inhibition of cellular DNA replication,¹⁸ if DNA is a target.

Although there are many studies of metal thiosemicarbazone complexes, only a few reports describe the biological activity of tetranuclear orthometallated complexes with thiosemicarbazone CNS donors.^{19–21} Herein, we have used potential CNS thiosemicarbazone ligands (Fig. 1A) to prepare tetranuclear cyclometallated palladium and platinum complexes (Fig. 1B) and investigated their biological activity against A2780 human ovarian cancer cells. Since selectivity is important for reducing the severity of undesired side-effects, we have also studied the activity of the most potent complexes against a non-tumourigenic cell line.

Experimental

Materials and measurements

1-Pyrenecarboxaldehyde, 4-ethyl-thiosemicarbazide, analytical reagents grade chemicals and solvents were purchased from Sigma-Aldrich and used without further purification. The 4-cyclohexyl-3-thiosemicarbazide and *trans*- $[PdCl_2(MeCN)_2]$ were prepared as described previously.^{22,23} CHN elemental analyses were carried out on a CE-440 elemental analyzer by Warwick Analytical (UK) Ltd or on PerkinElmer CHNS/O 2400 Series II equipment. FTIR spectra between 400 and 4000 cm^{-1} were recorded using KBr pellets on a Shimadzu IR Prestige-21

spectrophotometer. Ultraviolet-visible (UV-vis) spectra of solutions diluted in dichloromethane were recorded using a Shimadzu UV-1800 spectrophotometer. 1H and ^{195}Pt NMR spectra were acquired in 5 mm NMR tubes at 298 K on Bruker spectrometers (1H = 300 or 400 MHz, ^{195}Pt = 107 MHz). 1H NMR chemical shifts were internally referenced to $CHCl_3$ (7.27 ppm) for chloroform. All the $\delta(Pt)$ values were referenced to $[PtCl_6]^{2-}$ via $[PtCl_4]^{2-}$ = −1628 ppm. HPLC studies were performed on a HP 1100 Series HPLC System (Agilent) using a ZORBAX Eclipse Plus C-18 column. The mobile phases were A: water (HPLC grade, Aldrich) containing 0.1% trifluoroacetic acid (TFA) and B: methanol (HPLC grade, Aldrich) containing 0.1% TFA. A flow rate of 1 mL min^{-1} was used. Electrospray ionization mass spectrometry (ESI-MS) spectra were recorded on an Agilent 6130B single quadrupole detector instrument at 298 K with a scan range of m/z 50–2000 for positive ions. Samples were prepared in mixture of DCM/MeOH (25%/75%)+0.3% formic acid. The stability of the complexes (*ca.* 500 μM) was investigated by UV-vis spectroscopy over 24 h in 5% DMSO/95% H_2O . A Cary 300 UV-Vis recording spectrophotometer was used with 1 cm path-length quartz cuvettes (0.5 mL) and a PTP1 Peltier temperature controller. Spectra were processed using UVWinlab software. Experiments were carried out at 298 K from 600 to 200 nm. ICP-OES analyses were made on a PerkinElmer Optima 5300 DV series ICP-OES instrument. The water (18.2 M Ω) used for analysis was doubly deionized (DDW) using a Millipore Milli-Q water purification system and a USF Elga UHQ water deionizer. ICP standards for platinum ($1001 \pm 2 \mu g mL^{-1}$, Fluka) and palladium ($1015 \mu g mL^{-1}$, Aldrich) were diluted with 3.6% HNO_3 DDW to prepare freshly calibrants at concentrations of 50–700 ppb, which were spiked with NaCl to match the salt content of the samples being analysed. All Inductively Coupled Plasma-Mass Spectrometry (ICP-MS) analyses were carried out on an Agilent Technologies 7500 series ICP-MS instrument. ICP standards were diluted with 3.6% HNO_3 DDW to prepare fresh calibrants at concentrations of 0.1–1000 ppb. The ICP-MS instrument was set to detect Pd and Pt with typical detection limits of *ca.* 2 ppt using no-gas mode, with an internal calibration standard of Er (50 pp).

Cell culture

Cell maintenance. A2780 human ovarian carcinoma cells and MRC-5 human fetal lung fibroblasts were obtained from the European Collection of Cell Cultures (ECACC). A2780 human ovarian carcinoma cells were used between passages 5 and 18. Cell lines were grown in Roswell Park Memorial Institute medium (RPMI-1640) supplemented with 10% of fetal calf serum, 1% of 2 mM glutamine and 1% penicillin/streptomycin. The cells were grown as adherent monolayers at 310 K in a 5% CO_2 humidified atmosphere and passaged at *ca.* 70–80% confluence.

In vitro growth inhibition assay. 5000 cells were seeded per well (150 μL) in 96-well plates. The cells were pre-incubated in drug-free medium at 310 K for 48 h before adding various concentrations of the compounds to be tested. Stock solutions of



the palladium complexes were firstly prepared in DMSO (5% v/v) and a mixture 0.9% saline and RPMI-1640 medium (1 : 1 v/v) following serial dilutions in RPMI-1640. Drug exposure period was 24 h. After this, supernatants were removed by suction and each well was washed with PBS. A further 72 h was allowed for the cells to recover in drug-free medium at 310 K. SRB assay was used to determine cell viability.²⁴ Absorbance measurements of the solubilized dye (on a BioRad iMark microplate reader using a 470 nm filter) allowed the determination of viable treated cells compared to untreated controls. IC₅₀ values (concentrations which caused 50% of cell death) were determined as duplicates of triplicates in two independent sets of experiments and their standard deviations were calculated.

Flow cytometry. The flow cytometry experiments were carried out using a Becton Dickinson FACScan Flow Cytometer at Warwick University (School of Life Sciences).

Metal accumulation in cancer cells

Cell seeding. Briefly, 4×10^6 cells were seeded in a P100 Petri dish using 10 ml of culture medium. Cells were incubated at 310 K for 24 h.

Sample preparation. 100 μ M solution of each compound was prepared in cell culture medium containing 5% DMSO. The exact concentration of Pt/Pd was determined by ICP-OES (using freshly prepared calibration standards, 50–700 ppb, with standard addition of sodium chloride to match the sample matrix). The stock solutions were diluted with culture medium to achieve a final working concentration equal to IC₅₀.

Drug addition. The supernatant medium was removed, and 10 mL of each compound to be tested was added in triplicate to the Petri dishes. Cells were exposed to the drugs for 24 h.

Drug removal. After supernatants were removed by suction, cells were washed with PBS and detached using trypsin/EDTA and a single cell suspension was obtained using culture medium. Cells were counted using a hemocytometer, and centrifuged (1000 rpm, 5 min, 277 K) to obtain whole-cell pellets.

Sample digestion. Cell pellets were digested in 72% v/v nitric acid using a CEM Discovery SP microwave reactor (3 min, 393 K, 150 W, 250 psi) then diluted to a working acid concentration of 3.6% v/v. Samples were analysed using an Agilent 7500 series ICP-MS in both no-gas and He-gas mode. Calibration samples for Pt/Pd (0.1–1000 ppb) were freshly prepared in 3.6% v/v nitric acid. Total dissolved solids for ICP-MS analysis did not exceed 0.1% w/v. Final metal accumulation is reported as ng Pt/Pd $\times 10^6$ cells and standard deviations were calculated.

Cell cycle. Briefly, 4×10^6 A2780 cancer cells were seeded in a 6-well plate using 2 ml per well and incubated for 24 h. The supernatant was removed by suction, and cells treated with complex 3 using IC₅₀ concentration. After this time, the supernatant was removed by suction, cells washed with PBS and cell pellets obtained using trypsin/EDTA. After collection and centrifugation, the pellets were washed with PBS and stained in the dark with a mixture of propidium iodide and RNase. After

30 min staining, cell samples were washed and set up for flow cytometry reading on the red channel FL-2.

Induction of apoptosis. Flow cytometry analysis of apoptosis in A2780 cells caused by exposure to complex 3 was carried out using the Annexin V-FITC Apoptosis Detection Kit (Sigma Aldrich) according to the manufacturer's instructions. Briefly, A2780 cells were seeded in six-well plates (1.0×10^6 cells per well), pre-incubated for 24 h in drug-free media at 310 K, after which they were exposed to either, complex 3 (concentration equal IC₅₀ and 3 times IC₅₀). Cells were harvested using trypsin and stained using PI/Annexin V-FITC. After staining, cell pellets were analysed in a Becton Dickinson FACScan Flow Cytometer. For positive-apoptosis controls, A2780 cells were exposed for 2 h to staurosporine (1 mg ml⁻¹). Cells for apoptosis studies were used with no previous fixing procedure as to avoid nonspecific binding of the annexin V-FITC conjugate.

Cellular membrane integrity. Flow cytometry analysis of cellular membrane integrity of A2780 cells caused by exposure to complex 3, was carried out using the CytoPainter assay (Abcam) according to the manufacturer's instructions. Briefly, A2780 cells were seeded in 6-well plates (1.0×10^6 cells per well), pre-incubated for 24 h in drug-free media at 310 K, after which they were exposed to complex 3 at equipotent concentration equal to IC₅₀ and 3 times IC₅₀ values. Cells were harvested using trypsin and stained in the dark using CytoPainter Red (Ex/Em 583/603 nm) which reacts with cell surface amines in compromised membranes. After staining, cell pellets were analysed in a Becton Dickinson FACScan Flow Cytometer.

Syntheses

Preparation of H₂PrCh and H₂PrEt. The ligands were synthesized by refluxing a solution of 1-pyrenecarboxaldehyde (1 mmol) and the desired thiosemicarbazide (1 mmol) in ethanol (15 mL) with the addition of HCl for 1.5 h at 333 K, as described previously.²⁵ The purity of the free ligands was confirmed by ¹H-NMR.

Synthesis of the tetranuclear palladium and platinum complexes

Preparation of [M₄(μ -S-PrR- κ^3 -C,N,S)₄]. Two synthetic routes were explored to obtain the tetranuclear palladium complexes. Both methods A and B worked well. However, method B gave better yields.

Method A. H₂PrR (0.1 mmol) was added to a solution of *trans*-[PdCl₂(MeCN)₂] (0.1 mmol) in 3 mL of MeCN. The mixture was stirred for 24 h at room temperature. An orange precipitate was formed during this time, which was filtered off, washed with hexane and dried in a vacuum. Afterwards, MeCN (2 mL) and Et₃N (0.5 mL) was added to the solid and the resulting mixture was kept under stirring and reflux for 5 h. The solvent was removed *in vacuo* and the residue was washed with methanol and then filtered. Afterwards the red solid was recrystallized from a mixture of CHCl₃/MeOH (1 : 1 v/v), affording a crystalline powder.

Method B. To a mixture of 0.1 mmol of *trans*-[PdCl₂(MeCN)₂] and 0.1 mmol of 1-pyrenecarboxaldehyde,



R-thiosemicarbazone in MeCN (5 mL) 0.5 mL of Et₃N was added; the mixture was then heated at 358 K for 8 h. A crystalline powder was obtained as described for method A.

[Pd₄(μ-S-PrEt-κ³-C,N,S)₄](1). Colour: Red. Yield: 75% mg (33 mg). M.P.: decomposes at 311 °C. IR (ν_{max}/cm⁻¹): 3398 ν(N-H), 1560, 1517 ν(C=N) + ν(C=C), 715 ν(C-S). Anal. Calcd for C₂₀H₁₅N₃PdS (435.83 g mol⁻¹): C, 55.12; H, 3.47; N, 9.64%. Found: C, 55.08; H, 3.37; N, 9.52%. UV-Vis, CH₂Cl₂ solution concentration: 5.1 × 10⁻⁵ M [λ_{max} (ε, L mol⁻¹ cm⁻¹): 279.00 nm (11 710), 352.50 nm (7015), 438.00 nm (6719), 491.50 nm (5445). MS (ESI⁺): *m/z* for C₈₀H₆₀N₁₂Pd₄S₄[M]⁺: calcd 1743.0106, found 1743.0109. Molar conductivity (1 × 10⁻³ mol L⁻¹ in DMSO): 0.02 μS cm⁻¹. HPLC: *R*_t = 28.18 min (at 254 nm).

[Pd₄(μ-S-PrCh-κ³-C,N,S)₄](2). Colour: Red. Yield: 81% mg (40 mg). M.P.: decomposes at 290 °C. IR (ν_{max}/cm⁻¹): 3392 ν(N-H), 1575, 1562, 1517 ν(C=N) + ν(C=C), 715 ν(C-S). Anal. Calcd for C₂₄H₂₁N₃PdS (489.92 g mol⁻¹): C, 58.84; H, 4.32; N, 8.58%. Found: C, 58.06; H, 4.26; N, 8.57%. UV-Vis, CH₂Cl₂ solution concentration: 1.12 × 10⁻⁵ M [λ_{max} (ε, L mol⁻¹ cm⁻¹): 280.50 nm (26 714), 353.50 nm (17 232), 439.00 nm (15 357), 495.00 nm (2857). MS (ESI⁺): *m/z* for C₉₆H₈₄N₁₂Pd₄S₄[M]⁺: calcd 1959.2091, found 1959.1927. Molar conductivity (1 × 10⁻³ mol L⁻¹ in DMSO): 0.004 μS cm⁻¹. HPLC: *R*_t = 33.03 min (at 254 nm).

Preparation of [Pt₄(μ-S-PrR-κ³-C,N,S)₄]. The synthetic routes explored for the platinum tetranuclear complex are described below. Method C was the best since it provided higher purity and the formation of side products was not observed.

Method A. To a flask containing 0.1 mmol of [PtCl₂(COD)] dissolved in 2 mL of acetonitrile was added 0.1 mmol of the desired ligand H₂PrR. Et₃N (0.5 mL) and then the solution was refluxed for 8 h. After cooling to room temperature, a precipitate was formed which was filtered off, washed with methanol, hexane and dried under vacuum.

Method B. 0.1 mmol of H₂PrR dissolved in hot ethanol was added to a solution of the metal precursor K₂[PtCl₄] (0.1 mmol) in 1 mL of water. The formation of an orange coloured precipitate was observed. The reaction was kept under constant stirring at room temperature for 24 h in the absence of base. The precipitate was filtered, washed with methanol, hexane and dried under vacuum.

Method C. To a solution containing 0.1 mmol of K₂[PtCl₄] dissolved in 1 mL of water, a solution containing one mol equiv. H₂PrR in 3 mL of DCM/MeCN (1 : 2) mixture was slowly added. Thereafter, 0.5 mL of triethylamine was added. The reaction mixture became clear and a red solid formed. This solution was kept under constant stirring and under reflux for 24 h. Upon slow evaporation of the solvent at room temperature a microcrystalline solid was obtained. The red solid was filtered, washed with methanol and n-hexane, and dried under vacuum. The powder obtained was recrystallized from a mixture of chloroform and acetone (1 : 1 v/v).

[Pt₄(μ-S-PrEt-κ³-C,N,S)₄](3). Colour: Red. Yield: 57% mg (30 mg). M.P.: 337–339 °C. IR (ν_{max}/cm⁻¹): 3398 ν(N-H), 1560, 1517 ν(C=N) + ν(C=C), 713 ν(C-S). Anal. Calcd for

C₂₀H₁₅N₃PtS (524.50 g mol⁻¹): 45.80; H, 2.88; N, 8.01%. Found: C, 45.29; H, 2.91; N, 8.04%. UV-Vis, CH₂Cl₂ solution concentration: 5.95 × 10⁻⁵ M [λ_{max} (ε, L mol⁻¹ cm⁻¹): 281.00 nm (4638), 383.00 nm (4184), 436.00 (3411), 465.00 (3058). MS (ESI⁺): *m/z* for C₂₀H₁₅N₃PtS [M]⁺: calcd 2096.25, found 2098.76. Molar conductivity (1 × 10⁻³ mol L⁻¹ in DMSO): 0.003 μS cm⁻¹. ¹⁹⁵Pt NMR (CDCl₃): δ -3801.88 and -3815.21 ppm.

[Pt₄(μ-S-PrCh-κ³-C,N,S)₄](CH₃)₂COCHCl₃ (4). Colour: Red. Yield: 91% mg (53 mg). M.P.: 305–307 °C. IR (ν_{max}/cm⁻¹): 3392 ν(N-H), 1577, 1562, 1517 ν(C=N) + ν(C=C), 715 ν(C-S). Anal. Calcd for C₂₄H₂₁N₃PtS (578.59 g mol⁻¹): 54.10; H, 3.11; N, 9.96%. Found: C, 54.29; H, 3.17; N, 9.84%. UV-Vis, CH₂Cl₂ solution concentration: 6.04 × 10⁻⁵ M [λ_{max} (ε, L mol⁻¹ cm⁻¹): 287.00 nm (13 476), 386.00 nm (12 466), 409.00 (12 135). MS (ESI⁺): *m/z* for C₉₆H₈₄N₁₂Pt₄S₄ [M + H]⁺: calcd 2313.4430, found 2313.4262. Molar conductivity (1 × 10⁻³ mol L⁻¹ in DMSO): 0.009 μS cm⁻¹. ¹⁹⁵Pt NMR (CD₂Cl₂): δ -4053.34 and -4080.67 ppm.

X-ray crystallography

Single crystals of C₁₀₃H₉₅Cl₃N₁₂O₂Pt₄S₄ (4) were grown from acetone/chloroform solution. A suitable crystal was selected and mounted on a glass fibre with Fromblin oil and placed on a Bruker APEX-II diffractometer with a CCD area detector. The crystal was kept at 200(2) K during data collection. Using Olex2,²⁶ the structure was solved with the ShelXT²⁷ structure solution program using Intrinsic Phasing and refined with the ShelXL²⁸ refinement package using Least Squares minimisation. Besides tetrameric complex, the asymmetric unit contained two molecules of solvent, acetone and chloroform. There was further diffuse electron density and attempts were made to model the disordered solvent, but little progress was made using several partially occupied chloroform molecules refined with many restraints. Better progress was made using the Squeeze routine in Platon. Voids located by the Squeeze routine are shown in ESI.† Table 1 presents more detailed information about the structure determinations. The structural data have been deposited at the Cambridge Crystallographic Data Centre under the accession number CCDC 1990070.†

Results and discussion

Synthesis and characterisation

The reaction of 1-pyrenecarboxaldehyde with ethyl or cyclohexyl-thiosemicarbazide in the presence of a catalytic amount of HCl in methanol afforded yellow precipitates of composition H₂PrR, which were isolated in good yields, as described for a procedure reported earlier.^{25,29} The purity and identities of the free thiosemicarbazones were confirmed by IR, ¹H-NMR, ¹³C NMR, MS, HPLC and UV-vis. The ¹³C NMR spectra for the uncoordinated thiosemicarbazones had peaks in the expected regions, with those for C=N and C=S at *ca.* 140 ppm and 176 ppm, respectively, as observed for similar structures.³⁰



Table 1 Crystal data and structure refinement for [Pt₄(μ-S-PrCh-κ³-C, N, S)₄](CH₃)₂COCHCl₃ (4)

Empirical formula	C ₁₀₃ H ₉₅ Cl ₃ N ₁₂ O ₂ Pt ₄ S ₄
Formula weight	2547.85
Temperature/K	200(2)
Crystal system	Monoclinic
Space group	P2 ₁ /n
a/Å	18.8631(8)
b/Å	26.4584(12)
c/Å	21.3233(8)
α/°	90
β/°	91.045(3)
γ/°	90
V/Å ³	10 640.4(8)
Z	4
ρ _{calc} /g cm ⁻³	1.590
μ/mm ⁻¹	5.447
F(000)	4960.0
Crystal size/mm ³	0.2 × 0.15 × 0.15 red block
Radiation	MoK _α (λ = 0.71073)
2θ range for data collection/°	2.454 to 52.886
Index ranges	−22 ≤ h ≤ 23, −30 ≤ k ≤ 33, −26 ≤ l ≤ 26
Reflections collected	87 773
Independent reflections	21 672 [R _{int} = 0.1224, R _{sigma} = 0.1492]
Data/restraints/parameters	21 672/216/1181
Goodness-of-fit on F ²	1.024
Final R indexes [I ≥ 2σ(I)]	R ₁ = 0.0657, wR ₂ = 0.1458
Final R indexes [all data]	R ₁ = 0.1516, wR ₂ = 0.1768
Largest diff. peak/hole/e Å ⁻³	1.32/−1.46

Reactions of *trans*-[PdCl₂(MeCN)₂] or K₂[PtCl₄] with H₂PrR in the presence of trimethylamine in MeCN under reflux afforded cyclometallated palladium and platinum complexes (Scheme 1). Two methods were used to prepare the palladium tetramers. For method A, described in the Experimental section, a dimeric product was isolated. The structure of the palladium dimers was confirmed by high resolution mass spectrometry (example in Fig. S7†). In the second method, however, the dinuclear complex was generated *in situ*, providing higher yields. For the preparation of the platinum tetranuclear

complex, three methods were investigated, with method C being the most efficient, providing higher purity with no side products generated. The platinum dinuclear intermediate complexes could not be isolated, but they were generated *in situ* as described before.³¹ Generally, dinuclear complexes are used as intermediates in orthometallation reactions, representing the first step for the cyclometallation.³² However, since the aim was to obtain tetranuclear complexes, the isolation of such dimers was not necessary.

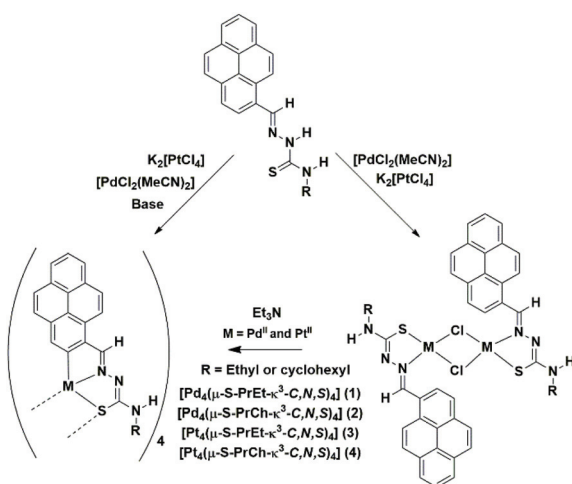
The IR spectra of the free ligands H₂PrEt and H₂PrCh show two absorptions for the ν(NH) stretching mode, while all the complexes show only one band (see Experimental section). This suggests that the ligands are deprotonated and coordinated in the thiolate form in the four complexes. Furthermore, the ν(C=N) absorption band at 1537 cm⁻¹ for H₂PrEt, shifts to lower frequencies upon complexation, being found at 1560 cm⁻¹ in the spectra of both complexes 1 and 3. This suggests coordination to the metal ion through the iminic nitrogen. The same bathochromic shift was observed for the H₂PrCh derived complexes after complexation, a *ca.* 30 cm⁻¹ shift.

The ¹H NMR spectrum of complex 3, recorded in CDCl₃ solution, shows two sets of signals, suggesting that the tetranuclear complex possesses two units in different environments. The ¹H-NMR spectrum shows two sets of peaks in the aromatic region from the four pyrene units and two sets of aliphatic hydrogen atoms related to the ethyl group from the thiosemicarbazone ligand. The spectra of the other complexes show a similar pattern. The formation of two different geometric isomers in solution probably arises from the flexibility of the eight-membered ring of alternating M-S atoms.³¹

The four metallocyclic complexes were analysed by nano-electrospray-ionisation MS (nESI-MS), after dissolution of a few crystals in a mixture DCM/MeOH (25/75 v/v) + 0.3% formic acid. Without the presence of acid, ionisation was poor. Both complexes 1 and 2 showed significant peaks for protonated species at *m/z* 1743.0109 and 1959.1927, respectively, consistent with the corresponding proposed molecular ion peaks of the palladium tetranuclear complexes (Fig. S8 and S9†). On the other hand, the solvent + acid system did not work well for the platinum tetranuclear complexes. It was possible to observe peaks for protonated complexes 3 and 4 at *m/z* 2098.76 and 2313.42 respectively, in accord with the formation of the platinum tetramers; however, other species were generated by this method (see Fig. S11,† as an example).

In order to verify the purity of the platinum complexes, we used ¹⁹⁵Pt NMR spectroscopy, since each fragment shows only one characteristic signal.³³ The ¹⁹⁵Pt NMR spectra of the complexes 3 and 4 were recorded in CDCl₃ and CD₂Cl₂, respectively, at ambient temperature. Two broad signals at −3801.9 and −3815.2 ppm were found for 3, and at −4053.3 and −4080.7 ppm for 4 (Fig. S12†), as expected from tetrameric species. The chemical shifts for square planar Pt(II) complexes are consistent with a PtCNS₂ coordination sphere (δ(¹⁹⁵Pt) −4000 to −3000 ppm), as reported in the literature.³⁴

The electronic absorption spectra of the tetranuclear complexes in CH₂Cl₂ solution at room temperature display high-

**Scheme 1** Synthesis route to the preparation of the tetranuclear complexes.

energy bands at $\lambda \approx 279\text{--}287\text{ nm}$, with extinction coefficients (ϵ) of 1×10^3 to $1 \times 10^4\text{ L mol}^{-1}\text{ cm}^{-1}$, and low-energy absorption bands at $\lambda = 352\text{--}495\text{ nm}$, with extinction coefficients of $1 \times 10^3\text{ L mol}^{-1}\text{ cm}^{-1}$. The high-energy absorption bands are assigned to the spin-allowed intraligand IL ($\pi \rightarrow \pi^*$) transitions of the thiosemicarbazone ligands.³⁵ The low-energy absorption bands of the tetranuclear complexes may be metal-to-ligand charge-transfer (MLCT) transitions [$d\pi(\text{Pt-Pd}) \rightarrow \pi^*(\text{TSC})$],³⁶ as observed previously for other organometallic complexes.³⁷

The luminescence properties of the complexes were determined for DMSO solutions at ambient temperature, since only very weak emission bands were observed in CH_2Cl_2 . The maximum emission for the complexes is blue-shifted by $\sim 270\text{ nm}$ when compared to their free ligand absorption spectra. The palladium complexes show green emission around 500 nm , while the platinum complexes exhibited two emission maxima around 450 nm . Based on such large Stokes shifts, these emissions are indicative of phosphorescence.³⁸ The large spin-orbit coupling caused by the palladium/platinum centre, indicates efficient spin-orbit coupling, often a prerequisite for phosphorescence of organometallic complexes.³⁹ Furthermore, the presence of a fluorophore can increase the luminescence properties of the complex.⁴⁰ The emission energy of the tetranuclear complexes is not significantly influenced by the R groups of the thiosemicarbazone ligands, since both ethyl and cyclohexyl substituents are electron-donors.

Reverse phase HPLC studies confirmed the purity of the tetranuclear palladium complexes. The chromatograms obtained show only one peak for complexes **1** and **2**. Complex **1** had a retention time of 28.18 min , while the cyclohexyl derivative (complex **2**) had a retention time of 33.03 min . From the retention times, it can be assumed that the cyclohexyl derivative is more lipophilic than the ethyl derivative. As indicated by the mass spectra, the platinum complexes are not stable in the acid system used for HPLC studies.

Suitable crystals of $[\text{Pt}_4(\mu\text{-S-PrCh-}\kappa^3\text{-C,N,S})_4](\text{CH}_3)_2\text{COCHCl}_3$ (**4**) were obtained by slow evaporation of a solution of complex **4** and were studied by single crystal X-ray crystallography. A perspective view of the molecule is shown in Fig. 2. Selected bond distances are summarized in Tables S1 and S2,[†] respectively. The asymmetric unit contains four tetrameric Pt complexes, and also two molecules of solvent acetone and a chloroform molecule. There was further diffuse electron density and attempts were made to model the disordered solvent, but little progress was made using several partially occupied chloroform molecules refined with many restraints. Better progress was made using the Squeeze routine in Platon. Voids located by the Squeeze routine are shown in detail in the ESI.[†]

The X-ray structure shows the tetranuclear complex $[\text{Pt}_4(\mu\text{-S-PrCh-}\kappa^3\text{-C,N,S})_4](\text{CH}_3)_2\text{COCHCl}_3$ is composed of four metallated monomers held together by bridging sulfur atoms. In this way, each platinum centre possesses a coordination sphere of donor atoms from the thiosemicarbazone ligand, the imine nitrogen, a sulfur atom and an *ortho* carbon from the benzyl ring, while the fourth position is occupied by a sulfur

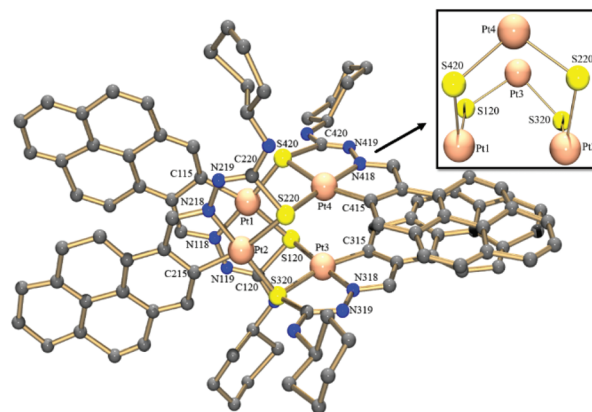


Fig. 2 Molecular structure of complex **4**. The solvent molecules and hydrogen atoms have been omitted for clarity.

atom from a neighboring monomer unit, resulting in a *CNSS* tetradentate coordination mode. A predominantly double bond is found for $\text{C}=\text{S}$ in the free ligand ($1.685(3)\text{ \AA}$), while in complex **4** this bond is longer, $1.820(14)\text{ \AA}$. The elongation of the C-S bond in the dianionic ligand is due to the deprotonation of the imine nitrogen followed by the thioenolization of the CS bond. Pt-N bond distances of $\sim 2.00\text{ \AA}$ and Pt-C of $\sim 2.03\text{ \AA}$ in the five-membered orthometallo cycle are analogous to those found in other similar complexes.⁴¹ The *trans* influence of the metallated carbon is reflected in the lengthening of the Pt-S distance *trans* to this carbon atom, $\text{Pt-S}_{\text{chelating}}$ (*ca.* 2.35 \AA), with respect to the Pt-S distance *trans* to the imine nitrogen, $\text{Pt-S}_{\text{bridging}}$ (*ca.* 2.03 \AA). This feature has also been observed for other polynuclear complexes with similar ligands.¹⁹ Therefore, the two distinct Pt-sulfur bonds, $\text{Pt-S}_{\text{chelating}}$ and $\text{Pt-S}_{\text{bridging}}$, show that the sulfur bridge is relatively strong. The strong $\text{M-S}_{\text{bridging}}$ bond was also apparent from treatment of the tetranuclear compounds with coordinating solvents such as DMSO, DMF and acetonitrile, which did not lead to opening of the eight-membered Pt_4S_4 ring core of the tetranuclear structure, nor to ring cleavage of the chelate. However, some papers have described the $\text{M-S}_{\text{bridging}}$ ($\text{M} = \text{Pt}$ or Pd) bond cleavage in reactions of orthometallated TSC tetranuclear complexes with strongly binding phosphine ligands.⁴² The two 5-membered chelate rings are planar to each other, this planarity being extended to the pyrene fragment. Moreover, the connections between the platinum ions and the bridging sulfur atoms generate a cavity in the centre of the structure (Fig. 3). The Pt_4S_4 core consists of an eight-membered ring of alternating Pt(II) and S atoms in a boat conformation. The size of this cavity can be assessed by the distance between two platinum centres of parallel monomers: a Pt-Pt distance of *ca.* 3.37 \AA . More inter-metal distances are listed in Table S3.[†] This Pt-Pt distance indicates little interaction between the platinum centres. This is in agreement with the reported values for tetranuclear Pt_4 complexes ($2.54\text{--}3.51\text{ \AA}$).⁴³ The geometry around the Pt ion is distorted square-planar, in accord with the bond angles C115-Pt1-S120 [$166.2(4)^\circ$], C315-Pt1-S120 [$166.2(4)^\circ$], C115-Pt1-S320 [$166.2(4)^\circ$], C315-Pt1-S320 [$166.2(4)^\circ$], C115-Pt1-S420 [$166.2(4)^\circ$], C315-Pt1-S420 [$166.2(4)^\circ$], C115-Pt1-S220 [$166.2(4)^\circ$], C315-Pt1-S220 [$166.2(4)^\circ$], C115-Pt1-S120 [$166.2(4)^\circ$], C315-Pt1-S120 [$166.2(4)^\circ$], C115-Pt1-S320 [$166.2(4)^\circ$], C315-Pt1-S320 [$166.2(4)^\circ$], C115-Pt1-S420 [$166.2(4)^\circ$], C315-Pt1-S420 [$166.2(4)^\circ$], C115-Pt1-S220 [$166.2(4)^\circ$], C315-Pt1-S220 [$166.2(4)^\circ$], C115-Pt1-S120 [$166.2(4)^\circ$], C315-Pt1-S120 [$166.2(4)^\circ$], C115-Pt1-S320 [$166.2(4)^\circ$], C315-Pt1-S320 [$166.2(4)^\circ$], C115-Pt1-S420 [$166.2(4)^\circ$], C315-Pt1-S420 [$166.2(4)^\circ$], C115-Pt1-S220 [$166.2(4)^\circ$], C315-Pt1-S220 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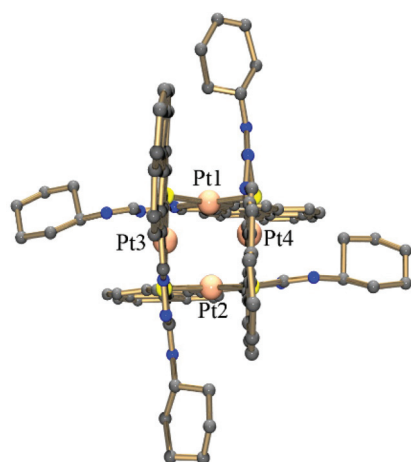


Fig. 3 View of the tetramer 4 showing the central cavity.

Pt3–S320 [164.4(4)°], C215–Pt2–S220 [164.8(4)°], C415–Pt4–S420 [164.0(4)°]. Moreover, the two ^{195}Pt NMR peaks are in agreement with the X-ray structure, which shows two pairs of Pt ions in different magnetic environments because of the conformation of the macrocyclic ring.¹⁶

The crystal structure is stabilized by intermolecular hydrogen bonds involving the NH groups which have short contacts to acetone solvent molecules. Distances and angles related to hydrogen bonds are summarized in Table S4.† There is possible π stacking,⁴⁴ but with only a very slight overlap between the pyrene rings. Their orientation may be dominated by the requirements of coordination.

Metal-based compounds are sometimes unstable in solution. For this reason, it is important to ensure that the tested compound is stable in aqueous solution. Therefore, as a model system, the stability of complex 1 was investigated and monitored by UV-vis spectra in H_2O with 5% of DMSO for at least 24 h period at 25 °C (see Fig. S13, ESI†). The spectra remained unchanged over the time for this assay, suggesting that compounds are stable in solution.

Antiproliferative activity and cellular uptake

The inhibition of cell proliferation in ovarian cancer cell A2780 line by the tetranuclear complexes was evaluated using the SRB method,²⁴ Table 2. For comparison purposes, the anticancer drug cisplatin was also assayed under the same experimental conditions. The formation of the cyclometallated complexes resulted in an enhancement of activity against A2780 human ovarian cancer cells for complexes 1 and 3 in comparison with the free ligand H_2PrEt ($\text{IC}_{50} > 50 \mu\text{M}$), whereas the cyclohexyl derivative complexes exhibited lower potency against the cancer cells. Comparing the two analogous complexes 1 (Pd_4) and 3 (Pt_4), it is notable that the simple exchange of palladium for platinum enhances the activity almost four times. The structural modification in the thiosemicarbazone ligand periphery from cyclohexyl to ethyl group also has a significant effect on the anticancer activity, with the

Table 2 Comparison of IC_{50} values (μM) for the thiosemicarbazone ligands, tetranuclear Pd and Pt complexes and cisplatin against A2780 cancer cells and MRC5 normal cells. Experiments included 24 h of drug exposure and 72 h of recovery time in drug-free medium. Cell viability results, determined by the SRB assay, are expressed as the average of duplicates of triplicates and together with standard deviations

	IC_{50}^a (μM)	
	A2780	MRC5
H_2PrEt	>50	—
H_2PrCh	>50	—
1	1.27 ± 0.38	>100
2	45 ± 2	—
3	0.37 ± 0.11	>100
4	28.1 ± 0.9	—
CDDP	1.20 ± 0.03	10.9 ± 0.3

^a IC_{50} concentration based on the molecular weights of the tetranuclear complexes.

cyclohexyl derivative being much less active in the A2780 cell line. Since the potency of the antiproliferative activity of the ethyl complexes 1 and 3 was the highest, these were then screened against the non-tumourigenic cell line MRC5, human embryonal lung fibroblasts. These two complexes exhibited much higher IC_{50} values in MRC5 human fibroblasts (>100 μM , Table 2) compared to A2780 cells. This cell line is used routinely as an indicator to investigate differences in toxicity between cancerous and non-cancerous cells in the early-stage development of anticancer complexes and as a preliminary predictor for cancer cell selectivity. In this case, the values obtained are highly promising as there are at least two and three orders of magnitude difference (for complexes 1 and 3 respectively) between the potency in the MRC5 and the A2780 lines.

The cellular accumulation of palladium or platinum from 1 or 3, respectively, using equipotent IC_{50} concentrations was determined in A2780 ovarian cells in order to correlate the amount of metal accumulated with antiproliferative activity and lipophilicity. For these experiments, drug exposure time was 24 h and cells were not allowed to recover. Values are expressed as ng of Pd and Pt per million cells and were determined as independent duplicates of triplicates. The cellular accumulation of Pd for complex 1 was 6.0 ± 0.4 ng of Pd per 10^6 cells, while for complex 3 was 3.4 ± 0.2 ng of Pt per 10^6 cells, which is equivalent to a molar uptake of 3.44 and 1.62 μmol respectively, this taking into account the molecular weight of the tetramer units for complexes 1 and 3 (Table 3).

Table 3 Cellular accumulation (ng) of Pd or Pt from complexes 1 and 3 in A2780 ovarian cancer cells treated with equipotent IC_{50} concentrations for 24 h with no recovery time in drug-free medium

	ng Pt/Pd $\times 10^6$ cells
1	6.0 ± 0.4
3	3.4 ± 0.2
CDDP	0.51 ± 0.07



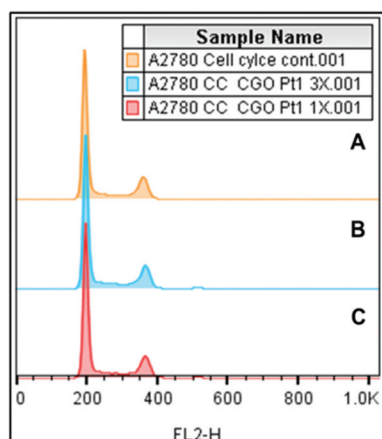


Fig. 4 FL2-H histograms obtained by flow cytometry for cell cycle analysis (A) control untreated cells; (B) cells treated using 3 times IC_{50} concentration of complex **3**; (C) cells treated using $0.4 \mu M$ of complex **3**. Experiments included 24 h drug exposure time.

Due to the potent cytotoxicity of complex **3**, we investigated its effects on the cell cycle (Fig. 4). Complex **3** was incubated with A2780 ovarian cancer cells for 24 h. For this study, cells were stained using propidium iodide (PI) as a fluorescent probe that interacts with DNA through intercalation. In comparison to the control population, data shown in Table S5 (ESI[†]) for complex **3** at two different concentrations, it is notable that there is no significant difference in the population of G1, G2/M and S phases after 24 h of exposure to the platinum complex (e.g., from $63.5 \pm 1.0\%$ for the control *versus* $61.8 \pm 0.6\%$ for G1 at the highest concentration tested). These results contrast dramatically with those for CDDP, which causes cell cycle arrest in the S phase given its DNA-based mechanism of action.⁴⁵

One cellular response to stress stimuli is programmed cell death (apoptosis). The possibility that treatment of A2780 cancer cells with the platinum tetranuclear complex **3** leads to apoptosis was investigated. Annexin V-fluorescein isothiocyanate (FITC) and propidium iodide (PI) dual-staining allowed the detection of four different cellular populations: viable cells, non-viable cells and early-stage, and late-stage apoptosis. Early-stage apoptosis is characterised by changes in the symmetry of the phospholipid membrane, and late-stage apoptosis by further disruption of the integrity of the cell membrane so it becomes permeable to PI. Staurosporine, a potent apoptosis-inducer, was used as a positive control.⁴⁶ After 24 h of drug exposure, the extent of non-viable, early and late apoptotic cells is close to zero. No population of cells in either of these two stages of apoptosis was detected (Fig. 5). The compound does not affect any of the apoptotic populations, showing only viable cells (see Table S2[†]).

In the absence of apoptosis after 24 h of drug exposure, we investigated the cellular membrane integrity by flow cytometry. A2780 cells exposed to complex **3** at two different concentrations ($1 \times IC_{50}$ concentration and $3 \times IC_{50}$ concentration) tested negative for increased fluorescence of the 7-ADD stain

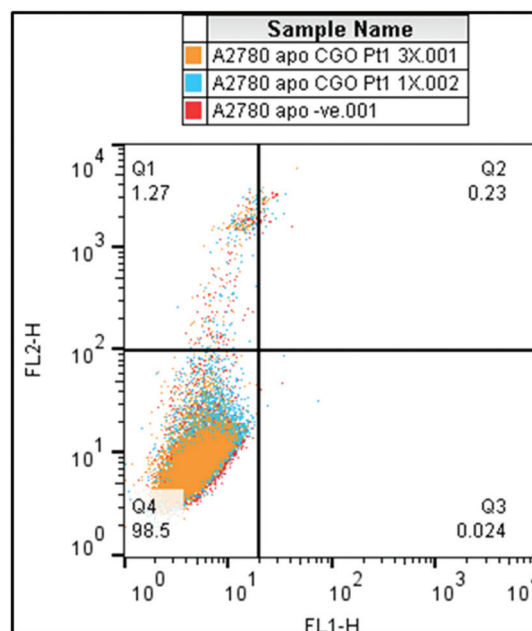


Fig. 5 FL2-H vs FL1-H dot plots obtained by flow cytometry for apoptosis analysis using FITC (FL1-H channel) and PI (FL2-H channel) staining. (■) cells treated using $3 \times IC_{50}$ concentration of complex **3**; (■) Cells treated using $0.4 \mu M$ of complex **3**. (■) Control untreated cells. For all experiments, drug exposure time was 24 h, cells were treated with RNase and stained using PI.

in the FL2-red channel, indicating that, under these experimental conditions, there is no cellular membrane integrity damage (Fig. 6). However, metal accumulation studies in A2780 cells indicated that complex **3** does accumulate in the cancer cells. We also attempted to investigate the interaction of complexes **1** and **3** with DNA (circular and linear dichroism) and with GSH. However, due to their poor solubility in buffer, it was not possible to perform these experiments despite many

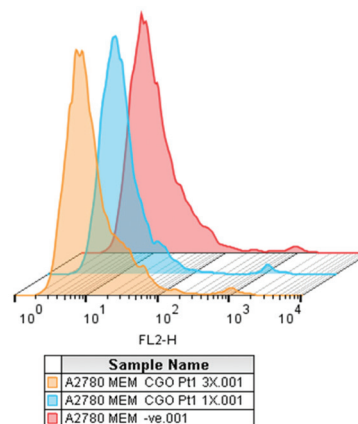


Fig. 6 Membrane integrity assessed by flow cytometry for A2780 cancer cells (red) and after treatment with complex **3** at $1 \times IC_{50}$ concentration (blue) and $3 \times IC_{50}$ concentration (orange) at 310 K. Viable cell membrane (FL2-). Non-viable cell membrane (FL2+). No changes were observed after 24 h drug exposure.



attempts at different concentrations of DMSO and water, but in all the cases precipitation of the compounds was observed.

Conclusions

Four tetrameric palladium(II) and platinum(II) complexes $[M_4(\mu-S-PrR-\kappa^3-C,N,S)_4]$ containing tridentate thiosemicarbazones (H_2PrR) derived from monocyclic aromatic aldehydes and thiosemicarbazides have been synthesised by *ortho*-metalation reactions *via* C–H activation of a pyrene ring substituent. These new cyclometallated complexes were fully characterised using various analytical techniques and also for one Pt_4 complex by X-ray crystallography. The crystal structure confirmed that the thiosemicarbazone ligands undergo cyclometallation reactions to give tetranuclear compounds with a central core consisting of an eight-membered ring of alternating platinum and sulfur atoms. The R = ethyl PrR thiosemicarbazone complexes are potentially cytotoxic to human ovarian cancer cells being as active (Pd , 1) or an order of magnitude more active (Pt , 3) than the clinical drug cisplatin. Moreover, the tetranuclear R = ethyl complexes appear more selective for cancer cells *versus* non-tumourigenic cells than cisplatin. In contrast to cisplatin, these complexes do not induce apoptosis, nor cause cell membrane damage. Hence, the difference in mechanism of action may provide a basis for combatting cisplatin resistance which is a major current problem and causes an unmet clinical need.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

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