






Multi-modal control over the assembly of a molecular motor bola-amphiphile in water†

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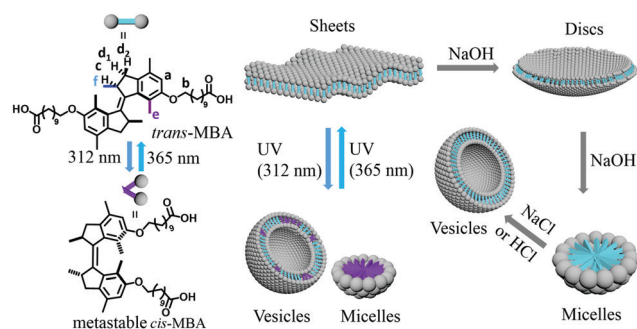
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We report multi-modal-control over the assembly behaviour of a first-generation molecular motor bola-amphiphile in water by light, pH and the choice of counter-ions. These findings open up opportunities for the development of materials that reconfigure enabling complex functions in response to different stimuli.

The variety of noncovalent interactions between assembled units found in nature provides a basis for designing artificial supramolecular systems.¹ The intrinsic dynamics, tunability of properties and stimuli responsiveness of synthetic supramolecular systems is controlled by the precise design of organic molecules,² mesogenic materials,³ polymers,⁴ and a variety of organic–inorganic hybrid composites,⁵ allowing modulation of, for example, assembly and functionality.⁶ Recent advances in supramolecular chemistry and soft materials design have enabled the creation of various supramolecular systems responsive to external stimuli,⁷ especially in aqueous media.^{1a,h,2a,6d} Encoding specific responses to different stimuli into self-assembling systems is a promising approach to achieve more complex dynamic and adaptive behaviour.⁸ Besides making use of multi-stimuli responsive polymers^{4a,8} or responsive organic–inorganic composites,^{5,9} it has also been demonstrated that supramolecular assemblies of small organic molecules in water can be controlled independently by external stimuli like light,¹⁰ pH,¹¹ and ions.¹² Notably, a few of these systems can be controlled by various external stimuli simultaneously.¹³ For example, the supramolecular assemblies of polyanionic dendritic peptide amphiphiles, reported by Besenius *et al.*,^{13b} were controlled systematically by pH and ionic strength. Among the various stimuli, light offers a non-invasive way with high spatial

as well as temporal precision for manipulating the structures of molecular assemblies.^{6c,14} However, implementing responsiveness to other stimuli into photo-responsive supramolecular systems based on organic molecules^{13c} to realize multi-modal control over their assembly in water remains highly challenging.

Taking advantage of rotary motors as light-driven multistate switches, we have previously developed a co-assembling supramolecular system, composed of a second-generation molecular motor and a lipid (1,2-dioleoyl-*sn*-glycero-3-phosphocholine), allowing upon irradiation,¹⁵ transformations from nanotubes into vesicles and multi-lamellar vesicles (in a fully reversible cycle) whereby comparable packing parameters were identified for both structures ($1/2 < P < 1$).¹⁶ In the present study, multi-modal control over the self-assembly morphology of a first-generation molecular motor is reported using light, pH and ions as triggering elements. This class of molecular motors offers advantages due to the distinct geometric differences between *cis*- and *trans*-isomer, leading to more pronounced changes of the packing parameter upon irradiation, and the absence of thermal back-isomerization. In the present study, the morphology of the molecular motor bola-amphiphile (**MBA**) aggregates could be switched between sheet-like structures and a mixture of micelles and vesicles upon irradiation (Scheme 1). By contrast, morphology transitions from sheet-like assemblies



Scheme 1 Schematic illustration of the photo-isomerization of **MBA** and multi-modal control over its assembly behaviour.

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to discs and ultimately micelles were found with increasing pH. Moreover, the addition of NaCl resulted in the formation of vesicles, while CaCl₂ led to macroscopic aggregates.

The structures of *trans*- and *cis*-isomers of **MBA** are shown in Scheme 1. Although the rotation of first-generation motors passes through four states,^{17a} the metastable *trans*-isomer is not discussed in this study due to its short half-life ($t_{1/2} < 30$ s) at room temperature.^{17b} Hence, *trans*-**MBA** in this paper refers to the stable *trans*-isomer. Stable *trans*- and metastable *cis*-isomer could be interconverted by irradiation with 312 nm and 365 nm light, respectively, while the stable *cis*-isomer could be obtained through thermal helix inversion from the metastable *cis*-isomer with a half-life of ~ 20 h.^{17a,c} The stable and metastable *cis*-isomer are expected to have the same assembly morphology, due to their related geometry. Therefore, we mainly focus on the photo-induced morphology transition between stable *trans*-**MBA** and metastable *cis*-**MBA** in the present study. Two carboxylic acid groups are connected to the motor core through alkyl-linkers whose relative orientation significantly changes upon *cis*-*trans* isomerization, potentially leading to distinct differences in self-assembly behaviour. The carboxylic acid functionality has recently also been used as the end group on a motor amphiphile forming electrostatic interactions with counter-ions, such as Ca²⁺ to allow the formation of macroscopic string micro-actuators.^{10b,18a} In the present study, we use this group as a cation binding as well as pH-responsive unit to gain additional handles for multi-modal control over **MBA**'s assembly behaviour in addition to photo-chemical *cis*-*trans* isomerization.

The synthesis and characterization of *trans*-**MBA** and stable *cis*-**MBA** are summarized in Scheme S1 (ESI[†]). The photo-responsive behaviour was studied by UV/Vis absorption and ¹H NMR spectroscopy. Irradiation of the *trans*-isomer with 312 nm light resulted in a decrease of the absorption bands at 286 nm and 312 nm with concomitant formation of a new band around 350 nm, indicating the formation of the metastable *cis*-isomer (Fig. S1a, ESI[†]). After irradiating for 18 min, no further changes were observed, meaning that the photostationary state (PSS) had been reached. Subsequent irradiation with 365 nm light induced the opposite spectral changes caused by the back-isomerization to *trans*-**MBA** (Fig. S1b, ESI[†]). An isosbestic point at 329 nm was observed during both irradiation processes, confirming the clean formation of metastable *cis*- and *trans*-isomer, respectively. As shown in the ¹H NMR spectra in Fig. 1, irradiation of a sample of *trans*-**MBA** with 312 nm light induced the formation of a new set of signals (e.g. $H_a = 6.55$ ppm, $H_e = 1.21$ ppm) belonging to the metastable *cis*-isomer. The *cis*:*trans* ratio at PSS was found to be 63:37. Subsequent irradiation at 365 nm led to full recovery of the initial spectrum, demonstrating that the *cis*-*trans* isomerization of **MBA** can be precisely controlled by light.

To investigate the critical aggregation concentration (CAC) of *trans*-**MBA** a Nile Red fluorescence assay (NRFA), which probes the internal hydrophobicity of assemblies,¹⁸ was performed. The results revealed a CAC of 4.0×10^{-6} M for *trans*-**MBA** (Fig. S2a, ESI[†]). The structure of aggregates of *trans*-**MBA**

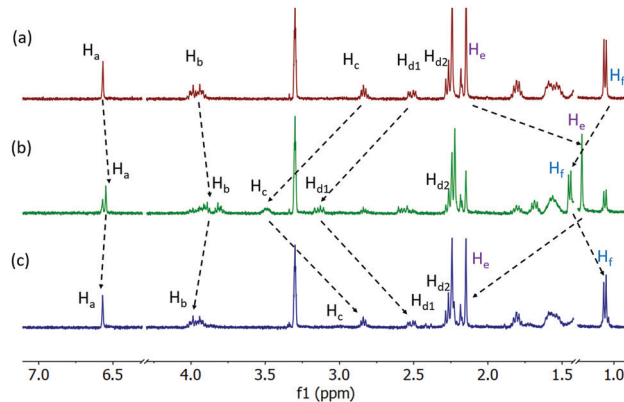


Fig. 1 ¹H NMR spectra (CD₃OD, 25 °C, 400 MHz) of *trans*-**MBA** (2.8×10^{-3} M) (a) before and (b) after irradiation to PSS with 312 nm light for 45 min at 5 °C and (c) after subsequent irradiation to PSS with 365 nm light for 15 min.

in water was imaged using cryogenic transmission electron microscopy (cryo-TEM) to accurately capture their solution-state morphology (see ESI[†]). Since self-assembly of *trans*-**MBA** is sensitive to pH and ionic strength (*vide infra*), irradiation experiments were carried out in sodium borate buffer (pH = 9.3, 0.1 M). As shown in Fig. 2a, *trans*-**MBA** was found to form sheet-like assemblies ($P \approx 1$). After irradiating with 312 nm light for 10 min to form metastable *cis*-**MBA**, vesicles with a diameter of around 20 nm and micelles with 5–6 nm diameter were observed in addition to sheet-like structures (Fig. 2b). As a comparison, we studied the aggregate morphology of stable *cis*-**MBA**, whose CAC (1.0×10^{-5} M, Fig. S2b, ESI[†]) was distinct from that of *trans*-**MBA** (4.0×10^{-6} M), implying possible different assembly morphologies between *trans*- and *cis*-isomers. Indeed, the stable *cis*-isomer showed the formation of micelles with 5–6 nm diameter ($P \leq 1/3$)

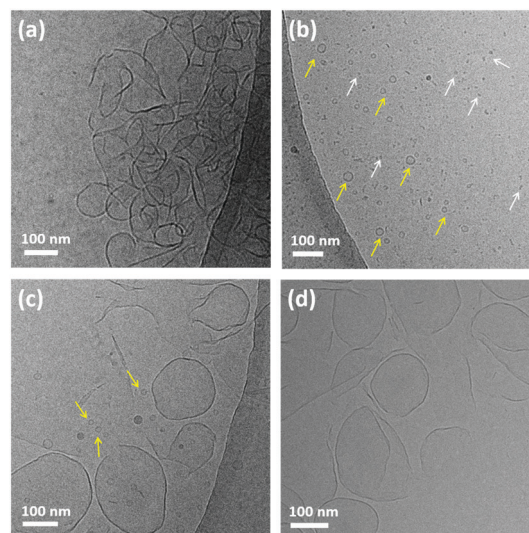


Fig. 2 Cryo-TEM images of (a) *trans*-**MBA** (7.0×10^{-3} M) in sodium borate buffer (pH = 9.3, 0.1 M) before irradiation, (b) after 312 nm irradiation for 10 min and (c) after subsequent 365 nm irradiation for 10 min (d) as well as a heating-cooling cycle. (yellow: vesicles, white: micelles).



(Fig. S4, ESI[†]). Due to its related geometry, metastable *cis*-MBA is expected to have the same assembly structure as stable *cis*-MBA. Therefore, the presence of micelles in the sample of *trans*-MBA after irradiation (mixture with metastable *cis*-MBA) can be attributed to the self-assembly of the metastable *cis* isomer. The formation of vesicles ($1/2 < P < 1$), we tentatively assigned to the co-assembly of both stable *trans*- and metastable *cis*-molecules. The remaining sheet-like assemblies (Fig. S5, ESI[†]) hint at an excess of self-assembling *trans*-MBA present in the mixture. This was supported by ¹H NMR measurement revealing a 6% share of metastable *cis*-MBA in the mixture (Fig. S7b, ESI[†]). Upon subsequent irradiation with 365 nm light for 10 min (metastable *cis*-MBA to *trans*-MBA isomerization), the original sheet-like assemblies were recovered, albeit with some small vesicles remaining in the mixture (Fig. 2c). ¹H NMR, however, revealed that there was no remaining *cis*-MBA (Fig. S7c, ESI[†]). This phenomenon had also been observed in earlier studies in our group.¹⁵ It was found that molecular motors could recover their original assembly morphologies only upon being subjected to conditions allowing them to reorganize. We therefore facilitated reorganisation by repeating the assembly preparation process (heating-cooling cycle) using the sample shown in Fig. 2c. Indeed, after this treatment, only sheet-like structures were observed (Fig. 2d).

Besides reversible control by irradiation, the assembly structure of *trans*-MBA also showed significant changes upon altering pH. As shown in Fig. 3, *trans*-MBA formed sheet-like structures ($P \approx 1$) at pH 8.8 (Fig. 3a) and disc-like structures ($1/2 < P < 1$) with a diameter of 30–40 nm and a thickness of 2–3 nm at pH 9.8 (Fig. 3b), as well as tiny micelles ($P \leq 1/3$) with a diameter of 4–5 nm at pH 11 (Fig. 3c). Therefore, as pH goes up structures with increasingly smaller packing parameters are formed, which we attribute to the increasing concentration of

deprotonated molecules at higher pH of the solution. It has been demonstrated that the head group area of long-chain fatty acids increases significantly upon dissociation of the carboxylic acid group.¹⁹ Thus, the ratio of molecules with larger head group area rises with increasing pH. Based on the equation of molecular packing parameters for amphiphiles,¹⁶ this increase of head group area leads to a decrease of the packing parameter, causing the self-assembly to change from sheets to micelles. In our case, the morphology transformations of *trans*-MBA were also in accordance with this analysis. Interestingly, when trace amounts of an aq. HCl stock solution (0.1 M) were added to the sample at pH 11 to decrease the pH to 9.4, the morphology changed from micelles to vesicles and sponge-like structures as congeries of vesicles (Fig. 3d). In this case, however, both pH as well as the concentration of NaCl were altered and the observed transformation of morphology could be the result of changing either one or both of these conditions.

In order to explore a possible new handle for control over the assembly morphologies of *trans*-MBA, the counter-ion effect of Na⁺, K⁺, Ca²⁺, and Zn²⁺ on the self-assembly structures formed by *trans*-MBA was studied by Nile Red fluorescence assay. Therefore, the blue shift of Nile Red in *trans*-MBA dicarboxylate solution (5.0×10^{-6} M) was determined in the presence of various concentrations of the chloride salts (1.0×10^{-6} – 5.0×10^{-1} M) (Fig. S3, ESI[†]). A gradual blue shift was observed with enhanced concentrations of NaCl, KCl, ZnCl₂ and CaCl₂, reflecting an increase of the internal hydrophobicity of the *trans*-MBA assemblies. Interestingly, the blue shift induced by increasing the concentration of CaCl₂ was significantly larger compared to other chloride salts, indicating that Ca²⁺ ions induce aggregate formation of *trans*-MBA more effectively.^{18a} Based on the results of NRFA, Ca²⁺ and Na⁺ were chosen for additional studies of the control of assembly morphology. Vesicles with diameters of 100–200 nm were observed in aq. 0.1 M NaCl solution (Fig. 4a) contrasting with the micelles observed in samples at high pH discussed earlier. The addition of NaCl likely reduces the repulsion between the deprotonated head groups due to charge screening.¹⁹ As a consequence, the decrease in head group area results in an increase of the packing parameter of MBA leading to the observed change in assembly structure. However, in 0.1 M aq. CaCl₂ solution *trans*-MBA formed macroscopic precipitates. In order to study the assembly structure in more detail, a sample of *trans*-MBA

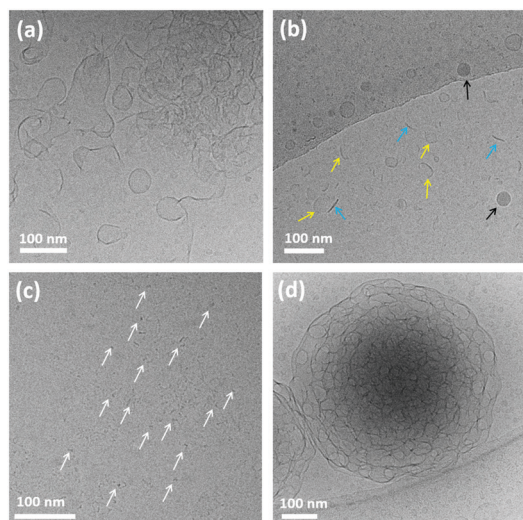


Fig. 3 Cryo-TEM images of *trans*-MBA (7.0×10^{-3} M) in aq. NaOH solution with (a) pH = 8.8, (b) pH = 9.8, (c) pH = 11 and (d) after adjusting its pH to 9.4 by adding an aq. HCl stock solution starting from pH 11. (blue: upright discs, yellow: discs at an angle with the substrate, white: micelles, black: contaminants).

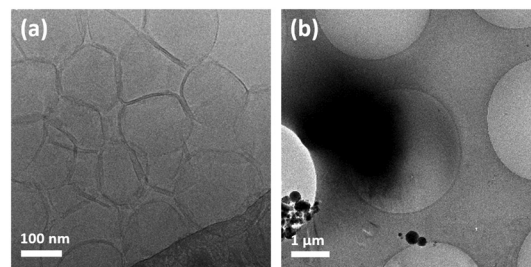


Fig. 4 Cryo-TEM images of *trans*-MBA dicarboxylate (7.0×10^{-3} M) in (a) 0.1 M NaCl solution and (b) 5.0×10^{-3} M CaCl₂ solution.



dicarboxylate at a lower CaCl_2 concentration (5.0×10^{-3} M) was characterized by cryo-TEM; micrometre-sized aggregates were observed as shown in Fig. 4b. The presence of Ca in these aggregates was confirmed by EDX analysis (Fig. S6, ESI[†]). The preferential induction of aggregate formation observed for Ca^{2+} may be related to the stronger interaction of Ca^{2+} with carboxylate groups.^{18a}

In summary, we realized multi-modal control over the assembly behaviour of a first-generation molecular motor in water. Transitions between sheet-like structures and a mixture of micelles and vesicles, indicating large changes in packing parameter, could be controlled by light in a reversible process. Dramatic changes of assembly morphology from sheet-like assemblies to discs and ultimately micelles were achieved by adjusting pH. Furthermore, the addition of NaCl led to the formation of vesicles, while CaCl_2 induced macroscopic aggregates. To the best of our knowledge, this is the first example of multi-responsive assembly of a molecular motor-based bola-amphiphile in water. This study significantly enhances our understanding of the self-assembly behaviour of multi-responsive systems in water and thereby paves the way to future applications like adaptive materials, delivery systems or supramolecular materials capable to perform various distinct tasks.

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Conflicts of interest

There are no conflicts to declare.

Notes and references

- (a) J. M. Lehn, *Supramol. Chem.*, John Wiley & Sons, Weinheim, 1995; (b) T. Aida, E. W. Meijer and S. I. Stupp, *Science*, 2012, **335**, 813–817; (c) E. Krieg, M. M. C. Bastings, P. Besenius and B. Rybtchinski, *Chem. Rev.*, 2016, **116**, 2414–2477; (d) E. Mattia and S. Otto, *Nat. Nanotechnol.*, 2015, **10**, 111–119; (e) L. Voorhaar and R. Hoogenboom, *Chem. Soc. Rev.*, 2016, **45**, 4013–4031; (f) F. Würthner, C. R. Saha-Möller, B. Fimmel, S. Ogi, P. Leowanawat and D. Schmidt, *Chem. Rev.*, 2016, **116**, 962–1052; (g) T. F. A. De Greef, M. M. J. Smulders, M. Wolffs, A. P. H. J. Schenning, R. P. Sijbesma and E. W. Meijer, *Chem. Rev.*, 2009, **109**, 5687–5754; (h) B. N. S. Thota, L. H. Uner and R. Haag, *Chem. Rev.*, 2016, **116**, 2079–2102.
- (a) L. L. K. Taylor, I. A. Riddell and M. M. J. Smulders, *Angew. Chem., Int. Ed.*, 2019, **58**, 1280–1307; (b) J. Raeburn, A. Z. Cardoso and D. J. Adams, *Chem. Soc. Rev.*, 2013, **42**, 5143–5156.
- (a) R. Eelkema and B. L. Feringa, *Org. Biomol. Chem.*, 2006, **4**, 3729–3745; (b) T. Kato, J. Uchida, T. Ichikawa and B. Soberats, *Polym. J.*, 2018, **50**, 149–166; (c) D. B. Amabilino, D. K. Smith and J. W. Steed, *Chem. Soc. Rev.*, 2017, **46**, 2404–2420.
- (a) F. D. Jochum and P. Theato, *Chem. Soc. Rev.*, 2013, **42**, 7468–7483; (b) X. Yan, F. Wang, B. Zheng and F. Huang, *Chem. Soc. Rev.*, 2012, **41**, 6042–6065.
- M. Grzelczak, L. M. Liz-Marzán and R. Klajn, *Chem. Soc. Rev.*, 2019, **48**, 1342–1361.
- (a) O. J. G. M. Goor, S. I. S. Hendrikse, P. Y. W. Dankers and E. W. Meijer, *Chem. Soc. Rev.*, 2017, **46**, 6621–6637; (b) K. Sato, M. P. Hendricks, L. C. Palmer and S. I. Stupp, *Chem. Soc. Rev.*, 2018, **47**, 7539–7551; (c) E. Busserson, Y. Ruff, E. Moulin and N. Giuseppone, *Nanoscale*, 2013, **5**, 7098–7140; (d) X. Ma and H. Tian, *Acc. Chem. Res.*, 2014, **47**, 1971–1981; (e) T. Muraoka and K. Kinbara, *Chem. Commun.*, 2016, **52**, 2667–2678; (f) C. Heinzmann, C. Weder and L. M. de Espinosa, *Chem. Soc. Rev.*, 2016, **45**, 342–358.
- (a) C. D. Jones and J. W. Steed, *Chem. Soc. Rev.*, 2016, **45**, 6546–6596; (b) B. L. Feringa and W. R. Browne, *Molecular Switches*, Wiley-VCH, Weinheim, 2011.
- J. Zhuang, M. R. Gordon, J. Ventura, L. Li and S. Thayumanavan, *Chem. Soc. Rev.*, 2013, **42**, 7421–7435.
- (a) O. Chovnik, R. Balgley, J. R. Goldman and R. Klajn, *J. Am. Chem. Soc.*, 2012, **134**, 19564–19567; (b) J. H. Schenkel, A. Samanta and B. J. Ravoo, *Adv. Mater.*, 2014, **26**, 1076–1080.
- (a) W. A. Velema, M. C. A. Stuart, W. Szymanski and B. L. Feringa, *Chem. Commun.*, 2013, **49**, 5001–5003; (b) J. Chen, F. K. C. Leung, M. C. A. Stuart, T. Kajitani, T. Fukushima, E. van der Giessen and B. L. Feringa, *Nat. Chem.*, 2018, **10**, 132–138.
- (a) T. J. Moyer, J. A. Finbloom, F. Chen, D. J. Toft, V. L. Cryns and S. I. Stupp, *J. Am. Chem. Soc.*, 2014, **136**, 14746–14752; (b) B. F. Lin, K. A. Megley, N. Viswanathan, D. V. Krogstad, L. B. Drews, M. J. Kade, Y. Qian and M. V. Tirrell, *J. Mater. Chem.*, 2012, **22**, 19447–19454.
- S. Shin, S. Lim, Y. Kim, T. Kim, T. Choi and M. Lee, *J. Am. Chem. Soc.*, 2013, **135**, 2156–2159.
- (a) H. A. M. Ardona and J. D. Tovar, *Chem. Sci.*, 2015, **6**, 1474–1484; (b) M. Von Gröning, I. de Feijter, M. C. A. Stuart, I. K. Voets and P. Besenius, *J. Mater. Chem. B*, 2013, **1**, 2008–2012; (c) F. K. C. Leung, T. Kajitani, M. C. A. Stuart, T. Fukushima and B. L. Feringa, *Angew. Chem., Int. Ed.*, 2019, **58**, 10985–10989; (d) E. Krieg, E. Shirman, H. Weissman, E. Shimoni, S. G. Wolf, I. Pinkas and B. Rybtchinski, *J. Am. Chem. Soc.*, 2009, **131**, 14365–14373.
- S. Yagai and A. Kitamura, *Chem. Soc. Rev.*, 2008, **37**, 1520–1529.
- D. J. van Dijken, J. Chen, M. C. A. Stuart, L. Hou and B. L. Feringa, *J. Am. Chem. Soc.*, 2016, **138**, 660–669.
- J. N. Israelachvili, S. Marcelja, R. G. Horn and J. N. Israelachvili, *Q. Rev. Biophys.*, 1980, **13**, 121–200.
- (a) T. van Leeuwen, G. H. Heideman, D. Zhao, S. J. Wezenberg and B. L. Feringa, *Chem. Commun.*, 2017, **53**, 6393–6396; (b) M. M. Pollard, A. Meetsma and B. L. Feringa, *Org. Biomol. Chem.*, 2008, **6**, 507–512; (c) D. Zhao, T. van Leeuwen, J. Cheng and B. L. Feringa, *Nat. Chem.*, 2016, **9**, 250–256.
- (a) F. K. C. Leung, T. van den Enk, T. Kajitani, J. Chen, M. C. A. Stuart, J. Kuipers, T. Fukushima and B. L. Feringa, *J. Am. Chem. Soc.*, 2018, **140**, 17724–17733; (b) F. Tantikitti, J. Boekhoven, X. Wang, R. V. Kazantsev, T. Yu, J. Li, E. Zhuang, R. Zandi, J. H. Ortony, C. J. Newcomb, L. C. Palmer, G. S. Shekhawat, M. O. de la Cruz, G. C. Schatz and S. I. Stupp, *Nat. Mater.*, 2016, **15**, 469–476.
- Y. Yan, W. Xiong, X. Li, T. Lu, J. Huang, Z. Li and H. Fu, *J. Phys. Chem. B*, 2007, **111**, 2225–2230.

