



Cite this: *Chem. Commun.*, 2020, **56**, 6814

Received 24th March 2020,
Accepted 13th May 2020

DOI: 10.1039/d0cc02171g

rsc.li/chemcomm

A novel approach for a temperature-sensitive stabilization of water-in-water (W/W) emulsions is described. Specifically, we leveraged the thermal induced conformation change of tailored thermo-responsive block copolymers to reversibly stabilize and destabilize water–water interfaces. In addition, we investigated our approach to reversibly tune the reaction kinetics of enzymes compartmentalized within aqueous two-phase systems.

Aqueous emulsions have wide-spread applications in a wide variety of biomedical, cosmetic, food, and performance products.^{1,2} Emulsions are kinetically stabilized multiphase systems. Traditionally, to prevent breaking of the droplet network by droplet coalescence and Ostwald ripening, emulsions are stabilized by surfactants or particles.³ Recently, aqueous two-phase systems (ATPS) and water-in-water (W/W) emulsions have received increased attention,^{4–9} which is based on an observation by Beijerinck in 1896.¹⁰ Phase separation in water systems is enabled for mixtures of two polymers but also for polymer–salt mixtures.¹¹ Due to high diversity and, more importantly biocompatibility, ATPS has gained broad attention in various application fields, including biomacromolecule extraction, purification and separation or to produce micro-reactors for biomimetic reactions.^{12,13} Although macroscopically similar to O/W and W/O emulsions, W/W emulsions exhibit significantly different properties as a result of their ultralow interfacial tensions at water–water interfaces (order of $\mu\text{N m}^{-1}$), in comparison to water–oil interfaces (order of mN m^{-1}).^{14–16}

Consequently, due to the related low interfacial tension small surfactant molecules and copolymers cannot accumulate at the water–water interface to sufficiently kinetically stabilize

Temperature sensitive water-in-water emulsions†

Marko Pavlovic,^a Alexander Plucinski,^b Lukas Zeininger  ^{*a} and Bernhard V. K. J. Schmidt  ^{*ab}

these emulsions.¹⁷ Larger particles, *i.e.* Pickering emulsifiers however, can stabilize such interfaces due to a different adsorption mechanism. Although aqueous multiphase systems have ultralow interfacial tensions, particles can reduce free energy at the interface by adsorption as particle kinetic energy is significantly lower than binding energy.¹⁸ In such systems ions, surfactants, water molecules or large biomolecules can freely pass through the interface, which possesses a considerable advantage over traditional oil-based emulsions.^{19–21} As such, enzymatic reactions in W/W droplets do not require expensive transmembrane proteins or any other sort of channels to enable substrate and/or product diffusion between the phases. This membrane-less separation and enrichment of functionalities in aqueous phases with different properties featuring permeable boundaries is employed also by biological cells to increase local concentration of substrates or enzymes while maintaining communication with the environment.

Recently, various types of particles were introduced in W/W emulsion systems, including latex particles,²² nanocrystals,⁷ montmorillonite clays,¹⁸ polydopamine particles,^{23,24} amphiphilic block copolymer micelles²⁵ or liposomes²⁰ with radius >10 nm. In order to reversibly tune the kinetics of chemical reactions or for using W/W emulsions as microcontainers for the delivery of active agents, recent interest has been drawn towards a controlled formation and disruption of W/W emulsions. In a recent example, de Freitas and coworkers reported on the stabilization of aqueous emulsions using pH responsive polysaccharide-coated protein particles leading to a switchable formation and destabilization of the droplet network.²⁶

Herein, we report on a temperature-triggered reversible and switchable stabilization of W/W emulsions (Scheme 1). We make use of water-soluble double hydrophilic block copolymer (DHBC) that can be switched to an amphiphilic block copolymer above the cloud point (T_{CP}). We synthesized poly(*N,N*-dimethylacrylamide)-*b*-poly(*N,N*-diethylacrylamide) (PDMA-*b*-PDEA) DHBC with different ratios of the polymer blocks (4:1 and 2:1), where the PDEA block can be switched from the coiled to the globule state above T_{CP} leading to the formation of micelles and,

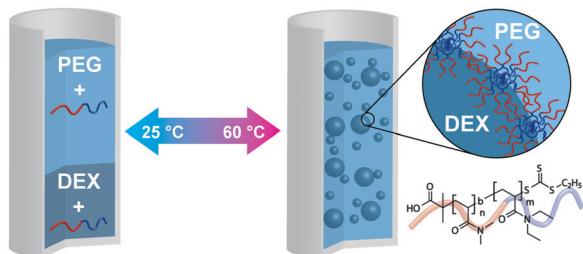
^a Max Planck Institute of Colloids and Interfaces, Department of Colloid Chemistry, Am Mühlenberg 1, 14476 Potsdam, Germany.

E-mail: Lukas.zeininger@mpikg.mpg.de

^b School of Chemistry, University of Glasgow, Joseph Black Building, G128QQ Glasgow, UK. E-mail: Bernhard.schmidt@glasgow.ac.uk

† Electronic supplementary information (ESI) available: Experimental Section, polymer characterization and additional microscopy images. See DOI: 10.1039/d0cc02171g





Scheme 1 Overview of the switchable emulsion process employing thermo responsive PDMA-*b*-PDEA block copolymers that feature a transition from coiled (unimer) to globule-based micellar state and non-stabilizing to stabilizing state with temperature.

subsequently, supramolecular aggregates. In the micellar state, the DHBCs successfully stabilized poly(ethylene glycol) (PEG) 35k-dextran (Dex) 40k W/W emulsions, which enabled tuning of the emulsion state by small temperature variations. The resulting stimuli-sensitive emulsions will help to pave the way towards dynamically triggerable purely aqueous based systems for drug delivery purposes or dynamic reaction control. To showcase this potential, we studied the kinetics of horseradish peroxidase (HRP) mediated oxidation of guaiacol within stable W/W emulsions in comparison to bulk ATPS.

We started by synthesizing switchable DHBCs *via* RAFT polymerization.²⁷ PDMA-*b*-PDEA block copolymers were synthesized with two ratios of PDMA/PDEA, *i.e.* 4:1 and 2:1 and characterized with size exclusion chromatography (SEC) and ¹H-NMR (Fig. S1–S4 and Table S1, ESI†). The PDEA block exhibits a T_{CP} value ranging from 32 °C to 41 °C, depending on polymer molar mass, heating rate and polymer concentration.²⁸ Therefore, initially T_{CP} of both block copolymers was measured *via* turbidimetry (Fig. 1a).

As it was expected, PDMA-*b*-PDEA (2:1) in comparison to PDMA-*b*-PDEA (4:1), exhibited 5 °C lower T_{CP} value. Interestingly, PDMA-*b*-PDEA (2:1) gave lower transmittance above T_{CP} in comparison to the other block copolymer, which was attributed to a certain degree of aggregation of the 4:1 copolymer at ambient temperature. Oppositely, supramolecular aggregates formed by 2:1 block copolymers are larger in diameter above T_{CP} leading to increased scattering intensity.

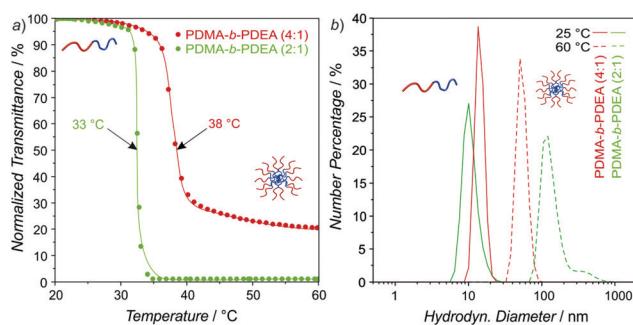


Fig. 1 Turbidimetry of PDMA-*b*-PDEA (2:1 or 4:1) at 50 mg mL⁻¹ (5 wt%) in MilliQ water (a). DLS measurement of PDMA-*b*-PDEA (2:1 or 4:1) at 2.5 mg mL⁻¹ (0.25 wt%) in MilliQ water. Both measurements were performed at ambient temperature and 60 °C (b).

To further evaluate the block copolymer behavior at different temperatures, the hydrodynamic diameter was monitored *via* dynamic light scattering (DLS) at 25 °C and 60 °C (Fig. 1b). DLS experiments correlated well with turbidimetry measurements (Fig. 1a). PDMA-*b*-PDEA (2:1) showed higher differences in radii below and above T_{CP} in comparison to the second copolymer, *i.e.* the 4:1 block copolymer increased in diameter from 14 nm to 52 nm, whereas 2:1 block copolymer increases from 10 nm to 119 nm. Approximately doubling in diameter of aggregated 2:1 block copolymer can be understood from the perspective of longer PDEA chains that form a larger hydrophobic micelle core compared to 4:1 block copolymer. In addition, it should be noted that 2:1 block copolymer also showed higher order aggregates illustrated by the shoulder towards higher diameters in the particle size distribution. For the purpose of emulsion stabilization such aggregates are advantageous due to the fact that larger spheres exhibit improved performances as a Pickering stabilizer.

Next, we investigated the preparation of switchable W/W emulsions. For this purpose, we chose W/W emulsion based on 7 w/v% of PEG 35k and 10 w/v% of Dex 40k that are characterized by a 1:1 volume ratio of the two aqueous phases, a comparatively fast phase separation in the absence of stabilizers and most importantly, the fact that this composition is far from the critical point of the system resulting in a high interfacial potential between the phases. At first, the macroscopic phase separation of PEG 35k-Dex 40k ATPS emulsion was followed over seven days (Fig. 2).

In order to prove the stability of the obtained W/W emulsion at elevated temperatures, we compared it to ambient temperature, but also to a system without any stabilizer (Fig. 2). Both block copolymers dissolve in water at ambient temperature (i_a, ii_a) or form a turbid dispersion above T_{CP} due to formation of polymeric supramolecular aggregates (i_b, ii_b). There is no change over time in the first two samples (i_a, ii_a, i_b, ii_b) regardless of the temperature. Mixed ATPS without any block copolymers (iii_a, iii_b) and with each of the block copolymers (iv_a, v_a, iv_b, v_b) were prepared. The control sample of pure ATPS (iii_a, iii_b) exhibited complete phase separation in less than 60 min and 5 min, at ambient and elevated temperatures, respectively. Increased rate of coalescence can be mainly attributed to higher diffusion rates of the polymers, but there is also a moderate effect of the increased interfacial tension caused by a temperature shift of binodal.¹⁶ In case of the last two samples (iv_a, v_a, iv_b, v_b), the stability of mixed ATPS containing each of PDMA-*b*-PDEA block copolymers is critically dependent on temperature. At ambient temperature polymeric chains are in a coiled-state, therefore they cannot cover W/W interface and prevent coalescence, and as a result they behaved as poor stabilizers (iv_a, v_a). On the other hand, at temperatures above T_{CP} , polymers tended to form micelles and supramolecular aggregates that are much higher in size and act as Pickering-stabilizer for aqueous emulsions (iv_b, v_b). Complete phase separation on macroscopic scale did not occur in these samples even after 7 days, although there were some initial stages of destabilization present in the sample stabilized by 2:1 block copolymer (iv_b) where the formation of a clear phase on top

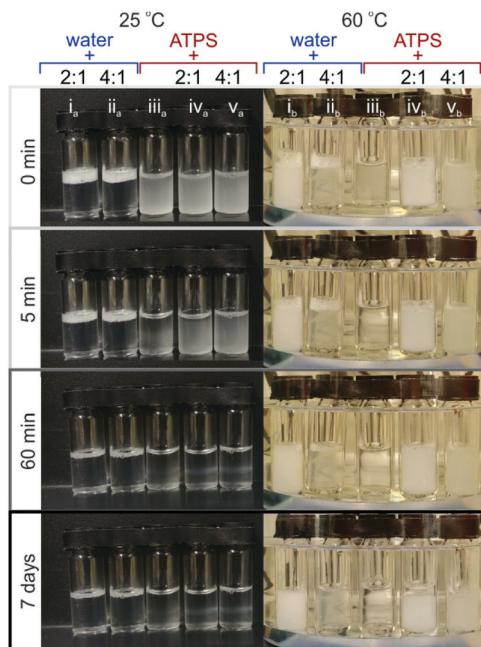


Fig. 2 Long-term emulsion stability test at 25 °C and 60 °C. First two samples are water solution of PDMA-*b*-PDEA (2:1 or 4:1) while the third one is a control sample of ATPS without any block copolymer. Last two samples are ATPS emulsion stabilized by PDMA-*b*-PDEA (2:1 or 4:1). Block copolymer concentration was set to 10 mg mL⁻¹ (1 wt%).

could be observed. This effect demonstrated that polymeric micelles showed preference to one of the phases.

The temperature-dependent emulsion stability was further investigated by optical microscopy (Fig. 3 and Fig. S5, ESI[†]). According to images taken at two different magnifications, droplet coalescence was prevented at temperatures above T_{cp} . As soon as samples reached temperatures below 38 °C (T_{cp} of PDMA-*b*-PDEA 4:1) droplet coalescence occurred, which is directly macroscopically reflected as phase separation (unstable emulsion) over time. In these experiments we observed only the beginning of the rapid coalescence at 30 °C, while at 20 °C large droplets were formed. The same effect was observed for PDMA-*b*-PDEA 2:1 (Fig. S5, ESI[†]).

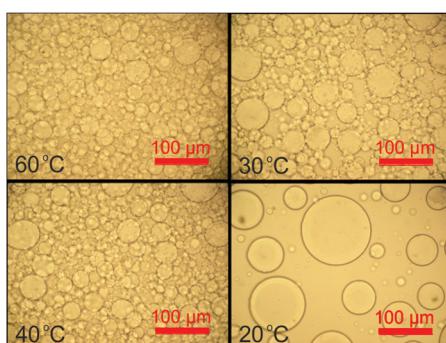


Fig. 3 Optical micrographs of W/W emulsion stabilized by PDMA-*b*-PDEA 4:1 at different temperatures. Experiment was performed at 20 \times magnification with a cooling rate of 5 °C min⁻¹. Block copolymer concentration was set to 10 mg mL⁻¹ (1 wt%).

We anticipated that the formed micelles prefer the PEG phase over the Dex phase and therefore would preferably stabilize a Dex in PEG emulsion. In order to prove this statement confocal laser scanning microscopy (CLSM) was employed. Accordingly, emulsions were prepared in presence of the block copolymers and stained with either FITC-labeled PEG or rhodamine-labeled Dex (Fig. 4a). As evident from confocal micrographs a Dex-in-PEG emulsion was formed regardless of the type of stabilizer used. Droplets exhibit moderate uniformity, except for the highly polydisperse sample of rhodamine-labeled Dex emulsion stabilized with 4:1 block copolymer. Because experiment temperature was very close to the T_{cp} of this stabilizer, we attributed this observation to a local cooling effect that promoted droplet coalescence. Nevertheless, as soon as temperature was equilibrated, droplets remained constant in radius of 54 μ m \pm 23 μ m and 24 μ m \pm 13 μ m for 2:1 and 4:1 block copolymers, respectively (measured from FITC-PEG labeled samples).

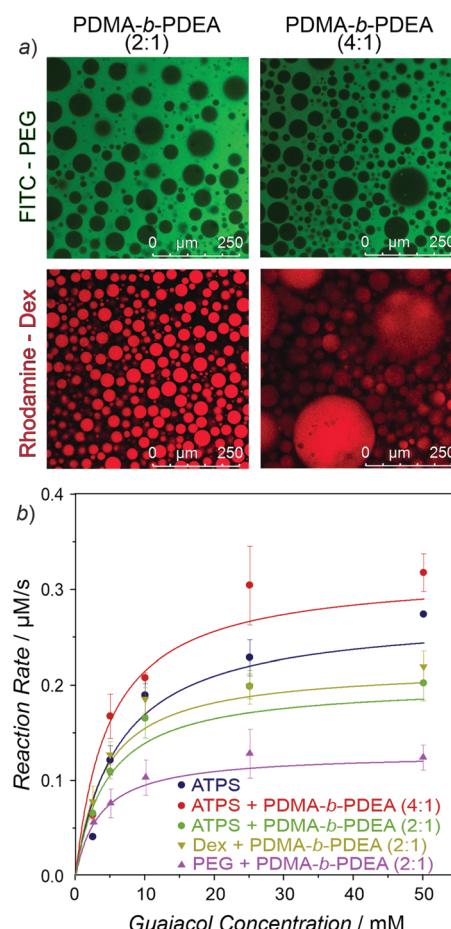


Fig. 4 CLSM micrographs of W/W emulsion stabilized by PDMA-*b*-PDEA (2:1 or 4:1) at 40 °C (samples stained with FITC-labeled PEG (green) or rhodamine-labeled Dex (red)) (a). Enzyme kinetics of guaiacol oxidation by HRP in ATPS in presence and absence of block copolymers (1 wt%) as well as controls in PEG or Dex solution. All experiments were performed in MilliQ water at 37 °C (b). Concentration of block copolymers and enzyme were set to 10 mg mL⁻¹ and 0.05 mg L⁻¹, respectively (1 wt%). The lines in the graph were obtained from Michaelis–Menten fits.



To showcase the potential of having control over the stability of aqueous emulsions we investigated enzyme kinetics in various W/W systems. Specifically, we introduced HRP to ATPSs with the different block copolymers (Fig. 4b). All experiments were performed at a temperature of 37 °C, *i.e.* in between the T_{CP} of the two different copolymers, and guaiacol was utilized as substrate for measuring the enzymatic activity (Scheme S3, ESI[†]). In our system, HRP preferably partitions to the Dex phase with a partitioning coefficient (K) of 0.63.⁸ In turn, the reaction substrate and product are more hydrophobic and consequently partition in the PEG phase (Fig. S6, ESI[†]). Therefore, we anticipated that the emulsion state and the associated different contact areas between the two phases would result in changed reaction kinetics. As displayed in Fig. 4b, when compared to the enzyme turnover rate in the pure ATPS system, the introduction of the hydrophilic block copolymer in the coiled form (PDMA-*b*-PDEA 4:1 with $T_{CP,4:1} = 38$ °C) promoted the enzymatic reaction. This increase in the maximum rate of the reaction was attributed to a higher contact area of two phases as a result of decelerated phase separation of the two phases. In contrast, when introducing the second block copolymer in the aggregated state (PDMA-*b*-PDEA 2:1 with $T_{CP,2:1} = 33$ °C), we observed a decrease of the reaction kinetics, despite a largely increased contact area between the two phases as a result of the formation of emulsion droplets. In this scenario, the presence of micelles caused encapsulation of the hydrophobic substrates, *i.e.* guaiacol,²⁹ and this confinement slowed down the mass exchange between the phases. The latter observation is supported by experiments where we investigated the enzymatic reactivity in the pure constituent phases of the ATPS which revealed a slowed down reaction rate even in the pure Dex phase. These experiments reveal that the enzymatic activity is mutually controlled by the interfacial area between the phases of an ATPS as well as by the substrate encapsulation within micelles. Thus, small temperature variations in a fine range of 5 °C around T_{CP} of the copolymers served to gain control over the reactivity of enzymatic transformations in solution.

In summary, we presented a novel approach towards stimuli-sensitive W/W emulsions. The emulsions were stabilized using synthesized PDMA-*b*-PDEA block copolymers with a thermoresponsive PDEA block. The block copolymers demonstrated utility as a competent long-term stabilizer for W/W emulsion based on PEG 35k and Dex 40k at temperatures above T_{cp} . However, decreasing the temperature below T_{CP} resulted in rapid droplet coalescence and macroscopic phase separation. In addition, enzymatic reaction kinetics of HRP were tested in the stable and unstable emulsion samples, which potentially serves as a promising path to tune enzymatic reactions in ATPS.

The authors are grateful for funding from the Max-Planck society. MP acknowledges financial support from the Swiss National Science Foundation (project no: P2GEP2_181528).

LZ is grateful for financial support through the Emmy-Noether program of the German Research Foundation (DFG) (grant no: ZE1121-3). BS and AP acknowledge funding from the DFG (grant no: SCHM 3282/3-1). Open Access funding provided by the University of Glasgow.

Conflicts of interest

There are no conflicts to declare.

Notes and references

- 1 C. C. Berton-Carabin and K. Schroën, *Annu. Rev. Food Sci. Technol.*, 2015, **6**, 263–297.
- 2 F. Goodarzi and S. Zendehboudi, *Can. J. Chem. Eng.*, 2019, **97**, 281–309.
- 3 V. Y. Grinberg and V. B. Tolstoguzov, *Food Hydrocolloids*, 1997, **11**, 145–158.
- 4 J. Esquena, *Curr. Opin. Colloid Interface Sci.*, 2016, **25**, 109–119.
- 5 L. Tea, T. Nicolai and F. Renou, *Langmuir*, 2019, **35**, 9029–9036.
- 6 Y. Beldengrün, J. Aragon, S. F. Prazeres, G. Montalvo, J. Miras and J. Esquena, *Langmuir*, 2018, **34**, 9731–9743.
- 7 K. R. Peddireddy, T. Nicolai, L. Benyahia and I. Capron, *ACS Macro Lett.*, 2016, **5**, 283–286.
- 8 M. Pavlović, A. Plucinski, J. Zhang, M. Antonietti, L. Zeininger and B. V. K. J. Schmidt, *Langmuir*, 2020, **36**, 1401–1408.
- 9 M. Pavlović, M. Antonietti, B. V. K. J. Schmidt and L. Zeininger, *J. Colloid Interface Sci.*, 2020, **575**, 88–95.
- 10 M. W. Beijerinck, *Z. Chem. Ind. Kolloide*, 1910, **7**, 16–20.
- 11 R. Hatti-Kaul, *Mol. Biotechnol.*, 2001, **19**, 269–277.
- 12 Y. Chao and H. C. Shum, *Chem. Soc. Rev.*, 2020, **49**, 114–142.
- 13 B. V. K. J. Schmidt, *Macromol. Chem. Phys.*, 2018, **219**, 1700494.
- 14 E. Scholten, J. E. Visser, L. M. C. Sagis and E. van der Linden, *Langmuir*, 2004, **20**, 2292–2297.
- 15 Y. A. Antonov, P. Van Puyvelde and P. Moldenaers, *Int. J. Biol. Macromol.*, 2004, **34**, 29–35.
- 16 D. Forciniti, C. K. Hall and M. R. Kula, *J. Biotechnol.*, 1990, **16**, 279–296.
- 17 B. T. Nguyen, W. Wang, B. R. Saunders, L. Benyahia and T. Nicolai, *Langmuir*, 2015, **31**, 3605–3611.
- 18 W. J. Ganley, P. T. Ryan and J. S. van Duijneveldt, *J. Colloid Interface Sci.*, 2017, **505**, 139–147.
- 19 B. Davis, W. Aumiller, N. Hashemian, S. An, A. Armaou and C. Keating, *Biophys. J.*, 2015, **109**, 2182–2194.
- 20 D. C. Dewey, C. A. Strulson, D. N. Cacace, P. C. Bevilacqua and C. D. Keating, *Nat. Commun.*, 2014, **5**, 4670.
- 21 S. D. Hann, K. J. Stebe and D. Lee, *ACS Appl. Mater. Interfaces*, 2017, **9**, 25023–25028.
- 22 H. Firoozmand, B. S. Murray and E. Dickinson, *Langmuir*, 2009, **25**, 1300–1305.
- 23 J. Zhang, B. Kumru and B. V. K. J. Schmidt, *Langmuir*, 2019, **35**, 11141–11149.
- 24 J. Zhang, J. Hwang, M. Antonietti and B. V. K. J. Schmidt, *Biomacromolecules*, 2019, **20**, 204–211.
- 25 D. M. A. Buzzia, P. D. I. Fletcher, T. K. Georgiou and N. Ghasdian, *Langmuir*, 2013, **29**, 14804–14814.
- 26 R. A. de Freitas, T. Nicolai, C. Chassenieux and L. Benyahia, *Langmuir*, 2016, **32**, 1227–1232.
- 27 B. V. K. J. Schmidt, M. Hetzer, H. Ritter and C. Barner-Kowollik, *Macromolecules*, 2011, **44**, 7220–7232.
- 28 I. Idziak, D. Avoce, D. Lessard, D. Gravel and X. X. Zhu, *Macromolecules*, 1999, **32**, 1260–1263.
- 29 A. Faridi Esfanjani and S. M. Jafari, *Colloids Surf., B*, 2016, **146**, 532–543.

