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Structural investigation of oxidized arabinoxylan oligosaccharides by negative ionization HILIC-qToF-MS†

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Owing to the strong structure–function relationship of polysaccharides, the targeted modification of polysaccharides is attracting widespread interest in various fields, such as food industry, nutritional science, and biomedical research. Apart from intended functionalization, polysaccharide degradation mediated by hydroxyl radicals (HO^\bullet) occurs in various industrial processes such as food processing. In particular, the oxidative degradation of feruloylated arabinoxylan (AX), a linearly-branched polysaccharide in cereals, causes chain scissions, and introduces new functional groups in the fiber, which can potentially modify the physicochemical properties and the functionalities of AX. However, the precise characterization of those structural modifications remains challenging due to the diversity of the oxidation products formed, the high molecular weight, and the relatively low quantity of newly formed functional groups. In this paper, selective (TEMPO-mediated) and random (Fenton) oxidations of several commercial xylo- and arabinoxylan oligosaccharides (AXOS) were studied as model systems by hydrophilic interaction UPLC-MS² in negative ion resolution mode to identify potential oxidation products. An in-depth identification of acidic (AXOS) oxidation products derived from TEMPO-mediated oxidation provided novel insights in the selective functionalization of isomeric oligosaccharides. Furthermore, MS² enabled the precise localisation of both glycosidic linkages and functional groups in oxidized (AXOS). An innovative combination of an enzymatic sample preparation combined with a subsequent HILIC-MS² analysis enabled the unprecedented comprehensive characterization of Fenton-induced oxidation products derived from AX. In future, this holistic analytical approach will enable the characterization of both selective and non-selective AX oxidation procedures in various applications.

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1. Introduction

In recent years, polysaccharides and their derived oligosaccharides have aroused great interest in research owing to their widespread biological functionalities.¹ The biological activity and the physicochemical properties of polysaccharides are highly affected by their complex structural characteristics such as molecular weight (M_w), varying monosaccharide building blocks, and substitution pattern. Therefore, the chemical modification of polysaccharides is commonly used to modulate specific properties and functionalities in different industrial applications such as drug delivery, materials science, or as functional food.² Besides this intended functionalization of polysaccharides, food processing modifies the native structure

of polysaccharides unspecifically, leading to a significant alteration of their health promoting and technological functionalities.^{3–5}

One of the major non-digestible polysaccharides within the human diet is feruloylated arabinoxylan (AX), a partially soluble fiber predominantly found in the cell wall of cereal grains such as rye, wheat, barley, and oat. The polysaccharide is composed of a β -(1→4)-linked D-xylopyranosyl backbone with random substitutions at O-2, O-3, or O-2 and O-3 with α -L-arabinofuranose units. Phenolic acids, primarily ferulic acid, can be esterified on the C(O)-5 position of arabinose units.⁶ Since cereals usually undergo extensive processing treatments such as grain milling, extrusion, hydrothermal treatments, or baking, AX is prone to degradation induced by those processes.⁵

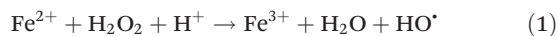
This degradation process includes hydrolytic reactions and chemically-induced oxidations. Thereby, the chemically-induced oxidative degradation takes place by hydroxyl radicals (HO^\bullet) randomly abstracting hydrogen atoms from any C–H groups within the monomer units, introducing new functional

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groups and causing – depending on the attack position within the monomers – also chain cleavage of the polymer backbone. HO[•] can be generated through the Fenton reaction (1), mediating the oxidation of AX.⁷ All in all, a combination of molecular oxygen, metal traces (Fe, Cu), and reducing agents build the catalytic cycle for HO[•] production.^{8–10}



As a consequence, carbonyl or carboxyl groups are formed, ring openings occur, and the M_w decreases.⁸ Since such structural features (new functional groups & chain length) are related to the solubility, viscosity and the derived functionalities of AX, the identification of those structural changes is crucial to understand AX's nutritional value.¹¹ However, the analysis of oxidized polysaccharide is challenging due to the high M_w and the relatively small number of newly formed functional groups. Prior research has focused on the application of multiple analytical techniques studying the oxidative state of polysaccharides such as AX. In prior studies, Bagdi *et al.* and Faure *et al.* used electron spin resonance spectroscopy (ESR) to monitor the HO[•] formation during polysaccharide oxidation.^{7,12} In addition, Bagdi *et al.* studied the influence of AX oxidation on molecular weight, viscosity, bile acid binding, and gel forming capacity. Similar studies have also been performed with β -glucan, in which the main focus was the determination of bulk properties and rheology.^{13,14} A deeper insight into the molecular level of polysaccharide oxidation was presented by Potthast *et al.*, who quantified carbonyls in polysaccharides through fluorescent labelling by high performance size exclusion chromatography (HPSEC) with multiple detectors.¹⁵ In a previous study, oligosaccharide products diagnostic for β -glucan oxidation were investigated by UPLC-MS² with and without prior functional group labelling, followed by selective enzymatic digestion and solid phase extraction.¹⁶ Among the analytical approaches mentioned, liquid chromatography (LC) combined with mass spectrometry (MS), is one of the most powerful techniques to analyse oligosaccharides diagnostic for oxidation.^{17–19} However, to the best of our knowledge, there is no reported study using this technology for the characterization of Fenton oxidized arabinoxylan, and thus the structural building blocks of oxidized AX have yet to be elucidated.^{20,21}

Hence, this study aims to elucidate the structural modification of diagnostic arabinoxylan oligosaccharides (A)XOS through selective TEMPO-mediated and non-selective Fenton oxidation in a holistic analytical approach and its subsequent application on non-selectively oxidized AX. For this purpose, a detailed characterization of oxidized (A)XOS as model compounds using hydrophilic interaction liquid chromatography (HILIC) coupled with MS² in negative ion resolution mode was performed. Our work is the first application of this established method for branched heterooligosaccharides. An in-depth identification of acidic (A)XOS oxidation products derived from TEMPO-mediated oxidation provided insights in the selective functionalization of isomeric oligosaccharides. Moreover, this paper presents an unprecedented comprehensive

characterization of the radical induced Fenton oxidation of (A)XOS. In a proof of concept experiment, the complex product mixture of Fenton-mediated arabinoxylan oxidation was characterized in detail using an innovative combination of an enzymatic sample preparation strategy and HILIC-MS² analysis.

2. Materials and methods

2.1. Materials

Linear XOS (X3, X4, X5, and X6), branched AXOS standards (A³X, A²XX, XA³XX, A²⁺³XX) both >95%, wheat arabinoxylan (medium viscosity; ~95%), and *endo*-1,4- β -xylanase from *Cellvibrio mixtus* (750 U mL⁻¹) were purchased from Megazyme International (Bray, Ireland), ammonium formate ($\geq 99.995\%$ trace metal basis), 25% aqueous ammonia (NH₃) and formic acid (both LC-MS grade), 35% hydrogen peroxide (H₂O₂; purum p.a.), iodine (I₂; $\geq 99.8\%$), iron(II) sulfate heptahydrate (FeSO₄·7H₂O; $\geq 99\%$), methanol (HPLC-grade), potassium hydroxide (KOH; $\geq 85\%$), sodium chlorite (NaClO₂; 80%), sodium hydroxide (NaOH; $\geq 97.0\%$), sodium hypochlorite solution (NaClO; 10–15%), and 2,2,6,6-tetramethylpiperidine 1-oxyl (TEMPO; 98%) were purchased from Sigma-Aldrich Chemie GmbH (Germany). L-Ascorbic acid (AA; $\geq 99.5\%$) was purchased from Fluka (Germany). Acetonitrile (ACN; ULC/MS grade) was purchased from Biosolve B.V. (Valkenswaard, The Netherlands) and the aqueous solutions were prepared with nanopure Milli-Q® water (H₂O; ≥ 18.2 M Ω cm at 25 °C).

2.2. Methods

2.2.1. Synthesis of xylonic acid-bearing oligosaccharide standards by I₂/KOH. Xylonic acids of A³X, X3, A²XX, X4, A³XX, A²⁺³XX, X5, and X6 were synthesised by selective C1-oxidation of the native oligomers. The reducing end was oxidized with hypoiodite based on a procedure presented by Hashimoto *et al.* with modifications.²² The oligomers (2 μ mol; 1 equiv.) were dissolved in H₂O (1.5 mL) and sonicated for 5 min. Afterwards, methanolic I₂ solution (0.1 M, 60 μ L; 3 equiv.) was added, followed by aqueous KOH solution (0.1 M, 160 μ L; 8 equiv.). The final reaction volume was adjusted to 3 mL with H₂O, mixed, and fractionized by solid phase extraction after 1 h at room temperature (see section 2.2.4).

2.2.2. Acidic oxidation products of (A)XOS by TEMPO-mediated oxidation. The preparation of acidic oxidation products was performed using the TEMPO/NaClO/NaClO₂ system.²³ (A)XOS (16 μ mol; 1 equiv.) were dissolved in acetate buffer (pH 4.7; 10 mM, 1 mL). Then, NaClO₂ (29 mM, 221 μ L; 40 equiv.), TEMPO (2 mM, 320 μ L; 4 equiv.), and NaOCl (13 mM, 615 μ L; 50 equiv.) were added. The final reaction volume was adjusted to 10 mL H₂O, mixed and stirred for 24 h at 20 °C. The reaction was stopped by SPE fractionation (see section 2.2.4).

2.2.3. Fenton degradations

2.2.3.1. (A)XOS. For the non-selective oxidation of (A)XOS, a reaction mixture of 1.5 mL final volume was prepared con-



taining 1 mM of the respective oligosaccharide, 100 μM FeSO_4 , 500 μM ascorbic acid (AA), and 10 mM H_2O_2 on the basis of Bagdi *et al.* with some modifications.²⁴ The oxidation was performed at room temperature (22–24 °C) with access to air for 24 h. Each oxidation was done in triplicates, including reagent controls that contained only the oligosaccharide in water. SPE fractionation followed as usual (see section 2.2.4.).

2.2.3.2. AX polymer. Arabinoxylan (132 mg) was dissolved in H_2O (5 mL) for 1 h at 80 °C and stirred at 20 °C overnight. Subsequently, the reagents were added to give final concentrations of 2% AX (w/v), 50 μM FeSO_4 , 50 μM AA, and 50 mM H_2O_2 in this order.²⁴ The oxidation was heated for the first 3 hours to 80 °C, then continued at RT with access to air for 24 h. The oxidation was done in triplicates, including reagent controls, and subsequently freeze dried. The oxidized AX (15 mg) was dissolved in H_2O (2.4 mL) for 1 h at 80 °C. Then, sodium phosphate buffer (0.5 M, 60 μL , pH 6.5) was added resulting in a final AX concentration of 0.5% (w/v) and a total volume of 3 mL. Subsequently, the oxidized AX was treated with *endo*-1,4- β -xylanase from *Cellvibrio mixtus* (750 U mL^{-1} , 144 μL) for 6 h at 40 °C to decrease the molecular weight.

2.2.4. Solid phase extraction (SPE). The reducing end oxidized (A)XOS were fractionated using the procedure as previously reported.²¹ The supelclean™ ENVI-Carb SPE cartridges (3.0 mL, 0.25 g) from SUPELCO® (Sigma-Aldrich, Germany) were preconditioned with 3.0 mL ACN and 3.0 mL H_2O . The reaction mixtures were loaded on the SPE cartridges and subsequently washed with 1.5 mL H_2O with a drop speed of 1–3 drops per second. The remaining native oligomers and neutral side products were eluted with 1.5 mL ACN/ H_2O 1 : 3 (v/v) (fraction 1). Following, 1.5 mL of 0.1% (v/v) formic acid in ACN/ H_2O 1 : 3 (v/v) was used to elute fraction 2 containing the desired C1 oxidized product. Fractions 2 were collected directly in HPLC vials and analysed by UPLC-MS².

2.3. UPLC-MS and MS²

2.3.1. Instrumentation. The Acquity UPLC system was equipped with an Acquity UPLC BEH Amide column (2.1 mm \times 150 mm, 1.7 μm). The UPLC was coupled to a Synapt G2 MS system with an electrospray ionisation (ESI) source and a quadrupole time-of-flight (qToF) analyser (Waters Corp., Milford, MA, USA). The calibration of the MS was done with sodium formate solution and leucine-enkephalin (m/z 554.2615) was used as lock mass. This lock mass was acquired every 45 s during the measurements in negative ion mode (correction applied with 5 scans averaged).

2.3.2. Chromatographic conditions. Both gradients applied were based on ACN and H_2O as eluents with a flow rate of 0.34 mL min^{-1} . Mixtures of ACN/ H_2O 8 : 2 and 2 : 8 (v/v) were used as weak and strong needle wash solutions, respectively. Injection volumes were 5 μL –10 μL of the undiluted and diluted sample (75% ACN) in partial loop mode, respectively. The column was maintained at 35 °C. The first gradient (Gradient I) for an overall analysis of the reaction mixture started with 20% H_2O and 80% ACN (both with 0.1% NH_3 additive), then increased linearly to 40% H_2O in 12 min. After

maintaining this for 4 min, it was reequilibrated with the initial conditions for 1 min.

The second gradient was applied for an improved separation of the acidic oxidation products (Gradient II) based on the procedure presented by Westereng *et al.*²⁵ Eluent A and B were a mixture of ACN/ H_2O in the ratio of 1 : 3 and 3 : 1, both with the addition of 60 mM ammonium formate buffer (NH_4HCO_2 ; pH 8). The gradient started isocratically with 100% B for 2 min (= 25% H_2O), increased linearly to 34% A for 5.5 min (= 42% H_2O), maintained for 1.5 min, and decreased back to 0% A within 1 min and reequilibrated for 2.5 min.

2.3.3. MS conditions. For both eluent systems, spectra were acquired in resolution mode and the voltages of capillary, sample cone, and extraction cone were set at 2000, 15, and 3 V for the NH_3 eluent system, respectively; whereas higher voltages were applied for the buffered ammonium formate eluent (2000, 25, and 4 V). The desolvation gas flow rate was 850 L h^{-1} at 350 °C, cone gas flow was 20 L h^{-1} and the source temperature 120 °C. The full scans were acquired from m/z 50 to 1200 with a scan rate of 1 Hz in centroid mode. In addition, with a selection of exact masses, MS² was performed through collision-induced dissociation (CID) with Argon as collision gas. For a narrow m/z isolation width, low mass (LM) resolution was set to 15, and the transfer collision energies were ramps ranging from 5–15 to 10–60 V depending on propensity for fragmentation. The data collected were processed in MassLynx software, version 4.1 (Waters Corp., Milford, MA, USA). Base peak ion chromatograms (BPI) were background-corrected by subtracting a blank injection, m/z signals that could be clearly assigned to contaminations were excluded. Peak integration was done in extracted ion chromatograms (XIC) with m/z of the analyte in question ± 0.05 Da.

2.3.4. Data processing. The data collected were processed using MassLynx software, version 4.1 (Waters Corp., Milford, MA, USA). BPI chromatograms and MS spectra were background-corrected by subtracting the signals of a blank from the respective measurement day. Retention time shifts were corrected by standard measurements of the native oligosaccharides and chromatograms displayed were smoothed (3 \times).

3. Results & discussion

3.1. Identification of selectively oxidized (A)XOS

3.1.1. UPLC-MS² differentiation of isomeric xylonic acid-bearing oligosaccharides. One of the main acidic products formed during Fenton-induced degradation of poly- and oligosaccharides are the result of lytic C1-oxidation, which for AX would correspond to xylonic acid-bearing oligosaccharides.^{20,21} In order to enable clear distinction of isomeric xylonic acid oxidation products, several commercially available (A)XOS were oxidized by means of selective reactions, namely I_2/KOH and $\text{TEMPO}/\text{NaClO}/\text{NaClO}_2$ and compared by MS². According to literature, $\text{TEMPO}/\text{NaClO}/\text{NaClO}_2$ preferentially oxidizes primary alcohols and reducing ends in sugar molecules which leads



among others to the formation of C1 oxidation products.²⁶ Those C1 oxidation products were further confirmed by the selective reducing end C1 oxidation using I_2/KOH (data not shown). In a first step, $NaClO$ initializes the oxidation of nitroxyl radicals such as TEMPO to their corresponding oxoammonium salt, which further oxidizes the primary hydroxyl groups of the sugar units to aldehydes. In a subsequent reaction, $NaClO_2$ oxidizes the resulting aldehydes to carboxylic acids, which regenerates $NaClO$.^{23,27} This general mechanism makes clear why the reducing end is readily oxidized as well as it is an aldehyde in the open-chain form, and hence easily oxidized by $NaClO_2$ to xylonic acids.

The obtained MS^2 spectra of the TEMPO-mediated oxidized oligosaccharides revealed mainly signals originating from the

fragmentation starting from the oxidized downstream end, as shown in Fig. 1 on the example of oxidized pentopentaoses (for -tetraoses and -trioses, see Fig. S1 in the ESI†). This observation is in accordance with negative ion ESI- MS^2 data reported for native (A)XOS by Juvonen *et al.*¹⁷ While linear, native XOS exclusively fragment from the reducing end (A, B, C fragments according to the nomenclature of Domon and Costello),²⁸ native AXOS also present fragments which can be assigned to Y fragments originating from the fragmentation starting at the non-reducing end. The Y type fragments are more pronounced in some xylonic acid-bearing oligosaccharides, which may be attributed to the high ionizability of the carboxylate group that is part of all Y fragments in these xylonic acid products.

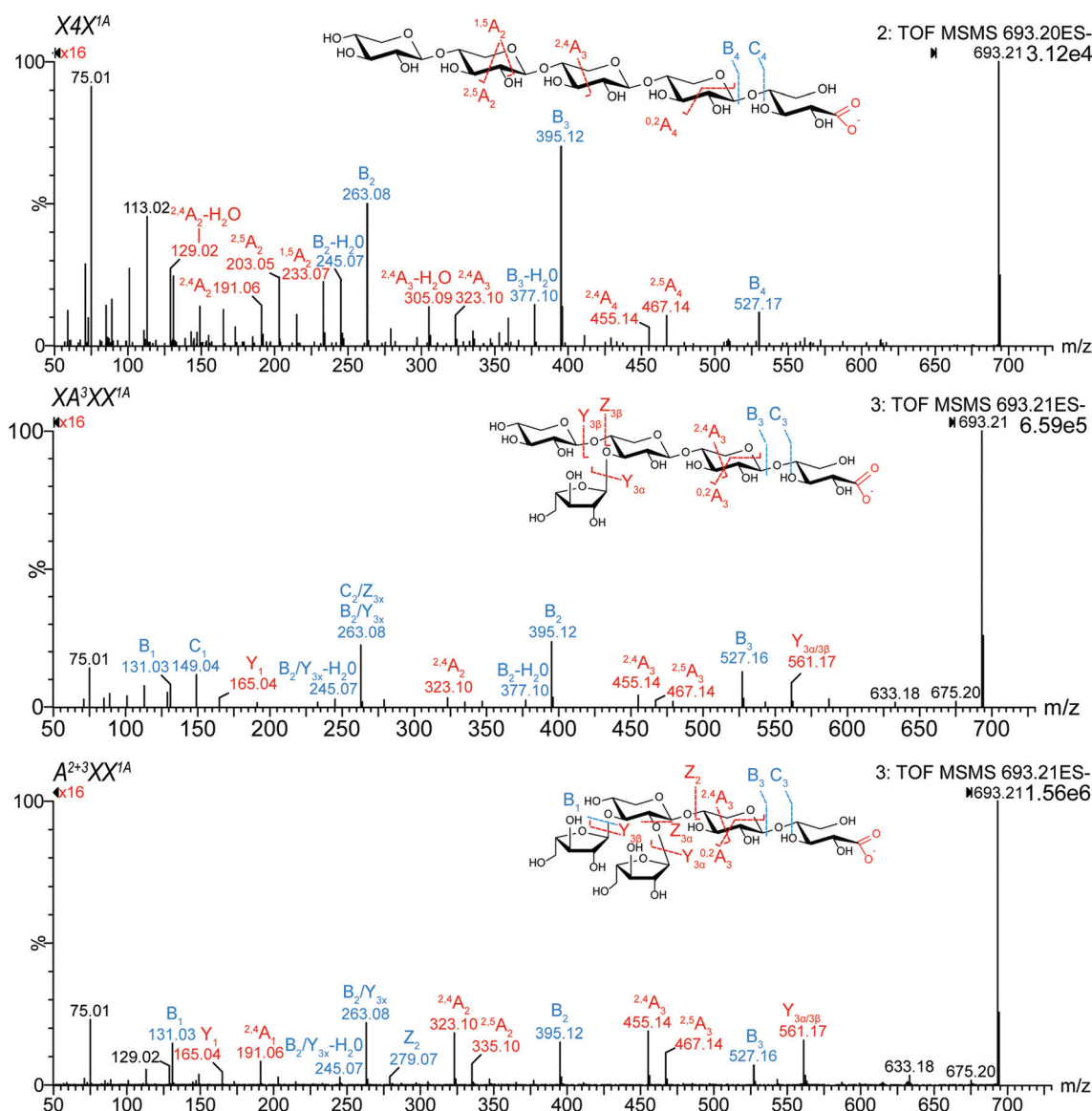


Fig. 1 MS^2 spectra of isomeric xylonic acid products with m/z 693.21, obtained by TEMPO-mediated oxidation of pentopentaose standards. TEMPO/ $NaClO$ / $NaClO_2$ preferentially oxidizes primary alcohols and reducing ends in sugar molecules, which leads among others to the formation of C1 oxidation products.²⁶ Those C1 oxidation products were further confirmed by the selective C1 oxidation using I_2/KOH (data not shown). The fragments were named according to the nomenclature of Domon and Costello.²⁸



All studied isomeric pentose oligosaccharides (Pent_n), namely C1-oxidized pentotrioses (X3 and A³XX), pentotetraoses (A²XX and X4), and pentopentaoses (XA³XX, A²⁺³XX, and X5) could be clearly distinguished and linkage positions of arabinose substitution could be easily identified (see Fig. 1, Table 1, and Fig. S1†). Linear XOS xylonic acids (Xyl_(n-1)Xyl1A = X(n-1)X^{1A}) could be mainly elucidated by their dominating B fragments and the absence of Y_(n-1) fragment. In contrast, both mono- and disubstituted oligomers presented a Y_(n-1) fragment, presumably due to the ease of loss of a side chain arabinose unit. For AXOS, the ^{1,5}A₂ fragment (*m/z* 233.07) was less abundant compared to XOS. Moreover, particular linkages of the arabinose substitution could be identified by the signal ratios of ^{2,4}A_n/^{2,5}A_n, namely *m/z* 455.14/467.14 (for *n* = 5), 323.10/335.10 (for *n* ≥ 4), and 191.06/203.06 (for *n* ≥ 3). While *m/z* 467.14 (^{2,5}A₄) was more abundant in oxidized linear X5 (X4X^{1A}), *m/z* 455.14 (^{2,4}A₄) was dominating in both substituted pentopentaoses (ratio >1). Similar ratios of ^{2,5}A₂ (203.06) and ^{2,4}A₂ (191.06) were found for pentotrioses. The opposite could be observed for pentotetraoses, where ^{2,4}A₂ (323.10) was more abundant in oxidized linear X4 (X3X^{1A}), and ^{2,5}A₃ (335.10) was dominating in A²XX^{1A}. Among the pentopentaoses evaluated, only the monosubstituted XA³XX^{1A} formed C₁ fragments (*m/z*

149.04). Therefore, the region below *m/z* 200 allows for a facilitated differentiation of arabinose substitution pattern.

These findings are in line with the previously published MS² differentiation of C1-oxidized *gluco*-oligosaccharides with gluconic acid downstream end.²¹ Consequently, all pento-oligosaccharides with the same *m/z* revealed unique MS² fragmentation, which enables a clear identification of isomeric AX, (A)XOS derived oxidation products.

3.1.2. Identification of acidic oxidation products of (A)XOS from TEMPO-mediated oxidation. In the random oxidation *e.g.* during food processing, acidic products are one of the major oxidation product classes formed through lytic and non-lytic reactions. In this study, acidic products were synthesized by TEMPO-mediated oxidation and subsequently investigated by UPLC-MS². The TEMPO/NaClO/NaClO₂ system converts primary hydroxyl groups in sugars to carboxylic acids.

Whereas linear XOS exclusively formed their corresponding xylonic acid product resulting from C1 xylose oxidation, AXOS created a mixture of acidic oxidation products resulting from both the C1 xylose oxidation and the C5 arabinose oxidation (see Fig. 2). Nevertheless, the product bearing the oxidized downstream end remained predominant during the AXOS oxidation as primary oxidation product. A variation of reagent equivalents corroborated the formation of reducing end oxidation prior to further oxidations at other positions (data not shown).²¹

Fig. 2 summarizes the successfully identified carboxylic acid-bearing oligosaccharides from A²⁺³XX and A²XX in the reaction mixture after TEMPO-mediated oxidation using UPLC-MS² with a buffered ammonium formate eluent (pH 8). Generally, this gradient separated acidic products according to their size and polarity on the UPLC BEH Amide column. The specific products formed could be detected by their unique *m/z* and even several isomeric products, *e.g.* of *m/z* 559.15, 575.15, and 707.19 achieved appropriate separation by the gradient applied. In particular, *m/z* 559.15 can be assigned to a product after C5 oxidation in the arabinose unit as well as to a product after C1 and C5 oxidation of the downstream xylose unit.

3.2. Identification of non-selective oxidized (A)XOS using Fenton oxidation

3.2.1. Acidic products. The complex product mixture of Fenton oxidized oligosaccharides was characterized using HILIC-MS². The general separation of acidic and neutral compounds could be achieved using a basic eluent system (0.1% NH₃). As already reported in a previous study as well as by Leijdekkers *et al.*, this basic eluent caused nearly no retention and partial coelution of all acidic products on the amide HILIC column.^{21,29} However, their anionic natures lead to strong signal strength in the negative ion mode, which is shown as collective MS spectrum of the respective region in the chromatogram in Fig. 3D–F for A²⁺³XX, XA³XX and X5 as substrates. In contrast to direct infusion MS, the data assembled already separated between neutral and charged products without a time-consuming sample preparation.

Table 1 MS² Overview of noteworthy diagnostic fragments (relative abundance ≥5%) enabling the distinction of isomeric xylonic acid products. The fragments were named according to the nomenclature of Domon and Costello²⁸

Oligosaccharide	Fragment ion (<i>m/z</i>)	Missing fragment ion (<i>m/z</i>)
Oxidized pentopentaoses X4X ^{1A}	Parent ion (<i>m/z</i> 693.21)	
	^{2,5} A ₃ (335.10)	Y _{3α/β} (561.17)
	^{1,5} A ₂ (233.07)	
	^{2,5} A ₂ (203.05)	
	^{2,4} A ₂ (191.06)	
	C ₁ (149.04)	
XA ³ XX ^{1A}	Y _{3α/β} (561.17)	^{2,5} A ₃ (467.14)
	C ₁ (149.04)	^{2,4} A ₃ –H ₂ O (305.09)
		^{2,5} A ₂ (335.10)
		^{2,5} A ₂ (203.05)
A ²⁺³ XX ^{1A}	Y _{3α/β} (561.17)	B ₂ –H ₂ O (377.10)
	^{2,5} A ₂ (335.10)	^{1,5} A ₂ (233.07)
	^{2,4} A ₂ (191.06)	^{2,5} A ₂ (203.05)
Oxidized pentotetraoses X3X ^{1A}	Parent ion (<i>m/z</i> 561.17)	
		Y ₃ (429.12)
		Z ₂ (279.07)
A ² XX ^{1A}	Y ₃ (429.12)	
	Z ₂ (279.07)	
	Y ₁ (165.04)	
Oxidized pentotrioses X2X ^{1A}	Parent ion (<i>m/z</i> 429.12)	
	^{1,5} A ₂ (233.07)	Y ₂ (297.08)
	^{2,5} A ₂ (203.06)	
A ³ X ^{1A}	Y ₂ (297.08)	B ₂ –H ₂ O (245.07)
		^{2,5} A ₂ (203.06)
		^{1,5} A ₂ (233.07)



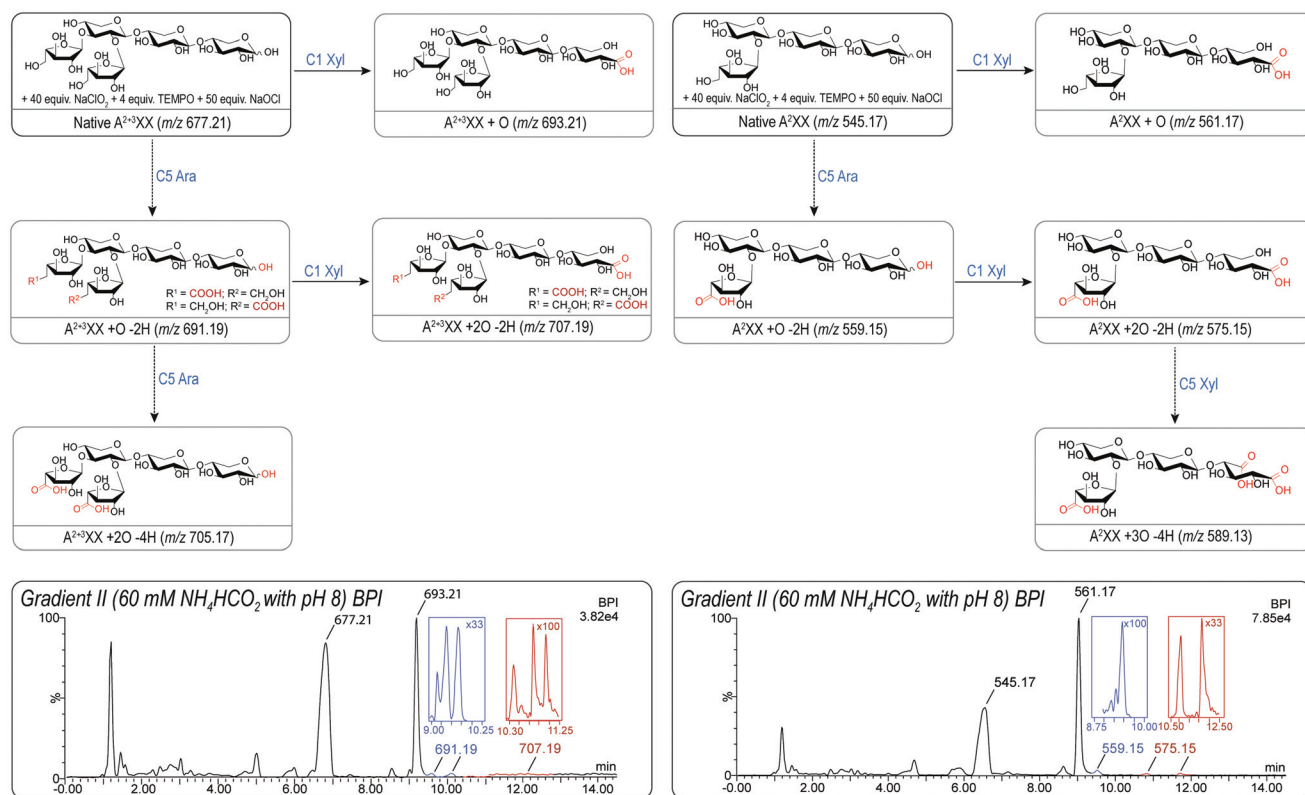
(A) TEMPO-mediated oxidation products of $A^{2+3}XX$ (B) TEMPO-mediated oxidation products of A^2XX 

Fig. 2 Exemplary overview of TEMPO-mediated oxidation products of $A^{2+3}XX$ and A^2XX . The shown m/z in parenthesis represent the deprotonated species $[M - H]^-$. The samples were analyzed by UPLC-MS using a buffered 60 mM ammonium formate eluent system at pH 8 and a BEH Amide column. Several isomeric products were detected for m/z 559.15, 575.15, and 707.19 differing in their oxidation position, e.g. m/z 559.15 can be assigned to a product after C5 oxidation in the arabinose unit as well as to a product after C1 and C5 oxidation of the downstream xylose unit. (A) TEMPO-mediated oxidation products of $A^{2+3}XX$. (B) TEMPO-mediated oxidation products of A^2XX .

The MS spectra from the oxidation of isomeric pentopen-taoses (with m/z 677.21) revealed signals clustered around the primary C1-oxidation products with m/z 165.04, 297.08, 429.12, 561.17, and m/z 693.21, representing $Pent_{(n-1)}Xyl1A$ with $n = 1, 2, 3, 4$, and 5 , respectively. Signals with the same number of monosaccharide units n , but higher m/z than their respective pentose (Pent) acid oligomers, such as m/z 707.19 ($n = 5$), are attributed to products with at least 1 additional oxygen, whereas products with lower m/z refer tetrose-, glyceric-, and glyconic acid oligomers formed by cross-ring cleavages during Fenton oxidation.

As expected, all products formed and identified in the TEMPO-mediated oxidation of oligomers were also present in the complex product mixture of Fenton oxidized XOS and AXOS and were therefore easily identified (see Fig. 3D–F). Furthermore, the product distribution with respect to the degree of polymerization (DP) varies between the oligosaccharides analyzed (see Fig. 3 for $A^{2+3}XX$, XA^3XX , and X5). This observation shows that the fine structure influences the chain scission and oxidation behaviour of oligosaccharides, which is in contrast to previous findings on β -glucan oligomer oxidation, where similar results for all four linear glucotetraose

isomers could be observed.²¹ The product profile obtained from the Fenton oxidation of arabinoxytan will strongly depend on the native fine structure of the polysaccharide.

3.2.2. Neutral products. Neutral degradation products formed during Fenton oxidation were reduced in size and eluted before the native oligosaccharide. In general, the neutral product profiles of the constitutionally isomeric substrates, namely X5, XA^3XX , and $A^{2+3}XX$ differed greatly and a clear differentiation of isomeric degradation products was possible by their retention time (see Fig. 3A–C for $A^{2+3}XX$, XA^3XX , X5). Linear XOS eluted in order of their size (DP), but slightly later than the respective substituted AXOS isomer (see Fig. 3, and Fig. S2†). The most abundant products of the neutral fraction were oligosaccharides with reduced DP from lytic oxidation (m/z 545.17 = $Pent_4$, m/z 413.13 = $Pent_3$) and their respective oxidized compounds with an additional carbonyl function (oxo- $Pent_n$ of m/z 675.20, 543.16, and 411.11 for $n = 5, 4$, and 3 , respectively). The identified neutral products with reduced DP revealed cleavage reactions occurring from both the non-reducing or the downstream end, as well as loss of arabinose side chains. These oxo-products eluted both earlier and later than their respective native oligomers.



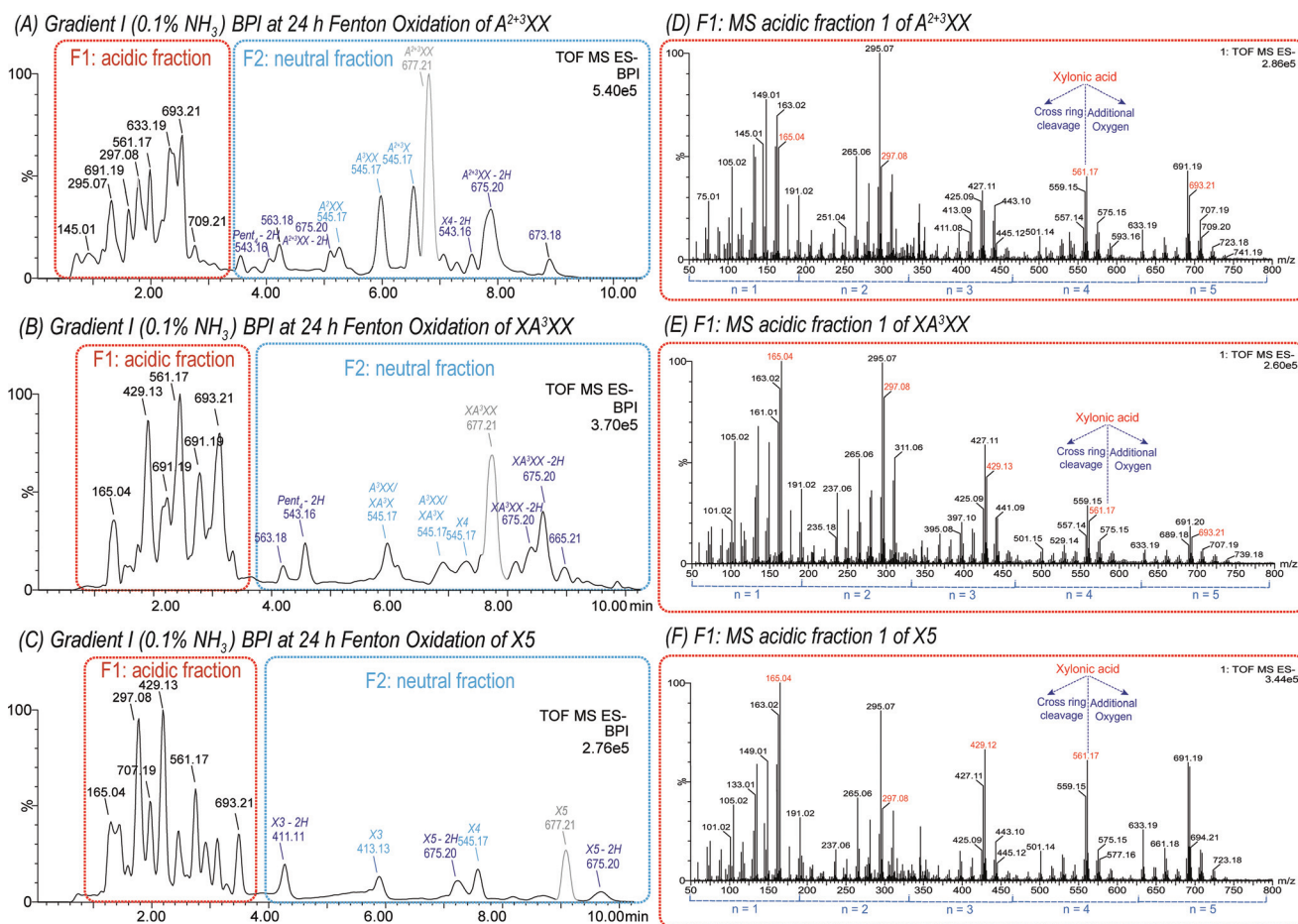


Fig. 3 UPLC-MS base peak ion chromatogram (BPI) using a BEH Amide column (ACN/ H_2O gradient I with NH_3 additive) and negative ion mode ESI of 1 mM $A^{2+3}XX$ (A), XA^3XX (B), and $X5$ (C) solution oxidized with 100 μM $FeSO_4$, 100 μM ascorbic acid and 500 mM H_2O_2 after 24 h at room temperature. Peak area of the starting material is indicated in grey. ESI-MS spectra of acidic fraction (F1) of the respective oligosaccharides (D–F), derived from the red marked area in (A–C). Product assignment was based on elution time of native, commercial oligosaccharides (see Fig. S2†).

Presumably, this depends on the additional carbonyl position. Similar neutral product types as the one identified by Fenton oxidized (A)XOS, have been determined in the Fenton induced oxidation of arabinotriose by Moreira *et al.* and of β -glucan oligosaccharides as previously reported by Boulos and Nyström.^{16,20,21} Consequently, the basic gradient applied enables a facilitated detection of oxidation product with distinct differentiation of neutral constitutionally isomeric products.

3.2.3. Summary of detected products of oxidized (A)XOS using negative ion MS. Table 2 summarizes the detected and assigned Fenton oxidation products of $A^{2+3}XX$, as example, in both gradients using negative ion MS in resolution mode. In contrast to the basic gradient I, where both neutral and acidic products achieved appropriate signal strength, the buffered gradient revealed lower sensitivity due to ion suppression by NH_4HCO_2 . However, isobaric oligosaccharides bearing carboxyl groups on different positions could be exclusively identified in the extracted ion chromatograms using the buffered eluent. In contrast to the previously reported method in which

high resolution was required, the differentiation of near isobaric products with a mass difference of $\Delta m \geq 0.03$ Da (e.g. $Xyl_3-CH_2O + 2O-2H$ with m/z 413.09 vs. Xyl_3 with m/z 413.13) was achieved already by the more sensitive resolution MS.²¹ As anticipated, the studied (A)XOS oxidation revealed a greater product variety, because of their numerous chain cleavage options. The majority of product types identified in this study are in accordance with products observed during hydroxyl radical-induced oxidation of arabinotriose, amylose, and amylopectin.^{20,30}

The assembled data suggest that resolution HILIC-MS² is a powerful technique for the rapid and precise structural identification of neutral and acidic oligomers derived from polysaccharide oxidation. Nevertheless, to characterize the complete product profile of hydroxyl radical-induced degradation, both gradients presented in this paper need to be applied. The basic gradient I enables the investigation of neutral oxidation products in combination with an MS scan of acidic products, whereas the buffered gradient II enables a distinct identification of acidic products and their structural variations.



Table 2 Fenton-induced oxidation products of $A^{2+3}XX$ identified by negative ion UPLC-MS in resolution mode as $[M - H]^-$ after 24 h oxidation. A BEH Amide column was used in combination with both gradient systems (ACN/H₂O gradient I with NH₃ additive and gradient II with buffered 60 mM ammonium formate eluent system at pH 8)^a

n^b	5	4	3	2	1
Neutral products					
Pentose _n (Pent _n)	677.21 ^{d*}	545.17*	413.13*	281.09*	149.05*
Pent _n -2H	675.20*	543.16*	411.11*	279.07*	147.03*
Pent _n -CH ₂ O	647.19	515.15			
Pent _n -CH ₂ O-2H	645.18	513.14			117.02
Pent _n -C ₂ H ₄ O ₂		485.15			89.03
Pent _n -C ₂ H ₄ O ₂ -2H					87.01
Acidic products					
<i>Pentose acid series: xylonic, xylaric, and arabinuronic acid^c</i>					
Pent _n + O	693.20*	561.16*	429.13*	297.08*	165.04*
Pent _n + O-2H	691.19*	559.15*	427.12*	295.07*	163.03*
Pent _n + O-4H	689.17	557.13	425.09	293.05	161.01
Pent _n + 2O	709.20	577.16*	445.12		
Pent _n + 2O-2H	707.18*	575.14*	443.10*	311.06*	
Pent _n + 2O-4H		573.13	441.09	309.05	
Pent _n + 3O-2H	723.18	591.14	459.10	327.06	
Pent _n + 3O-4H				325.04	
<i>Tetrose acid series: onic, uronic, and aric acids</i>					
Pent _n -CH ₂ O + O	663.19	531.15	399.11	267.07	135.03
Pent _n -CH ₂ O + O-2H	661.18	529.14	397.10	265.06	133.02
Pent _n -CH ₂ O + O-4H				263.04	
Pent _n -CH ₂ O + 2O-2H	677.17	545.13	413.09	281.05	149.01*
Pent _n -CH ₂ O + 2O-4H			411.08	279.04	
<i>Glyceric acid series</i>					
Pent _n -C ₂ H ₄ O ₂ + O	633.18	501.14	369.10	237.06	105.20
Pent _n -C ₂ H ₄ O ₂ + 2O	649.18				
Pent _n -C ₂ H ₄ O ₂ + 2O-2H	647.16	515.12			
Pent _n -C ₂ H ₄ O ₂ + 3O-2H				267.04	
Pent _n -C ₂ H ₄ O ₂ + 3O-4H	661.14				
<i>Glyconic acid series</i>					
Pent _n -C ₃ H ₆ O ₃ + O					75.01
Pent _n -C ₃ H ₆ O ₃ + 2O-2H			353.09		89.01
<i>Formic acid series</i>					
Pent _n -C ₄ H ₈ O ₄ + O-2H			323.06		

^a Experimental values of m/z (all within ± 2 mDa of theoretical values) with distinct peaks or $\geq 5\%$ intensity relative to the base peak of the respective fraction (neutral: m/z 663.199; acidic: m/z 193.035). An asterisk denotes products with $\geq 20\%$ relative signal intensity. ^b Indicates the number of total sugar units. ^c The names of the monosaccharides are given according to the oxidized unit of the oligosaccharides. ^d Starting material.

3.2.4. Differentiation of isomeric lytic pentose acid products by MS². The identification of carbonyl groups (apart from C1 at the reducing end) is crucial, since they are a clear indicator of polysaccharide oxidation. Thereby, the C=O

location can indicate if it was formed *via* a lytic or a non-lytic process. In this study, MS² analysis revealed interesting details about the location of carbonyl groups in the different evaluated oligosaccharides. As an example, the oxidation product m/z 559.15 can be attributed to deprotonated Pent₄ + O-2H, and was found in Fenton oxidized X5 and A²⁺³XX under loss of one pentose unit (for fragmentation pattern, see Fig. 4). Generally, the product contains either

(i) one uronic acid group (from oxidation of one of the pentose units at C5, hence -CH₂OH \rightarrow -COOH), which is then already fully responsible for + O-2H; or

(ii) one carboxylic acid at the downstream end's C1 (forming xylonic acid, hence -CHO \rightarrow -COOH), plus an additional carbonyl group to account for the -2H. Due to the indiscriminate nature of HO[•], various isomeric products would be expected for this option depending on the order of oxidation, and whether C1-oxidation or carbonyl formation was responsible for the chain scission.

Based on the acidic nature of the possible products, a clear separation of isomeric products could not be achieved by the basic gradient I. However, gradient II impeded acceptable MS² spectra due to extensive ion suppression caused by the eluent buffer. Only gradient I allowed for sufficient signal abundance for collision induced fragmentation analysis. Consequently, the MS² pattern of m/z 559.15 from both starting materials X5 and A²⁺³XX may represent a mixture of isomeric products. However, the fragments found allow for differentiation and identification of formed products. As shown for X5 oxidation, the pattern of Y_n fragments (m/z 429.12, 297.08, and 165.04 for $n = 3, 2$, and 1) produced through the loss of sugar units from the non-reducing end indicated the presence of a carboxylic acid group at the downstream end sugar unit, namely as a xylonic acid (Xyl1A) from C1 oxidation. In addition, these fragments corresponding to Xyl_(n-1)Xyl1A ($n = 1-3$) clearly showed that the additional carbonyl group is not located on the downstream end xylonic acid unit for this product. The neutral loss of -130 Da from the parent ion to the Y₃ fragment (m/z 559.15 \rightarrow 429.12) corresponds to an eliminated ^{oxo}Xyl unit, which indicates a product with a carbonyl group at the non-reducing end. Furthermore, the fragment m/z 129.02 can be assigned to the mentioned ^{oxo}Xyl as B₁ fragment itself.

There is some indication that the alternative possibility (i), namely a xyluronic acid-bearing oligosaccharide Xyl₃XylA, is at least a minor part of the product mixture for X5 oxidation: most notably, the fragment ion m/z 409, which could correspond to Z₃ of Xyl₃XylA (namely Xyl₂XylA-H₂O). And some fragments that could also be from ^{oxo}XylXyl₂Xyl1A, but then need two collision induced scission events to explain them, *vs.* only one if originating from Xyl₃XylA: m/z 263 being B₂ for the latter, and B₃/Y₃ for the former, as well as its dehydrated product m/z 245. Possibly, these fragments have contributions from both product isomers.

In the case of A²⁺³XX oxidation, the respective product with carbonyl at the non-reducing and C1-oxidized reducing end as found for X5 could be identified due to the occurrence of analogous fragment ions. However, additionally, the MS² spec-



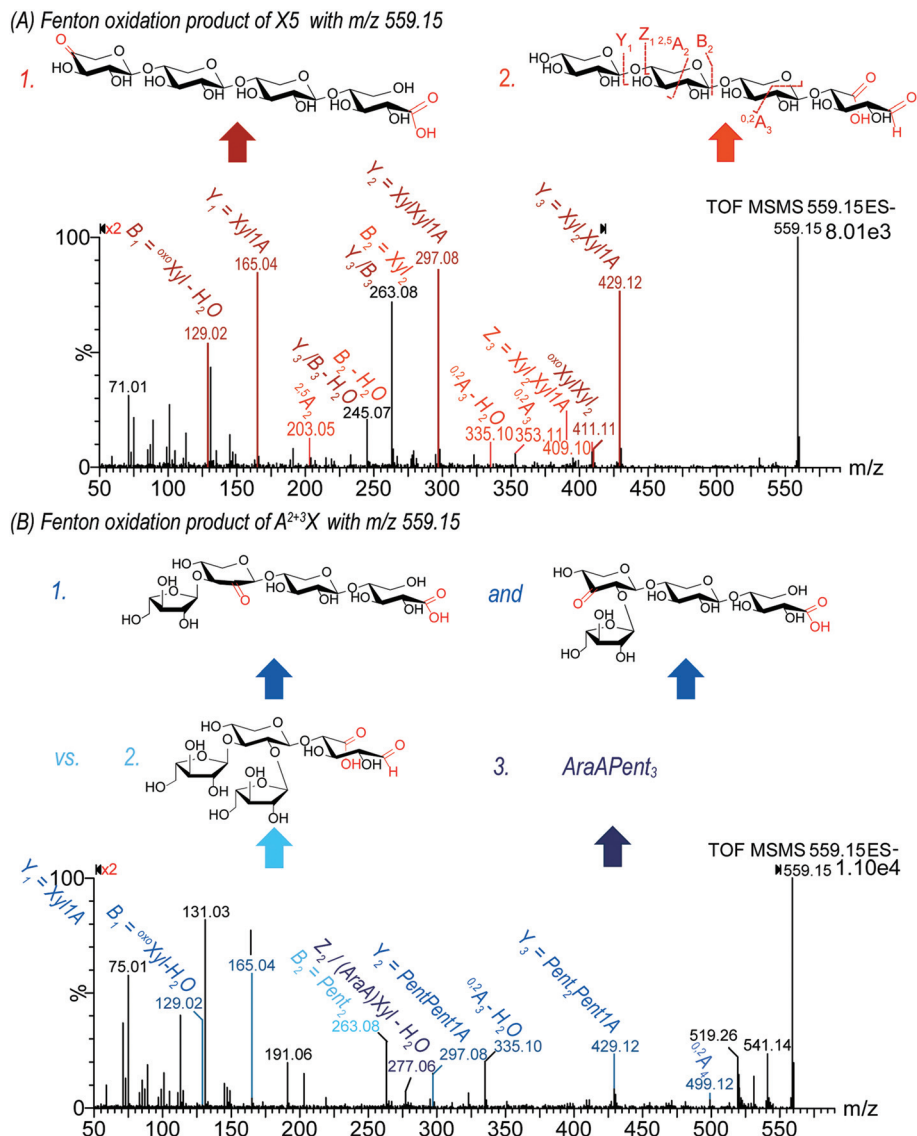


Fig. 4 MS^2 spectra of the oxidation product $Pent_4 + O-2H$ with m/z 559.15, found for Fenton oxidized (A) X5 and (B) $A^{2+3}XX$ to determine the location of carbonyl and carboxylic acid groups. Fragments highlighted in color are identifying the structural unit indicated with an arrow of the same color.

trum contained fragment m/z 277.06, which could be assigned to an arabinose unit bearing a carboxylic acid group at C5 (arabinuronic acid, AraA). Hence, for the Fenton-induced oxidation of both X5 and $A^{2+3}XX$, m/z 559.15 likely represents a mixture of $^{oxo}Xyl(Pent)_2Xyl1A$, and $Pent_3Xyl1A$ from lytic C5 oxidation at the unit next to the reducing end, whereas for $A^{2+3}XX$, some C5 oxidized arabinose products (AraA) $Pent_3$ is additionally present. Thus, MS^2 analysis is a powerful technique to determine the oxidation site on the oligomers.

Regarding reaction pathways, having the carbonyl group only at the non-reducing end strongly suggests that its formation is part of the lytic step, namely, on the example of X5 oxidation, C4-oxidation with scission of the last β -(1 \rightarrow 4)-linkage, leading to $^{4oxo}XylXyl_3$ (see Fig. 5). Introducing a carbonyl group is, in contrast to C1 oxidation, not selective – unless

it is formed through lytic action, then it must be localized at the former glycosidic linkage position, which is C4 for the xylan backbone, and C2 or C3 in the case of Ara sidechains. Under H_2O_2 conditions, it is known that reducing ends oxidize relatively easily to the corresponding onic acid species.²¹ This is in accordance with earlier studies, where MS^2 analysis was also used to localize the oxidation at the reducing or non-reducing end of glucose oligomers.^{16,21}

3.3. Characterization of the complex product mixture from Fenton oxidized AX polymer

Wheat arabinoxylan (AX) was oxidized by Fenton-induced oxidation using 100 μM $FeSO_4$, 500 μM ascorbic acid (AA), and 500 mM H_2O_2 . Subsequently, (A)XOS were produced by *endo*-1,4- β -xylanase from *Cellvibrio mixtus*, which selectively hydro-



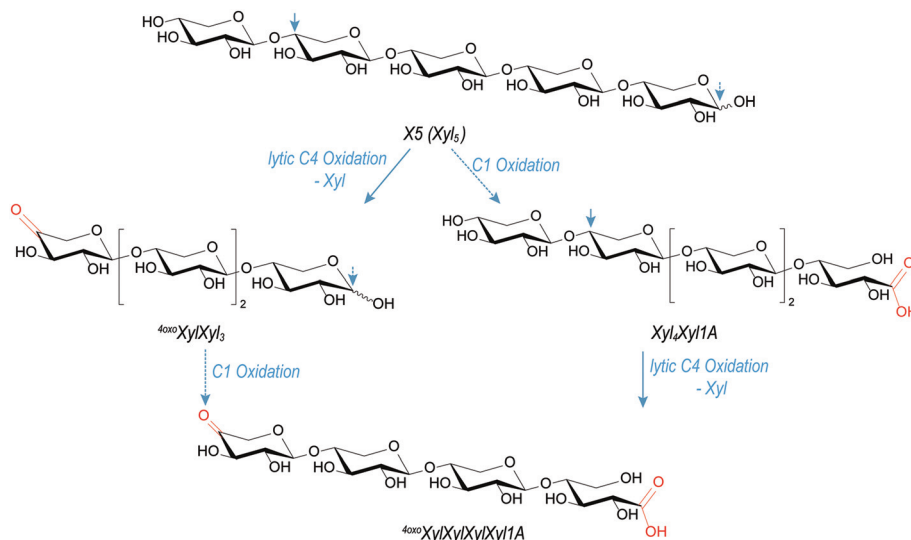
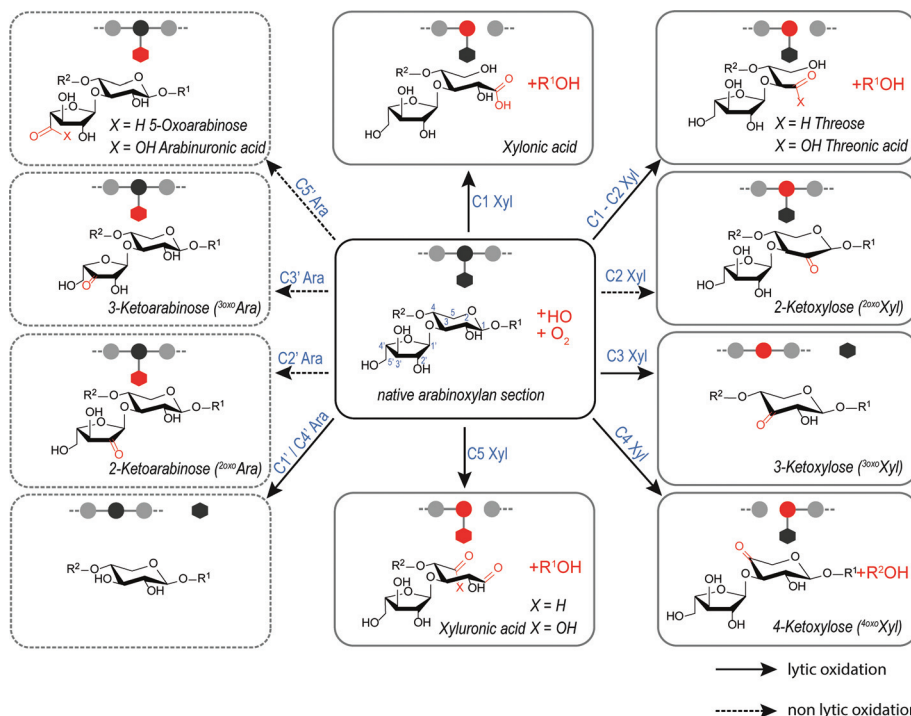


Fig. 5 Suggested likely pathways for the formation of m/z 559.15 product from Fenton-induced X5 oxidation through lytic C4 oxidation (HO^\bullet , O_2) and reducing end C1 oxidation (H_2O_2), with loss of the non-reducing end unit, forming $^{4\text{oxo}}\text{XylXylXyl1A}$ as product.

lyzes certain 1,4- β -glycosidic bonds within the β -(1 \rightarrow 4)-xylan backbone. Consequently, the product profile of oxidized AX derived oligosaccharides was characterized by hydrophilic interaction UPLC-MS². Scheme 1 provides structures, names, and abbreviations of expected AX oxidation products originating from HO^\bullet -attack on xylose and arabinose carbons C1–5

along the arabinoxylan backbone, based on our results from the (A)XOS model system experiments presented above. Reaction shown are categorized as lytic and non-lytic reactions, where lytic reactions cause chain scission within the polysaccharide backbone and hence depolymerization and loss of viscosity. In this study, loss of Ara sidechains is not



Scheme 1 Overview of expected AX oxidation products in the presence of molecular oxygen mediated by hydroxyl radicals (HO^\bullet), which attack at any of the five carbons of xylose (plane \square) and arabinose (hashed \square). Starting from α -(1 \rightarrow 3)-linked L-arabinofuranosyl (Araf) that is part of β -(1 \rightarrow 4)-xylan backbone, both lytic (plane arrows), which introduces chain scissions, and non-lytic oxidation (hashed arrows) are shown. Threose can be formed by a lytic cross-ring cleavage (C1–C2).



classified as lytic, as their loss has a negligible impact on molecular weight and viscosity of the polysaccharide material.

3.3.1. AX polymer oxidation – UPLC analysis with basic eluent. As similarly observed for the Fenton oxidized oligosac-

charides above, significant amounts of both acidic and neutral oxidation products were produced by the HO[•] induced degradation of arabinoxylan (see Fig. 6). The acidic fraction (F1) in Fig. 6B and C demonstrates that primary oxidation products were dominating, namely *m/z* 165.04, 297.08, 429.12, 561.17,

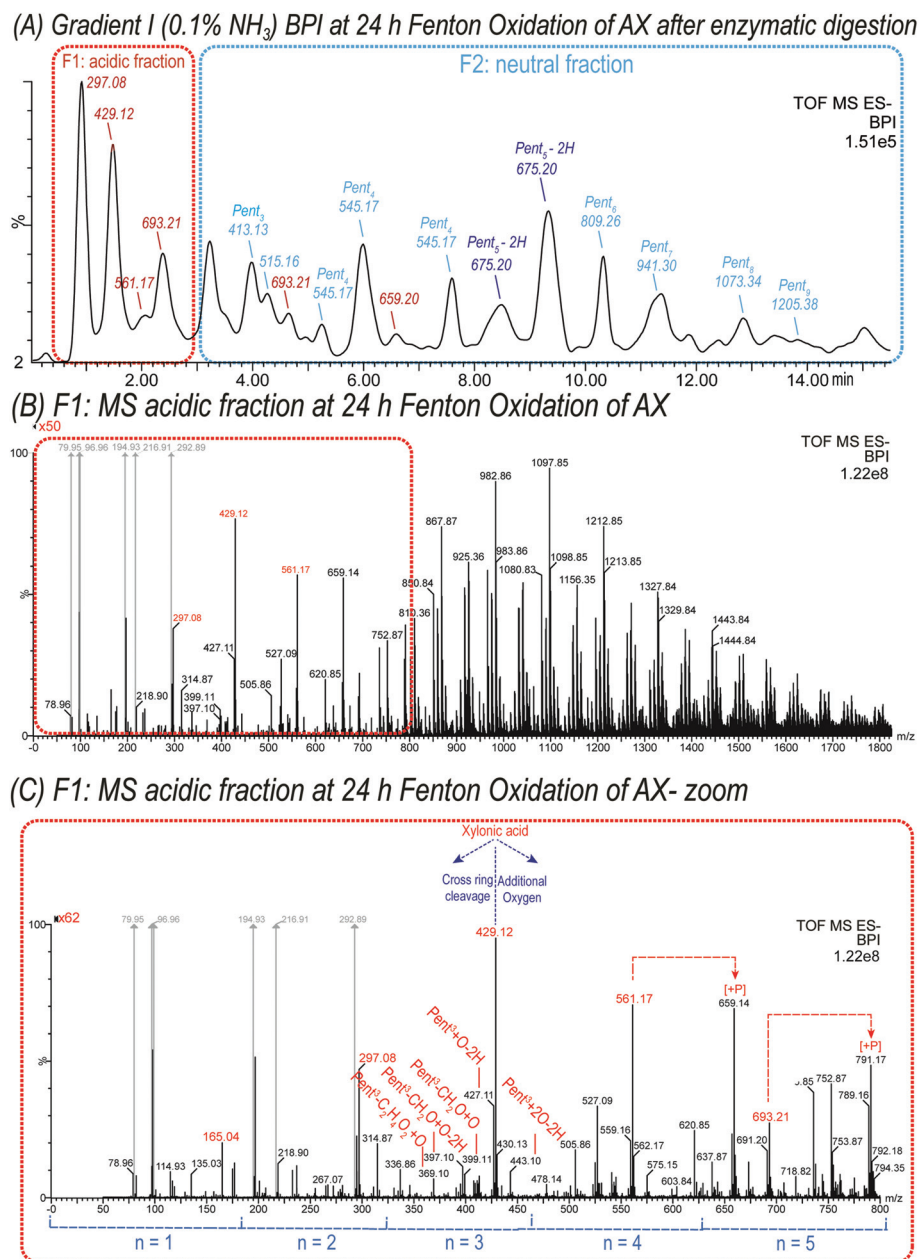


Fig. 6 (A) UPLC-MS base peak chromatogram of released oligosaccharides from 1 mM oxidized wheat arabinoxylan solution oxidized with 100 μ M FeSO₄, 100 μ M ascorbic acid and 50 mM H₂O₂ for 24 h at room temperature. The oxidized AX was digested with *endo*-1,4- β -xylanase to obtain diagnostic oligosaccharides suitable in *M_w* for UPLC-MS² analysis in negative ionization mode using a BEH Amide column and ACN/H₂O gradient (0.1% NH₃). Peaks are labelled with their respective base peak *m/z* and with *n* = number of monosaccharide units in blue for arabinoxylan oligomers Pent_{*n*}. (B) MS spectrum of acidic products obtained from the highlighted chromatogram region F1 in (A). Signals are labelled with *m/z*, and carboxylic acid-bearing oligomers are labelled in red. Signals indicated in grey are assigned to common background contamination ions such as buffer salt adducts. (C) MS spectrum of acidic products F1, zoomed in for the range of *m/z* 50 to 800. Carboxylic acid-bearing oligomer are labelled in red, products with additional oxygens and cross-ring cleavages are indicated. Several Xyl + O products, namely *m/z* 561.17, 693.21, 825.25, 957.29, 1089.33, and 1221.37 were additionally detected as the respective phosphate adduct (Δm = 97.96 Da) with *m/z* 659.13, 791.17, 923.21, 1055.25, 1187.29, and 1319.33, respectively (marked with [+P]).



and m/z 693.21 corresponding to $\text{Pent}_n + \text{O}$, and hence to xylonic acids $\text{Pent}_{(n-1)}\text{Xyl}1\text{A}$ with $n = 1, 2, 3, 4$, and 5 monosaccharide units, respectively. These xylonic acid-bearing oligomers are most likely the result of direct lytic C1-oxidation, although it cannot be excluded that an unknown proportion originates from other lytic action (such as lytic C4-oxidation) that releases an intact reducing end, which then oxidized later due to the high concentration of H_2O_2 in solution (50 mM), as observed previously for β -glucan oxidation under similar Fenton conditions.¹⁶ The second most abundant signals belong to $\text{Pent}_n + \text{O}-2\text{H}$. As discussed earlier on the case of oligosaccharide oxidation, this product class represents lytic C4-oxidation plus secondary C1-oxidation of the reducing end due to the high H_2O_2 concentration, (or lytic C1-oxidation plus a carbonyl somewhere from secondary oxidation), vs. simply the C5-oxidation of Ara-side chains to arabinuronic acid (AraA; only 1 oxidative event necessary), or a mixture of both. In contrast to the oligosaccharide oxidation case, where it is unlikely that two independent HO-radical steps are involved, here, with the oxidation of the polysaccharide, the enzymatic sample preparation is responsible for the production of smaller oligosaccharides, making the detection of AraA-bearing oligosaccharides more likely.

In addition, oxidation products carrying carboxylic acid groups at both C1 + C5 positions along the oligomer could also be detected, as well as the respective acidic cross-ring cleavage products such as tetrose-, glyceric-, and glyconic acid-bearing oligomers (e.g. $\text{Pent}_3-\text{CH}_2\text{O} + 2\text{O}-2\text{H}$ with m/z 413.09, and threonic acid $\text{Pent}_2\text{Thr}1\text{A}$ with m/z 399.11) (see Fig. 6 and Table 3).

The neutral fraction (see Fig. 6A) consisted mainly of pentose-oligosaccharides (DP 1 to DP 9), whereas their respective counterparts with one or more oxidized hydroxyl group (oxo- Pent_n) could also be detected, such as oxo- Pent_5 isomers (m/z 675.20) separated using basic gradient I (see Fig. 6A).

3.3.2. Summary of detected and identified products in resolution mode MS. The summary of the detected arabinoxylan derived oxidation products in both gradients using resolution MS mode revealed great similarities to the detected products from oligosaccharide oxidation (see Table 3).

The degree of polymerization obtained by the enzymatic digestion was ranging from DP1 to DP13, which was generally higher than the DP of oligosaccharides evaluated as oxidation model compounds in this study. A possible explanation for the limited production of smaller diagnostic (A)XOS by *endo*-1,4- β -xylanase could be the steric hindrance through side groups. This limitation may be eliminated by the implementation of a selective enzyme mixture, which may simplify the analysis. However, crucial structural information may be lost through a more extensive enzymatic degradation.

Furthermore, it could be observed that the non-selective oxidation of AX yielded lower product diversity than the oxidation of (A)XOS. This observation indicates that AX polymer was oxidized to a lesser extent than the oligomers. This is in line with the chosen oxidation conditions if the reagent con-

Table 3 Selected Fenton-induced oxidation products of AX in the range of DP1 to DP5 identified by negative ion, resolution UPLC-MS as $[\text{M} - \text{H}]^-$ after 24 h oxidation and xylanase digestion. A BEH Amide column was used in combination with both gradient systems (ACN/ H_2O gradient I with NH_3 additive and gradient II with buffered 60 mM ammonium formate eluent system at pH 8)^a

n^b	5	4	3	2	1
Neutral products					
Pentose _n (Pent _n)	677.21*	545.17*	413.13*	281.09	
Pent _n -2H	675.20*	543.16*	411.11*	279.07	
Pent _n -CH ₂ O		515.15*			
Pent _n -CH ₂ O-2H					
Pent _n -C ₂ H ₄ O ₂	617.19	485.15			
Acidic products					
<i>Pentose acid series: xylonic, xylaric, and arabinuronic acid^c</i>					
Pent _n + O	693.20*	561.16*	429.13*	297.08*	165.04
Pent _n + O-2H	691.19*	559.15*	427.12*	295.07	163.03
Pent _n + O-4H	689.17	557.13	425.09	293.05	
Pent _n + 2O					
Pent _n + 2O-2H	707.18	575.14	443.10	311.06	
Pent _n + 2O-4H			441.09	309.05	
<i>Tetrose acid series: onic, uronic, and aric acids^c</i>					
Pent _n -CH ₂ O + O				399.11	
Pent _n -CH ₂ O + O-2H	661.18	529.14	397.10		
Pent _n -CH ₂ O + 2O-2H			413.09*	281.05	
<i>Glyceric acid series^c</i>					
Pent _n -C ₂ H ₄ O ₂ + 2O-2H		515.12*			
Pent _n -C ₂ H ₄ O ₂ + 3O-4H	661.14				
<i>Glyconic acid series^c</i>					
Pent _n -C ₃ H ₆ O ₃ + O			339.09		
Pent _n -C ₃ H ₆ O ₃ + O-2H					
Pent _n -C ₃ H ₆ O ₃ + 2O-2H	617.17	485.13			

^a Experimental values of m/z (all within ± 2 mDa of theoretical values) with distinct peaks or $\geq 5\%$ intensity relative to the base peak of the respective fraction (neutral: m/z 663.199; acidic: m/z 193.035). An asterisk denotes products with $\geq 20\%$ relative signal intensity. ^b Indicates the number of total sugar units. ^c The names of the monosaccharides are given according to the oxidized unit of the oligosaccharides.

centrations are compared relative to the monomer units (AX: 150 mM monomers vs. 50 mM H_2O_2 ; (A)XOS: 5 mM monomers vs. 10 mM H_2O_2). Further oxidation of the AX material might have also produced a similar diversity of products as the (A)XOS.

The assembled data suggest that resolution HILIC-MS² is a powerful technique for the rapid and precise structural identification of neutral and acidic oligomers derived from arabinoxylan oxidation. Nevertheless, for the complete characterization of the product profile of hydroxyl radical-induced degradation, both gradients presented in this paper need to be applied as was the case in the study for β -glucan oxidation.^{16,21}

4. Conclusion & outlook

A complementary analytical approach using hydrophilic interaction UPLC-MS² in negative ion mode and a combination of



two gradient systems enabled the investigation of oxidized (A)XOS formed through selective TEMPO-mediated oxidation and non-selective Fenton-induced degradation.

Overall, our method application on branched heterooligosaccharides broadens the application field of the established method enabling the comprehensive characterization of functionalized branched heterooligo- and heteropolysaccharides. The in-depth analysis of acidic oxidation products synthesized by TEMPO-mediated oxidation provides mechanistic insights in the selective functionalization of oligosaccharides and polysaccharides. The chosen MS² condition allowed for the differentiation and linkage assignment of constitutionally isomeric native and synthesized C1-oxidation products. The implementation of higher collision energies facilitated the structural elucidation of stable C1-oxidized (A)XOS. Moreover, the localization of newly introduced functional groups by MS² could be shown on the example of a lytic C4 oxidation combined with reducing end oxidation. This analytical investigation enables the detailed elucidation of reaction pathways occurring during polysaccharide functionalizations such as Fenton oxidation. The implemented enzymatic sample preparation strategy revealed appropriate size distribution and improved signal strength in the HILIC-MS² analysis enabling the comprehensive characterization of Fenton-induced arabinoxylan oxidation. In further findings, MS resolution mode was sufficient for the separation of near isobaric products ($\Delta m = \sim 0.05$ Da). This feature prevents misassigning oxidation products due to a lack of resolution. In general, the complementary basic and buffer gradient systems yielded appropriate chromatographic resolution in combination with qTOF MS detection. However, to characterize the complex product mixture of hydroxyl radical-induced degradation, both gradients need to be considered. While gradient I enables the characterization of neutral products, gradient II is crucial to investigate acidic products and their isomeric species.

The analytical approach presented enables a molecular screening of selective and non-selective polysaccharide modifications providing valuable mechanistic insight within the broad field of polysaccharide applications.

Abbreviations

AX	Arabinoxylan
AXOS	Arabinoxylan oligosaccharides
(A)XOS	Xylo- and arabinoxylan oligosaccharides
DP	Degree of polymerization
HO [•]	Hydroxyl radicals
TEMPO	2,2,6,6-Tetramethylpiperidine 1-oxyl
XOS	Xylooligosaccharides

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Conflicts of interest

There are no conflicts to declare.

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