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Polyphosphazene-based nanocarriers for the release of agrochemicals and potential anticancer drugs†

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The synthesis and characterisation of novel polyphosphazene nanocarriers, based on hydrophilic polyalkylene oxide Jeffamine M1000 and hydrophobic steroids with a glycinate linker for pH-controlled release of diosgenin and two brassinosteroids (DI31 and S7) with agrochemical and potential anticancer activity, is hereby described. Polyphosphazenes carrying approximately 17 wt% of DI31 or S7 self-assembled in water to form 120–150 nm nanoaggregates, which showed an excellent plant growth effect on radish cotyledons due to sustained delivery of approximately 30% of the agrochemicals after 4 days. Cytotoxic evaluation showed that all polymers carrying steroids and Jeffamine M1000 resulted in strong to moderate toxicity to MCF-7 cancer cells and were non-toxic to primary human lung fibroblast cells at 0.1 to 0.025 mg mL⁻¹. Thus, DI31 and S7 bearing polymers applied at 10⁻⁴ to 10⁻⁶ mg mL⁻¹ for delivery of recommended DI31 or S7 quantities to crops should be harmless to humans. Particularly, DI31 and S7 bearing polymers with strong cytotoxicity on MCF-7 and non-toxicity on primary human lung fibroblasts, good cell uptake after 6 hours, proper hydrodynamic sizes between 100 and 200 nm, and slow sustained release of cytotoxic drugs (DI31, S7) in acidic conditions might potentiate their accumulation in cancer tissues with good antitumour effects and minor side effects. These results demonstrated that preparation of brassinosteroid bearing polymers is a promising strategy for the preparation of better agrochemicals with reduced pollutant impact on sustainable agriculture and potential anticancer formulations based on analogues of brassinosteroids.

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Introduction

Brassinosteroids (BR), a naturally occurring steroid plant hormone group found in all vegetal organs, regulate plant growth and development by eliciting several physiological effects in combination with other phytohormones.^{1,2} These phytohormones promote xylem differentiation, stem elongation, leaf bending, epinasty, and biosynthesis of nucleic acids, proteins and ethylene, regulate assimilation and allocation of carbohydrates and response mechanisms of plants to biotic and abiotic stress and activate photosynthesis.^{3–5} The content of BR in plants ranges from 10⁻¹ nmol g⁻¹ to 10⁻⁷ nmol g⁻¹, being 100 times smaller than other phytohormones.^{6,7} BR and some synthetic

analogues of brassinosteroids are widely used at concentrations of 5 to 100 mg ha⁻¹ as agrochemicals and pesticides, with a significant improvement in the quality and efficiency of crops.^{8,9} Particularly promising results are observed when the synthetic analogues of brassinosteroids DI31 and S7 are applied at 10–20 mg ha⁻¹ as agrochemicals, with demonstrated increases of 5–30% of crop yields.^{10,11} However, rapid metabolism of BR-based agrochemicals limits the efficacy of DI31 and S7, which results in periodic foliar applications being required to achieve their known benefits.¹¹ The hydrophobicity of DI31 and S7 also complicates their application in agriculture, it being necessary to prepare their commercial formulations as emulsions containing 100 ppm of DI31 in 50 vol% water/ethanol and surfactants (Biobras-16), with limited stability.^{12,13} To avoid the rapid metabolism of DI31 or S7 and provide a constant supply of these compounds with increased solubility in water, they would be incorporated in different polymer-based systems such as micelles, nanoparticles, vesicles, dendrimers or other carriers that protect them and assure controlled release for a longer period of time.^{14,15} However, relatively fewer drug delivery systems have been aimed at sustained release of agrochemicals

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than pharmaceuticals.¹⁶ In this sense, an ideal drug delivery system for application in agriculture should provide sustained release of the agrochemical, achieving a high drug loading with retained activity, good solubility and stability in water under normal usage conditions (pH 7.0, 25 °C), together with proper degradation and no toxicity of the polymer matrix and its metabolites in plants and animals.^{17,18} Hence, polyphosphazenes, a group of degradable synthetic polymers with highly tuneable composition and properties, appear to be promising candidates to this end. On the other hand, diosgenin was chosen as a model-drug to assess the pH-promoted delivery of steroids from polyphosphazene nanocarriers.¹⁹ This steroid sapogenin is a cheap substrate widely used for synthesis of steroids, corticosteroids and the agrochemical DI31,²⁰ providing a suitable model for the bulky steroid substituents on polyphosphazenes. Diosgenin itself has shown good antidiabetic, hypocholesterolemic, antithrombotic and anticancer activity, but very low aqueous solubility and reduced oral bioavailability limit its medical applications.^{19,21} Similarly, some brassinosteroids and synthetic analogues of brassinosteroids have displayed potent antiproliferative effects on different cancer cells with reduced haemolysis and negative effects on non-cancer cells.^{22,23} This is the reasoning behind designing an efficient polyphosphazene-based drug delivery system for diosgenin and brassinosteroids.

Polyphosphazenes, also named poly(organo)phosphazenes, are organic–inorganic hybrid polymers with a characteristic $[-R_2P=N-]$ repeating unit in which different nucleophiles attached to the phosphorus atom tailor the properties of the polymer.^{24,25} Then, different polyphosphazenes have been synthesised for wide diverse applications such as tissue engineering and drug delivery agents in biomedicine,^{25–34} catalysts,³⁵ flame retardants,³⁶ high performance polymers³⁷ and other materials.^{38,39} Polyphosphazenes are particularly attractive since their properties, architectures, functionality and degradability can be fine-tuned by precise control of the polymer composition and synthetic pathway (degree of polymerisation, distribution and ratio of hydrophobic/hydrophilic substituents, structure, etc.).^{24–38} Another advantage of the application of polyphosphazene-based biomaterials is that they degrade to phosphates and ammonium salts alongside the organic substituents, thus being benign degradation products if innocuous substituents have been selected.^{25,27,38} Therefore, smartly designed polyphosphazenes carrying brassinosteroids might fit the requirements to sustainably deliver agrochemicals once applied to plants with minimized environmental impact and increased plant growth effect.

In this research, five polyphosphazene-based nanocarriers for delivery of diosgenin and two brassinosteroids (DI31 and S7) with agrochemical and potential anticancer activity were synthesised by living cationic polymerisation of $Cl_3PNSiMe_3$ to obtain poly(dichloro)phosphazene, which was further functionalised *via* post-polymerisation substitution with the steroids and hydrophilic Jeffamine M1000. Polyphosphazenes carrying diosgenin were synthesised with two degrees of polymerisation and steroid content to optimize the synthesis of the polymers, to study the effect of the average chain length and chemical composition on

the nanoaggregates' hydrodynamic diameters and drug release behaviour. Sustained release of diosgenin from the nanocarriers in acidic medium *via* hydrolysis of ester bonds encouraged us to prepare pH-labile polyphosphazenes with DI31 and S7 for agrochemical applications. Controlled delivery of covalently linked DI31 and S7 from their nanocarriers increased their plant growth stimulant activity as assessed using *in vitro* radish cotyledon bioassay. Further, the cytotoxicity of all polymers to MCF-7 cancer cell line and primary human lung fibroblast cells was evaluated.

Materials and methods

Materials

An amine capped polyetheramine copolymer (PEO–PPO–NH₂) with an M_n of 1000 g mol⁻¹ and an ethylene oxide/propylene oxide ratio of 19/3, tradename Jeffamine M1000, was donated by Huntsman Performance Products and used as received. The diosgenin [(25R)-5-spirosten-3β-ol]⁴⁰ and synthetic analogues of brassinosteroids (25R)-3β,5α-dihydroxyspirostan-6-one (DI31) and (22R,23R)-22,23-epoxy-3β,5α-dihydroxystigmastan-6-one (S7) were kindly supplied by the University of Havana, Cuba. Triethylamine was distilled and stored over molecular sieves under argon. All other chemicals and solvents were purchased from Sigma-Aldrich and used without prior purification. All glassware was dried overnight in an oven at 120 °C prior to use. Spectra/Por 3 cellulose dialysis membranes (Spectrum Laboratories Inc., CA, USA) with a molecular weight cut off (MWCO) of 3.5 kDa were used for purification of the synthesised polymers.

Synthesis of steroid-glycine-Boc

The synthesis of steroid-glycine-Boc and deprotected steroid-glycine-NH₂ (diosgenin-glycine-NH₂ (1), DI31-glycine-NH₂ (2) and S7-glycine-NH₂ (3)) was carried out as previously reported, with minor modifications.^{26,27} The preparation of diosgenin glycinate is briefly described in the ESI.†

Synthesis of monomer trichlorophosphoranimine (Cl₃P=N–Si(CH₃)₃)

The monomer *N*-(trimethylsilyl)-trichlorophosphoranimine (Cl₃P=N–Si(CH₃)₃) was synthesised according to a reported procedure with slight modifications (ESI†).^{38,41}

Synthesis of the polymers

The synthesis of the poly(dichloro)phosphazene precursor was conducted *via* living cationic polymerisation of monomer trichlorophosphoranimine with triphenylphosphine dichloride.^{24,28,30,38} The synthesis of polymer **P1** is described in detail in the ESI.† For polymers **P2–P6** the ratio of monomer to initiator, and the ratio of substituent steroid to Jeffamine M1000 were adjusted differently (Table 1). All reactions were performed in a glovebox under an inert atmosphere.

In vitro drug release studies

The *in vitro* release studies of diosgenin, DI31 and S7 were performed in phosphate buffered saline (PBS, pH 6.0) solution



Table 1 Composition, yield and molecular weight of polymers P1–P6

| Sample | I:M | DP | S:J | Y (%) | M_n^{tht} (kg mol ⁻¹) | M_n^{exp} (kg mol ⁻¹) | D^{exp} |
|--------|------|-----------------|-----|-------|--|--|------------------|
| P1 | 1:25 | 23 ^a | 1:1 | 37 | 38 | 10 | 1.37 |
| P2 | 1:25 | 22 ^b | 1:2 | 38 | 42 | 15 | 1.47 |
| P3 | 1:50 | 34 | 1:1 | 35 | 76 | 12 | 1.31 |
| P4 | 1:25 | 20 | 1:2 | 34 | 43 ^c | 14 | 1.35 |
| P5 | 1:25 | 22 ^b | 1:2 | 30 | 43 ^c | 13 | 1.50 |
| P6 | 1:25 | 23 ^a | 0:2 | 49 | 51 | 11 | 1.44 |

Initiator to monomer ratio feed composition (I:M). Degree of polymerisation estimated by ¹H NMR (DP). Steroid to Jeffamine M1000 ratio feed composition (S:J). Theoretical number average molecular weight (M_n^{tht}). Experimental number average molecular weight and polydispersities measured by GPC against polystyrene standards (M_n^{exp} , D^{exp}).

at 25 °C. To this end, 2.0 mL of polymers P1–P5 (2.5 mg mL⁻¹) in PBS at pH 6.0 were placed in dialysis cups (MWCO 3.5 kDa, Slide-A-Lyzer mini dialysis devices, ThermoScientific, USA) and immersed in 40 mL of the release medium (PBS, pH 6.0) at 25 °C and stirred at 150 rpm. The release medium was replaced every required time point, and analysed with UV spectroscopy (diosgenin $\lambda_{\text{em}} = 280$ nm, DI31 and S7 $\lambda_{\text{em}} = 300$ nm). Release experiments were conducted in triplicate. Calibration curves of the steroids (ESI⁺) allowed quantification of the delivered drug (diosgenin $\epsilon_{280} = 688$ M⁻¹ cm⁻¹, DI31 $\epsilon_{300} = 1693$ M⁻¹ cm⁻¹, S7 $\epsilon_{300} = 1290$ M⁻¹ cm⁻¹).⁴²

In vitro agrochemical tests

The agrochemical bioactivity as a plant growth enhancer of the synthesised polymers carrying the brassinosteroid analogues DI31 and S7 (P4 and P5) was evaluated *in vitro* on radish (*Raphanus sativus*) plants. The increased weight of radish cotyledons was used to assess the agrochemical activity of the tested compounds. Radish seeds were sterilised with sodium hypochlorite solution prior to germination on wet filter paper in the dark at room temperature for 3 days.⁴³ Then, the hypocotyls were discarded and the cotyledons weighed and transferred to Petri dishes with 5 mL aqueous solutions of polymers P4, P5, DI31, S7, and P6 (polyphosphazene carrying only Jeffamine M1000) at 10⁻¹ to 10⁻⁷ mg mL⁻¹, and water (control). The cotyledons were allowed to grow for another 3 days and weighed again. The experiments were performed in triplicate and 10 cotyledons were used for each sample.

Cytotoxicity tests

The cytotoxicity of the samples against the human breast adenocarcinoma cell line MCF-7 and the non-cancerous primary human lung fibroblast cell line HLF was evaluated using the 2,3-bis(2-methoxy-4-nitro-5-sulphophenyl)-2H-tetrazolium-5-carboxanilide assay (XTT) (Cell Proliferation Kit II (XTT), Sigma-Aldrich). Cells were cultured in Dulbecco's modified Eagle's medium (DMEM)-high glucose supplemented with 10% fetal bovine serum, 1% penicillin–streptomycin and 1% L-glutamine. The media and supplements except L-glutamine (PAA Laboratories) were purchased from Sigma-Aldrich. Cells were grown in a humidified 5% CO₂ atmosphere at 37 °C. In all experiments, 70–80% confluence cells were used. Cells (1 × 10⁴ cells per well) in complete growth medium were seeded

into 96-well culture plates. The day after, the medium was changed to serum free medium with P1–P6 aggregate dispersions at different concentrations. The cells were incubated for another 48 hours, the medium was changed to full growth medium and 50 μL of XTT reagent was added to each well. Incubation was continued for another 3 hours at 37 °C in a humidified atmosphere with 5% CO₂ and 100% humidity. The absorbance was measured at 490 nm using a GloMax[®] Multimode Microplate Reader (Promega). Three independent experiments (with each sample in triplicate) were performed, and the cell viability was normalised to the untreated control. The cell viability values were analysed with Origin 2015 (Microcal Origin, OriginLab, MA, USA).

Cell uptake

Fluorescent dye safranin O ($\lambda_{\text{excitation}} = 495$ nm, $\lambda_{\text{emission}} = 587$ nm) was encapsulated in the polyphosphazene aggregates P1–P5 for fluorescence imaging. To this end, 1 mg of polyphosphazenes P1–P5 and 0.1 mg of safranin O were dissolved in 0.3 mL of MeOH and stirred for 1 hour at room temperature in darkness. After that, 1 mL of deionised water was added and the mixtures were further stirred for another 4 hours at room temperature in darkness. Lyophilisation afforded red coloured solids (safranin-loaded polyphosphazenes SAF-P1, SAF-P2, SAF-P3, SAF-P4 and SAF-P5).

For the cell uptake, cells were grown overnight in 8 well glass bottom μ-slides. The day after, the medium was changed to a serum free medium (control) or safranin-loaded polyphosphazene aggregate dispersions at 0.1 mg mL⁻¹. The cells were incubated for 6 hours and subsequently counterstained with 1 μg mL⁻¹ Hoechst 33342 (Fluka) for 10 minutes. Fluorescence imaging was performed on primary HLF non-cancer cells and MCF-7 cancer cells using an Olympus IX73 inverted microscope with the CY3 channel for the SAF-P1, SAF-P2, SAF-P3, SAF-P4 and SAF-P5 aggregates ($\lambda_{\text{excitation}} = 550$ nm, $\lambda_{\text{emission}} = 565$ nm), while the DAPI channel was used for the cell nuclei marker Hoechst 33342 ($\lambda_{\text{excitation}} = 345$ nm, $\lambda_{\text{emission}} = 455$ nm).

Statistics

Statistical evaluation of data was carried out using Statgraphics Plus 5.1, Professional Edition. Results were assessed using one-way analysis of variance (ANOVA), the Tukey *post hoc* test for between group comparisons, and multiple comparison and Kruskal–Wallis tests at a 95% confidence level ($p = 0.05$). Mean values with no significant differences are indicated with the same letter ($p > 0.05$). Significantly different means are marked with the same letter but different numbers following the letter ($p < 0.05$). Mean values with no letter used as a marker stand for significantly different means ($p < 0.05$).

Characterisation

Characterisation by ATR-FTIR spectroscopy was performed on a PerkinElmer Spectrum 100 FT-IR spectrophotometer using an ATR accessory, with 32 scans and 4 cm⁻¹ resolution from 4000 to 650 cm⁻¹. UV-vis spectra were obtained with a PerkinElmer Lambda 25 UV/vis spectrophotometer using quartz cuvettes.



^1H , ^{13}C and $^{31}\text{P}\{^1\text{H}\}$ NMR spectra were recorded at 298 K using a Bruker Avance 300 spectrometer operated at 300 MHz, 75 MHz and 121 MHz, respectively. CDCl_3 was used as an internal reference for ^1H and ^{13}C NMR measurements (^1H NMR signals were referenced to $\delta = 7.26$ ppm; ^{13}C NMR signals were referenced to 77.16 ppm),⁴⁴ while 85% phosphoric acid was used as an external standard for $^{31}\text{P}\{^1\text{H}\}$ NMR spectra. An Attached Proton Test (APT) ^{13}C NMR experiment was carried out to facilitate carbon assignment by separating carbons according to their number of attached protons (CH and CH_3 signals positive, C and CH_2 signals negative) with a single experiment, which is more sensitive than traditional $^{13}\text{C}\{^1\text{H}\}$ NMR. TopSpin 3.5 pl 7 (Bruker BioSpin GmbH) software was used for NMR spectra processing. Molecular weights were estimated using a Viscotek GPCmax gel permeation chromatograph (GPC) using a PFG column from PSS (Mainz, Germany) (300 mm \times 8 mm, 5 μm particle size), equipped with a Viscotek TDA 305 Triple Detector Array (Malvern, Germany), and calibrated with polystyrene standards from PSS. Samples were eluted with DMF containing 10 mM LiBr at a flow rate of 0.75 mL min^{-1} at 60 $^\circ\text{C}$. Dynamic light scattering (DLS) determinations were carried out on a Malvern Zetasizer Nano ZS (Malvern Instruments Ltd, UK) with a detection angle of 173 $^\circ$ and a 4 mW He-Ne laser operated at 633 nm for backscattering measurements. The samples were prepared at 1 mg mL^{-1} in deionised water and filtered through a 0.45 μm nylon film syringe filter and determinations of hydrodynamic diameters (d_h) were performed in triplicate using a folded capillary cell DTS1070 at 25 $^\circ\text{C}$. The critical micelle concentrations (CMC) of the **P1–P5** samples were determined in deionised water and PBS (pH 7.4) *via* the fluorimetry detection of a pyrene probe added to the samples.^{45,46} **P1–P5** samples were prepared at 1.3 to 0.000151 mg mL^{-1} in water or PBS. 70 μL of pyrene dissolved in acetone was added to each sample until a final pyrene concentration of 6×10^{-7} M was achieved. The samples were stirred overnight at room temperature in darkness. The samples were analysed by fluorimetry, with the emission wavelength fixed at 390 nm and excitation spectra recorded from 330 to 360 nm.⁴⁵ Calorimetric studies of the samples were conducted on a TA Instrument Q10 differential scanning calorimeter (DSC) using aluminium pans, with a sample weight of approximately 5 mg and a heating rate of 10 $^\circ\text{C min}^{-1}$ under a nitrogen flow of 20 mL min^{-1} . The samples were cooled and heated from -80 $^\circ\text{C}$ to 400 $^\circ\text{C}$. Atomic Force Microscopy (AFM) images (10 $\mu\text{m} \times 10 \mu\text{m}$ and 2 $\mu\text{m} \times 2 \mu\text{m}$) were taken with an MFP 3D-Stand Alone AFM (Asylum Research) with the cantilever OMCL-AC160TSA of Olympus, at a resonant frequency of 300 kHz and a spring constant of 26 N m^{-1} , a 50–70% set point and a scan rate of 1 Hz. An 80 μL droplet of a 1 mg mL^{-1} aqueous dispersion of the polymers was deposited on a silicon wafer and spin coated at 40 Hz for 6 s. Transmission Electron Microscopy (TEM) micrographs were recorded with a Jeol JEM-2011 FaSTEM (Jeol Ltd, Tokyo, Japan) operated at 100 kV. A drop of polymer dispersions in deionised water (1 mg mL^{-1}) was placed on a Pioloform coated 300 Mesh Cu grid (Plano GmbH, Germany). Excess solution was eliminated with filter paper; the samples were negatively stained

with a drop of uranyl acetate aqueous solution at 1%, and dried several hours before measurements.

Results and discussion

Synthesis and characterisation of polymers **P1–P6**

Polyphosphazenes with diosgenin as a model-drug and Jeffamine M1000 were first synthesised with a ratio of initiator to monomer of 1:25 and 1:50, and a ratio of substituents diosgenin to Jeffamine M1000 of 1:1 and 1:2 (Table 1). These preliminary experiments permitted us to assess the efficiency of post-polymerisation functionalisation with steroid molecules (bulky groups) on the poly(dichloro)phosphazene backbone, the effect of the hydrophilicity of the synthesised polyphosphazene (based on the Jeffamine M1000 content) on the aggregation in aqueous dispersions *via* hydrodynamic parameters and the effect of the steroid content on the size of the dried nanoaggregates using AFM and TEM imaging.

The synthetic approach followed in this work and the structures of the obtained polymers are presented in Fig. 1.

Living cationic polymerisation of trichlorophosphoranimine initiated with triphenylphosphine dichloride in solution (CH_2Cl_2) at room temperature allowed us to obtain linear poly(dichloro)phosphazenes with a controlled polymerisation degree (20 to 23 for **P1**, **P2**, **P4**, **P5**, and **P6** and 34 for **P3**) adjusted through the initiator to monomer ratio, and low polydispersities of 1.31–1.50 for the prepared polymers, consistent

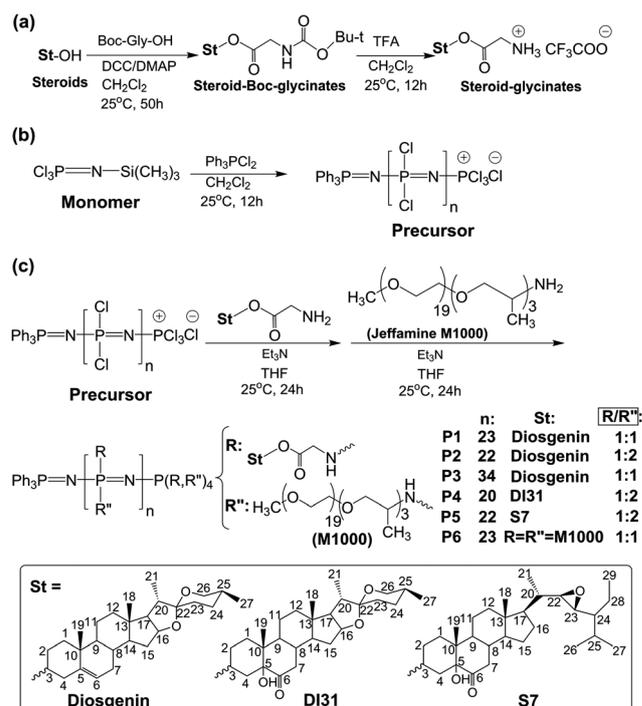


Fig. 1 Scheme of the synthetic route and structures of prepared polyphosphazenes **P1–P6**. (a) Synthesis of steroid glycinates (steroid-glycine- NH_2), (b) polymerisation of trichlorophosphoranimine to obtain poly(dichloro)phosphazene, (c) post-polymerisation substitution of chlorine atoms with steroid-glycine- NH_2 and Jeffamine M1000.



with previous reports (Fig. 1b).^{24,28,38} Different steroid-glycine-NH₂ and Jeffamine M1000 (PEO-PPO-NH₂) were later introduced *via* post-polymerisation substitution of the chlorine atoms in poly(dichloro)phosphazene, firstly introducing the bulky steroid groups and later the linear PEO-PPO-NH-substituent (Fig. 1c). Complete chlorine substitution of the poly(dichloro)phosphazene ([NPCl₂]_n), required to avoid later uncontrolled polymer degradation and hydrolysis due to highly labile P-Cl bonds,³⁸ was confirmed using ³¹P{¹H} NMR spectroscopy. Thus, the ³¹P{¹H} NMR spectra of polymers **P1–P6** only showed broad peaks at 0.5–1.0 ppm, associated with Jeffamine M1000 used as a substituent,^{24,26,28,33,38} while no characteristic peak of the P-Cl bond was observed (ESI†).²⁶ The presence of the hydrophilic Jeffamine M1000 substituent in the synthesised polymers **P1–P6** makes them suitable to be dispersed in aqueous solutions, allowing their later application as carriers for controlled release of agrochemicals. The presence of characteristic aromatic protons of the (C₆H₅)₃P=N- end group allowed the estimation of the attained degree of polymerisation (DP) for polymers **P1–P6** using ¹H NMR spectroscopy, which was expected to be near to 25 or 50 from the monomer to initiator feed in the mixture reaction. The integration of aromatic protons (C₆H₅)₃P=N- at 7.60 ppm of **P1–P6** was approximately 15/(20–23) or 15/(34) with respect to integration of -O-CH₂-CH₂-O- and CH₃O- protons of Jeffamine M1000 at 3.62–3.63 and 3.36 ppm, respectively. The degree of functionalisation with the two substituents (steroid-glycine-NH₂ and Jeffamine M1000) was similarly estimated (ESI†).²⁸ It is observed in Table 1 that the DP of polyphosphazene with diosgenin and Jeffamine M1000 was in good agreement with the initiator to monomer ratio feed composition for the reaction when the polymers were prepared with 25 repeating units (polymers **P1**, **P2**, and **P6**). However, polymer **P3** prepared with an initiator to monomer ratio of 1:50 only reached an estimated DP of 34, which is consistent with the results of Wilfert *et al.*²⁴ when conducting phosphine-mediated polymerisations. The polyphosphazenes carrying DI31 or S7 and Jeffamine M1000 (**P4** and **P5**) were synthesised with an initiator to monomer ratio of 1:25, allowing us to attain a DP of 20–22. As observed from ¹H NMR data (ESI†), the steroid to Jeffamine M1000 mole ratios in the feed (1:2) and in the obtained polymers were also in good agreement. The molecular weights of polymers **P1–P6** determined by GPC calibrated against linear polystyrene standards deviated by 2–4 orders of magnitude from the expected values

calculated for the polymers, as previously reported and ascribed to the different hydrodynamic volume of the branched polyphosphazenes and linear polystyrene standards.²⁸ Then, GPC measurements were used in this research as a guide of molecular weights for similarly related polyphosphazenes and to check the polydispersities of prepared polymers **P1–P6**.

Amphiphilic co-substituted polyphosphazenes might self-assemble or self-aggregate at molecular and supramolecular levels in a micelle-like array with a core formed of hydrophobic moieties and a shield of hydrophilic groups in contact with water molecules, as a result of favoured intramolecular and intermolecular interactions of hydrophobic groups and van der Waals and hydrogen bond interactions of hydrophilic groups and water molecules of the solvent.^{30,39,47,48} Furthermore, the hydrophilic Jeffamine M1000 shell, with a similar structure to poly(ethylene glycol), is expected to prolong the blood circulation time and to protect the lipophilic steroids in the micelle core of **P1–P5** polymers in aqueous media for potential anti-tumoral use.⁴⁹ The average hydrodynamic diameters (*d_h*) in water and PBS obtained using dynamic light scattering, and the sizes of dried nanoaggregates from AFM micrographs (*d_{AFM}*) and TEM micrographs (*d_{TEM}*) of polymers **P1–P6** are shown in Table 2.

P6 showed an average hydrodynamic diameter corresponding to predominant individual hydrated polymer chains in water (Table 2 and Fig. SI-39 and SI-45 in the ESI†). It might be due to the composition of **P6**, formed only for the hydrophilic Jeffamine M1000 bonded to the P-N backbone. **P1** and **P3** polymers with the same ratio of hydrophilic to hydrophobic substituents (a diosgenin to Jeffamine M1000 ratio of 1:1) exhibited similar average hydrodynamic diameters *ca.* 160–170 nm in water and PBS, even when the DP of **P3** was 1.4 times the one obtained for **P1**. Besides, polymer **P2**, with a higher content of hydrophilic Jeffamine M1000, exhibited an approximately 20% to 34% bigger average hydrodynamic diameter when compared to **P1** and **P3** in water and PBS respectively. However, polymers **P4** and **P5** with a similar DP and steroid to Jeffamine M1000 ratio to **P2** displayed lower average hydrodynamic sizes than **P2**. It is noteworthy that polymers **P1–P6** exhibited a bimodal size distribution by intensity or volume of their hydrodynamic diameters (ESI†), related to the dynamic formation of aggregates with different sizes in water, probably due to the self-assembly process. Particularly, polymers **P2**, **P4** and **P5** showed a unimodal size distribution by volume with sole peaks observed at 9–10 nm, which indicate

Table 2 Hydrodynamic sizes, average AFM and TEM diameters of dried nanoaggregates, and steroid weight contents of polymers **P1–P6**

| Sample | <i>d_h</i> ^a nm (PDI) | <i>d_h</i> ^b nm (PDI) | <i>d_{AFM}</i> nm | <i>d_{TEM}</i> nm | CMC ^a mg mL ⁻¹ | CMC ^b mg mL ⁻¹ | Wt% |
|-----------|--|--|---------------------------|---------------------------|--------------------------------------|--------------------------------------|------|
| P1 | 164 ± 2 ^a (0.21) | 160 ± 3 ^a (0.29) | 58 ± 5 ^{d1} | 53 ± 9 ^{d1} | 0.040 | 0.013 | 26.5 |
| P2 | 200 ± 5 (0.34) | 216 ± 1 (0.27) ^c | 88 ± 3 | 40 ± 7 ^{d2} | 0.0025 | 0.0030 | 16.2 |
| P3 | 169 ± 2 (0.23) | 162 ± 1 ^a (0.26) | 61 ± 6 ^{d1} | 35 ± 8 ^{d2} | 0.037 | 0.017 | 26.6 |
| P4 | 122 ± 1 (0.66) | 101 ± 2 (0.70) | 36 ± 8 ^{d2} | 42 ± 5 ^{d2} | 0.0040 | 0.0043 | 16.3 |
| P5 | 151 ± 2 (0.59) | 163 ± 4 ^a (0.63) | 52 ± 2 ^{d1} | 37 ± 8 ^{d2} | 0.0024 | 0.0035 | 17.6 |
| P6 | 6.0 ± 0.2 ^b (0.27) ^c | 6.5 ± 0.3 ^b (0.40) | — | — | — | — | 0 |

^a Average hydrodynamic diameter (*d_h*) in water by DLS, critical micelle concentration (CMC) in water. ^b Average hydrodynamic diameter (*d_h*) in PBS by DLS, critical micelle concentration (CMC) in PBS. Diameter of dried nanoaggregates by AFM (*d_{AFM}*). Diameter of dried nanoaggregates by TEM (*d_{TEM}*). Steroid weight contents (wt%).



the key contribution of single or a few polymer chains to the self-assembly process of these amphiphilic polyphosphazenes as small aggregates in water. However, further studies are required for a proper understanding of the factors controlling the self-assembly mechanism and the hydrodynamic sizes (d_h) of polyphosphazenes in water. Nevertheless, polymers **P1–P5** formed aggregates in aqueous medium with appropriate sizes (less than 200 nm) for possible medical or agrochemical applications.^{18,50,51}

The dried aggregates of **P1–P5** polymers appeared as aggregates and single rounded particles of approximately 35 to 90 nm when observed using AFM and TEM (Fig. 2 and Fig. SI-37 and SI-38 in the ESI[†]), with significant **P1–P5** shrinkage attributed to water removal during drying and collapse of the hydrophilic Jeffamine M1000 shell.^{49,52} The particle sizes determined using AFM followed the same trend of hydrodynamic sizes estimated in water with DLS.

The critical micelle concentrations (CMC) of polymers **P1–P5** were determined in water and in PBS according to their intended application as agrochemical and as anticancer drug carriers (Table 2 and Fig. SI-46 in the ESI[†]). Interestingly the less hydrophilic polymers **P1** and **P3** exhibited the highest CMC values in water and PBS. However, all the synthesised polymers showed a good ability to form aqueous aggregates with a micelle-like structure characterised by CMC values from 0.0024 to 0.04 mg mL⁻¹.

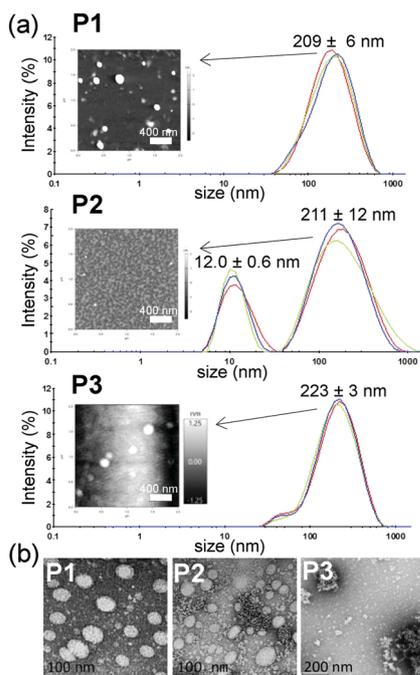


Fig. 2 (a) Hydrodynamic diameter by intensity distribution for polymers **P1–P3** in deionised water (concentration 1 mg mL⁻¹). Further DLS data can be seen in the ESI[†] (Fig. SI-37–SI-45). The inserts show AFM micrographs of polymers **P1–P3** dispersed in deionised water (concentration 1 mg mL⁻¹). (b) TEM micrographs of polymers **P1–P3** dispersed in deionised water (concentration 1 mg mL⁻¹).

Diosgenin and brassinosteroid release studies

In this research, the influence of the diosgenin content by weight (wt%) of polymers **P1** (26.5%) and **P3** (26.6%) compared to **P2** (16.2%), the hydrodynamic sizes of the polyphosphazenes **P1** and **P3** compared to **P2**, and the DP of polymers **P1** and **P2** ($n = 25$) compared to **P3** ($n = 50$) on the release profiles is studied.

All *in vitro* release experiments were carried out at pH 6.0 and 25 °C to simulate the slightly acidic environment found inside the vacuoles in vegetal cells, where the particles are stored once having entered the plant cells *via* endocytosis.^{53,54} It must be noted that hydrolysis of the ester bond in the steroid glycinato moiety and degradation of the polyphosphazene backbone in aqueous medium at neutral pH and 25 °C are minimal or very slow, but significant hydrolysis occurs under acidic conditions.^{25,26} However, *in vivo* vacuolar degradation of substances in plants is mostly enzyme-promoted and the loaded polymers **P1–P5** will suffer hydrolysis faster and more extensively due to esterases, phosphatases and other enzymes.^{55–57} Additionally, acid-promoted diosgenin and brassinosteroid release from the **P1–P5** aggregates would facilitate drug release and accumulation in acidic cancer tissues rather than in normal cells, for potential antitumour therapeutic applications.⁵⁸ The release experiments were extended up to 106 h (approximately 4 days), because the intended *in vitro* activity evaluation of agrochemical bearing polymers **P4** and **P5** will be performed 3 days after application. The *in vitro* release profiles of diosgenin bearing polymers **P1–P3** in PBS (pH 6.0) at 25 °C are shown in Fig. 3a.

The observed trend of the release rates was **P2** > **P3** ≈ **P1**, with a cumulative diosgenin release of ca. 33% (**P2**) and 21% (**P1** and **P3**) after 4 days. The release appeared faster and almost linear during the first 8 h, with approximately half of the total diosgenin released with a slope ranging from 1.34 to 2.5% h⁻¹, and an adjusted *R*-square of 0.98–0.99 (Table SI-1 in the ESI[†]). The more hydrophilic and bigger particles of polymer **P2**, with a lower content of hydrophobic diosgenin, exhibited faster and significantly higher drug release after 4 days than polymers **P1** and **P3**, both with 26.6% of diosgenin. It was expected that the more hydrophilic polymer **P2** with a diosgenin content by weight 40% lower than in **P1** and **P3** and with the capability to form particles with 34% bigger hydrodynamic diameters in PBS will occur with less compact and less dense hydrophobic cores (Fig. 3c), which are more suitable for ester hydrolysis and diosgenin release. On the other hand, polymers **P1** and **P3** with the same diosgenin content and quite similar hydrodynamic diameters of particles in aqueous medium showed similar release profiles and no significant difference of the total diosgenin released after 106 h at a 95% confidence level (approximately 21% of diosgenin contained within **P1** and **P3** was released).

The agrochemical content by weight (wt%) of polymers **P4** and **P5** was found to be 16.3 wt% of DI31 (**P4**) and 17.6 wt% of S7 (**P5**) respectively, as estimated using ¹H NMR data (ESI[†]). The release profile of S7 bearing polymer **P5** is characterised by an initial burst release up to 15.5% in the first 2 h (Fig. 3b). It was observed a fast S7 release rate during the first 8 h deviated



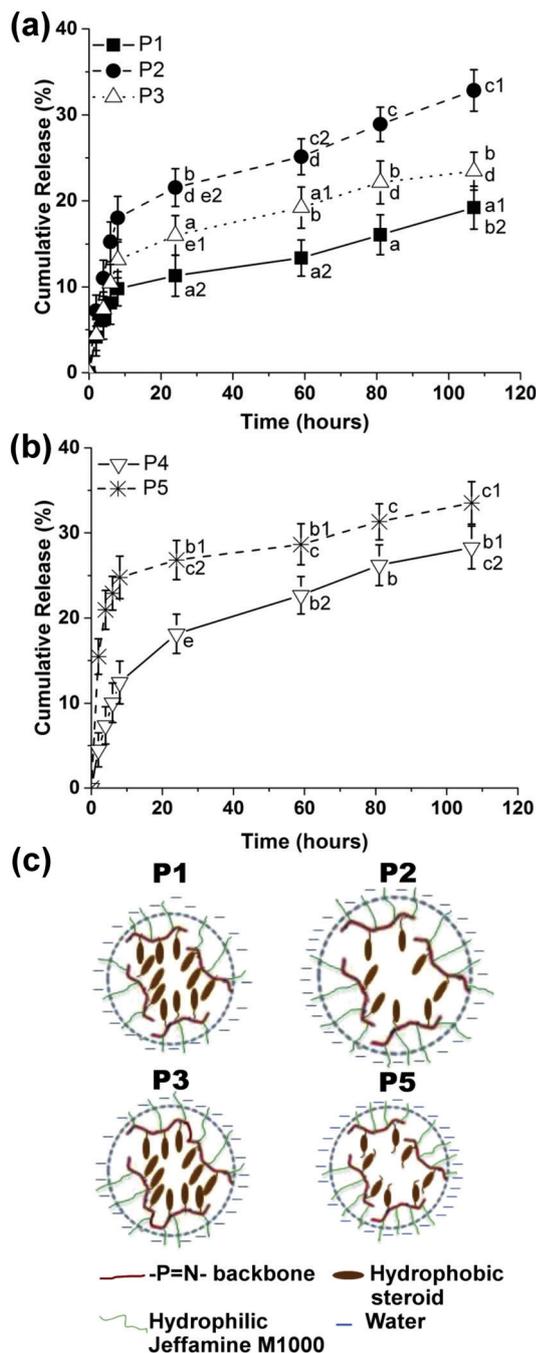


Fig. 3 (a) Cumulative diosgenin release from polymers **P1** to **P3** at 25 °C in PBS at pH 6.0; (b) cumulative steroid release from polymers **P4** and **P5** at 25 °C in PBS at pH 6.0. The amounts of diosgenin and steroids DI31 and S7 released were estimated using a calibration curve for the free drugs in PBS at pH 6.0. (c) Cross-section scheme of the proposed structure of polyphosphazene nanoaggregates **P1–P3** and **P5** in aqueous medium.

of linear behaviour (adjusted R -square 0.9132, Table SI-1 in the ESI[†]), followed by a slower release rate from 24 h to 106 h (Fig. 3b). The initial burst release of S7 from polymer **P5** might be due to the mobility of the lateral chain (carbons C20 to C29) in S7 combined with the superior hydrophilicity of polymers **P4** and **P5**, which aids H_3O^+ and water uptake (required for

generation of highly active acylium ions and further reaction with water during ester hydrolysis)⁵⁹ in the less hydrophobic core of **P5** aggregates (Fig. 3c).^{45,60} However, there is no significant difference of the total agrochemicals released after 106 h for polymers **P4** and **P5** (approximately 31% of DI31 or S7 contained within polymers **P4** and **P5** was released). It is hypothesised that hydrolysis of the ester bond between the steroids and the glycinate linker attaching the steroids to the polyphosphazene backbone determines the steroid release rates for polymers **P1–P5**, because drug release of steroid-grafted celluloses and chitosans was also extended over 3–4 days when carried out in similar conditions.^{15,48} On the other hand, the glycine-Jeffamine M1000 bearing polyphosphazene is known to degrade slower, with approximately 10% of the polymer degraded after 5 days in acetate buffer (pH 5.0) at 37 °C.²⁶

In vitro agrochemical activity

Brassinosteroids and auxin synergistically control vegetal cell division and elongation among other functions at the microscopic level, which is observed macroscopically as plant growth and accretion of biomass.⁶¹ The crosstalk between the brassinosteroid and auxin signaling pathways is exhaustively studied, with the BRI1 kinase brassinosteroid receptor overexpressed by auxin and brassinazole resistant 1 (BZR1) transcription factor of brassinosteroids capable of regulating auxin-controlled genes.⁶¹ It is accepted that the brassinosteroid and auxin pathways are highly interconnected, sharing transcriptional targets and controlling gene expression in plants.⁶¹ The radish cotyledon test, the radish hypocotyl elongation test, and the bending of the lamina joint of rice test are based on detection of brassinosteroid and auxin activities.⁶¹ These bioassays are frequently used to evaluate the agrochemical activity of different compounds as plant growth regulators.⁶¹ Recent advances in computer simulation of the BRI1 brassinosteroid receptor 3D structure and molecular docking studies of brassinosteroid–BRI1 interactions allow *in silico* screening of potential brassinosteroid mimetics.⁶² However, the traditional radish- and rice-based bioassays are still used to properly evaluate the plant growth activity and proper dosage of possible agrochemical candidates. Particularly, the proper concentration of brassinosteroids exogenously applied and the plant developmental stage for brassinosteroid application must be determined to obtain optimal yields.^{8,9}

The results as plant growth enhancers of DI31 bearing polymer **P4** and S7 bearing polymer **P5** against radish (*Raphanus sativus*) plants are shown in Fig. 4.

The methodology employed to evaluate the agrochemical activity of the studied compounds is based on detection of auxin type activity,^{43,61} expressed as the increased weight of radish cotyledons (Fig. 4a). The same agrochemical activity was observed for both polymers **P4** and **P5** when applied at the same concentrations, except at 10^{-3} and 10^{-6} mg mL⁻¹ with **P4** (DI31) being significantly more effective as a plant growth stimulator than **P5** (S7) at these two concentrations (Fig. 4b). The stimulatory effect of polymers **P4** and **P5** was maximal at 10^{-1} and 10^{-2} mg mL⁻¹ concentrations, with a three times increased radish cotyledon weight as compared to radish cotyledons



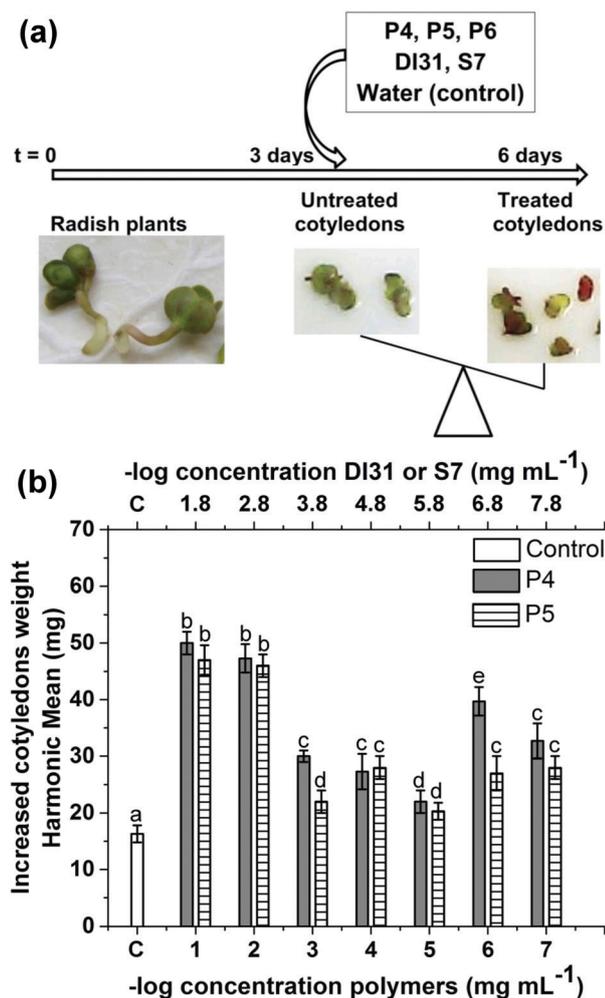


Fig. 4 (a) Scheme of the process to evaluate the agrochemical activity of polymers **P4** (DI31), **P5** (S7), **P6** (Jeffamine M1000), DI31 and S7 using radish cotyledons obtained from radish plants. (b) *In vitro* agrochemical activity expressed in terms of the increased weight of 10 radish cotyledons as a function of the concentration of applied polymers **P4** (DI31) and **P5** (S7) (bottom axis) and of the concentration of DI31 and S7 carried on the applied polymers **P4** (DI31) and **P5** (S7) (top axis), C refers to the control (radish cotyledons treated with water). See the graph showing the agrochemical activity of parent DI31 and S7 and Jeffamine M1000 bearing polymer **P6** in the ESI† (Fig. SI-53).

treated with water (control) (Fig. 4b). Further dilutions of **P4** and **P5** until 10^{-3} to 10^{-7} mg mL⁻¹ concentrations of the polymers still resulted in similar and good agrochemical activity, with almost twice the radish cotyledon weight increase as compared to the control experiment (Fig. 4b). Particularly attractive for further *in vivo* agrochemical application of polymers **P4** and **P5** is the slightly superior plant growth enhancer effect observed at the lowest polymer concentrations of 10^{-6} and 10^{-7} mg mL⁻¹ (content $10^{-6.8}$ and $10^{-7.8}$ mg mL⁻¹ of DI31 and S7) as compared to the agrochemical effect of parent DI31 and S7 evaluated at 10^{-7} mg mL⁻¹ (Fig. 4b and Fig. SI-53 in the ESI†). In the same way, polymers **P4** and **P5** displayed higher agrochemical activity at all concentrations as compared to DI31 and S7 tested at the same brassinosteroid concentration, except when **P4** and **P5**

were applied at 10^{-3} mg mL⁻¹ (content $10^{-3.8}$ mg mL⁻¹ of DI31 and S7) (Fig. 4b and Fig. SI-53 in the ESI†).

The agrochemical activity of parent DI31 and S7 (Fig. SI-53 in the ESI†) showed no significant difference of the plant growth enhancer effect for both the DI31 and S7 agrochemicals when evaluated at equal concentrations. The agrochemical activity of parent DI31 and S7 was diminished with dilutions, but DI31 and S7 still exhibited significant stimulatory activity when applied at 10^{-6} and 10^{-7} mg mL⁻¹ (nanomolar concentrations of synthetic analogues of brassinosteroids and natural brassinosteroids are applied as agrochemicals in crops).^{63,64} On the other hand, no stimulatory effect was observed for the Jeffamine M1000 bearing polymer **P6** at all concentrations. Therefore, the beneficial effects of DI31 bearing polymer **P4** and S7 bearing polymer **P5** observed on radish plants might be due to the sustained release of the incorporated agrochemicals DI31 and S7.

Cytotoxic activity

Diosgenin is known to exert antiproliferative and cytotoxic effects *via* activation of p53 proapoptotic protein and caspase-3 on MCF-7 breast cancer cells, as well as to promote the generation of reactive oxygen species that induce cell apoptosis *via* oxidative stress.¹⁹ Furthermore, diosgenin is capable of inhibiting the STAT3 signalling pathway in human hepatocellular carcinoma cells, which induces apoptosis and cell cycle arrest in the G1 phase *via* caspase-3 activation and PARP cleavage.¹⁹ Diosgenin also downregulates the NF-κB transcriptional activity, matrix metalloproteinase MMP-2 and MMP-9, Janus kinase JNK, protein kinase B AKT and other factors of several cancer cells.⁶⁵ Interestingly, anti-metastatic activity of diosgenin is reported against the human breast cancer cell line MDA-MB-231 and the human prostate cancer cell line PC-3 through suppression of Vav2 protein and VEGF angiogenic factors.¹⁹

On the other hand, studies of brassinolide and epibrassinolide natural brassinosteroids on human breast cancer cells and drug resistant small-cell lung carcinoma cells have shown potent anticancer activity and no cytotoxicity to non-cancer BEAS-2 lung epithelial cells.²³ It is proposed that brassinolide acts on the Wnt signaling pathway of small-cell lung carcinoma cells *via* a reduction of β-catenin-dependent genes related to cell division, protection against apoptosis and metastasis.²³ Similarly to that observed in diosgenin, the anticancer mechanism of action of brassinosteroids is dependent on the generation of reactive oxygen species, inhibition of angiogenic factors, and regulation of the cancer cell cycle and different cancer cell pathways.²² These facts and the cell-type and dose-dependent antitumour activity of these compounds motivated us to study the cytotoxic activity, and MCF-7 and primary HLF cell uptake of the diosgenin and brassinosteroid bearing polymers synthesised (**P1–P5**).

The relative cell viabilities of MCF-7 and primary HLF cells treated with the **P1–P5** polymers are shown in Fig. 5 and Fig. SI-54–SI-57 in the ESI†.

The Jeffamine M1000 bearing polymer **P6** appeared non toxic to MCF7 and HLF cells at 0.1 mg mL⁻¹, with relative cell viabilities of (97 ± 4)% and (94 ± 6)%, respectively (data not shown). The diosgenin bearing polymers **P1–P3** showed



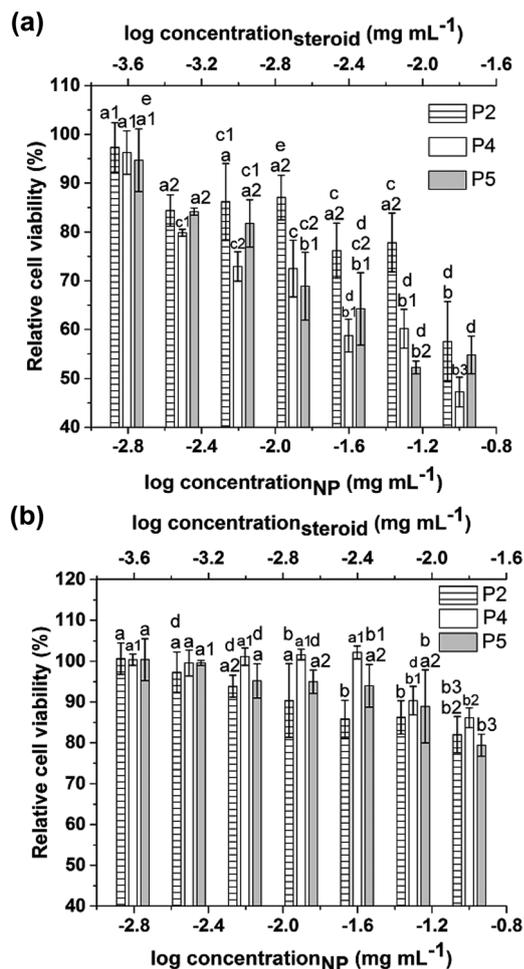


Fig. 5 Relative cell viability of (a) MCF-7 breast cancer cells and (b) non-cancer HLF cells treated with polymers **P2**, **P4**, and **P5**. Further relative cell viability data of MCF-7 and HLF cells treated with polymers **P1**, and **P3**, and parent steroids diosgenin, DI31 and S7 can be seen in the ESI† (Fig. SI-54–SI-57).

moderate to slight anticancer effects (relative cell viabilities of 60–80%) when evaluated on MCF-7 at 0.1 to 0.025 mg mL⁻¹ (a diosgenin content of 0.0266 to 0.00665 mg mL⁻¹ for **P1** and **P3**, and 0.0162 to 0.00405 mg mL⁻¹ for **P2**) (Fig. 5a and Fig. SI-54 in the ESI†), which were similar to the cytotoxic effect observed for parent diosgenin when evaluated on MCF-7 at 0.005 to 0.000625 mg mL⁻¹ (Fig. SI-56 in the ESI†). These polymers exhibited almost no toxicity on non-cancer HLF cells at 0.1 to 0.0015625 mg mL⁻¹, with observed relative cell viabilities of approximately 85–100% (Fig. 5b and Fig. SI-55 in the ESI†), and were similarly non-cytotoxic compared to parent diosgenin at 0.01 to 0.00015625 mg mL⁻¹ (a relative cell viability of 90–100%) (Fig. SI-57 in the ESI†). Interestingly, DI31 bearing polymer **P4** and S7 bearing polymer **P5** exhibited strong to moderate toxicity to MCF-7 cancer cells at 0.1 to 0.00625 mg mL⁻¹ (a DI31 and S7 content of approximately 0.017 to 0.0010625 mg mL⁻¹), with observed relative cell viabilities of 50–70% (Fig. 5a). However, polymers **P4** and **P5** appeared non toxic to HLF cells at all concentrations, with relative cell

viabilities of approximately 90–100% (Fig. 5b). Polymers **P4** and **P5** with an average hydrodynamic diameter of 101 nm and 163 nm respectively, might show extended circulation time and slow renal clearance, with accumulation in cancer tissues due to passive diffusion and an enhanced permeability and retention (EPR) effect.^{66,67} Then, polymers **P4** and **P5** seem to be potential candidates as anticancer drugs. On the other hand, polymers **P4** and **P5** with no toxicity observed at 0.1 to 0.00156 mg mL⁻¹ to primary HLF cells are safe to be used as agrochemicals at concentrations below 0.1 mg mL⁻¹, or particularly below 10⁻³ mg mL⁻¹ for delivery of recommended 10⁻⁴ to 10⁻⁷ mg mL⁻¹ of DI31 or S7 to crops.^{11,13,63,64}

Cell uptake

The uptake of safranin-loaded **P1–P5** aggregates in non-cancer primary HLF cells and MCF-7 cancer cells was assessed using fluorescence microscopy. Samples **SAF-P1**, **SAF-P2**, **SAF-P3**, **SAF-P4** and **SAF-P5** (Fig. 6 and Fig. SI-58–SI-65 in the ESI†) internalized well in both cell lines after 6 hours (safranin O related red fluorescence), and cell nuclei were counterstained with Hoechst 33342 (associated blue fluorescence). The safranin-loaded polyphosphazene aggregates seem to be distributed mostly in the cytoplasm after internalization into cells. Stronger red fluorescence of safranin is observed in the cytoplasm, whereas weaker red fluorescence due to slow diffusion of safranin from the safranin-loaded polymeric aggregates to the cell nuclei is visible for both non-cancer HLF and MCF-7 cancer cells (Fig. 6 and Fig. SI-58–SI-65 in the ESI†).^{49,58}

The experimental results discussed herein demonstrate that it is possible to tune the steroid release rate of polyphosphazene-based polymers with diosgenin and Jeffamine M1000 as co-substituents (**P1–P3**), as a function of the hydrodynamic size and hydrophilicity of the synthesised polymers. Later post-polymerisation functionalisation together with the right selection of substituents allows obtaining polymers with different hydrophilicities that

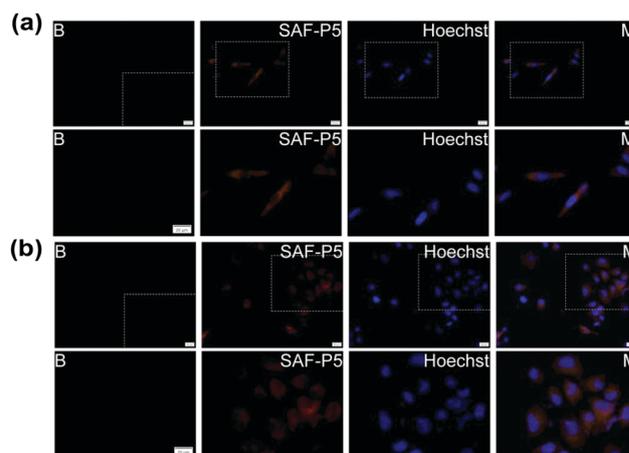


Fig. 6 HLF and MCF-7 cell fluorescence images and slices of cells without particles and Hoechst (B). (a) Primary HLF cells with 0.1 mg mL⁻¹ **SAF-P5** aggregates, 1 μg mL⁻¹ of Hoechst 33342 and merged pictures (M); (b) MCF-7 cells with 0.1 mg mL⁻¹ of **SAF-P5** aggregates, 1 μg mL⁻¹ of Hoechst 33342 and merged pictures (M), scale bars represent 20 μm.



result in aqueous aggregates ranging from nanoaggregates to microaggregates.^{28,29,31} The controlled and partial release of covalently linked DI31 from polymer **P4** and S7 from polymer **P5** caused very good *in vitro* agrochemical activity. Then, it is expected that polyphosphazenes **P4** and **P5** will exert an excellent and extended plant growth stimulator effect on crops using lower quantities than typical exogenous applications of DI31 and S7 when used as agrochemicals.^{68,69} Additional benefits of polymers like **P4** and **P5** are the lower environmental impact associated with their use in agriculture, as they do not need incorporation of harmful additives to ensure colloidal stability and proper dispersion in water as the commercial formulations of DI31, 24-epibrassinolide and other brassinosteroids on the market.^{68,70} On the other hand, diosgenin bearing polymers **P1–P3** exhibited moderate cytotoxicity to MCF-7 and no toxicity to primary HLF cells at medium to high concentrations. DI31 bearing polymer **P4** and S7 bearing polymer **P5** showed strong to moderate cytotoxicity to MCF-7 and no toxicity to primary HLF cells at almost all concentrations tested. Direct injection of **P4** and **P5** particles in solid tumours and/or their selective accumulation in cancer tissues *via* the EPR effect and passive diffusion might enhance the therapeutic effect with a reduction of side effects.^{66,67,71} Besides, polymers **P4** and **P5** are safe to be used in agriculture at concentrations below 10^{-3} mg mL⁻¹. Additionally, degradation of the polyphosphazene backbones to ammonia and phosphates in aqueous medium,³⁸ broad research about various polyphosphazenes in nanomedicine based on their biocompatibility,^{72,73} and no cytotoxic and inhibitory agrochemical effect observed for polymer **P6** put forward a potential application of polymers **P4** and **P5** for chemotherapy treatments and in agriculture.

Conclusions

Six different polyphosphazene-based nanocarriers were prepared for sustained release of diosgenin and the agrochemicals DI31 and S7, *via* controlled living cationic polymerisation and later post-polymerisation functionalisation with the steroids and hydrophilic Jeffamine M1000, an amine capped polyether-amine copolymer. Polyphosphazenes **P1–P3** showed moderate toxicity to cancer MCF-7 and no toxicity to non-cancer HLF cells when evaluated at medium to high concentration. Polyphosphazenes **P4** and **P5** carrying approximately 17 wt% of DI31 (**P4**) and S7 (**P5**) respectively, formed aqueous nanoaggregates that exerted an excellent stimulant plant growth effect on radish cotyledons due to the controlled delivery of the agrochemicals. Cytotoxic evaluation of polymers **P4** and **P5** showed that both polyphosphazenes were toxic to MCF-7 at concentrations above 0.003125 mg mL⁻¹ and harmless to HLF cells at concentrations below 0.1 mg mL⁻¹. MCF-7 and HLF cells showed good cell uptake of all synthesised steroid bearing polymers (**P1–P5**) after 6 hours. Therefore, the **P4** and **P5** polymers applied at 10^{-4} to 10^{-6} mg mL⁻¹ for delivery of the recommended DI31 or S7 quantities to plants should be harmless to humans. The results attained suggest that preparation of polyphosphazenes with

attached brassinosteroids (*i.e.* **P4** and **P5**) is a promising strategy for the synthesis of more efficient agrochemicals with reduced environmental impact, and of promising brassinosteroid-based antitumour candidates. However, exhaustive *in vivo* evaluation of these potential anticancer polyphosphazenes must be performed before their clinical use as antitumoral agents.

Conflicts of interest

There are no conflicts to declare.

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