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# A Janus-inspired amphichromatic system that kills two birds with one stone for operating a “DNA Janus Logic Pair” (DJLP) library†

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Although DNA computing has exhibited a magical power across diverse areas, current DNA logic gates with different functions are always separately operated and can only produce hard-to-visualize output. The fussy/obligatory gates' redesign/reconstruction and the non-intuitive output cause the wastage of time and costs, low efficiency and practicality. Herein, inspired by the ancient Roman mythical God Janus, for the first time, we propose the concept of “DNA Janus Logic Pair” (DJLP) to classify the DNA logic gates with contrary functions into “Positive + Negative” gates (DJLP = Pos + Neg). Based on the biocatalytic property of G-quadruplex DNAzyme (G4zyme) and the luminescence quenching ability of oxidized 3,3',5,5'-tetramethylbenzidine (OxTMB) towards the upconversion (UC) particles, we fabricated a universal amphichromatic platform that kills two birds with one stone for operating a versatile DJLP library. Different from the previous DNA logic systems, the “Pos + Neg” gates of each DJLP in this study were concomitantly achieved *via* the same one-time DNA reaction, which avoided the gates' redesign/reoperation and reduced the operating costs/time of the DNA gates by at least half. Besides, both the amphichromatic outputs (Visual-blue and UC luminescent-green) can be visualized under harmless-NIR, thus bringing greatly enhanced practicality to the method. Moreover, we constructed various concatenated logic circuits *via* logically modulating the G4zyme's biocatalytic property with glutathione, thus enabling the largely improved computing complexity. Furthermore, taking the circuit “YES-INH-1-2 decoder” as the “computing core”, we designed an “antioxidant indicator” with ratiometric logical responses that could recognize the presence of antioxidants smartly (output changed from “10” to “01”), which provided a typical prototype for potential intelligent bio-applications.

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## Introduction

Since the first molecular AND logic gate was pioneered by Prof. de Silva in 1993,<sup>1</sup> the field of molecular computing has flourished and made bioengineers think “outside-the-box”, leading to the development of various smart logical applications.<sup>2–11</sup> As molecular level computers that use biomolecules as building blocks to perform Boolean logic,<sup>12–26</sup> biocomputing has gained significant progress due to excellent biocompatibility, flexibility, and logical stringency.<sup>27–31</sup> In recent decades, different biocomputing systems have been designed that rely on various substrates, such as enzymes, proteins, peptides, antibodies, and DNA.<sup>32–45</sup> For instance, Katz *et al.* constructed a series of

enzymatic logic devices by using the biocatalytic property of different natural enzymes.<sup>31</sup> Fan *et al.* gained unique insights into DNA computing and made great progress in DNA tetrahedron/origami-based logic devices.<sup>43,44,48</sup> Besides, the subtle bridging of different substrates (enzymes and DNA, antibodies and DNA, and so on) for innovative biocomputing has also been reported.<sup>32–34</sup> Among these studies, DNA computing is considered as one of the most outstanding methods owing to its low-cost, predictable-design (Watson–Crick base pair), promising parallel-computing-ability and so on. Moreover, it is recognized that the multifunctional DNA logic systems could provide intelligent solutions for various bio-applications, such as logic-controlled biosensing/bio-diagnosis, drug load/release, genome editing, and stem-cell proliferation.<sup>46–51</sup>

Despite great advancements, there are still significant drawbacks in this area. On one hand, the current DNA logic gates with different functions are always separately constructed according to their operating principles, as shown in Scheme 1A. The fussy/ineluctable sequences' redesign and gates' reconstruction cause the wastage of costs/time and low efficiency. On the other hand, the current DNA gates can only produce

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monochromic (colorimetric) or even invisible (electrochemical, fluorescent, or other) outputs, which largely limit their potential practicality. These drawbacks have been the “Achilles’ heel” of this area. Considering the above background, exploring innovative multichromatic systems for operating a versatile DNA logic library in a facile and low-cost manner is urgently required.

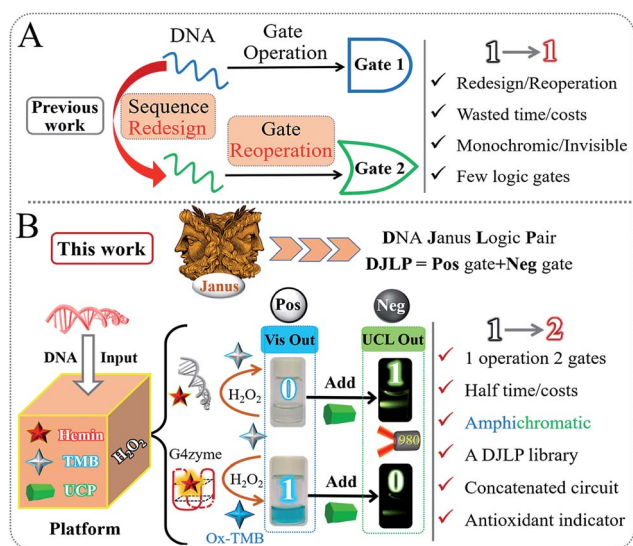
In ancient Roman mythology, Janus is the god of beginnings and endings. It is known that he has two faces that could see the past and the future at the same time, as shown in Scheme 1B. Descending from this mythical concept, there are various such forms in various scientific areas, such as the anode/cathode of a battery, two complementary DNA strands, and hydrophilic/hydrophobic interfaces. Coincidentally, for the logic gates, one characteristic point that researchers usually overlook is that they often coexist with corresponding gates of contrary functions. Herein, inspired by the mythical God Janus, we, for the first time, propose the concept of “DNA Janus Logic Pair” to classify the DNA logic gates with contrary functions into “Positive + Negative” gates (DJLP = Pos + Neg). For instance, the YES, AND, OR and INHIBIT gates are designated as the “Pos” gates, and the NOT, NAND, NOR and IMPLICATION gates with opposite functions belong to the “Neg” ones.

Subsequently, based on the peroxidase-like biocatalytic property of the G-quadruplex DNAzyme (G4zyme) and the luminescence quenching ability of oxidized 3,3',5,5'-tetramethylbenzidine (OxTMB) towards the upconversion particles (UCP),<sup>52–59</sup> we fabricated a universal amphichromatic platform (Scheme 1B) that kills two birds with one stone for operating a versatile DJLP library (YES + NOT, AND + NAND, OR + NOR, INHIBIT + IMPLICATION (INH + IMP), XOR + XNOR, Majority + Minority (MAJ + MIN), and so on). Different from the previous studies, the “Pos + Neg” gates of each DJLP were concomitantly

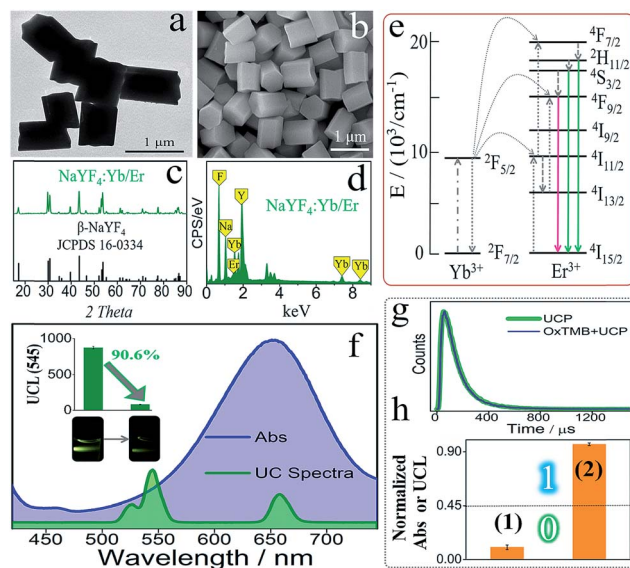
achieved *via* the same one-time DNA reaction, which avoided the gates’ redesign/reoperation and reduced the operation costs/time of the current DNA gates by at least half. Notably, both the amphichromatic outputs (Visual (Vis)-blue, UC luminescent (UCL)-green) were easily visualized by eyes under harmless-NIR (Scheme 1B), bringing greatly enhanced practicality to the method. Besides, the advanced logic devices (a parity checker for natural numbers 0–9, encoder/decoder) were also realized. Moreover, we constructed various concatenated logic circuits [(Pos-INH)/(Neg-INH)] *via* logically modulating the G4zyme’s biocatalytic property with glutathione, thus enabling significantly improved computing complexity. Furthermore, taking the circuit “YES-INH-1-2 decoder” as the “computing core”, we designed an “antioxidant indicator” with ratiometric logical responses, which could recognize the presence of anti-oxidants intelligently.

## Results and discussion

Scheme 1B illustrates the operating principle of the DJLP library. A mixture of hemin, TMB, UCP, and H<sub>2</sub>O<sub>2</sub> was used as the universal “platform-box” and different DNAs acted as the inputs to selectively “open” the box. For the outputs of DJLP, pale-blue and dark-blue colors of Vis outputs were defined as the “0” and “1” states of the Pos gates, respectively. Pale-green and dark-green colors of the UCL outputs were defined as the “0” and “1” states of the Neg gates, respectively. If the non-G4 DNA was the input, the mixture of non-G4 DNA/hemin presented a weak catalytic ability towards the oxidation reaction between TMB and H<sub>2</sub>O<sub>2</sub> due to the lack of G4zyme, thus



**Scheme 1** General operating principle of the DNA logic gates in (A) previous works and (B) this work. “DJLP”, “Pos”, and “Neg” are short forms for “DNA Janus Logic Pair”, “Positive”, and “Negative”, respectively. “UCP” represents “upconversion particles”.



**Fig. 1** From (a) to (e), the TEM image, SEM image, XRD pattern, EDS, and emission mechanism of the as-synthesized UCP; (f) the overlap between OxTMB's absorbance spectra and UCP's emission spectra, inset are the UCL column bars before and after adding OxTMB, and the corresponding photos under 980 nm NIR radiation; (g) luminescence decay curves of UCP and OxTMB/UCP; (h) high/low outputs of this DJLP library.



producing a pale-blue colored solution. After mixing with the UCP solution, it did not influence the UC emission, which remained dark-green under NIR. This phenomenon matches the amphichromatic outputs “Vis = 0, UCL = 1”. On the other hand, if DNA that could form G4 was the input, the formed G4zyme catalyzed the oxidation of TMB, yielding a dark-blue colored OxTMB. After mixing with UCP, it quenched the UC emission (pale-green) efficiently, thus matching the amphichromatic outputs “Vis = 1, UCL = 0”.

The UCP ( $\beta$ -NaYF<sub>4</sub>-doped) used in this DJLP library was synthesized according to previous reports.<sup>57–59</sup> As shown in the TEM and SEM images (Fig. 1a and b), UCP (NaYF<sub>4</sub>: 18%Yb/2% Er) exhibited mono-disperse hexagonal structures, which was further identified by the well-matched XRD patterns (Fig. 1c). Moreover, the length of most of the UCP was about 0.7–0.8  $\mu$ m (Fig. S1–S4†). For UC emission, Er-doped UCP (EDS in Fig. 1d) produced a clear green color (545 nm, <sup>4</sup>S<sub>3/2</sub> to <sup>4</sup>I<sub>15/2</sub> of Er<sup>3+</sup>, see mechanism in Fig. 1e) under a 980 nm NIR radiation, as shown in Fig. 1f. All of the above results proved the successful synthesis of UCP. After this, the reaction between UCP and OxTMB was explored. As shown in Fig. 1f, OxTMB showed absorption from 500 nm to 750 nm, exhibiting a broad overlap with the emission spectra of UCP, which provided the prerequisite for luminescence quenching. After mixing OxTMB with UCP, the green emission decreased remarkably with a quenching efficiency (QE) of 90.6%, which was calculated from the formula  $QE = (UCL_b - UCL_a)/UCL_b \times 100\%$ , where  $UCL_b$  and  $UCL_a$  were the UCL peak intensities (545 nm) before and after adding OxTMB, respectively. For the

quenching mechanism, the following observations were made: (1) the lifetimes (Fig. 1g) of UCP before (green line, 115.2  $\mu$ s) and after adding OxTMB (blue line) were almost unchanged, proving that dynamic quenching was not the main reason;<sup>60,61</sup> (2) after mixing with UCP, OxTMB's absorbance changed slightly (Fig. S5a†) and the reaction product that was catalyzed by non-G4 DNAs/hemin could not quench the UC emission (Fig. S5b†), thus eliminating the static quenching that was caused by the production of a non-fluorescent ground-state complex.<sup>60,61</sup> Thus, we speculated that the luminescence quenching between UCP and OxTMB could be attributed to the inner filter effect, which was also consistent with other recent reports.<sup>60,61</sup> To define the amphichromatic outputs more accurately, normalized Abs (650 nm) or UCL (545 nm) was used to discriminate the high/low outputs and 0.45 was taken as the universal threshold (Fig. 1h).

Based on the above design, we constructed various basic DJLPs initially. For the simplest “YES + NOT” pair (Fig. 2a), a complete G4 (PW<sub>17</sub>)<sup>62</sup> was used as the input. Its operation was similar to the basic principle of the DJLP library that has been described above (Scheme 1B). The visual, UCL photos and column bars under different input variations featured the characteristics of the YES and NOT gates subtly, indicating its favorable performance. It should be noted that the concomitant operation of the YES and NOT gates in the “YES + NOT” pair can also work as a combinatorial circuit. This intuitive amphichromatic system brings enhanced flexibility and practicality. Analogously, the “AND + NAND” pair (Fig. 2b) (PG3 and PG1 that form G4 through hybridization were the

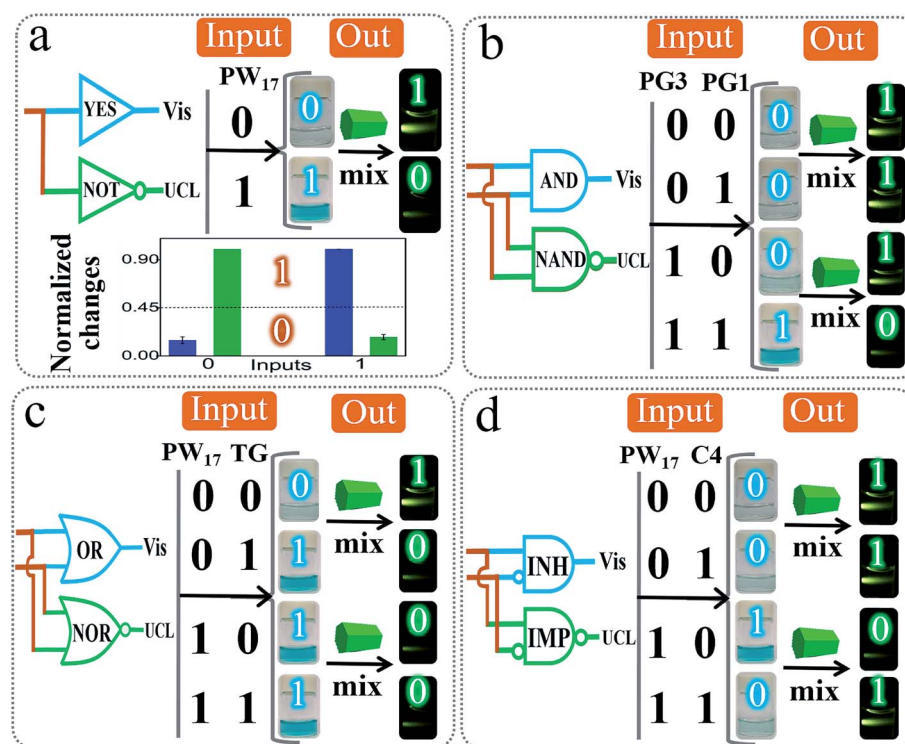


Fig. 2 Operation of 4 basic DJLPs. (a) The YES + NOT pair and the corresponding outputs under different input states: blue (Vis out, YES gate) and green (UCL out, NOT gate); (b) AND + NAND; (c) OR + NOR; and (d) INH + IMP pairs.



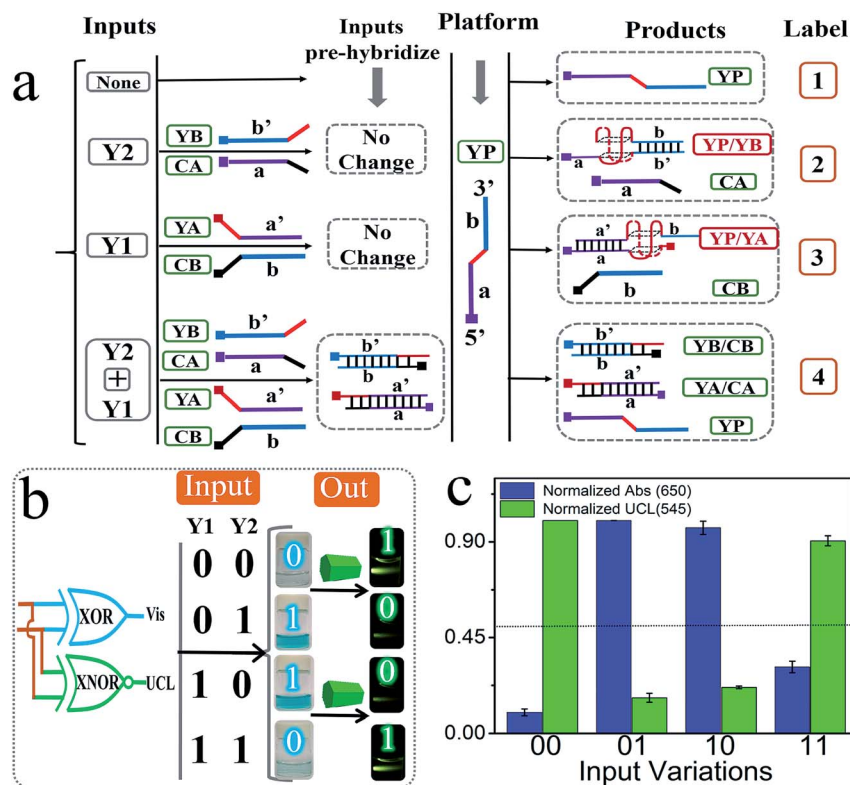


inputs, Fig. S6†), the “OR + NOR” pair (Fig. 2c) (two G4 strands, PW<sub>17</sub>, and T30695 (TG)<sup>62</sup> were the inputs), and the “INH + IMP” pair (Fig. 2d) (PW<sub>17</sub> and its complementary DNA C4 were the inputs, and C4 was the inhibitory element, Fig. S6†) were also fabricated. The output column bars of these DJLPs under different input variations are given in Fig. S7,† indicating their successful operation. The sequences of DNAs used in this study are given in Table S1.†

In addition to the above basic DJLPs, we also constructed the more complicated “XOR + XNOR” and “MAJ + MIN” DJLPs.<sup>54</sup> The XOR and XNOR gates are the core elements in the half-adder/subtractor, parity generator/checker,<sup>55</sup> Feynman gate, *etc.* The detailed operation is illustrated as below: the strand YP that integrated with GGGTGGG in the middle part (red region) was the platform of the “XOR + XNOR” pair. The mixtures of the strands YA + CB and that of YB + CA were the inputs Y1 and Y2, respectively. As shown in Fig. 3a, the purple part (a') of YA (with (TGGG)<sub>2</sub> at the 5' end, red region) could hybridize with the purple part of YP (a) to form the complete G4 (YP/YA). *N*-Methyl mesoporphyrin IX (NMM), the fluorescent dye that could exhibit high fluorescence after binding with G-quadruplex, was applied to confirm G4's formation. The significantly enhanced fluorescence intensity of NMM at 608 nm (Fig. S8a†) in the presence of YP and YA, and the corresponding circular dichroism (CD) peaks (Fig. S8b†) indicated the formation of parallel-G4.<sup>54</sup> Analogously, strand YB can hybridize with the blue part (b) of

YP to form the complete G4 (YP/YB), as shown in Fig. 3. By contrast, the purple part (a) of CA (with CCCA at the 3' end, green region) could hybridize with YA to produce more stable YA/CA, indicating that the pre-hybridization between YA and CA inhibits the formation of G4 (YP/YA), as shown in Fig. S8c.† Similarly, the pre-hybridization between YB and CB will also inhibit the formation of G4 (YP/YB). On the other hand, the mixture of CB and YA (or the mixture of CA and YB) will influence neither the reaction between YP and YA (or YB) nor the formation of G4, as shown in Fig. 3. The PAGE verification can be found in our previous study.<sup>54</sup> After reacting with the “platform-box”, the corresponding Vis and UCL outputs under different input states can be generated, as can be seen in Fig. 3c. The output columns matched the characteristics of the XOR and XNOR gates, indicating the successful operation of the “XOR + XNOR” DJLP.

Till now, all of the above logic pairs are the 2-input DJLPs. To further demonstrate the universality of our amphichromatic platform, the 3-input “MAJ + MIN” DJLP that was composed of the MAJ (majority) and MIN (minority) gates was also fabricated. Both the MAJ and MIN gates play important roles in the full-adder, full-subtractor, molecular voter and other logic devices.<sup>57</sup> For the operation of the “MAJ + MIN” DJLP, the middle region of the three platform DNAs (P1, P2, P3) were integrated with GGGTGGG, and the 3' and 5' ends of each input DNA (DA, DB, DC) were integrated with GGG. Through



**Fig. 3** (a) Operation of the XOR + XNOR pair. Y1 (the mixture of strands YA and CB), Y2 (the mixture of strands YB and CA) were the inputs and YP was the platform (the 5' ends of the DNAs are squared, the poly-G regions are colored in red, and G4 is illustrated with the red-basket structure); (b) schematic operation of the XOR + XNOR pair; (c) normalized Abs and UCL column bars of the XOR + XNOR pair under different input combinations. “01” represents the absence of Y1 and presence of Y2, others ditto.



reasonable design, the right part of DC can hybridize with P3's left part (pale-blue region) and the left part of DC can complement with P2's right part (pale-blue region), while DC does not interact with P1. DA could bridge P1 and P3 together while not interacting with P2, and DB could bridge P1 and P2 together while not hybridizing with P3 (Fig. 4a). The DNA hybridizations were adequately verified by the corresponding PAGE results (Fig. 4c and S9†). Thus, the formation of G4 can be easily modulated by the above DNA hybridizations. With the addition of one input, no G4 will be produced. On the other hand, in the presence of two or more inputs, the complete G4 will form (Fig. 4a). After reacting with the universal amphichromatic platform, the corresponding amphichromatic outputs can be obtained, as seen in Fig. 4b. The output states under different input combinations feature the operation principle of the MAJ and MIN gates properly, indicating the satisfactory performance of the "MAJ + MIN" DJLP.

Apart from the different elementary logic gates, the advanced logic devices with specific functions are also widely used in semiconductor circuits. Herein, we further designed several advanced DNA logic devices based on our universal platform, including a parity checker<sup>18,19,57</sup> that could discriminate the even/odd natural numbers (from 0 to 9), and 2-1 encoder/1-2 decoder<sup>20-24</sup> that could also transform the binary code (data) into the data (code) for molecular information processing. The parity checker could identify the even/odd numbers among the natural numbers less than 10. Unlike the previous parity checkers, this one could generate even (Vis) and odd (UCL) types of outputs simultaneously. Initially, ten decimal numbers were transformed into the 4 bit binary-coded decimal (BCD) numbers,<sup>18,19</sup> as shown in Fig. 5. Four strands, *viz.*, T20, T30, T40, and PW<sub>17</sub>, were assigned as the corresponding 4 bit inputs. As shown in Fig. 5, only when the input

combination corresponds to even decimal numbers (0, 2, 4, 6, 8) is, the Vis output will be "0", which belongs to an even type parity checker. On the other hand, the UCL output will be "0" only when the input combination corresponds to the odd decimal numbers (1, 3, 5, 7, 9), which matches an odd type parity checker.

For the encoder/decoder, it has been reported that an encoder can compress the information for transmission or even storage by transforming the data signal into a code and a decoder possesses the opposite functions.<sup>20-24</sup> Herein, we constructed a 2-1 encoder and a 1-2 decoder, as shown in Fig. S10.† The 2-1 encoder could transform the 2-input bits into the 1-output code.<sup>20-24</sup> The strands T30 and PW<sub>17</sub> were applied as two inputs and the Vis signal was taken as the output. In the presence of T30 and the absence of PW<sub>17</sub> (input "10"), the output will be "0" (pale-blue color) after reacting with the "platform-box". In contrast, in the absence of T30 and in the presence of PW<sub>17</sub> (input "01"), the output will be "1" (dark-blue color). This phenomenon matches the logical principle of a 2-1 encoder.<sup>20-24</sup> Contrary to a 2-1 encoder, a 1-2 decoder could convert the 1-input code into 2-output signals. The strand PW<sub>17</sub> was used as the input; in its absence and presence, the Vis and UCL output will be "0, 1" and "1, 0", respectively. This input-output pattern corresponds to a 1-2 decoder.<sup>20-24</sup>

Semiconductor circuits can perform sophisticated logic computing *via* the integration of the elementary gates.<sup>1-11</sup> Herein, we found that glutathione (GSH) could logically modulate the biocatalytic activity of G4zyme by utilizing its reduction ability ( $\text{GSH} + \text{OxTMB} = \text{GSSG} + \text{TMB}$ ). As shown in Fig. 6a, after adding GSH into the mixture of UCPs/OxTMB, the color of OxTMB changed from dark-blue to colorless. Accordingly, the emission of UCP exhibited significant recovery. Besides, both the high/low Vis and UCL signals can be reversibly

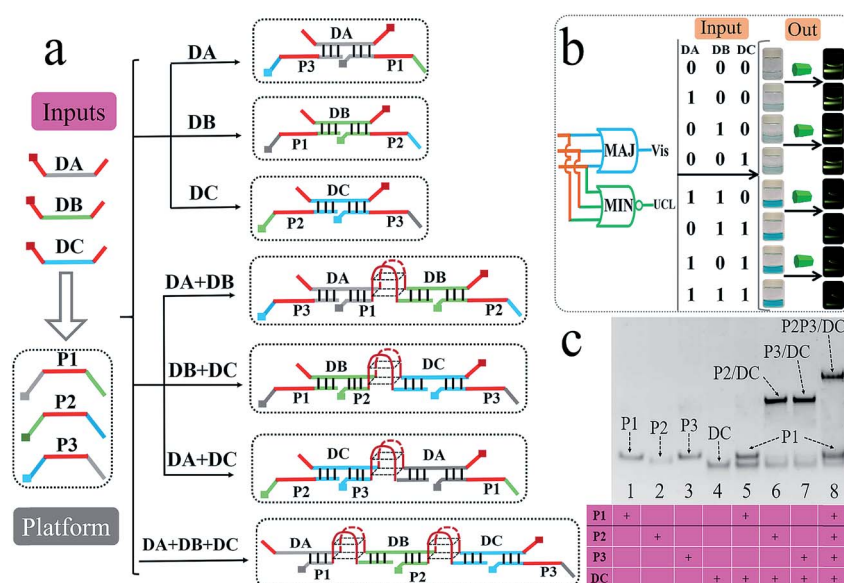


Fig. 4 (a) DNA hybridizations of the MAJ + MIN pair. The strands DA, DB, and DC were the inputs and the mixture of P1, P2, and P3 was the platform (the 5' ends of all the DNAs are squared, the poly-G regions are colored in red, and G4 is illustrated with a red-basket structure); (b) operation of the MAJ + MIN pair; (c) 15% PAGE analysis of the interactions between the strands P1, P2, P3, and DC.

switched several times by alternatively adding GSH/OxTMB under the assistance of centrifugation (Fig. 6b). By utilizing the above phenomenon, a series of concatenated logic circuits (Fig. 6c) were further achieved, including (YES-INH)/(NOT-INH), (AND-INH)/(NAND-INH), (OR-INH)/(NOR-INH), (INH-INH)/(IMP-INH), (XOR-INH)/(XNOR-INH), and (MAJ-INH)/(MIN-INH), in which GSH was the inhibitory element. We presented the output column bars and the truth tables of (YES-INH)/(NOT-INH) and (AND-INH)/(NAND-INH) (Fig. S11 and 12†) as examples to demonstrate the effective operation of these concatenated circuits, which largely improved the computing complexity and also laid a solid foundation for more powerful biocomputing logic networks.

It is recognized that versatile logic systems can provide intelligent solutions to various bio-applications such as logic-programmed biosensing/bio-diagnosis.<sup>42–48,63</sup> Antioxidants (such as GSH) play significant roles in many physiological functions such as helping to trap and neutralize the free radicals, thereby eliminating their harmful effects on the human body. The alternation in the level of antioxidants has been associated with many diseases, thus making the recognition of their absence/presence more significant. Based on the above platform and taking the circuit “YES-INH-1-2 decoder” as the “computing core”, we designed an “antioxidant indicator” with ratiometric logical responses, which could recognize the presence of antioxidants smartly. The first YES gate was used as the initial “Recognize module” to judge different targets (Yes for antioxidant and No for non-antioxidant). If an antioxidant was input, it was transferred to the second “Compute module”, the INHIBIT gate, another input of which was G4. The output of the INHIBIT gate is the mixture of G4 and GSH (the production of

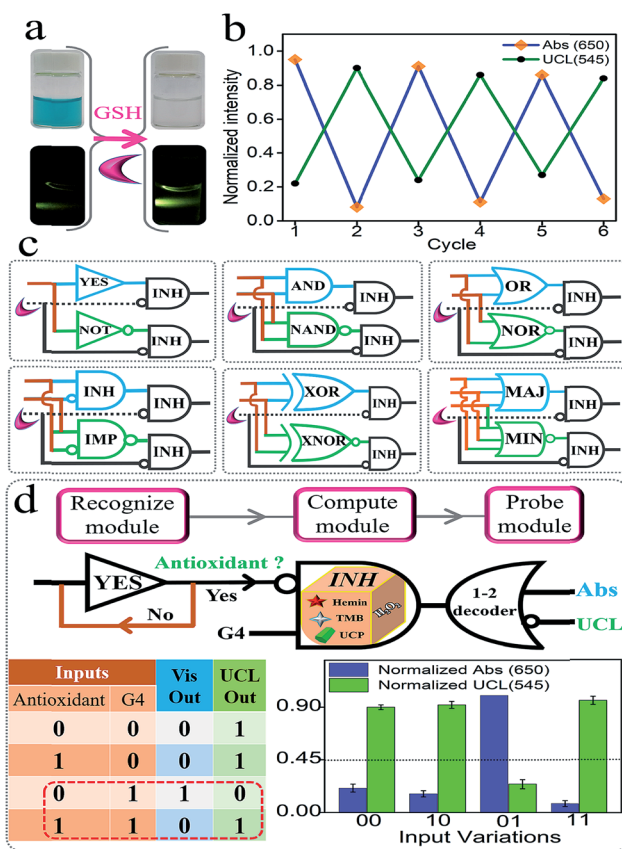


Fig. 6 (a) Photos of OxTMB and UCP before and after adding GSH; (b) normalized changes of Abs<sub>650</sub> and UCL<sub>545</sub> during the alternative addition of GSH and OxTMB assisted by centrifugation; (c) equivalent logic symbols of various concatenated logic circuits; it should be noted that the operation of the concatenated circuits does not need centrifugation as they just need the participation of GSH; (d) operation principle, truth table, and the normalized column bars of the “antioxidant indicator” that was programmed by logic circuit “YES-INH-1-2 decoder”.

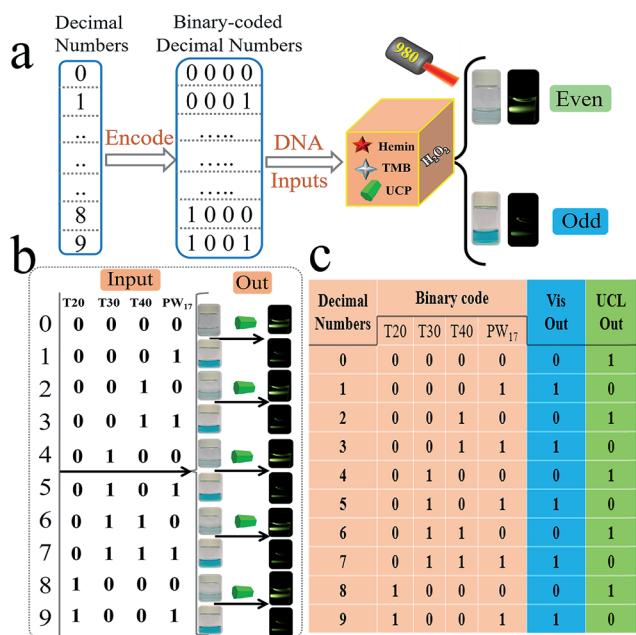


Fig. 5 Operation of the parity-checker for discriminating natural numbers (from 0 to 9). For the parity checker, the strands T20, T30, T40, and PW<sub>17</sub> were used as the 4 binary-coded inputs.

solely G4 is output “1”, while other products are “nothing”, and GSH and “GSH + G4” are output “0”). After reacting with the “platform-box”, the downstream “Probe module” (the 1-2 decoder,<sup>20–24</sup> the output of the INH gate was used as the input and the Vis/UCL signals were the 2-output) will produce ratiometric Vis/UCL outputs and the corresponding truth table is shown in Fig. 6d. In the absence of antioxidants and in the presence of G4, it produces the output “Vis = 1, UCL = 0” (the threshold value was set as “0.45” to distinguish the high/low outputs). On the other hand, in the coexistence of the antioxidants and G4, OxTMB will be reduced to TMB that cannot quench the UC emission, generating the output “Vis = 0, UCL = 1”. Thus, the presence of antioxidants can be easily recognized. This ratiometric dual-output operating-mode brought strengthened reliability and intelligence to the bio-diagnosis. Fig. S13† presents the normalized ratiometric column values (Vis<sub>650</sub>/UCL<sub>545</sub>) in the presence of antioxidants (GSH, AA, and Cys as the model targets) and other reactants, and the significant differences indicated the excellent selectivity of this “antioxidant indicator”.



## Conclusions

In summary, we, for the first time, proposed the concept of “DNA Janus Logic Pair” (DJLP) to classify DNA logic gates with contrary functions into “Positive + Negative” gates. Based on the biocatalytic property of G4zyme and the luminescence quenching phenomenon between OxTMB and upconversion particles, we constructed a universal amphichromatic platform that kills two birds with one stone by operating a versatile DJLP library. Various DJLPs, advanced logic devices (parity checker for distinguishing natural numbers less than 10, and encoder and decoder), the concatenated circuits and a logic-controlled “antioxidant indicator” were all successfully achieved. This study reduced the operating costs/time of the current DNA gates by at least half, and both the amphichromatic outputs (Visual-blue and UC luminescent-green) could be easily visualized by eyes under harmless-NIR, thus bringing largely enhanced computing complexity and practicality to the method. On combining with the genome editing technique, aptamers, nanozymes, DNA origami and other nanomaterials, numerous avenues for the development of more powerful biocomputing systems and intelligent logical bio-applications will open up.

## Experimental section

### Chemicals

The DNAs were synthesized by Shanghai Sangon Biotechnology Co. (Shanghai, China) and the sequences are in Table S1.† They were all dissolved in distilled-water and quantified by UV-Vis spectroscopy. Glutathione (GSH) was provided by Shanghai Sangon Biotechnology Co. (Shanghai, China) and  $\text{H}_2\text{O}_2$  (30%) was purchased from Beijing Xilong Chemical Industry (Beijing, China).  $\text{Yb}(\text{NO}_3)_3 \cdot 5\text{H}_2\text{O}$  (99.9%),  $\text{Y}(\text{NO}_3)_3 \cdot 6\text{H}_2\text{O}$  (99.9%), and  $\text{Er}(\text{NO}_3)_3 \cdot 5\text{H}_2\text{O}$  (99.9%) were provided by Aladdin (Shanghai, China). Sodium citrate and NaF were purchased from Beijing Chemical Works (Beijing, China). 3,3',5,5'-tetramethylbenzidine (TMB) and hemin were provided by Sigma-Aldrich (USA). Hemin (2 mM) and TMB (0.5% (w/v)) were dissolved in DMSO to prepare the stock solution. The reaction buffers used in this study are: 1× HEPES buffer (25 mM HEPES, 20 mM KCl, 100 mM NaCl, 0.05% (w/v) Triton X-100, 1% (v/v) DMSO, pH 7.5) was used to incubate G4zyme. 1× MES-Ac buffer (25 mM MES, 20 mM KAc, pH 4.5) was used in the oxidation reaction of TMB.

### Synthesis of UCP

The green colored UCP were synthesized according to a previous report. In general, the mixture of  $\text{Yb}(\text{NO}_3)_3 \cdot 5\text{H}_2\text{O}$ ,  $\text{Y}(\text{NO}_3)_3 \cdot 6\text{H}_2\text{O}$ , and  $\text{Er}(\text{NO}_3)_3 \cdot 5\text{H}_2\text{O}$  with different molar ratio (total concentration of the rare-earth element was 1 mmol) was added to 5 mL distilled water to achieve a homogeneous solution. Then, 8.5 mL of aqueous sodium citrate solution (3 g) was added and the above mixture was ultrasonicated for 15 min. After this, 16 mL NaF (0.52 g) was mixed with the above solution. After adding 1.5 mL 98% nitric acid to change the pH to 3, the mixture was stirred for another 40 min and added into a 50 mL Teflon autoclave. After reaction at 180 °C for about

2.5 h, the final solution was centrifuged and washed using ethanol and distilled water 6 times to obtain the white products. Finally, the UCP were dissolved in 20 mL distilled water as the stock solution (the concentrations of the UCP were 0.01475 g  $\text{mL}^{-1}$ ).

### SEM, TEM, XRD

TEM images of the UCP were obtained on a JEOL 2100F transmission electron microscope (TEM) (accelerating voltage = 200 kV) and the SEM images were taken on a Philips XL-30 ESEM (accelerating voltage = 20 kV). The XRD powder patterns were recorded on a D8 ADVANCE Wide Angle X-ray diffraction (BRUKER, Germany).

### Upconversion luminescence and UV-Vis spectra

The UCL spectra were collected from 400 to 750 nm on a Cary Eclipse Fluorescence Spectrophotometer (Agilent Technologies, USA) equipped with a 980 nm NIR-laser (Hi-Tech Optoelectronics Co., Ltd.). A constant working power was used for all the UCNPs samples and the emission slit width was 20 nm. The absorbance spectra of OxTMB were measured on a Cary 50 Scan UV/Vis/NIR Spectrophotometer (Varian, USA). The luminescence decay curves were obtained on a FLSP-920 Steady State and Time-resolved Fluorescence Spectrometer (Edinburgh, England).

### Native polyacrylamide gel electrophoresis (PAGE) and circular dichroism (CD)

The PAGE experiments were performed according to our previous studies. 1× TE buffer (50 mM Tris, 5 mM EDTA, and 50 mM  $\text{MgCl}_2$ , pH 8.2) was used as the DNA hybridizing buffer and the 1× TBE buffer (18 mM Tris, 18 mM boric acid, and 1 mM EDTA, pH 8.02) was applied as the working buffer for PAGE. The CD spectra were obtained from 220 to 320 nm on a JASCO J-820 spectropolarimeter (Tokyo, Japan). The spectra were recorded in 1 mm path length cuvettes and averaged from three scans.

### Formation of G4zyme

The solutions of different DNAs were heated at 88 °C for 10 min and slowly cooled down to room temperature.

(1) For the “YES + NOT”, “AND + NAND”, “OR + NOR”, “INH + IMP” DJLPs: 500 nM  $\text{PW}_{17}$  for “YES + NOT”; 500 nM PG3 and PG1 for “AND + NAND”; 500 nM  $\text{PW}_{17}$  and 500 nM TG for “OR + NOR” and 500 nM for  $\text{PW}_{17}$ , 1  $\mu\text{M}$  C4 for “INH + IMP” were selectively used and reacted at room temperature for about 10 min.

(2) For the “XOR + XNOR” DJLP, in the input combination of (00), 500 nM YP was used; in that of (01, 10), 500 nM YP was mixed with 500 nM YB, 1  $\mu\text{M}$  CA (or 500 nM YA, 1  $\mu\text{M}$  CB); in that of (11), 500 nM YB and 1  $\mu\text{M}$  CA were premixed with 500 nM YA, 1  $\mu\text{M}$  CB, and reacted at room temperature for about 10 min, and then 500 nM YP was added at last.

(3) For the “MAJ + MIN” DJLP, 400 nM DA, 300 nM DB, and 350 nM DC were alternatively added to the platform strands



(mixture of 300 nM P1/P2/P3) and reacted at room temperature for about 10 min.

(4) For the parity-checker: 200 nM of T20, T30, T40, and 500 nM PW<sub>17</sub> were selectively mixed and reacted at room temperature for about 10 min.

(5) For the 2-1 encoder, 1-2 decoder, different concatenated circuits (1 mM GSH was used), and the antioxidant-nose (200 μM, different targets or antioxidants), the concentrations of the DNAs were similar to the above devices.

After the above DNA reactions, equal volume of 1 μM hemin (DMSO) was added and the above mixture was incubated at room temperature for 1.5 h to form G4zyme.

### The visual and UCL photos

20 μL 0.5% (w/v) TMB and 15 μL 30% (w/v) H<sub>2</sub>O<sub>2</sub> was mixed with 30 μL of G4zyme and 935 μL 1× MES-Ac buffer in 1 mL centrifuge tube. After they reacted at room temperature for 10 min, a dark-blue colored OxtMB was obtained. Then, 1 mL OxtMB solution was mixed with 1 mL UCP (50 μL UCP in 950 μL 1× MES-Ac buffer). The visual photos were taken under natural light and the ones for UCL were taken under 980 nm NIR-laser (about 3.5 W for all the experiments) at dark. All the photos were taken using a digital Canon G12 camera.

## Conflicts of interest

There are no conflicts to declare.

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