A catalytic protein–proteomimetic complex: using aromatic oligoamide foldamers as activators of RNase S†

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Foldamers are abiotic molecules that mimic the ability of bio-macromolecules to adopt well-defined and organised secondary, tertiary or quaternary structure. Such templates have enabled the generation of defined architectures which present structurally defined surfaces that can achieve molecular recognition of diverse and complex targets. Far less explored is whether this mimicry of nature can extend to more advanced functions of biological macromolecules such as the generation and activation of catalytic function. In this work, we adopt a novel replacement strategy whereby a segment of protein structure (the S-peptide from RNase S) is replaced by a foldamer that mimics an α-helix. The resultant prosthetic replacement forms a non-covalent complex with the S-protein leading to restoration of catalytic function, despite the absence of a key catalytic residue. Thus this functional protein–proteomimetic complex provides proof that significant segments of protein can be replaced with non-natural building blocks that may, in turn, confer advantageous properties.

Introduction

Nature uses a stunning selection of molecular architectures to carry out complex tasks such as catalysis and cell signalling; proteins adopt a specific three-dimensional compact conformation using defined secondary structure elements to spatially position chemical groups so as to perform work. However, our ability to reproduce functions such as catalysis, using synthetic systems remains rudimentary.1–7 Chemists have readily accepted the challenge of designing and synthesising architectures of comparable complexity to biomacromolecules using non-natural building blocks, specifically through development of foldamers,8–18 sequences of non-natural monomers that adopt both biomimetic and abiotic secondary, tertiary and quaternary structures that topologically29–32 and topographically13–15 mimic native structures. This has allowed creation of foldamers that sterically occlude the binding of natural ligands e.g. to inhibit protein–protein interactions,11,13,16–18 and which are processed by biomacromolecular machinery.19–21 However, the control, plasticity and exquisite selectivity of proteins also lies within their dynamic properties, particularly during the catalysis of chemical reactions. We posed the question “Can structural mimics also recapitulate the intrinsic properties required for regulation and catalysis?” If so, a tantalizing alternative to bottom-up design of foldamers would be realizable by replacing components of proteins piece-by-piece22,23 with non-natural building blocks whilst retaining function.24 Such an approach has been termed “protein-prosthesis”25 and may even lead to “bionic proteins”.26 So far, largely conservative single residue mutations to protein sequence have been made.27,28 Replacement of entire secondary structural motifs with topological mimics (i.e. foldamers mimicking local conformation) is less established.29–32 Replacement of up to three amino acids with peptoid residues in RNase A afforded a protein-conjugate which, at ten-fold higher concentration than the native protein afforded 10% of the activity. Sequence based replacement of amino acid residues in chorismate mutase afforded proteins with comparable $k_{cat}/K_m$ values to the wild type protein (although mutations at the active site further diminished efficiency). Finally, replacement of an α-helix in IL-8 with a designed β-peptide variant resulted in a synthetic protein that retained IL-8 signalling ability. In this work, we study the classic model enzyme complex RNase S, replacing the S-peptide at the active site with a small-molecule topographical mimic (i.e. a foldamer mimicking the shape and surface) of protein secondary structure (Fig. 1a).22,23 The resulting non-covalent complex recovers the ability to bind and cleave the cleavage of RNA elicited by the natural RNase S complex despite the unprecedented extent of protein-structure replacement.
and the foldamer highlighting e
S-peptide
results in restoration of catalytic competence; S-protein is catalytically inert but addition of the S-peptide
short N-terminal S-peptide. In the absence of the S-peptide, the
structure of an oligoamide foldamer designed to mimic
(expansion) upon which to design a helix mimetic; (e) chemical
+ 7 constellation of side chains within the S-peptide sequence
mentation system has thus found use in several
results in restoration of catalytic competence;28 this complement-
ment system has thus found use in several
applications.39–42 The S-peptide is bound in an \( \alpha \)-helical conformation within a cleft on the S-protein37 and bears one of
the two histidine residues required for acid-base catalysed
hydrolysis of the phosphodiester linkage in RNA (Fig. 1b–d).42 The helical nature of this N-terminal segment of the protein
thus renders it an ideal motif for replacement with secondary
structure mimics. Our group previously introduced a series of
proteomimetic scaffolds that mimic the spatial projection of \( i, i + (3 \pm 4) \) and \( i + 7 \) residues on an \( \alpha \)-helix (Fig. 1e–h).14,43,44 These scaffolds, accessible via solid-phase synthesis,14,44 can be deco-
 rated with side chains that mimic hot-spot residues46 on template helices found at protein–protein interfaces to achieve inhibition of protein–protein interactions.14,43,44 We hypothe-
sized that helix mimetics capable of reproducing the spatial
orientation and composition of the \( i, i + 3 \) and \( i + 7 \) side chains
found on the S-peptide should bind to the S-protein and were
curious if these mimics would also activate the RNA processing
function.

An RNase activity screen identifies activators of RNase S
Using the S-protein obtained from an established biochemical
protocol35,47 and synthetic S-peptide (obtained via solid-phase
synthesis), we established an enzymatic assay to assess the
ability of reconstituted RNase S to hydrolyse RNA. The assay
relies upon binding of ethidium bromide to unhydrolysed
RNA.48 Using the remaining fluorescence as an approximation of
activity, we used the assay to screen libraries of previously
prepared aromatic oligoamide helix-mimetics (see ESI for
structures; ESI Tables S1 and S2†) derived from O-alkyl-
lated,49,49,50 N-alkylated14 and hybrid scaffolds,14,51 for reac-
tivation of the S-protein (ESI Fig. S1†). Despite the limited
prevailence of helix mimetics in our library bearing histidine-
mimicking side chains, we were intrigued as to whether
binding and global structural emulation would be sufficient to
support enzymatic hydrolysis of RNA upon binding to the S-
protein. Indeed, a number of the compounds elicited dose-
dependent reactivation of the RNase S-protein (Fig. 2a and
b and ESI Fig. S2† for representative examples), although the
solubility plateau of the mimetics prevented saturation and
therefore determination of the maximum concentration at
which maximal enzymatic activity is attained. Analysis of the
screening hits point to a requirement for a hybrid helix-mimetic
scaffold comprising a 3-O-alkylated benzamide at the N-
terminus with an aromatic R\(^1\) group, a central phenylalanine
in the R\(^2\) position, a 2-O-alkylated benzamide with an R\(^3\) iso-
propyl group and a C-terminal glycine (Fig. 2a).

**Results**

**Selection of RNase S and aromatic oligoamides for
construction of bionic proteins**

RNase S can be obtained by subtilisin mediated cleavage of
RNase A to yield the non-covalently associated S-protein35 and
short N-terminal S-peptide. In the absence of the S-peptide, the
S-protein is catalytically inert but addition of the S-peptide
results in restoration of catalytic competence;28 this comple-
mentation system has thus found use in several

**hDM2 can be used to negatively regulate RNase S-protein
function through competition for proteomimetics**

To further establish that a non-covalent interaction between
the foldamer and S-protein is responsible for restoration of catal-
ysis, we used a competition experiment with a second protein
target of the identified foldamers (Fig. 3). The hybrid helix-
mimetic scaffolds had previously been developed as p53/
hDM2 inhibitors,43 thus hDM2 was selected for this purpose.
Compound 1a was shown by fluorescence anisotropy
competition to act as a low micromolar inhibitor of the p53/hDM2 interaction (IC50/C24 10 μM, ESI Fig. S3 and Table S3† for additional compounds). Addition of excess hDM2 suppressed the foldamer-dependent activation of RNA hydrolysis (Fig. 3c and d). Notably, the addition of hDM2 to S-protein/S-peptide complex had no effect on catalysis (Fig. 3c) and S-peptide did not compete with fluorescently labelled p53 peptides in an assay against the hDM2/p53 interaction (ESI Fig. S4†). It should be noted, RNase S activation appears to be suppressed with sub-stoichiometric hDM2 concentrations relative to helix mimetic. Such behaviour could arise due to ternary complex formation between hDM2, the helix mimetic 1a and RNase S, opening up an untapped mechanism of non-covalent RNase S regulation. In addition, the tested oligoamides exhibit a degree of non-specific interaction (see later), which could result in lower effective free mimetic concentration for RNAse S activation when hDM2 is present. A more detailed analysis is beyond the scope of the current manuscript and will be the focus of future studies.

Proteomimetics bind directly to RNase S-protein at the S-peptide binding site

To further characterize the RNase S-protein/helix mimetic interaction we performed more detailed analyses focussing on mimetic 1a. In kinetic analyses, the rate of RNA hydrolysis for the S-protein/1a complex increased at higher RNA concentration (Fig. 2c, ESI Fig. S5–S9† and Table S4†) allowing kinetic parameters to be assessed (Fig. 2d and ESI Fig. S10†). $k_{cat}$ values could not be extracted from the kinetic data due to the unknown

Fig. 2  (a) Helix mimetics 1a, 3, and 4 as activators of RNase S-protein; (b) RNA degradation at 3 concentrations of selected foldamers (200, 50 and 1.25 μM) shown to be active in the screen (see ESI† for full data set at single concentration and dose response curves) RNA (0.15 mg mL$^{-1}$), S-protein (0.05 μM), S-peptide (0.4 μM), RNase A (0.05 μM) in Tris buffer (50 mM Tris, 100 mM NaCl, pH = 6.5). Fluorescence was measured immediately and every 30 s for 45 minutes of incubation at 25 °C. Remaining fluorescence was calculated using the 45 minute time-point; (c) variation in rate of RNA hydrolysis with substrate concentration for S-protein, S-protein/S-peptide and S-protein/1a; S-protein (0.05 μM), S-peptide (0.4 μM), 1a (100 μM) (for progression curves including RNase A see ESI Fig. S5–S9†); (d) Michaelis–Menten plots for S-protein, S-protein/S-peptide and S-protein/1a (see ESI Fig. S10† for Michaelis–Menten plots including RNase A).
molar concentration of RNA cleavage sites. In order to numerically compare the enzymatic efficiency we therefore fitted $V_{\text{max}}/K_M$ values using the first 3 data points of the Michaelis–Menten plots. Comparing at constant S-protein concentration (0.05 μM), the data for S-protein/1a ($V_{\text{max}}/K_M = 9.52 \pm 0.48 \times 10^{-4}$ s$^{-1}$) indicate that $V_{\text{max}}/K_M$ (the first order rate constant at low substrate concentration) decreases in comparison to S-protein/ S-peptide ($V_{\text{max}}/K_M = 1.36 \pm 0.07 \times 10^{-3}$ s$^{-1}$), but increases in comparison to the S-protein alone ($V_{\text{max}}/K_M = 5.17 \pm 0.84 \times 10^{-4}$ s$^{-1}$), indicating that 1a effectively activates the S-protein. ITC analysis provided further evidence for a direct interaction between the proteomimetic 1a and S-protein (Fig. 4a). This experiment was performed by titration of S-protein into mimetic; binding was observed to occur with low micromolar affinity ($K_D = 1.85 \pm 0.97$ μM, n = 0.1, Table S5†).

The affinity is only one order of magnitude weaker than the interaction of the native S-peptide with S-protein ($K_D = 0.13 \pm 0.02$ μM, ESI Fig. S11†), whilst the non-stoichiometric interaction may arise in part due to some non-specific binding which we discuss further below. Further evidence of an interaction between the S-protein and helix mimetic 1a was obtained via native ESI-MS. Using a 5-fold excess of the mimetic relative to the S-protein, a 1:1 complex and a lower intensity 1:2 S-protein/mimetic 1a complex could be detected (Fig. 4b). This minor species may again imply a degree of nonspecific binding to S-protein and suggests that the observed apparent $K_D$ from ITC may be an average of the simultaneously occurring binding events. Further native MS competition-binding experiments were performed to probe the binding site for the mimetic by keeping the concentrations of S-protein and 1a constant while using increasing concentrations of S-peptide; no ternary complex could be detected (Fig. 4c), suggesting the S-peptide and mimetic 1a compete for the same binding site on S-protein.

To provide additional insight into the mode of recognition between the mimetics and S-protein a range of docking analyses were performed (see ESI† for full details of the analyses). Helix mimetics were energy-minimized and subjected to Monte Carlo conformational analysis and then lowest energy conformers retained for docking experiments. These lowest energy conformers were docked into the rigid-body structures of either (i) the S-protein or (ii) RNase A (lacking the S-peptide residues) in complex with a tetranucleotide. All input ligands gave a pose

![Chemical Structure](Image)

**Fig. 4** Analyses of binding interaction between helix mimetics and RNase S: (a) ITC thermogram of S-protein titrated to compound 1a. Native MS analysis of 2 μM S-protein in complex with (b) 10 μM 1a showing S-protein/1a complexes having 1 : 1 and 1 : 2 ratio and (c) 10 μM 1a and 2.5 μM S-peptide showing S-protein/1a and S-protein/S-peptide complexes, no ternary complex detected. Modelling studies on helix mimetics as activators of RNase S: (d) overlay of molecular models of 123 conformers of hybrid helix mimetics docked with S-protein (green) using rigid docking. Lowest energy poses of (e) 1a, and (f) 4 using flexible docking optimized by induced-fit with S-protein; (g) distribution of the different orientations of the docked mimetics (top 20% lowest energy poses) relative to RNA binding site using rigid docking; (h) distribution of the different orientations of the top 20% lowest energy poses of the docked mimetics relative to RNA binding site after induced fit optimization.
in the S-protein binding groove in the region occupied by His12 of the S-peptide (e.g. Fig. 4d). For docking to the S-protein alone, ligands were observed in an extended form in both the S-peptide and the RNA-binding pocket while docking against the nucleotide-RNase A complex (lacking the S-peptide residues) led to binding in the S-peptide pocket but without a well-defined orientation (ESI Fig. S12†). In contrast, flexible docking followed by induced fit optimization gave a more consistent orientation with the majority of the lowest energy poses aligned with the C-terminus of the aromatic oligoamide close to the RNA (Fig. 4e and f for representative examples 1a and 4 and ESI Fig. S13† for the results without induced fit-optimization). This suggests that the C-terminal carboxylate might fulfil the role of an ionisable group in mediating acid-base catalysed hydrolysis of RNA. To test this hypothesis, we prepared derivatives of 1a bearing an N-terminal acetonilide 1b, a C-terminal carboxamide 1c and with both modifications 1d (see ESI†). Unfortunately, the solubility of these derivatives was such that inconsistent and therefore inconclusive behaviour was observed in the kinetic studies although docking afforded similar results to those observed with the parent hybrid mimetics, i.e. with the C-terminus of the mimic oriented close to the RNA (ESI Fig. S14†). Finally, the docking analyses highlight some imperfections in the original design: generally, the aromatic side-chains of the mimetics occupy the designed binding sites in the majority of the docked ligands, however, we observed diverse conformations and multiple different orientations of the ligand due to the increased conformational flexibility originating from the central \(\alpha\)-amino acid. This points towards an induced fit mechanism of binding, where the aromatic side-chain acts as anchor residue (see ESI Fig. S15† for an overlay of the S-peptide with docked poses obtained by flexible docking).

With the goal of improving catalytic activity we also: (i) attempted to prepare helix mimetics bearing imidazole groups on the \(\sigma\)-alkyl side chains, and (ii) helix mimetics bearing \(\sigma\)- or L-histidine instead of glycine at the C-terminus; our synthesis was not compatible with the former approach due to facile elimination of the imidazole side chain during synthesis whilst the later compounds did not reactivate S-protein. It is noteworthy however that removal of the catalytic histidine residues need not abrogate catalysis; RNase A variants with either His12Ala and His119Ala changes to primary sequence were previously shown to retain some catalytic activity.\(^{22}\) Loss of the His12 imidazole was shown to decrease the affinity for the transition state by \(10^4\) fold for a range of substrates (Poly(C), Upa and UpOC\(\sigma\)H\(_4\)p-NO\(_2\) hydrolysis), whereas eliminating the imidazole in His119 had a comparable effect on Poly(C) and Upa hydrolysis, but not the cleavage of UpOC\(\sigma\)H\(_4\)p-NO\(_2\) indicating the role of general acid for His119 to protonate the leaving group.\(^{22}\) Assuming a water fulfils the role of general base in lieu of His12, the rate of RNA hydrolysis would be anticipated to be reduced significantly – such behavior is thus consistent with our observations i.e. it may be the case that 1a binds to the S-protein so as to form a structurally competent site for catalysis whilst the role of general base is fulfilled by water.

Finally, closer analysis of the kinetic data allow formulation of a plausible hypothesis that can rationalize the ITC and ESI-MS binding data. At low concentrations of RNA, it appears that the mimic acts as an inhibitor of RNA hydrolysis (Fig. 5a), whereas at higher RNA concentrations (Fig. 5b), presence of the mimic accelerates hydrolysis. These data imply that the mimic can bind to both the S-protein-binding site, and competitively with the RNA binding site (Fig. 5c), the observation that helix mimetics can interact with the RNA binding site in the docking analyses performed in the absence of RNA (see ESI Fig. S12†), support this hypothesis.

**Conclusion**

In this work we have demonstrated that aromatic oligoamide foldamers can act as a component of a functional quaternary structure that performs RNA hydrolysis. Screening a panel of helix mimetics in an RNase assay led to identification of compounds that upon binding S-protein led to partial recovery of enzymatic function observed for the S-protein/S-peptide complex. The proteomimetics were shown to display first-order kinetics with enzymatic efficiency dependent on both concentration of proteomimetic, and RNA, and, directly bind to S-protein with low \(\mu\)M affinity. As further evidence of the potential to generate new capabilities using this approach, we demonstrate that the S-protein/proteomimetic enzyme function can be readily regulated using an additional protein – hDM2 – which competitively binds to the proteomimetics, a property not observed for the S-protein/S-peptide complex. Non-covalent MS analyses suggest the mimetics bind to the S-peptide binding site on the S-protein, a conclusion supported by molecular modelling. Crucially none of the proteomimetics bear an
imidazole side chain to facilitate catalysis; consequently, the efficiency of RNA hydrolysis is >10^4-fold lower than for fully functional RNase S-protein/S-peptide complex. Such an observation is consistent with prior studies on RNase A which established that His12 (within the S-peptide sequence) could be removed without complete loss of catalytic competence. Significantly, our experimental and in silico analyses, point to opportunities to address imperfections in the design through future studies; the solubility threshold, and lower selectivity of the mimetics (exploited advantageously here in the hDM2 competition experiment), together with improved potency should be addressed to achieve superior designs. We note that aromatic oligoamides of this nature have shown a measure of selectivity previously^44,45 and that solubility and selectivity can be improved using a number of strategies including the use of peptide-foldamer hybrids. Incorporation of further functionality that allows mimicry of more than the i, i + 3 and i + 7 constellation on an z-helix should further aid these objectives.

A significant goal in biomimetic chemistry is to reproduce structure and functionality of biomacromolecules using synthetic molecules. Foldamers do so firstly by mimicking the conformations adopted by bio-macromolecules and then their synthetic molecules. Foldamers do so structure and functionality of biomacromolecules using constellation on an.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

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