Alchemilla vulgaris agg. (Lady’s mantle) from central Balkan: antioxidant, anticancer and enzyme inhibition properties

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The current study was designed to evaluate the phytochemical profile and biological properties (antioxidant, enzyme inhibitory and cytotoxic activity) of methanolic, ethanolic, ethyl-acetate and water extracts of Alchemilla vulgaris from Southeast Serbia (central Balkan) which was traditionally used to alleviate and treat many diseases. Bioactive compounds were characterized by LC-MS/MS technique and biological properties were evaluated using antioxidant, enzyme inhibitory and cytotoxic assays. Twenty-six phenolic compounds were quantified in methanolic, ethanolic, ethyl acetate and water extracts, whereas the highest yields were found in ethyl-acetate (EA) extract (gallic acid, caffeic acid, catchin, quercetin). This extract has also shown the greatest antioxidant, anticancer and inhibitory enzyme activities which were demonstrated for the first time in this study. The obtained results indicated that Alchemilla vulgaris from South Serbia possesses high potential for pharmaceutical applications.

Introduction

Nature has been an inexhaustible source of bioactive compounds which have been used since ancient times for treatment of many diseases. Today more interest is being paid to understanding the complex role of plant nutrition where medicinal plants have particular importance. These plants include various types of plants for medicinal purposes. They are considered as a rich source of bioactive ingredients. Serbia is one of the world’s most biodiverse centers, rich in medical plants, where some of them have not been determined yet or there is no information on their ingredients and bioactivity.¹³

Alchemilla vulgaris, an aggregate species, commonly known as Lady’s mantle or lion’s foot, is a perennial herbaceous plant belonging to the rose family (Rosaceae).

The status of the species Alchemilla vulgaris is still not completely resolved and there are a few opinions. However, most of them treat the taxon as an aggregate of microspecies, which were also considered as subspecies or varieties of the species.

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According to Flora of Serbia (Genus Alchemilla, Gajić, 1972) name of the species – Alchemilla vulgaris L. is valid; the species is present in Serbia with three subspecies (subsp. montana, subsp. pratensis and subsp. vulgaris).¹⁵

Among ethnobotanical reports on Alchemilla vulgaris for Southeast Europe and the territory of Balkan peninsula, it is to mention a comprehensive work of Tucakov (1997), and his reports on astringent and anti diarrheal, as well as effects against pruritus vulvae, bleeding from uterus and menstrual pains. The most used folk names according to Simonovic (1959) are: “virak”, “vrkuta”, “rosanica”, “biserak”, “virkovina”, “virić” and “gospinplašt” (it is interesting that the latter literally means Lady’s mantle).¹²,¹³

It is 30 to 50 cm long half-rosette herbaceous plant with corrugated and lobed kidney-shaped to semicircular leaves and the yellowish-green flowers form clusters. The plant blooms from June through to September and can be found on roadsides, grasslands bank, and mountain slopes.

Aerial parts of the plant are often used against gastrointestinal diseases and inflammatory processes, as well as for improvement wound healing due to its strong antimicrobial and anti-inflammatory effects.² The plant is very popular especially due to its great effect against women’s problems and diseases such as fibroids, cysts, endometriosis, infertility, relief of menstrual problems and regulation of the menstrual cycle and reproductive and thyroid hormones balancing.² Extract of the plant exhibits high scavenger potential, especially towards superoxide anion radical. The results obtained in some studies indicate that presence of some phenolics, especially tannins, may contribute to the antioxidant activity.¹⁴ Furthermore, Takir...
et al. (2015) demonstrated that methanolic extract of *Alchemilla vulgaris* has shown vasorelaxant and hypotensive effects.\(^7\)

The extracts of leaves and roots of *Alchemilla vulgaris* have been reported to inhibit the orthopox viruses in Vero cell lines.\(^6\) Bahadir et al., 2017 have performed in their study that extracts of *Alchemilla vulgaris* demonstrated a hepatoprotective activity increasing the aminotransferase (ALT) level in carbon tetrachloride induced mice serum.\(^7\)

In the Southeast Europe and the Balkan peninsula, the Lady’s mantle is very popular and acknowledged medicinal plant, which is traditionally used in folk medicine and in range of herbal remedies. The herb is mostly used for preventing of woman illnesses, such as menstrual problems, the oligomenorrhea and dysmenorrhea, the menopausal complaints, for regulation of vaginal secretion, polycystic ovaries and even for infertility.\(^8\)–\(^10\) Moreover, the Lady’s mantle is used as diuretic, anti-anemic and anti-diabetic, for wounds and ulcers, hernias and muscle atrophy,\(^11\) and it is readily used for wound healing, especially when used externally for ulcers, eczema and skin. There are also notes on use for mild and nonspecific diarrhea and for slow metabolism.\(^12\) As reported for other regions, the most records refers to woman illnesses and disorders uses, in addition to astrinrent, antihemorrhagic, tonic, cardiotonic and diuretic effects.\(^13\) In general, there are not many reports on detailed phytochemical profile of *Alchemilla* and related biological properties, especially focusing on anticancer properties and in particular of the estrogen depended cancer lines. The current study focuses on phytochemical composition assessment, targeting the phenolic compounds, in different extracts of *Alchemilla vulgaris* collected from the central Balkan (South-east Serbia) region. The chosen populations originated from sites where the plant material is collected from the wild for Serbian and regional market needs. The main goal of the present study was to verify the existing ethnobotanical data by performing several antioxidant activity tests, as well as the anticancer and enzyme activity analyses. Evaluation of antioxidant capacity has been also performed using different tests such as 2,2-diphenyl-1-picrylhydrazyl (DPPH), 2,2’-azinobis-(3-ethylbenzothiazoline-6-sulfonic acid (ABTS), Cupric Reducing Antioxidant Capacity (CUPRAC), phosphomolibdenum, metal chelating and Ferric Reducing Antioxidant Power (FRAP). Concerning already known health beneficial effects of the plant, in first uses for gynecological disorders 8, we decided to perform investigation on anticancer activity of plant extracts on cancer cells obtained from reproductive tissues, such as human breast adenocarcinoma (MCF7 cell line), cervical adenocarcinoma (HeLa) and ovarian carcinoma (A2780) in addition to prostate adenocarcinoma (PC-3 cell line). The efficacy of different *Alchemilla vulgaris* extracts against cancer cell lines was evaluated using two viability tests, MTT based on the measurement of mitochondrial respiration and crystal violet (CV), on the number of adherent cells in culture that retained after the treatment.

There are also reports which have shown that plants from *Alchemilla* genus were traditionally used for skin and neurodegenerative disorders and against diabetes and gastrointestinal problems of this plant,\(^14\)–\(^17\) its ability to inhibit anti-cholinesterase,\(^18\) anti-tyrosinase and anti-amylase activity of *A. vulgaris* extracts will be investigated in this study.

These investigations were performed to verify whether there are some other biological performances of Lady’s mantle extracts which have been neglected so far, as there is not such ethnopharmacological information.

For best of our knowledge, the evaluation of Lady’s mantle antancer activity, especially on estrogen depended tumors, is reported for the first time. Summarily, the phytochemical characterization in relation to antioxidant activity, enzyme inhibitory effects and anticancer activity of different *Alchemilla vulgaris* extracts are presented to clarify already known ethnopharmacological records, as well as to identify some novel possible biological effects of the plant.

### Material and methods

#### Plant material

Plant material was collected at the flowering stage in midsummer, 2017 from the site situated in south Serbia, the Vlasina lake (E 22.36446000 N 42.73636300, 1234 m above sea level). The habitat of the population is wet grassland of the Calthion type. Plant vouchers were deposited at Herbarium of Applied Botany Department of the Faculty of Agriculture, University of Belgrade, under voucher code 85000 BEO. The collected material was air dried.

#### Extracts preparation

Dry plant material was macerated for 24 h in 80% methanol (v/v), 70% ethanol (v/v), 70% ethylacetate (v/v) and distilled water (dH\(_2\)O) at room temperature using incubator shaker and filtrated (IKA\(^\circ\)). The supernatants were taken for drying. The obtained residues were dissolved in dimethyl sulfoxide (DMSO) to the concentration of 10 mg mL\(^{-1}\). The extracts (methanolic-MeOH, ethanolic-ETO\(_{H}\), ethylacetate-EA, water-H\(_2\)O) were stored in dark at +4 °C until further analyses.

#### Quantification of bioactive compounds using liquid chromatography tandem-mass spectrometry (LC-MS/MS) technique

Determination of 45 selected phenolic compounds was performed according to previously described method.\(^19\)

Data were acquired in dynamic multiple reaction monitoring (MRM) mode, using the optimized compound-specific parameters. For all the compounds, peak areas were determined using Agilent Mass Hunter Workstation software – Qualitative Analysis (ver. B.03.01.). Calibration curves were plotted and samples’ concentrations calculated using the OriginLabs Origin Pro (ver. 9.0) software.

#### Assessment of antioxidant activity and total content of phenolic compound

**DPPH assay.** Colorimetric DPPH radical scavenging activity was measured according to the method described by Espin et al., 2000.\(^20\) The obtained results were expressed as milligrams
of Trolox equivalents per gram of dry weight (mg TE per g d.w.), calculated according to the Trolox standard calibration curve.

**ABTS assay.** The ABTS radical scavenging ability was determined according to the method reported by Re et al. (1999). The results were evaluated as Trolox equivalents per gram of dry weight (mg TE per g d.w.).

**CUPRAC assay.** The cupric reducing power ability was detected by CUPRAC assays as reported by Apak et al. (2006). The results were expressed as Trolox equivalents per gram of dry weight (mg TE per g extract d.w.).

**Phosphomolybdenum assay.** The phosphomolybdenum assay was performed according to the method reported by Prieto et al. (1999). The results were evaluated as Trolox equivalents per gram of dry weight (mmol TE per g extract).

**Metal chelating.** Ferrozine assay was performed to detect the metal chelating ability as reported by Dinis et al. (1994). The results were provided as ethylenediaminetetraacetic acid (EDTA) equivalent (mg EDTAE/g extract).

**FRAP assay.** The FRAP assay was conducted according to Benzie and Strain (1996), adapted for 96 well plate. The results were expressed as mg of ascorbic acid equivalents per g of dry weight (mg AAE per g d.w.) using the calibration curve of ascorbic acid.

Each antioxidant assay was carried out in triplicate and results were expressed as mean values.

**TPC assay.** Total phenolic compounds were determined with Folin-Ciocalteu reagent using the method of Medina-Remon et al. (2009). The results were expressed as milligrams of gallic acid equivalents per g dry weight (mg GAE per g d.w.) using standard calibration curve of gallic acid.

**Enzyme inhibitory activity**

The extracts were tested as sources of several enzyme inhibitors, including α-amylase, cholinesterases (AChE and BChE), and tyrosinase. The procedures of these assays were reported in our earlier work. The enzyme inhibitor effects were evaluated as equivalents of acarbose for α-amylase, galantamine for acetyl cholinesterase (AChE) and butyryl cholinesterase (BChE), and kojic acid (KAE) for tyrosinase.

**Reagents and cells**

Culture medium RPMI-1640 and Dulbecco’s Modified Eagle Medium (DMEM) were purchased from Biowest (MO, USA). Dimethyl sulfoxide (DMSO), phosphate-buffered saline (PBS), fetal calf serum (FCS) were acquired from Sigma (St. Louis, MO, USA). 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) was bought from AppliChem (MO, USA) while crystal violet was obtained from Mol (Belgrade, Serbia). The Penicillin Streptomycin solutionsolution was bought from Biological Industries (Cromwell, CT, USA). MCF7 (breast adenocarcinoma), HeLa (cervical adenocarcinoma) and PC-3 prostate (adenocarcinoma) cell lines were obtained from American Type Culture Collection (Rockville, MD, USA) while the A2780 (ovarian carcinoma) cells were from European Collection of Authenticated Cell Cultures (Salisbury, UK). Cell lines MCF7 and PC-3 were routinely cultivated in HEPESTM-buffered RPMI-1640 medium while HeLa and A2780 cells were cultured in DMEM. Both mediums were supplemented with 10% heat-inactivated FCS, 2 mM l-glutamine, 0.01% sodium pyruvate, penicillin (100 units per mL) and streptomycin (100 μg mL⁻¹) and cells were maintained at 37 °C in a humidified atmosphere with 5% CO₂. For viability screening MCF7 and HeLa cells were seeded at 2 x 10³ cells per well, PC-3 cells at 5 x 10³ cells per well and A2780 in at 3 x 10³ cells per well overnight and then treated with a wide range of concentration of A. vulgaris extracts for 72 h. Peritoneal exudates cells were collected from the peritoneal cavity of C57BL/6 mice by rinsing with ice-cold PBS. Mice were originated from the facility at the Institute for Biological Research “Siniša Stanković” (IBISS), University of Belgrade (Belgrade, Serbia). Cells were washed with PBS, seeded at 1.5 x 10⁵ cells per well in 96-well plates and allowed to adhere overnight. Before treatment, non-adherent cells were removed and cells were then exposed to the same concentration range of isolated extracts.

**Determination of cell viability by mitochondrial dehydrogenase (MTT) and crystal violet (CV) assays**

The activity of mitochondrial dehydrogenase was defined by a reduction of MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) to formazan. Cells were seeded overnight and treated with experimental compounds for 72 hours. Afterward, cells were cultivated in the presence of 0.5 mg mL⁻¹ MTT (Sigma, ST Louis, MO, USA) from 30 min 1 h. Thereafter, the medium was discarded and cells were lysed in DMSO. The conversion of MTT to formazan by metabolically viable cells was measured at 540 nm and cell viability was calculated as a percentage of control, untreated cells.

For CV test, cells were fixed with 4% of paraformaldehyde (PFA) and stained with 1% CV solution for 15 min at RT. Afterward, cells were washed and dried. Dye was dissolved in acetic acid and the absorbance was measured at 540 nm.

**Statistical analysis**

For data analysis, the Statistica 12 software was used. To analyze the significance of the differences between applied treatments the analysis of variance (ANOVA), followed by the Student-Newman-Keuls test was used. A p value less than 0.05 were considered significant.

**Results and discussion**

**LC-MS/MS characterization of phenolic compounds**

It is well-known that the flavonoids and their glycosides are responsible for antioxidant, anticancer, and cardioprotective activity. LC-MS/MS analysis was carried out to get insight into polyphenolic profile of A. vulgaris extracts and results are presented in the Table 1. The study showed variations in the amounts of phenolic compounds among the investigated extracts. The obtained results have shown clearly that EA extract was the richest in phenolic compounds. On the other hand, the number of quantified phenols and their amounts was notable lower than in the water extract. Phenolic acids, mainly the gallic...
and caffeic acids were the most presented and recorded for EA extract only. Generally, the amount of gallic and caffeic acids was about ten times higher comparing with the other phenolic acids in all tested extracts. Also, we can notice that chlorogenic acid was quantified in the highest amount in EtOH extract but in a lowest amount in EA extract. The health-benefits of this phenolic acid is correlated to several health benefits including antioxidant, anticancer, antimutagenic, anti-diabetic effects.\(^{29,30}\) Duckstein \textit{et al.}\(^{31}\) have detected in \textit{Alchemilla vulgaris} extracts a significant concentration of phenolic acids, such as chlorogenic, gallic, ellagic acids in lower concentrations.

Besides phenolic acids, the flavonoids catechin, and quercetin were recorded in significant concentration in EA extract, where catechin was the most dominant (8144.98 \(\mu \text{g g}^{-1}\)). The catechin was also determined in MeOH and EtOH extract, but in a much lower value (704.55 \(\mu \text{g g}^{-1}\) and 396.16 \(\mu \text{g g}^{-1}\)) than in EA extract. Quercetin was found in significant concentration in EA extract, where the EtOH (1186.34 \(\mu \text{g g}^{-1}\)) and MeOH (903.94 \(\mu \text{g g}^{-1}\)) extracts and apigenin-7-O-\(\beta\)-glucoside was detected in much lower concentration in all extracts. Further, kaempferol-3-O-glucoside and quercitrin-hexosides were noticed in ET extract in plentiful amount (1038.52 \(\mu \text{g g}^{-1}\); 2274.85 \(\mu \text{g g}^{-1}\)). Due to the fact that the information related to phenol profile of this plant is very scattered and that there are no reports for \textit{A. vulgaris} from this region (locality), the results obtained in this study represent a significant contribution to ethnobotany and phytomedicine.

### Total phenolic content and antioxidant activity

The results presented in Table 2 have shown the means of the total phenolics content and antioxidant activity of \textit{A. vulgaris} extracts using several in vitro assays. The total phenolic content were strongly correlated with antioxidant activity, whereas the EA extract showed the highest content of total phenol compounds (9.65 mg GAE per g) of extract and therefore the highest antioxidant activity.

Uniformly, it can be concluded that the EA extract demonstrated the highest antioxidant potential, where the most pronounced and significant antioxidant effect of tested extracts was observed for DPPH and FRAP assays (DPPH: 502.56 mg TE per g extract; FRAP: 8745.31 mg EAA per g of dry extract), followed by MeOH extract. The \(\text{H}_2\text{O}\) extract has shown significantly low anti-radical activity compared to the other extracts. It is not unusual for antioxidant assays to be less sensitive and accurate to some bioactive compounds that are presented in the testes.

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**Table 1** The concentrations of selected phenols in \textit{Alchemilla vulgaris} extracts obtained by LC-MS/MS

<table>
<thead>
<tr>
<th>Compounds(^a)</th>
<th>MeOH</th>
<th>EtOH</th>
<th>EA</th>
<th>(\text{H}_2\text{O})</th>
</tr>
</thead>
<tbody>
<tr>
<td>(p)-Hydroxybenzoic acid</td>
<td>&lt;\text{loq}</td>
<td>&lt;\text{loq}</td>
<td>135.45 ± 0.21</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Protocatechuic acid</td>
<td>&lt;\text{loq}</td>
<td>&lt;\text{loq}</td>
<td>255.94 ± 0.36</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>2,5-Dihydroxybenzoic acid</td>
<td>&lt;\text{loq}</td>
<td>74.00 ± 1.52</td>
<td>45.74 ± 0.01</td>
<td>3.35 ± 2.21</td>
</tr>
<tr>
<td>(p)-Coumaric acid</td>
<td>25.68 ± 1.12</td>
<td>23.02 ± 0.33</td>
<td>470.21 ± 0.44</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Gallic acid</td>
<td>&lt;\text{loq}</td>
<td>&lt;\text{loq}</td>
<td>2465.79 ± 0.01</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Esculetin</td>
<td>&lt;\text{loq}</td>
<td>&lt;\text{loq}</td>
<td>353.03 ± 0.11</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Caffeic acid</td>
<td>&lt;\text{loq}</td>
<td>69.19 ± 1.11</td>
<td>1138.54 ± 0.30</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Quinic acid</td>
<td>284.56 ± 0.17</td>
<td>544.85 ± 0.90</td>
<td>&lt;\text{loq}</td>
<td>157.99 ± 0.45</td>
</tr>
<tr>
<td>Ferulic acid</td>
<td>&lt;\text{loq}</td>
<td>&lt;\text{loq}</td>
<td>346.87 ± 0.12</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Genistein</td>
<td>&lt;\text{loq}</td>
<td>&lt;\text{loq}</td>
<td>94.01 ± 0.10</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Apigenin</td>
<td>&lt;\text{loq}</td>
<td>16.30 ± 0.01</td>
<td>501.97 ± 0.41</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Naringenin</td>
<td>&lt;\text{loq}</td>
<td>&lt;\text{loq}</td>
<td>15.40 ± 0.70</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Luteolin</td>
<td>23.15 ± 0.44</td>
<td>50.88 ± 1.08</td>
<td>638.09 ± 0.02</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Kaempferol</td>
<td>&lt;\text{loq}</td>
<td>&lt;\text{loq}</td>
<td>364.00 ± 0.22</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Catechin</td>
<td>704.55 ± 0.54</td>
<td>396.16 ± 0.01</td>
<td>8144.98 ± 0.01</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Chrysoeriol</td>
<td>&lt;\text{loq}</td>
<td>6.714 ± 1.36</td>
<td>222.63 ± 0.99</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Quercetin</td>
<td>&lt;\text{loq}</td>
<td>&lt;\text{loq}</td>
<td>4541.70 ± 0.04</td>
<td>&lt;\text{loq}</td>
</tr>
<tr>
<td>Chlorogenic acid</td>
<td>258.01 ± 1.05</td>
<td>510.43 ± 1.66</td>
<td>23.13 ± 0.11</td>
<td>74.69 ± 0.33</td>
</tr>
<tr>
<td>Apigenin-7-O-(\beta)-glucoside</td>
<td>40.20 ± 2.11</td>
<td>119.61 ± 0.10</td>
<td>141.16 ± 0.15</td>
<td>6.43 ± 0.01</td>
</tr>
<tr>
<td>Luteolin-7-O-(\beta)-glucoside</td>
<td>903.94 ± 0.03</td>
<td>1186.34 ± 0.24</td>
<td>329.71 ± 0.04</td>
<td>106.65 ± 1.05</td>
</tr>
<tr>
<td>Quercitrin</td>
<td>84.58 ± 1.05</td>
<td>168.56 ± 2.02</td>
<td>70.37 ± 2.15</td>
<td>19.72 ± 0.01</td>
</tr>
<tr>
<td>Kaempferol-3-O-glucoside</td>
<td>195.55 ± 0.33</td>
<td>151.52 ± 1.05</td>
<td>1038.52 ± 1.01</td>
<td>30.75 ± 2.30</td>
</tr>
<tr>
<td>Quercitrin-hexosides</td>
<td>424.27 ± 0.55</td>
<td>853.40 ± 1.10</td>
<td>2274.85 ± 1.12</td>
<td>86.27 ± 1.45</td>
</tr>
<tr>
<td>Rutin</td>
<td>71.06 ± 0.05</td>
<td>64.63 ± 1.17</td>
<td>&lt;\text{loq}</td>
<td>32.66 ± 2.05</td>
</tr>
</tbody>
</table>

\(^a\) Values are means ± SD of three measurements; <\text{loq} — less than limit of quantification.
samples. Therefore, more than two assays should be used in order to obtain more completed results of antioxidant effect. Using DPPH, ABTS•⁺ and OH assays, Borja et al., 2018 have demonstrated a great antioxidant potential of the Lady’s mantle methanol extract which has been higher than synthetic antioxidant.

**Enzyme inhibitory effects of the *A. vulgaris* extracts**

To determine enzyme inhibitory effects of *A. vulgaris* extracts, cholinesterases, tyrosinase and amylase were selected as target enzymes. The results are presented in Table 3. The highest AChE inhibition was provided by EA extract (5.21 mg GALAE per g) and other extracts had almost similar inhibitory effects on the enzyme. The activity for EA may be linked with the higher level of phenolics that are presented in the tested samples.

<table>
<thead>
<tr>
<th>Enzyme inhibition assays</th>
<th>MeOH</th>
<th>EtOH</th>
<th>EA</th>
<th>H₂O</th>
</tr>
</thead>
<tbody>
<tr>
<td>AChE inhibition (mg GALAE per g extract)</td>
<td>5.17 ± 0.02²</td>
<td>5.14 ± 0.01²</td>
<td>5.21 ± 0.05²</td>
<td>5.17 ± 0.02²</td>
</tr>
<tr>
<td>BChE inhibition (mg GALAE per g extract)</td>
<td>9.59 ± 0.14²</td>
<td>9.71 ± 0.09²</td>
<td>9.61 ± 0.02²</td>
<td>10.19 ± 0.16²</td>
</tr>
<tr>
<td>Tyrosinase inhibition (mg KAE per g extract)</td>
<td>79.84 ± 1.61³</td>
<td>78.55 ± 3.83³</td>
<td>77.66 ± 2.64³</td>
<td>73.68 ± 2.11³</td>
</tr>
<tr>
<td>Amylase inhibition (mmol ACAE per g extract)</td>
<td>0.34 ± 0.04³</td>
<td>0.32 ± 0.03³</td>
<td>0.41 ± 0.03³</td>
<td>0.22 ± 0.03³</td>
</tr>
</tbody>
</table>

Values expressed are means ± S.D. of three parallel measurements. Different letters mean significant differences (p < 0.05); GALAE: galatamine equivalent; KAE: kojic acid equivalent; ACAE: acarbose equivalent.

| **Table 2** Evaluation of antioxidant potential and total phenolic content³ |

<table>
<thead>
<tr>
<th>Extract</th>
<th>MeOH</th>
<th>EtOH</th>
<th>EA</th>
<th>H₂O</th>
</tr>
</thead>
<tbody>
<tr>
<td>DPPH (mg TE per g extract)</td>
<td>153.30 ± 0.013⁴</td>
<td>95.99 ± 0.003⁴</td>
<td>502.56 ± 0.01⁤</td>
<td>89.25 ± 0.02⁤</td>
</tr>
<tr>
<td>ABTS•⁺ (mg TE per g extract)</td>
<td>143.55 ± 3.65⁴</td>
<td>119.62 ± 3.20⁴</td>
<td>174.05 ± 0.90⁣</td>
<td>37.50 ± 0.39⁣</td>
</tr>
<tr>
<td>CUPRAC (mg TE per g extract)</td>
<td>216.14 ± 6.86⁣</td>
<td>203.53 ± 9.29⁣</td>
<td>283.16 ± 12.36⁣</td>
<td>78.56 ± 0.26⁣</td>
</tr>
<tr>
<td>PhosphomolibdCV (mmol TE per g extract)</td>
<td>1.77 ± 0.01⁣</td>
<td>1.57 ± 0.02⁣</td>
<td>2.22 ± 0.07⁣</td>
<td>0.53 ± 0.03⁣</td>
</tr>
<tr>
<td>Metal chelating (mg EDTAE per g extract)</td>
<td>42.58 ± 0.26⁣</td>
<td>42.32 ± 0.05⁣</td>
<td>37.96 ± 1.29⁣</td>
<td>39.23 ± 0.32⁣</td>
</tr>
<tr>
<td>Reducing power (mg EAA per g of extract)</td>
<td>7831.45 ± 0.43⁣</td>
<td>6405.75 ± 0.08⁣</td>
<td>8745.31 ± 0.04⁣</td>
<td>2420.09 ± 0.08⁣</td>
</tr>
<tr>
<td>Total phenolics content (mg GAE per g of extract)</td>
<td>7.71 ± 0.01³</td>
<td>7.40 ± 0.03³</td>
<td>9.65 ± 0.02³</td>
<td>6.89 ± 0.03³</td>
</tr>
</tbody>
</table>

 Values expressed are means ± S.D. of three parallel measurements; a–d means in the same row not sharing the same superscript are significantly different (p < 0.05); TE: trolox equivalent; EDTAE: EDTA equivalent; GAE: gallic acid equivalent.

**Table 4** IC₅₀ values of effect *Alchemilla vulgaris* extracts in cytotoxicity

<table>
<thead>
<tr>
<th>Cell line</th>
<th>Assay</th>
<th>MeOH</th>
<th>EtOH</th>
<th>EA</th>
<th>H₂O</th>
</tr>
</thead>
<tbody>
<tr>
<td>A2780</td>
<td>MTT</td>
<td>27.9 ± 1.9</td>
<td>40.3 ± 0.4</td>
<td>43.3 ± 0.9</td>
<td>67 ± 2.4</td>
</tr>
<tr>
<td></td>
<td>CV</td>
<td>38.3 ± 1.1</td>
<td>43.9 ± 0.1</td>
<td>42.6 ± 1.3</td>
<td>78.4 ± 1.9</td>
</tr>
<tr>
<td>HeLa</td>
<td>MTT</td>
<td>80.6 ± 6.3</td>
<td>84.75 ± 5</td>
<td>46.4 ± 5.1</td>
<td>&gt;100</td>
</tr>
<tr>
<td></td>
<td>CV</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>53.8 ± 3.9</td>
<td>&gt;100</td>
</tr>
<tr>
<td>MCF7</td>
<td>MTT</td>
<td>41.9 ± 0.6</td>
<td>58.7 ± 1.6</td>
<td>31.3 ± 1.5</td>
<td>94.1 ± 1.2</td>
</tr>
<tr>
<td></td>
<td>CV</td>
<td>47.2 ± 0.4</td>
<td>64.4 ± 0.1</td>
<td>37.5 ± 5</td>
<td>95 ± 7.1</td>
</tr>
<tr>
<td>PC-3</td>
<td>MTT</td>
<td>54.4 ± 2.5</td>
<td>43.5 ± 1.1</td>
<td>18.7 ± 0.9</td>
<td>&gt;100</td>
</tr>
<tr>
<td></td>
<td>CV</td>
<td>73.5 ± 4.2</td>
<td>61.1 ± 4.7</td>
<td>31.7 ± 0.3</td>
<td>&gt;100</td>
</tr>
</tbody>
</table>

Values expressed are means ± S.D. of three parallel measurements. Differences of IC₅₀ for extract were compared using Kruskal-Wallis test, and different letters mean significant differences (p < 0.05).
ethnobotanical notes for the Balkan region\textsuperscript{15} and as a novel report presented in our current study – for treating the neurodegenerative disorders, including Alzheimer and Parkinson’s disease.

**Anticancer activity of \textit{A. vulgaris} extracts using different cancer cell lines**

To evaluate the impact of isolated extracts on tumor cell growth, human breast MCF7, ovarian A2780, cervical HeLa and prostate cancer PC-3 cell lines were used. All selected cell lines were isolated from tumors developed in tissues regulated by hormones and accordingly, highly sensitive to hormonal status. During tumor progression, some of them like A2780, HeLa and PC-3 lost their hormone receptors and became independent to the treatment with hormone antagonist.\textsuperscript{45–48} This qualified them as invasive and, in general, resistant to conventional chemotherapeutics. Having all this in mind, it was intriguing to assess sensitivity of hormone-dependent (MCF-7) as well as independent cell lines (A2780, HeLa, PC-3) to different extracts of \textit{A. vulgaris}, whose composition varied depending on the type of the solvent used in extraction procedure. Cells were cultivated with isolated extracts for 72 h when the number of viable cells in the culture was assessed by MTT and CV tests. As presented in Table 4 and Fig. 1, H$_2$O extract was almost inefficient

![Fig. 1](image_url)

**Fig. 1** Effect of \textit{Alchemilla vulgaris} extracts on different tumor cell lines.
in tumor cell growth suppression. Three other extracts displayed the different potential to decrease the number of viable cells depending on the cell type. In general, despite different origin and sensitivity to hormone stimulation of tested cell lines the strongest potential to decrease tumor cells’ viability possessed EA extract. While cervical adenocarcinoma HeLa was almost insensitive to MeOH and EtOH, ovarian A2780 and breast carcinoma MCF7 displayed better response to treatment with MeOH than EtOH extract. To evaluate the selectivity toward malignant phenotype, peritoneal exudates cells isolated from C57BL6 mice were exposed to the same range of doses as those used for evaluation of the cytotoxicity against malignant cells. Differently, to tested cancer cell lines, MeOH and EtOH extracts were not able to reduce the viability of normal cells for 50%. Since IC_{50} value was not reached upon the treatment with indicating extracts, it can be concluded that the inttested range of doses they are non-toxic for normal phenotype and therefore, selective to malignant cells. Differently, to them, IC_{50} value for EA was determined at 100 µg mL^{-1} but, regarding to high sensitivity of all tested lines to this extract, selectivity index varies from 2–3, depending on the type of tumor cells. Taking together, apart from its confirmed antioxidant and anti-inflammatory action, A. vulgaris extracts might directly impact the growth of malignant cells. Especial importance represents the fact the even hormone independent cell lines originating from tumors known as highly aggressive and nonresponsive to conventional therapy were sensitive to tested extracts. Despite to ethno-botanical data showing the anticancer potential of A. vulgaris against female reproductive tissues we might speculate that male reproductive cancers, like in this case prostate cancer, or other type of tumors can be targeted with this plant constituents. Varying in efficacy of used A. vulgaris extract against tested cell lines might be ascribed to the differences in content, concentration and ratio of biologically active compounds with proven anticancer activity. Namely, it is well recognized that both caffeic and gallic acid, that are present in EA extract, possess strong anti-proliferative and cytotoxic properties in wide range of tumor types such as those used in this study (breast, cervical, ovarian, prostate carcinoma). These phenolic acids affected main signaling pathways involved in cell cycle progression and cell death. Moreover, they interfered with migrative and invasive properties, epithelial/mesenchimal transition and even angiogenesis. Very important property of mentioned phenolic compounds is their ability to sensitize tumor cells to conventional therapeutics or even to affect cells initially resistant to chemotherapy. In addition, genistein, protein tyrosine kinase inhibitor, and quercetin, present in EA extract only, displayed strong anticancer potential realized preferentially through inhibition of cell cycle progression, induction of apoptosis affecting MAP-ERK signaling pathway, inhibition of metastatic properties of malignant cells etc. The anticancer properties of these molecules were even proved in several in vivo tumor models. Concordantly, with described data these isoflavones might sensitize malignant cells to conventional therapies. Since quercetin inhibited stem cell proliferation, its presence might be highly beneficial in terms of treatment of aggressive malignant tumors. The rest of active compounds for example, catechin, are also compounds with potential to limit tumor cell growth. Apart from discussed antitumor properties, catechin modulated epigenetic events such as DNA methylation and demethylation, as well as acetylation deacetylation of histones, influence tubulin-microtubule equilibrium and changed the expression of enzymes relevant for migration and invasion of malignant cells. Plenty of literature data confirmed that efficacy of any herbal extract is defined by the mixture and interplay of its constituents. Despite the lack of scientific data about the anticancer features of A. vulgaris the panel of biologically active substances inside of it indicated its enormous potential in the field of anticancer therapy and needs of further evaluation.

Conclusion

Alchemilla vulgaris is a generally poorly studied plant, apart from its long and known traditional use world wide. The results obtained by the phytochemical examination and the assessment of the biological activities of A. vulgaris extracts, indicated the justification of use of the plant in traditional medicine, mainly for woman related diseases due to hormone dependent anticancer activity (breast and ovarian anticancer effects), as well as amylase and tyrosinase inhibitory effects, which might be related to antidiabetic and skin-related diseases, as previously reported for Lady’s mantle of the Balkans region. Moreover, potential use in prevention of neuro-degenerative illnesses was shown by inhibitory effects on cholinesterases. Great biological activity of A. vulgaris extracts is linked with its phenolic profile, rich in catechin, quercetin and its hexoside, luteolin, apigenin, gallic and caffeic acids. Our results present a first report on anticancer activity of hormone dependent cancer lines, for this plant. The ethylacetate extract should be emphasized due its exceptional antioxidant, enzyme inhibitory and antitumor activity upon the content of the phenolic compounds such as catechin, quercetin and phenolic acids. Further research will be focused on mechanisms of anticancer activity and modes of action of the key most efficient plant metabolites. Overall this Alchemilla vulgaris from Vlasiška lake (central Balkan) is a promising a valuable source of different bioactive compounds that could have great health benefits.

Conflicts of interest

The authors declare there is no conflict of interest.

Acknowledgements

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