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First total synthesis of ampullosine, a unique isoquinoline alkaloid isolated from *Sepedonium ampullosporium*, and of the related permethylampullosine†

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A straightforward and convenient approach toward the first total synthesis of ampullosine, a structurally unique 3-methylisoquinoline alkaloid isolated from *Sepedonium ampullosporium*, is reported. Access to the related *O*-methyl ampullosine methyl ester from a common intermediate is also disclosed. The synthetic sequence toward the natural product comprised a Kolbe-type carboxylation of 3,5-dihydroxybenzoic acid and further esterification of the diacid, followed by masking of one of the phenols through selective ester reduction and subsequent acetonide formation. Installation of the three-carbon atom required for the 3-methylpyridine ring was performed by triflation of the remaining free phenol and a Pd-catalyzed Suzuki–Miyaura reaction with potassium *E*-propenyltrifluoroborate. Deprotection of the acetonide, followed by partial oxidation of the benzylic alcohol to the salicylaldehyde, *O*-methylation of the free phenol and hydrazone formation of the resulting *ortho*-anisaldehyde derivative gave a hydrazone-based 1-azatriene. This was further subjected to 6 π -aza-electrocyclization to afford permethylampullosine (11 steps, 14% overall yield), whereas exhaustive demethylation with AlI₃ generated *in situ* gave ampullosine (12 steps, 3.2% global yield).

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Introduction

The genus *Sepedonium* (Ascomycetes) is sapro-parasitic and comprises asexual states of fungi that parasitize on other fungi; more specifically basidiocarps of the mushroom order Boletales.¹ They operate by first settling on their fleshy living hosts, which they kill to employ the organic matter as nutrients to grow and develop. These fungicolous species have a mould-like aspect, with thick-walled gold-yellow colored aleurioconidia and colorless thin-walled phialoconidia.

The genus was established at the beginning of the XIX century. However, the seminal works of Petch and Pieschel² described variations of *S. chrysospermum*, namely *S. ampullosporium*,^{3a} *S. chalcipori*,^{3b} and *S. microspermum*,^{3c} which along with *S. laevigatum* ended up conforming autonomic species on the basis of sequence analysis of the internal transcribed spacer region of the nuclear ribosomal RNA genes.¹

Fungi of the genus *Sepedonium* have been studied as producers of antibiotics and pigments, as well as for their properties as antagonist agents of various plant pathogens.

Chemical investigations of *Sepedonium* spp. revealed that this species is an interesting source of various natural products with potential biopharmaceutical applications (Fig. 1), including peptaibols, uncommon peptides with a broad spectrum of biological activities,⁴ the unusual cyclic peptide chrysosporide,⁵ the azaphilone chrisodin (1),⁶ as well as some mono- and bis-anthraquinones (skyrin, rugulosin, chrysophanol)^{1,7} and tropolone derivatives (2–3),⁸ among others.⁵

Infected organisms are rarely colonized by secondary fungal parasites, to compete with *Sepedonium* for host nutrients. This observation supports the speculation that *Sepedonium* species produce small defense molecules that prevent further infections.

Recently, the group of Arnold examined the culture extract of *S. ampullosporium* and isolated ampullosine (4).⁹ This alkaloid, responsible for the deep yellow color of its culture fluid, is the first and only isoquinoline isolated from the genus *Sepedonium*. Its structure was established by spectroscopic means.

In addition, methylation of the heterocycle afforded *O*-methyl ampullosine methyl ester (5), which contributed to the identification of the natural product. Employing LC/ESI, the authors were able to detect ampullosine in ten out of twelve strains of *Sepedonium*, belonging to eight different species. However, they observed that the two most phylogenetically distant species lacked the alkaloid.

The structure of ampullosine is unique. Despite some 3-methylisoquinolines have been synthesized, the combination of this

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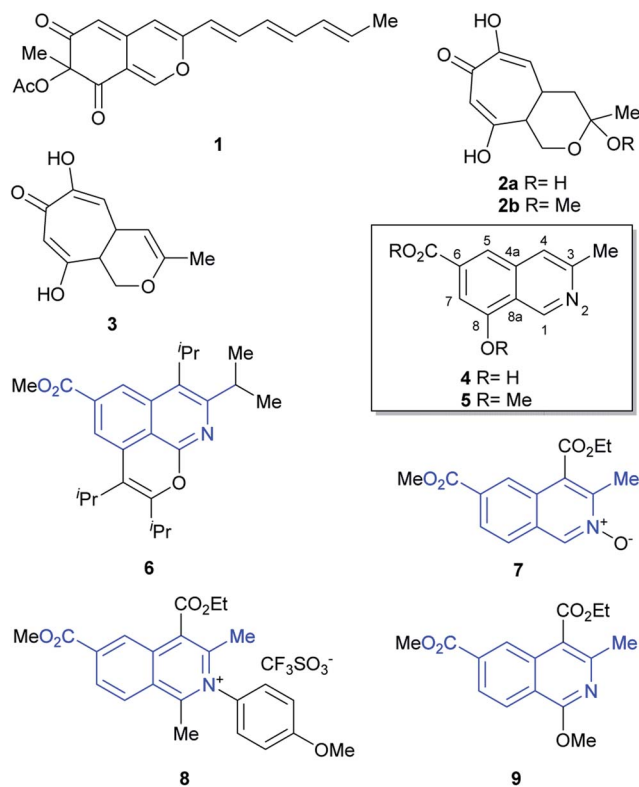


Fig. 1 Chemical structures of some polyketides (1–3) and ampuulosine (4), isolated from *Sepedonium* species, as well as the semisynthetic ampuulosine derivative (5) and structurally related, synthetic 3-methyl-6-carboxymethyl isoquinolines (6–9).

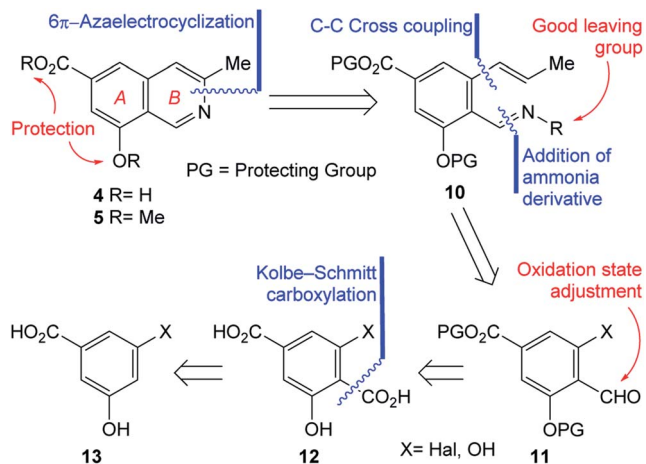
motif with a 6-carboxylic acid moiety is rare; further, although they are seldom found in patents,¹⁰ a few and scattered examples started to appear in the literature just recently, favored by the flourishing of research into C–H activation reactions based on ruthenium (6)¹¹ and rhodium (7–9)¹² catalysis. In many cases, however, the current limitations of this novel chemistry still turn unfavorable its use to approach ampuulosine.

We have been interested in the synthesis of structurally unique heterocyclic natural products¹³ and have used the 6 π -electrocyclization of 1-azatrienes as an efficient and atom-economic strategy toward the synthesis of natural products or their analogs, containing the 3-methyl isoquinoline motif.¹⁴

In pursuit of these interests, herein we wish to report the first total synthesis of ampuulosine (4) and the synthesis of *O*-methyl ampuulosine methyl ester (5), from a common intermediate, using the easily available 3,5-dihydroxybenzoic acid and employing a 6 π -azaelectrocyclization reaction as the key transformation, according to the retrosynthetic analysis depicted in Scheme 1.

Results and discussion

The initial N–C3 disconnection of the analysis was performed on 4 and 5 and unveiled the polysubstituted benzenoid 10 as a fit key intermediate for the proposed 6 π -azaelectrocyclization based ring closure toward both targets. The main features of 10 are a properly masked carboxyl group, a differentially protected



Scheme 1 Retrosynthetic analyses of ampuulosine (4) and its derivative 5.

phenolic moiety, according to the final targets and an *ortho*-disubstituted carbonyl condensed with an ammonia derivative which carries a suitable leaving group.

A couple of additional disconnections were carried out on 10 to uncover the salicylaldehyde derivative 11. The reasoning was that condensation with an ammonia derivative (oxime, hydrazone, etc.) could provide the required activated nitrogen atom for the pyridine ring of the target; in addition, it was conjectured that a suitable cross-coupling reaction with an allyl/propenyl derivative¹⁵ could perform the substitution of a properly located halogen or activated phenol, enabling the installation of the three carbon atoms chain, required to build the heterocyclic ring.

Further analysis of 11 determined that its substituent pattern could be advantageously simplified towards a suitable starting material, by considering an oxidation state adjustment of the aldehyde. This placed terephthalic acid derivative 12 in the path toward the natural product. In turn, the *ortho*-disubstituted carboxylic acid was disconnected, to reveal compound 13 as an apt starting material, considering the availability of protocols to access salicylic acid derivatives from phenols.¹⁶

Interestingly, despite different 3-halobenzoic acids have been reported in the literature,¹⁷ they exhibited some accessibility drawbacks; among them, lack of suitable precursors or efficient transformations from commercial materials. Therefore, it was concluded that among the possible candidates to fulfil the odd structural requirements imposed by the retrosynthetic analysis, only 3,5-dihydroxybenzoic (14) is inexpensive, easily available and displays the most ideal substitution pattern.

On the other hand, despite various methodologies are available for the *ortho* formylation of phenols, some protocols such as Duff, Skattebøl, Reimer–Tiemann, Vilsmeier–Haack and Gattermann–Koch require activated substrates, while others (*ortho* metalation) demand protection of the sensitive groups, such as the carboxyl moiety, and all of them may afford mixtures of isomeric products.¹⁸ Furthermore, some literature



precedents suggested that formylation of **13** may result in the undesired formyl derivative, and in low yield.¹⁹

Therefore, the synthesis commenced with **14** (Scheme 2), which was submitted to a Kolbe–Schmitt type carboxylation with KHCO_3 at 180 °C, under a CO_2 atmosphere (balloon).²⁰ It was observed that the use of glycerol as reaction medium resulted in a deep red coloured gummy mass, which diffculted extraction and product purification, affording **15** in poorly reproducible yields, below 60%. Luckily, however, employing a minimum amount of propylene glycol (final concentration of **14** was $\sim 4 \text{ mol L}^{-1}$) as solvent provided **15** as an easily recoverable pale yellow solid in better yield (80%).

Furthermore, similar performances were observed when the reaction was carried out in a closed vessel, purged with CO_2 , a requirement of the reaction mechanism.²¹ Without purification, the resulting solid mass containing the diacid product **15** was submitted to selective esterification with Me_2SO_4 providing the dimethyl ester **16** (70% yield for the two steps).

In turn, the diester was selectively reduced with NaBH_4 , under anchimeric assistance provided by the neighbouring phenol moieties to the *ortho*-disubstituted C4 carboxylate, and the resulting benzyl alcohol was converted to the acetonide **17** in 75% overall yield.

In order to install the projected three carbon atom chain for the heterocyclic ring, compound **17** was next activated toward a Stille cross-coupling reaction, by triflate ester formation with PhNTf_2 . The latter reagent was elected over the more reactive Tf_2O after considering previous failures of the anhydride in similar transformations.^{22a}

Luckily, the reaction of **17** with *N*-phenyl triflimide, employing Et_3N as base and DMAP as nucleophilic substitution

catalyst, furnished 96% of the triflate **18**.^{22b,c} Interestingly chromatographic elution with hexane: CH_2Cl_2 mixtures proved critical to obtain the product free from *N*-phenyltriflamide, a by-product of the transformation.

In continuation of the sequence, the triflate was submitted to a Stille cross-coupling reaction with ${}^n\text{Bu}_3\text{SnCH}_2\text{CH}=\text{CH}_2$ in diglyme, in the presence of LiCl and under $\text{PdCl}_2(\text{PPh}_3)_2$ catalysis. This afforded the expected allyl-substituted compound **19** in mixture (95 : 5) with the propenyl derivative **E-20** in combined 70% yield, as observed in its ${}^1\text{H}$ NMR spectrum. However, despite our experience, it was found comparatively difficult to obtain clean samples of **19**, devoid of undesirable contamination with tin residues, which somehow hindered a proper development of the projected isomerization step toward **20**.

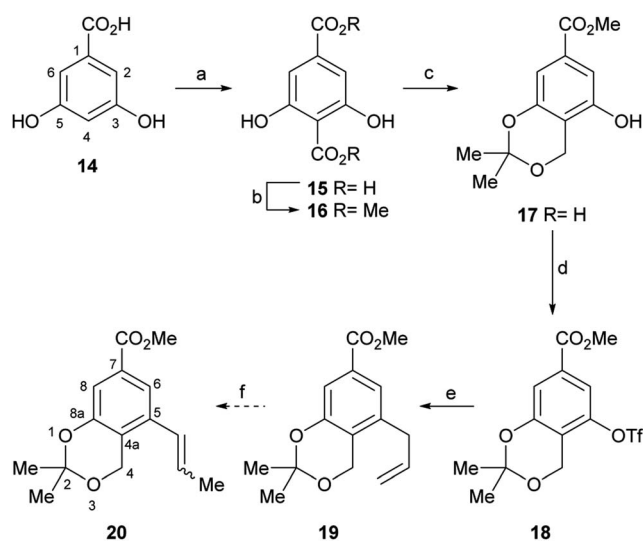
Therefore, an alternative was sought. Fortunately for our needs, due to their improved air-stability, reduced cost and improved nucleophilicity, the use of organotrifluoroborate salts has emerged²³ as an excellent alternative to the boronates and boronic acids, expensive conventional substrates for the Suzuki–Miyaura reactions.²⁴ Their advantages are more evident when it is also considered that vinylboronic acids often require excess reagent to achieve better yields²⁵ and suffer from stability issues, tending to polymerize.²⁶

Therefore, the modification of the Suzuki–Miyaura reaction pioneered by Molander was implemented, and the cross-coupling of **18** was performed with potassium propenyl trifluoroborate (**21**)²⁷ and Cs_2CO_3 as base. The reaction required optimization (Table 1), since the initial results with $\text{Pd}(\text{OAc})_2$ as the metal source and DavePhos as ligand afforded only moderate yields of **20** (53–58%) when the reaction was run in a toluene–water system (4 : 1, v/v) at 90–100 °C during 16–24 h, respectively (entries 1 and 2).

Furthermore, under these conditions, the required product was observed in mixture with minor amounts of the *Z*-isomer **22** (3–4%) and with the allyl derivative **19** (9–13%). This was somewhat surprising, since alkenyltrifluoroborates are known to retain the double bond geometry with a high degree of fidelity.^{24b}

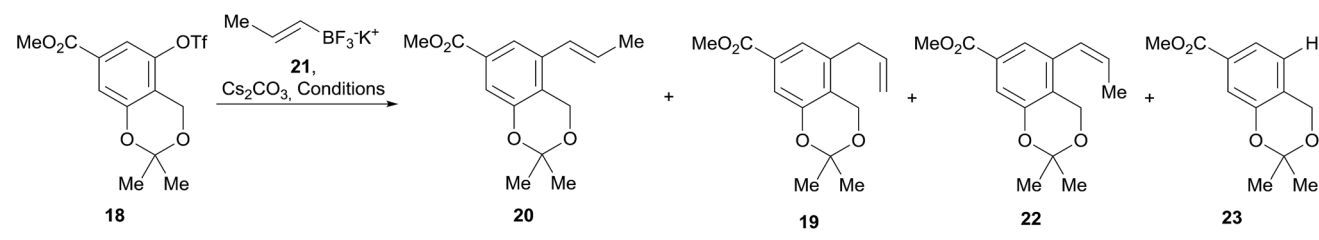
Changing the palladium source to $\text{PdCl}_2(\text{PPh}_3)_2$ did not met with much better success (entries 3 and 4) when DPPF was employed as ligand (36% yield) or in the presence of DavePhos in dioxane/ H_2O (10 : 1, v/v). Similar moderate results (53–56% yield) were obtained with the use of $\text{Pd}(\text{OAc})_2$ and DavePhos in $\text{THF}/\text{H}_2\text{O}$ (4 : 1, v/v) at 70 °C for 15 h (entry 5), where some unreacted starting material (10%) was concomitantly isolated, along with the proto-deoxygenated product **23** (12%), recognized through the diagnostic signals in its ${}^1\text{H}$ NMR spectrum [δ 7.55 (dd, $J = 1.5$ and 7.8, H-2), 7.50 (d, $J = 1.5$, H-6) and 7.03 (d, $J = 7.8$, H-3)].

Employing a toluene/water solvent in association with $\text{PdCl}_2(\text{PPh}_3)_2$ was found to afford significantly improved results, attaining 85% yield of **20** (entry 6). This performance could not be surpassed by the use of *t*-BuPhos as ligand (entry 7) nor by microwave irradiation (entry 8), which gave **20** in 68–69% yield. In all these cases, mixtures of **19**, **20** and **22** were observed.



Scheme 2 Reagents and conditions: (a) CO_2 (sealed tube), KHCO_3 , 1,2-propanediol, 180 °C, 6 h; (b) Me_2SO_4 , KHCO_3 , acetone, reflux, 10 h, (70% overall); (c) (1) NaBH_4 , $\text{THF}/\text{H}_2\text{O}$ (4 : 1), 0.1 M phosphate buffer pH 7.5, 0 °C \rightarrow r.t., 1.5 h; (2) 2,2-DMP, TsOH , r.t., 5 h (75% overall); (d) PhNTf_2 , DMAP, NEt_3 , CH_2Cl_2 , r.t. 16 h, (96%); (e) ${}^n\text{Bu}_3\text{SnCH}_2\text{CH}=\text{CH}_2$, $\text{PdCl}_2(\text{PPh}_3)_2$, PPh_3 , LiCl , diglyme, 125 °C, 15 h (**19** and **20**, 95 : 5, 70% global); (f) $\text{RuCl}(\text{CO})(\text{PPh}_3)_3$ (cat.), PhMe , 80 °C.



Table 1 Optimization of the propenylation of the triflate **18**^a


Run no.	Catalyst	Ligand	Solvent	Temp. (°C)	Time (h)	Yield (%)				
						18	19	20	22	23
1	Pd(OAc) ₂	DavePhos	PhMe/H ₂ O ^b	100	16	—	13	53	4	—
2	Pd(OAc) ₂	DavePhos	PhMe/H ₂ O ^b	90	24	—	9	58	3	—
3	PdCl ₂ (PPh ₃) ₂	DPPF	PhMe/H ₂ O ^b	100	15	—	—	36	—	—
4	PdCl ₂ (PPh ₃) ₂	DavePhos	Dioxane/H ₂ O ^c	100	15	—	3	47	3	2
5	Pd(OAc) ₂	DavePhos	THF/H ₂ O ^d	70	15	10	—	56	—	12
6	PdCl ₂ (PPh ₃) ₂	DavePhos	PhMe/H ₂ O ^b	100	15	—	4	85	4	—
7	PdCl ₂ (PPh ₃) ₂	^t BuPhos	PhMe/H ₂ O ^b	100	15	4	3	68	4	—
8	PdCl ₂ (PPh ₃) ₂	DavePhos	PhMe/H ₂ O ^{b,e}	100	0.7	—	5	69	6	—
9	PdCl₂(PPh₃)₂	DavePhos	THF/H₂O^{d,f}	80	24	—	—	95	—	—

^a The experiments were carried out at a final substrate concentration of 0.1 M in the mixture of solvents, employing 1.6 equiv. of the trifluoroborate salt, 3 equiv. Cs₂CO₃, 5 mol% of the Pd catalyst and 10 mol% of the ligand. ^b A PhMe/H₂O 4 : 1 (v/v) medium was employed. ^c The reaction was carried in a dioxane/H₂O 10 : 1 (v/v) medium. ^d A THF/H₂O 4 : 1 (v/v) medium was used. ^e The reaction was carried out under microwave irradiation (250 W). ^f LiCl (0.4 equiv.) was added.

Examining the whole set of results, it was noticed that the deoxygenated product **23** was obtained only in ethereal solvents. It was conjectured that this outcome may be the result of the lack of reactivity or stability of some reaction intermediate. Thus, it was thought that addition of LiCl may improve the results.

It has been shown that LiCl is a required additive when the transformation is run in ethereal solvents, because it may act as a source of chloride ligand, stabilizing the Pd intermediates and making the cross-coupling stage more efficient.²⁸ It has also been demonstrated that the lithium salt turns the Pd catalyst more active towards transmetalation and increases its proneness to undergo oxidative addition, resulting in a powerful accelerant of the reaction rate. Furthermore, LiCl enhances both the polarity of the solvent, and the leaving ability of anionic ligands.

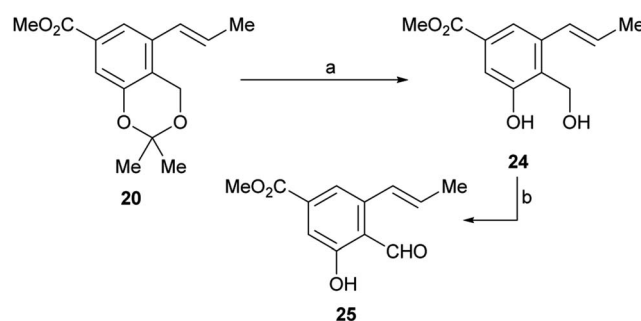
Therefore, LiCl was added to a THF/H₂O mixture of the starting triflate, Cs₂CO₃, PdCl₂(PPh₃)₂ and DavePhos. Rewardingly, the LiCl supplemented reaction cleanly afforded 95% yield of the expected derivative **20**, free of isomers and under milder conditions (80 °C), as shown in entry 9. Next, the acetone moiety of **20** was smoothly hydrolysed by exposure to *p*-toluenesulfonic acid in THF/H₂O, affording the benzylic alcohol **24** in almost quantitative yield (Scheme 3).

However, oxidation of the latter to the corresponding aldehyde **25**, which is a key step of the synthesis, proved to be tricky. The starting alcohol demonstrated to exhibit some instability, probably favoured by an easy benzylic protonation, followed by dehydration and further quinomethide formation; on the other

hand, the tested oxidants were also able to oxidize the neighbouring phenol moiety to the related quinones.

As shown in Table 2, a series of trial and error experiments with PDC,²⁹ IBX³⁰ and activated MnO₂ (ref. 31) were executed. After initial discouraging results (entries 1–3) where **25** was obtained (20–34% yield) along with unidentifiable products, presumably resulting from overoxidation,³² it was found that MnO₂ provided a significantly better yield when the solvent was changed from CH₂Cl₂ to EtOAc (entry 4).

Additionally, it was observed that warming to 50 °C improved the performance to 54% yield (entry 5), also shortening the reaction time from 16 h to 5 h, whereas the best results (55% yield) were achieved in boiling EtOAc for 3 h (entry 6), when it was observed complete consumption of the starting material.



Scheme 3 Reagents and conditions: (a) TsOH (cat.), THF/H₂O, 60 °C, 48 h, (98%) or 1 M HCl, THF, 45 °C, 18 h, (90%); (b) see Table 2.



After having secured a satisfactory access to the key propenyl salicylaldehyde **25**, the latter was submitted to our recently developed one-pot hydrazonation/ 6π -aza-electrocyclization protocol.³³ Thus, **25** was exposed to 1,1-dimethylhydrazine in PhCF₃ with the addition of AcOH as a promoter, to afford the hydrazone intermediate **26** (Scheme 4). Without purification, the hydrazone was heated at 160 °C for 45 min in the same medium, under microwave irradiation. Unexpectedly, however, the process resulted in recovery of 50% of the starting aldehyde **25**, and no signs of the expected product **27** could be detected.

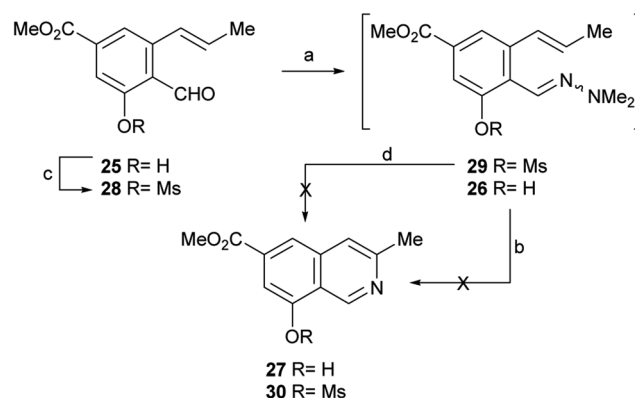
Under the hypothesis that the presence of a free phenolic hydroxyl group could have caused the observed outcome, **25** was protected with a readily removable group. Therefore, **25** was treated with MeSO₂Cl and Et₃N and DMAP in CH₂Cl₂, to afford 55% yield of the unstable mesylate ester **28**. The latter exhibited diagnostic signals of the methanesulfonate group, as a singlet resonating at δ_{H} 3.32 ppm and a resonance at δ_{C} 38.6 ppm.

Unfortunately, however, application of the one-pot hydrazonation/ 6π -aza-electrocyclization protocol also met with failure; neither the expected mesylate **30** nor the isoquinoline **27** were found.³³

In view of these discouraging results, an alternate protecting group was sought. Considering our previous results with the benzyl ether and MOM as protecting groups,³³ and taking into account our objective, to synthetically access permethylampullosine (**5**), the intermediate **25** was *O*-methylated with MeI in DMF at 0 °C, employing K₂CO₃ as base, to give **31** in 85% yield (Scheme 5). Then, the latter was submitted to our hydrazonation/ 6π -aza-electrocyclization/elimination protocol,³³ affording **5** in 64% overall yield, through the intermediacy of **32a**, which was not isolated because of its poor hydrolytic stability.

Interestingly, the 3,4-dihydroisoquinoline **33** was isolated concomitantly in 26% yield. The latter was recognized by its pattern of signals in the ¹H NMR spectrum, related to H-3 (m), H-4 (dd) and Me-3 (d). A reaction mechanism to explain the generation of 3,4-dihydroisoquinolines as side products of 6π -aza-electrocyclization of similar heterocyclic systems has been proposed.³³

It has been shown that, in aqueous solution, aliphatic hydrazones are 10²–10³ times more sensitive to hydrolysis than the analogous oximes.³⁴ Therefore, conjecturing that the use of a more stable ammonia derivative for condensation with **31** may improve the performance of the 6π -aza-electrocyclization, **31**

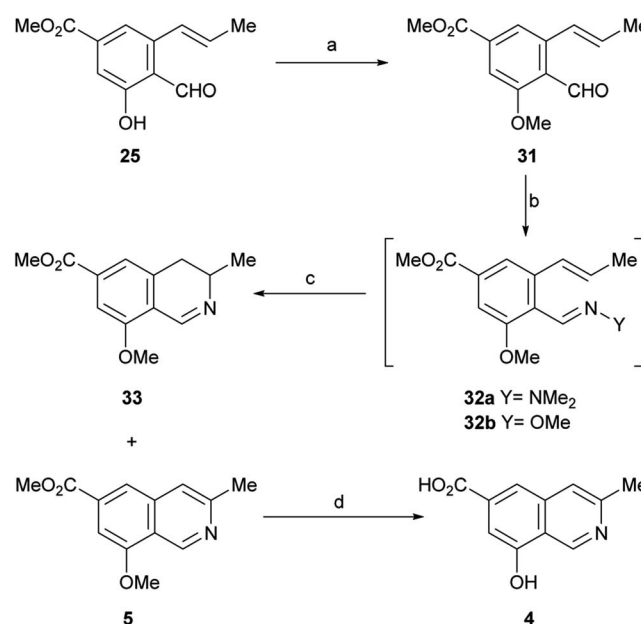


Scheme 4 Reagents and conditions: (a) Me₂N–NH₂, AcOH, PhCF₃, r.t., 1.5 h; (b) MW, 160 °C, 45 min (**25**, 50% recovered); (c) MsCl, Et₃N, CH₂Cl₂, 0 °C → r.t., 24 h (55%); (d) MW, 160 °C, 45 min.

was converted into the methoxime intermediate **32b** by reaction with methoxylamine in absolute EtOH. After replacing the solvent with PhCF₃, compound **32b** was heated under microwave irradiation at 180 °C for 45 min to undergo the 6π -aza-electrocyclization, furnishing **5** in 50% yield, accompanied by the related 3,4-dihydroisoquinoline **33** (15% yield).

With these results it was concluded that the hydrazone pathway should be preferred to the methoxime alternative. The ¹H and ¹³C NMR spectroscopic data of the synthetic compound **5** fully matched those reported by the group of Arnold for their semisynthetic *O*-methyl ampullosine methyl ester.⁹

It was also found that treatment of **5** with aluminium and iodine (which generates *in situ* the powerful Lewis acid AlI₃),³⁵



Scheme 5 Reagents and conditions: (a) MeI, K₂CO₃, DMF, 0 → r.t., 1.5 h (85%); (b) Me₂N–NH₂, AcOH, PhCF₃, r.t., 1.5 h or MeO–NH₂, EtOH, r.t., 1 h; (c) MW, 160 °C (via **32a**: 1 h, **5**, 64%; **33**, 26%; via **32b**: 45 min, **5**, 50%; **33**, 15%); (d) AlI₃, DMSO, MeCN, 80 °C, 20 h (23%).

Table 2 Optimization of the oxidation of benzyl alcohol **24** to **25**

Run no.	Oxidizing agent	Solvent	Temperature (°C)	Time (h)	Yield (%)
1	IBX	EtOAc	50	1.5	23
2	PDC	CH ₂ Cl ₂	r.t.	2	34
3	MnO ₂	CH ₂ Cl ₂	r.t.	16	20
4	MnO ₂	EtOAc	r.t.	16	50
5	MnO ₂	EtOAc	50	5	54
6	MnO ₂	EtOAc	78	3	55



effected the simultaneous removal of both methyl groups, affording ampullosine (**4**) in 23% yield.

Experimental

General information

All the reactions were carried out under dry argon atmospheres, using oven-dried glassware and freshly distilled anhydrous solvents. Anhydrous CH_2Cl_2 was obtained from an M. Braun solvent purification and dispenser system. Anhydrous EtOAc was obtained by a 3 h reflux of the PA grade product over P_2O_5 and further distillation of the liquid. Anhydrous DMF was obtained by heating the PA grade product over BaO for 4 h, followed by distillation under reduced pressure. Anhydrous Et_3N was prepared by distillation of the commercial product from CaH_2 . The anhydrous solvents were stored in dry Young ampoules. All other solvents and reagents were used as received.

The reactions were monitored by TLC (silica gel 60 GF₂₅₄) run in different hexanes:EtOAc mixtures. The chromatographic spots were revealed by exposure to UV light (254 and 365 nm) and spraying with ethanolic *p*-anisaldehyde/sulfuric acid reagent, followed by careful heating to improve selectivity; to detect the isoquinolines, the Dragendorff reagent (Munier and Macheboeuf modification)^{36a} was used. The flash column chromatographies were performed under conditions that entailed a modification of known procedures.^{36b-d} They were run under positive pressure with slurry-packed silica gel 60 H for thin layer chromatography (particle size < 55 μm), employing gradient of solvent polarity techniques with hexane : EtOAc, hexane : CH_2Cl_2 and EtOAc : EtOH, as required. The column size (height and diameter), amount of silica gel, solvent polarity variation (gradient) step, solvent volume per step and fraction volume were adjusted for each case (compound and reaction), taking into account mainly the amount of sample, its polarity and the presence of impurities, especially if their R_f value in the corresponding TLC analysis was close to that of the product. The chromatographic variables were adjusted aiming to elute the product approximately after the 15th tube, when the solvent polarity was equivalent to that yielding a $R_f \approx 0.50$ in the TLC plate. The chromatographies were continued after elution of the expected product, in order to ensure recovery of all the relevant compounds.

Equipment

The melting points (uncorrected) were measured on an Ernst Leitz Wetzlar model 350 hot-stage microscope. The FT-IR spectra were recorded on a Shimadzu Prestige 21 spectrophotometer, as solid dispersions in KBr disks (solid samples). The NMR spectra were recorded in CDCl_3 unless informed otherwise with a Bruker Avance 300 NMR spectrometer (300.13 MHz for ^1H , 75.48 MHz for ^{13}C and 282.38 MHz for ^{19}F). In addition, some spectra were acquired with a Bruker Avance 400 MHz spectrometer (400.13 MHz for ^1H , 100.61 MHz for ^{13}C). The chemical shifts are reported in ppm on the δ scale, and TMS was used as the internal standard ($\delta = 0.0$ ppm); the residual solvent

peaks of CDCl_3 ($\delta_{\text{H}} = 7.26$ ppm, $\delta_{\text{C}} = 77.16$ ppm), $\text{DMSO}-d_6$ ($\delta_{\text{H}} = 2.50$ ppm, $\delta_{\text{C}} = 39.52$ ppm) and CD_3OD ($\delta_{\text{H}} = 3.31$ ppm, $\delta_{\text{C}} = 49.00$ ppm) as well as the signal of CFCl_3 for ($\delta_{\text{F}} = 0.00$ ppm), were used as internal reference. The magnitudes of the coupling constant (J) values are given in hertz. To complete the elucidation, NOE and 2D NMR experiments (HSQC and HMBC) were employed. HRMS was obtained with a Bruker MicroTOF-Q II instrument from UMyMFOR (Buenos Aires, Argentina). The microwave-assisted reactions were carried out in a CEM Discover microwave reactor. The UV-Vis spectrum of ampullosine (**4**) was acquired in an Agilent model 8453 spectrophotometer equipped with a diode-array detector, in the range 205–700 nm and matched quartz cells (10 mm optical path). The sample was dissolved in MeOH, and the spectrum was taken against a blank of the solvent.

Dimethyl 2,6-dihydroxyterephthalate (16). A mixture of 3,5-dihydroxybenzoic acid (**14**) (380 mg, 2.5 mmol) in 1,2-propanediol (700 μL) was heated at 100 °C in a pressure tube until the mixture was completely homogenized. After cooling, KHCO_3 (1.25 g, 12.5 mmol) was added, and the resulting suspension was purged with CO_2 , sealed and heated at 180 °C for 6 h. After cooling, water (50 mL) and concentrated HCl (50 mL) were added to the resulting whitish residue, the mixture was extracted with Et_2O (6 \times 50 mL). The combined extracts were dried over MgSO_4 and the solvent was evaporated *in vacuo* on a rotary evaporator, affording the product dicarboxylic acid product **15** (381 mg, 80%) as a pale yellow solid. Without further purification, this solid was dissolved in acetone (5 mL), KHCO_3 (500 mg, 5 mmol) was added and the mixture was stirred for 15 min at room temperature under an argon atmosphere. Then, Me_2SO_4 (470 μL , 5 mmol) was added dropwise, and the suspension was heated at reflux for 10 h. After completion of the reaction, the volatiles were evaporated, saturated NaHCO_3 (50 mL) was added to the residue and the products were extracted with EtOAc (3 \times 45 mL). The combined organic phases were dried over MgSO_4 , filtered, and concentrated under reduced pressure. The residue was purified by column chromatography (hexane:EtOAc, 1 : 0 \rightarrow 2 : 8 v/v) to afford the desired product **16** (394 mg, 70%, two steps), as a white solid, R_f : 0.53 (hexane : EtOAc, 6 : 4 v/v), mp: 146–148 °C (lit. 20a and 20d 146–149 °C); ^1H NMR δ : 9.67 (br s, $w_{1/2} = 10.2$, 2H, ArOH), 7.11 (s, 2H, H-3 and H-5), 4.11 (s, 3H, 1-CO₂Me) and 3.90 (s, 3H, 4-CO₂Me); ^{13}C NMR δ : 169.6 (1-CO₂Me), 165.8 (4-CO₂Me), 160.8 (C-2 and C-6), 137.3 (C-4), 109.2 (C-3 and C-5), 102.9 (C-1), 53.4 (1-CO₂Me) and 52.6 (4-CO₂Me).

Methyl 5-hydroxy-2,2-dimethyl-4H-benzo[d][1,3]dioxine-7-carboxylate (17).^{20a,20d} A mixture of NaBH_4 (195 mg, 5.16 mmol) in 0.05 M phosphate buffer, pH 7.5 (1.5 mL) was added dropwise to a stirred solution of **16** (390 mg, 1.72 mmol) in THF (9 mL) cooled to 0 °C in an ice-water bath. The resulting solution was stirred for 1.5 h, when the reaction was quenched with 10% citric acid solution (5 mL) at 0 °C. Then, brine (20 mL) was added and the organic products were extracted with EtOAc (3 \times 20 mL). The combined organic layers were dried over MgSO_4 and concentrated *in vacuo*. The crude residue was purified by column chromatography to give methyl 3,5-dihydroxy-4-(hydroxymethyl)benzoate (286 mg), as a whitish solid. The product was filtered through a short silica gel column and



without further purification, this solid was dissolved in 2,2-dimethoxypropane (5 mL), *p*-TsOH·H₂O (32 mg, 0.17 mmol) was added at 0 °C, the ice bath was removed, and the mixture was stirred for 5 h at room temperature. The reaction mixture was placed again in an ice bath and NaHCO₃ (20 mg) was added. After stirring for 5 min, the resulting solution was purified by silica gel column chromatography (hexane : EtOAc, 1 : 0 → 1 : 1 v/v), furnishing **17** (307 mg, 75%, two steps), as a white solid, *R*_f: 0.50 (hexane : EtOAc 7 : 3 v/v), mp: 141–142 °C; IR (KBr, ν): 3327, 2995, 2846, 1697, 1595, 1431, 1365, 1276, 1149, 1060, 950, 862, 763 and 642 cm⁻¹; ¹H NMR δ : 7.15 (d, *J* = 1.2, 1H, H-8), 7.11 (d, *J* = 1.2, 1H, H-6), 5.92 (s, 1H, ArOH), 4.86 (s, 2H, H-4), 3.89 (s, 3H, CO₂Me) and 1.55 (s, 6H, Me-2); ¹³C NMR δ : 167.2 (CO₂Me), 152.5 (C-8a), 152.1 (C-5), 130.0 (C-7), 112.7 (C-4a), 111.0 (C-6), 107.7 (C-8), 99.8 (C-2), 58.1 (C-4), 52.5 (CO₂Me) and 24.7 (Me-2); HRMS (ESI-TOF): found *m/z*: 261.0729; C₁₂H₁₄NaO₅ ([M + Na]⁺) requires *m/z*: 261.0733.

Methyl 2,2-dimethyl-5-((trifluoromethyl)sulfonyloxy)-4H-benzo[d][1,3]dioxine-7-carboxylate (18). A stirred suspension of phenol **27** (303 mg, 1.27 mmol) in anhydrous CH₂Cl₂ (4 mL) was cooled to 0 °C under argon and successively treated with DMAP (152 mg, 0.12 mmol), Et₃N (322 mg, 445 μ L, 3.18 mmol) and PhNTf₂ (635 mg, 1.77 mmol). The reaction mixture was left to acquire room temperature and further stirred overnight. Then, the reaction was diluted with CH₂Cl₂ (50 mL), the organic phase was washed with saturated NaHCO₃ (2 × 50 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was chromatographed (hexane : CH₂Cl₂, 1 : 0 → 3 : 2 v/v), furnishing **18** (451 mg, 96%), as a white solid, *R*_f: 0.20 (hexane : CH₂Cl₂, 7 : 3 v/v), mp: 63–64 °C; IR (KBr, ν): 3086, 2997, 2962, 1724, 1629, 1583, 1423, 1328, 1253, 1138, 1029, 925, 829, 767 and 638 cm⁻¹; ¹H NMR δ : 7.55 (d, *J* = 1.4, 1H, H-6), 7.51 (d, *J* = 1.4, 1H, H-8), 4.93 (s, 2H, H-4), 3.92 (s, 3H, CO₂Me) and 1.56 (s, 6H, Me-2); ¹³C NMR δ : 165.1 (CO₂Me), 153.1 (C-8a), 145.1 (C-5), 131.3 (C-7), 118.7 (q, *J* = 320, CF₃), 118.6 (C-6), 118.0 (C-4a), 113.7 (C-8), 100.8 (C-2), 57.4 (C-4), 52.8 (CO₂Me) and 24.6 (Me-2); ¹⁹F NMR δ : -73.3 (s, CF₃SO₂Ar); HRMS (ESI-TOF): found *m/z*: 393.0226; C₁₃H₁₃F₃NaO₇S ([M + Na]⁺) requires *m/z*: 393.0226.

Methyl 5-allyl-2,2-dimethyl-4H-benzo[d][1,3]dioxine-7-carboxylate (19). A mixture of triflate **28** (29 mg, 0.08 mmol), LiCl (3 mg, 0.07 mmol), Ph₃P (10 mg, 50 mol%), and PdCl₂(PPh₃)₂ (6 mg, 10 mol%), was placed in a tube, capped with a septum, purged with dry argon for 5 minutes and then diluted with anhydrous diglyme (0.5 mL). After stirring for 5 min, the resulting mixture was treated with Bu₃SnCH₂CH=CH₂ (31 μ L, 0.10 mmol), the reaction tube was sealed and the mixture was heated at 125 °C for 15 h. After cooling to room temperature, the reaction mixture was diluted with Et₂O (15 mL), a saturated solution of KF (10 mL) was added and the mixture was stirred for 20 min in order to quench the organotin-derivatives. The aqueous phase was separated and the organic layer was washed with brine (3 × 8 mL). The organic solvent was filtered through a short pad of Florisil and Celite (1 : 1 w/w) and the filtrate was dried over MgSO₄ prior to concentration under reduced pressure. Column chromatography (hexane : EtOAc, 1 : 0 → 8 : 2 v/v) of the residue gave the corresponding allyl substituted

compound **19** as a 95 : 5 mixture with the related β -propenyl derivative **E-20** (14.6 mg, 70%), as a colourless solid (hexane : EtOAc, 9 : 1 v/v); ¹H NMR δ : 7.43 (d, *J* = 1.4, 1H, H-6), 7.39 (d, *J* = 1.4, 1H, H-8), 5.90 (ddt, *J* = 17.0, 10.1 and 6.2, 1H, ArCH₂CH=CH₂), 5.10 (ddt, *J* = 10.1, 1.4 and 1.4, 1H, ArCH₂CH=CH₂_{cis}), 5.00 (ddt, *J* = 17.0, 1.6 and 1.6, 1H, ArCH₂CH=CH₂_{trans}), 4.83 (s, 2H, H-4), 3.88 (s, 3H, CO₂Me), 3.25 (dt, *J* = 6.2 and 1.6, 2H, ArCH₂CH=CH₂) and 1.52 (s, 6H, Me-2); ¹³C NMR δ : 166.9 (CO₂Me), 151.5 (C-8a), 136.2 (C-5), 135.1 (ArCH₂CH=CH₂), 130.0 (C-7), 123.2 (C-4a), 122.3 (C-6), 116.8 (ArCH₂CH=CH₂), 116.7 (C-8), 99.3 (C-2), 59.7 (C-4), 52.2 (CO₂Me), 36.2 (ArCH₂CH=CH₂) and 24.7 (Me-2).

Methyl (E)-2,2-dimethyl-5-(propen-1-yl)-4H-benzo[d][1,3]dioxine-7-carboxylate (20). A mixture of the triflate **18** (100 mg, 0.27 mmol), potassium *trans*-1-propenyltrifluoroborate (68 mg, 0.45 mmol), Cs₂CO₃ (264 mg, 0.81 mmol), LiCl (5 mg, 0.12 mmol), DavePhos (10 mg, 10 mol%), and PdCl₂(PPh₃)₂ (9 mg, 5 mol%), was placed in a pressure tube capped with septum. The solids were treated with THF (2 mL) and degassed water (0.5 mL) with stirring and then were purged with dry argon for 5 minutes. The tube was sealed and the reaction mixture was heated at 80 °C for 24 h in an oil bath. After cooling to room temperature, the reaction mixture was diluted with water (6 mL) and extracted with CH₂Cl₂ (3 × 10 mL). The combined organic phases were dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by column chromatography (hexane : EtOAc, 1 : 0 → 8 : 2 v/v), to afford the desired product **20** (67 mg, 95%), as a white solid, *R*_f: 0.47 (hexane : EtOAc, 9 : 1 v/v), mp: 70–71 °C; IR (KBr, ν): 3030, 2999, 2850, 1722, 1577, 1431, 1338, 1224, 1141, 1012, 904, 819 and 767 cm⁻¹; ¹H NMR δ : 7.67 (d, *J* = 1.4, 1H, H-6), 7.36 (d, *J* = 1.4, 1H, H-8), 6.20–6.33 (m, 2H, ArCH=CHMe and ArCH=CHMe), 4.88 (s, 2H, H-4), 3.89 (s, 3H, CO₂Me), 1.91 (d, *J* = 4.7, 3H, ArCH=CHMe) and 1.53 (s, 6H, Me-2); ¹³C NMR δ : 167.0 (CO₂Me), 151.4 (C-8a), 135.2 (C-5), 130.2 (ArCH=CHMe), 130.0 (C-7), 125.6 (ArCH=CHMe), 121.2 (C-4a), 118.8 (C-8), 116.6 (C-6), 99.2 (C-2), 60.2 (C-4), 52.2 (CO₂Me), 24.7 (Me-2) and 18.9 (ArCH=CHMe); HRMS (ESI-TOF): found *m/z*: 285.1098; C₁₅H₁₈NaO₄ ([M + Na]⁺) requires *m/z*: 285.1097.

Methyl (E)-3-hydroxy-4-(hydroxymethyl)-5-(propen-1-yl)benzoate (24)

Method A. A stirred solution of the acetone **20** (187 mg, 0.71 mmol) and *p*-TsOH·H₂O (26 mg, 0.14 mmol) in a THF : H₂O mixture (1 : 1 v/v, 14 mL) was heated at 60 °C for 48 h. The reaction was cooled to 0 °C, quenched with saturated aqueous NaHCO₃ (30 mL), and the resulting mixture was extracted with EtOAc (3 × 50 mL). The combined organic layers were dried over MgSO₄, and concentrated *in vacuo*. The crude residue was purified by column chromatography (hexane : EtOAc, 1 : 0 → 4 : 6 v/v), to give **24** (154 mg, 98%), as a white solid, *R*_f: 0.25 (hexane : EtOAc, 7 : 3 v/v), mp: 95–97 °C; IR (KBr, ν): 3502, 3385, 3169, 2953, 2850, 1689, 1581, 1427, 1340, 1247, 1099, 1001, 964, 883 and 761 cm⁻¹; ¹H NMR δ : 8.12 (br s, 1H, W_{1/2} = 15.3 Hz, ArOH), 7.54 (d, *J* = 1.6, 1H, H-6), 7.35 (d, *J* = 1.6, 1H, H-2), 6.48 (dq, *J* = 15.5 and 1.7, 1H, ArCH=CHMe), 6.08 (dq, *J* = 15.5 and 6.6, 1H, ArCH=CHMe), 4.96 (s, 2H, ArCH₂OH), 3.88 (s, 3H, CO₂Me), 3.04 (br s, 1H, W_{1/2} = 21.8 Hz, ArCH₂OH) and 1.87 (dd, *J*



= 6.6 and 1.7, 3H, ArCH=CHMe); ^{13}C NMR δ : 167.3 (CO₂Me), 156.4 (C-3), 138.2 (C-5), 130.7 (ArCH=CHMe), 130.3 (C-4), 127.2 (ArCH=CHMe), 126.5 (C-1), 120.0 (C-6), 115.8 (C-2), 60.1 (ArCH₂OH), 52.4 (CO₂Me) and 18.8 (ArCH=CHMe); HRMS (ESI-TOF): found m/z : 245.0782; C₁₂H₁₄NaO₄ ([M + Na]⁺) requires m/z : 245.0784.

Method B. A stirred solution of **20** (98 mg, 0.37 mmol) in THF (4.5 mL) was treated with 1 M HCl (3 mL) and the system was heated at 45 °C for 18 h. The reaction was cooled to 0 °C and quenched with saturated NaHCO₃ (10 mL); the organic products were extracted with EtOAc (3 × 15 mL). The combined organic layers were dried over MgSO₄, and concentrated *in vacuo*. The crude residue was purified by column chromatography to give **24** (74 mg, 90%). The spectroscopic data of the product were in full agreement with those of the product synthesized according to Method A.

Methyl (E)-4-formyl-3-hydroxy-5-(propen-1-yl)benzoate (25).

A mixture of the benzylic alcohol **24** (50 mg, 0.22 mmol) and freshly activated MnO₂ (287 mg, 3.30 mmol) was slurred in anhydrous EtOAc (3 mL) and the system was heated to reflux for 3 h. After cooling, the reaction mixture was filtered through a Celite pad, and the filtrate was concentrated *in vacuo*. The resulting yellow residue was purified by column chromatography (hexane : EtOAc, 1 : 0 → 7 : 3 v/v), to afford **25** (27 mg, 55%), as a yellow solid, R_f : 0.50 (hexane : EtOAc, 8 : 2 v/v), mp: 140–141 °C; IR (KBr, ν): 3043, 2990, 2848, 1716, 1647, 1558, 1417, 1348, 1247, 1174, 1097, 999, 887 and 766 cm⁻¹; ^1H NMR δ : 11.74 (s, 1H, ArOH), 10.36 (s, 1H, CHO), 7.53 (d, $J = 1.4$, 1H, H-6), 7.46 (d, $J = 1.4$, 1H, H-2), 6.88 (dq, $J = 15.5$ and 1.7, 1H, ArCH=CHMe), 6.21 (dq, $J = 15.5$ and 6.6, 1H, ArCH=CHMe), 3.93 (s, 3H, CO₂Me) and 1.97 (dd, $J = 6.6$ and 1.7, 3H, ArCH=CHMe); ^{13}C NMR δ : 195.8 (CHO), 165.9 (CO₂Me), 162.5 (C-3), 143.8 (C-5), 137.3 (C-1), 135.0 (ArCH=CHMe), 125.2 (ArCH=CHMe), 119.5 (C-4), 119.4 (C-6), 117.4 (C-2), 52.7 (CO₂Me) and 19.1 (ArCH=CHMe); HRMS (ESI-TOF): found m/z : 243.0633; C₁₂H₁₂NaO₄ ([M + Na]⁺) requires m/z : 243.0628.

Methyl (E)-4-formyl-3-((methylsulfonyl)oxy)-5-(prop-1-en-1-yl) benzoate (28). A stirred solution of **25** (35 mg, 0.16 mmol) in CH₂Cl₂ (1 mL) was cooled in an ice-water bath and successively treated with Et₃N (34 μL , 0.24 mmol) and MeSO₂Cl (15 μL , 0.19 mmol). The reaction mixture was stirred at room temperature for 16 h. Then, brine (5 mL) was added to the reaction mixture and extracted with EtOAc (3 × 20 mL). The combined organic phases were washed successively with 0.2 M HCl (10 mL), saturated NaHCO₃ (10 mL) and then dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the residue was chromatographed (hexane : EtOAc, 1 : 0 → 2 : 8 v/v) to afford **28** (27 mg, 55%; 77% br s m), as an unstable colorless oil R_f : 0.56 (hexane : EtOAc, 1 : 1 v/v); ^1H NMR δ : 10.43 (s, CHO), 8.13 (d, $J = 1.4$, 1H, H-2), 7.88 (d, $J = 1.4$, 1H, H-6), 7.06 (dd, $J = 15.6$ and 1.7, 1H, ArCH=CHMe), 6.30 (dq, $J = 15.6$ and 6.6, 1H, ArCH=CHMe), 3.96 (s, 3H, CO₂Me), 3.32 (s, 3H, MeSO₃⁻) and 1.93 (dd, $J = 6.6$ and 1.7, 3H, ArCH=CHMe); ^{13}C NMR δ : 189.7 (CHO), 164.9 (CO₂Me), 149.3 (C-3), 142.5 (C-5), 134.8 (C-1), 134.4 (ArCH=CHMe), 129.0 (ArCH=CHMe), 127.6 (C-4), 126.6 (C-6), 122.1 (C-2), 52.8 (CO₂Me), 38.6 (MeSO₃⁻) and 18.9 (ArCH=CHMe).

Methyl (E)-4-formyl-3-methoxy-5-(propen-1-yl)benzoate (31).

A mixture of the methyl 3-hydroxybenzoate **25** (53 mg, 0.24 mmol) and K₂CO₃ (46 mg, 0.33 mmol) in anhydrous DMF (1 mL) was stirred for 15 min at 0 °C. The system was treated with iodomethane (20 μL , 0.31 mmol) and the resulting reaction mixture was stirred at room temperature for 1.5 h. Then, the mixture was poured into cold water (10 mL), and the products were extracted EtOAc (3 × 15 mL). The combined organic phases were washed with brine (25 mL), dried over MgSO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography (hexane : EtOAc, 1 : 0 → 7 : 3 v/v) to afford **31** (48 mg, 85%), as a yellowish-white solid R_f : 0.52 (hexane : EtOAc, 8 : 2, v/v), mp: 72–73 °C; IR (KBr, ν): 3086, 2954, 2873, 2785, 1718, 1681, 1566, 1404, 1330, 1238, 1103, 1010, 964, 819 and 765 cm⁻¹; ^1H NMR δ : 10.61 (d, $J = 0.5$, CHO), 7.77 (ddd, $J = 1.4$, 0.7 and 0.7, 1H, H-6), 7.48 (d, $J = 1.4$, 1H, H-2), 7.20 (dq, $J = 15.5$ and 1.7, 1H, ArCH=CHMe), 6.30 (dq, $J = 15.5$ and 6.6, 1H, ArCH=CHMe), 3.95 (s, 3H, ArOMe), 3.95 (s, 3H, CO₂Me) and 1.93 (dd, $J = 6.6$ and 1.7, 3H, ArCH=CHMe); ^{13}C NMR δ : 192.3 (CHO), 166.3 (CO₂Me), 162.3 (C-3), 141.2 (C-5), 134.8 (C-1), 131.7 (ArCH=CHMe), 128.5 (ArCH=CHMe), 124.6 (C-4), 120.8 (C-6), 110.1 (C-2), 56.3 (ArOMe), 52.7 (CO₂Me) and 18.9 (ArCH=CHMe); HRMS (ESI-TOF): found m/z : 235.0960; C₁₃H₁₅O₄ ([M + H]⁺) requires m/z : 235.0965.

Methyl 8-methoxy-3-methylisoquinoline-6-carboxylate (per-methyl ampullosine, 5)⁹ and methyl 8-methoxy-3-methyl-3,4-dihydroisoquinoline-6-carboxylate (33)

Method A. Methyl 4-formylbenzoate **31** (22 mg, 0.09 mmol), glacial AcOH (5 μL , 0.09 mmol) and 1,1-dimethylhydrazine (7 μL , 0.09 mmol) were added to a microwave tube and diluted with PhCF₃ (1 mL). The mixture was purged with argon and stirred at room temperature until TLC analysis indicated complete consumption of the starting aldehyde (~1.5 h). Then, the vessel was capped and irradiated in the microwave reactor (160 °C, *ca.* 250 W, 1 h). After cooling to room temperature, the solvent was recovered by careful distillation under atmospheric pressure, and the residue was purified by column chromatography (hexane/EtOAc, 1 : 0 → 0 : 1 v/v), to afford **5** (13.3 mg, 64%), as a yellowish solid, R_f : 0.60 (hexane/EtOAc, 6 : 4 v/v), mp: 126–129 °C; IR (KBr, ν): 3068, 2947, 2848, 1716, 1633, 1570, 1433, 1354, 1238, 1122, 1035, 927, 825 and 762 cm⁻¹; ^1H NMR (300 MHz, CDCl₃) δ : 9.56 (s, 1H, H-1), 8.03 (s, 1H, H-5), 7.50 (s, 1H, H-4), 7.37 (d, $J = 1.2$, 1H, H-7), 4.07 (s, 3H, ArOMe), 3.99 (s, 3H, CO₂Me) and 2.71 (s, 3H, Me-3); ^{13}C NMR (75 MHz, CDCl₃) δ : 166.9 (CO₂Me), 156.9 (C-8), 153.5 (C-3), 147.4 (C-1), 137.2 (C-4a), 132.2 (C-6), 121.2 (C-5), 120.6 (C-8a), 119.1 (C-4), 103.6 (C-7), 56.0 (ArOMe), 52.7 (CO₂Me) and 24.3 (Me-3); ^1H NMR (400 MHz, CD₃OD) δ : 9.31 (s, 1H, H-1), 7.94 (s, 1H, H-5), 7.58 (s, 1H, H-4), 7.34 (s, 1H, H-7), 4.05 (s, 3H, ArOMe), 3.97 (s, 3H, CO₂Me) and 2.63 (s, 3H, Me-3); ^{13}C NMR (100 MHz, CD₃OD) δ : 167.8 (CO₂Me), 158.0 (C-8), 153.7 (C-3), 147.5 (C-1), 138.6 (C-4a), 134.0 (C-6), 121.8 (C-5), 121.5 (C-8a), 121.0 (C-4), 104.7 (C-7), 56.5 (ArOMe), 53.1 (CO₂Me) and 23.5 (Me-3); HRMS (ESI-TOF): found m/z : 254.0798; C₁₃H₁₃NNaO₃ ([M + Na]⁺) requires m/z : 254.0788. The ^1H and ^{13}C NMR spectral data of compound **5** were in agreement with the literature.⁹



Increasing solvent polarity afforded **33** (5.5 mg, 26%), as an amber solid, mp: 99–101 °C; IR (KBr, ν): 3005, 2951, 2837, 1724, 1618, 1570, 1446, 1319, 1222, 1112, 1001, 968, 864 and 767 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 8.72 (d, $J = 2.6$, 1H, H-1), 7.45 (s, 1H, H-5), 7.42 (s, 1H, H-7), 3.93 (s, 6H, ArOMe and CO_2Me), 3.60–3.70 (m, 1H, H-3), 2.78 (dd, $J = 16.2$ and 5.6, 1H, H_{B-4}), 2.49 (dd, $J = 16.2$ and 12.1, 1H, H_{B-4}) and 1.39 (d, $J = 6.9$, 3H, Me-3); ^{13}C NMR (75 MHz, CDCl_3) δ : 166.6 (CO_2Me), 156.9 (C-8), 154.1 (C-1), 138.3 (C-4a), 133.1 (C-6), 121.2 (C-7), 120.2 (C-8a), 110.6 (C-5), 55.9 (ArOMe), 52.5 (CO_2Me), 52.0 (C-3), 32.5 (C-4) and 21.6 (Me-3); HRMS (ESI-TOF): found m/z : 256.0949; $\text{C}_{13}\text{H}_{15}\text{NNaO}_3$ ($[\text{M} + \text{Na}]^+$) requires m/z : 256.0944.

Method B. A mixture of *O*-methylhydroxylamine hydrochloride (16 mg, 0.19 mmol) and NaOAc (17 mg, 0.21 mmol) in absolute EtOH (1.5 mL) was stirred at room temperature for 30 min under argon, when the resulting suspension was allowed to settle. An aliquot of the solution (1 mL) was transferred to a microwave tube containing the methyl 4-formylbenzoate **31** (23 mg, 0.1 mmol) and the resulting mixture was stirred at room temperature for 1 h under argon. Then, the solvent was removed under reduced pressure and the residue was dissolved in PhCF_3 (1 mL). The tube was purged with Argon and the mixture was irradiated in the microwave reactor (180 °C, ca. 250 W, 45 min). After cooling to room temperature, the solvent was recovered by careful distillation under atmospheric pressure, and the residue was purified through a short column chromatography, to afford a separable mixture of **5** (11.5 mg, 50%) and **33** (3.5 mg, 15%). Their spectroscopic data were in full agreement with those of the corresponding products, synthesized employing Method A.

Ampullosine (4). Iodine (91 mg, 0.36 mmol) was added in one portion to a stirred mixture of permethylampullosine **5** (20 mg, 0.08 mmol), small pieces of aluminium foil (12 mg, 0.44 mmol) and dry DMSO (20 μL , 0.28 mmol) in acetonitrile (1 mL). The mixture was further stirred at 80 °C for 20 h and monitored by TLC until the completion of conversion. After cooling to room temperature, the reaction was acidified to pH 3 with solution of citric acid (0.2 M, 5 mL) and was treated with Na_2SO_3 (15 mg) before extraction with EtOAc (5 \times 10 mL). The organic phases were combined, and dried over MgSO_4 . After removal of organic solvents under reduced pressure, the residue was purified by column chromatography (EtOAc : EtOH, 1 : 0 \rightarrow 2 : 8 v/v), furnishing **4** (3.7 mg, 23%), as a yellow solid, R_f : 0.40 (EtOAc : EtOH, 1 : 1 v/v). UV-Vis (MeOH): 227 nm ($\log \epsilon = 4.3$) and 356 nm ($\log \epsilon = 3.5$). ^1H NMR (300 MHz, $\text{DMSO}-d_6$) δ : 10.88 (bs, 1H, OH), 9.37 (s, 1H, H-1), 7.85 (s, 1H, H-5), 7.64 (s, 1H, H-4), 7.45 (s, 1H, H-7), 3.35 (s, 1H, OH) and 2.59 (s, 3H, Me-3); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ : 168.3 (6- CO_2H), 154.8 (C-8), 152.2 (C-3), 147.2 (C-1), 137.3 (2C, C-4a and C-6), 119.5 (C-8a), 119.1 (C-4), 118.3 (C-5), 109.2 (C-7) and 24.2 (3-Me). HRMS (ESI-TOF): found m/z : 204.0659; $\text{C}_{11}\text{H}_{10}\text{NO}_3$ ($[\text{M} + \text{H}]^+$) requires m/z : 204.0655; found m/z : 226.0471; $\text{C}_{11}\text{H}_9\text{NNaO}_3$ ($[\text{M} + \text{Na}]^+$) requires m/z : 226.0475. The UV-Vis, ^1H and ^{13}C NMR spectral data were in agreement with the literature.⁹

Conclusion

We have developed a straightforward and convenient approach toward the first total synthesis of ampullosine, a structurally unique 6-carboxy-3-methylisoquinoline. The heterocycle is the first and only alkaloid isolated from the saproparasitic fungus *Sepedonium ampullosporum*. The synthesis of *O*-methyl ampullosine methyl ester, from a common intermediate, is also reported.

The synthesis of the natural product used 3,5-dihydroxybenzoic acid as starting material and its key steps included a Kolbe–Schmitt type carboxylation for introduction of C1 of the heterocyclic ring, a Molander cross-coupling with potassium propenyl trifluoroborate to install the remaining three carbon atoms, a carbonyl hydrazonation to add the heterocyclic nitrogen and a final 6π -azaelectrocyclization to build the isoquinoline system.

The total synthesis of ampullosine was achieved in 12 steps and 3.2% overall yield, whereas that of its permethyl derivative was accomplished in 14% overall yield, after 11 steps. Hydrolytic demethylation of *O*-methyl ampullosine methyl ester also afforded ampullosine. To the best of our knowledge, this is the first report of an electrocyclization reaction toward isoquinolines bearing a 6-carboxylic acid derivative.

Conflicts of interest

There are no conflicts of interest to declare.

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Notes and references

- 1 T. Sahr, H. Ammer, H. Besl and M. Fischer, *Mycologia*, 1999, **91**, 935–943.
- 2 (a) T. Petch, *Trans. Br. Mycol. Soc.*, 1938, **21**, 243–305; (b) E. Pieschel, *Mykol. Mitteilungsblatt*, 1962, **6**, 62–68.
- 3 (a) S. C. Damon, *Mycologia*, 1952, **44**, 86–96; (b) M. S. Foster, G. F. Bills and G. M. Mueller, *Biodiversity of Fungi: Inventory and Monitoring Methods*, Academic Press, New York, USA, 2004; (c) H. Besl, A. Hagn, A. Jobst and U. Lange, *Z. Mykol.*, 1998, **64**, 45–52.
- 4 (a) T. Neuhof, A. Berg, H. Besl, T. Schwecke, R. Dieckmann and H. von Döhren, *Chem. Biodiversity*, 2007, **4**, 1103–1115; (b) A. Otto, A. Laub, L. Wendt, A. Porzel, J. Schmidt, G. Palfner, J. Becerra, D. Krüger, M. Stadler,



- L. Wessjohann, B. Westermann and N. Arnold, *J. Nat. Prod.*, 2016, **79**, 929–938; (c) M. Kronen, P. Kleinwächter, B. Schlegel, A. Härtl and U. Gräfe, *J. Antibiot.*, 2001, **54**, 175–178; (d) M. I. Mitova, A. Murphy, G. Lang, J. W. Blunt, A. L. J. Cole, G. Ellis and M. H. G. Munro, *J. Nat. Prod.*, 2008, **71**, 1600–1603; (e) T. Degenkolb, J. Kirschbaum and H. Brückner, *Chem. Biodiversity*, 2007, **4**, 1052–1067.
- 5 M. I. Mitova, B. G. Stuart, G. H. Cao, J. W. Blunt, A. L. J. Cole and M. H. G. Munro, *J. Nat. Prod.*, 2006, **69**, 1481–1484.
- 6 (a) A. Closse and D. Hauser, *Helv. Chim. Acta*, 1973, **56**, 2694–2698; (b) H. Haraguchi, M. Taniguchi, K. Motoba, K. Shibata, S. Oi and K. Hashimoto, *Agric. Biol. Chem.*, 1990, **54**, 2167–2168.
- 7 S. Shibata, J. Shoji, A. Ohta and M. Watanabe, *Chem. Pharm. Bull.*, 1957, **5**, 380–382.
- 8 (a) P. V. Divekar, H. Raistrick, T. A. Dobson and L. C. Vining, *Can. J. Chem.*, 1965, **43**, 1835–1848; (b) K. Nagao, N. Yoshida, K. Iwai, T. Sakai, M. Tanaka and T. Miyahara, *Environ. Sci.*, 2006, **13**, 251–256; (c) J. L. C. Wright, A. G. McInnes, D. G. Smith and L. C. Vining, *Can. J. Chem.*, 1970, **48**, 2702–2708.
- 9 D. N. Quang, J. Schmidt, A. Porzel, L. Wessjohann, M. Haid and N. Arnold, *Nat. Prod. Commun.*, 2010, **5**, 869–872.
- 10 (a) A. McDonald, WO Pat., 2015103317, 2015; (b) D. A. Claremon, L. W. Dillard, Y. Fan, L. Jia, S. B. Singh, C. M. Tice, Z. Xu, J. Yuan and L. Zhuang, WO Pat., 201787608, 2017.
- 11 M. Shankar, K. Ghosh, K. Mukherjee, R. K. Rit and A. K. Sahoo, *Org. Lett.*, 2018, **20**, 5144–5148.
- 12 (a) J. Wang, S. Zha, K. Chen, F. Zhang and J. Zhu, *Org. Biomol. Chem.*, 2016, **14**, 4848–4852; (b) Z. Shi, D. C. Koester, M. Bouladakakis–Arapinis and F. Glorius, *J. Am. Chem. Soc.*, 2013, **135**, 12204–12207; (c) M. Tian, G. Zheng, X. Fan and X. Li, *J. Org. Chem.*, 2018, **83**, 6477–6488.
- 13 (a) J. L. Pergomet, A. B. J. Bracca and T. S. Kaufman, *Org. Biomol. Chem.*, 2017, **15**, 7040–7049; (b) S. O. Simonetti, E. L. Larghi and T. S. Kaufman, *Org. Biomol. Chem.*, 2016, **14**, 2625–2636; (c) M. V. Méndez, D. A. Heredia, E. L. Larghi, A. B. J. Bracca and T. S. Kaufman, *RSC Adv.*, 2017, **7**, 28298–28307; (d) J. L. Pergomet, E. L. Larghi, T. S. Kaufman and A. B. J. Bracca, *RSC Adv.*, 2017, **7**, 5242–5250; (e) D. F. Vargas, E. L. Larghi and T. S. Kaufman, *Synthesis*, 2019, **51**, 2030–2038.
- 14 (a) D. A. Heredia, E. L. Larghi and T. S. Kaufman, *Eur. J. Org. Chem.*, 2016, 1397–1404; (b) C. C. Silveira, E. L. Larghi, S. R. Mendes, A. B. J. Bracca, F. Rinaldi and T. S. Kaufman, *Eur. J. Org. Chem.*, 2009, 4637–4645; (c) D. F. Vargas, E. L. Larghi and T. S. Kaufman, *Nat. Prod. Rep.*, 2019, **36**, 354–401; (d) I. Cortés, C. M. Borini Etichetti, J. E. Girardini, A. B. J. Bracca and T. S. Kaufman, *Synthesis*, 2019, **51**, 433–440.
- 15 (a) C. D. Campbell, R. L. Greenaway, O. T. Holton, P. R. Walker, H. A. Chapman, C. A. Russell, G. Carr, A. L. Thomson and E. A. Anderson, *Chem.–Eur. J.*, 2015, **21**, 12627–12639; (b) A. Barbero, P. Cuadrado, I. Fleming, A. M. González and F. J. Pulido, *J. Chem. Soc., Chem. Commun.*, 1992, 351–353; (c) H. Tanaka, Y. Tokumaru and S. Torii, *Synlett*, 1999, 774–776.
- 16 I. Bonneau-Gubelmann, M. Michel, B. Besson, S. Ratton and J.-R. Desmurs, Carboxylation of Hydroxy Aromatic Compounds. In: *Industrial Chemistry Library*, Elsevier, Amsterdam, The Netherlands, 1996, vol. 8, pp. 116–128.
- 17 (a) J. Gràcia, M. A. Buil, J. Castro, P. Eichhorn, M. Ferrer, A. Gavalda, B. Hernández, V. Segarra, M. D. Lehner, I. Moreno, L. Pagès, R. S. Roberts, J. Serrat, S. Sevilla, J. Taltavull, M. Andrés, J. Cabedo, D. Vilella, E. Calama, C. Carcasona and M. Miralpeix, *J. Med. Chem.*, 2016, **59**, 10479–10497; (b) T. Inghardt, A. Johansson and A. Svensson, *US Pat.*, 200419033, 2004; (c) A.-C. Roth-Rosendahl and E. Svernhage, WO Pat., 2003101956, 2003; (d) R. Dominique, R. A. Goodnow Jr, A. Kowalczyk, Q. Qiao, A. Sidduri and J. W. Tilley, *US Pat.*, 2010240678, 2010; (e) J. Gracia Ferrer, M. A. Buil Albero, I. M. Moreno Mollo, L. M. Pages Santacana, R. S. Roberts, S. Sevilla and J. Taltavull Moll, EP Pat., 20102196465, 2010; (f) R. E. Counsell, T. D. Smith, V. V. Ranade, O. P. D. Noronha and P. Desai, *J. Med. Chem.*, 1973, **16**, 684–687.
- 18 M. B. Smith, *March's Advanced Organic Chemistry: Reactions, Mechanisms, and Structure*, Wiley, New York, USA, 7th edn, 2013.
- 19 (a) W. S. Leal, Y. Kuwahara and T. Suzuki, *Agric. Biol. Chem.*, 1990, **54**, 2593–2597; (b) N. Ueberschaar, Z. Xu, K. Scherlach, M. Metsä-Ketelä, T. Bretschneider, H.-M. Dahse, H. Görls and C. Hertweck, *J. Am. Chem. Soc.*, 2013, **135**, 17408–17416; (c) M. Sato, Y. Kuwahara, S. Matsuyama and T. Suzuki, *Biosci., Biotechnol., Biochem.*, 1993, **57**, 1299–1301.
- 20 (a) K. Watanabe, J. Sakurai, H. Abe and T. Katoh, *Chem. Commun.*, 2010, **46**, 4055–4057; (b) C. M. Harris, J. J. Kibby, J. R. Fehlner, A. B. Raabe, T. A. Barber and T. M. Harris, *J. Am. Chem. Soc.*, 1979, **101**, 437–445; (c) X. Wu, J. Zhou and B. B. Snider, *Angew. Chem., Int. Ed.*, 2009, **48**, 1283–1286; (d) J. Sakurai, T. Kikuchi, O. Takahashi, K. Watanabe and T. Katoh, *Eur. J. Org. Chem.*, 2011, 2948–2957.
- 21 Z. Markovic, S. Markovic, N. Manojlovic and J. Predojevic–Simovic, *J. Chem. Inf. Model.*, 2007, **47**, 1520–1525.
- 22 (a) Y.-S. Yao and S.-J. Yao, *J. Org. Chem.*, 2008, **73**, 5221–5225; (b) H. Morita, Y. Tomizawa, J. Deguchi, T. Ishikawa, H. Arai, K. Zaima, T. Hosoya, Y. Hirasawa, T. Matsumoto, K. Kamata, W. Ekasari, A. Widyawaruyanti, T. S. Wahyuni, N. C. Zaini and T. Honda, *Bioorg. Med. Chem.*, 2009, **17**, 8234–8240; (c) S. O. Simonetti, E. L. Larghi, A. B. J. Bracca and T. S. Kaufman, *Org. Biomol. Chem.*, 2012, **10**, 4124–4134.
- 23 (a) G. A. Molander and N. Ellis, *Acc. Chem. Res.*, 2007, **40**, 275–286; (b) H. Doucet, *Eur. J. Org. Chem.*, 2008, 2013–2030; (c) G. A. Molander and M. Rodríguez Rivero, *Org. Lett.*, 2002, **4**, 107–109.
- 24 (a) E. Alacid and C. Nájera, *J. Org. Chem.*, 2009, **74**, 8191–8195; (b) G. A. Molander, T. Barcellos and K. M. Traister, *Org. Lett.*, 2013, **15**, 3342–3345.
- 25 (a) Y. Duan, J. Zhang, J. Yang and M. Deng, *Chin. J. Chem.*, 2009, **27**, 179–182; (b) Z.-Y. Peng, J.-P. Wang, J. Cheng, X.-M. Xie and Z. Zang, *Tetrahedron*, 2010, **66**, 8238–8241.



- 26 D. S. Matteson, *J. Am. Chem. Soc.*, 1960, **82**, 4228–4233.
- 27 M. D. Hammers, L. Singh, L. A. Montoya, A. D. Moghaddam and M. D. Pluth, *Synlett*, 2016, **27**, 1349–1353.
- 28 (a) M. Kosugi and K. Fugami, *J. Organomet. Chem.*, 2002, **19**, 10–16; (b) V. Farina, V. Krishnamurthy and W. J. Scott, *The Stille Reaction*, Wiley, New York, USA, 2004.
- 29 C. Wei, R. Wang, C. Zhang, G. Xu, Y. Li, Q.-Z. Zhang, L.-Y. Li, L. Yi and Z. Xi, *Chem.-Asian J.*, 2016, **11**, 1376–1381.
- 30 (a) D. H. Dethe and G. Murhade, *Org. Lett.*, 2013, **15**, 429–431; (b) D. H. Dethe, B. D. Dherange, S. Ali and M. M. Parsutkar, *Org. Biomol. Chem.*, 2017, **15**, 65–68.
- 31 (a) K. C. Nicolaou, T. Montagnon, G. Vassilikogiannakis and C. J. N. Mathison, *J. Am. Chem. Soc.*, 2005, **127**, 8872–8888; (b) K. S. MacMillan, T. Nguyen, I. Hwang and D. L. Boger, *J. Am. Chem. Soc.*, 2009, **131**, 1187–1194; (c) M. Moreno, W. A. M. Elgaher, J. Herrmann, N. Schläger, M. M. Hamed, S. Baumann, R. Müller, R. W. Hartmann and A. Kirschning, *Synlett*, 2015, **26**, 1175–1178.
- 32 H. Cong, T. Yamato and Z. Tao, *New J. Chem.*, 2013, **37**, 3778–3783.
- 33 D. F. Vargas, E. L. Larghi and T. S. Kaufman, *Eur. J. Org. Chem.*, 2018, 5605–5614.
- 34 J. Kalia and R. T. Raines, *Angew. Chem., Int. Ed.*, 2008, **47**, 7523–7526.
- 35 (a) J. Tian, H. Yue, P. Yang and D. Sang, *ChemistrySelect*, 2019, **4**, 38–41; (b) D. Sang, X. Tu, J. Tian, Z. He and M. Yao, *ChemistrySelect*, 2018, **3**, 10103–10107; (c) D. Sang, C. Yi, Z. He, J. Wang, J. Tian, M. Yao and H. Shi, *Tetrahedron Lett.*, 2018, **59**, 1469–1472.
- 36 (a) H. Wagner, S. Bladt and V. Rickl, *Drug Analysis: A Thin Layer Chromatography Atlas*, Springer, Heidelberg, Germany, 2nd edn. 2009, p. 360; (b) W. C. Still, M. Kahn and A. Mitra, *J. Org. Chem.*, 1978, **43**, 2923–2925; (c) J. D. Fair and C. M. Kormos, *J. Chromatogr. A*, 2008, **1211**, 49–54; (d) D. F. Taber, *J. Org. Chem.*, 1982, **47**, 1352–1354.

