

Cite this: *RSC Adv.*, 2019, 9, 33678

Molecular imprinting on PtPd nanoflowers for selective recognition and determination of hydrogen peroxide and glucose†

Caini Fan,^a Junjia Liu,^b Haiying Zhao,^a Ling Li,^a Min Liu,^{*a} Jing Gao^b and Li Ma ^{*b}

PtPd nanoflowers (PtPd NFs) exhibit intrinsic peroxidase-like activity as nanozymes, but the nanozymes lack substrate specificity and have low catalytic activity. Herein, a molecularly imprinted nanogel on PtPd NFs was prepared by using 3,3',5,5'-tetramethylbenzidine (TMB) as the template through the aqueous precipitation polymerization method. After the TMB was washed out, many substrate binding pockets were retained in the PtPd NFs. Scanning electron microscopy (SEM), transmission electron microscopy (TEM) and powder X-ray diffraction (XRD) were employed to characterize the molecularly imprinted polymer (MIP) PtPd nanoflowers (T-MIP-PtPd NFs). The obtained T-MIP-PtPd NFs exhibited enhanced catalytic activity and specific recognition for TMB. Compared with PtPd NFs, T-MIP-PtPd NFs showed a linear range from 0.01–5000 μM and a detection limit of 0.005 μM toward the detection of H_2O_2 . Glucose can also be sensitively detected through cascade reaction by the T-MIP-PtPd NFs and glucose oxidase. Therefore, molecular imprinting on nanozymes technology shows promising application in biocatalysis and sensing fields.

Received 23rd July 2019

Accepted 2nd October 2019

DOI: 10.1039/c9ra05677g

rsc.li/rsc-advances

Introduction

Nanozymes, a nanomaterial with enzyme-like activity, are attracting more and more attention thanks to their outstanding properties.^{1–3} To date, various nanozymes, including metal-organic frameworks (MOFs),^{4–7} metal nanoparticles,⁸ oxidized metal nanoparticles^{9,10} and some other nanomaterials,¹¹ have been successfully prepared and used as enzyme mimics for catalytic applications.² As one of the enzyme mimics, Pt nanoparticles have been demonstrated to exhibit versatile catalytic activity like peroxidase, catalase and superoxide dismutase.^{2,12} Recent studies reported that the Pt-based bimetallic nanoparticles (*i.e.* PtPd nanodendrites) can exhibit peroxidase-like activity.^{13,14} For this kind of nanozyme, the creation of mesoporous nanostructures is considered to be an effective method to improve the enzyme mimetic activity.¹⁴ With the presence of a mesoporous structure, specific surface area and accessible pores of the Pt-based nanozymes will be increased, and the activity will be improved. Recently, a surfactant-directing method for synthesizing multimetallic spheres with ultra-large mesoporous was reported^{15,16} and the obtained

mesoporous multimetallic spheres exhibited enhanced electrochemical activity thanks to the high surface area and mesoporous structure. Despite the progress made in the synthesis of multi-metal nanospheres, their enzyme mimetic properties are rarely reported and the mechanism remains elusive. In addition, there are still disadvantages for these nanozymes, such as complicated synthesis process, low specificity and low catalytic activity.¹ Thus, the synthesis of the PtPd bimetallic nanomaterials with high peroxidase-mimic activity and substrate specificity are still challenges for further investigation.

Compared to natural enzymes, nanozymes are stable to denaturation, low in cost, highly resistant to high concentrations of substrates, flexible storage conditions and easy to manipulate catalytic activity. However, most nanozymes have no substrate specificity and can catalyze a lot of substrates leading to the lack of molecular recognition function. Molecularly imprinted polymers (MIPs) could create functional polymer materials with selective recognition properties.¹⁷ MIPs are usually prepared by bulk polymerization, photopolymerization, surface grafting polymerization and sol-gel mechanisms to generate imprinting specific cavities.^{18,19} Thus, combining MIPs with nanozymes provides a new class of nanozymes with improved selectivity and activity.^{20–22} For example, Zhang *et al.*²³ imprinted substrate binding pockets gels on three different nanozymes and the experimental results showed that the molecular imprinting on nanozymes remarkably increased the selectivity and catalytic activity.

Thus, in this study, PtPd nanoflowers (PtPd NFs) were synthesized by a surfactant-directing method. Then, the MIP

^aDepartment of Hypertension, Henan Provincial People's Hospital, Zheng Zhou, 450003, China. E-mail: liumin136@126.com

^bSchool of Chemical Engineering and Technology, Hebei University of Technology, Tianjin 300130, China. E-mail: mali0502@hebut.edu.cn

† Electronic supplementary information (ESI) available: Color evolution of TMB and ABTS in different reaction systems, optimization of experimental parameters, the selectivity of glucose detection. See DOI: 10.1039/c9ra05677g



layer was created on the surface of PtPd NFs and the obtained nanocomposites were named as T-MIP-PtPd NFs. The structure and composition of PtPd and T-MIP-PtPd were analyzed by scanning electron microscopy (SEM), transmission electron microscope (TEM), energy dispersive X-ray spectroscopy (EDX) and powder X-ray diffraction (XRD). Their intrinsic peroxidase-like activities were investigated. The T-MIP-PtPd NFs was used to detect H_2O_2 and glucose through a sensitive colorimetric method. To the best of our knowledge, it is the first report regarding the preparation of TMB imprinted PtPd NFs and its application in detecting hydrogen peroxide and glucose.

Materials and methods

Chemicals

Chloroplatinic acid (H_2PtCl_6) was purchased from Shanghai Macklin Biochemical Co., Ltd. (Shanghai, China). Potassium tetrachloroplatinate(II) (K_2PtCl_4), sodium tetrachloropalladate(II) (Na_2PdCl_4), ascorbic acid, 3,3',5,5'-tetramethylbenzidine (TMB), 2,2'-azino-bis(3-ethylbenzthiazoline-6-sulphonate) (ABTS), F127 and glucose oxidase (GOD) were purchased from Shanghai Aladdin Bio-Chem Technology Co., Ltd. (Shanghai, China). Glacial acetic acid, anhydrous sodium acetate, and ethanol were purchased from Tianjin Fengchuan Chemical Reagent Technologies Co., Ltd.

Preparation of PtPd NFs

Typically, the solutions of H_2PtCl_6 (20 mM, 1.2 mL), K_2PtCl_4 (20 mM, 1.8 mL) and Na_2PdCl_4 (20 mM, 0.6 mL) were mixed uniformly in a 10 mL centrifuge tube. Then F127 (60 mg) and HCl solution (6 M, 60 μL) were added. After F127 was completely dissolved, ascorbic acid (0.1 M, 3 mL) was added with ultrasonic at 40 °C. After 4 hours, the obtained PtPd NFs were centrifuged and washed with Milli-Q water and ethanol, respectively.

Molecular imprinting on PtPd NFs

The aqueous precipitation polymerization method is used to prepare TMB imprinted PtPd NFs (T-MIP-PtPd NFs). Typically, PtPd NFs (1 mg mL^{-1} , 0.5 mL) and TMB solution (7.5 mM, 20 μM) were mixed in HEPES buffer solution (20 mM, pH 7.6) that was purged with N_2 . Acrylamide (2.9 mg, 42 μM), *N*-isopropylacrylamide (NIPAAm) (4.6 mg, 42 μM), *N,N'*-methylenebisacrylamide (MBAAm as crosslinker) (2.4 mg, 16 μM) and SDS (0.8 mg) were dissolved in 0.5 mL HEPES buffer solution to prepare a monomer solution. Polymerization of the monomer solutions was initiated by adding ammonium persulfate (0.2 mg) and tetramethyl-ethylenediamine (0.3 μL). The monomer and the PtPd NFs-containing solutions were mixed after 20 min of initiation, and then the mixture was continued to react for 1 h at room temperature. The resulting imprinted gels were collected by centrifugation at 5000 rpm for 5 min. Then 1 mM H_2O_2 was added to react with the imprinted substrates so they can be easily removed by subsequent washing. As a control experiment, non-imprinted PtPd NFs nanozymes (NIP-PtPd NFs) were also prepared in the same way except that no substrate template was added.

Determination of peroxidase-like activity of PtPd NFs, NIP-PtPd NFs and T-MIP-PtPd NFs

Typically, 10 μL (0.1 mg mL^{-1}) nanozymes (PtPd NFs, NIP-PtPd NFs and T-MIP-PtPd NFs) and 50 μL of substrate (0.375 mM, TMB or ABTS) were added to 0.93 mL HAc–NaAc buffer solution (0.05 M, pH 4.0) containing 10 μL H_2O_2 (0.2 M). The reaction mixture was incubated at 25 °C for 10 min, and the peroxidase-like activity was investigated by detecting the absorbance (652 nm for TMB and 420 nm for ABTS). To measure the steady-state kinetics, various concentrations of substrates (TMB and ABTS) were used, respectively. For colorimetric detection, 0.1 mg mL^{-1} of nanozymes (PtPd NFs, NIP-PtPd NFs and T-MIP-PtPd NFs) and 0.1 mM of H_2O_2 were added to 9.3 mL acetate buffer solution (0.1 M, pH 4) containing substrates (TMB and ABTS) with various concentrations for 10 min at 25 °C. The kinetic parameters were calculated by the Michaelis–Menten equation (eqn (1)):

$$\frac{1}{v} = \frac{K_m}{V_{\max}} \left(\frac{1}{[S]} + \frac{1}{K_m} \right) \quad (1)$$

where $[S]$ is the concentrations of TMB or ABTS.

Detection of H_2O_2 and glucose

The detection of H_2O_2 by a colorimetric assay was carried out as follows: 10 μL of nanozyme (0.1 mg mL^{-1}) (PtPd NFs, NIP-PtPd NFs and T-MIP-PtPd NFs), 50 μL of TMB (7.5 mM), and 10 μL of H_2O_2 (0.2 M) were added into 930 μL of HAc–NaAc buffer solution (0.05 M, pH 4.0). Afterward, the absorbance was measured at 652 nm after 15 min of incubation at room temperature. For comparison, ABTS (50 μL 7.5 mM) was also used as colorimetric substrate to detect H_2O_2 , and the absorbance was measured at 420 nm after 15 min of incubation at room temperature.

PtPd NFs, NIP-PtPd NFs and T-MIP-PtPd NFs were also used to detect glucose through catalyzing the production of H_2O_2 from the oxidation of glucose by glucose oxidase. In a typical process, 300 μL of PBS (phosphate buffer solution, pH 7.0) containing a serials concentration of glucose and 100 μL of PBS (pH 7.0) containing 1 mg mL^{-1} glucose oxidase were mixed at 37 °C for 30 min. Then 540 μL of HAc–NaAc buffer solution, 50 μL of TMB (0.35 mM) and 10 μL of (0.1 mg mL^{-1}) nanozymes (PtPd NFs, NIP-PtPd NFs and T-MIP-PtPd NFs) were added to the above mixture, and incubated at 25 °C for 10 min to oxidize the reaction solution from colorless to blue.

Results and discussion

Characterization of T-MIP-PtPd NFs

As shown in Fig. 1, PtPd NFs were synthesized firstly and then the monomers including acrylamide, NIPAAm and MBAAm were mixed with PtPd NFs to perform precipitation polymerization. TMB was used as a template for imprinting. After the TMB imprinted polymer layer was formed on the PtPd NFs surface, the TMB molecules were washed away, leaving behind the binding pockets that can selectively rebinding TMB. As shown in Fig. 2A and C, SEM and TEM images of PtPd NFs



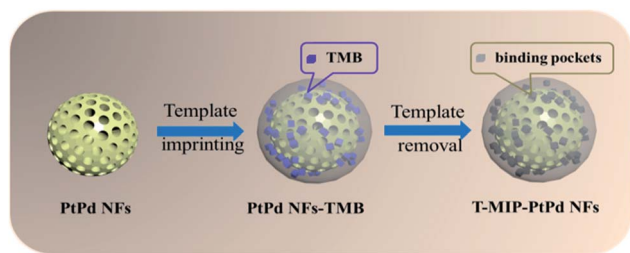


Fig. 1 The schematic diagram of preparing T-MIP-PtPd NFs.

indicated a spherical nanostructure with flower-like morphology and mesoporous on the surface. The particle size of PtPd NFs was approximately 120 nm (Fig. 2A and C). Compared to PtPd NFs, T-MIP-PtPd NFs had a bigger size due to the polymer coating on the surface of PtPd NFs (Fig. 2B and D). XRD pattern of T-MIP-PtPd NFs showed four strong diffraction peaks at 39.7° , 46.4° , 67.5° and 81.2° , which can be assigned to the (111), (200), (220) and (311) crystalline planes of the standard pattern of Pt and Pd, respectively^{16,24} (Fig. 2E). EDX analysis of the T-MIP-PtPd NFs showed obvious Pt, Pd, C, N and O peaks

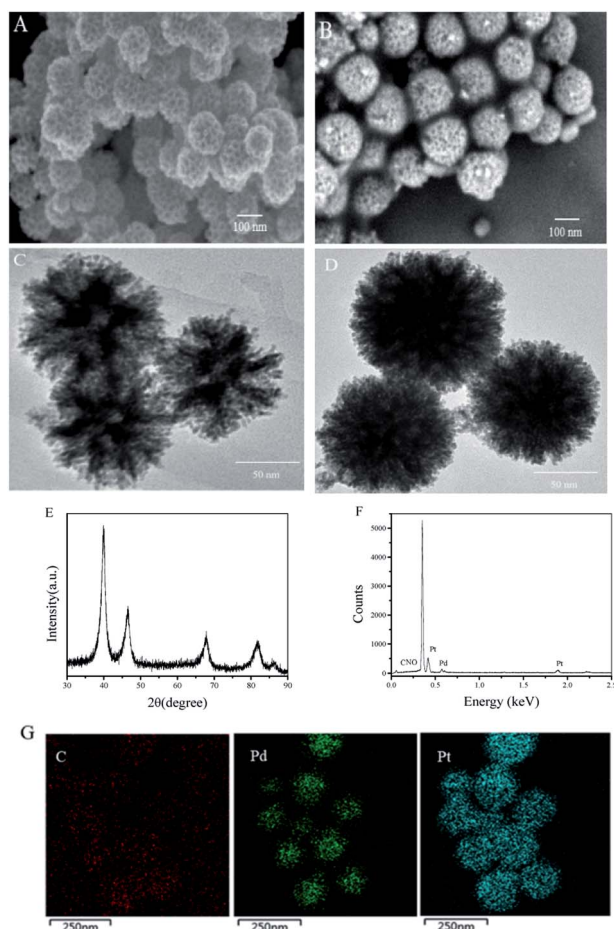


Fig. 2 SEM images of (A) PtPd NFs, (B) T-MIP-PtPd NFs, TEM images of (C) PtPd NFs, (D) T-MIP-PtPd NFs, (E) XRD pattern of T-MIP-PtPd NFs, (F) EDX spectrum of T-MIP-PtPd NFs and (G) elemental mapping images of T-MIP-PtPd NFs.

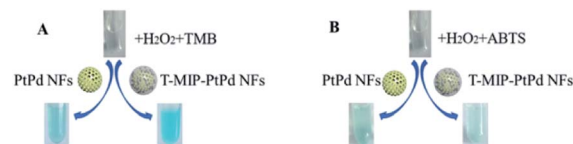


Fig. 3 Photographs with color change showing the activity and specificity of T-MIP-PtPd NFs for oxidation of (A) TMB and (B) ABTS with H_2O_2 .

(Fig. 2F). Moreover, elemental mapping images of T-MIP-PtPd NFs (Fig. 2G) showed that C, Pt and Pd element were homogeneously distributed through the T-MIP-PtPd NFs structure. The content of C element is much lower than Pt and Pd, and the boundary between C and Pt (and Pd) is unclear, indicating the polymer layer coating on the PtPd NFs is thin.

Peroxidase-like activity of T-MIP-PtPd NFs

The peroxidase activity of PtPd NFs, NIP-PtPd NFs and T-MIP-PtPd NFs was investigated at $25^\circ C$ using TMB as the substrate (Fig. S1†). Besides, similar reaction was also carried out using ABTS as reductant (Fig. S2†). These results confirmed that PtPd NFs, NIP-PtPd NFs and T-MIP-PtPd NFs were peroxidase mimicking nanozyme (Fig. 3A and B). The mechanism for the oxidation of TMB may be as follows: firstly, the d-band of platinum will become nearly bare when palladium atoms combine with platinum atoms. Then, the lone electron pairs of H_2O_2 will fill the d-band. After that, H_2O_2 cleave into $-OH$, and $-OH$ can oxidize TMB to TMB^+ .²⁵ Although both TMB and ABTS can be oxidized by T-MIP-PtPd NFs, it showed poor catalytic activity for ABTS.

Imprinting enhanced the activity of nanozyme

To compare the catalytic activity of PtPd NFs, NIP-PtPd NFs and T-MIP-PtPd NFs, the value changes in absorption spectra during the catalytic process was measured using UV-vis

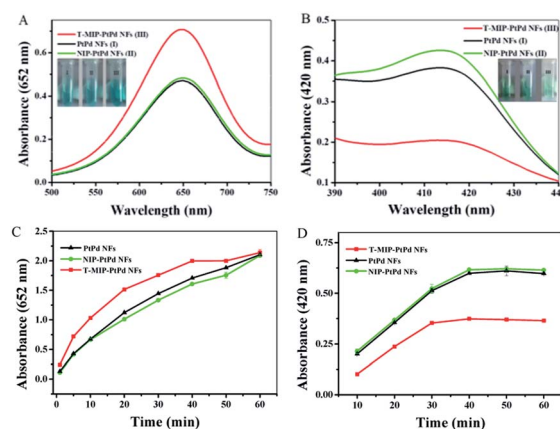


Fig. 4 UV-vis spectra after 30 min of reaction for (A) TMB and (B) ABTS oxidation, inset: photographs with color change showing peroxidase-like enzyme activity, the kinetics of (C) TMB oxidation monitored at 652 nm and (D) ABTS oxidation monitored at 420 nm by PtPd NFs, NIP-PtPd NFs and T-MIP-PtPd NFs.



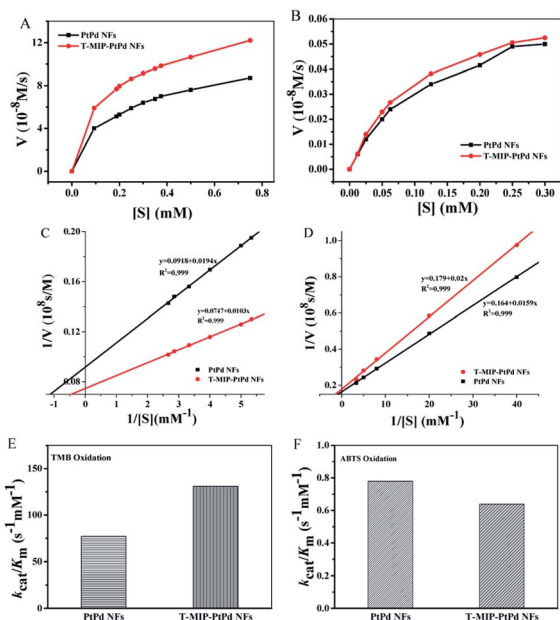


Fig. 5 Reaction rates with various concentrations of (A) TMB and (B) ABTS, kinetic parameters for oxidizing (C) TMB and (D) ABTS, and the catalytic efficiency (k_{cat}/K_m) of PtPd NFs and T-MIP-PtPd NFs for (E) TMB oxidation and (F) ABTS oxidation.

spectrophotometer. When PtPd NFs was used as catalyst, the absorbance at 652 nm reached 0.47 after reaction for 10 min (Fig. 4A, black trace). In contrast, when T-MIP-PtPd NFs were used as catalyst, the absorbance reached 0.71 (Fig. 4A, red trace), indicating the enhanced activity. According to these results, T-MIP-PtPd NFs could enhance the catalytic activity for TMB oxidation. The improved catalytic activity can be attributed to the TMB pockets, which imprinted in the PtPd NFs and had selective recognition for TMB molecules (Fig. 4C). Moreover, the increased local substrate concentration after MIP may also enhance the catalytic rate according to the mass action law.²⁶ The system of NIP-PtPd NFs with TMB and H_2O_2 exhibited similar absorption value to PtPd NFs system (Fig. 4A, green trace), indicating similar catalytic activity of NIP-PtPd NFs and PtPd NFs. Because the polymer layer around the PtPd NFs was thin, the polymer layer might have no obvious influence on the activity. In contrast, when ABTS was used as another substrate to investigate the selectivity of T-MIP-PtPd NFs, the catalytic activity of T-MIP-PtPd NFs for ABTS was lower than that of PtPd NFs and NIP-PtPd NFs (Fig. 4B and D). Therefore, after TMB imprinting, the selectivity of T-MIP-PtPd NFs was improved for TMB oxidation and inhibited for ABTS oxidation. In order to

achieve an optimal colorimetric effect, the effect of pH, temperature, reaction time, H_2O_2 concentration, and quantity of T-MIP-PtPd NFs on relative activity were investigated (Fig. S4†). The highest absorbance was reached when 0.1 mg mL^{-1} of T-MIP-PtPd NFs, 0.35 mM TMB, and 2.0 M H_2O_2 were used. Therefore, the above condition was selected in the following experiments.

Kinetic analysis

In order to investigate the mechanism of peroxidase-mimetic activity, the reaction kinetics of PtPd NFs and T-MIP-PtPd NFs were measured (Fig. 5A and B). Reaction rates with PtPd NFs and T-MIP-PtPd NFs as catalysts were tested by varying the concentration of TMB while fixing H_2O_2 concentration. For comparison, K_m of free HRP was also determined. The K_m value for PtPd NFs and T-MIP-PtPd NFs with TMB as substrate was 0.211 mM and 0.137 mM, respectively (Fig. 5C and D). The K_m value for T-MIP-PtPd NFs is much lower than those of PtPd NFs and free HRP (Table 1), indicating that T-MIP-PtPd NFs had stronger affinity to TMB than PtPd NFs and free HRP. However, the K_m for T-MIP-PtPd NFs with ABTS as substrate was higher than that of PtPd NFs, implying that the imprinting process only offer accessible site for TMB and then enhance substrate specificity.

Using TMB as substrate, the catalytic efficiency (k_{cat}/K_m) of PtPd NFs and T-MIP nanozymes were also investigated. Compared to PtPd NFs, the k_{cat} value of the T-MIP-PtPd NFs increased by 0.2-fold, while the K_m value dropped by 10% for

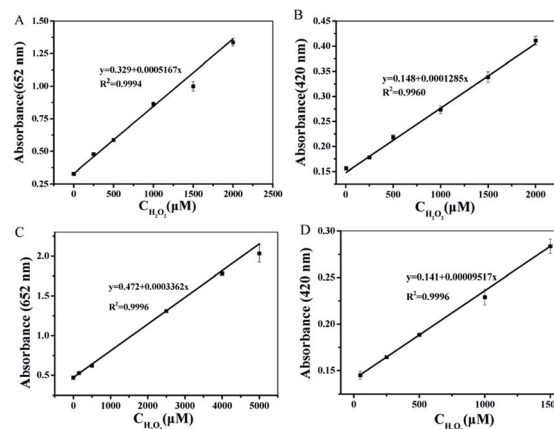


Fig. 6 Linear calibration plot for H_2O_2 with different concentration in absorbance of 652 nm reacted with (A) PtPd NFs and TMB, (B) PtPd NFs and ABTS, (C) T-MIP-PtPd NFs and TMB, (D) T-MIP-PtPd NFs and ABTS.

Table 1 Comparison of catalytic parameters of the PtPd NFs, T-MIP-PtPd NFs and HRP with TMB and ABTS as substrate

Nanozyme	Substrate	K_m (mM)	V_{max} ($10^{-8} M s^{-1}$)	k_{cat} (s^{-1})	k_{cat}/K_m ($s^{-1} mM^{-1}$)
PtPd NFs	TMB	0.211	10.87	16.28	77.2
T-MIP-PtPd NFs	TMB	0.137	13.30	17.94	131
HRP	TMB	0.434 (ref. 27)	10.00 (ref. 27)	—	—
PtPd NFs	ABTS	0.097	6.180	0.0756	0.78
T-MIP-PtPd NFs	ABTS	0.112	5.603	0.0715	0.639



Table 2 Comparison of different H₂O₂ sensors in terms of linear range and LOD

Materials	Substrate	Linear range (μM)	LOD (μM)	Reference
Fe ₃ O ₄ @C YSNs	TMB	1–20	0.39	28
PtPd NDs/GNs	TMB	0.5–150	0.1	13
PtPd NRs	TMB	0.02–50 000	0.0086	14
HRP	TMB	0.43	3.70	29
PtPd NFs	TMB	0.05–2000	0.02	This article
T-MIP-PtPd NFs	TMB	0.01–5000	0.005	This article

ABTS oxidation. We plotted k_{cat}/K_m of T-MIP-PtPd NFs for TMB and ABTS oxidation to compare PtPd NFs (Fig. 5E and F). The catalytic efficiency of T-MIP-PtPd NFs showed 0.7-fold higher than that of PtPd NFs for oxidizing TMB, but 0.15-fold lower for oxidizing ABTS. The imprinting in PtPd NFs can selectively enrich TMB substrate near the T-MIP-PtPd NFs and then result in enhanced catalytic activity.

Detection of H₂O₂

Under the optimized experimental conditions (see ESI†), a series concentration of H₂O₂ were detected based on the peroxidase-like activity of T-MIP-PtPd NFs through colorimetric method. When TMB was used as substrate, the absorbance of oxidized TMB is linearly correlated to concentration of H₂O₂ from 0.05–2000 μM (limit of detection (LOD) was 0.02 μM) catalyzed by PtPd NFs (Fig. 6A). The absorbance ranged from 0.01–5000 μM (LOD was 0.005 μM) catalyzed by T-MIP-PtPd NFs. This result proved that molecular imprinted nanozyme enhances its activity thanks to the pockets of TMB molecule (Fig. 6C). When ABTS was used as the substrate, the linear range between the absorbance and concentration of H₂O₂ was ranged from 0.05–2000 μM by PtPd NFs (Fig. 6B). For T-MIP-PtPd NFs, the linear range for H₂O₂ sensing was ranged from 0.01–1500

μM (Fig. 6D), indicating that the special pockets of TMB will hinder the approach of ABTS molecule. It can be seen from Table 2 that detection of H₂O₂ based on the T-MIP-PtPd NFs exhibited good performance, which further confirmed that the molecular imprinted method can enhance the catalytic activity.

Detection of glucose

Through combining T-MIP-PtPd NFs and GOx, we further developed a colorimetric method for the determination of glucose. Glucose can be catalyzed by GOx to generate gluconic acid and H₂O₂. In the presence of H₂O₂, TMB can be oxidized through the catalytic action of peroxidase-like nanozymes. The combination of two catalytic reactions was used to detect glucose through colorimetric method. With the increasing concentrations of glucose, the absorbance will increase from 0.05 to 1500 μM and a LOD of 0.023 μM for PtPd NFs was obtained (Fig. 7A). The T-MIP-PtPd NFs exhibits a linear detection range from 0.02 to 4000 μM and a LOD of 0.017 μM (Fig. 7B). Compared with the reported glucose sensors, T-MIP-PtPd NFs showed better catalytic performance for detecting glucose owing to the molecularly imprinted process (Table 3). This efficient detection performance was mainly ascribed to: (i) the T-MIP-PtPd NFs had excellent selectivity to TMB; (ii) the T-MIP-PtPd NFs provided an optimal microenvironment to stabilize GOx.

To evaluate the selectivity of the established method for glucose detection, the influence of lactose, galactose, fructose, and arabinose were investigated (Fig. S5†). All above interferences showed negligible effect on glucose detection. This phenomenon demonstrates that the reaction system is highly selective for glucose. The reproducibility of the T-MIP-PtPd NFs was investigated by analyzing 0.6 mM glucose for six times in parallel. The relative standard deviation (RSD) was 4.8%, which indicates that the testing results had acceptable reproducibility. To verify the feasibility of the as-prepared colorimetric method,

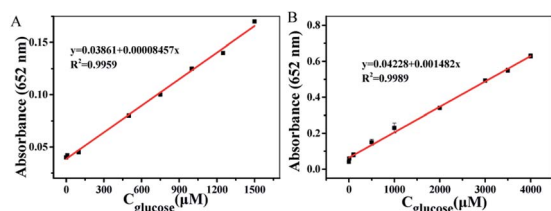


Fig. 7 Linear calibration plot for glucose with different concentration of glucose in absorbance of 652 nm with (A) PtPd NFs and (B) T-MIP-PtPd NFs.

Table 3 Comparison of colorimetric detection of glucose with different nanomaterials

Materials	Substrate	Linear range (μM)	LOD (μM)	Reference
PB NPs	ABTS	0.1–50	0.03	30
PB/MIL-101(Fe)	TMB	0.1–1000	0.04	4
Fe ₃ O ₄ @C YSNs	TMB	1–10	1.12	28
Au NPs	TMB	18–1100	4	31
PtPd NFs	TMB	0.05–1500	0.023	This article
T-MIP-PtPd NFs	TMB	0.02–4000	0.017	This article



glucose concentration in human serum samples were measured by the standard addition method (ESI†). All the obtained results were listed in Table S1.† It was found that the colorimetric method has a favorable recovery of 92.3–98.7% to detect glucose, which indicates that T-MIP-PtPd NFs, TMB and H₂O₂ system could be reliable for glucose detection in the biological environment.

Conclusions

In summary, molecular imprinting PtPd nanoflowers were successfully fabricated. The integration of PtPd nanozyme and molecular imprinting can enhance the peroxidase like activity and specificity of T-MIP-PtPd NFs. The obtained T-MIP-PtPd NFs exhibit better catalytic efficiency than that of PtPd NFs and HRP. Based on T-MIP-PtPd NFs, colorimetric sensor was developed for detecting hydrogen peroxidase and glucose. The T-MIP-PtPd showed better catalytic performance as peroxidase mimetic in the optical detection of hydrogen peroxidase and glucose. Thanks to these benefits, T-MIP-PtPd NFs provides a facile, low-cost, sensitive and selective method for colorimetric detection of H₂O₂ and glucose molecules. With good enzyme-like activity and specificity, the method of combining molecular imprinting with nanozyme can be extended to fabricate other kinds of nanozymes.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by Science and technology projects in Henan province (182102311235), the National Natural Science Foundation of China (No. 21576068, 21276060 and 21878068), the Natural Science Foundation of Hebei Province, China (B2017202056), the Program for Top 100 Innovative Talents in Colleges and Universities of Hebei Province, China (SLRC2017029) and Hebei High level personnel of support program, China (A2016002027).

Notes and references

- 1 D. Jiang, D. Ni, Z. T. Rosenkrans, *et al.*, *Chem. Soc. Rev.*, 2019, **48**, 3683–3704.
- 2 H. Wei and E. Wang, *Chem. Soc. Rev.*, 2013, **42**, 6060–6093.
- 3 S. Jin, C. Wu, Z. Ye, *et al.*, *Sens. Actuators, B*, 2019, **283**, 18–34.
- 4 F. Cui, Q. Deng and L. Sun, *RSC Adv.*, 2015, **5**, 98215–98221.
- 5 Y. Zhang, C. Dai, W. Liu, *et al.*, *Microchim. Acta*, 2019, **186**, 340–349.
- 6 W. Xu, L. Jiao, H. Yan, *et al.*, *ACS Appl. Mater. Interfaces*, 2019, **25**, 22096–22101.
- 7 Z. Zhao, T. Lin, W. Liu, *et al.*, *Spectrochim. Acta, Part B*, 2019, **219**, 240–247.
- 8 X. Liu, X. Wang, C. Qi, *et al.*, *Appl. Surf. Sci.*, 2019, **479**, 532–539.
- 9 X. Cheng, L. Huang, X. Yang, *et al.*, *J. Colloid Interface Sci.*, 2019, **535**, 425–435.
- 10 X. Cao and N. Wang, *Analyst*, 2011, **136**, 4241–4246.
- 11 X. Wang, Y. Hu and H. Wei, *Inorg. Chem. Front.*, 2016, **3**, 41–60.
- 12 T. Yu, W. Wang, J. Chen, *et al.*, *J. Phys. Chem. C*, 2012, **116**, 10516–10521.
- 13 X. Chen, B. Su, Z. Cai, *et al.*, *Sens. Actuators, B*, 2014, **201**, 286–292.
- 14 S. Ge, W. Liu, H. Liu, *et al.*, *Biosens. Bioelectron.*, 2015, **71**, 456–462.
- 15 B. Jiang, C. Li, M. Imura, *et al.*, *Adv. Sci.*, 2015, **2**, 1500112.
- 16 L. Ma, L. Zhou, Y. He, *et al.*, *Electroanalysis*, 2018, **30**, 1801–1810.
- 17 M. Dinc, C. Esen and B. Mizaikoff, *TrAC, Trends Anal. Chem.*, 2019, **114**, 202–217.
- 18 H. Zhang, *Adv. Mater.*, 2019, e1806328.
- 19 M. Zarejousheghani, W. Lorenz, P. Vanninen, *et al.*, *Polymers*, 2019, **11**, 888–905.
- 20 W. Zhao, H. Yang, S. Xu, *et al.*, *Colloids Surf., A*, 2018, **555**, 95–102.
- 21 T. Kajisa and T. Sakata, *ACS Appl. Mater. Interfaces*, 2018, **10**, 34983–34990.
- 22 D. Duan, H. Yang, Y. Ding, *et al.*, *Electrochim. Acta*, 2018, **261**, 160–166.
- 23 Z. Zhang, X. Zhang, B. Liu, *et al.*, *J. Am. Chem. Soc.*, 2017, **139**, 5412–5419.
- 24 L. Wang and Y. Yamauchi, *J. Am. Chem. Soc.*, 2013, **135**, 16762–16765.
- 25 R. Mandal, A. Baranwal, A. Srivastava, *et al.*, *Biosens. Bioelectron.*, 2018, **117**, 546–561.
- 26 Z. Zhang, Y. Li, X. Zhang, *et al.*, *Nanoscale*, 2019, **11**, 4854–4863.
- 27 V. K. Singh, P. K. Yadav, S. Chandra, *et al.*, *J. Mater. Chem. B*, 2018, **6**, 5256–5268.
- 28 N. Lu, M. Zhang, L. Ding, *et al.*, *Nanoscale*, 2017, **9**, 4508–4515.
- 29 J. Liu and Y. Lu, *Angew. Chem., Int. Ed.*, 2005, **45**, 90–94.
- 30 W. Zhang, D. Ma and J. Du, *Talanta*, 2014, **120**, 362–367.
- 31 Y. Jv, B. Li and R. Cao, *Chem. Commun.*, 2010, **46**, 8017–8019.

