Cite this: *RSC Adv.*, 2019, 9, 26043

Two AIEE-active α -cyanostilbene derivatives containing BF_2 unit for detecting explosive picric acid in aqueous medium†

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Two novel α -cyanostilbene derivatives bearing triphenylamine and BF_2 groups are synthesized (named TPE-B and TPE-BN). The fluorescent emissions of compounds TPE-B and TPE-BN are hypochromatically shifted and bathochromically shifted, respectively, with increasing polarity of the solvents, suggesting that the two compounds have characteristic polarity-dependent solvatochromic effects. Furthermore, they show obvious aggregation-induced emission enhancement (AIEE) phenomenon in THF/water mixture solutions. Meanwhile, compounds TPE-B and TPE-BN emit orange and yellow fluorescence in their solid states, respectively. Most significantly, in aqueous medium, compounds TPE-B and TPE-BN can selectively and sensitively detect picric acid (PA) among a number of nitroaromatic compounds, and their limits of detection (LOD) are calculated as 1.26×10^{-6} M and 1.51×10^{-6} M, respectively. The recognition mechanism for PA can be attributed to the photo-induced electron transfer (PET) process and this is supported by density functional theory (DFT) calculation. This research provides two novel compounds for the rational design of AIEE-active materials for sensing systems.

Received 5th July 2019
Accepted 12th August 2019

DOI: 10.1039/c9ra05116c

rsc.li/rsc-advances

1. Introduction

During the past decade, aesthetic organic fluorophores with aggregation induced emission (AIE) or aggregation-induced enhanced emission (AIEE) properties have received intense attention by virtue of their extensive applications as imaging agents and sensors, components of light-emitting diodes, and data recording and storage, among others.^{1–8} Developing functional AIE/AIEE materials can provide several advantages over conventional materials, such as their high quantum yields, controlled self-assembly effects, photostabilities, controllable emission colour, and flexible post-functionalization such as tetraphenylethene, α -cyanostilbene, silole, diphenyldibenzofulvene, organoboron, phosphindole oxide, pyrazine, phenothiazine derivatives, *etc*.^{9–13} The twisted molecular conformations of organic fluorophores can emit strong fluorescence in the aggregated state by suppressing a dense face-to-face packing and immense π - π stacking interaction. Also, in the aggregated state, the molecules interlock themselves which

restricts the intramolecular rotation (RIR) and vibration (RIV) and relaxes through radiative pathways.^{14–21} Therefore, it is of great significance to design and understand the relationships between structure and property for their further applications.^{22–24}

For the purpose of obtaining eminent organic molecules with strong fluorescence in aggregated states, the molecular design according to organic fluorophores is critical for adjusting their photophysical properties. Triphenylamine (TPA) is strong electron-donating structural units with good luminescence properties, high thermal, photostability, good absorptivity, and high absolutely quantum yields.^{25–27} Furthermore, an organoboron compound allowing trigonal-planar geometry, endows high extinction coefficients and high absolutely fluorescence quantum yields.^{28–31,55} The combination of α -cyanostilbene modified with triphenylamine and BF_2 groups were used to design AIEE-active compounds TPE-B and TPE-BN (Scheme 1).^{32–34} In the present investigation, it was found that picric acid (PA) is a powerful explosive as TNT, only a few available PA

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† Electronic supplementary information (ESI) available: Photophysical properties; the Sterne–Volmer plots; UV-vis spectra in THF/H₂O mixtures. PL emission spectra changes of TPA-B in THF/H₂O mixtures; photophysical properties of PA detection for compound TPA-BN; normalized absorption spectrum of PA and the fluorescence spectra of TPE-B and TPE-BN. See DOI: 10.1039/c9ra05116c

Scheme 1 Synthetic routes of compounds TPE-B and TPE-BN.

sensing reports in aqueous medium. Meanwhile, PA is a seriously environmental pollutant, which consequently causes damage of human health and aquatic systems.^{35–39} We expect two compounds to exhibit brilliant PA detection in aqueous medium.

Based on the above considerations, two α -cyanostilbene derivatives (named **TPE-B** and **TPE-BN**) were synthesized *via* facile reaction. The UV-vis absorption spectra and fluorescence spectra of compounds **TPE-B** and **TPE-BN** were systematically investigated in different polar solvents. Meanwhile, the aggregates emission behaviors of compounds were studied in THF/water mixture solution. In addition, the recognition properties of **TPA-B** and **TPA-BN** to PA were studied and the corresponding sensor mechanism was analyzed by density functional theory (DFT) calculations.

2. Experimental section

2.1 Materials and instruments

The chemicals and solvents were purchased from a commercial supplier and used directly without any further refinement. The ¹H and ¹³C NMR spectra were identified by a Bruker Avance 400 MHz spectrometer with tetramethylsilane as the internal standard. FT-IR spectra were obtained by Nicolet 380 spectrometer (4000–400 cm^{−1}, KBr pellets). UV-vis absorption spectra were measured using a TU-1901 spectrometer and the fluorescence spectra were collected by Hitachi FL-7000 spectrofluorimeter from Hitachi High Technologies Corporation (Tokyo, Japan). Photo images were obtained using a digital camera (Nikon D7000).

2.2 Synthesis

The synthetic routes of compounds **TPE-B** and **TPE-BN** are shown in Scheme 1.

2.2.1. Synthesis of compound 1. The 4-*N*,*N*-diphenylaminobenzaldehyde (410 mg, 1.50 mmol) and *p*-nitrophenylacetonitrile (240 mg, 1.50 mmol) were dissolved in absolute ethanol (40 mL). After adding 2 drops of piperidine, the reaction mixture was refluxed for 5 h. After cooling to room temperature, the red precipitate was filtered, washed with ethanol for three times. The final product was dried in vacuum and collected as red powder **1** (290 mg, 0.69 mmol, yield: 46%). ¹H NMR (DMSO-*d*₆, 400 MHz) δ (ppm): 6.95 (d, *J* = 7.9 Hz, 2H), 7.17–7.23 (m, 6H), 7.42 (t, *J* = 7.9 Hz, 4H), 7.92 (d, *J* = 8.4 Hz, 2H), 7.98 (d, *J* = 8.0 Hz, 2H), 8.15 (s, 1H), 8.32 (d, *J* = 8.4 Hz, 2H). ¹³C NMR (DMSO-*d*₆, 100 MHz) δ (ppm): 103.70, 118.01, 119.21, 124.29, 125.10, 125.22, 125.96, 126.30, 129.94, 131.52, 140.84, 145.56, 145.69, 146.75, 150.37.

2.2.2 Synthesis of compound 2. Compound **1** (250 mg, 0.60 mmol) and SnCl₂·2H₂O (680 mg, 3.01 mmol) were reflux under absolute ethanol (40 mL) for 2 hours. After cooling to room temperature, the mixture was poured into a saturated K₂CO₃ aqueous solution and extracted with dichloromethane. The organic layer was dried with MgSO₄ and concentrated by vacuum evaporation. The crude product was purified by column chromatography (petroleum ether : ethyl acetate = 2 : 1). The

final product was dried in vacuum and collected as orange powder **2** (180 mg, 0.46 mmol, yield: 76%). ¹H NMR (DMSO-*d*₆, 400 MHz) δ (ppm): 5.57 (s, 2H), 6.55 (d, *J* = 8.0 Hz, 1H), 6.63 (d, *J* = 8.4 Hz, 1H), 6.73 (d, *J* = 7.9 Hz, 1H), 6.96 (d, *J* = 8.3 Hz, 2H), 7.03–7.08 (m, 2H), 7.10–7.19 (m, 5H), 7.35–7.40 (m, 4H), 7.57 (s, 1H), 7.76 (d, *J* = 8.4 Hz, 2H). ¹³C NMR (DMSO-*d*₆, 100 MHz) δ (ppm): 107.71, 113.90, 118.78, 120.84, 121.25, 124.15, 125.09, 126.45, 127.40, 129.73, 129.95, 136.21, 146.31, 148.35, 149.67. MALDI-TOF calcd for C₂₇H₂₁N₃, 387.170, found, 387.947.

2.2.3 Synthesis of compounds 3 and 4. A 50 mL round bottom flask was charged with compound **2** (580 mg, 1.50 mmol), aldehyde (1.80 mmol) and absolute ethanol (70 mL). A dropwise glacial acetic acid was added into the mixture solution. The mixture solution was refluxed for 12 h. After cooling to room temperature, the precipitate was filtered. The final product was dried in vacuum and collected.

Red powder **3** (530 mg, 1.08 mmol, yield: 72%). FT-IR (cm^{−1}, KBr) ν : 3423, 2211, 1618, 1581, 1535, 1332, 1285, 1196, 1178, 902, 829, 753, 696, 621, 535, 511. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 6.94–6.98 (m, 2H), 7.04 (d, *J* = 9.2 Hz, 2H), 7.19–7.23 (m, 7H), 7.38–7.47 (m, 5H), 7.62 (d, *J* = 7.6 Hz, 1H), 7.79–7.87 (m, 4H), 7.92 (d, *J* = 9.2 Hz, 2H), 8.99 (s, 1H), 13.06 (s, 1H). ¹³C NMR (CDCl₃, 100 MHz) δ (ppm): 106.78, 117.31, 118.63, 119.19, 120.74, 121.80, 124.45, 125.44, 125.74, 126.25, 126.67, 129.57, 130.71, 132.44, 133.43, 133.62, 141.38, 146.52, 148.41, 150.03, 161.22, 162.78. MALDI-TOF calcd for C₃₄H₂₅N₃O, 491.200, found, 491.671.

Red powder **4** (590 mg, 1.04 mmol, yield: 69%). FT-IR (cm^{−1}, KBr) ν : 3430, 2965, 2924, 2210, 1633, 1558, 1520, 1375, 1193, 1177.69, 1133, 1076, 874, 972, 845, 751, 727. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 1.18–1.22 (m, 6H), 3.45–3.50 (m, 4H), 6.15 (s, 1H), 6.37 (d, *J* = 8.7 Hz, 2H), 7.03 (d, *J* = 8.8 Hz, 2H), 7.16–7.20 (m, 6H), 7.31 (d, *J* = 8.8 Hz, 2H), 7.37–7.43 (m, 5H), 7.77 (d, *J* = 8.6 Hz, 3H), 7.91 (d, *J* = 8.3 Hz, 2H), 8.70 (s, 1H), 13.40 (s, 1H). ¹³C NMR (CDCl₃, 100 MHz) δ (ppm): 12.73, 44.65, 97.78, 104.05, 107.36, 109.16, 118.74, 120.98, 121.35, 124.34, 125.68, 126.15, 126.62, 129.56, 130.59, 132.69, 133.78, 133.99, 140.62, 146.67, 149.85, 152.12, 160.49, 164.21. MALDI-TOF calcd for C₃₈H₃₄N₃O, 562.273, found, 562.995.

2.2.4 Synthesis of compounds TPA-B and TPA-BN. The compound **3** or **4** (1.0 mmol) and *N,N*-diisopropylethylamine (DIPEA, 1 mL) were dissolved in CH₂Cl₂ (10 mL) and the reaction was stirred at room temperature for 2 h. Then, BF₃·OEt₂ (1.5 mL) was dropwise added into the mixture solution. The reaction mixture was stirred and monitored by TLC until completed. The mixture was poured into cold water (80 mL), extracted with CH₂Cl₂ and dried with MgSO₄. After the solvent is evaporated under reduced pressure. The solid was obtained by chromatography on a silica gel column using DCM as the eluent.

Orange red powder **TPA-B** (400 mg, 0.75 mmol, yield: 75%). FT-IR (cm^{−1}, KBr) ν : 3056, 2923, 2853, 2217, 1627, 1583, 1556, 1534, 1459, 1389, 1331, 1329, 1270, 1216, 1179, 1153, 1047, 758, 698, 530, 512. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 5.53 (s, 3H), 6.83 (d, *J* = 7.5 Hz, 1H), 7.03–7.18 (m, 8H), 7.29–7.35 (m, 3H), 7.48 (s, 1H), 7.53 (t, *J* = 7.4 Hz, 1H), 7.59 (s, 1H), 7.63 (d, *J* = 7.9 Hz, 1H), 7.69 (t, *J* = 8.0 Hz, 1H), 7.76–7.81 (m, 3H), 8.48 (s,

¹H). ¹³C NMR (CDCl₃, 100 MHz) δ (ppm): 105.76, 118.35, 120.53, 120.61, 124.12, 124.42, 124.66, 125.54, 125.76, 125.90, 126.83, 129.47, 129.54, 129.63, 130.22, 131.02, 132.26, 136.09, 139.58, 143.02, 146.42, 150.50. MALDI-TOF calcd for C₃₄H₂₄BF₂N₃O, 538.202, found, 538.595.

Yellow powder **TPA-BN** (410 mg, 0.66 mmol, yield: 66%). FT-IR (cm⁻¹, KBr) ν : 3035, 2975, 2929, 2204, 1626, 1589, 1509, 1459, 1352, 1313, 1297, 1278, 1241, 1217, 1192, 1145, 1075, 1030, 971, 961, 898, 842, 750, 729, 529. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 1.23–1.27 (m, 6H), 3.49–3.44 (m, 4H), 6.25 (s, 1H), 6.38 (d, J = 9.2 Hz, 1H), 6.82 (d, J = 8.6 Hz, 1H), 7.04–7.17 (m, 9H), 7.28–7.34 (m, 4H), 7.43 (s, 1H), 7.51 (s, 1H), 7.56 (d, J = 8.4 Hz, 1H), 7.23 (s, 1H), 7.69 (d, J = 8.5 Hz, 1H), 7.79 (d, J = 8.6 Hz, 1H), 8.06 (s, 1H).

¹³C NMR (CDCl₃, 100 MHz) δ (ppm): 12.69, 45.24, 98.03, 106.40, 106.97, 107.17, 118.58, 120.75, 123.49, 124.51, 125.79, 126.22, 126.57, 129.62, 130.86, 134.17, 134.20, 142.03, 143.11, 146.55, 150.16, 156.48, 157.52, 161.96. MALDI-TOF calcd for C₃₈H₃₃BF₂N₄O, 609.271, found, 609.830.

3. Results and discussion

3.1 Solvatochromic effect

The UV-vis absorption and photoluminescence (PL) spectra of compounds **TPA-B** and **TPA-BN** were measured in various polar solvents and demonstrated in Fig. S1† and Fig. 1, respectively, and the corresponding photophysical data were summarized in Table 1. All the spectroscopic studies were performed at room temperature and the concentration of the solutions were 1.0×10^{-5} mol L⁻¹. The UV-vis absorption spectra of **TPA-BN** showed two characteristic absorption bands at 297 nm and 420 nm in various polarity solvents, respectively. The absorption peak at 297 nm is ascribed to the π - π^* transition in TPA moiety corresponding to locally excited (LE) state,^{27,40,41} while the latter peak at 420 nm denotes intramolecular charge transfer (ICT) process. The UV-vis absorption of **TPA-B** has a similar peak shapes. And the emission wavelengths of two compounds revealed larger shift with increasing polarity. For compound **TPA-B**, the emission peak was hypochromatic-shifted from 541 nm in benzene to 530 nm in ethanol, indicated that **TPA-B** molecule possibly had a larger polarity in the ground state than in the excited state. However, the fluorescent emission peak of compound **TPA-BN** was bathochromic-shifted from 516 nm in benzene to 540 nm in DMF, indicated that **TPA-BN** molecule might have a larger polarity in the excited state than in the

Table 1 Photophysical properties of compounds **TPA-B** and **TPA-BN** in different polar solvents

Compounds	Solvents	λ_{abs}^a	ϵ_{max}^b	λ_{fl}^c	$\Delta\nu^d$	Φ^e (%)
TPA-B	Benzene	419	4.59	541	5382	8.8
	DCM	427	7.10	538	4832	1.0
	Ethyl acetate	420	6.64	570	6262	2.3
	THF	420	6.76	531	4977	1.5
	Ethanol	408	6.06	530	5642	1.2
	Acetonitrile	420	7.19	535	5118	0.1
TPA-BN	DMF	423	7.07	540	5122	0.1
	Benzene	423	7.78	516	4260	9.1
	DCM	418	7.93	533	5161	6.1
	Ethyl acetate	415	7.81	519	4828	8.8
	THF	416	7.75	520	4807	4.2
	Ethanol	414	7.59	526	5143	6.9
	Acetonitrile	422	8.76	542	5246	1.2
	DMF	425	7.66	540	5010	3.1

^a Absorption peak position in nm (1×10^{-5} mol L⁻¹). ^b Maximum molar absorbance in 10^4 mol⁻¹ L cm⁻¹. ^c Peak position of fluorescent (1.0×10^{-5} mol L⁻¹), excited at the absorption maximum. ^d Stokes shift in cm⁻¹. ^e Quantum yields determined (RhB as the standard).

ground state.^{42–44} Meanwhile, the evident shift of emission peaks manifested the existence of ICT process. The slopes of the fitting line in Lippert–Mataga plots (Fig. S2†), which were –646.5 and 2887.8, respectively, indicated that the compound **TPA-BN** introduced into the electron-donating group significantly possessed the solvatochromic effects. And, Fig. 5 shows that the TPA group has a larger density in the highest occupied frontier molecular orbital (HOMO) of **TPA-B** molecular. As the electron density of the lowest unoccupied molecular orbital (LUMO) of the TPA unit decreases, the other moieties achieve larger electron density. For **TPA-BN**, the HOMO and LUMO were mostly distributed over the molecular and TPA groups, respectively, which further manifested that the ICT process might occur in **TPA-B** and **TPA-BN**.⁶⁰ It was indicated that two compounds were characteristic of polarity-dependent solvatochromic effects.

3.2 Aggregation-induced emission enhancement (AIEE) property

Aiming to investigate the AIEE characteristics of **TPA-B** and **TPA-BN**, their UV-vis absorption spectra and PL spectra with different water fraction were performed. The concentrations were kept at 1.0×10^{-5} mol L⁻¹. For UV-vis absorption spectra of compound **TPA-BN** (Fig. S3b†), with the increasing water fraction, the solubility of the compound gradually decreased and the molecules gradually shifted from the monomolecular state to the aggregation state, forming the molecular aggregated particles. The level-off tails attributed to Mie scattering were responsible for the aggregated forms in the UV-vis absorption spectra.^{45–50} A similar change trend was also observed from the UV-vis absorption spectra of **TPA-B** with the change of water contents. For compounds **TPA-B** and **TPA-BN**, dynamic light scattering (DLS) experiment displays the formation of aggregates (the hydrodynamic diameter of 267.2 nm and 390.2 nm, respectively). For PL spectra (Fig. 2), in pure THF solution,

Fig. 1 UV-vis absorption spectra (A) and PL spectra (B) of the compound **TPA-BN** in different solvent.

compound **TPA-BN** emitted weak yellow-green fluorescence. Compound **TPA-BN** started gradually aggregate with the increasing water fraction, the PL intensity showed a remarkable enhancement accompanied with a bathochromic-shift of fluorescent emission peak. This emission behavior may be accounted for the restriction of intramolecular rotation (RIR) and forms of *J*-aggregation,⁵¹ which blocks the channel of non-radiative decay in the aggregates. Furthermore, when the water fraction is higher than 80%, the PL intensities gradually decreased, because the settlement of large aggregated particles resulted in a lower fluorescence intensity. Moreover, we found that the PL spectra of the compound **TPA-B** (Fig. S4†) displayed a similar behavior. The PL intensity of the compounds **TPA-B** and **TPA-BN** in the aggregated state were obviously stronger than that of the THF solution, the PL intensities were 1.6 and 2.0 fold, respectively. It was easily concluded that compounds **TPA-B** and **TPA-BN** were typical AIEE-active molecules.

Furthermore, PL spectra of compounds **TPA-B** and **TPA-BN** were investigated to explore the difference of solid emission. As was shown in Fig. 3, **TPA-B** exhibited orange emission, whereas **TPA-BN** containing an additional electron-donating group has a obvious influence on hypsochromic shift of fluorescent peaks, emitting yellow fluorescence. The solid state fluorescent quantum yields of **TPA-B** and **TPA-BN** were measured as 13.5% and 19.4%, respectively in the solid state. The possible fluorescence emission discrepancy was caused by the role of intermolecular arrangement and stacking in solid state. Thus, the introduction of additional electron-donating group could effectively affect their fluorescence emission behavior and result in different spectral shifts and absolute quantum yields.

3.3 The detection for picric acid

Based on brilliant AIEE behavior of compounds **TPA-B** and **TPA-BN** in mixtures of THF/H₂O. We next explore whether compounds **TPA-B** and **TPA-BN** can detect common nitroaromatic compounds (*p*-methyl phenol (*p*-MP), *o*-nitrophenol (*o*-NP), 2,4,6-trinitrotoluene (TNT), *m*-dihydroxybenzene (*m*-DOB), *p*-nitrophenol (*p*-NP), *p*-dihydroxybenzene (*p*-DOB), phenol (PhOH), *p*-nitroaniline (*p*-NA), nitrotoluene (NT), nitrobenzene (NB), 2,4,6-trinitrophenol (PA)) in aqueous medium, the PL spectra were measured with addition of 10 equivalents nitroaromatic compounds. As shown in Fig. 4a and S6a,†

Fig. 3 Normalized PL spectra of two solids. Inset: the corresponding fluorescence images of **TPA-B** and **TPA-BN** under illumination.

compared with other analytes, it was clearly observed that the positive interactions of **TPA-B** and **TPA-BN** with PA showed significant fluorescent quenching effect. Experimental results exhibited that other nitroaromatics exhibited relatively little effect on emission quenching compared with that of PA, suggesting the selective sensing behaviors of **TPA-B** and **TPA-BN** towards PA. Furthermore, it is worth noting that further additions of 10 equivalents PA into other nitroaromatic systems leads to remarkable quenching efficiency of fluorescence intensity (Fig. 4b and S6b†), means that other compounds have negligible effect on the detection of PA by **TPA-B** and **TPA-BN**. According to the above results, compounds **TPA-B** and **TPA-BN**

Fig. 2 PL emission (a) spectra changes of **TPA-BN** (5.0×10^{-5} M) in THF/H₂O mixtures with different water volume fractions; plots of PL intensity determined in THF–H₂O solutions versus water fractions (b). Insets: photos of **TPA-BN** in THF–H₂O mixtures ($f_w = 0\%$, 80% and 99%) taken under 365 nm UV lamp.

Fig. 4 (a) PL spectra obtained for different analytes; (b) quenching percentages of compound **TPA-BN** ($10.0 \mu\text{M}$) with different analytes in THF/water ($v/v = 2 : 8$) mixtures before (black) and after (red) the addition of 10 equiv. a: *p*-MP, b: *o*-NP, c: TNT, d: *m*-DOB, e: *p*-NP, f: *p*-DOB, g: PhOH, h: *p*-NA, i: NT, j: NB; (c) PL spectra of **TPA-BN** ($10.0 \mu\text{M}$) in THF/water ($v/v = 2 : 8$) containing different amounts of PA ($100.0 \mu\text{M}$); (d) corresponding Stern–Volmer plot for PA detection. Inset: Stern–Volmer plot obtained at a lower concentration of PA.

are regarded as promising candidates for PA detection in aqueous medium.

The key to performing better practical application, the experiment was carried out to better explore the sensitivity of compounds **TPA-B** and **TPA-BN** towards PA in aqueous medium. As shown in Fig. 4c and Fig. S6c,† the fluorescent quenching efficiencies of **TPA-B** and **TPA-BN** were 82.7% and 81.6% in the presence of 10 equivalents PA, respectively. In order to evaluate the quenching efficiency of **TPA-B** and **TPA-BN** in response to the PA in solution, a quenching process can be analyzed by Stern–Volmer equation,⁵² the equation was as follow:

$$I_0/I = K_{sv}[A] + 1$$

where I_0 and I are the initial and final PL intensity after addition of PA, respectively; $[A]$ is the molar concentration of analytes; and K_{sv} is the Stern–Volmer quenching constant. By fitting the linear curves, the slopes of the plots gave the K_{sv} of **TPA-BN** and **TPA-B**, which were $1.28 \times 10^4 \text{ M}^{-1}$ and $1.07 \times 10^4 \text{ M}^{-1}$, respectively. The limit of detection (LOD)^{53,54} was obtained from the equation of $\text{LOD} = 3\sigma/k$, wherein σ is the standard deviation of the blank test, k represents the slope of the linear calibration curve, the LOD were calculated as $1.26 \times 10^{-6} \text{ M}$ for **TPA-B** and $1.51 \times 10^{-6} \text{ M}$ for **TPA-BN**, respectively (Fig. S7†). Compounds exhibiting brilliant PA detection in aqueous medium are comparable with organoboron chemosensors in the Table S1.†^{56–59}

Density functional theory (DFT) calculation was used to investigate possible PA sensing mechanism. As shown in Fig. 5, the LUMO level (−2.79 eV) of **TPA-B** is higher than that of PA (−3.90 eV), which suggested when excited **TPA-B** was exposed to PA, the excited electron was transferred from the LUMO of fluorophores to that of the electron-deficient PA. The radiative electronic transition was blocked due to the electron transfer from the chromophore to PA, resulting in fluorescence quenching. Moreover, the other driving force is the energy difference between the LUMO of the fluorophore and the LUMO of analytes.^{61–65} The LUMO orbital energy typically indicates how easily an electron can be transferred from excited fluorophore to the electron-deficient analytes. As the LUMO orbital energies of PA and TNT calculated by DFT is −3.90 eV and −3.33 eV, respectively. As expected, because of its small LUMO value, the

PET driving force of PA is larger than that of TNT. This may partly explain the sensitivity and selectivity of **TPA-B** for PA detection. Similarly, for **TPA-BN**, the LUMO energy level of PA is between the LUMO energy level and the HOMO energy level of **TPA-BN**, indicating that the fluorescence quenching of **TPA-BN** with PA may suggest that the possibility of electron transfer from the LUMO of the chemosensor to the lower LUMO of PA. Based on the above results, the main driving force was attributed to the photo-induced electron transfer (PET) process, resulting in fluorescence quenching. Meanwhile, as was showed in the Fig. S8,† the UV-vis absorption spectra of PA and the PL spectra of **TPA-B** and **TPA-BN** were recorded and presented to examine whether fluorescence resonance energy transfer (FRET) occurs in detection process. The absorption spectrum of PA has no overlap with the PL spectra of the two chemosensors, indicating that FRET was neglectable in the quenching process. Therefore, the PET process may be main reason for recognition mechanism between the chemosensors and PA.

4. Conclusions

In conclusion, two novel α -cyanostilbene derivatives (**TPA-B** and **TPA-BN**) containing triphenylamine and BF_2 units are synthesized and characterized that displayed obvious solvatochromic effect. Their ICT behaviors are certified by DFT analysis on molecular orbitals. Meanwhile, the incorporation of α -cyanostilbene unit in the backbone endows molecules an obvious AIEE behaviour in THF–water mixture solvent. Compounds showed excellent fluorescence emission in the solid state with high absolute quantum yield. Most impressing, **TPA-B** and **TPA-BN** were used as highly selectively and sensitively chemosensor to detect PA in aqueous medium and the LODs were calculated as $1.26 \times 10^{-6} \text{ M}$ and $1.51 \times 10^{-6} \text{ M}$, respectively. The efficient sensing mechanism of two chemosensors in the presence of PA contributed to the PET process, verified by DFT calculations on molecular orbitals. This research provides two novel compounds for the rational design of AIEE-active materials for sensing application in aqueous medium.

Conflicts of interest

We declare that we do not have any commercial or associative interest that represents a conflict of interest in connection with the work submitted.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (51673001, 51432001), the Educational Commission of Anhui Province of China (KJ2014ZD02).

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Fig. 5 HOMO and LUMO energy–level diagram of **TPA-B/TPA-BN** and PA.

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