



Cite this: *RSC Adv.*, 2019, 9, 26856

^{15}N labeling and analysis of ^{13}C – ^{15}N and ^1H – ^{15}N couplings in studies of the structures and chemical transformations of nitrogen heterocycles

Sergey L. Deev,^a Igor A. Khalymbadzha,^a Tatyana S. Shestakova,^a Valery N. Charushin^{ab} and Oleg N. Chupakhin^{ab}

This review provides a generalization of effective examples of ^{15}N labeling followed by an analysis of ^{13}C – ^{15}N (J_{CN}) and ^1H – ^{15}N (J_{HN}) coupling constants in solution as a tool to study the structural aspects and pathways of chemical transformations (e.g., rearrangements and ring-chain tautomerisms) in monocyclic and fused nitrogen heterocycles. This approach allows us to significantly expand and supplement the scope of NMR techniques for heterocyclic compounds. Moreover, methods for the incorporation of ^{15}N atoms into the cores of various N-heterocycles have been collected in this work.

Received 26th June 2019
Accepted 19th August 2019

DOI: 10.1039/c9ra04825a

rsc.li/rsc-advances

1. Introduction

Nitrogen heterocycles are a ubiquitous class of organic compounds that include azirine, azetidione, azole, azine and azepine derivatives and their fused analogs. These structures are inherent in drug design^{1–7} and natural compounds,^{8,9} in catalysis for cross-coupling and asymmetric synthesis reactions,^{10,11} in materials science as metal complexes with luminescent properties,^{12–14} in ligands for the separation of lanthanides¹⁵ and in high-density energy materials.^{16,17}

At first glance, it seems that establishing the structure of poly-nitrogen-containing compounds can be achieved by general NMR methods suitable for carbon-containing compounds; however, this statement is not correct. It should be noted that heterocycles have low densities of hydrogen and carbon atoms. Therefore, the application of conventional NMR methods (1D ^1H and ^{13}C spectroscopy, 2D HMQC, HMBC, INADEQUATE, etc.) might be ineffective for the structural estimation of poly-nitrogen compounds. Another important consequence of an increased nitrogen atom content is a decrease in the aromaticity and the tendency to undergo ring-chain transformations. Thus, the confirmation of the structure can be complicated by the azide–tetrazole equilibrium, ANRORC (Addition of Nucleophile, Ring Opening, Ring Closure) reaction, Dimroth rearrangement, etc., which are observed in the azolo and azine series under the action of nucleophilic reagents and solvents. In this case, researchers usually use X-ray crystallography or comparison of UV-vis and

^1H and ^{13}C NMR spectra with the data of model compounds with unambiguously confirmed structures. Unfortunately, the first approach gives information about the structure of compounds only in the solid state and is not suitable for the analysis of mixtures of compounds. The second method allows for the determination of structural characteristics in solution, but it can lead to incorrect conclusions.¹⁸ There are several approaches to solve these issues. For example, the use of 2D H–(C)–N multiple bond correlation (HNCMBC) experiments was described for the structural confirmation of *N*-alkylated azolo derivatives based on the natural ^{15}N abundance.^{19,20} However, these NMR procedures rely on magnetization transfer through ^{13}C – ^{15}N *J*-coupling, and although they allow the determination of some other structures of simple monocyclic azoles, they cannot be considered as general.

The selective ^{15}N -labeling of organic molecules leads to the appearance of additional ^1H – ^{15}N and ^{13}C – ^{15}N spin-spin coupling constants (SSCCs) that significantly expand the application of NMR methods in the determination of molecular structures. This approach is widely used in the chemistry of proteins and nucleic acids.^{21–24} Although ^{15}N -labeling and the subsequent analysis of J_{HN} and J_{CN} couplings in structural studies of nitrogen heterocycles have been described in a few articles, such a method can be effective for the determination of the structure and studying of the mechanism of chemical transformations of azoles, azines and their fused derivatives. Most of these articles were presented in early published review papers and books that described the highlight achievements in various areas of chemistry.

However, the incorporation of a ^{15}N atom into structures leads to the appearance of isotope shifts.²⁵ These data may only be considered as additional features to confirm the structure. The chemical shifts of labeled nitrogens in 1D ^{15}N NMR spectra

^aUral Federal University, 19 Mira Street, 620002 Yekaterinburg, Russian Federation. E-mail: deevsl@yandex.ru

^bI. Ya. Postovsky Institute of Organic Synthesis, 22 S. Kovalevskoy Street, 620219 Yekaterinburg, Russian Federation



can also be used as additional characteristics of enriched compounds.²⁶ However, information on chemical shifts is relative and can only be used in the context of the already obtained detailed data. In contrast, the measurement of ^{13}C - ^{15}N and ^1H - ^{15}N coupling constants provides unambiguous information about the molecular structures.

Despite the great opportunities for selective ^{15}N incorporation followed by an analysis of J_{HN} and J_{CN} couplings, no systematic review devoted to this tool for structural studies of heterocycles has been presented until today. This is the first attempt to generalize the literature data on ^{15}N -labeling and show the capacity of the usage of J_{HN} and J_{CN} couplings for the determination of the molecular structure and the chemical transformations of nitrogen-containing heterocycles. The relevance of the topic has been confirmed by a recently published article describing the Dimroth-type ring transformation of an azine fragment in the imidazo[1,2-*a*]pyrimidine scaffold, which is of significant interest for modern medicinal chemistry.²⁷ The authors showed the efficiency and universality of the use of ^{15}N -labeling and the analysis of ^1H - ^{15}N coupling constants for ring-chain transformations, which require more detail to determine the structures of heterocyclic compounds. It was this work that drove us to summarize the literature data on the incorporation of ^{15}N isotopes and the use of J_{HN} and J_{CN} couplings in the chemistry of heterocycles. Analysis of the literature showed that this approach is general and covers different series of heterocyclic systems.

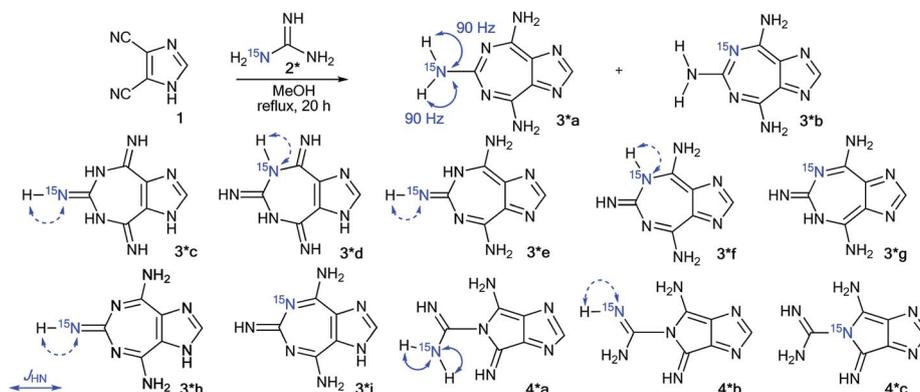
This review includes three sections. The first section describes the application of ^1H - ^{15}N and ^{13}C - ^{15}N constants for confirmation of the ways to fuse azole, azine and azepine rings during the synthesis of bicyclic, tricyclic and polycyclic compounds and the use of J_{HN} and J_{CN} in the studies of tautomeric rearrangements of heterocyclic derivatives. The next section reports the ^{15}N -labeling and analysis of ^1H - ^{15}N and ^{13}C - ^{15}N spin-spin interactions in the determination of the mechanism of ring transformation rearrangements. The last section includes information about the use of J_{HN} and J_{CN} couplings for the determination of the sites and mechanisms of interaction of heterocycles with electrophilic and nucleophilic reagents, which occurs without the transformation of the heterocyclic framework.

2. Use of J_{HN} and J_{CN} for establishing methods for the heterocyclization of azole, azine or azepine to azole and azine fragments and studying ring-chain tautomerism in a series of heterocycles

A combination of ^{15}N -labeling and an analysis of J_{HN} and J_{CN} couplings is one of the approaches for the confirmation of the structure of a fused heterocyclic system that can be formed by different methods for the cyclization of an azole, azine or azepine ring to various five- or six-membered heterocycles. Moreover, this method may be effective for the study of ring-chain tautomerisms that are observed in various classes of heterocycles.

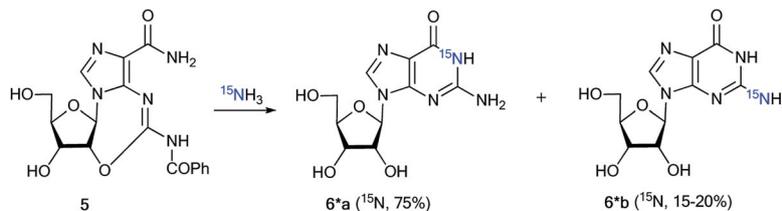
2.1. Determination of methods of heterocyclization based on an analysis of ^1H - ^{15}N and ^{13}C - ^{15}N spin-spin interactions

The measurement of ^{13}C - ^{15}N and ^1H - ^{15}N spin-spin interactions can be used for the determination of alternative heterocyclization pathways, including monocyclic derivatives. This method can be efficient even for the analysis of isotopomer mixtures. For example, the interaction of 4,5-dicyanoimidazole **1** with ^{15}N -guanidine **2*** led to compounds **3*a** and **3*b** (Scheme 1).²⁸ It should be noted that the reaction could give tautomers **3*a-i** and isomers **4*a-c**. Nonetheless, the registration of the signals of the two labeled nitrogen nuclei as a triplet and singlet in the ^{15}N NMR spectrum provided solid evidence of the formation of isotopomers **3*a** and **3*b** and excluded the formation of tautomers **3*c-i**. Furthermore, structure **3*a** was confirmed by observation of the single ^1H - ^{15}N spin-spin coupling ($J_{\text{HN}} = 90$ Hz) between a ^{15}N atom and the protons of an NH_2 group in the ^1H NMR spectrum. If a mixture of compounds **4*a-c** were produced, additional J_{HN} splitting for the ^{15}NH imino group would appear in both the ^1H and ^{15}N NMR spectra. However, such ^1H - ^{15}N SSCCs were absent, which excluded the formation of **4*a-c**. This result also showed that the measurement of J_{HN} couplings can be effective in the



Scheme 1 Establishing the structure of the condensation product of 3,4-dicyanoimidazole with guanidine. The observed J_{HN} couplings in the ^1H NMR spectra are shown by blue arrows. The expected couplings (J_{HN}) in the excluded structures are indicated by dashed arrows.





Scheme 2 Synthesis of ^{15}N -labeled guanosines **6*a** and **6*b** using $^{15}\text{NH}_3$.

determination of the structure of prototropic tautomers instead of an analysis of the chemical shifts of ^{13}C and ^{15}N atoms in the NMR spectra.^{29–31}

An analysis of direct ^{13}C – ^{15}N coupling constants allowed for the determination of the method of cyclization in the reaction of guanosine with glycidaldehyde.³² Two methods were used for the incorporation of ^{15}N atoms in the pyrimidine fragment of guanosine. The first procedure was based on the treatment of compound **5** with [^{15}N]-ammonia (Scheme 2). This approach gave an isotopomeric mixture containing 75% **6*a** and 15–20% **6*b**. Another synthesis of **6*a** and **6*b** that included obtaining the ^{15}N -labeled amide **5#** by the interaction of [^{15}N]-benzoyl isothiocyanate and imidazolecarboxamide derivative **7** is depicted in Scheme 3. The reaction of compound **5#** with ammonia yielded a mixture containing **6#a** (15–20%) and **6#b** (75%).

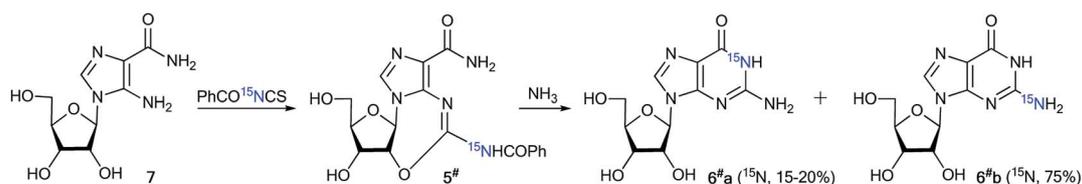
The use of the mixture of ^{15}N -guanosines **6*a**/**6#a** and **6*b**/**6#b** in a reaction with glycidaldehyde led to obtaining samples that had different concentrations of isotopomers **8*a** (^{15}N , 75%)/**8#a** (^{15}N , 15–20%) and **8*b** (^{15}N , 15–20%)/**8#b** (^{15}N , 75%) (Scheme 4). Differences in the enrichment of products **8*a**/**8#a** and **8*b**/**8#b** allowed for the direct measurement of ^{13}C – ^{15}N coupling constants ($^1J_{\text{C}7-\text{N}8}$ 9 Hz and $^1J_{\text{C}6-\text{N}5}$ 11 Hz) and the assignment of the C6 and C7 signals in the ^{13}C NMR spectra.

The values of $^1J_{\text{C}7-\text{N}8}$ and $^1J_{\text{C}6-\text{N}5}$ were obtained from the carbon spectra of compounds **8*a** and **8*b** with 75% ^{15}N enrichment. This characteristic permitted the determination of

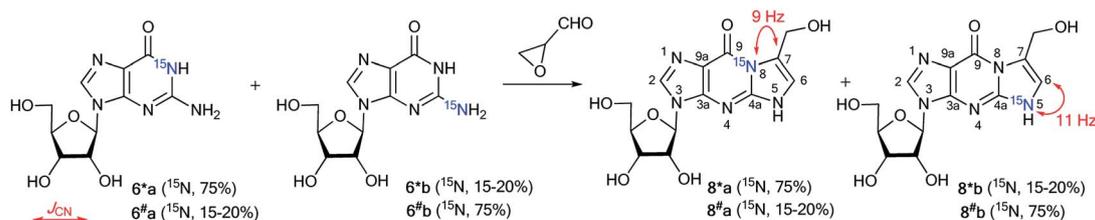
sites for the attachment of the hydroxymethyl group in compounds **8*a** and **8*b**. However, the alternative isomers **9*a**/**9#a** and **9*b**/**9#b** were not observed (Fig. 1).

The values of the ^1H – ^{15}N and ^{13}C – ^{15}N couplings can be applied as diagnostic features for confirmation of the ring closure of heterocycles. This approach was described in 1986 by Villarasa *et al.*, where a ^{15}N -labeled derivative of diazazole **11*** was prepared.³³ The synthesis comprises the diazotation of 2-aminoimidazole **10** with $\text{Na}^{15}\text{NO}_2$ having 25% ^{15}N enrichment (Scheme 5). The following reaction between compound **11*** and 1,1-dimethoxyethene **12** can lead to either open-chain azoalkene **13*** or fused azoloazine **14***. It should be noted that compound **14*** is the product of the cyclization of **13***. The observed $^1J_{\text{CN}}$ of 2.3 Hz and $^2J_{\text{HN}}$ of 14.2 Hz for the obtained compound were in good agreement with the NMR spectral data of pyridine and other azines. Thus, they confirmed the formation of bicycle **14*** in the reaction of imidazole **11*** with dimethoxyethene compound **12**.

The use of labeled azole derivatives in the synthesis of fused heterocycles is one method for the incorporation of ^{15}N atoms in the azoloazine series. Obtaining ^{15}N -azoles can require several steps. For example, the synthesis of [2- ^{15}N]-5-amino-1,2,4-triazole **18*** began by the interaction between labeled potassium nitrate (87%, ^{15}N) and guanidine sulfate **15** (Scheme 5).³⁴ Then, the N-nitrated product **16*** was reduced to ^{15}N -aminoguanidine bicarbonate **17***, which was transformed into [1- ^{15}N]-3-amino-1,2,4-triazole **18*** by reaction with formic acid.



Scheme 3 Synthesis of ^{15}N -labeled guanosines **6*a** and **6*b** using ^{15}N -benzoyl isothiocyanate.



Scheme 4 Reaction of guanosines **6*a** and **6*b** with glycidaldehyde. The observed $^1J_{\text{CN}}$ couplings in the ^{13}C NMR spectra are shown by red arrows.



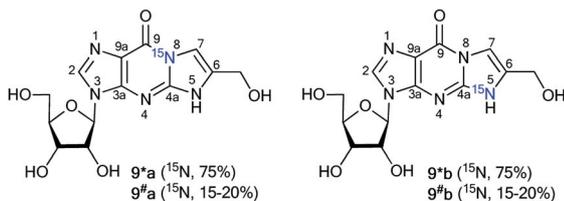
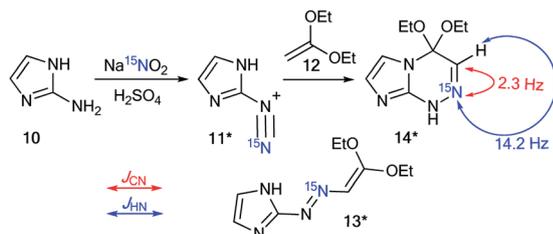


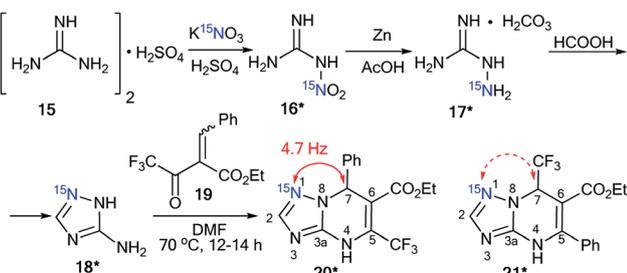
Fig. 1 Alternative isomers 9*a/9#*a and 9*b/9#*b.



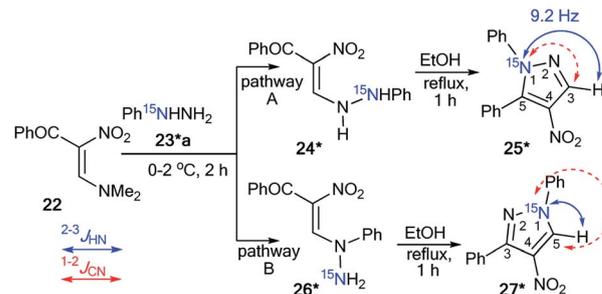
Scheme 5 The choice between open chain azoalkene 13* and fused azoloazine 14*. The observed $^1J_{CN}$ and $^2J_{HN}$ couplings are shown by red and blue arrows, respectively.

Compound 18* (~87%, ^{15}N) with one equivalent of unlabeled 3-amino-1,2,4-triazole was used in a condensation reaction with 2-benzylidene-2-fluoroacetyl ester 19 (Scheme 6).³⁵ The analysis of the J_{CN} couplings showed that a ^{13}C - ^{15}N spin-spin interaction was observed for the C7 atom bonded to a phenyl fragment ($J_{CN} = 4.7$ Hz). Moreover, a $^2J_{H2-N1}$ of 15.3 Hz was detected in the ^1H NMR spectrum of compound 20*. These data unequivocally prove the formation of structure 20* (~43%, ^{15}N). In the case of obtaining the alternative product 21*, ^{13}C - ^{15}N splitting should be observed for the signal of the carbon atom coupled with a CF_3 group.

The use of ^{15}N -phenylhydrazine 23*a in the interaction with nitroenamine 22 allowed for the determination of a reaction pathway that can be route A or B (Scheme 7).³⁶ According to pathway A, product 25* is formed through 24*, while the alternative pathway B involves the formation of compound 26* that transforms into resulting pyrazole 27*. The comparison of the detected J_{HN} of 9.2 Hz with the early described amplitudes of the ^1H - ^{15}N SSCCs for the pyrazole series showed that the observed coupling is a vicinal one. Thus, it was found that the reaction between 22 and 23*a gave compound 25*. Moreover,



Scheme 6 Structural elucidation of a series of 1,2,4-triazolo[1,5-a]pyrimidines. The observed $^2J_{CN}$ coupling is shown by a red arrow. The expected coupling ($^2J_{CN}$) in the excluded structure is indicated by a dashed arrow.



Scheme 7 Establishing the structure of isomeric nitropyrazole 25*. The observed J_{HN} couplings are shown by blue arrows. The expected but unobserved J_{CN} are couplings indicated by dashed red arrows.

the formation of azole 27* would lead to the appearance of $^1J_{C5-N1}$ with values of 8–11 Hz, whereas a carbon coupled with a hydrogen did not show the splitting of ^{13}C - ^{15}N due to the small amplitude of $^2J_{C3-N1}$ (>2 Hz). The weak $^{13}\text{C3-}^{15}\text{N1}$ constant was used as an additional criterion for the confirmation of the structure of 25*.

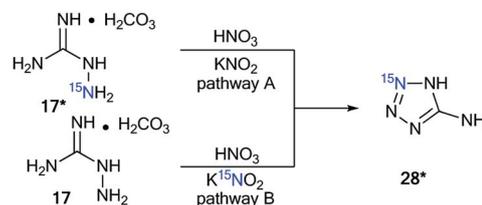
2.2. Study of ring-chain tautomerism using J_{CN} and J_{HN} coupling constants

Aminoguanidine bicarbonate 17* (^{15}N , 86%) was also applied as a labeled starting material for the incorporation of ^{15}N atoms in aminotetrazole 28* (pathway A, Scheme 8). The synthesis of 28* was based on the Thiele method,³⁷ which includes the interaction of aminoguanidine salts with nitro acid produced *in situ* from potassium nitrite and nitric acid.

The reaction of ^{15}N -potassium nitrite (^{15}N , 86%) with unlabeled aminoguanidine 17 is an alternative method that is suitable for obtaining compound 28* (pathway B, Scheme 8).

The use of ^{15}N -aminoguanidine bicarbonate 17* and ^{15}N -potassium nitrite allowed for the incorporation of two stable isotopes into aminotetrazole 28** (Scheme 9).³⁸

Compounds 28* and 28** were used for the synthesis of labeled tetrazolo[1,5-a]pyrimidines and tetrazolo[1,5-b][1,2,4]triazines. Because ^{15}N -aminotetrazole 28* exists in two tautomeric forms in solution, 28*a and 28*b, the reaction of this compound should result in the formation of a mixture of



Scheme 8 Synthesis of ^{15}N -labeled aminotetrazole 28*.



Scheme 9 Synthesis of ^{15}N double-labeled aminotetrazole 28**.

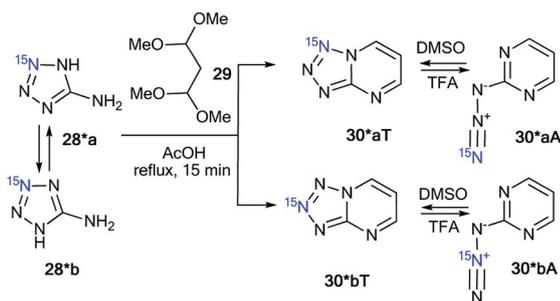


isotopomers (Scheme 10).³⁹ As expected, the condensation of tetrazole **28*** with 1,1,3,3-tetramethoxypropane **29** led to a mixture of tetrazolopyrimidines **30*aT** and **30*bT** in a 1 : 1 ratio.

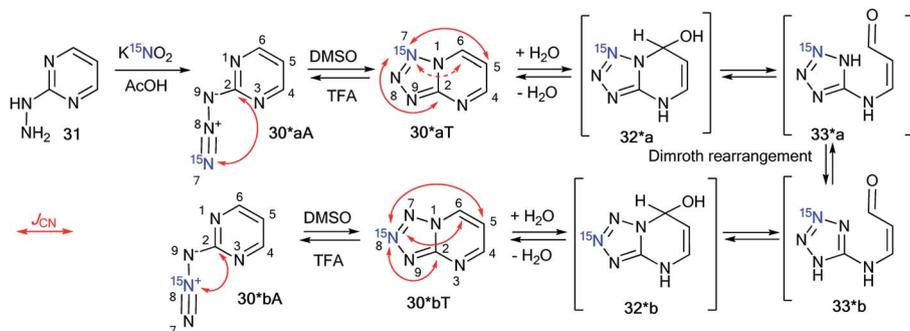
A similar result was obtained by the reaction of 2-hydrazinopyrimidine **31** with ¹⁵N-enriched nitrous acid that was generated from labeled potassium nitrite in acetic acid (Scheme 11).³⁷ A mixture of isotopomers **30*aT** and **30*bT** in a 5 : 1 ratio was obtained. The Dimroth rearrangement is one of the most likely sources of the observed isomerization.

The NMR spectra of the isotopomer mixture in DMSO-d₆ solution showed that the tetrazole forms **30*a,bT** prevail over the azides **30*a,bA**, which were detected in trace amounts (~5%). The detection of *J*_{CN} couplings for the C2, C5, and C6 nuclei (Scheme 11 and Table 1) proved the formation of a fused tetrazole (Scheme 10). Interestingly, small amounts of the azide forms **30*a,bA** (~5%) were detected in both the ¹³C and ¹H NMR spectra. Similarly, *J*_{CN} splitting, which was only observed for the C2 nucleus, confirmed the azide structure of **30*a,bA**. In TFA solution, the azide–tetrazole equilibrium was rapidly shifted toward the azides **30*a,bA**. As expected for the azide forms **30*a,bA**, the presence of ¹³C–¹⁵N *J*-couplings was detected only for the C2 nucleus (Scheme 11 and Table 2).

The cyclization of the diazonium salt **34***, derived from ¹⁵N-labeled aminotetrazole **28***, with ethyl α-formylphenylacetate **35** gave a mixture of tetrazolo[1,5-*b*][1,2,4]triazines **37*aT** and **37*bT** at a 1 : 1 ratio (Scheme 12).³⁷ Similarly, a 1 : 1



Scheme 10 The formation of two isotopomeric ¹⁵N-labeled azides **30*A** and tetrazoles **30*T** in the reaction of aminotetrazole **28***.



Scheme 11 The Dimroth rearrangement of tetrazolopyrimidines **30*aT** and **30*bT**. The azide–tetrazole equilibrium is shifted to the tetrazole/azide form in DMSO-d₆/TFA-d solution. The observed *J*_{CN} couplings from the ¹⁵N7 and ¹⁵N8 nuclei are shown by red arrows. The expected coupling (²*J*_{CN}) is indicated by a dashed arrow. The measured *J*_{CN} values are presented in Tables 1 and 2.

Table 1 *J*_{CN} couplings (Hz) of tetrazolo[1,5-*a*]pyrimidines and tetrazolo[1,5-*b*][1,2,4]triazines^a

Compound	Solvent	<i>J</i> _{C2-N7}	<i>J</i> _{C2-N8}	<i>J</i> _{C5-N7} / <i>J</i> _{C6-N7}	<i>J</i> _{C5-N8} / <i>J</i> _{C6-N8}
30*a,bT ^b	DMSO-d ₆	2.3	2.5	1.0/3.8 ^c	0.5/<0.3 ^d
37*a,bT	DMSO-d ₆	3.3	2.1	1.6	0.9
37*a,bT	TFA-d	ND ^e	ND ^e	1.6	0.7
37**T ^c	DMSO-d ₆	3.3	2.0	1.5	0.8
40*a,bT	DMSO-d ₆	3.0	2.2	1.1	1.0
40*a,bT ^f	TFA-d	~0.5	~0.5	~1.0	~1.0
44*T	DMSO-d ₆	3.1		1.4	
44*T	TFA-d	<0.3 ^e		1.7	

^a Unless otherwise stated, the listed *J*_{CN} values represent the average between two independent measurements using ¹³C line-shape analysis and amplitude-modulated spin-echo experiments. ^b The mixture of isotopomers **30*a/30*b** (5 : 1) synthesized by Scheme 11 was used for the *J*_{CN} measurements. ^c The *J*_{CN} value was measured only by ¹³C line-shape analysis. ^d The *J*_{CN} coupling constant was not detected, probably due to the low abundance of the **30*b** isotopomer. ^e Measurements of the *J*_{CN} couplings were impossible due to the significant broadening of the corresponding ¹³C resonance. ^f The *J*_{CN} values were estimated from the ¹³C line-shape analysis. Precise measurements were impossible due to the fast conversion of tetrazoles **40*a,bT** to azides **40*a,bA** in TFA solution.

isotopomeric mixture of tetrazolo[1,5-*b*][1,2,4]triazines **40*aT** and **40*bT** was obtained by the reaction of **34*** with ethyl cyanoacetate **38** (Scheme 12).

Compounds **37*a,bT** were only registered in the tetrazole form in DMSO-d₆. The observation of the ¹³C–¹⁵N splitting for the C2 and C5 carbon signals in the 1D ¹³C NMR spectra of **37*a,bT** unambiguously confirmed the [1,5-*b*] fusion between the tetrazole and 1,2,4-triazine rings.

A small amount of the azide forms **40*a,bA** (~4%) in DMSO-d₆ solution was detected in the ¹³C and ¹H NMR spectra for the sample that was obtained by the interaction of compounds **34*** and **38** (Table 2). The low concentration of the azide form did not allow for the detection or measurement of the corresponding *J*_{CN} couplings. This form was only characterized by a relatively large downfield shift of the C2 resonance. Moreover, the main tetrazole isomers **40*a,bT** were characterized by *J*_{CN} couplings for signals C2 and C5. These characteristics allowed for the determination of the type of fusion between the tetrazole and 1,2,4-triazine rings in heterocycles **40*a,bT** as [1,5-*b*].



Table 2 J_{CN} couplings (Hz) of azidopyrimidines and azido-1,2,4-triazines^a

Compound	Solvent	$J_{\text{C2-N7}}$	$J_{\text{C2-N8}}$
30*a,bA ^b	DMSO-d ₆	0.6 ^c	ND ^d
30*a,bA ^b	TFA-d	0.5	0.7
37*a,bA	DMSO-d ₆	ND ^e	ND ^e
37*a,bA	TFA-d	0.5	0.8
40*a,bA	DMSO-d ₆	ND ^f	ND ^f
40*a,bA	TFA-d	0.5	0.8
44*A	TFA-d	0.5	

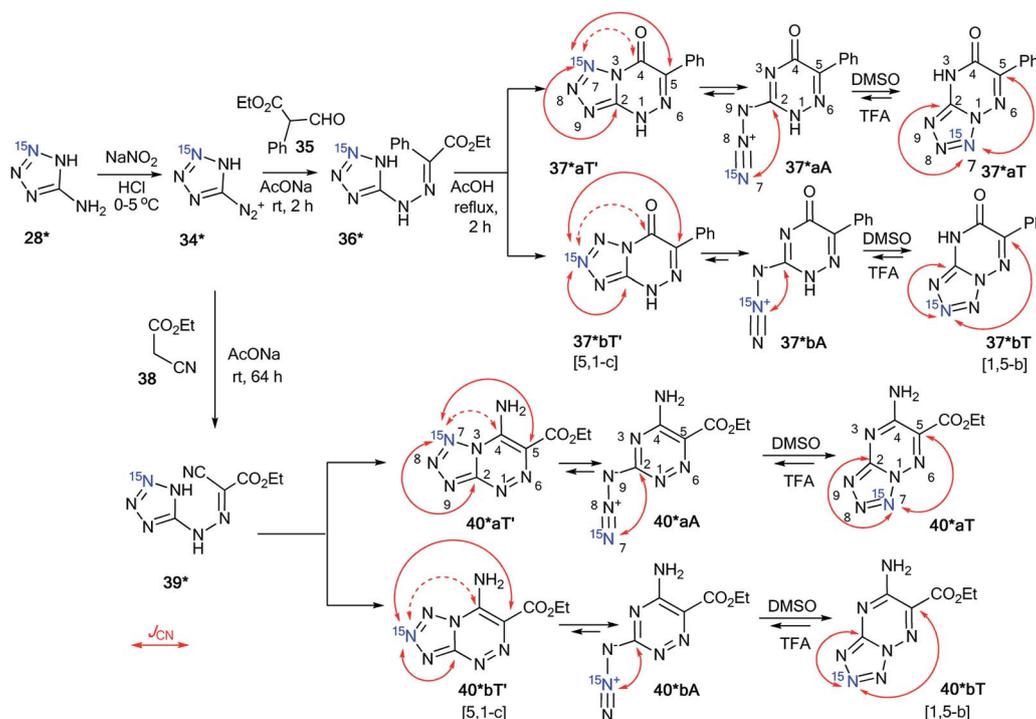
^a Unless otherwise stated, the listed J_{CN} values represent the average between two independent measurements using ¹³C line-shape analysis and amplitude-modulated spin-echo experiments. ^b The mixture of isotopomers 30*a/30*b (5 : 1) synthesized by Scheme 11 was used for the J_{CN} measurements. ^c The J_{CN} value was measured only by ¹³C line-shape analysis. ^d The measurement of the J_{CN} coupling was impossible due to the low abundance of the 30*b isotopomer. ^e Signals from the azide forms 37*a,bA were not observed in DMSO-d₆ solution. ^f The measurement of J_{CN} was impossible due to the low concentration of the azide forms 40*a,bA in DMSO-d₆ solution.

The NMR spectral parameters of compounds 37*a,bT were determined in TFA immediately after dissolving the tetrazole form and 30 days after the preparation of the solution. During this period of time, the relative population of the azide forms of 37*a,bA increased from 0% to 60%. Significant broadening of the C2 signal observed in the ¹³C spectra of 37*a,bT in TFA did not allow for the identification of the J_{CN} couplings from this carbon atom (Table 1). The ³ $J_{\text{C5-N7}}$ and ⁴ $J_{\text{C5-N8}}$ values for the C5

nucleus were measured easily. The obtained values of 1.6 and 0.7 Hz, respectively, correspond nicely to the J -couplings observed for the C5 carbon of 37*a,bT in DMSO-d₆ solution (1.6 and 0.9 Hz, respectively). This similarity confirms the retention of the bicyclic structure with a [1,5-*b*] type of fusion between the azole and azine fragments for 37*a,bT in TFA (Scheme 12). An analysis of the ¹³C multiplets of 37*a,bA in the 1D NMR spectra revealed the presence of J_{CN} interactions only for the C2 nucleus, which confirmed the azide structure of this compound (Table 2 and Scheme 12).

The rearrangement of 40*a,bT to 40*a,bA in TFA solution was relatively fast when compared to that of compounds 40*a,bT. The ¹³C NMR spectra were measured 1, 2 and 12 h after dissolving 40*a,bT in TFA, and the concentrations of the azide forms 40*a,bA were 18%, 40% and 97%, respectively. The J_{CN} interactions for the C2 and C5 carbon atoms of tetrazole 40*a,bT were observed in the ¹³C NMR spectra (Scheme 12 and Table 1), thus proving that the tetrazole structure was retained and that the [1,5-*b*] type of fusion for 40*a,bT occurred. The precise measurement of the J_{CN} values for 40*a,bT was impossible due to the fast conversion of tetrazoles to azides. A simplified line-shape analysis, however, revealed that the ² $J_{\text{C2-N7}}$ and ² $J_{\text{C2-N8}}$ interactions in 40*a,bT have significantly lower values in TFA than in DMSO-d₆ (Table 1). The azides 40*a,bA in TFA solution were characterized by the presence of ¹³C-¹⁵N J -coupling constants for only the C2 carbon (Scheme 12 and Table 2).

The use of double-labeled aminotriazole 28** is one way to avoid the formation of isotomeric mixtures. For example, the



Scheme 12 The azide–tetrazole equilibrium in the series of azido-1,2,4-triazines. The azide–tetrazole equilibrium is shifted to the tetrazole/azide form in DMSO-d₆/TFA-d solution. The observed J_{CN} couplings from the ¹⁵N7 and ¹⁵N8 nuclei are shown in red. The expected but unobserved couplings (² J_{CN} and ³ J_{CN}) are indicated by dashed arrows. The measured J_{CN} values are presented in Tables 1 and 2.

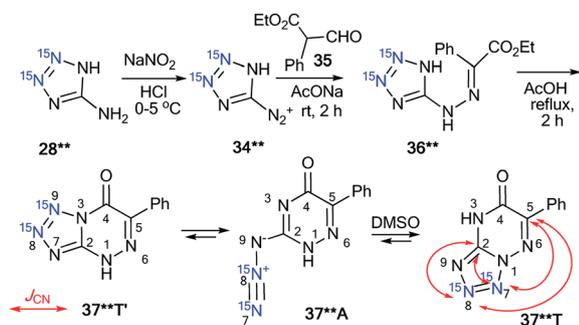


reaction of diazonium salt **34****, obtained from **28****, with ethyl α -formylphenylacetate **35** gave compound **36****, which underwent cyclization in acetic acid (Scheme 13).³⁹ As a result, $^{15}\text{N}_2$ -tetrazolo[1,5-*b*][1,2,4]triazine **37**T** was synthesized. An analysis of the J_{CN} couplings confirmed the type of fusion between the tetrazole and 1,2,4-triazine rings in compound **37**T** (Table 1 and Scheme 13).

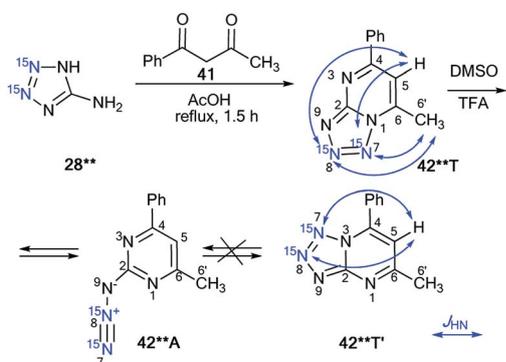
A similar approach was applied for the incorporation of two ^{15}N atoms in the structure of tetrazolo[1,5-*a*]pyrimidine derivatives. The interaction of **28**** with benzoylacetone **41** led to product **42**T** (Scheme 14).³⁸ The structure of compound **42**T** in DMSO- d_6 was determined by the analysis of the long-range ^1H - ^{15}N coupling constants, which were measured by spin-echo experiments with the selective inversion of ^{15}N nuclei. The observed $^nJ_{\text{HN}}$ values are presented in Table 3. The $^4J_{\text{H6}'-\text{N7}}$ and $^5J_{\text{H6}'-\text{N8}}$ couplings can only be observed in the proton spectrum of compound **42**T** (Scheme 14). In the case of the formation of tetrazoloazine **42**T'**, these spin-spin interactions should be absent.

This result indicated that the analysis of $^4J_{\text{HN}}$ and $^5J_{\text{HN}}$ can be used for the determination of the method of fusion between azole and azine fragments in tetrazolo[1,5-*a*]pyrimidines.

It was also found that tetrazole isomer **42**T** underwent complete rearrangement into the azide form **42**A** in TFA



Scheme 13 The azide-tetrazole equilibrium in double-labeled 1,2,4-triazine **37****. The observed ^{13}C - ^{15}N coupling constants in DMSO- d_6 solution are shown by red arrows.



Scheme 14 The azide-tetrazole equilibrium in double-labeled pyrimidine **42****. The observed long-range ^1H - ^{15}N J coupling constants ($^{4-5}J_{\text{H6}'/\text{H5}-\text{N7}/\text{N8}}$) are shown by blue arrows. The measured J_{HN} values are presented in Table 3.

solution. This was confirmed by the disappearance of the long-range ^1H - ^{15}N 7/8 correlations in the proton spectrum.

In contrast to 2-hydrazinopyrimidine **31**, the reaction of compound **43** with ^{15}N -potassium nitrite (^{15}N , 86%) in phosphoric acid led to the single isotopomer **44**T** (Scheme 15).³⁷ Registration of the 1D ^{13}C NMR spectra of **44**T** in DMSO- d_6 showed J_{CN} splittings for the signals from the C2 and C5 nuclei (Scheme 15 and Table 1).

These spin-spin interactions confirmed the formation of a cyclic structure with a [1,5-*b*] type of fusion between the tetrazole and 1,2,4-triazine fragments. If the alternative isomer **44**T'** was present, then additional J_{CN} splitting for the C4 signal should be observed in the carbon spectrum (Scheme 15). The azide-tetrazole equilibrium was shifted to the azide form **44**A** in TFA solution. The rearrangement of tetrazole **44**T** to the azide did not occur quickly.

The NMR spectra for compound **44**T** were measured in TFA-*d* solution immediately after dissolving the tetrazole form, after 16 days, and again after 30 days. During this period, the relative population of isomer **44**A** increased gradually from 0% to 87%. An analysis of the J_{CN} couplings allowed for the detection of both **44**T** and **44**A** in TFA-*d* (Tables 1 and 2).

Similar to compound **43**, the treatment of hetarylhydrazines **45** and **47** with labeled sodium nitrite (^{15}N , 98%) in acetic acid selectively yielded compounds **46**A** and **48**A**, respectively (Schemes 16 and 17).⁴⁰ The selective incorporation of a ^{15}N label into the azolo core of the tetrazoloazine leads to the appearance of long-range ^1H - ^{15}N J couplings, which can be easily measured in a quantitative fashion using 1D spin-echo experiments.

Compound **46**** in DMSO- d_6 underwent a transformation to an isomeric mixture of 1,2,4-triazine derivatives **46**T'**, **46**A** and **46**T** in a ratio of 79 : 12 : 9, respectively. The addition of TFA-*d* to DMSO- d_6 solution led to an increase in the relative concentration of the minor forms A and T.

In pure TFA, compound **46**** underwent an almost complete rearrangement to yield the open-chain azide **46**A**. In this case, the minor isomer **46**T'** was also found with a concentration of approximately 1%. The measurement of $^4J_{\text{HN}}$ for tetrazole **46**T'** was possible in DMSO- d_6 , TFA-*d* and different mixtures of these solvents (Table 3). The detection of $^{15}\text{N7}$ - $^1\text{H13}$ and $^{15}\text{N7}$ - $^1\text{H12}$ spin-spin interactions provides evidence for the [5,1-*c*] type of fusion between the azole and azine rings in **46**T'**. It should be noted that other protons signals (H11 and H12) can have $J_{\text{H-N7}}$ couplings. The disappearance of the $J_{\text{H13-N7}}$ and $J_{\text{H12-N7}}$ couplings and the measurement of the values of $J_{\text{H10-H7}}$ and $J_{\text{H11-N7}}$ allowed the confirmation of the structure of **46**T** (Table 3 and Scheme 16).

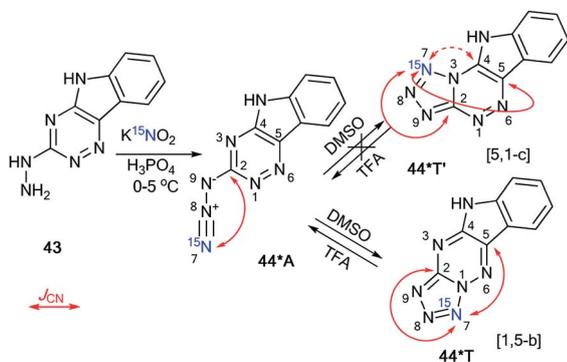
Two tetrazole forms, **48**T'** and **48**T**, in a ratio of 1 : 1 were detected upon dissolution of **48**A** in DMSO- d_6 . The signals from azide **48**A** in the NMR spectra became detectable only in the mixture DMSO- d_6 /TFA-*d* at a concentration of the acid of more than 50%. For example, in DMSO- d_6 /TFA-*d* (3 : 1) solution, a 24 : 29 : 47 concentration ratio of **48**T'** : **48**A** : **48**T** was found. In TFA-*d*, a mixture of the isomers **48**A** and **48**T** (96 : 4) was obtained. The observed $J_{\text{H5-N7}}$ and $J_{\text{H6}'-\text{N7}}$ patterns enable the unambiguous determination of the cyclization method of the azole fragment in tetrazolopyrimidines **48**T'** and **48**T** in



Table 3 Long-range J_{HN} couplings (Hz) of tetrazolo[1,5-a]pyrimidines and tetrazolo-1,2,4-triazines^a

Compound	Solvent	$J_{\text{H5-N7}}/J_{\text{H10-N7}}$	$J_{\text{H5-N8}}/J_{\text{H11-N7}}$	$J_{\text{H6-N7}}/J_{\text{H12-N7}}$	$J_{\text{H6-N8}}/J_{\text{H13-N7}}$
42**T	DMSO-d ₆	0.20	0.82	0.09	0.08
46*T	DMSO-d ₆ /TFA-d (3/1)	0.06	0.06		
46*T'	DMSO-d ₆	0.11	0.04	0.04	0.15
46*T'	DMSO-d ₆ /TFA-d (3/1)	0.11	0.04	0.04	0.16
46*T'	TFA-d	0.13	ND ^b	ND ^b	0.17
48*T	DMSO-d ₆	0.6		0.2	
48*T	DMSO-d ₆ /TFA-d (2/3)	0.6		0.2	
48*T	TFA-d	0.7		0.2	
48*T'	DMSO-d ₆	0.6			
48*T'	DMSO-d ₆ /TFA-d (2/3)	0.6			

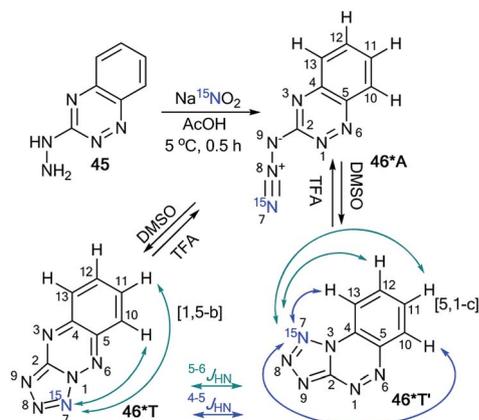
^a The ^1H - ^{15}N J coupling constants (J_{HN}) were measured by amplitude modulated spin-echo experiments. ^b Not determined due to overlap.



Scheme 15 The azide–tetrazole equilibrium of 1,2,4-triazine **44***. The azide–tetrazole equilibrium is shifted to the tetrazole/azide form in DMSO-d₆/TFA-d solution. The observed J_{CN} couplings from the ^{15}N nucleus are shown in red. The expected but unobserved coupling ($^2J_{\text{CN}}$) is indicated by a dashed arrow. The measured J_{CN} values are presented in Tables 1 and 2.

DMSO-d₆, TFA-d and a mixture of DMSO-d₆/TFA-d (Table 3 and Scheme 17).

The treatment of nitrobenzofuroxan **49** with ^{15}N -nitric acid (^{15}N , 98%) gave enriched compound **50*** (Scheme 18).⁴¹ The incorporation of the isotope label in the 6-position of

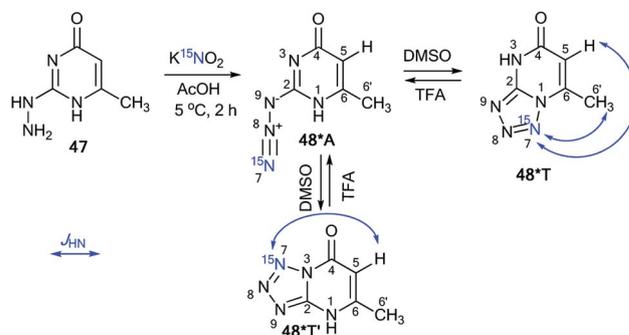


Scheme 16 The azide–tetrazole equilibrium of azido-1,2,4-triazine **46***. The observed ^1H - ^{15}N J coupling constants (J_{HN}) with magnitudes of $J \geq 0.1$ Hz and $J < 0.1$ Hz are indicated by the blue and green arrows, respectively.

dinitrobenzofuroxan **50*** permitted the determination of the specificity of the reaction with indene **51**. The appearance of $^3J_{\text{H5-N6}} = 1.9$ Hz, $^3J_{\text{H7-N6}} = 3.1$ Hz and $^3J_{\text{H1'-N6}} = 7.1$ Hz in the proton spectrum showed that the expected σ -adduct **52*** underwent a transformation into N-oxide **53***. If the isomerization of **52*** into **53*** had not occurred, the $^3J_{\text{H1'-N6}}$ coupling would have been absent.

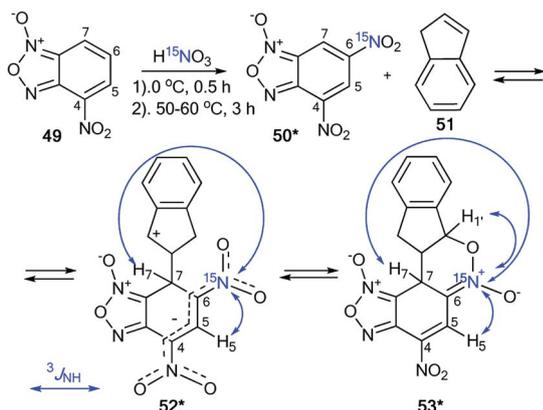
The incorporation of the ^{15}N isotope in structure **55*** was achieved by the nitration of compound **54** with labeled nitric acid (Scheme 19).⁴² The appearance of the ^{15}N atom provided the opportunity to study the interaction of **55*** with sulfite ions (Scheme 19). It was found that the initial reaction occurs at the 5-position to give the σ -adduct **56*a**, which was characterized by a geminal ^1H - ^{15}N coupling constant of 2.0 Hz. Then, isomerization of intermediate **56*a** occurred to yield adduct **56*b** carrying a sulfite at the 7-position. $^4J_{\text{H7-15N}} = 1.2$ Hz and $^3J_{\text{H6-15N}} = 1.2$ Hz were observed in the proton spectrum of compound **56*b**. These ^1H - ^{15}N coupling constants showed that the authors observed the intramolecular Boulton–Katritzky rearrangement rather than the SO_3^{2-} transfer, which should lead to the formation of compound **56*c**.

The use of ^{15}N -labeled compounds allowed the examination of the interaction of 3-aryl-5-methylisothiazoles with **57a-f** with aromatic nitriles in the presence of lithium diisopropylamide (LDA) (Scheme 20). The reaction between compounds **57b,c,e,f** and nitriles with different aromatic fragments ($\text{Ar}_1 \neq \text{Ar}_2$)

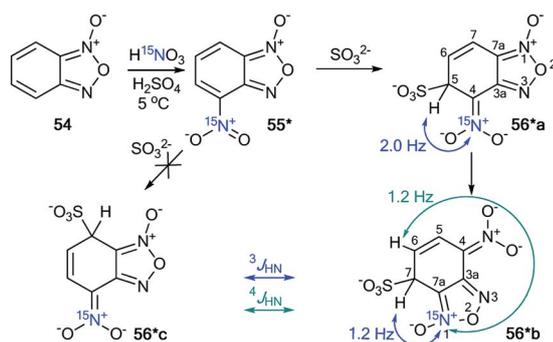


Scheme 17 The azide–tetrazole equilibrium in pyrimidine **48***. The observed ^1H - ^{15}N J coupling constants are indicated by the blue arrows.





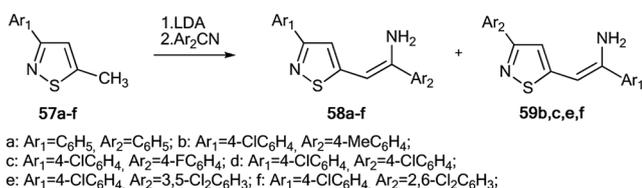
Scheme 18 The study of the interaction between dinitrofurazone 50* and indene using a ^{15}N label. The observed ^1H - ^{15}N J coupling constants are indicated by the blue arrows.



Scheme 19 The study of the interaction between nitrofurazone 55* and SO_3^{2-} using a ^{15}N label. The observed $^3J_{\text{HN}}$ and $^4J_{\text{HN}}$ couplings are shown by blue and green arrows, respectively.

afforded the formation of a mixture of isomers 58b,c,e,f and 59b,c,e,f. In the cases where $\text{Ar}_1 = \text{Ar}_2$, the reaction gave a single product.⁴³ The application of ^{15}N -enriched *p*-chlorobenzonitrile in these experiments allowed the authors to study the structures of the obtained compounds and the mechanism of formation of these products.

The treatment of 3-(4-chlorophenyl)-5-methylisothiazole with LDA and ^{15}N -labeled *p*-chlorobenzonitrile afforded a mixture of isotopomers 58*d and 59*d containing ^{15}N atoms in the amino group and thiazole fragment, respectively (Scheme 21). The positions of the isotope label in heterocycles 58*d and 59*d were provided by the measurement of the corresponding direct ^{13}C - ^{15}N coupling constants (11.7 Hz and 6.9 Hz for 58*d and 59*d, respectively). The value of $^3J_{\text{HN}} = 4.3\text{ Hz}$ for the vinyl



Scheme 20 Reaction between 3-aryl-5-methylisothiazoles 57a-f and aromatic nitriles.

proton in isotopomer 58*d showed that the obtained compound had a *trans* geometry with respect to the two aromatic rings Ar_1 and Ar_2 .

For an explanation of the formation of the isotopomeric mixture 58*d and 59*d, the selectively labeled compound 58*d was prepared, and the ring-chain transformation for this labeled structure was studied by NMR. Compound 58*d was obtained by treatment of 60 with ^{15}N -labeled *p*-chlorobenzonitrile.

This procedure led to compound 61*, which then underwent desilylation to form isotopomer 58*d (Scheme 22). Compound 60 was obtained by the interaction of methylisothiazole with *tert*-butyldimethylsilyl chloride in the presence of LDA. Then, heating the C_6D_6 solution of 58*d at 50°C for 50 h resulted in an equilibrium of a 1 : 1 mixture of isotopomers 58*d and 59*d.

Hence, two possible mechanisms (A and B) for the reversible equilibrium between 58*d and 59*d were presented (Scheme 23). It should be noted that the key intermediate in each path is sulfurane-2, 62*b. Pathway A involved the formation of sulfurane-1, 62*a, which transformed into structure 62*b via a [1,5]-sigmatropic hydrogen shift. Alternative pathway B suggests that thiazoles 63* and 64* are involved in the isomerization process.

Thus, the incorporation of ^{15}N labels and the analysis of the J_{CN} and J_{HN} couplings allow the determination of the formation mechanism of different heterocyclic systems and the observation of ring-chain tautomerism in a series of heterocycles.

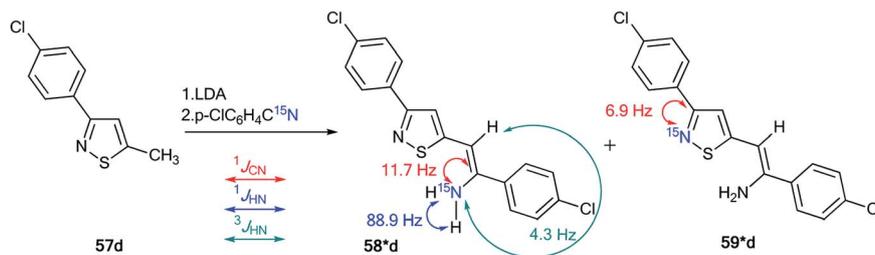
3. Analysis of J_{CN} and J_{HN} couplings as a method for the study of ring-chain transformations under the action of nucleophilic and electrophilic reagents

Some reactions of nitrogen heterocycles are simple at first glance, but they can hide a complex and completely unobvious mechanism. Isotopic labels allow a look deep inside into the details of such reaction mechanisms. Several heterocycles undergo ring transformations under the action of nucleophilic reagents (ANRORC, Dimroth rearrangement, *etc.*). Another type of recyclization is associated with intramolecular attack by the electron-deficient terminal atoms of the open-chain form on the ring system heteroatoms bearing a lone pair of electrons. It should be noted that ring opening or recyclization processes in both cases can occur and lead to the formation of a new type of heterocyclic structure. Additionally, determination of the structure of the obtained product always remains an important issue in organic chemistry. Moreover, solution of this task can help provide evidence for the mechanisms of ring-chain transformations.

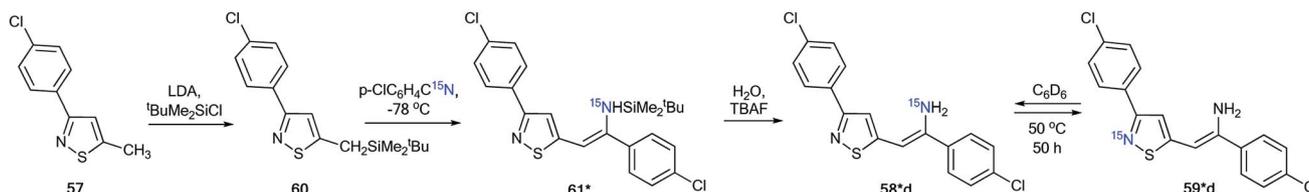
3.1. Study of ANRORC and Dimroth rearrangements and similar chemical transformations by analysis of J_{CN} and J_{HN} couplings

The analysis of J_{CN} and J_{HN} is an efficient method for studying different rearrangements that include ring-chain and ring

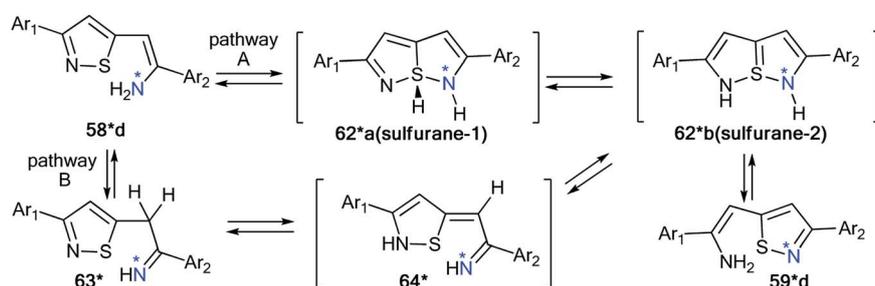




Scheme 21 Evidence of the formation of two isomeric 3-arylisothiazoles **58*d** and **59*d**. The observed $^1J_{CN}$, $^1J_{HN}$ and $^3J_{HN}$ couplings are shown by red, blue and green arrows, respectively.



Scheme 22 Selective synthesis of 3-arylisothiazole **58*** and ^{15}N label redistribution.

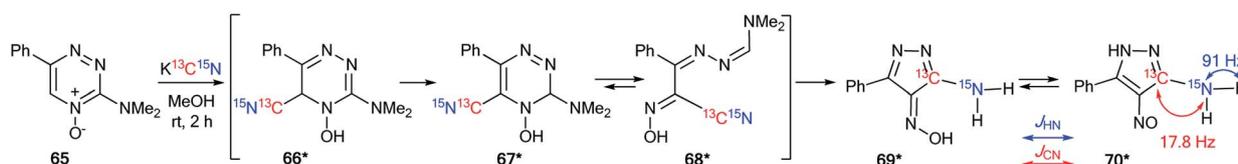


Scheme 23 The possible mechanism of the ^{15}N label redistribution in a series of isotope-enriched 3-aryl-5-enaminoisothiazoles.

opening transformations. It should be noted that the use of ^{15}N labeling and the estimation of ^{13}C - ^{15}N and ^1H - ^{15}N coupling constants are a single approach for the determination of one-pot cascade reactions; for example, the study of the ANRORC mechanism in the reaction of *N*-oxide-1,2,4-triazine **65** with potassium cyanide (Scheme 24).⁴⁴ The use of $\text{K}^{13}\text{C}^{15}\text{N}$ showed that the product of nucleophilic addition **66*** transformed into pyrazole **70*** through the formation of intermediates **67***-**69***. The structure of compound **70*** was confirmed by the measurements of $J_{CN} = 17.8 \text{ Hz}$ and $J_{HN} = 91 \text{ Hz}$. These characteristics show the unambiguous incorporation of the ^{13}C atom in the pyrazole ring of product **70***.

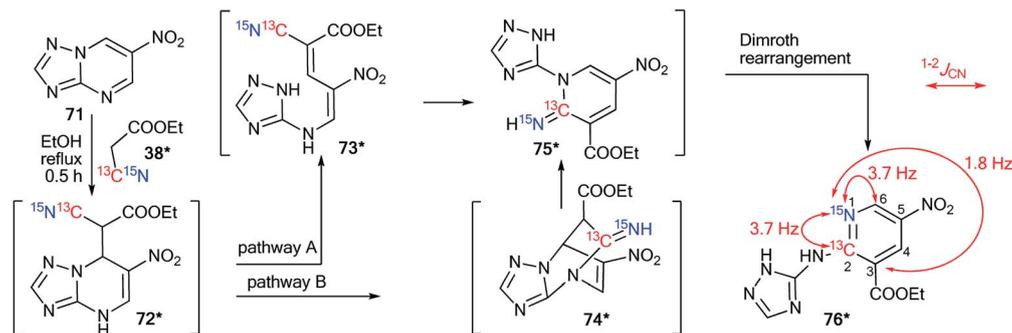
The application of double-labeled ^{13}C , ^{15}N -ethyl cyanoacetate **38*** permitted the detection of the Dimroth rearrangement for nitro-1,2,4-triazolo[1,5-*a*]pyrimidine **71** (Scheme 25).^{45,46}

It was found that ^{13}C , ^{15}N -ethyl cyanoacetate **38*** demonstrated binucleophilic properties in the reaction with 6-nitro-1,2,4-triazolo[1,5-*a*]pyrimidine **71**, and the mechanism of this reaction included several consecutive transformations (Scheme 25). The first step gave adduct **72***, which underwent isomerization into the open form **73*** (pathway A) or structure **74*** (pathway B). It should be noted that both intermediates **73*** and **74*** can transform into imine **75***. The following Dimroth rearrangement led to compound **76***. The usage of ^{13}C - and ^{15}N -



Scheme 24 The structural determination of the interaction products of 1,2,4-triazine oxide with potassium cyanide using ^{15}N and ^{13}C labels. The observed ^{13}C - ^{15}N and ^1H - ^{15}N coupling constants are shown by red and blue arrows, respectively.





Scheme 25 Establishment of the products and the mechanism of the reaction between nitrotriazolopyrimidine **71** and ethyl cyanoacetate. The observed ^{13}C – ^{15}N coupling constants are shown by red arrows.

labeled atoms and the analysis of ^{13}C – ^{15}N coupling constants allowed for the detection of the C– ^{13}C – ^{15}N –C fragment in the pyridine ring of molecule **76***. This result was confirmed by the registration of ^{13}C – ^{15}N splitting for signals C2 ($^1J_{\text{C-N}} = 3.7$ Hz), C6 ($^1J_{\text{C-N}} = 3.7$ Hz) and C3 ($^2J_{\text{C-N}} = 1.8$ Hz).

Recently, a representative study of a Dimroth-type ring-chain rearrangement in the imidazo[1,2-*a*]pyrimidine series using ^{15}N -labeled samples and an analysis of ^1H – ^{15}N couplings was described.²⁷

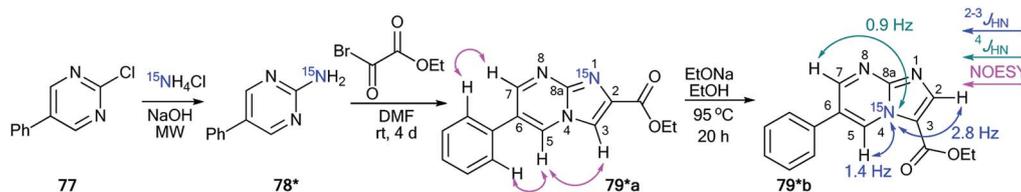
The incorporation of a ^{15}N atom in molecule **79*a** involved the substitution at C2 in compound **77** using $^{15}\text{NH}_4\text{Cl}$ under mildly basic conditions and cyclization of the obtained amine **78*** with ethyl bromopyruvate (Scheme 26). The structure of imidazo[1,2-*a*]pyrimidine **79*a** was determined by a 2D NOESY spectrum that showed a ^1H – ^1H NOE interaction of the H5 atom with the phenyl *ortho* protons and proton H3 of the imidazole fragment. The formation of isomer **79*b** was achieved by the treatment of compound **79*a** with sodium ethylate. Overall, azoloazine **79*b** containing a ^{15}N label in position 4 of the imidazo[1,2-*a*]pyrimidine scaffold was obtained. The appearance of the labeled nitrogen atom at the bridgehead position in compound **79*b** was confirmed by the additional ^1H – ^{15}N splitting for the signals of H2, H5 and H7 (2.8 Hz, 1.4 Hz and 0.9 Hz, respectively) in the 1D ^1H NMR spectrum. The values of the $^2J_{\text{HN}}$, $^3J_{\text{HN}}$ and $^4J_{\text{HN}}$ couplings were in good agreement with the data from the 2D ^1H – ^{15}N HMBC NMR experiment, which allowed the observation of same spin–spin interactions for heterocycle **79*b**. Thus, it was shown that structure **79*a** underwent a Dimroth rearrangement, which involves the addition of a hydroxide/alkoxide ion followed by ring opening (Scheme 27).

A similar result was obtained from the treatment of compound **79*a** with a solution of sodium hydroxide in a mixture of EtOH and THF. In this case, the rearrangement was accompanied by hydrolysis of the ester group.

An unusual transformation was found from the study of Vorbrüggen glycosylation in a series of aminopyrimidines **82a,b** and **89a–e** using ^{15}N -labeled reagents (Schemes 28 and 29).⁴⁷ It was revealed that the silylated 4-aminopyrimidines **83a,b** or **90a–e** after interaction with **84** yielded compounds **88a,b** and **94b–e**, which are products of the Dimroth rearrangement of nucleosides **85a,b** and **91a–e** in basic conditions. The mechanisms of these processes involve nucleophilic addition, ring opening and ring closure. Furthermore, the formation of structures **88a** and **94a–e** from **91a–e** presumes the nucleophilic substitution of a chlorine atom in open intermediates **92a–e**.

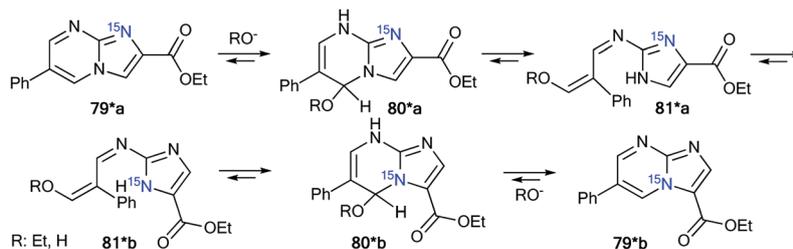
The synthesis of double-labeled diaminopyrimidine **82*b** and the use of this compound in the glycosylation reaction led to product **88*b** (Scheme 30). The observed $^1J_{\text{H8-N8}} = 93$ Hz, $^2J_{\text{H2-N1}} = 16$ Hz and $^3J_{\text{H7-N1}} = 3.9$ Hz unambiguously confirmed the structure of compound **88*b**. These characteristics showed that the pyrimidine derivative **88*b** was formed by the Dimroth rearrangement of the ribosylated azine **85*b**. The mechanism presented in Scheme 29 was confirmed by the application of ^{15}N -ammonium chloride in the process of the glycosylation of compound **89a** (Scheme 31). In this case, the incorporation of an isotope label in the pyrimidine ring was also observed. The values of the ^1H – ^{15}N coupling constants ($^2J_{\text{H2-15N}} = 15.1$ Hz and $^3J_{\text{H7-15N}} = 2.2$ Hz) unequivocally proved the incorporation of a ^{15}N atom into a pyrimidine fragment in compound **88*a**.

The application of ^{15}N -benzylamine (prepared by the reduction of $\text{PhCO}^{15}\text{NH}_2$) in the reaction with *N*-nitrouridine **96**

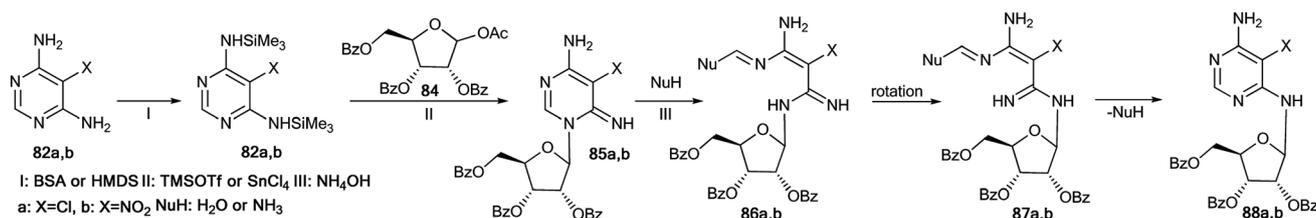


Scheme 26 The Dimroth rearrangement in a series of imidazo[1,2-*a*]pyrimidines. The measured $^3J_{\text{HN}}$ and $^4J_{\text{HN}}$ couplings in the 1D ^1H NMR spectrum of compound **79*b** are shown by blue and green arrows, respectively. The observed ^1H – ^1H NOE interactions between H-atoms in structure **79*a** are shown by magenta arrows.





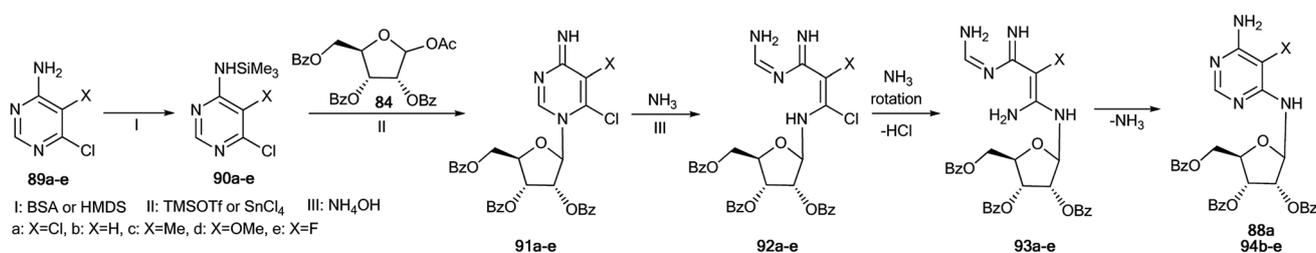
Scheme 27 Mechanism of the Dimroth rearrangement of compound 79*a to isomer 79*b.



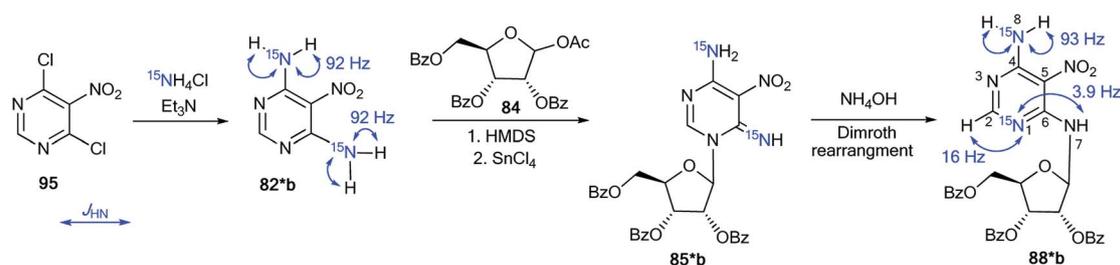
Scheme 28 The Dimroth rearrangement under the action of nucleophiles in a pyrimidine series accompanied by ribose migration.

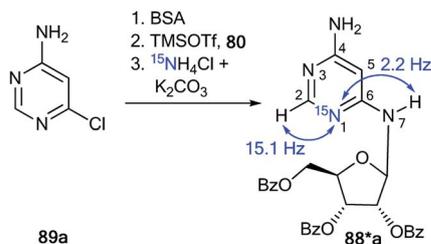
showed that the nucleophilic attack mainly occurs at the C4 atom of the uridine ring.⁴⁸ Moreover, this interaction afforded ring-chain intermediate 97*, which transformed into compound 98* in the presence of K₂CO₃ (Scheme 32). The production of structure 97* was confirmed by analysis of the ¹³C–¹⁵N coupling constants. Indeed, the *J*_{CN} splittings were only detected for C4 (*J*_{CN} = 16.4 Hz) and C5 (*J*_{CN} = 7.7 Hz) atoms, which unambiguously proved the production of the ring opening product 97*. The formation of intermediate 97* was monitored by an NMR experiment in CDCl₃ solution. This reaction is one example that allows the incorporation of a ¹⁵N-labeled atom in the core of a uridine derivative.

A previously mentioned article⁴⁸ also describes the study of the features of the interaction of *N*-nitroinosine with an amine by using ¹⁵N-labeled benzylamine (Scheme 33). The reaction of 99 with PhCH₂¹⁵NH₂ was monitored by NMR in CDCl₃ solution. It was shown that the attack of the amine takes place at the C2 atom of the inosine nitro derivative 99. This process leads to the ring opened intermediate 100*, which may follow two pathways, either 101*, or, in the presence of an excess of ¹⁵N-labeled benzylamine, compound 102*. In the 1D ¹³C NMR spectrum of 100*, the interaction ¹³C–¹⁵N was only observed for signal C2 (*J*_{CN} = 18.3 Hz) and the carbon atom of the benzyl fragment (*J*_{CN} = 9.8 Hz). The signal of C6 did not show any ¹³C–¹⁵N

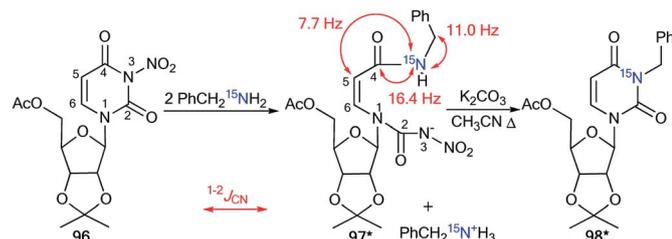


Scheme 29 The Dimroth rearrangement under the action of ammonia in the pyrimidine series accompanied by ribose migration.

Scheme 30 The Dimroth rearrangement in double-labeled nitroindine. The observed ¹H–¹⁵N coupling constants are shown by blue arrows.



Scheme 31 The ribosyl migration during the Dimroth rearrangement. The observed ^1H - ^{15}N coupling constants are shown by blue arrows.

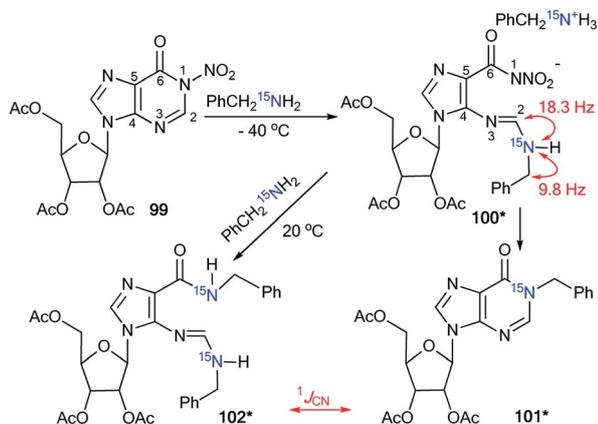


Scheme 32 The incorporation of ^{15}N atom into the structure of the uridine derivative. The observed $^{1-2}J_{\text{CN}}$ couplings are shown by red arrows.

splitting. Thus, analysis of the J_{CN} couplings allowed the straightforward determination of the structure of intermediate **100***.

The measurement of direct ^{13}C - ^{15}N SSCCs permitted the identification of the conversion of thiadiazoles **106*a-c** and **107*a-c** into 1,2,4-triazoles **108*a-c** and **109*a-c**, respectively (Scheme 34).⁴⁹ This work is one example of the successful use of ^{15}N -labeled samples for the investigation of the Dimroth-like rearrangement.

The three differently ^{15}N -enriched 1,4-diphenylthiosemicarbazides **103*a-c** in reaction with two different aroyl chlorides **104** and **105** were used as labeled starting materials. Thiosemicarbazide **103*a** was synthesized by treatment of



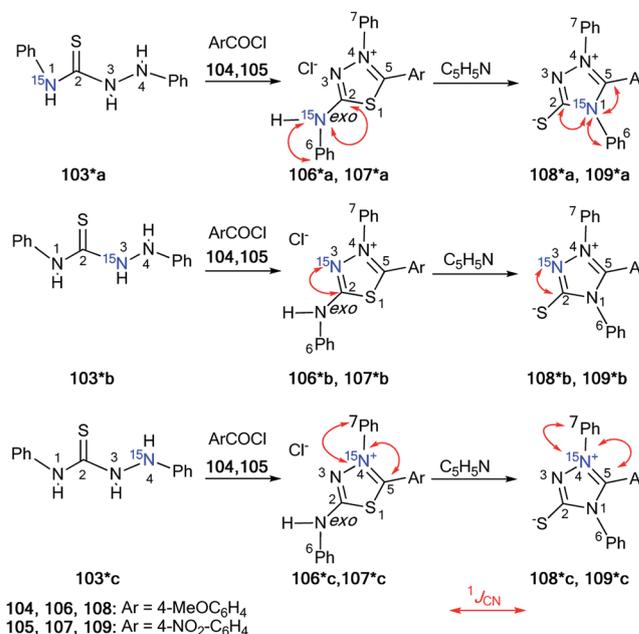
Scheme 33 The incorporation of ^{15}N atom into the structure of a hypoxanthine derivative. The observed $^1J_{\text{CN}}$ couplings are shown by red arrows.

phenylhydrazine with ^{15}N -phenyl isothiocyanate, which was the product of the interaction between ^{15}N -aniline, carbon disulfide and ethyl chloroformate (equivalent quantities). To obtain compound **103*b/103*c**, it was necessary to use labeled hydrazine obtained by the diazotization of aniline/ ^{15}N -aniline with $\text{Na}^{15}\text{NO}_2/\text{NaNO}_2$ in the reaction with phenyl isothiocyanate and perform the subsequent reduction in a mixture of Na_2SO_3 - $\text{Na}_2\text{S}_2\text{O}_3$. The formation of structures **106*a-c/107*a-c** from compounds **108*a-c/109*a-c** was observed by refluxing in pyridine solution. The measured values of $^1J_{\text{CN}}$ for heterocycles **106*a-c** and **109*a-c** were collected in Table 4.

The main characteristics confirming the formation of structures **108*** and **109*** were the appearance of three direct ^{13}C - ^{15}N coupling constants that were observed in the carbon spectra of isotomers **108*a** and **109*a**. The starting heterocycles **106*a** and **107*a** were only characterized by two ^{13}C - $^{15}\text{N}^{\text{exo}}$ and ^{13}C - $^{15}\text{N}^{\text{exo}}$ spin-spin interactions. The detection of the $^1J_{\text{C5-N1}}$ couplings verified the transformation of compound **106*/107*** into **108*/109*** under reflux in pyridine.

3.2. Analysis of J_{CN} and J_{HN} couplings in the study of other ring-chain rearrangements and transformations occurring with changes to the heterocyclic scaffold

The analysis of the $^1J_{\text{CN}}$ value was used for the determination of the structure of the nitrosation product obtained from compound **110** (Scheme 35).⁵⁰ The data from the 1D ^1H and ^{13}C NMR spectra did not permit the identification of the ring opening of the pyrrole fragment in **110** under the action of sodium nitrite in acetic acid. The use of the ^{15}N -labeled sodium nitrite and analysis of the direct ^{13}C - ^{15}N coupling constant showed that the tetrazole derivative **111*** underwent



Scheme 34 The use of ^{15}N labels to establish the structure of triazoles and thiadiazoles. The observed $^1J_{\text{CN}}$ couplings are shown by red arrows. The measured $^1J_{\text{CN}}$ values are presented in Table 4.



Table 4 Observed direct ^{13}C - ^{15}N coupling constants for compounds 106–109*a–c

Compound	$^1J_{\text{CN}}$, Hz	
106*a	C2-N ^{exo}	23.1 ± 1.5
	C6-N ^{exo}	14.1 ± 1.5
106*b	C2-N3	<2
106*c	C5-N4	14.1 ± 1.5
	C7-N4	16.1 ± 1.5
107*a	C2-N ^{exo}	23.1 ± 1.5
	C6-N ^{exo}	14.1 ± 1.5
107*b	C2-N3	<1
107*c	C5-N4	16.1 ± 0.8
	C7-N4	16.1 ± 0.8
108*a	C2-N1	4.0 ± 1.5
	C5-N1	18.1 ± 1.5
108*b	C6-N1	16.1 ± 1.5
	C2-N3	3.0 ± 0.8
108*c	C5-N4	19.1 ± 0.8
	C7-N4	19.1 ± 0.8
109*a	C2-N1	4.0 ± 1.5
	C5-N1	20.1 ± 1.5
109*b	C6-N1	16.1 ± 1.5
	C2-N3	3.0 ± 0.8
109*c	C5-N4	20.1 ± 0.8
	C7-N4	18.1 ± 0.8

a transformation into the open form **112***. The conclusion was based on comparison of the ^{13}C - ^{15}N spin-spin interaction ($^1J_{\text{CN}} = 81$ Hz) that was observed in the carbon spectrum of compound **112*** to that ($^1J_{\text{CN}} = 77$ Hz) obtained for the earlier structure **113***.⁵¹

The use of ^{15}N -phenylhydrazine **23*b** in the reaction of 2,3-dihydrofuro[3,2-c]coumarin-3-one **114** is another example of the use of labeled compounds to study chemical conversions in a series of heterocyclic compounds (Scheme 36).⁵² The analysis of the ^1H - ^{15}N coupling constants allowed the determination of the positions of the labeled atoms in product **118*** and proved the mechanism of the transformation. In the proton spectrum of compound **118***, direct and long-range ^1H - ^{15}N spin-spin interactions were detected for two signals of ^{15}NH -groups ($^1J_{\text{HN}} = 89.7$ Hz, $^4J_{\text{HN}} \sim 4$ Hz and $^1J_{\text{HN}} = 93.4$ Hz, $^4J_{\text{HN}} \sim 4$ Hz). Moreover, additional splitting was observed for the protons from the $\text{CH}=\text{N}$ and $\text{NH}-\text{Ph}$ fragments. Thus, compound **118*** was formed by the double addition of $^{15}\text{NH}_2-\text{NH}-\text{Ph}$ (**23*b**) and the

elimination of aniline in the last step of the reaction. This mechanism involves the formation of intermediates **115***–**117*** (Scheme 36).

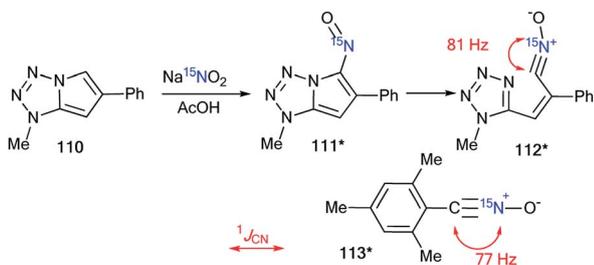
The revision of the result for the reduction reaction of diazonium salt **120** that was obtained by the diazotation of amine **119** was described in another work.⁵³ Previously, it was considered⁵⁴ that the product of this transformation is 1,2,3-triazine **121** (Scheme 37), while the determination of structure **121** was based on the detection of two direct ^1H - ^{15}N coupling constants ($^1J_{\text{H-N12}} = 107$ Hz and $^1J_{\text{H-N6}} = 97$ Hz) in the 1D ^{15}N NMR spectrum of the unlabeled sample. However, using ^{15}N -labeled diazonium salt **120*** in the reaction with sodium sulfite shed light on the real method of this transformation. The signal from the ^{15}N -nitrogen atom in the resulting compound was only characterized by one ^1H - ^{15}N coupling constant, $^2J_{\text{HN}} = 8.2$ Hz. Therefore, the 1,2,3-triazine structure of **121*** was rejected, and it was shown that the reduction of compound **120*** led to heterocycle **122***.

An unusual transformation was detected from the use of ^{15}N -enriched compound **125*** in a reaction with phenyltetrazole **126** (Scheme 38).⁵⁵ The synthesis of chloride **125*** included the interaction of benzoyl chloride and labeled ethylammonium chloride (^{15}N , 99%) in the presence of triethylamine in dichloromethane. Then, amide **124*** was treated with thionyl chloride. The interaction of **125*** and **126** led to heterocycles **129*** and **131***, which were characterized by ^{13}C - ^{15}N and ^1H - ^{15}N SSCCs.

These data confirmed the structures of compounds **129*** and **131***. In the proton NMR spectra of **129*** and **131***, the signals of the *N*-ethyl fragments showed additional splittings ($^2J_{\text{HN}} = 1.2$ Hz, $^3J_{\text{HN}} = 2.9$ Hz and $^2J_{\text{HN}} = 0.5$ – 1.0 Hz, and $^3J_{\text{HN}} = 3.2$ Hz, respectively). Moreover, in the ^{13}C NMR spectra, the signals of the carbons of the tetrazole and 1,2,4-triazine fragments were detected as doublets with magnitudes of 12.2 Hz and 12.1 Hz, respectively. These results indicate that the formation of structures **129*** and **131*** occurred over two different mechanisms (pathways A and B, Scheme 38). However, these transformations started with the common intermediate **127***. Pathway A included the elimination of benzonitrile, obtaining azide **128***, which underwent cyclization into tetrazole **129***. Compound **131*** may be formed according to the mechanism demonstrated by route B. In this case, structure **127*** transformed into **130*** by the elimination of nitrogen. Then, the isomerization of intermediate **130*** led to 1,2,4-triazine **131***.

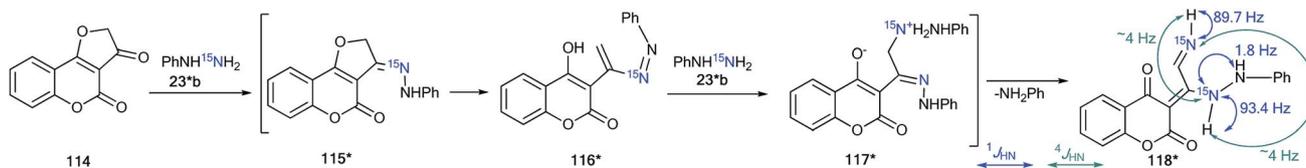
The incorporation of the ^{13}C and ^{15}N atoms in compound **136*** allowed determination of the method of the photo-sensitized oxidation of imidazole derivatives (Scheme 39).⁵⁶ This interaction with singlet oxygen was considered a model reaction for natural and biologically active structures containing the imidazole fragment (guanosine, xanthine, theophylline, histidine, etc.). The production of **136*** involved heating a mixture of $^{15}\text{N}_2$ -urea **132*** (^{13}C , 99% and ^{15}N , 98%) and ^{13}C -formic acid **133*** (^{13}C , 99%) at 150 °C for 4 h (Scheme 39). Then, ^{13}C -enriched benzoin **135*** was introduced, and the reaction mixture was heated at 180 °C.

The synthesis of labeled benzoin **135*** was based on coupling 2 mol of ^{13}C -benzaldehyde (^{13}C , 99%) in the presence of NaCN.

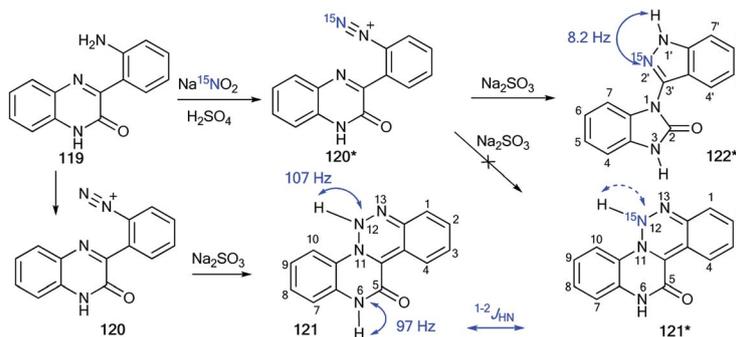


Scheme 35 Determination of the structure of the nitrosation product using a ^{15}N label. The observed $^1J_{\text{CN}}$ couplings are shown by red arrows.





Scheme 36 Determination of the positions of the ^{15}N atoms in 118^* . The observed direct and long-range ^1H - ^{15}N coupling constants are shown by blue and green arrows, respectively.



Scheme 37 Establishment of the correct structure of 122^* by using the ^{15}N label. The observed $^{1-2}J_{\text{HN}}$ couplings are shown by blue arrows. The expected but unobserved $^1J_{\text{HN}}$ coupling is indicated by a dashed blue arrow.

The photosensitized oxidation of $^{13}\text{C}_3$, $^{15}\text{N}_2$ -labeled 4,5-diphenylimidazole 136^* was carried out in an NMR tube. This experiment permitted the observation of the formation of several intermediates 137^* - 141^* that were obtained from 136^* under the action of singlet oxygen.

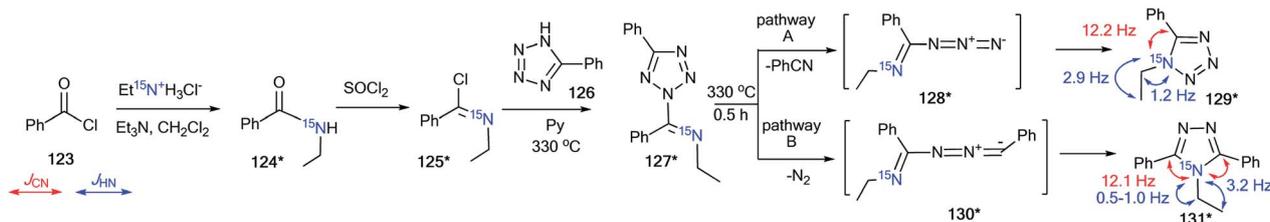
Oxidation of labeled imidazole 136^* gave one intermediate 137^* at -100°C . The observation of splitting of C2 ($^1J_{\text{C-N}1} = 5.1\text{ Hz}$ and $^1J_{\text{C-N}3} = 4.9\text{ Hz}$) showed that this carbon bonds with two nonequivalent, labeled nitrogen atoms. Moreover, a ^1H - $^{15}\text{N}1$ SSCC of 69.7 Hz was detected in the proton spectrum for this intermediate. These data and an analysis of the chemical shifts of the labeled atoms with the measurement of J_{CN} couplings confirmed the formation of 2,5-endoperoxide 137^* in the reaction of 136^* with singlet oxygen. The warming of 136^* led to decomposition, which could occur in two pathways: A and B (Scheme 39).

Both paths contained general intermediate 138^* , which was obtained by the transformation of 137^* . In the first pathway, 138^* lost water to give 139^* , which underwent hydrolysis to form 140^* . Another pathway included the transformation of 138^* into diol 141^* . Then, intermediate 141^* rearranged to 143^* by the opening and reclosing of the imidazole ring. This

transformation occurred through the open form 142^* . The formation of intermediates 138^* - 140^* and 141^* , 143^* was monitored by ^1H , ^{13}C and ^{15}N NMR spectroscopy.

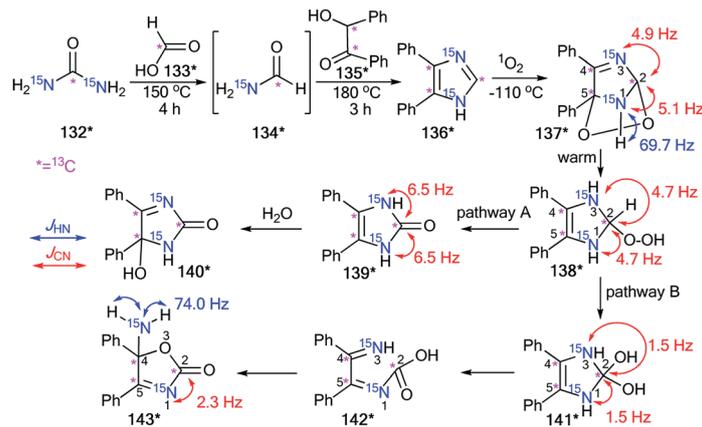
It should be noted that the estimation of the ^{13}C - ^{15}N and ^1H - ^{15}N coupling constants with analysis of the chemical shifts for ^1H , ^{13}C and ^{15}N NMR spectra and the measurement of the ^1H - ^{13}C spin-spin interaction allowed for the determination of the structures of these compounds. For example, the detection of only one $^1J_{\text{CN}} = 2.3\text{ Hz}$ for C2 in the carbon spectra of 143^* showed that one bond of C2- ^{15}N was broken. The chemical shift of C4 was characteristic of a sp^3 -carbon connected to two heteroatoms. These data indicated that C4 was bonded to an oxygen and the N1 nitrogen. Moreover, the signal from the amino group was observed as a triplet split by two bonded hydrogens ($^1J_{\text{H-N}1} = 74.0\text{ Hz}$) in the proton spectrum. This information suggested that the five-membered imidazole ring in 141^* opened and reclosed to form intermediate 143^* .

The application of ^{15}N -azirine 146^* (^{15}N , 50%) with one equivalent of unlabeled 2,2-dimethyl-3-(dimethylamino)-2H-azirine 146 provided an opportunity to illustrate the capacity of ^{15}N -labeling and ^1H - ^{15}N and ^{13}C - ^{15}N analysis in studying the mechanisms of chemical transformation in a series of three-



Scheme 38 Analysis of ^{13}C - ^{15}N and ^1H - ^{15}N SSCCs in the NMR spectra of ^{15}N -labeled products 129^* and 131^* to establish the reaction pathways. The observed ^{13}C - ^{15}N and ^1H - ^{15}N coupling constants are shown by red and blue arrows, respectively.





Scheme 39 The analysis of the ^{13}C - and ^{15}N -labeled intermediates for the choice between possible reaction pathways. The observed direct ^{13}C - ^{15}N and ^1H - ^{15}N coupling constants are shown by red and blue arrows, respectively.

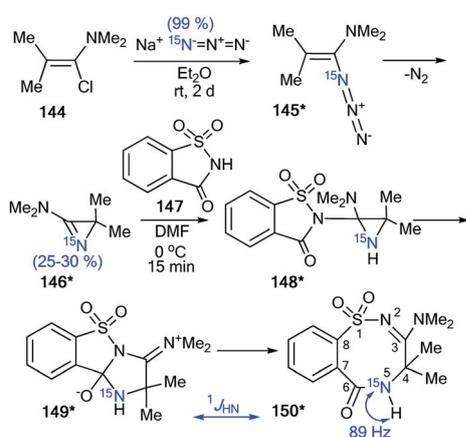
membered heterocyclic compounds (Schemes 40 and 41).⁵⁷ The synthesis of compound **146*** was based on the treatment of trimethyl-1-propenylamine **144** with enriched sodium azide (^{15}N , 99%). The interaction of **146*** (^{15}N , 25–30%) with 1,1-dioxo-1,2-benzothiazol-3-one (saccharine) **147** led to compound **150***, which was characterized by $^1J_{\text{HN}} = 89$ Hz (Scheme 40). This result confirmed the position of the ^{15}N -labeled atom in **150*** and showed that the transformation of **144*** into **150*** occurs *via* aziridine **148*** and ring expansion to zwitterion **149*** (Scheme 40).

In the case of the reaction between azirine derivative **146*** and phthalimide, the tricyclic product **153***⁵⁷ was isolated. The structure of **153*** was identified by measurements of ^{13}C - ^{15}N SSCCs ($^1J_{\text{C9b-N2}} = 7$ Hz and $^1J_{\text{C3-N2}} = 8$ Hz), which were registered for sp^3 -hybridized carbon atoms (Scheme 41). Additionally, the $^1J_{\text{HN}}$ -couplings were not detected in the 1D ^1H and ^{15}N NMR spectra. The authors suggested that compound **153*** is the product of the transformation of 2,3-dihydro-2,5-benzodiazocine-1,6-dione **152***. Moreover, the analysis of chemical shifts for the carbon atoms and ^{13}C - ^{15}N coupling constants of signals C2 ($^1J_{\text{CN}} = 5$ Hz) and C4 ($^1J_{\text{CN}} = 5$ Hz)

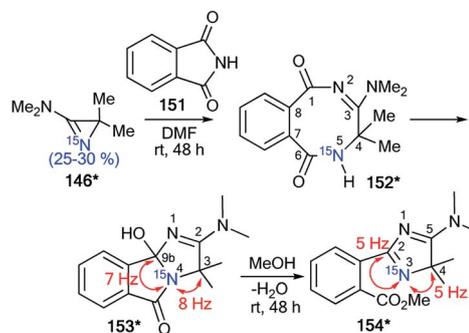
verified the formation 2-(4*H*-imidazol-2-yl)benzoic acid derivative **154***, which was obtained by treatment of **153*** with methanol.

An interesting example of the use of ^{15}N -labeled compounds for determining the ring open reaction pathway in a series of diazirines was reported by Creary and co-workers.⁵⁸ The incorporation of ^{15}N in structure **157*** was based on the reaction of methyl benzimidate **155** with enriched ammonium chloride (^{15}N , 99%) (Scheme 42). Then, oxidation of the resulting benzimidine **156*** with NaOBr yielded ^{15}N -phenylbromodiazirine **157***. Compound **157*** reacted with tetrabutylammonium azide to give a mixture of benzonitriles **160** and **160*** in a ratio of 1 : 1. The ^{13}C NMR spectrum confirmed partial ^{15}N labeling for the obtained product. The nitrile carbon of **160*** appeared as a doublet ($^1J_{\text{CN}} = 17.8$ Hz), while the carbon of the cyano group of **160** registered as a singlet. A comparison of the intensity of these signals showed a 50% excess of the ^{15}N isotope of **160***. This result proved that the formation compounds **160** and **160*** from diazirine **157*** and BuN^+N_3^- occurred *via* the structures **158*a,b** and **159*a,b**.

Thus, analysis of the literature shows that the measurement of the ^1H - ^{15}N and ^{13}C - ^{15}N coupling constants can be successful for the determination of different mechanisms of chemical

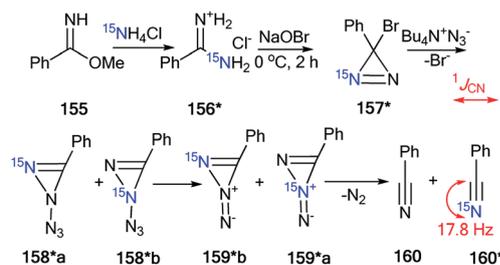


Scheme 40 Determination of the ^{15}N atom positions in **150***. The observed direct ^1H - ^{15}N coupling constant is shown by the blue arrow.



Scheme 41 Establishment of the reaction mechanism for the reaction of azirine **146*** with phthalimide **151**. The observed ^{13}C - ^{15}N coupling constants are shown by red arrows.



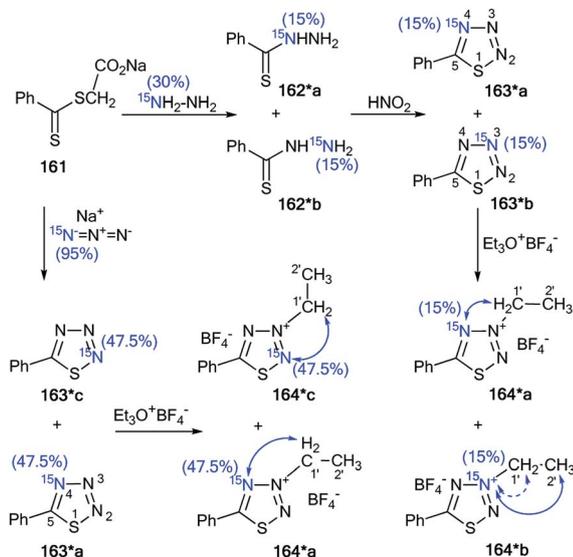


Scheme 42 The use of ^{15}N -labeled diazirine **157*** for the determination of the reaction pathway. The observed ^{13}C - ^{15}N coupling constant is shown by the red arrow.

transformations and rearrangements in nitrogen-containing heterocycles.

4. ^{13}C - ^{15}N and ^1H - ^{15}N coupling constants as an approach to the determination of structural reaction/isomerization products occurring without changes to the heterocyclic core

Nitrogen-enriched heterocycles are molecules containing several reaction centers. This class of compounds can exhibit dual behaviors, expressed by the fact that they can react with both nucleophilic and electrophilic reagents. As a result, such interactions often give a mixture of isomers even when the heterocyclic scaffold does not undergo rearrangement. This situation requires that researchers use different methods for



Scheme 43 The use of ^{15}N -labeled thiazotriazoles to determine the site of alkylation. The observed $^3J_{\text{H}1'-\text{N}2}$ couplings are shown by blue arrows. The expected but unobserved $^2J_{\text{H}1'-\text{N}2}$ coupling is indicated by a dashed blue arrow. The values of the observed vicinal ^1H - ^{15}N coupling constants are shown in Table 5.

the determination of structures obtained from similar products. One of the effective approaches that can be used to solve these problems is the measurement of the J_{CN} and J_{HN} couplings in ^{15}N -labeled samples. In this part of the review, examples of ^{15}N labeling and the use of the analysis of ^1H - ^{15}N and ^{13}C - ^{15}N coupling constants for the determination of functionalization methods that do not lead to changes in the heterocyclic core were collected. Such chemical transformations can be observed in reactions of *N*-alkylation, nitration, amination and obtaining azomethines based on different heterocycles.

4.1. Using J_{CN} and J_{HN} couplings for the investigation of *N*-alkylation reactions

The incorporation of ^{15}N atoms in different positions of 5-phenyl-1,2,3,4-thiazotriazole **163** allowed for determination of the structure of the products obtained by alkylation with triethylxonium tetrafluoroborate (Scheme 43).⁵⁹ The synthesis of isotopomers **163*a,b** was based on the reaction of a sodium salt of carboxymethyl dithiobenzoate **161** with ^{15}N -hydrazine (^{15}N , enrichment 30%) and the subsequent interaction with nitric acid. A mixture of 15% isotopically labeled compounds **163*a,b** was obtained.

Treatment of salt **161** with ^{15}N -labeled sodium azide (95%, ^{15}N) led to the formation of a mixture of compounds **163*a** (^{15}N , 47.5%) and **163*c** (^{15}N , 47.5%). The use of the mixture of **163*a** and **163*b/163*a** and **163*c** in a reaction with $\text{EtO}_3^+\text{BF}_4^-$ gave *N*-alkylated derivatives **164*a** and **164*b/164*a** and **164*c** (Scheme 43). The position of the alkylation of 1,2,3,4-thiazotriazole with ethyl fragments in **164*a-c** was determined by analysis of the ^1H - ^{15}N coupling constants. It should be noted that vicinal ($^3J_{\text{HN}}$) couplings were only registered in the proton spectra of **164*a-c**, and the expected geminal coupling ($^2J_{\text{HN}}$) was not detected by 1D ^1H NMR spectroscopy due its small amplitude (approximately 1.2 Hz). For compound **164*a/164*c**, $^3J_{\text{H}1'-\text{N}2}/^3J_{\text{H}1'-\text{N}2}$ was measured (Table 5). The detection of an additional splitting $^3J_{\text{H}2'-\text{N}3}$ (3.7 Hz) for the methyl signal of **164*b** unambiguously confirmed the structures of the products obtained by the interaction of compound **164*a-c** with $\text{EtO}_3^+\text{BF}_4^-$ because the formation of the alternative isomers **164*d,e** should lead to the appearance of $^3J_{\text{H}2'-\text{N}2}$ and $^3J_{\text{H}2'-\text{N}4}$ couplings, respectively (Fig. 2).

Moreover, the authors developed a selective synthesis for isotopomer **164*c**. This approach included the interaction of unlabeled 2-ethylthiobenzoylethylhydrazine **162** with ^{15}N -nitric acid. Then, the obtained **163*c** underwent alkylation with triethylxonium tetrafluoroborate (Scheme 44). The detection of $^2J_{\text{C}1'-\text{N}2} = 5.0$ Hz was further evidence for the binding of the *N*-ethyl fragment with the N2 atom in compounds **164*a-c**.

Another example of the application of ^{13}C - ^{15}N SSCC has been presented for the determination of the structures of *N*-alkylated derivatives **172*a** and **172*b** that were obtained by the interaction of ^{15}N -labeled 8-methylthioimidazo[4,5-*g*]quinazoline **171*** with benzyl bromide.⁶⁰ The synthesis of compound **171*** included the treatment of 7-chloro-4-quinazolone **165** with ^{15}N -nitric acid (^{15}N , 99%) (Scheme 45) to obtain ^{15}N -labeled product **166***. The remainder of the synthesis followed



Table 5 Vicinal ^1H - ^{15}N coupling constants measured in the 1D ^1H NMR spectra of **147* a-c**

Compound	$^3J_{\text{HN}}$, Hz
164* a	1.64 ± 0.05
164* b	3.74 ± 0.1
164* c	2.3 ± 0.05

Scheme 45 through heterocycles **167***, **168***, **169*** and **170***. The chosen method for the synthesis of **171*** determined the position of the ^{15}N -labeled atom.

The use of **171*** in the *N*-benzylation reaction led to the formation of two isomers, **172* a** and **172* b**, that were separated by column chromatography. The determination of the site of the *N*-alkyl fragments in compounds **172* a** and **172* b** was based on data from the proton-decoupled ^{13}C NMR spectra. In the case of isomer **172* a**, a signal of atom C1' of a benzyl moiety was registered as a singlet. The benzylic carbon of **172* b** showed splitting and appeared as a doublet ($^1J_{\text{CN}} = 8.6$ Hz) due to the spin-spin interaction $^{13}\text{C}1'-^{15}\text{N}1$ (Scheme 45).

The obtained results allowed the authors to reach the conclusion that this method for determining the site of alkylation may well prove applicable in other heterocyclic systems where ^{15}N can be selectively incorporated.

Indeed, the selective ^{15}N -labeling of compound **178*** and analysis of the ^{13}C - ^{15}N spin-spin interaction allowed the unambiguous determination of *N*-methylation sites in a series of imidazo[2,1-*c*][1,2,4]triazin-4-ones and related azolo[5,1-*c*][1,2,4]triazin-4-ones.⁶¹ In this work, compound **178*** was prepared by the coupling of isopropylidene malonate **175** (Meldrum's acid) with diazoazole **174*** (Scheme 46). The incorporation of the isotope label in structure **174*** was based on the treatment of 2-aminoimidazole **173** with ^{15}N -enriched sodium nitrite (^{15}N , 36%) in an acidic medium. The reaction between **175** and **174*** gave product **176***. The heating of **176*** in acetic acid led to azoloazine **178***. Cyclization and decarboxylation occurred in one pot, as shown in Scheme 46.

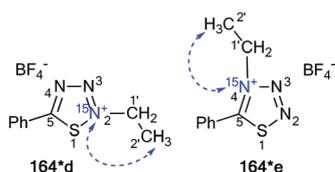


Fig. 2 The two possible alternative *N*-alkylation products are illustrated by structures **164* d** and **164* e** (*N*-2 and *N*-4-alkylation, respectively). The expected but unobserved $^3J_{\text{HN}}$ couplings are indicated by dashed blue arrows.



Scheme 44 The use of ^{15}N -labeled thiazotriazoles to determine the site of alkylation. The observed $^2J_{\text{CN}1'-\text{N}2}$ coupling is shown by a red arrow.

The methylation of compound **178*** yielded isomers **179* a** and **179* b**, which were separated. The detection of the direct ^{13}C - ^{15}N coupling of 11.0 Hz for the signal of the carbon of the *N*-methyl group confirmed the formation of betaine-like structure **179* b** in the alkylation reaction of imidazo[2,1-*c*][1,2,4]triazin-4-ones and their analogs.

It should be noted that today, for the determination of the site(s) of the benzylation/methylation of an unlabeled analog **171*/178***, it is possible to use conventional ^1H , ^{13}C -NMR methods because for the resulting products, the ^1H - ^{13}C spin-spin interactions between the protons of the *N*- CH_2 /*N*- CH_3 fragment and the carbons of the heterocyclic part (8-methylthioimidazo[4,5-*g*]quinazoline/imidazo[2,1-*c*][1,2,4]triazin-4-one) can be identified. That is, these cases do not require ^{15}N labeling and the analysis of J_{CN} couplings. However, the determination of *N*-adamantylation site(s) in unenriched analogs of heterocycle **178*** such as azolo[5,1-*c*][1,2,4]triazine, azolo[1,5-*a*]pyrimidine and related azoloazines using well-established ^1H and ^{13}C NMR methods (such as 1D, 2D COSY, HMQC, HMBC, and INADEQUATE spectra) are difficult because the heterocyclic moiety is covalently attached to the adamantane tertiary carbon that has no bond with hydrogen atoms. Nuclear Overhauser effect spectroscopy (NOESY or ROESY) also does not provide unequivocal structures for *N*-adamantylated derivatives. In this case, the application of ^{15}N labeling and the measurement of ^{13}C - ^{15}N and ^1H - ^{15}N coupling constants allows for the determination of the *N*-adamantylation sites in the azolo-1,2,4-triazine and 1,2,4-triazolo[1,5-*a*]pyrimidine series.

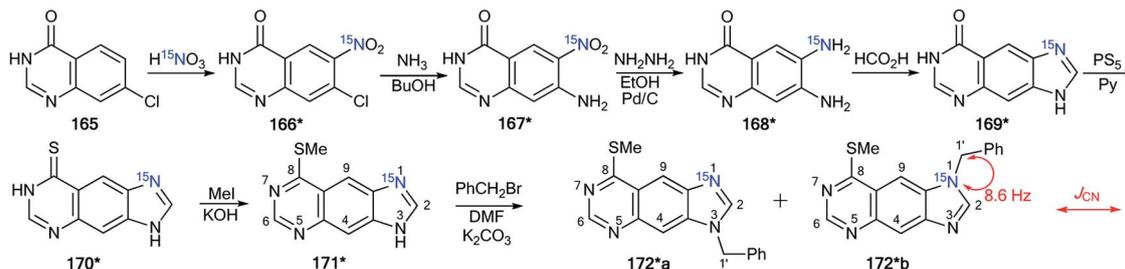
The first time efficiency of this approach was shown in the study of the interaction of 6-nitro-1,2,4-triazolo[5,1-*c*][1,2,4]triazin-7-one **183*** (Scheme 47).⁶² The synthesis of **183*** was based on the treatment of amine-1,2,4-triazole **18*** (^{15}N , 87%) with sodium nitrite in acidic medium and a subsequent interaction of compound **180*** with ethyl nitroacetate **181**.

Then, the resulting sodium salt **182*** was transformed into azoloazine **183***, which was treated with adamantanol **184** in a solution of sulfuric acid. The adamantylation of **183*** gave a mixture of isomers **185* a** and **185* b**. The appearance of the ^{15}N label in structure **185* a** permitted the determination of the conformation of the additional adamantane fragment in the *N*3-atom azole rings by the observation of the $^3J_{\text{CN}}$ couplings (0.5 Hz) in the carbon spectrum.

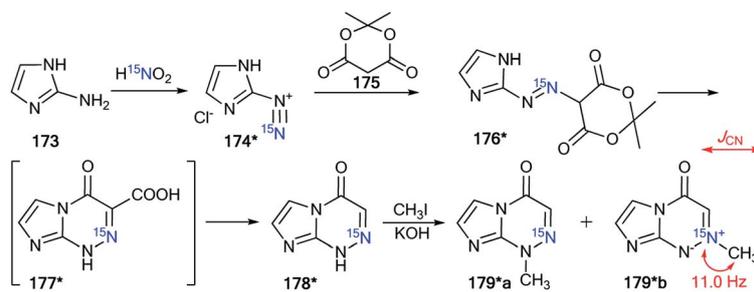
The structure of the adamantylated derivative **185* b** was determined by ^{13}C NMR spectroscopy *via* comparison with a model compound, *N*-methylated 1,2,4-triazolo[5,1-*c*][1,2,4]triazin-7-one. Moreover, the use of ^{15}N -enriched azoloazine **183*** in the investigation of adamantylation showed that compound **185* b** is a product of the reversible isomerization of **185* a**; this rearrangement occurs *via* the formation of an adamantyl cation and heterocyclic base **183***.

The selective incorporation of two ^{15}N atoms in different positions of the 1,2,4-triazolo[5,1-*c*][1,2,4]triazin-7-ones and other nitrogen-containing heterocycles together with a combined analysis of the J_{HN} and J_{CN} coupling constants was more effective for the structural determination of heterocyclic *N*-adamantylated derivatives.³⁹ For example, the use of this approach allowed for the determination of the structures of





Scheme 45 The use of ^{15}N -labeled annulated imidazole to determine the site of alkylation. The observed $^1J_{\text{C}'_1-\text{N}_1}$ coupling is shown by a red arrow.

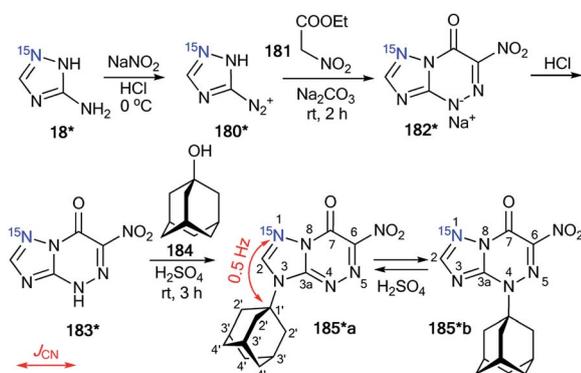


Scheme 46 The use of ^{15}N -labeled triazine to determine the site of alkylation. The observed $^1J_{\text{CN}}$ coupling is shown by a red arrow.

compounds **187**a** and **187**b** obtained by the coupling between **186**** and **184** in trifluoroacetic acid (TFA) solution under reflux (Scheme 48).

The incorporation of two labeled atoms in compound **186**** was based on the treatment of ^{15}N -3-amino-1,2,4-triazole **18*** (^{15}N , 98%) with labeled sodium nitrite (^{15}N , 98%). The reaction of **18*** and ^{15}N -sodium nitrite in acidic medium allowed the *in situ* production of diazonium salt **180****, which reacted with ethyl nitroacetate **181** in a sodium carbonate solution. This reaction led to the formation of $^{15}\text{N}_2$ -1,2,4-triazolo[5,1-*c*][1,2,4]triazinone **182****. The interaction of **182**** with hydrogen bromide generated from acetyl bromide and ethanol gave compound **186****.

It should be noted that the formation of an isomeric mixture in the adamantylation reaction of **186**** could explain the rearrangement of the N3-isomer **170**a** into the N4-isomer



Scheme 47 The use of ^{15}N -labeled triazolotriazine to determine the site of adamantylation. The observed $^3J_{\text{CN}}$ coupling is shown by a red arrow.

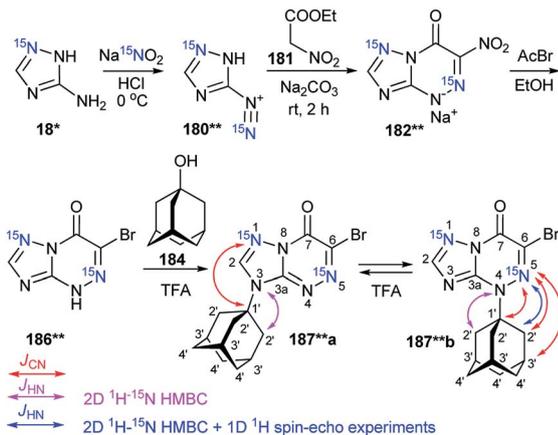
170b**. This process could include the production of an adamantyl cation and the base of **186****. The structures of compounds **187**a,b** were unambiguously confirmed by analysis of the J_{CN} and J_{HN} couplings. The detection of a single $^3J_{\text{C}'_1-\text{N}_1}$ coupling (0.4 Hz) for the adamantane carbon in compound **187**a** indicated that the substituent group is attached to the N3 atom of the 1,2,4-triazole ring (Table 6 and Scheme 48).

The attachment of the adamantane fragment to the N4 atom of the triazine ring in compound **187**b** led to a large set of observable J_{CN} couplings, including geminal ($^2J_{\text{C}'_1-\text{N}_5} = 5.0$ Hz) and vicinal ($^3J_{\text{C}'_2-\text{N}_5} = 1.7$ Hz) (Table 6 and Scheme 45). The ^{15}N -HMBC spectra of compounds **187**a,b** allowed for $^3J_{\text{HN}}$ couplings of the nitrogen atoms at a natural isotopic abundance. The analysis of the $^1\text{H}-^{15}\text{N}_3/^{15}\text{N}_5$ spin-spin interactions also confirmed the structures of the products of adamantylation of **186**** (Table 7 and Scheme 48). Moreover, the appearance of the ^{15}N isotope in the azine ring of **187**b** permitted the observation of the long-range $^1\text{H}'_2-^{15}\text{N}_5$ coupling constant (Table 7 and Scheme 48). This characteristic was additional evidence for the attachment of the adamantane substituent to the unlabeled N4 atom.

In the abovementioned work,³⁹ the selective incorporation of two ^{15}N atoms and combined analysis of J_{CN} and J_{HN} were also used for the determination of the adamantylation site of 1,2,4-triazolo[1,5-*a*]pyrimidine and tetrazolo[1,5-*b*][1,2,4]triazine derivatives (Schemes 49 and 50).

In the first case, double-labeled amino-1,2,4-triazole **18**** was used for to incorporate ^{15}N in the core of 1,2,4-triazolo[1,5-*a*]pyrimidine. Compound **18**** was synthesized by the interaction of $^{15}\text{N}_2$ -hydrazine sulfate (98%, ^{15}N) with *S*-methyl isothiurea sulfate **188** and the consecutive cyclization of $^{15}\text{N}_2$ -aminoguanidine **17**** with formic acid (Scheme 49). The use of





Scheme 48 The use of double ^{15}N -labeled triazolotriazine to determine the site of adamantylation. The measured J_{CN} couplings are shown by red arrows. The J_{HN} couplings measured by amplitude-modulated 1D ^1H spin-echo experiments and detected in the 2D ^{15}N -HMBC spectra are shown by blue and magenta arrows (see the legend in the scheme).

18** in the reaction with ethyl 4,4,4-trifluoroacetate **189** yielded azoloazine **190****, containing two isotopic labels. The adamantylation of **190**** in TFA solution was regioselective and led to the formation of only the N3-isomer **191****. The observed $^{13}\text{C}'\text{-}^{15}\text{N}1/\text{N}8$ coupling in the 1D ^{13}C NMR spectra were acquired with selective decoupling from $^{15}\text{N}1/^{15}\text{N}8$ and the detected $^1\text{H}2'\text{-}^{15}\text{N}3$ cross-peak in the ^{15}N -HMBC spectrum that unambiguously confirmed the position of the adamantyl fragment in compound **191*** (Tables 6, 7 and Scheme 49).

Table 6 J_{CN} couplings (Hz) observed from the signals of the adamantyl fragment in the spectra of azoloazines **187**a,b**, **191**** and **192**a,b**^a

Compound	$^n J_{\text{CN}}$, Hz
187**a	$^3 J_{\text{C}1'\text{-N}1}$ 0.4
	$^2 J_{\text{C}1'\text{-N}5}$ 5.0
187**b	$^3 J_{\text{C}2'\text{-N}5}$ 1.7
	$^4 J_{\text{C}3'\text{-N}5}$ 0.4
	$^3 J_{\text{C}1'\text{-N}1}$ 0.4
	$^3 J_{\text{C}1'\text{-N}8}$ 0.6
191**	$^1 J_{\text{C}1'\text{-N}2}$ 6.5
	$^2 J_{\text{C}1'\text{-N}3}$ 3.8
	$^2 J_{\text{C}2'\text{-N}2}$ 0.4
	$^3 J_{\text{C}2'\text{-N}3}$ 1.2
	$^3 J_{\text{C}3'\text{-N}2}$ 1.6
	$^4 J_{\text{C}3'\text{-N}3}$ 0.2
	$^4 J_{\text{C}4'\text{-N}2}$ 0.3
192**a	$^2 J_{\text{C}1'\text{-N}2}$ 6.5
	$^2 J_{\text{C}1'\text{-N}3}$ 3.8
	$^2 J_{\text{C}2'\text{-N}2}$ 0.4
	$^3 J_{\text{C}2'\text{-N}3}$ 1.2
	$^3 J_{\text{C}3'\text{-N}2}$ 1.6
192**b	$^4 J_{\text{C}3'\text{-N}3}$ 0.2
	$^4 J_{\text{C}4'\text{-N}2}$ 0.3
	$^2 J_{\text{C}1'\text{-N}2}$ 2.7
	$^3 J_{\text{C}1'\text{-N}3}$ 0.3
	$^3 J_{\text{C}2'\text{-N}2}$ 1.1
	$^4 J_{\text{C}3'\text{-N}2}$ 0.3

^a The $^{13}\text{C}\text{-}^{15}\text{N}$ J coupling constants were measured by line-shape analysis in the 1D ^{13}C spectra and acquired with selective ^{15}N decoupling and broadband ^1H decoupling. The estimated error in the J_{CN} values is 0.1 Hz, and the lower limit of reliable J_{CN} measurements is 0.2 Hz.

In contrast to **190****, the adamantylation of tetrazolo-1,2,4-triazine **37**T** in TFA solution led to N2- and N1-regioisomers (compounds **192**a** and **192**b**, respectively, Scheme 50). It was found that compound **192**b** is the product of the reversible isomerization of **192**a**. This process occurs *via* the formation of an adamantyl cation and heterocyclic base **37**T**.

Compound **192**a** was characterized by a set of $^1\text{H}\text{-}^{15}\text{N}$ coupling constants that were detected for the H2' ($^3/4 J_{\text{H}2'\text{-N}2/\text{N}3} = 0.83/0.06$ Hz), H3' ($^4/5 J_{\text{H}3'\text{-N}2/\text{N}3} = 0.60/0.11$ Hz) and H4' ($^5 J_{\text{H}4'\text{-N}2} = 0.23$ Hz) atoms of the adamantane fragment (Table 7 and Scheme 50). These spin-spin interactions, with the exception of $^5 J_{\text{H}3'\text{-N}3}$, were also observed in the 2D ^{15}N -HMBC spectrum. However, analysis of the J_{HN} couplings did not allow the determination of the adamantylation site. This problem was solved by measuring the $^{13}\text{C}\text{-}^{15}\text{N}$ coupling constant (Table 6 and Scheme 50). Observation of the direct $^1 J_{\text{C}1'\text{-N}2}$ (6.5 Hz) and other $^{13}\text{C}\text{-}^{15}\text{N}$ interactions for the C1' ($^2 J_{\text{C}\text{-N}3} = 3.8$ Hz), C2' ($^2 J_{\text{C}\text{-N}2} = 0.4$ Hz and $^3 J_{\text{C}\text{-N}3} = 1.2$ Hz), C3' ($^3 J_{\text{C}\text{-N}2} = 1.6$ Hz and $^4 J_{\text{C}\text{-N}3} = 0.2$ Hz) and C4' ($^4 J_{\text{C}\text{-N}2} = 0.3$ Hz) atoms of the adamantane group in **192**a** indicated that the initial adamantylation of **37**T** occurred on the N2 atom of the tetrazole ring (Table 6 and Scheme 50).

A similar situation with the determination of the adamantylation site in isomer **192**b** had arisen. The detected $^{13}\text{C}\text{-}^{15}\text{N}$ spin-spin interactions of the $^2 J_{\text{C}1'\text{-N}2}$ (2.7 Hz), $^3 J_{\text{C}2'\text{-N}2}$ (1.1 Hz), $^3 J_{\text{C}1'\text{-N}3}$ (0.3 Hz) and $^4 J_{\text{C}3'\text{-N}2}$ (0.3 Hz) couplings in the 1D ^{13}C NMR spectra of **192**b** revealed the attachment of the adamantane fragment to the N1 atom of the tetrazole ring (Table 6 and Scheme 50). However, an analysis of the $^1\text{H}\text{-}^{15}\text{N}$ coupling constants in the ^{15}N -HMBC spectrum ($^3 J_{\text{H}2'\text{-N}1}$ and $^5 J_{\text{H}2'\text{-N}3}$) and the obtained values of J_{HN} from spin-echo experiments in 1D ^1H NMR did not allow the unambiguous establishment of the structure of **192**b** (Table 7 and Scheme 50).

4.2. J_{CN} and J_{HN} coupling constants as an approach for the confirmation of the mechanisms of nitration

The application of ^{15}N -labeled compounds and the measurement of the $^1\text{H}\text{-}^{15}\text{N}$ and $^{13}\text{C}\text{-}^{15}\text{N}$ coupling constants permitted the investigation of the mechanism for nitration in a series of different heterocycles. An unusual method for the incorporation of the nitro group into the core of 6-chloro-9-Boc-purine was found by using a mixture of trifluoroacetic anhydride (TFAA) with labeled tetrabutylammonium nitrate ($\text{Bu}_4\text{N}^+^{15}\text{NO}_3^-$) and freezing the nitration reaction (Scheme 51).⁶³ The transformation of intermediate **194*** was detected at -50 °C by NMR spectroscopy. In the corresponding ^1H spectrum, the signal of proton H8 was split ($^3 J_{\text{HN}} = 2.7$ Hz).

Moreover, in the carbon spectrum of an experiment with $\text{Bu}_4\text{N}^+^{15}\text{NO}_3^-$, the $^2 J_{\text{CN}}$ (1.8 Hz) coupling was observed for C8, and the signal of the carbon of the carbonyl group for the trifluoroacetoxy fragment was characterized by a $^1\text{H}\text{-}^{13}\text{C}$ spin-spin interaction (2.8 Hz). These results unambiguously confirmed the formation of intermediate **194***, which underwent rearrangement into structure **196*** at -10 °C. Notably, this process occurred *via* the formation of intermediate **195*** (Scheme 51).



Table 7 J_{HN} couplings (Hz) of adamantylated azoloazines **187**a,b**, **191**** and **192**a,b**^{a,b}

Compound	N1/N2	N3/N5/N8	¹⁵ N at natural abundance N1/N3/N4
187**a			³ $J_{\text{H}2'-\text{N}3}$ (m)
187**b		⁴ $J_{\text{H}2'-\text{N}5}$ 0.06 (w)	³ $J_{\text{H}2'-\text{N}4}$ (m)
191**			³ $J_{\text{H}2'-\text{N}3}$ (m)
192**a	³ $J_{\text{H}2'-\text{N}2}$ 0.83 (s), ⁴ $J_{\text{H}3'-\text{N}2}$ 0.60 (m), ⁵ $J_{\text{H}4'-\text{N}2}$ 0.23 (m)	⁴ $J_{\text{H}2'-\text{N}3}$ 0.06 (w), ⁵ $J_{\text{H}3'-\text{N}3}$ 0.11 (-)	
192**b	⁴ $J_{\text{H}2'-\text{N}2}$ < 0.04 ^c (-), ⁵ $J_{\text{H}3'-\text{N}2}$ 0.04 (-)	⁵ $J_{\text{H}2'-\text{N}3}$ < 0.04 ^c (w)	³ $J_{\text{H}2'-\text{N}1}$ (m)

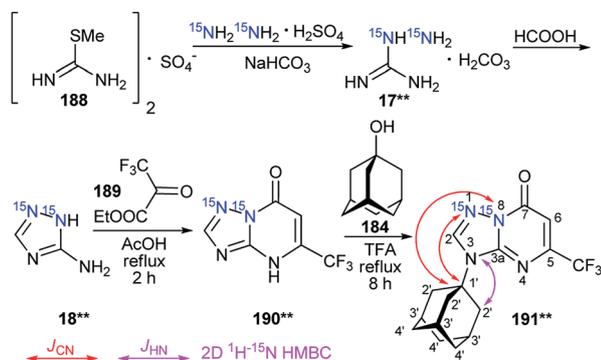
^a The J_{HN} values were measured using amplitude-modulated 1D ¹H spin-echo experiments with delays for the evolution of J_{HN} up to 1 s. The estimated error in the J_{HN} values is 0.02 Hz, and the lower limit of reliable J_{HN} measurements is 0.04 Hz. ^b The cross-peaks in the 2D ¹⁵N-HMBC spectra were classified into three categories (weak - w; medium - m; strong - s). Weak peaks approximately correspond to $J_{\text{HN}} < 0.5$ Hz, strong peaks approximately correspond to $J_{\text{HN}} > 2$ Hz, and medium peaks correspond to the remaining values. ^c The measurement of the J_{HN} values was impossible due to the fast transverse relaxation of the corresponding ¹H nuclei.

The use of ¹⁵N-enriched nitric acid in the nitration of nitroimidazole **197** allowed the detection of the rearrangement occurring in this process.⁶⁴ It was found that the interaction of **197** with ¹⁵N-nitric acid led to N-¹⁵NO₂ dinitroimidazole **198***, which underwent isomerization into compound **199*** in chlorobenzene at 115 °C (Scheme 52). It should be noted that the carbon NMR spectrum of dinitroimidazole **199*** was only characterized by one direct ¹³C-¹⁵N coupling constant at the C2 atom with a magnitude of 30.6 Hz (Scheme 52). Thus, the fact that a doublet was observed for the ¹³C resonance at the 2-position not only confirms the assignment of the C-2 resonance but also excluded the formation of intermediate **200*** in the process of the isomerization of **198*** into **199***. If this transformation included the production of structure **200***, statistically only 50% of the ¹⁵N would move to the 2-position. Moreover, this result showed that the isomerization included a [1,5]-sigmatropic rearrangement.

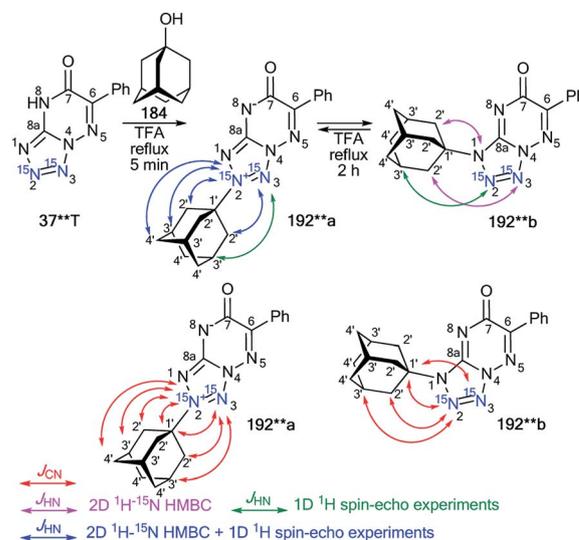
4.3. Analysis of ¹H-¹⁵N and ¹³C-¹⁵N constants for the confirmation of the structures of amine and imine derivatives

The appearance of labeled nitrogen atoms in different amines simplifies the determination of the structures of the reaction products exploiting the analysis of ¹³C-¹⁵N and ¹H-¹⁵N coupling constants. In that vein, the specifics of the interaction of 2,3-

diketopyrido[4,3,2-*de*]quinolines **201** with ¹⁵N-propylamine were studied⁶⁵ (Scheme 53). The ¹H NMR spectrum of the final product **206*** showed signals from two protons that are attached to the ¹⁵N-labeled atom (J_{HN} 92.5 Hz, Scheme 53). These data unambiguously confirmed the production of compounds **206***. Then, the author shed light on the mechanism of this transformation after the isolation of intermediate **203***. The structure of **203*** was confirmed by ¹H and ¹³C NMR spectroscopy. Indeed, the proton spectrum showed that proton H1' and an *n*-propyl group are attached to the ¹⁵N atom. The signal of atom H1' was characterized by a direct ¹H-¹⁵N' coupling constant with a magnitude of 92.5 Hz (Scheme 53). The aliphatic C2' and the aromatic C4 are directly linked to the labeled nitrogen atom ($J_{\text{C}2'-\text{N}1'} = 8.5$ Hz and $J_{\text{C}4-\text{N}1'} = 15.8$ Hz, Scheme 53). This result suggested that the transformation of **201** into **206*** under the action of ¹⁵N-propylamine includes an amination-elimination reaction that involves the formation of structures **202*-205*** (Scheme 53).

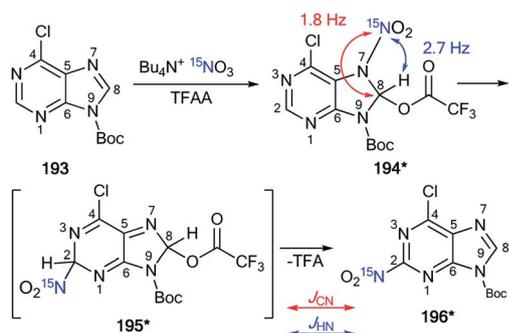


Scheme 49 The use of double ¹⁵N-labeled triazolopyrimidine to determine the site of adamantylation. The measured J_{CN} couplings are shown by red arrows. The observed J_{HN} coupling in the 2D ¹⁵N-HMBC spectra is shown by a magenta arrow (see the legend in the scheme).

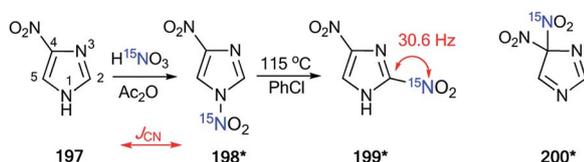


Scheme 50 The use of double ¹⁵N-labeled triazolotetrazine to determine the site of adamantylation. The observed J_{CN} couplings are shown by red arrows. The measured J_{HN} couplings by amplitude-modulated 1D ¹H spin-echo experiments and detected ¹H-¹⁵N spin-spin interactions in the 2D ¹⁵N-HMBC spectra are shown by blue, green and magenta arrows (see the legend in the scheme).



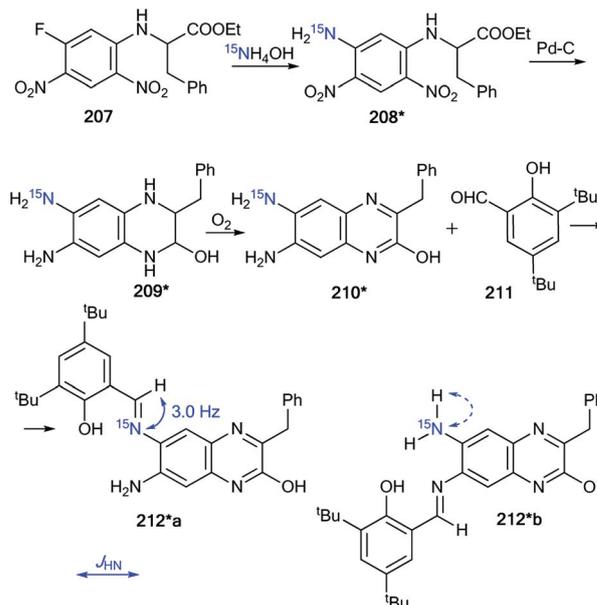


Scheme 51 The unusual mechanism of nitration using ^{15}N -labeled compounds. The observed J_{CN} and J_{HN} couplings are shown by red and blue arrows, respectively.



Scheme 52 An example of the application of ^{15}N -labeled compounds for establishing a nitration mechanism in a series of imidazoles. The observed direct J_{CN} coupling is shown by a red arrow.

The selective incorporation of the ^{15}N -label into diamino-2-quinoxalino **210*** permitted the analysis of the ^1H - ^{15}N coupling constants for the confirmation of the structure of product **212***, which was isolated from the reaction of **210*** with salicylaldehyde **211** (Scheme 54).⁶⁶ A synthesis of compound **210*** was based on several steps. Initially, an interaction occurred between compound **207** and ^{15}N -labeled ammonium hydroxide (^{15}N , 98%), and then **208*** was transformed into **209***.

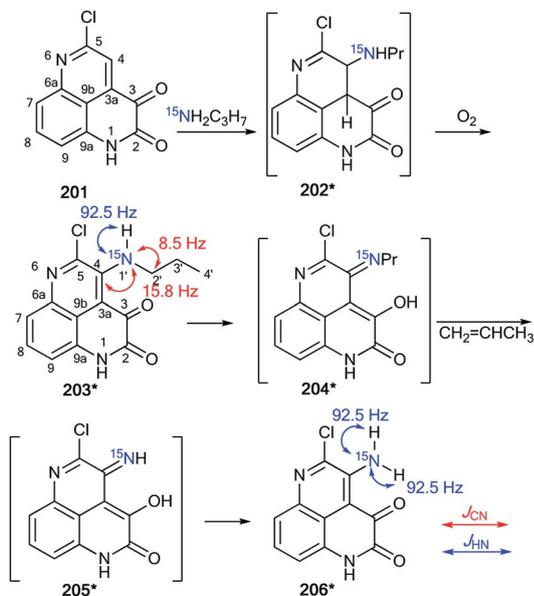


Scheme 54 The use of ^{15}N -labeled *o*-phenylenediamine for establishing the site of reaction with salicylic aldehyde. The observed $^2J_{\text{HN}}$ coupling is shown by a blue arrow. The expected but unobserved coupling $^1J_{\text{HN}}$ is indicated by a dashed arrow.

Oxidation of **209*** led to diamine **210***, which had two amino groups with different reactivities. Next, reaction of compound **210*** with aldehyde **211** could give an imine **212*a** or the isomer **212*b**. The observation of the ^1H - ^{15}N coupling constant for the proton signal of the imine group ($^2J_{\text{HN}} = 3.0 \text{ Hz}$) confirmed the formation of structure **212*a**.

5. Conclusion

We have reviewed examples of the incorporation of ^{15}N atoms and subsequent analyses of the J_{HN} and J_{CN} couplings in labeled samples. The above approach can be considered an effective, general, and convenient tool for establishing realistic structures of nitrogen-containing compounds in solution, including mixtures of compounds and equilibrium mixtures. The measurement of the ^1H - ^{15}N and ^{13}C - ^{15}N coupling constants allows the investigation of ring-chain tautomerism, the mechanisms of chemical transformation and other structural aspects of azoles, azines, azepines and their fused derivatives. Although this approach is not widely used, ^{15}N labeling and estimation of the ^1H - ^{15}N and ^{13}C - ^{15}N spin-spin interactions may be a single method that can be efficient for the determination of the molecular structure or method of chemical transformation in the chemistry of poly-nitrogen heterocycles. In structural studies, two types of coupling constants are used, namely, long-range and near (direct, geminal, vicinal) constants. While early publications used only near constants, modern works exploit both long-range and near constants, as well as 2D spectra. For example, 2D ^1H - ^{15}N HMBC spectra and spin-echo experiments were used for the determination of the *N*-adamantylation site of azolo-1,2,4-triazines and confirmation of the structure of



Scheme 53 The use of ^{15}N -labeled propyl amine to determine the oxidative amination mechanism. The observed direct J_{CN} and J_{HN} couplings are shown by red and blue arrows, respectively.



imidazo[1,2-*a*]pyrimidine. This situation is closely related to the development of the NMR technique. Thus, it can be expected that this approach will be expanded in the chemistry of nitrogen-containing compounds since ^{15}N labeling and the analysis of the ^{13}C - ^{15}N and ^1H - ^{15}N spin-spin coupling constants shines light on new and known chemical transformations, rearrangements and ring-chain tautomerism in a series of nitrogen heterocycles.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

This work was supported by the Ministry of Science and Higher Education of the Russian Federation (State contract 4.6351.2017/8.9) and the Russian Foundation for Basic Research (grant 17-03-01029).

References

- M. Baumann and I. R. Baxendale, *Beilstein J. Org. Chem.*, 2013, **9**, 2265–2319.
- T. Che, Y.-Q. Wang, Z.-L. Huang, J.-H. Tan, Z.-S. Huang and S.-B. Chen, *Molecules*, 2018, **23**, 493–511.
- M. Krasavin, *Chem. Heterocycl. Compd.*, 2017, **53**, 240–255.
- P. Martins, J. Jesus, S. Santos, L. R. Raposo, C. Roma-Rodrigues, P. V. Baptista and A. R. Fernandes, *Molecules*, 2015, **20**, 16852–16891.
- Y.-J. Wu, in *Progress in Heterocyclic Chemistry*, ed. G. W. Gribble and J. A. Joule, Elsevier, Amsterdam, 2012, vol. 24, pp. 1–53.
- L. Wang, Y. Tian, W. Chen, H. Liu, P. Zhan, D. Li, H. Liu, E. De Clercq, C. Pannecouque and X. Liu, *Eur. J. Med. Chem.*, 2014, **85**, 293–549.
- S. Cascioferro, B. Parrino, V. Spano, A. Carbone, A. Montalbano, P. Barraja, P. Diana and G. Cirrincione, *Eur. J. Med. Chem.*, 2017, **142**, 74–86.
- E. K. Davison and J. Sperry, *J. Nat. Prod.*, 2017, **80**, 3060–3079.
- D. C. G. A. Pinto, C. M. M. Santos and A. M. S. Silva, *Advanced NMR techniques for structural characterization of heterocyclic structures*, in *Recent Research Developments in Heterocyclic Chemistry*, ed. T. M. V. D. Pinho e Melo and A. M. R. Gonsalves, Research Signpost, Kerala, India, 2007, ch. 8, pp. 397–475.
- A. K. Verma, R. R. Jha, R. Chaudhary, R. K. Tiwari and A. K. Danodia, *Adv. Synth. Catal.*, 2013, **355**, 421–438.
- G. Desimoni, G. Faita and P. Quadrelli, *Chem. Rev.*, 2003, **103**, 3119–3154.
- J. N. McPherson, B. Das and S. B. Colbran, *Coord. Chem. Rev.*, 2018, **375**, 285–332.
- H. Xu, R. Chen, Q. Sun, W. Lai, Q. Su, W. Huang and X. Liu, *Chem. Soc. Rev.*, 2014, **43**, 3259–3302.
- V. W.-W. Yam, V. K.-M. Au and S. Y.-L. Leung, *Chem. Soc. Rev.*, 2015, **115**, 7589–7728.
- P. J. Panak and A. Geist, *Chem. Rev.*, 2013, **113**, 1199–1236.
- H. Gao and J. M. Shreeve, *Chem. Rev.*, 2011, **111**, 7377–7436.
- Y. Tang, C. He, G. H. Imler, D. A. Parrish and J. M. Shreeve, *Chem. Commun.*, 2018, **54**, 10566–10569.
- S. L. Deev, T. S. Shestakova, V. N. Charushin and O. N. Chupakhin, *Chem. Heterocycl. Compd.*, 2017, **53**, 963–975.
- S. Cheatham, M. Kline and E. Kupče, *Magn. Reson. Chem.*, 2015, **53**, 363–368.
- S. Cheatham, P. Gierth, W. Bermel and Ě. Kupče, *J. Magn. Reson.*, 2014, **247**, 38–41.
- J. Cavanagh, W. J. Fairbrother, A. G. Palmer III, N. J. Skelton and M. Rance, *Protein NMR Spectroscopy: Principles and Practice*, Academic Press, San Diego, CA, 2nd edn, 2006.
- A. Bax, G. W. Vuister, S. Grzesiek, F. Delaglio, A. C. Wang, R. Tschudin and G. Zhu, *Methods Enzymol.*, 1994, **239**, 79–105.
- F. H. T. Nelissen, M. Tessari, S. S. Wijmenga and H. A. Heus, *Prog. Nucl. Magn. Reson. Spectrosc.*, 2016, **96**, 89–108.
- M. Nowakowski, S. Saxena, J. Stanek, S. Žerko and W. Koźmiński, *Prog. Nucl. Magn. Reson. Spectrosc.*, 2015, **90–91**, 49–73.
- P. E. Hansen, *Magn. Reson. Chem.*, 2000, **38**, 1–10.
- R. Marek and A. Lyka, *Curr. Org. Chem.*, 2002, **6**, 35–66.
- M. Chatzopoulou, R. F. Martínez, N. J. Willis, T. D. W. Claridge, F. X. Wilson, G. M. Wynne, S. G. Davies and A. J. Russell, *Tetrahedron*, 2018, **74**, 5280–5288.
- L. Wang, A. Bhan and R. S. Hosmane, *Nucleosides Nucleotides*, 1994, **13**, 2307–2320.
- E. Kleinpeter, *NMR Spectroscopic Study of Tautomerism in Solution and in the Solid State*, in *Tautomerism: Methods and Theories*, ed. L. Antonov, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, Germany, 2013, pp. 103–143.
- E. Kolehmainen and B. Osmialowski, *Int. Rev. Phys. Chem.*, 2012, **31**, 567–629.
- L. I. Larina, *Adv. Heterocycl. Chem.*, 2018, **124**, 233–321.
- B. T. Golding, P. K. Slaich and W. P. Watson, *J. Chem. Soc., Chem. Commun.*, 1986, 515–517.
- J. Farras and J. Vilarrasa, *J. Chem. Soc., Chem. Commun.*, 1986, 1127–1129.
- O. N. Chupakhin, E. N. Ulomsky, S. L. Deev and V. L. Rusinov, *Synth. Commun.*, 2001, **31**, 2351–2355.
- M. V. Pryadeina, Ya. V. Burgart, V. I. Saloutin, M. I. Kodess, E. N. Ulomskii and V. L. Rusinov, *Russ. J. Org. Chem.*, 2006, **40**, 902–907.
- M. M. Maslova, V. I. Pol'shakov, O. S. Anisimova, N. B. Marchenko and R. G. Glushkov, *Pharm. Chem. J.*, 1993, **26**, 889–893.
- S. L. Deev, Z. O. Shenkarev, T. S. Shestakova, O. N. Chupakhin, V. L. Rusinov and A. S. Arseniev, *J. Org. Chem.*, 2010, **75**, 8487–8497.
- I. A. Khalymbadzha, T. S. Shestakova, S. L. Deev, V. L. Rusinov, O. N. Chupakhin, Z. O. Shenkarev and A. S. Arseniev, *Russ. Chem. Bull.*, 2013, **62**, 521–528.
- S. L. Deev, A. S. Paramonov, T. S. Shestakova, I. A. Khalymbadzha, O. N. Chupakhin, J. O. Subbotina, O. S. Eltsov, P. A. Slepukhin, V. L. Rusinov, A. S. Arseniev



- and Z. O. Shenkarev, *Beilstein J. Org. Chem.*, 2017, **13**, 2535–2548.
- 40 T. S. Shestakova, Z. O. Shenkarev, S. L. Deev, O. N. Chupakhin, I. A. Khalymbadzha, V. L. Rusinov and A. S. Arseniev, *J. Org. Chem.*, 2013, **78**, 6975–6982.
- 41 P. MacCormack, J.-C. Hallé, M.-J. Pouet and F. Terrier, *J. Org. Chem.*, 1988, **53**, 4407–4409.
- 42 M. R. Crampton, L. M. Pearce and L. C. Rabbitt, *J. Chem. Soc., Perkin Trans. 2*, 2002, 257–261.
- 43 K. Akiba, K. Kashiwagi, Y. Ohyama, Y. Yamamoto and K. Ohkata, *J. Am. Chem. Soc.*, 1985, **107**, 2721–2730.
- 44 V. N. Kozhevnikov, D. N. Kozhevnikov, V. L. Rusinov and O. N. Chupakhin, *Chem. Heterocycl. Compd.*, 1999, **25**, 470–473.
- 45 O. N. Chupakhin, V. L. Rusinov, A. A. Tumashov, E. O. Sidorov and I. V. Karpin, *Tetrahedron Lett.*, 1992, **33**, 3695–3696.
- 46 O. N. Chupakhin, V. L. Rusinov, T. L. Pilicheva and A. A. Tumashov, *Synthesis*, 1990, 713–717.
- 47 R. Y. Baiazitov, N. Sydorenko, H. Ren and Y.-C. Moon, *J. Org. Chem.*, 2017, **82**, 5881–5889.
- 48 X. Ariza, V. Bou and J. Vilarrasa, *J. Am. Chem. Soc.*, 1995, **117**, 3665–3673.
- 49 C. A. Montanari, J. P. B. Sandall, Y. Miyata and J. Miller, *J. Chem. Soc., Perkin Trans. 2*, 1994, 2571–2575.
- 50 D. Moderhack and D. Decker, *Heterocycles*, 1994, **37**, 683–688.
- 51 M. Christ, J. P. Warren, B. L. Hawkins and J. D. Roberts, *J. Am. Chem. Soc.*, 1973, **95**, 4392–4397.
- 52 N. A. Kondratova, N. P. Solov'eva and V. F. Traven, *Russ. Chem. Bull.*, 2010, **59**, 1612–1620.
- 53 A. Lyčka, I. Fryšova and J. Slouka, *Magn. Reson. Chem.*, 2007, **45**, 46–50.
- 54 I. Wiedermannová, J. Slouka, O. Humpa and K. Lemr, *J. Heterocycl. Chem.*, 2003, **40**, 357–358.
- 55 O. R. Gautun and P. H. J. Carlsen, *Acta Chem. Scand.*, 1992, **46**, 469–473.
- 56 P. Kang and C. S. Foote, *J. Am. Chem. Soc.*, 2002, **124**, 9629–9638.
- 57 S. M. Ametamey, R. Hollenstein and H. Heimgartner, *Helv. Chim. Acta*, 1988, **71**, 521–530.
- 58 X. Creary and A. F. Sky, *J. Am. Chem. Soc.*, 1990, **112**, 368–374.
- 59 A. Holm, K. Schaumburg, N. Dahlberg, C. Christophersen and J. P. Snyder, *J. Org. Chem.*, 1975, **40**, 431–436.
- 60 D. F. Wiemer, D. I. C. Scopes and N. J. Leonard, *J. Org. Chem.*, 1976, **41**, 3051–3053.
- 61 J. Farràs, E. Fos, R. Ramos and J. Vilarrasa, *J. Org. Chem.*, 1988, **53**, 887–891.
- 62 E. N. Ulomskii, S. L. Deev, A. V. Tkachev, I. K. Moiseev and V. L. Rusinov, *Russ. J. Org. Chem.*, 2002, **38**, 272–280.
- 63 B. Rodenko, M. Koch, A. M. van der Burg, M. J. Wanner and G.-J. Koomen, *J. Am. Chem. Soc.*, 2005, **127**, 5957–5963.
- 64 S. Bulusu, R. Damavarapu, J. R. Autera, R. Behrens, L. M. Minier and T. Axenrod, *J. Phys. Chem.*, 1995, **99**, 5009–5015.
- 65 Q. Ding and J. W. Lown, *J. Org. Chem.*, 1999, **64**, 7965–7972.
- 66 X. V. Wu, A. E. Gorden, S. A. Tonks and J. Z. Vilseck, *J. Org. Chem.*, 2007, **72**, 8691–8699.

