



Cite this: *RSC Adv.*, 2019, 9, 21265

Utilization of catecholic functionality in natural safrole and eugenol to synthesize mussel-inspired polymers†

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Naturally occurring safrole I upon epoxidation gave safrole oxide II, which underwent ring opening polymerization using a Lewis acid initiator/catalyst comprising of triphenylmethylphosphonium bromide/triisobutylaluminum to afford new polyether III in excellent yields. Epoxy monomer II and allyl glycidyl ether IV in various proportions have been randomly copolymerized to obtain copolymer V. A mechanism has been proposed for the polymerization reaction involving chain transfer to the monomers. A strategy has been developed for the deprotection of the methylene acetal of V using Pb(OAc)₄ whereby one of the methylene protons is replaced with a labile OAc group to give VI. The pendant allyl groups in VI have been elaborated *via* a thiol–ene reaction using cysteamine hydrochloride and thioglycolic acid to obtain cationic VII and anionic VIII polymers, both containing a mussel-inspired Dopa-based catechol moiety. During aqueous work up, the protecting group containing OAc was deprotected under mild conditions. Cationic VII and anionic VIII were also obtained *via* an alternate route using epoxide IX derived from 3,4-bis[*tert*-butyldimethylsilyloxy]allylbenzene. Monomer IX was homo- as well as copolymerized with IV using Lewis acid initiator/catalyst system to obtain homopolymer X and copolymer X1. Copolymer XI was then elaborated using a thiol–ene reaction followed by F[−] catalysed silyl deprotection to obtain mussel inspired polymers VII and VIII, which by virtue of having charges of opposite algebraic signs were used to form their coacervate.

Received 23rd June 2019
 Accepted 1st July 2019

DOI: 10.1039/c9ra04719k

rsc.li/rsc-advances

1 Introduction

The performance of synthetic adhesive polymers in aquatic environments is most often frustrating because of the various adverse effects of water or moisture in inducing hydrolysis, swelling, interfacial wicking, *etc.*^{1,2} Rapid, strong and tough moisture-resistant adhesion to solid surfaces in the sea by the holdfast of marine mussels have lured many a researcher to mimic the essential features of the adhesive chemistry practiced by the mussels. Roughly 25–30 different adhesive proteins of varying molar masses used by the mussels contained a considerable proportion of randomly placed 3,4-dihydroxyphenyl-L-alanine (L-DOPA) (≈ 1 –28 mol%) and 4-hydroxyarginine resulting from a post translational modification of amino acid tyrosine and arginine, respectively.¹ To be an effective adhesive, it must exert more favourable interaction with the wet polar surfaces than a layer of water is able to offer. Polymers

covalently bonded into the interfaces may mitigate the deleterious effects of moisture, but it is a costly process.³ As such the situation demanded a closer look at the way mussel's adhesive chemistry works. The byssus is a bundle of filaments of DOPA decorated proteins secreted by mussels that function to attach the mussels to a solid surface whereby the catecholic functionalities provide bidentate or covalent interfacial interactions. The proteins themselves gain cohesiveness through metal chelation, and covalent coupling.

The cohesive and adhesive roles played by the catechol functionality of DOPA in byssal proteins and their effective adhesive performance in turbulent wet environments have now been firmly established. The biomimetic efforts have thus targeted the attractive mussel byssal proteins. However, the performance of biological adhesion, *e.g.*, from mussels is much superior to biomimetic adhesives. Recent advances demonstrate that judicious biomimetic design incorporating key elements in the natural adhesive system is essential to replicate the wet adhesion of the marine organism.^{4,5} Several reviews deal with the recent progress of mussel-inspired underwater adhesives polymers having catechol-functional motifs for their potential applications in anti-biofouling, biological adhesives, and drug delivery.^{6,7} A review specifically provides an overview of the various applications of poly[2-(3,4-dihydroxyphenyl)ethylamine] (*i.e.* polydopamine (PDA)) in tumor targeted drug

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† Electronic supplementary information (ESI) available. See DOI: 10.1039/c9ra04719k



delivery systems and discusses the release behavior of the drug-loaded PDA-based nanocarriers.⁸

The synthesis of mussel mimetic polymers include DOP-Apolypeptides^{9,10} and DOPA and lysine polypeptide copolymers.^{11,12} Hydrogen bonding by phenolic hydroxyls of bidentate DOPA leads to its bridging adhesion to the polysiloxane surface of mica.¹³ The mussel mimicking adhesives has also been applied as complex coacervates,^{14,15} which are prepared using aqueous solutions of polyanions (*e.g.* DOPA-containing proteins having phosphoserine residues) and polycations (*e.g.* Dopa-containing proteins having 4-hydroxyarginine residues). With charge symmetry, the polymers of different algebraic signs undergo phase separation in a complex coacervation process, which is essential for wet adhesion. Synthesis of mussel mimetic polymers have been achieved through polymerization of catechol-based monomers.^{16,17} Dopamine methacrylamide and 2-methoxyethyl acrylate or poly(ethylene glycol) methyl ether methacrylate has been copolymerized to give adhesive polymers having divergent physical properties.¹⁸

A review article discusses the mussel-inspired chemistry in the surface engineering of polymer membranes to improve their performance. Catecholamines, deposited on the membrane surface, serve as a surface component for membrane modification or fabrication.¹⁹ Polydopamine capped graphene oxide sheets are crosslinked by polyethylenimine leading to ultrahigh modulus and high strength of macroscopic graphene oxide papers, which broaden the potential applications of graphene.²⁰ The adhesive mechanism of mussel-inspired polymers has been investigated using the terpolymers of *N*-(3,4-dihydroxyphenethyl) methacrylamide (DMA), acrylic acid and butyl acrylate. The bulk adhesion was found to increase with a corresponding increase in DMA content reaching a maximum at around 40 mol%.²¹ Inspired by strong adhesion of mussel adhesive proteins, a high-performance nanocomposite is generated by assembling polydopamine-coated montmorillonite with corn starch.²² Underwater-superoleophobic materials have been successfully developed by a two-step dip-coating method with mussel-inspired coatings of polydopamine and subsequent zwitterionic sulfobetaine methacrylate grafting onto stainless steel meshes and used in oil/water separation.²³ Two functional catechols were incorporated into backbones of polyacrylate and polyurethane, which showed excellent coatability on various surfaces.²⁴

Homo- and copolymers of catecholic monomer 3,4-dihydroxystyrene with styrene or styrene sulfonate has been reported to give polymers having a wide range of physical properties.^{18,25} A complex coacervate adhesive has been synthesized from a mixture of catechol containing synthetic polyanion and a synthetic polycation.^{14,26} Coating of polyethylene glycol containing Dopa as an anchor has been demonstrated to prevent bacterial adhesion and biofilm formation on a variety of surfaces.^{27,28} One of the serious drawbacks of mussel proteins is the oxidation of DOPA at or above neutral pH to DOPA quinone which has been shown to be less sticky than DOPA. While the mussels have the talents^{29,30} to control the deleterious oxidation, it remains a challenge to the researchers to safeguard the integrity of the catecholic

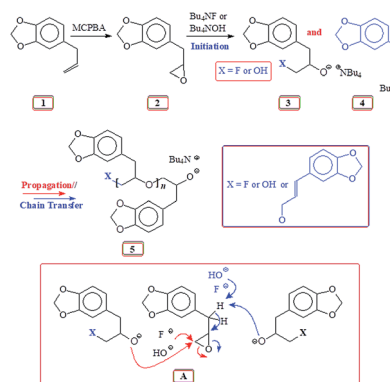
functionality in DOPA for facile bidentate surface binding. Even in the more adverse pH of 8.2 of seawater, the mussels secrete their proteins at a low pH of 5–6 in the confined space of a reducing environment where the redox balance is provided by the thiol functionalities of a cysteine-rich protein.

It is our intention to synthesize mussel-inspired polymers using ring opening polymerization of epoxide (oxirane). Several review articles summarize the developments in the ring opening polymerization of alkylene oxides *via* anionic, coordination and cationic polymerization using a variety of catalysts including metal-free organocatalysts to synthesize linear homo- and amphiphilic block as well as branched, hyperbranched, and dendrimer like polyethers.^{31–33} Herein we report the use of readily available naturally occurring safrole (1) (Scheme 1) or eugenol (10) (Scheme 5) to synthesize epoxy functionalized monomers safrole oxide (2) (Scheme 1) and eugenol-derived 14 (Scheme 5) and their ring opening homo- and copolymerization with allyl glycidyl ether (AGE) 7 (Scheme 2). The allyl pendants of the mussel mimicking polymers decorated with catechol functionalities would then offer the latitude of transformation to ionic polymer backbones of both algebraic signs. The work would thus pave the way to study these polymers from the perspective of coacervate adhesive.

2 Experimental

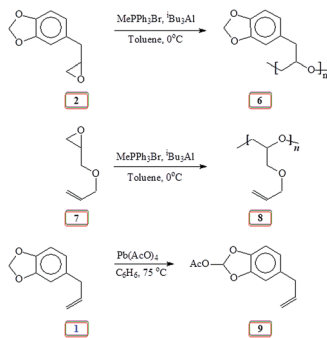
2.1. Materials

m-Chloroperbenzoic acid (MCPBA), LiCl, *tert*-butylammonium fluoride (TBAF · 3H₂O), *tert*-butylammonium hydroxide (TBAH), 4-dimethylaminopyridine (DMAP), ¹Bu₃Al solution (25% in toluene), eugenol, 2,2-dimethoxy-2-phenylacetophenone (DMPA), *t*-butyldimethylsilyl chloride (TBDMSCl), thioglycolic acid, cysteamine and safrole were purchased from Sigma Aldrich. MePPh₃Br and imidazole from Fluka, 1 M *tert*-butylammonium fluoride (TBAF) solution in THF from ChemCruz, and diphenylsilane and B(C₆F₅)₃ from Alfa Aesar were used as received without any further purification. 4-Allylcatechol was prepared by reacting eugenol with LiCl in dimethyl formamide (DMF) as described.³⁴ Allyl glycidyl ether (AGE) from Sigma Aldrich was dried over CaH₂ and distilled. Pb(AcO)₄ was freshly



Scheme 1 Base catalyzed polymerization of safrole oxide 2 using Bu₄NF and Bu₄NOH.





Scheme 2 Lewis acid catalyzed polymerization of safrole oxide 2, AGE 7 and activation of methylene group.

prepared.³⁵ Membrane (Spectra/Por) with a MWCO of 6000–8000 Daltons was purchased from Spectrum Laboratories, Inc.

2.2. Physical methods

Elemental analyses were performed on a PerkinElmer (Series II Model 2400). FTIR and NMR were recorded using a PerkinElmer 16F PC and a 500 MHz JEOL LA spectrometer, respectively. Tetramethylsilane (TMS) in CDCl₃ and residual H in D₂O at δ 4.65 ppm were used as internal standards. The ¹³C chemical shifts in D₂O were referenced against ¹³C peak of external standard dioxane at δ 67.4 ppm. Gel Permeation Chromatographic analysis was performed at 30 °C using a PL-GPC 220 manufactured by Agilent Technologies having two detectors (*i.e.* Refractive Index & Light Scattering), equipped with two columns (PL aquagel-OH 8 μ m; 300 \times 7.5 mm). Deionized water with 0.02 wt% NaN₃ was used as solvent for water soluble polymer samples. A sample solution of 5 mg/1.5 mL was prepared at ambient temperature in water having 0.02 wt% NaN₃. The resulting solution (100 μ l) was injected into the GPC columns and chromatographic data was analyzed using GPC/SEC software by Agilent. In case of THF soluble polymer samples, GPC was equipped with two columns (PLgel Olexis 300 \times 7.5 mm). The sample solution was prepared using tetrahydrofuran (THF) containing 0.0125 wt% antioxidant, [2,6-di-*tert*-butyl-4-methyl phenol (BHT)]. The equipment was calibrated using polyethylene oxide (PEO) and polystyrene (PS) standards for water-soluble and THF-soluble polymers, respectively.

2.3. Synthesis of safrole oxide (2) from safrole (1)

Safrole was converted to safrole oxide using a modified procedure.^{36,37} Safrole 1 (32.4 g) was added to a solution of MCPBA (54 g, 0.22 mol, 70% purity) in chloroform (400 mL) and stirred at 25 °C for 24 h. After adding additional MCPBA (20 g, 0.082 mol), the reaction mixture was stirred for a further 24 h and was washed with 10% sodium hydroxide solution (3 \times 70 mL) and water (2 \times 30 mL). The organic layer was dried (MgSO₄) and concentrated. The residual liquid was purified by silica gel chromatography using hexane/ether as eluent to give safrole oxide (2) as a colorless oil (28.8 g, 67%). The oil was then distilled over calcium hydride to exclude any moisture $b_{p,0.8}$ mbarHg 92 °C. δ_{H} (CDCl₃): 2.52 (1H, m), 2.83–3.73 (3H, m), 3.10 (1H, m), 5.91 (2H, s), 6.69 (1H, d, *J* 7.6 Hz), 6.74 (1H, s), 6.75 (1H,

d, *J* 7.9 Hz); δ_{C} (CDCl₃): 28.39, 46.77, 52.53, 100.87, 108.26, 109.43, 121.85, 130.79, 146.28, 147.66 (CDCl₃ middle carbon: 77.00).

2.4. Lewis acid catalyzed polymerization of safrole oxide 2 using ¹Bu₃Al^{38,39}

As described in entry 10 (Table 1), methyl-triphenylphosphonium bromide (11.9 mg, 0.0333 mmol) was placed in an RB flask and closed with a rubber septum. Safrole oxide 2 (445 mg, 2.5 mmol) and anhydrous toluene (1.6 mL) were added under Ar by a syringe. The mixture was cooled to 0 °C, then 1 M triisobutylaluminium solution (0.376 mL of 25 wt% in toluene, 0.403 mmol) was injected by a syringe under Ar. The polymerization was quenched after 2 h by adding 4 : 1 MeOH/H₂O (v/v). After removal of the solvents, the residue was dissolved in CH₂Cl₂ (50 mL) and filtered over Celite 545. The filtrate was dried (over Na₂SO₄) and evaporated to obtain polymer 6.

For purification, polymer 6 (Scheme 2) was dissolved in minimum amount of dichloromethane and then precipitated in methanol; the polymer was then separated by centrifuge. This process was repeated to remove the initiator and unreacted monomer (found: C 67.0; H 5.6%. C₁₀H₁₀O₃ requires C 67.41; H 5.66%); ν_{max} (KBr) (Fig. S7[†]): 3444, 3066, 2899, 2772, 1608, 1475, 1435, 1248, 1090, 1026, 742, and 693 cm⁻¹.

2.5. Polymerization of AGE 7 (ref. 40 and 41)

A solution of AGE 7 (296 mg, 2.60 mmol), methyl-triphenylphosphonium bromide (8.2 mg, 0.023 mmol) in anhydrous toluene (1.6 mL) under Ar was cooled by ice bath, then triisobutylaluminium (0.24 mL, 0.26 mmol of 25 wt% in toluene) was added by a syringe under Ar. The polymerization was stopped after 2 h by adding 4 : 1 MeOH/H₂O (10 mL). After removing the solvents, the residue was dissolved in a CH₂Cl₂ (50 mL) and filtered over Celite 545. The filtrate was dried (Na₂SO₄) and evaporated to obtain polymer 8.

2.6. Activation of methylene acetal of safrole 1 as a model case^{42,43}

Pb(OAc)₄ (4.23 g, 9.5 mmol) was added under N₂ to a solution of safrole 1 (1.05 g, 6.48 mmol) in benzene (35 mL) at 75 °C and stirred for 3 h. The reaction mixture was taken up in EtOAc (30 mL) and washed with H₂O (3 \times 25 mL). The organic layer was dried (Na₂SO₄) and concentrated. The residual liquid upon silica gel chromatography using EtOAc/hexane afforded 9 as a faint yellow oil (1.08 g, 82%) (found: C 65.2; H 5.4%. C₁₂H₁₂O₄ requires C 65.45; H 5.49%); δ_{H} (CDCl₃): 2.09 (3H, s), 3.34 (2H, d, *J* 6.7 Hz), 5.05–5.09 (2H, m), 5.89–5.96 (1H, m), 6.77 (1H, dd, *J* 7.9 Hz, *J* 1.6 Hz), 6.83 (1H, d, *J* 1.6 Hz), 6.88 (1H, d, *J* 7.9 Hz), 7.66 (1H, s); δ_{C} (CDCl₃): 20.92, 39.80, 108.85, 109.64, 112.64, 115.92, 122.30, 134.88, 134.92, 137.16, 143.05, 144.83, 168.99, (CDCl₃ middle carbon: 77.00); ν_{max} (KBr) (Fig. S8[†]): 3079, 3005, 2979, 2914, 1838, 1768, 1639, 1495, 1445, 1376, 1255, 1216, 1181, 1100, 1009, 963, and 760 cm⁻¹.



Table 1 Polymerization^a of Safrole Oxide (SO) (2) initiated with MePPh₃Br (I) and catalyzed by ^tBu₃Al (C)^b

Entry	I (mmol)	C (mmol)	[SO]/[I]	[C]/[I]	Time (h)	Yield ^c (%)	<i>M</i> _{n,Theor} ^d	<i>M</i> _{n,Exp} ^e	PDI
1	0.0256	0.263	98	10.3	3	94	17 500		
2	0.0116	0.261	216	22.5	3	91	38 500	14 450	1.38
3	0.0458	0.264	55	5.76	6	95	9800	7500	1.45
4	0.0220	0.239	114	10.9	2	93	20 300	11 150	1.23
5	0	0.418	—	—	2	0			
6 ^f	0	0.418	—	—	24	0			
7 ^g	0.0331	0.416	76	12.6	3	69			
8	0.0227	0.725	110	31.9	12	70			
9	0.0222	0.235	113	10.6	12	65			
10	0.0328	0.403	76	12.3	2	99	17 000	9970	1.32

^a Polymerization was carried out at 0 °C using 2.5 mmol of monomer 2 with 1.8 mL of additional toluene except in entry 4 where no additional toluene was added. ^b 25 wt% solution in toluene (≈ 1 M ^tBu₃Al). ^c NMR indicates complete conversion to polymer where isolated yields are over 90%. ^d For entry 2: molar mass = [molar mass of SO] × [SO]/[I] = 178.19 × 216 = 38 489 (assuming 100% conversion). ^e GPC using light scattering detector. ^f Carried out at 20 °C. ^g 1 M ^tBu₃Al solution in hexane.

2.7. Synthesis of [(4-allyl-1,2-phenylene)bis(oxy)]bis(*tert*-butyldimethylsilane) (13) (Scheme 5)⁴⁴

TBDMSCl (25.0 g, 166 mmol) and DMAP (2.11 g, 17 mmol) were added under N₂ to a mixture of allylcatechol 12 (10.86 g, 72.4 mmol) (Scheme 5) and imidazole (23.6 g, 347 mmol) in DMF (130 mL) at 0 °C. After stirring at 25 °C for 16 h, the reaction mixture was taken up in water (300 mL) and extracted with ether (2 × 150 mL). After washing with water (4 × 200 mL), the ether layer was dried (Na₂SO₄). The removal of the solvent followed by distillation (bp_{0.2 mbarHg} 122 °C) afforded 13 as a colorless oil (20.5 g, 75%) (found: C 66.2; H 10.0%. C₂₁H₃₈O₂Si₂ requires C 66.60; H 10.11%); δ_H (CDCl₃): 0.17 (12H, s), 0.97 (18H, s), 3.24 (2H, d, *J* 6.7 Hz), 4.99–5.03 (2H, m), 4.99–5.03 (1H, m), 6.59 (1H, dd, *J* 1.8 Hz, *J* 8.3 Hz), 6.64 (1H, d, *J* 1.8 Hz), 6.72 (1H, d, *J* 8.3 Hz). δ_C (CDCl₃): -4.10, 18.44, 25.97, 39.45, 115.33, 120.83, 121.31, 121.47, 133.07, 137.83, 145.02, 146.59 (CDCl₃ middle carbon: 77.01). IR KBr (Fig. S9†): 2957, 2930, 2896, 2859, 1640, 1606, 1577, 1510, 1473, 1463, 1421, 1294, 1254, 1228, 1155, 1125, 986, 912, 839, and 781 cm⁻¹.

2.8. Synthesis of [(4-(oxiran-2-ylmethyl)-1,2-phenylene)bis(oxy)]bis(*tert*-butyldimethylsilane) (14)⁴⁵ (Scheme 5)

MCPBA (19.8 g, 80 mmol, 70% purity, 30% water) was dissolved in 150 mL CH₂Cl₂, and aqueous layer was removed. Silyl derivative 13 (16.3 g, 43 mmol) was added to the MCPBA/dichloromethane solution at 0 °C and stirred at 25 °C for 24 h. The reaction mixture was washed with 10% NaOH solution (3 × 20 mL) and water (2 × 25 mL). The CH₂Cl₂ layer was dried (Na₂SO₄), concentrated and distilled (bp_{0.2 mbarHg} 127–130 °C) to obtain 14 (10.7 g, 63%) as a colorless liquid (found: C 63.7; H 9.6%. C₂₁H₃₈O₃Si₂ requires C 63.90; H 9.70%); δ_H (CDCl₃): 0.17 (6H, s), 0.18 (6H, s), 0.966 (9H, s), 0.971 (9H, s), 2.49 (1H, q), 2.66 (1H, dd, *J* 14.7 Hz, *J* 5.5 Hz), 2.74–2.68 (2H, m), 3.06–3.09 (1H, m), 6.65 (1H, dd, *J* 8.3 Hz, *J* 2.2 Hz), 6.71 (1H, d, *J* 2.2 Hz), 6.74 (1H, d, *J* 8.3 Hz). δ_C (CDCl₃): (-) 4.11, 18.43, 25.94, 37.99, 46.72, 52.57, 120.93, 121.82, 121.89, 130.14, 145.56, 146.65, (CDCl₃ middle carbon: 77.02); ν_{max} (KBr) (Fig. S10†): 3044, 2930, 2897,

2859, 1765, 1727, 1701, 1607, 1577, 1514, 1473, 1464, 1423, 1292, 1256, 1224, 1159, 1127, 988, 907, 842, and 783 cm⁻¹.

2.9. Lewis acid catalyzed polymerization of 14

As described in Table 2, methyltriphenylphosphonium bromide was taken in an RB flask under Ar and closed with a rubber septum. Epoxidized silyl derivative 14 and anhydrous toluene were injected under Ar by a syringe. The mixture was cooled to 0 °C, then triisobutylaluminium solution (25 wt% in toluene) was injected. After the specified time (Table 2), the reaction mixture was quenched by adding 4 : 1 MeOH/H₂O mixture (10 mL). After extracting with CH₂Cl₂ (2 × 25 mL), the organic layer was dried (MgSO₄), filtered over Celite 545 and concentrated to obtain polymer 15a (Scheme 5). The polymer was purified by dissolving in ether and precipitating in MeOH; the process was repeated three times (found: C 63.6; H 9.8%. C₂₁H₃₈O₃Si₂ requires C 63.90; H 9.70%); ν_{max} (KBr) (Fig. S11†): 2930, 2896, 2859, 1607, 1578, 1518, 1473, 1427, 1422, 1362, 1305, 1254, 1224, 1160, 1128, 983, 852, 778, and 666 cm⁻¹.

2.10. Random copolymerization of safrole oxide 2 and allyl glycidyl ether 7

All the experiments were performed under argon atmosphere. As described in (Table 3), methyltriphenylphosphonium bromide was placed in an RB flask under Ar and closed with a rubber septum. Safrole oxide 2, allyl glycidyl ether 7 and anhydrous toluene were injected under Ar by a syringe. The mixture was cooled to 0 °C, and then triisobutylaluminium solution (25 wt% in toluene) was added under Ar in one portion by a syringe. The polymerization was stopped after complete polymerization (as indicated by ¹H NMR spectrum) by adding 4 : 1 MeOH/H₂O mixture (10 mL); the mixture was extracted with CH₂Cl₂, dried (MgSO₄), and filtered over Celite 545. The filtrate upon evaporation afforded random copolymer 16 (Scheme 6). The following IR data belong to a 1 : 1 copolymer of 2 and 7. ν_{max} (KBr) (Fig. S13†): 3076, 2869, 2773, 1646, 1608, 1500, 1443, 1353, 1249, 1121, 928, 808, and 774 cm⁻¹ (found: C



Table 2 Polymerization^a of silyl protected (SP) monomer **14** initiated with MePPh₃Br (**I**) and catalysed by ⁱBu₃Al (**C**)^b

Entry	I (mmol)	C (mmol)	[SP]/[I]	[C]/[I]	Time (h)	Yield ^c (%)	M _{n,Theor} ^d	M _{n,Exp} ^e	PDI
1	0.0213	0.24	117	11.3	1	0	—	—	
2	0.0213	0.70	117	32.9	2	60	27 700	16 300	1.8
3	0.0495	0.53	50	10.7	20	90	17 700	12 900	1.5
4	0.0092	0.53	272	57.6	40	75	80 200	11 500	1.6
5	0.0370	0.53	68	14.3	18	85	22 800	10 300	1.8

^a Polymerization was carried out at 0 °C for 6 h using 2.5 mmol of monomer **14** with 1.8 mL of additional toluene except in entry 4 where no additional toluene was used. ^b 25 wt% solution in toluene (≈ 1 M ⁱBu₃Al). ^c Isolated and NMR yields are similar within 2%. ^d M_n for entry 2 = [molar mass of SP] × [SP]/[I] × % conversion/100 = 394.70 × 117 × 0.60 = 27 708. ^e GPC using light scattering detector.

65.1; H 6.7%. Repeat units **2** and **7** in 1 : 1 requires C 65.74; H 6.90%).

2.11. Conversion of copolymer **16** to **17** (Scheme 6) using lead tetraacetate

The procedure as described under Section 2.7 was followed.^{42,43} A solution of polymer **16** (containing 15 mol% safrole oxide **2** repeating unit) (1.1 g, 8.9 mmol of total repeating units and 1.33 mmol of safrole oxide units) in benzene (25 mL) was heated to 75 °C in a round bottom flask. After the addition of Pb(OAc)₄ (0.90 g, 2.0 mmol) under N₂, the mixture was stirred at 75 °C for 4 h. Then, it was cooled, diluted with EtOAc (30 mL) and washed with H₂O (3 × 25 mL). The organic layer was dried (Na₂SO₄) and concentrated to give **17**. For a copolymer having 1 : 1 ratio of the monomer units: ν_{\max} (KBr) (Fig. S14†): 3080, 3013, 2919, 2867, 1763, 1645, 1496, 1446, 1352, 1215, 1105, 923, 786, 758 cm⁻¹. The polymers having various compositions gave satisfactory elemental analysis.

2.12. Synthesis of **18** (Scheme 6) via thiol-ene reaction

The copolymer **17** (prepared from entry 5, Table 3) (containing 85 mol% repeating unit of allyl glycidyl ether **7**) (274 mg, 2.2 mmol, containing 1.9 mmol of alkene motifs) was dissolved in THF (2 mL) and methanol (1.5 mL). Cysteamine·HCl (1.4 g, 12 mmol) and photoinitiator 2,2-dimethoxy-2-phenylacetophenone (DMPA) (227 mg, 0.9 mmol) were added. The mixture was purged with N₂ for 10 min, then irradiated with a 365 nm UV lamp until the completion of the reaction as revealed by ¹H NMR spectrum. The reaction mixture was taken up in 1 : 1 MeOH/0.1 M HCl (5 mL) and dialyzed against water.

The polymer solution upon freeze-drying afforded the deprotected polymer **18** (394 mg, 86%). (15 mol% catechol units) ν_{\max} (KBr) (Fig. S15†): 3469, 2960, 2923, 2857, 1624, 1494, 1384, 1259, 1105, 808, and 587 cm⁻¹ (found: C 45.6; H 7.9; N, 5.5; S, 12.1%). Repeat units **2** (diol form) and **7** (after thiolene reaction) in 15 : 85 requires C, 44.79; H, 7.75; N, 5.45; S, 12.47%).

2.13. Synthesis of **19** (Scheme 6) via thiol-ene reaction

The copolymer **17** (prepared from entry 5, Table 3) (containing 85 mol% repeating unit of allyl glycidyl ether **7**) (240 mg, 1.9 mmol, containing 1.6 mmol alkene motifs) was dissolved in THF (3 mL). Thioglycolic acid (1.00, 11 mmol) and photoinitiator 2,2-dimethoxy-2-phenylacetophenone (DMPA) (200 mg, 0.8 mmol) were added. After purging with N₂ for 10 min, the reaction mixture was irradiated with a 365 nm UV lamp until the completion of the reaction as revealed by ¹H NMR spectrum. The product dissolved in methanol was precipitated in chloroform. The process was repeated to give **19** (292 mg, 81%). The polymer was found to be insoluble in water. (15 mol% catechol units) ν_{\max} (KBr) (Fig. S16†): 3500 (br), 2916, 2869, 1734, 1601, 1520, 1471, 1111, 682, and 523 cm⁻¹ (found: C 48.1; H 6.5; S, 13.2%). Repeat units **2** (diol form) and **7** (after thiolene reaction) in 15 : 85 requires C, 48.88; H, 6.75; S, 13.61%).

2.14. Copolymerization of silyl protected **14** and AGE **7**

As described in (Table 4), silyl protected monomer **14**, allyl glycidyl ether **7** and anhydrous toluene were added onto methyltriphenylphosphonium bromide contained in a RB flask under Ar. The mixture was cooled to 0 °C and polymerization

Table 3 Random copolymerization^a of Safrole Oxide (SO) **2** and allyl glycidyl ether **7** initiated/catalyzed by MePPh₃Br/ⁱBu₃Al^b

Entry	SO ^c (mol%)	MePPh ₃ Br (mmol)	Toluene (mL)	ⁱ Bu ₃ Al (mmol)	Temp. (°C)	Time (h)	Yield (%)
1	50	0.063	2.5	0.835	20	20	91
2	50	0.065	2.0	0.860	20	24	93
3	50	0.025	1.0	0.484	0	2	89
4 ^d	50	0.042	3.0	0.513	0	18	92
5 ^e	15	0.040	2.5	0.534	0	20	93

^a Polymerization was carried out using a total of 5.0 mmol of monomer **2** and **7**. ^b 25 wt% solution in toluene (≈ 1 M ⁱBu₃Al). ^c Mol% of the monomer in the mixture of the two monomers. ^d This reaction was run using a total of 35 mmol of **2** and **7**, however, calculations are shown for the total of 5.0 mmol. ^e This reaction was run using a total of 50 mmol of **2** and **7**, however, calculations are shown for the total of 5.0 mmol.



Table 4 Random copolymerization^a of silyl protected oxide (SP) **14** and allyl glycidyl ether (AGE) **7** initiated with MePPh₃Br (**I**) and catalyzed by ⁱBu₃Al (**C**)^b

Entry	SP ^c (mol%)	I (mmol)	[M] ^d /[I]	C (mmol)	[C]/[I]	Time (h)	Conv. ^{e,f} (%)	M _{n,Theor} ^g	M _{n,Exp} ^h	PDI
1	50	0.045	111	0.76	17	3	88 (88)			
2	50	0.053	94	0.76	14	5	95 (99)	23 900	14 300	1.5
3	25	0.045	111	0.50	11	12	99 (99)	20 500	12 700	1.3
4	10	0.045	111	0.50	11	12	96 (99)	15 800	6100	2.0
5	10	0.092	54	0.50	5	3	84 (99)			
6	10	0.012	417	0.50	42	15	78 (93)			
7 ⁱ	10	0.032 in	156	0.50	16	17	99 (99)	22 200	7900	1.8
8	5	0.045	111	0.50	11	12	98 (99)	14 200	5900	2.1

^a Polymerization was carried out at 0 °C using a total of 5.0 mmol of monomer **14** and **7** with an additional amount toluene added (3.5 mL). ^b 25 wt% solution in toluene (≈ 1 M ⁱBu₃Al). ^c Mol% of the monomer the mixture of the two monomers. ^d Total monomers of silyl protected oxide **14** and allyl glycidyl ether **7** is 5 mmol. ^e % conversion monomer **14** as determined by ¹H NMR, the number in parentheses belongs to % conversion of AGE **7**. ^f Isolated yield was in the range 85–95%. ^g M_n for entry 4 = [molar mass of SP × 0.10 + molar mass of AGE × 0.90] × [M]/[I] = 142.2 × 111 = 15 784 (assuming 100% conversion). ^h GPC with a light scattering detector. ⁱ This reaction was run using a total of 30 mmol of **14** and **7**; however, calculation is based on a total of 5.0 mmol of monomers **14** and **7**.

was started with the addition of triisobutylaluminium solution (25 wt% in toluene). The polymerization was stopped (after its completion as indicated by ¹H NMR spectrum) by adding 4 : 1 MeOH/H₂O mixture (10 mL) and extracted with CH₂Cl₂ (25 mL). The organic layer was dried (MgSO₄), filtered over Celite 545, and concentrated to obtain **20** (Scheme 8). The polymer was dissolved in diethyl ether, precipitated in methanol and separated by centrifuging. This process was repeated to obtain pure **20**. (25 mol% silyl protected, entry 3, Table 4): ν_{max} (KBr) (Fig. S17†): 3081, 3014, 2855, 1646, 1605, 1576, 1512, 1469, 1422, 1358, 1298, 1255, 1224, 1160, 984, 839, 782, 696, and 667 cm⁻¹ (found: C 62.3; H 9.0%. Repeat units **14** and **7** in 25 : 75 requires C, 63.55; H, 9.30%).

2.15. Silyl deprotection of polymer 15a

TBAF (1.0 M in THF) (1.26 mL, 1.26 mmol) was added to a solution of polymer **15a** (entry 4, Table 2) (227 mg, 0.575 mmol) in THF (4 mL) under N₂ at 0 °C. After stirring at 0 °C for 1 h, a 1 : 1 mixture of H₂O/acetic acid (2 mL) was added to the reaction mixture. The work-up procedures included the evaporation of the solvents, redissolving the residue in MeOH and precipitation in water. The process was repeated two times to wash out remaining TBAF to obtain deprotected polymer **15b** (Scheme 5), (yield: 72 mg, 75%). The fluoride ion could chelate with catechol moiety according to literature.^{46,47} The polymer was soluble in methanol but insoluble in H₂O, Et₂O and CH₂Cl₂ (found: C 64.7; H 5.8%. C₉H₁₀O₃ requires C 65.05; H 6.07%); ν_{max} (KBr) (Fig. S12†): 3445, 2921, 2870, 1608, 1521, 1446, 1363, 1285, 1190, 1113, 1052, 958, 871, 811, 788, and 754 cm⁻¹.

2.16. Synthesis of 21 (Scheme 8) via thiol-ene reaction (10 mol% catechol units)⁴⁸

The copolymer **20** (entry 7, Table 4) (690 mg, containing 4.32 mmol of alkene motifs) was dissolved in THF (18 mL) and methanol (4.5 mL). Cysteamine·HCl (3.6 g, 33 mmol) and photoinitiator 2,2-dimethoxy-2-phenylacetophenone (DMPA) (582 mg, 2.4 mmol) were added. After purging with N₂ for 10 min, the mixture was irradiated with a 365 nm UV lamp at

25 °C until the reaction is completed. The reaction mixture was dialyzed against MeOH (to remove DMAP) for 30 min and then against deionized water (1 h). The resultant polymer solution was freeze-dried to obtain polymer **21** (1.11 g, 90%). For a sample of **21** derived from entry 2 (Table 4) (50% silyl): (found: C, 55.1; H, 8.9; N, 2.1; S, 4.8%. **21** in a 1 : 1 ratio of the repeating units requires C, 55.96; H, 9.07; N, 2.25; S, 5.15%); ν_{max} (KBr) (Fig. S18†): 3430, 2962, 2927, 2857, 1637, 1511, 1467, 1422, 1385, 1301, 1256, 1221, 1162, 1100, 981, 910, 840, 778, 664, and 614 cm⁻¹.

2.17. Synthesis of 22 (Scheme 8) via thiol-ene reaction (10 mol% catechol units)

The reaction was performed for polymer samples **20** having SP **14** and AGE **7** repeating units in mole ratios of 50 : 50 and 10 : 90 to obtain polymer **22** using UV light in dark room.

Thioglycolic acid (1.88 g, 20.4 mmol) and photoinitiator DMPA (522 mg, 2.1 mmol) were added to a solution of copolymer **20** (entry 7, Table 4) (618 mg, 4.05 mmol alkene motifs) in THF (12 mL). After purging the mixture with N₂ for 10 min, it was then irradiated with a 365 nm UV lamp at 25 °C until the reaction is completed. The THF-soluble product was precipitated with the addition of ether to the reaction mixture. The process of dissolving in THF followed by precipitation in ether was repeated three times to obtain **22** (885 mg, 84%). For a sample of **22** derived from entry 2 (Table 4) (50% silyl): (found: C, 57.2; H, 8.8; S, 5.1%. **22** in a 1 : 1 ratio of the repeating units requires C, 57.96; H, 8.72; S, 5.33%); IR for a 50 : 50 copolymer **22** is given: (50% silyl) ν_{max} (KBr) (Fig. S19†): 3458, 2931, 2858, 2511, 1727, 1632, 1511, 1467, 1421, 1385, 1301, 1255, 1223, 1122, 983, 910, 841, 790, 669, and 552 cm⁻¹.

2.18. Synthesis of 18 from 21 (catechol 10 mol%)

The copolymer **21** as obtained from **20** (1.0 mmol) (entry 7, Table 4) using procedure described under Section 2.16, was dissolved in MeOH/0.5 M HCl (6 mL) and stirred under N₂ at 25 °C for 18 h. The reaction mixture was dialyzed against water and then freeze-dried to obtain polymer **18** in (90%) (found: C,



43.0; H, 8.0; N, 5.5; S, 12.7%. **18** containing 10 mol% catechol units requires C, 43.90; H, 7.82; N, 5.69; S, 13.02%). IR and NMR spectra of **18** (10 mol% catechol repeating unit) were found to be like the one prepared earlier from saffrole-oxide under Section 2.14. Acid hydrolysis of silyl groups in 1 : 1 copolymer **21** (*i.e.* containing 50 mol% catechol units) to get polymer **18** could not be achieved because of solubility problem.

2.19. Synthesis of **19** from **22** (catechol 10 mol%)

TBAF (1.0 M in THF) (0.25 mL, 0.25 mmol) was added to a solution of polymer **22** (50 mg, 0.22 mmol) in THF (1.0 mL) under N₂ at 0 °C and stirred for 12 h. After the elapsed time, a 2 : 1 mixture of H₂O/acetic acid (2 mL) was added. The crude polymer was soluble in methanol but insoluble in H₂O. Therefore, it was purified by dissolving in methanol and precipitating in water. The process was repeated two times to obtain polymer **19** (yield: 90%) (found: C, 47.5; H, 6.9; S, 13.9%. **19** containing 10 mol% catechol units requires C, 48.10; H, 6.78; S, 14.27%). Removal of the acid hydrolysis of silyl groups in 1 : 1 copolymer **22** (*i.e.* containing 50 mol% catechol units) to get polymer **19** could not be achieved because of solubility problem. IR and NMR spectra of **19** (10 mol% silyl) were found to be like the one prepared earlier from saffrole-oxide under Section 2.15.

2.20. Coacervate formation (10% catechol unit)

A sample of **19** (514 mg) was dissolved in MeOH and dialyzed against NaHCO₃ solution. After 1 h, the polymer became soluble and the dialysis was continued against water for 24 h; the polymer remained soluble. A solution of **18** (494 mg) in water was added to the dialyzed solution; the resultant turbid mixture was dialyzed against water for 24 h. The turbid solution was freeze-dried to obtain the **23** (Scheme 8), the coacervate of **18** and **19** (1.00 g). ν_{\max} (KBr) (Fig. S20[†]): 3456, 2919, 2869, 1733 (w), 1620 (overlapping), 1578, 1461, 1384, 1280, 1226, 1110, 780, 900 and 701 cm⁻¹.

3 Results and discussion

Natural saffrole **1** upon oxidation with MCPBA was converted to its epoxide **2** (Scheme 1). Fig. 1a and 2a displays the ¹H and ¹³C NMR spectra of **2**. Numerous attempts were made to polymerize saffrole oxide **2** using various basic catalyst (see ESI[†]). The results are given in (Table S1[†]). The ring opening polymerization monomer **2** as neat or in solvent THF was carried out under argon using TBAF (1–6 mol%) or TBAH (2–5 mol%) or NaO^tBu as initiators at 50–100 °C for 2–72 h. Under the reaction conditions, the monomer was recovered unreacted in the case of initiator NaO^tBu (Table S1,† entry 14). TBA-initiated polymerization also failed to give any polymer (entries 1 and 2). In other cases, we were unable to obtain the expected polymer **5** cleanly; extensive base catalysed elimination reaction occurred. Use of TBAH as the initiator led to the formation of polymer in high yields in the temperature range 70–100 °C; however, the proton NMR analysis revealed the presence of alkene motifs (\approx 10 mol%) on the polymer terminal (Fig. S1b[†]). Here, the F⁻ or OH⁻ or the ring opened alkoxide ion (RO⁻) is involved in

a competing reaction between nucleophilic ring opening and base catalysed elimination reaction *via* abstraction of labile benzylic protons as depicted in Scheme 1. The elimination process was very extensive with F⁻ as confirmed by appearance of ¹H signal at δ 6.2, 6.5 and 4.1 ppm attributed to the –CH=CH–CH₂–O motifs (Fig. S1b[†]). For the sake of comparison, the ¹H NMR spectrum of cinnamyl alcohol is shown in Fig. S1c.† The signals for the protons marked ‘a’, ‘b’ and ‘c’ are readily identifiable for comparison, thereby confirming the base catalysed chain transfer to monomer **2** by abstraction of benzylic protons leading to alkene motifs akin to the motifs of cinnamyl alcohol. With TBAH, ring opening polymerization happened accompanied by chain transfer *via* elimination process as depicted by **A** (Scheme 1). The extensive chain transfer to monomer led us to pursue this important polymerization using other protocols with the objective of minimizing the chain transfer, which puts a limit on the maximum molar mass possible for the polymers.

In the precedent literature, considerable efforts have been exerted to control the living character of the polymerization of propylene oxide (PO). In most cases, alkali metal alkoxides and hydroxides are widely used as anionic polymerization initiators.^{49–51} The high basicity of propagating species (akin to an alkoxide, RO⁻) leads to the abstraction of proton from methyl group of the PO, thereby constituting a chain transfer to the monomer. This process also occurred in the current work involving the base catalysed polymerization of saffrole oxide (SO) **2**, thereby resulting in the extensive formation of SO oligomers possessing a terminal cinnamyl unsaturation (Scheme 1). One significant problem inherent in SO is the much higher acidity of the benzylic protons than that of CH₃ protons in PO; the labile H in SO is, therefore, much more prone to base catalysed abstraction than the methyl protons in PO.

Polymerization reaction of **2** using Lewis acid triisobutylaluminum as a catalyst and methyltriphenylphosphonium bromide (**I**) as an initiator^{38,39,52} gave ring opened polymer **6** in excellent yields (Scheme 2), (Table 1). The ¹H NMR spectra (Fig. 1c) revealed the formation of ring opened polymer without any indication of chain transfer reaction *via* abstraction of allylic proton.

Experimental evidence suggests the requirement of [AlR₃]/[I] \geq 1 for successful polymerization. Trialkylaluminum participates in the formation of an aluminate complex as depicted by ‘A’ (Scheme 3) which by itself is not reactive enough to effectively initiate and/or propagate the current SO polymerization. An excess of ⁱBu₃Al ensures a fast polymerization at 0 °C owing to activating effect of the “free” triisobutylaluminum derivative as depicted by **B**. Note that the experimental molar masses of poly(saffrole oxide) (PSO) are not close to theoretical values based on the formation of one polymer chain per Ph₃MeP⁺Br⁻ (see Table 1). In the context of molar masses by GPC, it is worth mentioning that the chemical structures of the standards (PEO or PS) and the polymers/copolymers are different. As such (for a note of caution), the investigated polymers/copolymers and standards may not have the same hydrodynamic radii for the same molar mass. However, using the same standards, GPC has been extensively used to determine the molar masses of



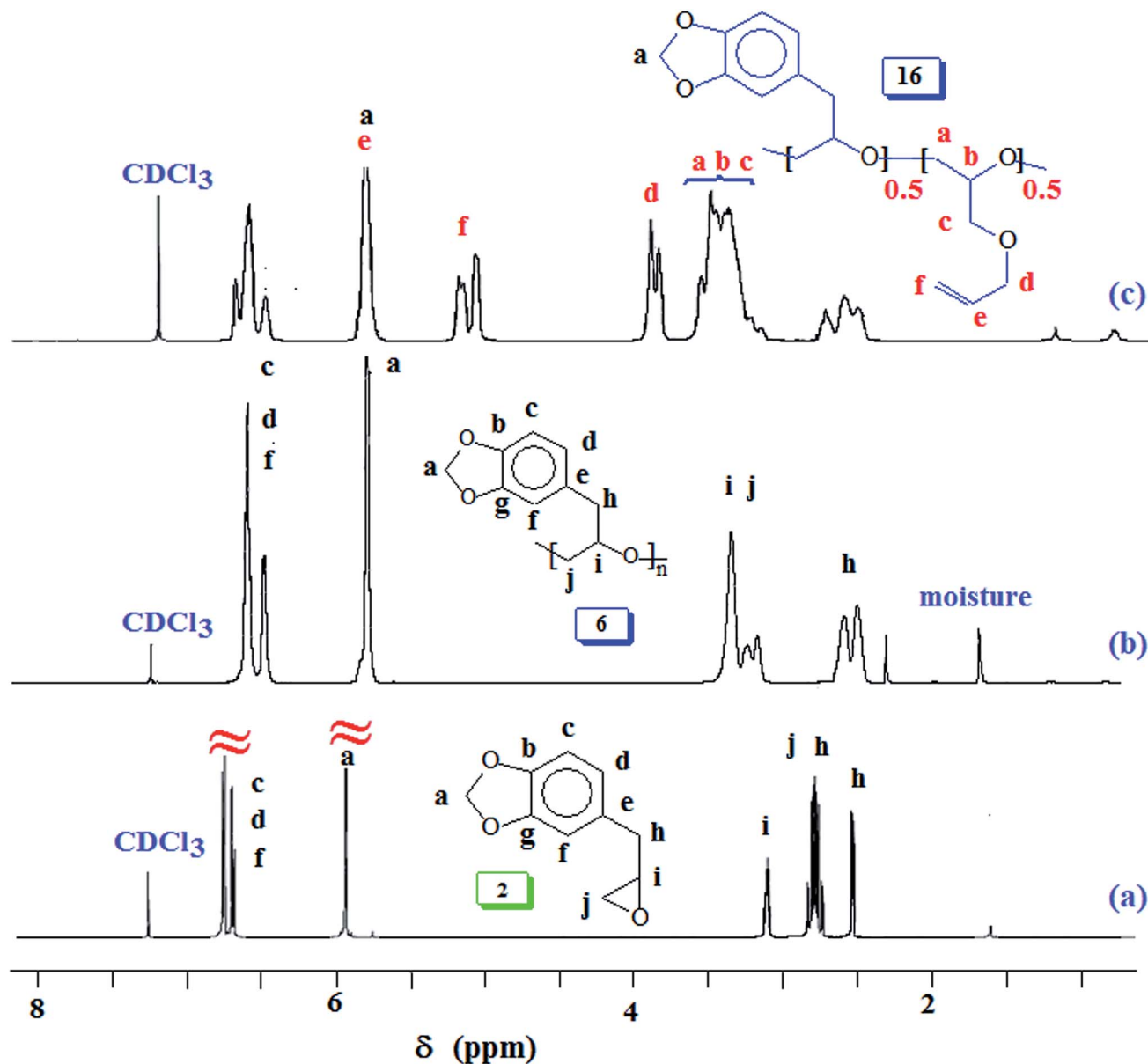


Fig. 1 ^1H NMR spectra of (a) safrole oxide 2, (b) polymer 6, and (c) copolymer 16.

polymers/copolymers derived from epoxide containing a range of substituents.^{38,39,53,54} The measured molar masses have then been compared with the theoretical values as described for the current polymers.

Thus, the living-like polymerization of SO is jeopardized by a significant contribution of the chain transfer process to the monomer. For the PO polymerization, it has been reported, however, that the number of PPO chains remains identical to the number of $\text{Ph}_3\text{MeP}^+\text{Br}^-$ molecules.³⁸

The polymerization mechanism may thus involve the electrophilic complex **B** which is inserted into the nucleophilic species **C** (Scheme 3). The involvement of reactive complex **B** minimizes the transfer process to monomer SO, as observed in the cases involving alkali metal alkoxide initiators. The greater electron-withdrawing effect of R_3Al in **B** imparts greater positive

charges on the ring-carbons involved in the ring opening process, than on the benzyl hydrogens involved in the transfer process to monomer (Scheme 1). Also, note that the basicity of alkoxide species in alkoxy aluminate complexes **B** or **C** is greatly reduced as compared to alkali metal alkoxide species, thereby inhibiting the proton abstraction reaction leading to chain transfer to the monomer.

It is reported that the initiation process involved the attack by not only bromide but also by the isobutyl group and hydride as shown in Scheme 3,^{39,52} thereby leading to three polymers having bromide, isobutyl, and hydride end groups.⁵² $^i\text{Bu}_3\text{Al}$ can work both as a hydride as well as an isobutyl anion in the ring-opening reaction.^{55,56}

Generally, propagation *via* nucleophilic attacks on the less hindered methylene carbon of epoxides leads to regioregular



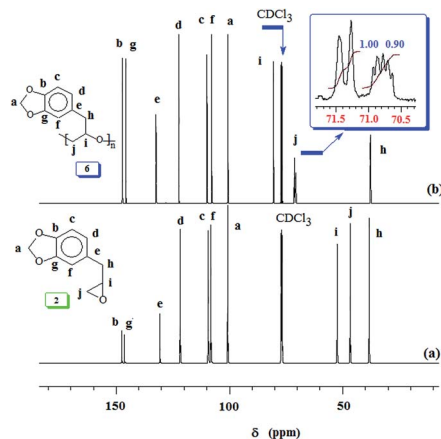
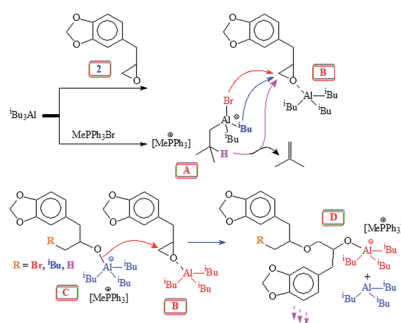
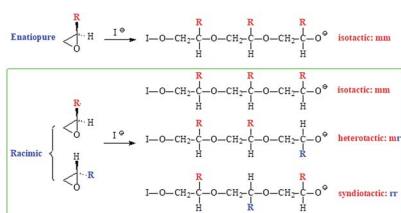


Fig. 2 ^{13}C NMR spectra of (a) safrole oxide 2 and (b) polymer 6 in CDCl_3 .



Scheme 3 Mechanism of Lewis acid catalyzed polymerization.

polymers.^{57,58} Optically pure (*i.e.* enantiopure) epoxides are thus expected to provide an easy access to isotactic polymers since the methine carbon retains its configuration (Scheme 4). However, for a racemic epoxide, formation of an atactic polymer is expected with equal percentages of mm, mr, rm and rr triads (Scheme 4). ^{13}C NMR MR spectrum revealed the regioregularity and stereochemistry of monomer insertion. The observed methine and methylene carbon signals ensure the exclusive head-to-tail (HT) chain propagation.⁵⁹ In the current work, while the methine carbon marked 'i' appeared as single signal, the methylene carbon marked 'j' resolved into triads and diads (Fig. 2b). The ^{13}C NMR spectrum thus showed the current polymer 6 as atactic (Scheme 4).^{38,59,60}



Scheme 4 Regioregular ring opening polymerization of enantiopure and racemic epoxide.

Our research plan includes the synthesis of copolymers of monomers safrole oxide 2 and allyl glycidyl ether 7 (Scheme 2). In this context, triisobutylaluminum-methyltriphenylphosphonium bromide catalyst-initiator system was used to homopolymerize 7 to obtain polymer 8 (Scheme 2). The ^1H and ^{13}C NMR spectra of 7 and 8 are shown in respective Fig. S2 and S3.† The spectral data is consistent with the regioregular ring opening to give 8. As in the case of safrole oxide polymer 6, the splitting of carbon marked 'a' in the ^{13}C NMR spectrum (Fig. S3b†) pointed toward the formation of an atactic microstructure.

At this stage, we were apprehensive about the deprotection of methylene acetal protecting group in PSO 6. Methylene acetal is indeed a robust protective group which does not respond to acid catalyzed deprotection. As a model case, safrole 1 was used for examining the deprotection aspect using lead tetraacetate.^{42,61} To our relief, safrole 1 on treatment with the oxidizing agent afforded 9 having labile acetoxy group (Scheme 2). In fact, the similar acetoxy derivatives are usually removed during aqueous work up leading to catechol motifs. The ^1H and ^{13}C NMR spectra of 1 and 9 are shown in respective Fig. 3 and S4.† The methylene protons of 1 marked 'a' (Fig. 3a) is shifted downfield in the spectrum of 9 (Fig. 3b) owing to the presence of electron withdrawing AcO substituent. Similar downfield shift is observed for the carbon marked 'j' in the ^{13}C NMR spectrum (Fig. S4b†).

Unaware of the AcO group's ability in 9 (Scheme 2) to survive during subsequent chemical transformation, we set out to explore the related chemistry using silyl protecting group. In this context, natural product eugenol 10 was demethylated and protected by reacting with diphenylsilane in the presence of catalyst tris-pentafluorotriphenylborane to give 11 (Scheme 5). To our dismay, silyl protected 11 was found to be extremely moisture sensitive; it broke down during silica gel chromatography to give allylcatechol 12 quantitatively. Thereafter, we decided to prepare 12 *via* demethylation using LiCl. The 'OH' groups in 12 were then protected to give 13 by reacting with *tert*-butyldimethylsilyl chloride in the presence of imidazole. Epoxidation using *m*-chloroperbenzoic acid transformed 13 to 14

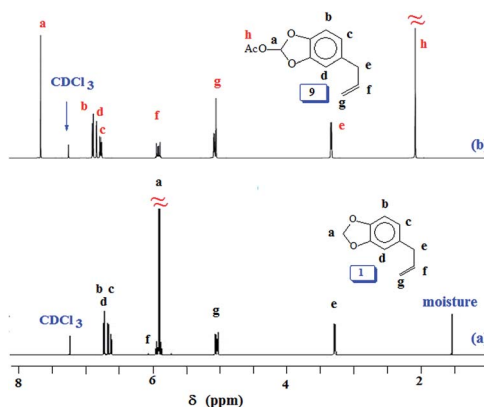
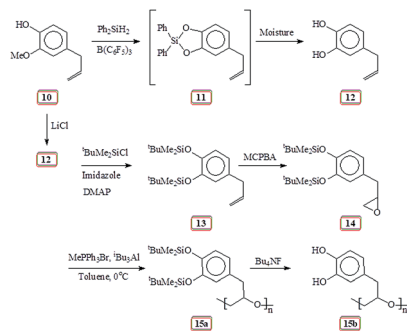


Fig. 3 ^1H NMR spectra of (a) safrole oxide 1 and (b) acetoxy safrole 9 in CDCl_3 .



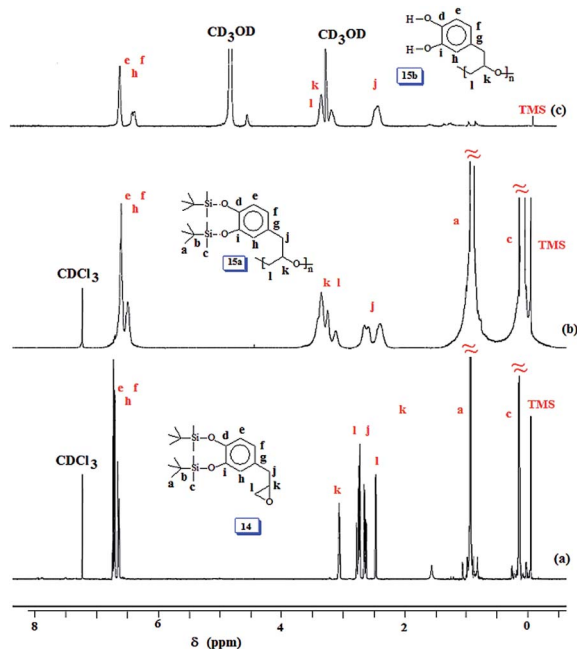
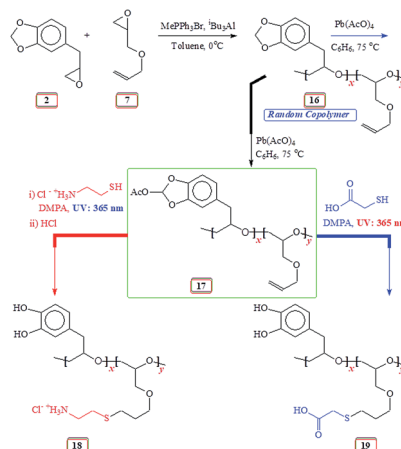


Scheme 5 Lewis acid catalyzed polymerization of silylated epoxide 14.

which was then subjected to Lewis acid catalysed polymerization to afford **15a**. The results of the polymerization reaction are given in (Table 2). The *tert*-butyldimethylsilyl groups in **15a** were then deprotected using tetrabutylammonium fluoride to obtain **15b** containing the catechol motifs.

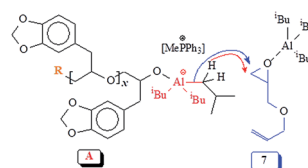
The ^1H and ^{13}C NMR spectra of **14**, **15a** and **15b** are shown in respective Fig. 4 and S5.† The spectra confirmed the structures of the monomer and polymers. The silyl protons and carbons marked 'a', 'b' and 'c' in Fig. 4a, b, S5a and b† disappeared upon deprotection of the silyl groups as indicated by the absence of the corresponding signals in Fig. 4c and S5c.† However, it was very difficult to remove the F^- as it is known to bind strongly with catechol motifs *via* H-bonding.^{46,47} In such a scenario, the ammonium counterion remained with the polymer sample as can be seen as minor peaks in the region around δ 1 ppm (Fig. 4c). Minor carbon signals are also observed in Fig. S5c.†

Next, we focussed our attention to Lewis acid catalysed copolymerization reaction of SO (**2**) and AGE (**7**) as outlined in

Fig. 4 ^1H NMR spectra of (a) **14** and (b) **15a** in CDCl_3 ; and (c) **15b** in CD_3OD .Scheme 6 Lewis acid catalyzed copolymerization of epoxides **2/7** and thiol-ene reaction.

Scheme 6. Numerous attempts to obtain block copolymers by sequential addition of the monomers resulted in failures. At every instance, the reaction resulted in the formation of a mixture of homopolymers which were separated as explained in the Experimental section. A plausible rationale is illustrated in Scheme 7 where the hydride or butyl transfer to the second monomer might be able to initiate new chain thereby resulting in the formation of two homopolymers. We, therefore, shifted our attention to obtain the random copolymer from monomers **2** and **7** as shown in Scheme 6. Lewis acid catalysed polymerization of **2** and **7** afforded random copolymer **16** in excellent yields. The results are given in (Table 3). The ^1H spectra of homopolymer **6** and the 1 : 1 **2/7** random copolymer **16** are displayed in Fig. 1b and c. The feed ratio of the monomers matched with incorporated ratio of the corresponding repeating units as determined by integration of several non-overlapping proton signals Fig. 1c. The finding thus implies that the two monomers have equal reactivity ratio.

The 1 : 1 random copolymer **16** was then subjected to lead tetraacetate oxidation thereby giving copolymer **17** containing acetoxy group (*vide supra*, Scheme 6). The ^1H and spectra of **16** and **17** are displayed in Fig. 1c and 5a, respectively. The proton marked 'a' (Fig. 1c) is shifted downfield at 'c' (Fig. 5a) and the new CH_3CO protons marked 'd' appeared at δ 2 ppm as expected. The ^{13}C NMR spectrum (Fig. S6†) also revealed the formation of acetoxy derivative **17**. The acetoxy carbons marked 'r' and 'q' appeared at δ 20.5 and 170.3 ppm, respectively (Fig. S6b†). Also note that carbon marked 'a' in **17** is shifted downfield as compared to 'a' of **16** respectively (Fig. S6a†).

Scheme 7 A plausible hydride or isobutyl transfer to activated monomer **7**.

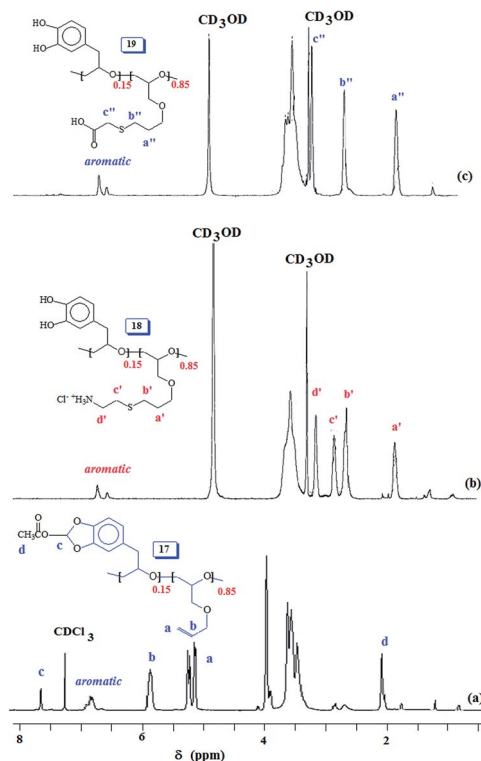
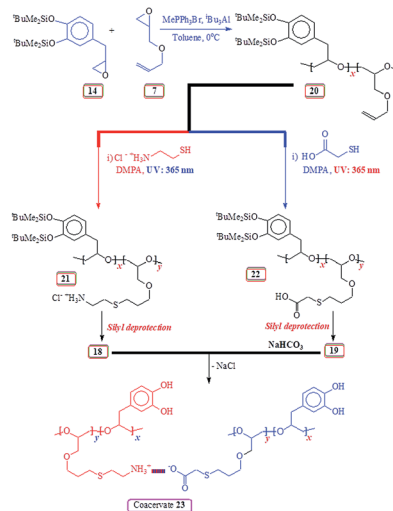


Fig. 5 ^1H NMR spectra of 15 : 85 copolymer (a) 17 in CDCl_3 , (b) 18 and (c) 19 in CD_3OD .

The acetoxy derivative 17 was subjected to thiol-ene reaction using photoinitiator 4-dimethylaminopyridine (DMAP) using UV light ($\lambda = 365 \text{ nm}$). The addition of cysteamine·HCl ($\text{H}_3\text{N}^+\text{CH}_2\text{-CH}_2\text{SH Cl}^-$) and thioglycolic acid ($\text{HO}_2\text{CCH}_2\text{SH}$) to 17 with a 15 : 85 ratio of SO 2/AGE 7 repeating units converted 17 to 18 and 19, respectively (Scheme 6). The ^1H NMR spectra of 17, 18 and 19, displayed in Fig. 5 confirms the successful transformation. The acetoxy proton signal (marked 'd', Fig. 5a) disappeared in the spectra of 18 and 19 (Fig. 5b and c) as a result of hydrolysis of the functional motifs during aqueous workup under mild acidic condition. This is further confirmed by the absence of the signal (for proton marked 'c', Fig. 5a) in the spectra of 18 and 19 (Fig. 5b and c). The spectra are consistent with the addition of the thiol motifs into the alkene double bond as confirmed the absence of alkene proton signals in Fig. 5b and c. The aromatic protons are visible; chemical shifts of some of the readily identifiable protons are assigned. The work presented in the previous paragraph are repeated using silyl protected epoxide 14 which was copolymerized with allyl glycidyl ether 7 (Scheme 8) to give copolymer 20 having various proportions of the x/y units. The results of the polymerization are given in (Table 4). The silyl protected copolymer was then elaborated using thiol-ene reaction as discussed before to give 21 and 22, which on treatment with HCl and Bu_4NF respectively afforded 18 and 19 after deprotection of the silyl groups. The spectral analysis revealed the identical nature of the polymers to the polymers derived *via* acetoxy protective groups (*vide supra*). The ^1H NMR spectra of random copolymer 20 with SP/AGE ratio of 1 : 1 and 0.10 : 0.90 are shown in Fig. 6a and b, respectively. Fig. 6c and



Scheme 8 Lewis acid catalyzed copolymerization of epoxides 14/7 and thiol-ene reaction.

d display the spectra for 21 and 22; the absence of alkene protons in the range δ 5–6 ppm clearly confirms the addition of the thio group onto the double bonds.

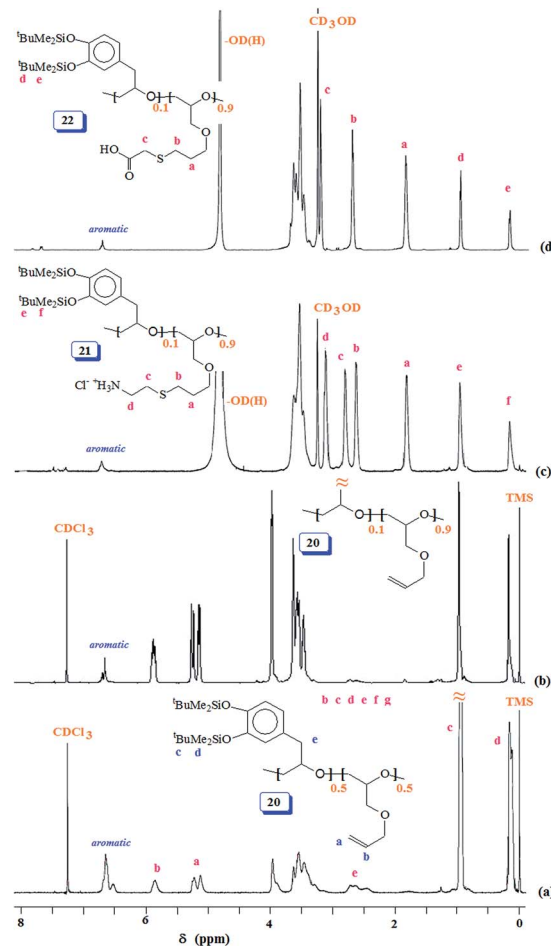


Fig. 6 ^1H NMR spectra of (a) 50 : 50 copolymer 20 in CDCl_3 , (b) 10 : 90 copolymer 20 in CDCl_3 , (c) 10 : 90 copolymer 21 in CD_3OD , and (d) 10 : 90 copolymer 22 in CD_3OD .



Coacervate **23**, obtained by treating aqueous solutions of HCl salt **18** and Na-salt **19**, was found to be insoluble in water (Scheme 8). The strong interaction leading to the **23** is evinced by inability of various concentration of NaCl to dislodge the component polymers from the coacervate.

4 Conclusions

The ring opening polymerization of safrole oxide **2**, derived from naturally occurring safrole **1**, using basic catalyst led to the formation of intractable materials as a result of extensive chain transfer reaction. However, safrole oxide **2**, for the first time, underwent homo- as well as copolymerization with AGE **7** using the Lewis acid initiator/catalyst comprising of triphenylmethylphosphonium bromide/triisobutylaluminum. A mechanism has been proposed for the chain transfer reaction to monomer and the inability to form block copolymers. The random copolymer **16** of various compositions (from SO **2**/AGE **7**), obtained in excellent yields, were elaborated by activation of methylene acetal with acetylation to give **17** followed by incorporating anionic (CO_2^-) and cationic (NH_3^+) group on the terminal of the pendants by thiol-ene reaction to generate. The polymer backbones of **18** and **19** having charges of opposite algebraic signs was used for the formation of complex coacervate by electrostatic attraction. Such complex is expected to be adsorbed on a surface where the catechol motifs would stabilize the adhesion. A way has been found to remove the acetal protective group under mild conditions using lead tetraacetate to generate catechol moieties on the polymer backbone. Use of *tert*-butyldimethyl groups as a protective group for the catechol motifs has also been explored.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

The authors would like to acknowledge the support provided by King Abdulaziz City for Science and Technology (KACST) through the Science & Technology Unit at King Fahd University of Petroleum & Minerals (KFUPM) for funding this work through project no. 12-ADV2397 as part of the National Science, Technology and Innovation Plan. The authors would like to acknowledge the valuable guidance of Prof. Craig Hawker in conducting this research work.

References

- J. Comyn, The relationship between joint durability and water diffusion, in *Developments in Adhesives*, ed. A. J. Kinloc, Appl. Sci., Barking UK, 1981, vol. 2, pp. 279–313.
- P. Lee Bruce, P. B. Messersmith, J. N. Israelachvili and J. H. Waite, *Annu. Rev. Mater. Res.*, 2011, **41**, 99–132.
- A. V. Pocius, *Adhesives and Adhesives Technology*, Hanser/Gardner, Cincinnati, 1997.
- B. K. Ahn, *J. Am. Chem. Soc.*, 2017, **139**, 10166–10171.
- K. Zhang, F. Zhang, Y. Song, J. B. Fan and S. Wang, *Chin. J. Chem.*, 2017, **35**, 811–820.
- K. Zhang, F. Zhang, Y. Song, J. Fan and S. Wang, *Chin. J. Chem.*, 2017, **35**, 811–820.
- N. K. Kaushik, N. Kaushik, S. Pardeshi, J. G. Sharma, S. H. Lee and E. H. Choi, *Mar. Drugs*, 2015, **13**, 6792–6817.
- Z. Wang, Y. Duan and Y. Duan, *J. Controlled Release*, 2018, **290**, 56–74.
- H. Yamamoto, S. Yamauchi and S. Ohara, *Biomimetics*, 1992, **1**, 219–238.
- H. Tatehata, A. Mochizuki, T. Kawashima, S. Yamashita and H. Yamamoto, *J. Appl. Polym. Sci.*, 2000, **76**, 929–937.
- M. Yu, J. Hwang and T. J. Deming, *J. Am. Chem. Soc.*, 1999, **121**, 5825–5826.
- M. Yin, Y. Yuan, C. Liu and J. Wang, *Biomaterials*, 2009, **30**, 2764–2773.
- T. H. Anderson, J. Yu, A. Y. Estrada, M. U. Hammer, J. H. Waite and J. N. Israelachvili, *Adv. Funct. Mater.*, 2010, **20**, 4196–4205.
- H. Zhao, C. J. Sun, R. J. Stewart and J. H. Waite, *J. Biol. Chem.*, 2005, **280**, 42938–42944.
- H. Shao and R. J. Stewart, *Adv. Mater.*, 2010, **22**, 729–733.
- H. Lee, B. P. Lee and P. B. Messersmith, *Nature*, 2007, **448**, 338–341.
- B. P. Lee, K. Huang, F. N. Nunalee, K. R. Shull and P. B. Messersmith, *J. Biomater. Sci., Polym. Ed.*, 2004, **15**, 449–464.
- G. Westwood, T. N. Horton and J. J. Wilker, *Macromolecules*, 2007, **40**, 3960–3964.
- H. C. Yang, J. Luo, Y. Lv, P. Shen and Z. K. Xu, *J. Membr. Sci.*, 2015, **483**, 42–59.
- Y. Tian, Y. Cao, Y. Wang, W. Yang and J. Feng, *Adv. Mater.*, 2013, **25**, 2980–2983.
- F. Zhang, S. Liu, Y. Zhang, Y. Wei and J. Xu, *RSC Adv.*, 2012, **2**, 8919–8921.
- J. Li, M. Zhou, F. Cheng, Y. Lin and P. Zhu, *Carbohydr. Polym.*, 2019, **221**, 113–119.
- X. Chen, Y. Zhai, X. Han, H. Liu and Y. Hu, *Appl. Surf. Sci.*, 2019, **483**, 399–408.
- J. Duan, W. Wu, Z. Wei, D. Zhu, H. Tu and A. Zhang, *Green Chem.*, 2018, **20**, 912–920.
- X.-D. Pan, Z. Qina, Y.-Y. Yana and P. Sadhukhana, *Polymer*, 2010, **51**, 3453–3461.
- H. Shao, K. N. Bachus and R. J. Stewart, *Macromol. Biosci.*, 2009, **9**, 464–471.
- R. Ko, P. A. Cadieux, J. L. Dalsin, B. P. Lee, C. N. Elwood and H. Razvi, *J. Endourol.*, 2008, **22**, 1153–1160.
- B. P. Lee, J. L. Dalsin and P. B. Messersmith, *Biomacromolecules*, 2002, **3**, 1038–1047.
- J. Yu, W. Wei, E. Danner, R. K. Ashley, J. N. Israelachvili and J. H. Waite, *Nat. Chem. Biol.*, 2011, **7**, 588–590.
- X. X. Qin, K. J. Coyne and J. H. Waite, *J. Biol. Chem.*, 1997, **272**, 32623–32627.
- J. Herzberger, K. Niederer, H. Pohlitz, J. Seiwert, M. Worm, F. R. Wurm and H. Frey, *Chem. Rev.*, 2016, **116**, 2170–2243.
- Y. Sarazin and J. F. Carpentier, *Chem. Rev.*, 2015, **115**, 3564–3614.



- 33 A. L. Brocas, C. Mantzaridis, D. Tunc and S. Carlotti, *Prog. Polym. Sci.*, 2013, **38**, 845–873.
- 34 P. Kraft and W. Eichenberger, *Eur. J. Org. Chem.*, 2003, **2003**, 3735–3743.
- 35 J. C. Bailar Jr, W. C. Fernelius and H. A. Skinner, *Inorg. Synth.*, 1939, **1**, 47–49.
- 36 L.-C. Shen, S.-Y. Chiang, I.-T. Ho, K.-Y. Wu and W.-S. Chung, *Eur. J. Org. Chem.*, 2012, **4**, 792–800.
- 37 Z. Baoxiang, W. Dawei, Z. Hua, Q. Guangtao and M. Junying, *Chin. J. Org. Chem.*, 2003, **23**, 1026–1028.
- 38 C. Billouard, S. Carlotti, P. Desbois and A. Deffieux, *Macromolecules*, 2004, **37**, 4038–4043.
- 39 K. Sakakibara, K. Nakano and K. Nozaki, *Macromolecules*, 2007, **40**, 6136–6142.
- 40 A. L. Brocas, G. Cendejas, S. Caillol, A. Deffieux and S. Carlotti, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**, 2677–2684.
- 41 B. F. Lee, M. J. Kade, J. A. Chute, N. Gupta, L. M. Campos, G. H. Fredrickson, E. J. Kramer, N. A. Lynd and C. J. Hawker, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**, 4498–4504.
- 42 K. C. Nicolaou, J. Wang, Y. Tang and L. Botta, *J. Am. Chem. Soc.*, 2010, **132**, 11350–11363.
- 43 K. C. Nicolaou, Y. Tang and J. Wang, *Angew. Chem., Int. Ed.*, 2009, **48**, 3449–3453.
- 44 S. Strych and D. Trauner, *Angew. Chem., Int. Ed.*, 2013, **52**, 9509–9512.
- 45 B. K. Ahn, D. W. Lee, J. N. Israelachvili and J. H. Waite, *Nat. Mater.*, 2014, **13**, 867–872.
- 46 X. Wu, X.-X. Chen, B.-N. Song, Y.-J. Huang, W.-J. Ouyang, Z. Li, T. D. James and Y.-B. Jiang, *Chem. Commun.*, 2014, **50**, 13987–13989.
- 47 K. J. Winstanley, A. M. Sayer and D. K. Smith, *Org. Biomol. Chem.*, 2006, **4**, 1760–1767.
- 48 L. M. Campos, K. L. Killops, R. Sakai, J. M. J. Paulusse, D. Damiron, E. Drockenmuller, B. W. Messmore and C. J. Hawker, *Macromolecules*, 2008, **41**, 7063–7070.
- 49 S. Boileau, in *Comprehensive Polymer Science, Chain Polymerization*, ed. G. C. Eastmond, A. Ledwith, S. Russo and P. Sigwalt, Pergamon Press, Oxford, 1989, vol. 3, Part 1, pp. 467–487.
- 50 S. D. Gagnon, in *Encyclopedia of Polymer Science and Engineering*, ed. R. P. Quirk, Wiley-Interscience, New York, 1985, vol. 6, pp. 273–307.
- 51 R. P. Quirk and G. M. Lizarraga, *Macromol. Chem. Phys.*, 2000, **201**, 1395–1404.
- 52 K. Sakakibara, K. Nakano and K. Nozaki, *Chem. Commun.*, 2006, **57**, 3334–3336.
- 53 S. S. Müller, C. Moers and H. Frey, *Macromolecules*, 2014, **47**, 5492–5500.
- 54 J. Herzberger and H. Frey, *Macromolecules*, 2015, **48**, 8144–8153.
- 55 G. Boireau, D. Abenhaim and E. Henry-Basch, *Tetrahedron*, 1980, **36**, 3061–3070.
- 56 J. J. Eisch, Z. R. Liu and M. Singh, *J. Org. Chem.*, 1992, **57**, 1618–1621.
- 57 T. Hagiwara, Y. Terasaki, H. Hamana, T. Narita, J. Umezawa and K. Furuhashi, *Makromol. Chem. Rapid Commun.*, 1992, **13**, 363–370.
- 58 K. Maruoka and T. Ooi, *Chem.–Eur. J.*, 1999, **5**, 829–833.
- 59 B. Antelmann, M. H. Chisholm, S. S. Iyer, J. C. Huffman, D. Navarro-Llobet, M. Pagel, W. J. Simonsick and W. Zhong, *Macromolecules*, 2001, **34**, 3159–3175.
- 60 M. H. Chisholm and D. Navarro-Llobet, *Macromolecules*, 2002, **35**, 2389–2392.
- 61 M. Lee, W. Zhang and B. C. Noll, *Angew. Chem., Int. Ed.*, 2009, **48**, 3449–3453.

