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Narjatamanins A and B, a pair of novel epimers possessing a 2,3-*seco*-iridoid skeleton with an unusual 1,10-oxygen bridge from *Nardostachys jatamansi* and evaluation of their effects on worm paralysis in AD *C. elegans*†

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Narjatamanins A (1) and B (2), a pair of epimers possessing a novel 2,3-*seco*-iridoid skeleton with an unusual 1,10-oxygen bridge, were isolated from the whole plants of *Nardostachys jatamansi*. Their structures were elucidated by a combination of various spectroscopic methods, including HRESIMS, IR and 1D and 2D NMR techniques. The absolute configurations of 1 and 2 were established by electronic circular dichroism (ECD) calculations. The pharmacological activities of 1 and 2 to alleviate AD-like symptoms were also evaluated using the *Caenorhabditis elegans* Alzheimer's disease (AD) pathological model, and narjatamanins A (1) and B (2) showed statistically significant delay in the worm paralysis phenotype of AD-like symptoms.

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1. Introduction

Nardostachys jatamansi DC, which belongs to the genus *Nardostachys* (Valerianaceae), is a perennial herb mainly distributed in China and India.¹ As a traditional medicine, its rhizomes and roots have been widely used as sedatives and analgesic in folk lore.^{1,2} It's worth mentioning that the rhizomes of *N. jatamansi* are also a key ingredient in ayurvedic formulations as an anticonvulsant.³ Several biological effects of the extracts from this species were reported, such as stress remedies, nervous headaches, epilepsy, intestinal colic, improving learning and memory and liver protection.⁴⁻⁷ Besides, recent studies have showed chemical constituents of *N. jatamansi* possess positive anti-Alzheimer's disease activity.^{8,9} To the best of our knowledge, previous phytochemical investigations on *N. jatamansi* led to the isolation of iridoids, sesquiterpenoids, and lignans.¹⁰⁻¹³ In an attempt to further explore structurally and biologically interesting natural products,¹⁴⁻¹⁷ we investigated the chemical constituents of the whole plants of *N. jatamansi*, which has resulted in the isolation and structural characterization of a pair of epimers possessing a novel 2,3-*seco*-iridoid skeleton with an unusual 1,10-oxygen bridge, named narjatamanins A (1) and B (2) (Fig. 1). In this paper, we describe the

isolation, structural elucidation, and the pharmacological activities of narjatamanins A (1) and B (2) towards alleviating Alzheimer's disease like symptoms in AD *Caenorhabditis elegans*.

2. Results and discussion

Compound 1 was obtained as a colorless oil. Its molecular formula of C₁₂H₁₈O₄ was determined by HRESIMS at *m/z* 249.1110 [M + Na]⁺ (calcd for C₁₂H₁₈O₄Na, 249.1097), which required four degrees of unsaturation. Its IR spectrum revealed absorption bands for hydroxyl (3445 cm⁻¹), double bond (1625 cm⁻¹), and aldehyde carbonyl (1690 cm⁻¹) groups. Its ¹H NMR spectroscopic data (Table 1) showed one oxygenated methine [δ_{H} 4.01 (1H, m)], one acetal methine [δ_{H} 5.03 (1H, d, *J* = 4.8 Hz)], an aldehyde proton [δ_{H} 9.58 (1H, s)], one ethoxyl group [δ_{H} 3.89 (1H, m), 3.62 (1H, m), and 1.27 (3H, t, *J* = 7.2

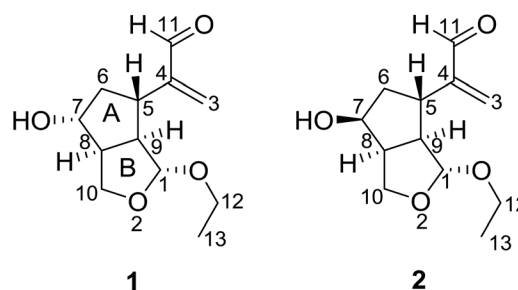


Fig. 1 Structures of narjatamanins A (1) and B (2).

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Table 1 ^1H (600 MHz) and ^{13}C (150 MHz) NMR data of compounds **1** and **2** in CDCl_3 (δ in ppm)

Position	1		2	
	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}
1	5.03 (d, 4.8)	101.5	5.06 (brs)	108.1
3	6.01 (brs)	132.8	6.07 (brs)	133.5
	6.28 (brs)		6.34 (brs)	
4		152.9		151.8
5	3.29 (m)	34.2	2.95 (m)	39.2
6 α	2.01 (m)	43.9	1.76 (m)	40.6
6 β	2.19 (m)		1.95 (m)	
7	4.01 (m)	72.1	4.29 (m)	72.9
8	2.96 (m)	48.7	2.90 (m)	47.5
9	2.56 (m)	54.9	2.52 (m)	55.6
10 α	3.92 (t, 9.0)	64.6	3.84 (t, 8.4)	65.2
10 β	4.04 (m)		4.20 (d, 9.0)	
11	9.58 (s)	194.7	9.57 (s)	194.3
12	3.89 (m)	62.7	3.43 (m)	62.3
	3.62 (m)		3.66 (m)	
13	1.27 (t, 7.2)	14.8	1.17 (t, 7.2)	14.9
7-OH	4.03 (brs)		2.58 (brs)	

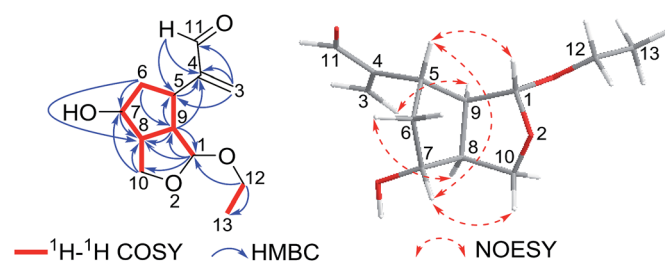


Fig. 2 Key ^1H - ^1H COSY, HMBC and NOESY correlations of **1**.

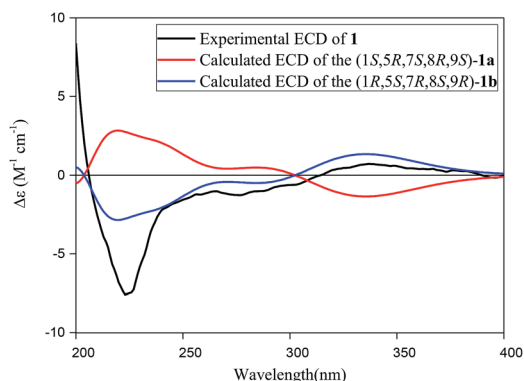


Fig. 3 Calculated and experimental ECD spectra of **1**.

Hz)]. Furthermore, a pair of isolated AB protons [δ_{H} 6.01 (1H, brs) and 6.28 (1H, brs)] indicated the presence of a terminal double bond. The ^{13}C NMR (Table 1) and DEPT spectra of **1** exhibited 12 carbon signals, which were sorted into one methyl, four methylenes (including two oxygenated carbons and one olefinic carbon), six methines (including one acetal carbon, one oxygenated carbon, and one aldehyde carbonyl carbon). Two

degrees of unsaturation represent one double bond and one aldehyde group, and the remaining two degrees of unsaturation pointed to two ring systems for compound **1**.

Interpretation of the 2D NMR data, containing ^1H - ^1H COSY, HSQC, and HMBC spectra, led to the construction of the planar structure of **1** (Fig. 2). The ^1H - ^1H COSY revealed the presence of two structural fragments [$\text{CH}(1)$ - $\text{CH}(9)$ - $\text{CH}(8)$ / $\text{CH}(5)$ - $\text{CH}_2(6)$ - $\text{CH}(7)$ - $\text{CH}(8)$ - $\text{CH}_2(10)$ and $\text{CH}_2(12)$ - $\text{CH}_3(13)$], which demonstrated the presence of a cyclopentane moiety (ring A). Given that C-1 and C-10 (one acetal methine carbon and one oxygenated methylene carbon, δ_{C} 101.5 and 64.6) were dramatically shifted downfield compared to typical hydroxylated carbons, an epoxy group was proposed between C-1 and C-10. Moreover, the HMBC correlations from acetal proton H-1 to C-8, C-9 and C-10, from H₂-10 to C-7 and C-8, from H-9 to C-1, C-5 and C-8 demonstrated the presence of a tetrahydrofuran ring (ring B) moiety fused to a cyclopentane (ring A) at C-8 and C-9. Other HMBC correlations from olefinic protons H₂-3 to C-4, C-5 and C-11, from aldehyde proton H-11 to C-4 and C-5 indicated the existence of an acrolein moiety that was located at C-5. Finally, the location of the ethoxyl group at C-1 was implied by the HMBC cross peaks of H₂-12 with C-1 and C-13. Considering the molecular formula, the downfield chemical shift of C-7 (δ_{C} 72.1) suggested a hydroxyl group suited at C-7, which could be proposed to share the remaining one oxygen atom. Thus, the planar structure of **1** was confirmed to be a novel 2,3-*seco*-iridoid with an oxygen bridge connecting C-1 and C-10.

The relative stereochemistry of compound **1** was resolved by NOESY spectrum as well as a computer-generated lower energy conformation using MM2 force field calculations (Fig. 2). The NOESY correlations of H-7/H-10 β , H-7/H-5, and H-5/H-1 indicated that H-1, H-5, H-10 β and H-7 were cofacial and randomly assigned as β -orientation. The NOESY correlations of H-9/H-3 α , H-8/H-6 α combined with the unobserved correlations of H-8/H-5 and H-8/H-1 revealed that H-9, H-6 α and H-8 were in the same orientation and assigned as α -orientation.

The absolute configuration of **1** was established by comparison of its experimental electronic circular dichroism (ECD) spectrum with calculated the computational spectra of the (1*S*,5*R*,7*S*,8*R*,9*S*)-**1a** and (1*R*,5*S*,7*R*,8*S*,9*R*)-**1b** using the time-dependent density functional theory (TD-DFT) method. As shown in Fig. 3, spectrum calculated for the (1*R*,5*S*,7*R*,8*S*,9*R*)-**1b** was nearly identical with the experimental data of **1** over the whole range of wavelengths under investigation, which confirmed the assignment of the absolute configuration of **1** as 1*R*, 5*S*, 7*R*, 8*S* and 9*R*. Thus, the absolute configuration of compound **1** could be defined unequivocally as shown in Fig. 1, and it was given the trivial name of narjatamanin A.

Compound **2** was obtained as a colorless oil. The HRESIMS indicated a molecule formula of $\text{C}_{12}\text{H}_{18}\text{O}_4$ (m/z 249.1098, calcd for $\text{C}_{12}\text{H}_{18}\text{O}_4\text{Na}$, 249.1097), the same as that of **1**. After the analysis of their ^1H and ^{13}C NMR data, compound **2** was found to share the same planar structure with compound **1**. Furtherly, the planar structure of compound **2** was confirmed by 2D NMR, including HSQC, ^1H - ^1H COSY, and HMBC spectra. In comparison of their ^1H -NMR, the most remarkable change was the downfield shift of 0.28 ppm for H-7 in **2** which suggested the



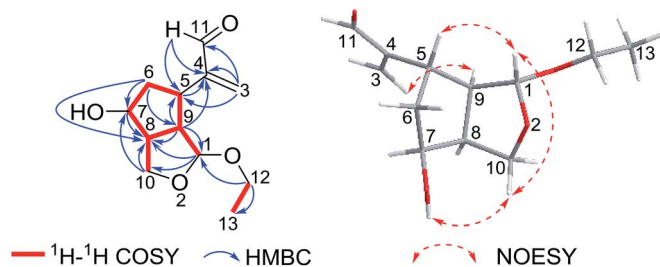


Fig. 4 Key ^1H – ^1H COSY, HMBC and NOESY correlations of **2**.

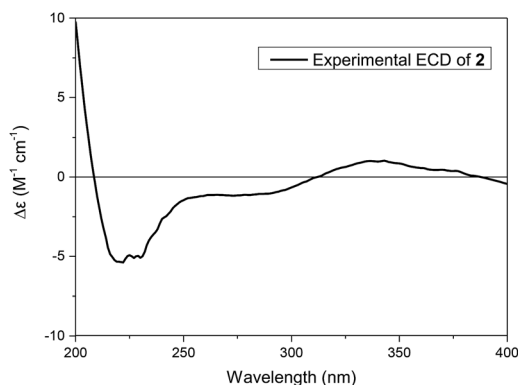


Fig. 5 Experimental ECD spectrum of **2**.

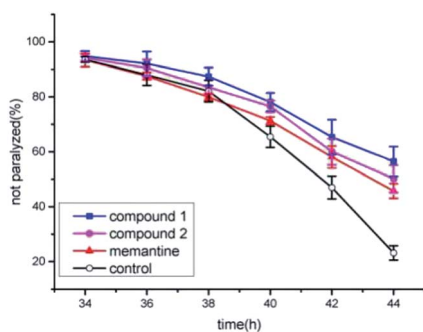


Fig. 6 Anti-Alzheimer's disease (AD) activities of compound **1** and **2** positive control and blank control.

configuration at C-7 could be different with compound **1**. The relative configuration was ensured by NOESY examination. The NOESY correlations of H-1/H-10 β , H-10 β /OH-7, and H-1/H-5 implied that H-1, H-5 and OH-7 were cofacial with β -oriented. The NOESY correlation of H-9/H-3a indicated that H-9 and H-5 were adverse direction, and H-9 was settled as α -orientation. Finally, H-8 was determined as α -oriented on the basis of the analysis that unobserved NOESY correlations of H-8/H-5 and H-8/H-1 (Fig. 4).

The absolute configuration of **2** was established by its experimental electronic circular dichroism (ECD) spectrum. The electronic circular dichroism (ECD) curve of **2** showed a negative Cotton effect at 219 nm and a positive Cotton effect at 343 nm, ascribed to the π – π^* transition and n – π^* transition of

α,β -unsaturated ketone, respectively. Further, the positive Cotton effect at 343 nm in its ECD spectrum (Fig. 5) was same as the positive Cotton effect at 337 nm in the ECD spectrum of **1**, which suggested the same configurations of the chiral center adjacent aldehyde carbonyl of **2** as **1**. Thus, the assignment of the absolute configuration of **2** as $1R, 5S, 7S, 8S,$ and $9R,$ and **2** was given the trivial name of narjatamanin B.

Previous works have showed that the effects of compounds tested were linked to their anti-amyloidogenic activity which have been indicated by a readout of worm paralysis being delayed,^{18,19} and the pharmacological actions of compounds tested has been correlated with *in vivo* amyloid aggregation and toxicity by using transgenic AD worms.^{20,21} Transgenic AD *Caenorhabditis elegans* CL4176 was used to evaluate whether new compounds of narjatamanins A (**1**) and B (**2**) have pharmacological activities for fighting against AD worm paralysis. The worms had been transferred with a human $\text{A}\beta_{1-42}$ gene downstream of promoter *myo-3*, and the overexpression of $\text{A}\beta_{1-42}$ in worm muscular tissues under nonpermissive condition will lead to the AD like symptom of paralysis phenotype. Previous work has demonstrated that memantine can significantly delay worms paralysis and acts as a disease modifying agent.²² Here, memantine was used as positive control. The result showed that 50 μM of memantine significantly delayed AD worm paralysis (Fig. 6). Further, 100 μM of narjatamanins A (**1**) and B (**2**) were also significantly alleviated AD like symptoms ($p < 0.05$). In contrast to 50 μM of memantine, the tested compounds **1** and **2** tended to have a higher pharmacological activity of delaying AD worm paralysis ($p < 0.05$). Additionally, as the hallmark of AD is amyloid β accumulation, and it is widely accepted to be the centre of AD pathology.²⁰ Taken together, our results provided clues that the compounds tested may be potential to be served as anti-AD agents. To study AChE enzyme, radicals scavenging activity and to perform amyloid β protein assay will give us the whole anti-AD activity profile of the compounds tested here, we will perform these experiment in our further research.

3. Conclusion

In summary, this work discovered a pair of novel epimers named narjatamanins A (**1**) and B (**2**) from *N. jatamansi*. These two compounds possessed a novel 2,3-*seco*-iridoid skeleton with an unprecedented oxygen bridge, which is a structural feature not previously reported in natural products. Their structures were determined by their spectroscopic data. They possessed multiple chiral centers, which could attract great interest from organic synthetic chemists. These two compounds showed statistically significant pharmacological activity of delaying AD worm paralysis, indicating that they might be potential to act as anti-AD agent candidate.

4. Experimental section

4.1 General experimental procedures

Optical rotations were determined on a PerkinElmer 341 polarimeter. IR spectra were recorded with a Nicolet NEXUS 670 FT-IR spectrometer in KBr. 1D ($^1\text{H}, ^{13}\text{C}, \text{DEPT}$) NMR and 2D



(^1H - ^1H COSY, HSQC, HMBC, NOESY) NMR were recorded with a Varian INOVA-600 and a Bruker AVANCE NEO-600 spectrometer with TMS as an internal standard. HRESIMS spectra were obtained on a Thermo LTQ Orbitrap Elite mass spectrometer. ECD spectra were recorded on a JASCO J-810 spectrophotometer using MeOH as the solvent. Silica gel (200–300 mesh) used for column chromatography (CC) and silica gel GF₂₅₄ (10–40 μm) used for thin layer chromatography (TLC) were purchased from the Qingdao Marine Chemical Factory in China. Sephadex LH-20 was supplied by GE Healthcare Bio-Sciences AB. MCI GEL CHP 20P (75–150 μm) was purchased from Mitsubishi Chemical Holdings in Japan.

4.2 Plant material

The whole plants of *N. jatamansi* were collected in Lijiang, Yunnan province, in August of 2017. The plant material was identified by Dr Jian-Yin Li, School of Pharmacy, Lanzhou University. A voucher specimen (no. 201708NJ) was deposited at School of Pharmacy, Lanzhou University, China.

4.3 Extraction and isolated

The air-dried and milled whole plants of *N. jatamansi* (10 kg) were extracted with 95% EtOH three times at room temperature to give a crude extract (1.8 kg). The extract was suspended in H₂O and partitioned with EtOAc. The EtOAc extract (890 g) was applied to column chromatography over silica gel eluted with petroleum ether–acetone (from 40 : 1 to 1 : 1) gradient elution to obtain six fractions (Fr.1–Fr.6). Fr.3 (71 g) was chromatographed using MCI gel CC eluted with EtOH–H₂O gradient (30%, 50%, 70%, 90%, 100%, v/v) to get five subfractions (Fr.3.1–Fr.3.5). Fr.3.2 was purified by thin layer chromatography (TLC) with CH₂Cl₂–acetone (20 : 1) to afford compound **1** (8 mg). Fraction 5 was separated to a Sephadex LH-20 CC (CHCl₃–CH₃OH, 1 : 1) to give six subfractions (Fr.5.1–Fr.5.6). Fr.5.2 was separated by silica gel (CH₂Cl₂–acetone, 40 : 1 to 2 : 1) to afford five subfractions (Fr.5.2.1–Fr.5.2.5). Fr.5.2.3 was purified by TLC to yield compound **2** (11 mg).

Narjatamanin A (1). Colorless oil; $[\alpha]_{\text{D}}^{25}$ –55 (*c* 0.60, CH₂Cl₂); IR (KBr) ν_{max} 3445, 3056, 2947, 1690, 1625, 1272, 1097, 735 cm^{-1} ; ^1H and ^{13}C NMR data, see Table 1; HRESIMS *m/z* 249.1100 [M + Na]⁺ (calcd for C₁₂H₁₈O₄Na, 249.1097).

Narjatamanin B (2). Colorless oil; $[\alpha]_{\text{D}}^{25}$ –47 (*c* 1.10, CHCl₃); IR (KBr) ν_{max} 3449, 3088, 2936, 1690, 1625, 1272, 1099, 735 cm^{-1} ; ^1H and ^{13}C NMR data, see Table 1; HRESIMS *m/z* 249.1098 [M + Na]⁺ (calcd for C₁₂H₁₈O₄Na, 249.1097).

4.4 Bioactivity assay for delaying the AD like symptom of paralysis phenotype in transgenic Alzheimer's disease *C. elegans* procedure

The transgenic *C. elegans* strain CL4176 (smg-1ts [myo-3/ $\text{A}\beta_{1-42}$ long 3'-untranslated region (UTR)]) was obtained from Caenorhabditis Genetics Center (CGC) (University of Minnesota, Minneapolis, MN). Worms were cultured at 16 °C on nematode growth medium (NGM) plates, which were seeded with *E. coli* (OP50) as standard food resource. About 100 synchronized worms at L1 larval stage were transferred onto NGM containing

50 μM tested compound in 0.1% DMSO, and 0.1% DMSO was used as negative control. The animals were kept at 16 °C till they grew into L3 larval stage, then transferred into a 25 °C incubator for 34 h. Paralyzed worms were counted under a dissecting microscope at 2 h intervals until all animals did not moved autonomously and got into paralysis. The pharmacological activity was described as the capacity of tested compounds to delay worm paralysis. For pharmacological activity of delaying AD worm paralysis assay, log rank survival test was used to compare the significance among treatments.¹⁹ The *p* value at a level of 0.05 or less was considered as statistical significance.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

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References

- 1 L. Q. Qiu, *Flora Reipublicae Popularis Sinicae (Zhongguo Zhiwu Zhi)*, Science Press, Beijing, 1986, vol. 73, pp. 25–26.
- 2 A. Chatterjee and S. C. Pakrashi, *The Treatise on Indian Medicinal Plants*, National Institute of Science Communication, New Delhi, 1997, vol. 5, pp. 99–100.
- 3 A. Chatterjee, B. Basak, M. Saha, U. Dutta, C. Mukhopadhyay, J. Banerji, Y. Konda and Y. Harigaya, *J. Nat. Prod.*, 2000, **63**, 1531–1533.
- 4 H. Joshi and M. Parle, *J. Med. Food*, 2006, **9**, 113–118.
- 5 A. Ali, Y. Dua, A. W. Siddiqui, S. Sultana and M. R. M. Rafiullah, *Pharm. Biol.*, 2005, **43**, 533–539.
- 6 V. Prabhu, K. S. Karanth and A. Rao, *Planta Med.*, 1994, **60**, 114–117.
- 7 S. Ali, K. A. Ansari, M. A. Jafry, H. Kabeer and G. Diwakar, *J. Ethnopharmacol.*, 2000, **71**, 359–363.
- 8 Q. F. Liu, Y. Jeon, Y. W. Sung, J. H. Lee, H. Jeong, Y. M. Kim, H. S. Yun, Y. W. Chin, S. Jeon, K. S. Cho and B. S. Koo, *Biol. Pharm. Bull.*, 2018, **41**, 470–477.
- 9 P. Q. Wu, Y. F. Yu, Y. Zhao, C. X. Yu, D. J. Zhi, F. M. Qi, D. Q. Fei and Z. X. Zhang, *Org. Biomol. Chem.*, 2018, **16**, 9038–9045.
- 10 A. Bagchi, Y. Oshima and H. Hikino, *Tetrahedron*, 1990, **46**, 1523–1530.
- 11 K. Rekha, R. R. Rao, R. Pandey, K. R. Prasad, K. S. Babu, J. R. Vangala, S. V. Kalivendi and J. M. Rao, *J. Asian Nat. Prod. Res.*, 2013, **15**, 111–116.



- 12 R. S. Chauhan, M. K. Kaul, A. Kumar and M. C. Nautiyal, *Sci. Hortic.*, 2008, **117**, 78–81.
- 13 G. Rücker, J. Tautges, M. L. Maheswari and D. B. Saxena, *Phytochemistry*, 1976, **15**, 224.
- 14 Z. X. Zhang, H. H. Li, D. J. Zhi, P. Q. Wu, Q. L. Hu, Y. F. Yu, Y. Zhao, C. X. Yu and D. Q. Fei, *Tetrahedron Lett.*, 2018, **59**, 4028–4030.
- 15 Z. X. Zhang, P. Q. Wu, H. H. Li, F. M. Qi, D. Q. Fei, Q. L. Hu, Y. H. Liu and X. L. Huang, *Org. Biomol. Chem.*, 2018, **16**, 1745–1750.
- 16 Z. Y. Li, F. M. Qi, D. J. Zhi, Q. L. Hu, Y. H. Liu, Z. X. Zhang and D. Q. Fei, *Org. Chem. Front.*, 2017, **4**, 42–46.
- 17 D. Q. Fei, L. L. Dong, F. M. Qi, G. X. Fan, H. H. Li, Z. Y. Li and Z. X. Zhang, *Org. Lett.*, 2016, **18**, 2844–2847.
- 18 C. D. Link, *Proc. Natl. Acad. Sci. U. S. A.*, 1995, **92**, 9368–9372.
- 19 C. D. Link, *Genes, Brain Behav.*, 2005, **4**, 147–156.
- 20 V. Dostal, C. M. Roberts and C. D. Link, *Genetics*, 2010, **186**, 857–866.
- 21 L. Diomedea, G. Cassata, F. Fiordaliso, M. Salio, D. Ami, A. Natalello, S. M. Doglia, A. De Luigi and M. Salmona, *Neurobiol. Dis.*, 2010, **40**, 424–431.
- 22 P. Martorell, E. Bataller, S. Llopis, N. Gonzalez, B. Álvarez, F. Montón, P. Ortiz, D. Ramón and S. Genovés, *PLoS One*, 2013, **8**, e63283.

