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## The first demonstration of a novel isolated fungus *Eutypella* sp. BJ associated with the biodegradation of polyvinyl alcohol

Yiling Deng, Chenyuan Wang, Yalan Liu, Ping Chen, Xiaoshan Lin and Yi Zhang \*

The aim of this work is to study the potential degradation of polyvinyl alcohol (PVA) by a novel fungus *Eutypella* sp. BJ isolated from soil compost. When it was cultured on a semi-synthetic medium containing PVA at 30 °C and 160 rpm for 8 days, the removal rates of PVA 1788, 1799 and 2488 reached 87.40%, 86.31% and 44.80%, respectively. Gel permeation chromatography (GPC) analysis revealed significant reductions of the number average molecular weight and the weight average molecular weight of PVA 1788 from 47 358 to 13 345 and from 71 387 to 24 238, respectively, after 8 days. Fourier transform infrared spectroscopy (FTIR) indicated that some substances containing carbonyl groups (likely aldehydes or ketones) might have been produced during the biodegradation process. These results indicate that the isolate has potential for degrading PVA. This study provides the first demonstration that *Eutypella* has the ability to assimilate PVA.

## 1. Introduction

Polyvinyl alcohol (PVA), a synthetic water-soluble polymer, is widely used in textile, food, leather, medical and paper industries for its excellent properties, such as good adhesion, film toughness and wear resistance.<sup>1</sup> However, PVA is not readily degraded in the natural environment, resulting in serious environmental problems. For example, once dispersed in aqueous systems, PVA can directly interfere with the life cycles of aquatic organisms and convert them into a major organic pollution source that is subsequently preyed upon by fish, which may then die due to toxicity. In addition, when mixed into the soil, it can affect the absorption of nutrients and water by crops, leading to reduced crop yields. Therefore, in recent years, researchers have been investigating environmentally friendly methods to degrade PVA.

Research on the biodegradation of PVA can be traced back to 1936, when Nord found that PVA can be degraded under extracellular attack by a dehydratase of *Fusarium lini*.<sup>2</sup> Since Suzuki and co-workers successfully isolated the first strain capable of utilizing PVA as the sole carbon source in 1973, researchers have conducted extensive research on the biodegradation of PVA.<sup>3,4</sup> *Pseudomonas* sp. 113P3 isolated by Hatanaka was able to grow in a medium with PVA as the sole carbon source and produce a PQQ-dependent PVA dehydrogenase.<sup>5</sup> Sakai *et al.* found that *Pseudomonas* secreted PVA oxidase, which could catalyse the oxidation of PVA and make it hydrolyse into

a small molecular compound.<sup>6</sup> However, most research to date has focused on the degradation of PVA by bacteria, and limited information is available regarding PVA degradation by fungi. Actually, the use of fungi and other versatile microbes to remove or mitigate the effects of macromolecular contaminants in the environment is especially important for environmental protection.<sup>7</sup>

In recent years, the degradation of PVA by fungi has been investigated. Yeast and basidiomycetes able to degrade lignin were founded to assimilate PVA.<sup>8,9</sup> Furthermore, *Flammulina velutipes* was shown to exhibit PVA degradation ability and to preferentially assimilate the syndiotactic structure of PVA, suggesting that the mycelia produced PVA-degrading enzymes.<sup>10</sup> A brown-rot fungus, *Fomitopsis pinicola*, could degrade PVA via the Fenton reaction in unsubmerged culture.<sup>11</sup> Mejía *et al.* found that enzymatic extracts of the *Phanerochaete chrysosporium* fungus could promote the degradation of PVA and reduce the average molecular weight by approximately 80% by forming carbonyl groups and double bonds.<sup>12</sup> Following infrared and ultraviolet analysis, they proposed a degradation route: modification of the free radical into an epoxide, followed by hydrolysis to form a double bond. A fungal strain belonging to *Penicillium* sp. isolated from activated sludge was reported able to completely degrade PVA after 12 days and secrete PVA oxidase to decompose H<sub>2</sub>O<sub>2</sub>.<sup>13</sup> In addition, the fungus *Aspergillus niger* was demonstrated to grow vigorously on the surface of PVA films and assimilate them by SEM techniques.<sup>14</sup> Two white-rot fungi, *Pycnoporus cinnabarinus* and *Phanerochaete chrysosporium*, have been reported able to degrade PVA.<sup>15</sup> Following their discovery of a bacterial strain (BX1) and a fungal strain (WF9101) in activated sludge, Mori *et al.* studied the biodegradation mechanism of PVA oligomers and found that

School of Biology and Biological Engineering, South China University of Technology, No. 382 Waihuan East Rd., Guangzhou University City, Panyu District, Guangzhou 510006, Guangdong, China. E-mail: [btzhang@scut.edu.cn](mailto:btzhang@scut.edu.cn); Fax: +86-20-39380602; Tel: +86-20-39380601



the strains could synergistically degrade PVA to vinyl alcohol oligomers.<sup>16–18</sup> They proposed that the hydroxyl groups of PVA oligomers were oxidized by secondary alcohol oxidase to carbonyl groups, and subsequently,  $\beta$ -diketone hydrolase catalysed the cleavage of C–C bonds of oxidized vinyl alcohol oligomers.<sup>19</sup> Lignolytic fungi might contribute to non-metabolic assimilation of PVA by extracellular oxidizing systems in soil environments.<sup>20</sup> The biodegradation of composites based on PVA have also been studied. *Trichothecium roseum* fungus was found able to degrade PVA and PVA-graft-lactic acid copolymers, and copolymers with greater numbers of lactic acid grafted units were more conducive to attack by this fungus than those with fewer units.<sup>21</sup>

For this study we isolated and identified a novel fungus capable of degrading PVA from soil compost. To assess the potential of the isolate in assimilating PVA, we (1) compared the degradation characteristics of PVA 1788, 1799 and 2488, which exhibit different degrees of polymerization and saponification; (2) characterized the chemical structure changes of PVA 1788 during degradation by FTIR analysis; and (3) investigated the molecular weight and the molecular weight distribution of PVA 1788 at different times. Compared with fungi able to degrade PVA (Table 1), the isolated fungus *Eutypella* sp. BJ showed the ability to assimilate PVA with high efficiency in a shorter period of time. This study provides the first demonstration that *Eutypella* can successfully degrade PVA.

## 2. Materials and methods

### 2.1 Materials

PDA, MEA, CYA medium and mold liquid medium were purchased from Guangdong HuanKai Microbial Sci. & Tech. Co., Ltd. The commercial PVA powders (degree of polymerization, 1700 or 2400; degree of saponification, 88 or 99%) were purchased from Chang-Chun Petrochemical, Taiwan, China. Poly vinyl formal (PVF) compounded from PVA and formaldehyde was generously supplied by Professor Youyi Zhang from Tunghai University in Taiwan, China. Compost was obtained from Junde Biotech Co., Ltd. (Zhucheng, China). The main components and properties of the compost were as follows: chicken dung, 52.02% (w/w); straw, 28.45% (w/w); water-holding capacity, 68%; pH, 6.73.

### 2.2 Isolation of PVA degrading strain

Microbial samples were extracted from the surfaces of degraded PVF samples that had been buried in compost for three years in

our laboratory.<sup>22</sup> PVF particles were placed in Erlenmeyer flasks containing 50 mL sterile deionized water and shaken gently. Three millilitres of the suspension was incubated in mold liquid medium (100 mL). Then, 50  $\mu$ L of culture was sprayed onto the screening agar medium using PVA as the sole carbon source and incubated at 30 °C for 7 days. Strains that successfully grew on the screening agar medium were isolated, purified, and then incubated again on the screening agar medium on sterile filter paper. Next, a 3 mL mixture of iodine and boric acid (1 : 4 by volume) was added to the plates, and the reaction was allowed to occur for 15 min in the dark.<sup>22</sup> The assay was performed in three replicates. The colonies surrounded by a large transparent circle were selected. The composition of the screening agar medium was as follows: PVA, 1.00 g L<sup>-1</sup>; (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 1.00 g L<sup>-1</sup>; K<sub>2</sub>HPO<sub>4</sub>, 2.50 g L<sup>-1</sup>; KH<sub>2</sub>PO<sub>4</sub>, 0.20 g L<sup>-1</sup>; CaCl<sub>2</sub>, 0.05 g L<sup>-1</sup>; MgSO<sub>4</sub> · 7H<sub>2</sub>O, 0.10 g L<sup>-1</sup>; FeSO<sub>4</sub> · 7H<sub>2</sub>O, 0.05 g L<sup>-1</sup>; NaCl, 0.50 g L<sup>-1</sup>; chloramphenicol, 0.30 g L<sup>-1</sup>; agar, 15.00 g L<sup>-1</sup>.

### 2.3 DNA extraction, PCR amplification and phylogenetic analysis

Strains that grew on the screening medium and formed large transparent circles were cultured in mold liquid medium in an Erlenmeyer shaker at 30 °C and 160 rpm for 2 days. DNA of the isolated fungus was extracted from cultures by using the Ezup Column Fungi Genomic DNA Purification Kit. Two universal fungal primers ITS4 (5'-TCCTCCGCTTATTGATATGC-3') and ITS5 (5'-GGAAGTAAAGTCGTAACAAGG-3') were used to amplify the DNA by PCR. The amplified PCR product was purified by electrophoresis in a 1.0% agarose gel. Purified DNA amplicons were sent to Guangzhou Qingke Biotechnology Co., Ltd., for sequencing. The DNA sequence from the fungal isolate was aligned with the sequences of closely related species from the NCBI nucleotide database by the CLUSTAL W program.<sup>23</sup> The evolutionary tree inferred from the data was constructed using the neighbour-joining method in MEGA version 7.0.26. The stability of each branch was estimated by performing bootstrap analyses of the neighbour-joining data based on 1000 trials.

### 2.4 Morphology of the isolated PVA degrading fungus

The isolated fungus was separately spotted on PDA, MEA, and CYA medium and cultured at 30 °C for 7 days to observe colony morphology. The strain was inoculated into a mold liquid medium, cultured at 30 °C for 3 days, and then stained with

Table 1 The comparation of fungi reported able to degrade PVA

Fungus	Efficiency of PVA degradation	Degradation time (days)	Researchers
<i>Flammulina velutipes</i>	53.5%	14	Tsujiyama <i>et al.</i> <sup>10</sup>
<i>Phanerochaete chrysosporium</i>	79.3%	15	Mejia <i>et al.</i> <sup>12</sup>
<i>Penicillium</i> sp. WSH02-21	100%	12	Ding <i>et al.</i> <sup>13</sup>
<i>Aspergillus niger</i>	The surface being eroded	7	Jecu <i>et al.</i> <sup>14</sup>
<i>Pycnoporus cinnabarinus</i>	Nearly 50%	30	Larking <i>et al.</i> <sup>15</sup>
<i>Geotrichum</i> sp. WF9101	80%	8	Sakimoto <i>et al.</i> <sup>16–18</sup>



a lactic acid carbonic acid blue staining solution to observe the morphology of the isolated fungus.

## 2.5 PVA degradation test

The PVA degradation test was performed in liquid cultures. The ability of the isolated strain to degrade PVA was investigated using PVA with two different degrees of polymerization and saponification. Prior to inoculum preparation, a seed culture was prepared by culturing the isolated fungus in 100 mL of mold liquid medium on a rotary shaker at 30 °C and 160 rpm for 3 days. Next, one percent of the fungal inoculum was added to the semi-synthetic medium (100 mL) and incubated in an Erlenmeyer shaker at 30 °C and 160 rpm for 8 days. Cultures were centrifuged at 10 000 rpm for 15 min at 4 °C to remove the cells for degradation analyses. The composition of the semi-synthetic medium was as follows: PVA, 1.00 g L<sup>-1</sup>; (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 1.00 g L<sup>-1</sup>; K<sub>2</sub>HPO<sub>4</sub>, 2.50 g L<sup>-1</sup>; KH<sub>2</sub>PO<sub>4</sub>, 0.20 g L<sup>-1</sup>; CaCl<sub>2</sub>, 0.05 g L<sup>-1</sup>; MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.10 g L<sup>-1</sup>; FeSO<sub>4</sub>·7H<sub>2</sub>O, 0.05 g L<sup>-1</sup>; NaCl, 0.50 g L<sup>-1</sup>; chloramphenicol, 0.30 g L<sup>-1</sup>. The initial pH was neutral.

## 2.6 Analysis methods

The biodegradation of PVA by the isolated strain was comprehensively investigated by analysing the changes in biomass, PVA concentration, functional groups and molecular weight among time points before, as well as during and after the degradation of PVA.

**2.6.1 Measurements of dry cell weight (DCW).** DCW was determined to measure cell growth. The cells collected by centrifugation were washed three times with deionized water and dried to a constant weight at 105 °C.

**2.6.2 Measurement of concentration of PVA.** The analysis of PVA concentration was performed using a modified Finley method.<sup>22</sup> Aqueous boric acid and iodine were added to the samples for 15 min at room temperature at a proportion of KI-I<sub>2</sub> (2.5 + 12.7 g L<sup>-1</sup>): boric acid (4 g L<sup>-1</sup>): sample, which was equivalent to 2 : 3 : 2 (v/v/v) for PVA 1799, 1 : 3 : 1 (v/v/v) for PVA 1788 and 1 : 2 : 1 (v/v/v) for PVA 2488. The OD of each mixture was measured at 500 nm for PVA 1799, at 495 nm for PVA 1788, and at 520 nm for PVA 2488. Three measurements were taken for each sample.

**2.6.3 Gel permeation chromatography analysis.** Molecular weight distribution was analysed by GPC. The centrifugal supernatant was sent to Guangzhou Zhebo Testing Technology Co., Ltd., for testing by gel permeation chromatography Waters 5510, equipped with a refractive index detector/triple detector. The conditions for GPC were as follows: 10 μm mixed-b column, 1,2,4-trichlorobenzene as the elution solvent, and an effluent flow rate of 1.0 mL min<sup>-1</sup>. Calibration was performed using a polystyrene standard.

**2.6.4 Fourier transform infrared spectroscopy analysis.** The culture supernatant was filtered through a membrane filter (pore size, 0.22 μm). The PVA-containing supernatant was dried by a freeze dryer (Guangzhou Kepeng Scientific Instrument Co., Ltd.), and its infrared spectra at different biodegradation times were measured by using a Bruker FTIR Vector 33

spectrophotometer with the spectral regions from 400 to 4000 cm<sup>-1</sup> at room temperature.

## 3. Results and discussion

### 3.1 Identification of the isolated PVA degrading strain

A novel fungus forming a large transparent circle on the screening agar medium with the ability to degrade PVA was isolated (Fig. 1). Identification of the fungus was performed through ITS sequence analysis and the observation of cell morphology.

The isolated fungus was identified as *Eutypella* sp. by ITS sequence analysis, which had 100% similarity. Its position in a phylogenetic tree is shown in Fig. 2. The ITS sequence of *Eutypella* sp. BJ was submitted to GenBank under accession number MK651593.

After one week of growth, *Eutypella* sp. BJ was densely spread on the surfaces of the CYA, MEA and PDA media, with its colonies exhibiting a white, cotton-like morphology. The original inoculation point on the central of CYA medium had turned black and hard (Fig. 3(a)), and branching hyphae were visible on the edges of MEA medium (Fig. 3(b)). The mycelium of *Eutypella* sp. BJ incubated on the PDA medium tended to spread to the cover of the culture dish, and the mycelium was densely distributed at the centre of the medium and sparsely distributed at the edges (Fig. 3(c)). No spores were detected when *Eutypella* sp. BJ was cultured in mold liquid medium. After staining *Eutypella* sp. BJ, mycelia were observed with an optical microscope (Fig. 4).

Recent research on *Eutypella* has mainly focused on its metabolites, which have excellent antibacterial and antitumour activities.<sup>24–26</sup> However, little research on the degradation of materials by *Eutypella* has been conducted. One report concerning its degradation ability was that Lee Jung-Bok *et al.* isolated *Eutypella* sp. KEF-1 from endosulfan-polluted soils, and it had the potential to degrade organochlorine insecticide endosulfan.<sup>27</sup> To the best of our knowledge, *Eutypella* sp. BJ is the first fungus discovered to have the ability to independently degrade PVA. In addition to having metabolites with antitumour, immunosuppressive and other biological activities, *Eutypella* sp. may have great potential for polymer degradation.

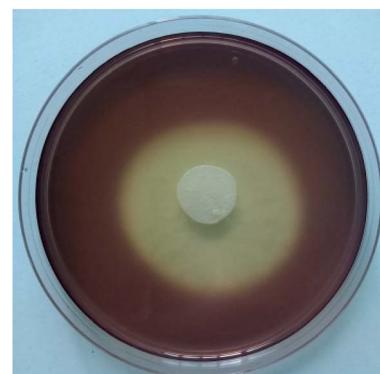


Fig. 1 Agar plate assay for PVA degradation by *Eutypella* sp. BJ.



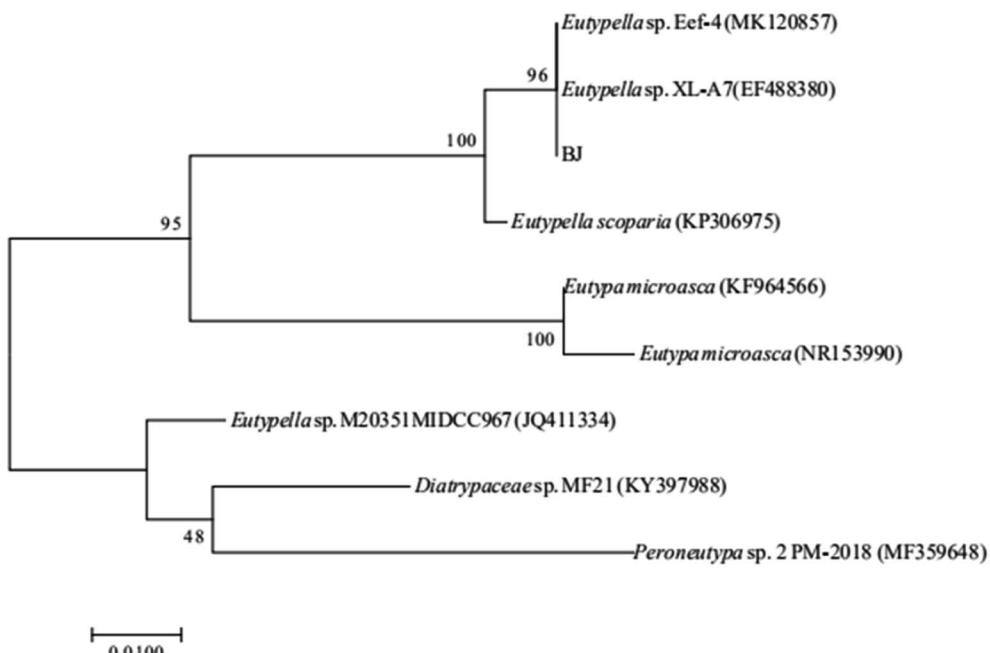


Fig. 2 Phylogenetic analysis of the *Eutypella* sp. BJ based on ITS sequence.

### 3.2 Degradation of different types of PVA

*Eutypella* sp. BJ was cultivated in semi-synthetic medium at 30 °C to study its biodegradation of PVA with different degrees of polymerization and saponification. As shown in Fig. 5, degradation speed of *Eutypella* sp. BJ to three types of PVA on the first day was faster than that on other 7 days, and the slow growth of *Eutypella* sp. BJ accompanied with the tardy increase of PVA removal on the 2nd to 8th day. With the increase of DCW, the removal of the three types of PVA increased. These results indicated that the growth of *Eutypella* sp. BJ was closely related to the biodegradation of PVA. *Eutypella* sp. BJ degraded 87.40% and 86.31% of PVA 1788 and PVA 1799, respectively, after 8 days. When *Eutypella* sp. BJ was cultivated on PVA 1788 and PVA 1799, more cell growth was observed compared to cells grown on PVA 2488, which was consistent with their high removal rates. The inconspicuous degradation difference indicated that the degree of saponification had little effect on the

degradation of PVA by *Eutypella* sp. BJ. Similarly, Tomoo Suzuki *et al.* found no marked difference in the degradation of PVA by crude enzyme of *Pseudomonas* O-3 among PVAs with different degrees of saponification.<sup>28</sup> The removal rate of PVA 2488 (44.80%), which has a higher degree of polymerization than PVA 1788, was lower than PVA 1788, mainly because when the molecular chains of PVA are stretched in semi-synthetic medium, entanglement is more likely to occur between molecular chains, and higher polymerization causes greater molecular chain entanglement. In addition, PVA samples with high molecular weight had stronger intermolecular hydrogen bonding, which was beneficial to crystallization, making it more difficult to the stretch of PVA molecular chains in liquid cultures. Therefore, it was not conducive to the invasion of *Eutypella* sp. BJ, causing poor efficiency to degrade PVA 2488 with high molecular weight.

Previous investigations have also reported effects of the degrees of polymerization and saponification on PVA

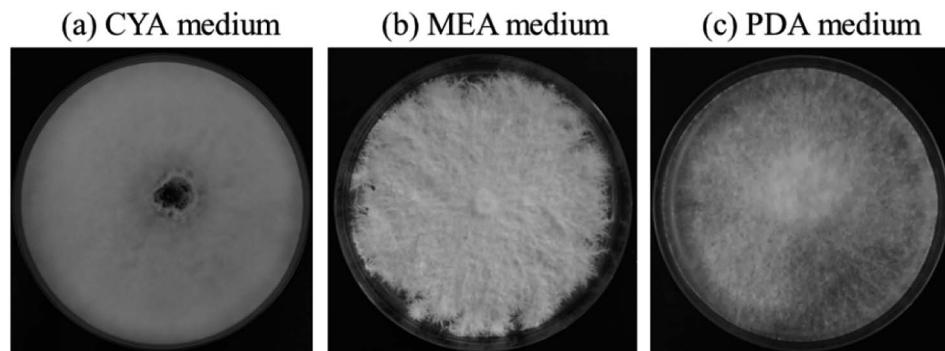
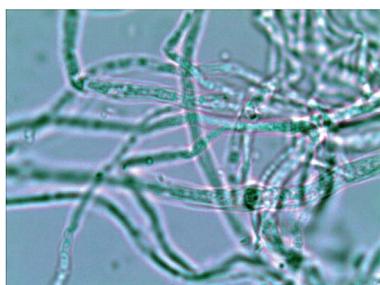


Fig. 3 Photographs of 7 days fungal cultures incubated in Petri plates. (a) Growth of *Eutypella* sp. BJ on CYA medium; (b) growth of *Eutypella* sp. BJ on MEA medium; (c) growth of *Eutypella* sp. BJ on PDA medium.



Fig. 4 Micrographs of *Eutypella* sp. BJ ( $\times 1000$ ).

biodegradation. Roberto Solaro *et al.* demonstrated a moderate dependence of saponification on the degradation of PVA by a selected PVA-degrading mixed bacterial culture isolated from the sewage sludge of a paper mill treatment plant.<sup>29</sup> Emo Chiellini found no significant differences in degradation among three PVA samples (PVA72, PVA88, PVA98) with different degrees of saponification during the biodegradation process.<sup>30</sup> In addition, when studying the effect of PVA structure on the dehydrogenation reaction by PVA dehydrogenase (PVADH) from *Pseudomonas* sp. 113P3, Tadashi Hatanaka found that as the degree of polymerization decreased, the Michaelis constant of PVADH-S decreased.<sup>31</sup> Combining with the above analysis and results of this study, the degradation of PVA depends little on the degree of saponification and is more influenced by the degree of polymerization.

**Table 2** The molecular weight and the molecular weight distribution of PVA 1788 in the semi-synthetic medium inoculated with *Eutypella* sp. BJ by GPC analysis

Time (days)	$M_n$	$M_w$	$d$
0	47 358	71 387	1.51
3	43 420	62 254	1.43
6	16 561	28 599	1.73
8	13 345	24 238	1.82

### 3.3 GPC analysis of PVA

As PVA 1788 exhibited a relatively high removal rate, it was chosen for GPC and FTIR analysis. The GPC analysis revealed changes in the molecular weight and the molecular weight distribution of PVA 1788 over different biodegradation times. As shown in Table 2, the number average molecular weight ( $M_n$ ) and the weight average molecular weight ( $M_w$ ) of the degraded PVA 1788 decreased from 47 358 and 71 387 to 13 345 and 24 238, respectively, representing significant decreases of 71.82% and 66.05% over the eight-day incubation. Increases in the polydispersity index ( $d$ ,  $d = M_w/M_n$ ) of PVA 1788 was also observed, which changed from 1.51 (unbiodegraded PVA 1788 at 0 day) to 1.81 (biodegraded PVA 1788 at 8 days). The increase of  $d$  indicated that the molecular weight distribution of PVA 1788 had broadened. In addition, with increasing degradation time, the GPC profiles of PVA 1788 shifted towards the lower molecular weight area (Fig. 6). These behaviours can be

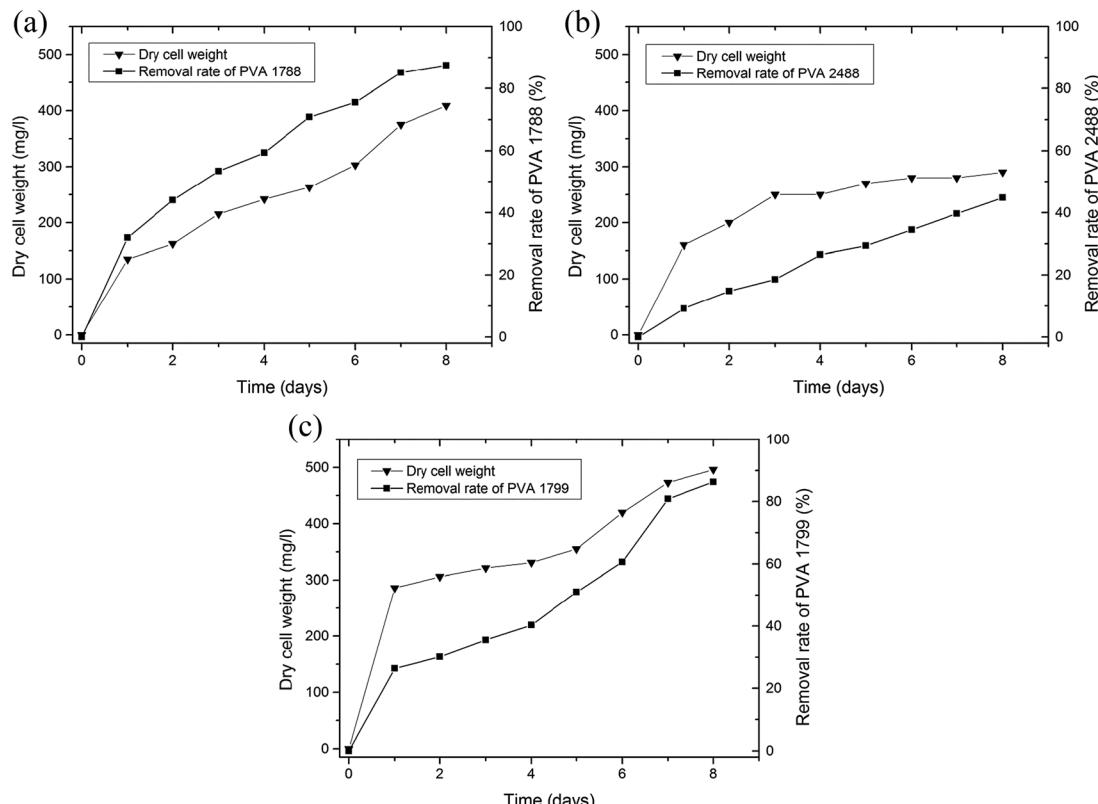


Fig. 5 Biodegradation curves of PVA samples with different degrees of polymerization and saponification recorded in biodegradation tests. (a) biodegradation curve of PVA 1788; (b) biodegradation curve of PVA 2488; (c) biodegradation curve of PVA 1799.



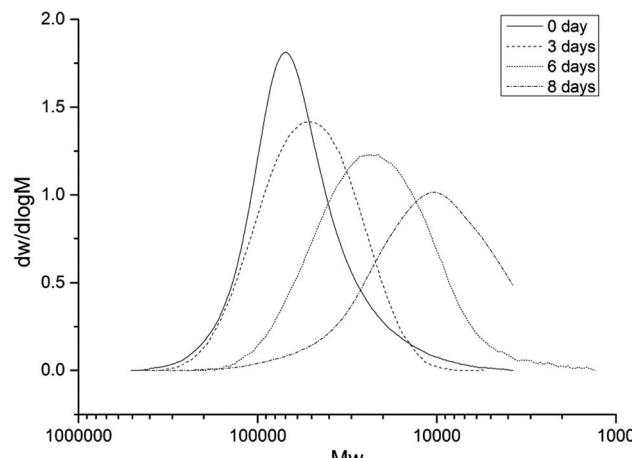


Fig. 6 Molecular weight distribution of PVA 1788 in the semi-synthetic medium as recorded at different incubation times.

attributed to the preferential assimilation of low molecular weight fractions of PVA 1788, which is in accordance with the literature.<sup>32</sup> Relative to the low molecular weight fraction, the high molecular weight fraction of PVA 1788 has severe chain entanglement and strong intermolecular force, making the segments and molecular chains difficult to move; these conditions are not conducive to the invasion of *Eutypella* sp. BJ. Therefore, there was a tendency for *Eutypella* sp. BJ to assimilate the low molecular weight part of PVA 1788.

#### 3.4 FTIR characterization

FTIR spectra analysis was performed to study the structural changes of PVA 1788 at different culture times. Compared with unbiodegraded PVA 1788 (0 day), the degraded PVA 1788 (at 3 days, 6 days, and 8 days) exhibited some differences in the infrared spectrum, as shown in Fig. 7. An absorption peak at  $1715\text{ cm}^{-1}$  was observed, which represents the carbonyl group, indicating an increase in macromolecule unsaturation. The

stretching vibration band of the hydroxyl group ( $3700\text{--}3000\text{ cm}^{-1}$ ) became widened and less peaked, and superimposed with the  $\nu\text{CH}$  stretching vibration band, which appeared at the tail of the  $\nu\text{OH}$  stretching vibration peak. These results indicated that some substances containing carbonyl groups (likely aldehydes or ketones) might have been produced during the biodegradation of PVA 1788, which was in accordance with the literature data.<sup>32,33</sup> Proposed mechanisms of PVA biodegradation reported until now are mainly based on two steps: hydroxyl groups of PVA are firstly oxidized to  $\beta$ -diketone structures, and then C-C bonds at diketone structures are cleaved by hydrolysis. Formation of compounds containing carbonyl groups may indicate that related PVA-degrading enzymes of the fungus may act on the hydroxyl groups in PVA by oxidation or dehydrogenation, leading to the formation of carbonyl groups during the degradation. It has been reported that after microorganisms invade material, they can release enzymes that accelerate the degradation of polymers by destroying the stability of the internal structure.<sup>34,35</sup> It could be explained that the hyphae of fungus *Eutypella* sp. BJ may penetrate into the interior of PVA 1788 and cause a chemical breakdown.

## 4. Conclusions

The present study explored the potential of a novel fungus (*Eutypella* sp. BJ) in the biodegradation of PVA. From the results, *Eutypella* sp. BJ showed little difference in degradation between PVA 1788 and 1799, which have different degrees of saponification, and it exhibited relatively poor degradation ability for PVA 2488, which has a high degree of polymerization. The GPC analysis showed that *Eutypella* sp. BJ can assimilate PVA, decreasing its molecular weight and increasing its molecular weight distribution. The FTIR analysis indicated that some substances containing carbonyl groups may be produced during the degradation process. Unexpectedly, we discovered that *Eutypella* has never been reported to have the ability to degrade PVA so far, which may have promising application in the assimilation of macromolecular pollutants in the environment. With regard to research on PVA degradation, further work is needed to study its biochemistry and physiology: what enzymes and related genes have acted on the biodegradation of PVA? What are the biochemical pathways of PVA degradation by fungi? In addition, the production of industrial enzyme preparations is also an efficient strategy for the biological treatment of polymer wastes.

## Conflicts of interest

All contributing authors declare that they have no conflicts of interest.

## Acknowledgements

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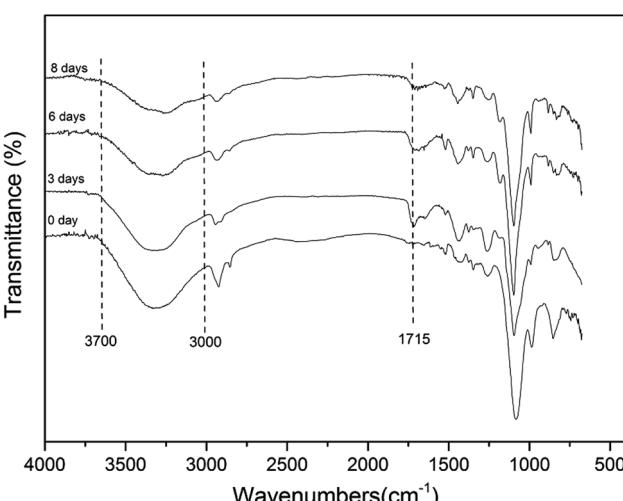


Fig. 7 FTIR spectra of PVA 1788 degradation at different incubation times.



## References

1 E. Chiellini, A. Corti, G. D. Sarto and S. D'Antone, Oxo-biodegradable polymers-effect of hydrolysis degree on biodegradation behaviour of poly(vinyl alcohol), *Polym. Degrad. Stab.*, 2006, **91**, 3397–3406.

2 F. F. Nord, Über die Dehydrierungsleistungen von *fusarium lini* B, *Naturwissenschaften*, 1936, **24**, 763.

3 T. Suzuki, Y. Ichihara, M. Yamada and K. Tonomura, Some Characteristics of *Pseudomonas* O-3 which Utilizes Polyvinyl Alcohol, *J. Agric. Chem. Soc. Jpn.*, 2008, **37**, 747–756.

4 Y. Watanabe, M. Morita, N. Hamada and Y. Tsujisaka, Formation of Hydrogen Peroxide by a Polyvinyl Alcohol Degrading Enzyme, *Agric. Biol. Chem.*, 1975, **39**, 2447–2448.

5 T. Hatanaka, N. Asahi and M. Tsuji, Purification and Characterization of Poly(vinyl alcohol) Dehydrogenase from *Pseudomonas* sp. 113P3, *Biosci., Biotechnol., Biochem.*, 1995, **59**, 1813–1816.

6 M. Morita, N. Hamada, K. Sakai and Y. Watanabe, Purification and Properties of Secondary Alcohol Oxidase from a Strain of *Pseudomonas*, *Agric. Biol. Chem.*, 1979, **43**, 1225–1235.

7 D. Jendrossek, in *Biopolymers*, ed. W. Babel and A. Steinbüchel, Springer, Heidelberg, 2001, vol. 71, ch. 10, pp. 293–325.

8 M. Amann and O. Minge, Synthetic Biodegradable Polymers, in *Advances in Polymer Science*, ed. B. Rieger, A. Künkel, G. W. Coates, R. Reichardt, E. Dinjus and T. A. Zevaco, Springer, Heidelberg, 2011, ch. 5, vol. 245, pp. 137–172.

9 M. Shima, S. Kishida, S. Harayama and T. Tamogami, The gene pvaB encodes oxidized polyvinyl alcohol hydrolase of *Pseudomonas* sp. strain VM15C and forms an operon with the polyvinyl alcohol dehydrogenase gene pvaA, *Microbiology*, 2000, **146**, 649–657.

10 S.-i. Tsujiyama, T. Nitta and T. Maoka, Biodegradation of polyvinyl alcohol by *Flammulina velutipes* in an unsubmerged culture, *J. Biosci. Bioeng.*, 2011, **112**, 58–62.

11 S. Tsujiyama and A. Okada, Biodegradation of polyvinyl alcohol by a brown-rot fungus, *Fomitopsis pinicola*, *Biotechnol. Lett.*, 2013, **35**, 1907–1911.

12 G. A. I. Mejía, O. B. L. López and P. A. Mulet, Biodegradation of poly(vinyl alcohol) with enzymatic extracts of *phanerochaete chrysosporium*, *Macromol. Symp.*, 2015, **148**, 131–147.

13 Q. Ding, G. Du and J. Chen, Isolation and Culture Characterization of a New Polyvinyl Alcohol-Degrading Strain: *Penicillium* sp. WSH02-21, *World J. Microbiol. Biotechnol.*, 2004, **20**, 587.

14 L. Jecu, A. Gheorghe, A. Rosu, I. Raut, E. Grosu and M. Ghirea, Ability of Fungal Strains to Degrade PVA Based Materials, *J. Polym. Environ.*, 2010, **18**, 284–290.

15 L. D. C. Rj, C. Gby and G. T. Lonergan, Enhanced degradation of polyvinyl alcohol by *Pycnoporus cinnabarinus* after pretreatment with Fenton's reagent, *Appl. Environ. Microbiol.*, 1999, **65**, 1798–1800.

16 T. Mori, M. Sakimoto, T. Kagi and T. Sakai, Enantioselective Oxidation of Diols by Secondary Alcohol Dehydrogenase from *Geotrichum* sp. WF9101, *Biosci., Biotechnol., Biochem.*, 1996, **60**, 1191–1192.

17 T. Mori, M. Sakimoto, T. Kagi and T. Sakai, Isolation and Characterization of a Strain of *Bacillus megaterium* That Degrades Poly(vinyl alcohol), *Biosci., Biotechnol., Biochem.*, 1996, **60**, 330–332.

18 T. Mori, M. Sakimoto, T. Kagi and T. Sakai, Degradation of Vinyl Alcohol Oligomers by *Geotrichum* sp. WF9101, *Biosci., Biotechnol., Biochem.*, 1996, **60**, 1188–1190.

19 T. Mori, M. Sakimoto, T. Kagi and T. Sakai, Secondary alcohol dehydrogenase from a vinyl alcohol oligomer-degrading *Geotrichum fermentans*; stabilization with Triton X-100 and activity toward polymers with polymerization degrees less than 20, *World J. Microbiol. Biotechnol.*, 1998, **14**, 349–356.

20 F. Kawai and X. Hu, Biochemistry of microbial polyvinyl alcohol degradation, *Appl. Microbiol. Biotechnol.*, 2009, **84**, 227.

21 R. Lipsa, N. Tudorachi, V. C. Grigoraş and C. Vasile, Degradation of poly(vinyl alcohol)-graft-lactic acid copolymers by *Trichothecium roseum* fungus, *J. Appl. Polym. Sci.*, 2015, **132**, DOI: 10.1002/app.41777.

22 Y. Liu, Y. Deng, P. Chen, M. Duan, X. Lin and Y. Zhang, Biodegradation analysis of polyvinyl alcohol during the compost burial course, *J. Basic Microbiol.*, 2019, **59**, 368–374.

23 G. P. Rédei, in *Encyclopedia of Genetics, Genomics, Proteomics and Informatics*, Springer, Dordrecht, 3rd edn, 2008, ch. 3187, DOI: 10.1007/978-1-4020-6754-9\_3188.

24 M. Isaka, S. Palasarn, W. Prathumpai and P. Laksanacharoen, Pimarane Diterpenes from the Endophytic Fungus *Eutypella* sp BCC 13199, *Chem. Pharm. Bull.*, 2011, **59**, 1157–1159.

25 X. L. Wang, K. L. Sun and B. Wang, Bioactive Pimarane Diterpenes from the Arctic Fungus *Eutypella* sp D-1, *Chem. Biodivers.*, 2018, **15**, 7.

26 Y. Zhou, Y.-X. Zhang, J.-P. Zhang, H.-B. Yu, X.-Y. Liu, X.-L. Lu and B.-H. Jiao, A new sesquiterpene lactone from fungus *Eutypella* sp. D-1, *Nat. Prod. Res.*, 2017, **31**, 1–6.

27 J.-B. Lee, S.-Y. Park, K. Shin, C.-P. Jeon, J.-E. Kim and G.-S. Kwon, Biodegradation of Organochlorine Insecticide Endosulfan by the Fungus *Eutypella* sp. KEF-1, *Korean Journal of Environmental Agriculture*, 2011, **30**, 202–208.

28 T. Suzuki, Y. Ichihara, M. Yamada and K. Tonomura, Some Characteristics of *Pseudomonas* O-3 which Utilizes Polyvinyl Alcohol, *Agric. Biol. Chem.*, 1973, **37**, 747–756.

29 R. Solaro, A. Corti and E. Chiellini, Biodegradation of poly(vinyl alcohol) with different molecular weights and degree of hydrolysis, *Polym. Adv. Technol.*, 2000, **11**, 873–878.

30 E. Chiellini, A. Corti and R. Solaro, Biodegradation of poly(vinyl alcohol) based blown films under different environmental conditions, *Polym. Degrad. Stab.*, 1999, **64**, 305–312.

31 T. Hatanaka, T. Kawahara, N. Asahi and M. Tsuji, Effects of the Structure of Poly(vinyl alcohol) on the Dehydrogenation Reaction by Poly(vinyl alcohol) Dehydrogenase from



*Pseudomonas* sp. 113P3, *Biosci., Biotechnol., Biochem.*, 1995, **59**, 1229–1231.

32 R. Solaro, A. Corti and E. Chiellini, Biodegradation of poly(vinyl alcohol) with different molecular weights and degree of hydrolysis, *Polym. Adv. Technol.*, 2000, **11**, 873–878.

33 M. Ullah, C. H. Weng, H. Li, S. W. Sun, H. Zhang, A. H. Song and H. Zhu, Degradation of polyvinyl alcohol by a novel bacterial strain *Stenotrophomonas* sp. SA21, *Environ. Technol.*, 2017, **39**, 1–20.

34 E. Chiellini, A. Corti, S. D'Antone and R. Solaro, Biodegradation of poly(vinyl alcohol) based materials, *Prog. Polym. Sci.*, 2003, **28**, 963–1014.

35 H. C. Flemming, Relevance of biofilms for the biodeterioration of surfaces of polymeric materials, *Polym. Degrad. Stab.*, 1998, **59**, 309–315.

