

Cite this: *RSC Adv.*, 2019, 9, 25405

On the road to cost-effective fossil fuel desulfurization by *Gordonia alkanivorans* strain 1B †

Marta Pacheco,  Susana M. Paixão, * Tiago P. Silva and Luís Alves *

Biodesulfurization (BDS) is an ecofriendly process that uses microorganisms to efficiently remove sulfur from fossil fuels. To make the BDS process economically competitive with the deep hydrodesulfurization process, which is currently used in the oil industry, it is necessary to improve several factors. One crucial limitation to be overcome, common within many other biotechnological processes, is the cost of the culture medium. Therefore, an important line of work to make BDS scale-up less costly is the optimization of the culture medium composition aiming to reduce operating expenses and maximize biocatalyst production. In this context, the main goal of this study was on the minimization of inorganic key components of sulfur-free mineral (SFM) medium in order to get the maximal production of efficient desulfurizing biocatalysts. Hence, a set of assays was carried out to develop an optimal culture medium containing minimal amounts of nitrogen (N) and magnesium (Mg) sources and trace elements solution (TES). These assays allowed the design of a SFMM (SFM minimum) medium containing 85% N-source, 25% Mg-source and 25% TES. Further validation consisted of testing this minimized medium using two carbon sources: the commercial C-source (glucose + fructose) *versus* Jerusalem artichoke juice (JAJ) as a cheaper alternative. SFMM medium allowed microbial cells to almost duplicate their specific desulfurization rate (q_{2-HBP}) for both tested C-sources, namely from 2.15 to 3.39 $\mu\text{mol g}^{-1}$ (DCW) h^{-1} for Fru + Glu and from 1.91 to 3.58 $\mu\text{mol g}^{-1}$ (DCW) h^{-1} for JAJ, achieving a similar net 2-hydroxybiphenyl produced per g of consumed sugar ($\sim 17 \mu\text{mol g}^{-1}$). These results point out the great advantage of using cheaper culture medium that in addition enhances the bioprocess effectiveness, paving the way to a sustainable scale-up for fossil fuel BDS.

Received 13th May 2019
Accepted 7th August 2019

DOI: 10.1039/c9ra03601f

rsc.li/rsc-advances

1. Introduction

With the emergence of hybrid and non-petroleum-based engines, it was expected that oil consumption would decrease drastically. However, this effect was only observed in developed countries where alternative energy sources are well established and the population economic power allows access to these new technologies.¹ In developing countries, the demand for cheaper, reliable and comfortable means of transportation is high; therefore, vehicles with internal combustion engines are the preferred choice. Moreover, the recent advances of the technology necessary to remove oil from difficult-to-drill sites, such as shale formations or oil sands, have revolutionized the energy market, bringing oil prices to a historical low in 2015,² making oil an attractive and well-known energy source. However, the combustion of fossil fuels releases many hazardous components such as CO_2 , NO_x , SO_x and other sulfur compounds. As a response to the increasing concern with the environmental

and health problems related with the sulfur levels in fossil fuels, countries have begun to implement strict sulfur concentration limits in on-road and off-road gasoline and diesel requirements. These will become stricter in the foreseeable future towards ultra-low sulfur fuels, approaching zero sulfur emissions from burned fuels. Therefore, the efficiency of the desulfurization technologies becomes a key point.³

Sulfur (S) is a major component of crude oil, along with carbon and hydrogen, ranging from 0.03 to 7.89% (g g^{-1}), and between 500 and 5000 mg L^{-1} in the distillate fraction used to make diesel oil.^{4,5} High viscosity and density crudes (such as shale oils) tend to have higher sulfur concentrations 3.09% (g g^{-1}), especially in its organic forms (*e.g.* sulfides and thiophenic compounds) that can account for 75% of total sulfur. Nowadays, the most used sulfur removing process in the refineries is the hydrodesulfurization (HDS), which uses high temperatures; high pressures conjugated with elemental hydrogen and advanced metal catalysts to remove sulfur from crude oil. However, some types of organic sulfur molecules are very recalcitrant to this process, because the sulfur molecule is enclosed within an aromatic ring. Biodesulfurization (BDS) is a biotechnological process corresponding to HDS, since it removes sulfur from complex compounds without the need for

LNEG – Instituto Nacional de Energia e Geologia, IP, Unidade de Bioenergia, Estrada do Paço do Lumiar, 22, 1649-038, Portugal. E-mail: susana.alves@lneg.pt; luis.alves@lneg.pt

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c9ra03601f



high temperatures or pressures, elemental hydrogen or metal catalysts. In the last decades, BDS has drawn wide attention because of its green processing of fossil fuels. Since BDS is more efficient and less expensive than HDS in removing sulfur from refractory heterocyclic compounds present in crude oil, it could be used in oil refineries as a complement to achieve ultra-low sulfur diesel.³ Indeed, BDS can be used to desulfurize heavy oils, like shale oils, which have high thiophene concentration.

The preferred microorganisms for BDS are those that remove the sulfur from the hydrocarbons through 4S pathway metabolism, regulated by a *dsz* operon, in which the calorific value of the fuel is preserved. One of those microorganisms is the bacterium *Gordonia alkanivorans* strain 1B, isolated by Alves *et al.*,⁶ from oil contaminated soil samples from Parque das Nações (former Petrogal location, Lisbon, Portugal). In the last decade, several works have demonstrated the great potential of *G. alkanivorans* strain 1B towards fossil fuel BDS.^{3,7–12}

However, this bioprocess still has a few limitations, such as the cost of the culture medium for the biocatalysts production.^{12–16} In order to reduce the production costs, it is important to search for cheaper carbon sources which can contribute to produce less expensive microbial biomass. Due to their low cost, the utilization of agroindustrial residues or other abundant materials as carbon sources is one of the most followed strategies in biotechnology processes.^{10–12,17} Silva *et al.*¹² described an optimized dibenzothiophene desulfurization process by *G. alkanivorans* strain 1B using Jerusalem artichoke juice (JAJ) as carbon source ($\sim 25 \text{ g L}^{-1}$ total sugars), achieving high desulfurization results (2-hydroxybiphenyl (2-HBP) production rate = $28.2 \mu\text{M h}^{-1}$ and $q_{2\text{-HBP}} = 5.06 \mu\text{mol g}^{-1} (\text{DCW}) \text{ h}^{-1}$), and highlighted JAJ potential as a promising alternative carbon source towards further scale-up assays.

Indeed, Jerusalem artichoke (*Helianthus tuberosus*) is an herbaceous perennial tuberous plant that accumulates high concentrations of inulin in its tubers. Inulin is a linear polymer in which a variable number of D-fructose units are linked by $\beta(2 \rightarrow 1)$ bonds, terminated by a D-glucose residue through a sucrose-type linkage at the reducing end $\alpha(2 \rightarrow 1)$. So, this polysaccharide can be easily converted into a sugar-rich juice ($\text{JAJ} \geq 80\%$ fructose) and further used as carbon source.

In this context, this work consisted on the minimization of inorganic key components (N-source; Mg-source and micronutrients) of *G. alkanivorans* strain 1B culture medium aiming to get a total sugar consumption (C-source: 10 g L^{-1} total reducing sugars) and by the further validation of Jerusalem artichoke juice as a cheaper alternative C-source in order to achieve a significant decrease on the overall production costs of BDS biocatalysts.

II. Materials and methods

A. Jerusalem artichoke juice

Jerusalem artichoke tubers cultivated in a forest soil at Oleiros, Portugal were harvested and further processed, as described in Silva *et al.*¹² and in Paixão *et al.*,¹⁸ to produce Jerusalem artichoke juice (JAJ). The JAJ was hydrolyzed at pH 2 and 55°C for 48 hours to convert inulin into fructose and glucose and then

filtered ($0.20 \mu\text{m}$ membrane filters). For total sulfate precipitation, sterile BaCl_2 at 0.5% (w/v) was added to a filter sterilized juice adjusted to pH 8.73.¹² This JAJ was shaken vigorously, incubated at 30°C for 36 hours and then filtered to remove all the BaSO_4 ($0.45 \mu\text{m}$ membranes) and again filter sterilized ($0.20 \mu\text{m}$ membrane filters). So, the precipitated JAJ contained $\sim 0 \text{ mg L}^{-1}$ of sulfate, and a total sugar concentration of 160 g L^{-1} , of which 80% was fructose and 20% glucose, indicating complete inulin conversion.

B. Microorganism and culture media

The microorganism used in this study was the bacterium *G. alkanivorans* strain 1B, isolated in our laboratory.⁶ The basal salts medium used for cultivation of this microorganism was a sulfur-free mineral (SFM) culture medium containing $1.22 \text{ g L}^{-1} \text{NH}_4\text{Cl}$, $2.55 \text{ g L}^{-1} \text{KH}_2\text{PO}_4$, $2.55 \text{ g L}^{-1} \text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$ and $0.17 \text{ g L}^{-1} \text{MgCl}_2 \cdot 6\text{H}_2\text{O}$. This medium was supplemented with 0.50 mL L^{-1} of a sulfur-free trace elements solution (TES)⁹ and its final pH was adjusted to 7.5, before being autoclaved at 121°C , 1 atm for 15 min. Pure carbon sources, namely glucose and fructose, were dissolved in Millipore water in 50% (w/v) concentrated solutions. Pure carbon sources and Jerusalem artichoke juice (JAJ) were filter sterilized before being added to the culture medium in aseptic conditions, to an initial concentration of about 10 g L^{-1} total reducing sugars.

Prior the BDS assays, the bacterial inoculum was prepared by growing strain 1B with fructose (10 g L^{-1}) as the only carbon source and $250 \mu\text{M}$ DBT as the only sulfur source, within 150 mL culture medium in 500 mL Erlenmeyer, at 30°C and 150 rotations per minute (rpm) for 72 hours, in an orbital-shaker.

C. Biotransformation assays: culture medium minimization

Aiming to minimize the concentrations of key inorganic components for a full consumption of 10 g L^{-1} of C-source without limiting neither the growth nor the desulfurization ability of strain 1B, a set of BDS assays were performed testing different formulations of the SFM culture medium aforementioned. The key culture medium inorganic components minimized were: nitrogen-source (NH_4Cl), magnesium-source ($\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$) and TES, ranging from 0% to 75% of the SFM original amounts. The details of the tested medium formulations, F#1 to F#13, are presented in Table 1, being F#1 the original SFM medium (control assay). In all tested formulations, the carbon and sulfur sources concentration were maintained constant [10 g L^{-1} of total reducing sugars in a ratio of 2 : 8 glucose/fructose (to mimic the sugar concentration of the Jerusalem artichoke juice); $250 \mu\text{M}$ of dibenzothiophene (DBT) as sulfur source]. Following, validation assays (F#14 and F#15) were performed for the designed minimized culture medium (SFMM medium) using mix glucose/fructose vs. JAJ as C-source.

BDS assays were performed in 500 mL Erlenmeyer-flasks with about 150 mL of culture medium formulations (F#1 to F#15), inoculated with 2% (v/v) of bacterial inoculum prepared as above described, and incubated at 30°C and 150 rpm from about 96 to 124 h, in an orbital-shaker. All BDS assays were carried out in triplicates.



Table 1 Different formulations of the SFM medium, with the amounts of the key components (NH_4Cl , $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$ and TES) tested in shake-flask assays towards the design of the optimal minimal culture medium to produce effective desulfurizing biocatalysts

		SFM culture medium composition				
	Medium formulations	Components minimized				Sulfur source
Set assays	(F#)	NH ₄ Cl (g L ⁻¹)	MgCl ₂ ·6H ₂ O (g L ⁻¹)	TES (mL L ⁻¹)	Carbon source (10 g L ⁻¹) ^c	(250 μM)
Control (^a SFM medium)	#1	1.22	0.17	0.50	8 Fru + 2 Glu	DBT
	#1'				JAJ	
N-source minimization	#2	0.92 (75%)	0.17	0.50	8 Fru + 2 Glu	DBT
	#3	0.61 (50%)				
	#4	0.31 (25%)				
Mg-source minimization	#5	1.22	0.13 (75%)	0.50		
	#6		0.09 (50%)			
	#7		0.04 (25%)			
TES minimization	#8	1.22	0.17	0.38 (75%)		
	#9			0.25 (50%)		
	#10			0.13 (25%)		
	#11			0.03 (5%)		
	#12			0.01 (2.5%)		
	#13			0 (0%)		
Minimized medium (^b SFMM)	#14	1.04 (85%)	0.04 (25%)	0.13 (25%)	8 Fru + 2 Glu	DBT
	#15				JAJ	

^a SFM medium – sulfur-free mineral medium (default medium for shake-flask assays). ^b SFMM medium – SFM minimum medium for shake-flask.

^c 10 g L^{-1} Total sugars; 8 Fru: fructose 8 g L^{-1} ; 2 Glu: glucose 2 g L^{-1} ; JAJ: sulfate precipitated Jerusalem artichoke juice.

D. Analytical methods

The culture growth was monitored by determining the sugars consumption (glucose/fructose) using high-performance liquid chromatography (HPLC) instrumentation, the optical density at 600 nm ($\text{OD}_{600\text{nm}}$) and the dry cell weight (DCW), as described by Fernandes *et al.*¹⁹

DBT desulfurization was evaluated by measuring 2-HBP production, which is the final product of the DBT desulfurization through the 4S-pathway. The samples (0.750 mL) were acidified with 0.025 mL of HCl (4 M) and then a liquid–liquid extraction with ethyl acetate was performed on a vortex (5 minutes), in order to extract 2-HBP and DBT. After phase separation, the organic phase was analyzed by gas-chromatography (GC) in a gas-chromatograph (Chrompack, Model CP9001, Middelburg, The Netherlands) equipped with a flame ionization detector. A 10% CP-5 CB on 100/120 MESH Chromosorb W-HP column was used with nitrogen as the carrier gas. The chromatograph oven start temperature was 130 °C for 3 minutes and the end temperature 230 °C maintained for 3 minutes (heating rate of 6 °C min^{-1}). The injector and detector temperatures were set for 280 and 290 °C, respectively. In all GC measurements 4-methyl-DBT was used as internal standard to minimize variations.

E. Statistical validation

Unless indicated otherwise, the results presented in graphs are mean values derived from three replicates (*i.e.* 3 independent BDS assays performed in the same conditions). The corresponding standard deviation ($n = 3$) is represented as error bars.

To validate the statistical significance of the results observed with the cultures grown with the final minimized formulations (F#14 and F#15), and compare them with the initial formulations (F#1 and F#1'), an ANOVA (single factor) test was used, and a significant difference was considered at a level of $p < 0.05$.

III. Results and discussion

A. Biodesulfurization assays: culture medium minimization

This study consisted on the optimization of the SFM culture medium for the maximum production of efficient desulfurizing biocatalysts (*G. alkanivorans* strain 1B active cells) taking into account the lowest operation costs towards a future BDS process scale-up. In this context, key components of the SFM medium used to grow strain 1B (NH_4Cl , MgCl_2 and TES) were reduced to a minimal amount while aiming to have no limitation both on the full consumption of the carbon source (10 g L^{-1}) and on the biodesulfurization ability. The phosphate compounds of the culture medium were not altered since, in the shake-flask growth assays, their presence is crucial for pH maintenance (buffer effect). Indeed, the reduction of their concentration would imply a sharp pH decrease on culture medium and consequently a decrease of BDS effectiveness.

A.1. Nitrogen-source. Nitrogen (N) is a major intervenient in cellular growth, representing 14% of the dry cell weight. It is found in the composition of amino acids, nucleic acids and enzymes/coenzymes, therefore, it is one of the major macronutrients of most culture media.²⁰

Hence, in the first set of BDS assays, three formulations with different amounts of N-source (NH_4Cl), namely 75% (F#2), 50% (F#3) and 25% (F#4) of the amount in the original medium (1.22 g L^{-1} = 100% N) were tested against the original SFM



medium as a control (F#1, default SFM medium), for comparison. Fig. 1A and B shows the time course profiles of cellular growth (OD_{600nm}) and sugar consumption for the four formulations (F#1 to F#4, Table 1) containing different N-source concentrations. The bacterial culture attained maximal OD_{600nm} (9.98 – F#1; 10.39 – F#2) with complete sugar consumption (10 g L^{-1}) for both F#1 and F#2 formulations. However, for the remaining formulations (F#3 and F#4) a lower growth was observed. For F#3, an OD_{600} of 7.87 was attained, with a complete fructose consumption but with a remaining of 0.9 g L^{-1} of glucose; and for F#4 was attained an OD_{600nm} of only 4, remaining more than half of the total sugars (3.7 g L^{-1} Fru and 1.5 g L^{-1} Glu) in culture medium.

These results indicate that the amount of the N-source greatly influenced the bacterial behavior. F#4 (25% N) was clearly insufficient to ensure normal cell growth, resulting in an abrupt growth stop when all other nutrients were still in excess. Comparatively to F#4, F#3 (50% N) allowed a growth

enhancement, almost doubling the maximum OD_{600nm} attained (from 4 to 7.87), but without the full consumption of the available sugars. In F#3, fructose was fully consumed but most of the glucose was left untouched. This preferential consumption of fructose was expected since strain 1B is a fructophilic microorganism.⁹ F#2 (75% N) was the only formulation that allowed complete growth and sugar consumption, achieving a similar growth curve to the one obtained with 100% N (F#1, original medium).

The desulfurization ability of the strain 1B depends mostly on cell physiology, which is influenced by the amount of nutrients and carbon in the culture medium. In Fig. 1C is presented the DBT desulfurization by strain 1B grown in the formulations F#1 to F#4, with decreasing amounts of N-source (100–25%). These results point out for the limitation of desulfurization ability by strain 1B due to the lack of N-source in the culture medium. In F#1, the bacterial culture presented a maximum 2-HBP production of $227.2\text{ }\mu\text{M}$ and a maximum specific 2-HBP production rate ($q_{2\text{-HBP}}$) of $2.5\text{ }\mu\text{mol g}^{-1}\text{ (DCW) h}^{-1}$, achieving a total consumption of the $250\text{ }\mu\text{M}$ of DBT by the end of the growth. In F#2, the $q_{2\text{-HBP}}$ obtained was $2.4\text{ }\mu\text{mol g}^{-1}\text{ (DCW) h}^{-1}$ with a maximum 2-HBP of $167.1\text{ }\mu\text{M}$ and a remaining $27\text{ }\mu\text{M}$ of DBT. In F#3 and F#4, the $q_{2\text{-HBP}}$ obtained was $2.25\text{ }\mu\text{mol g}^{-1}\text{ (DCW) h}^{-1}$ and $1.41\text{ }\mu\text{mol g}^{-1}\text{ (DCW) h}^{-1}$, and the maximum 2-HBP produced was $110.1\text{ }\mu\text{M}$ and $47.8\text{ }\mu\text{M}$, respectively. Relatively to DBT, $69\text{ }\mu\text{M}$ and $202\text{ }\mu\text{M}$ were not consumed in F#3 and F#4, respectively. The desulfurization results for both F#3 and F#4 formulations can be supported by their respective data for cellular growth and sugar consumption, showed in Fig. 1A and B. In fact, a NH_4Cl concentration $\leq 75\%$ of the original amount (F#2 to F#4) seems to not be sufficient for strain 1B to achieve maximum growth with total C-source consumption and full desulfurization of the $250\text{ }\mu\text{M}$ DBT. The overall growth, sugar consumption profiles and $q_{2\text{-HBP}}$ were very similar for F#1 and F#2; however a maximum 2-HBP production of $167.1\text{ }\mu\text{M}$ was observed in F#2. In contrast, in F#1 the maximum 2-HBP production was $227.2\text{ }\mu\text{M}$ ($\sim 60\text{ }\mu\text{M}$ higher), despite of the consumption of almost all DBT ($223\text{ }\mu\text{M}$) in the F#2. Hence, these results point to the need of using a concentration of NH_4Cl $> 75\%$ of the original amount as the minimal N-source towards a minimized SFM medium.

Since nitrogen is a major component of enzymes and functions as a co-factor for innumerable essential proteins, it is expected that insufficient amounts will lead to deficiencies in cellular processes.^{20,21} This fact can support the sequential decrease in the desulfurization rate and, consequently, the maximum 2-HBP production observed in the tested formulations, namely from F#1 (100% N) to F#4 (25% N). Only in F#1 (default medium), the N amount was sufficient to support the complete cell growth, using the totality of C-source available, and the biodesulfurization of all the DBT. In the other formulations (F#2 to F#4), with decreasing NH_4Cl concentrations, increasing limitations at different levels (e.g. growth rate, sugar consumption, 2-HBP production or $q_{2\text{-HBP}}$) were observed.

Therefore, based on results from Fig. 1B and C, 85% N was extrapolated as an appropriate amount to be further tested as minimal N-source within a SFM minimized medium (F#14 and F#15 in Table 1).

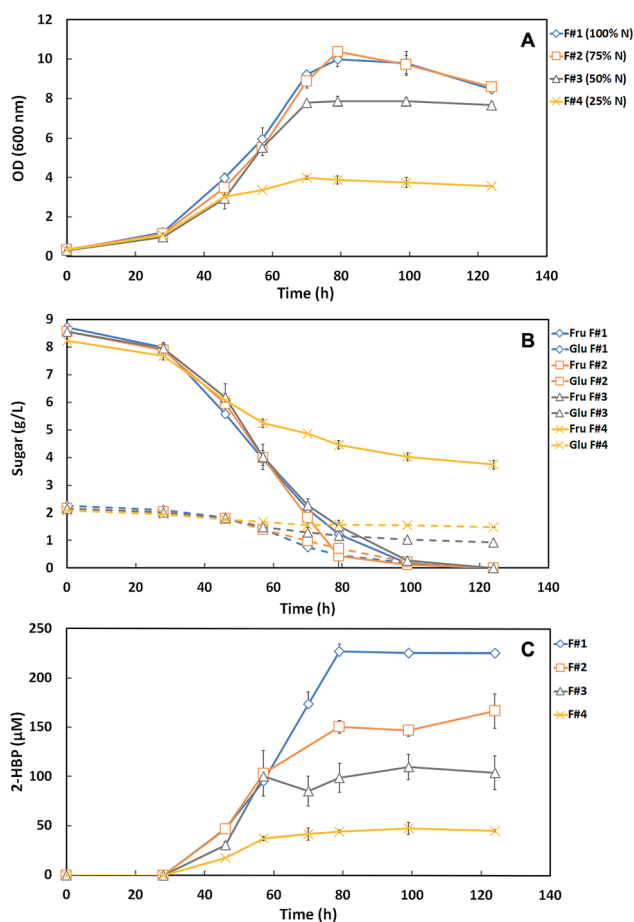


Fig. 1 Time course profiles for cellular growth (A), sugar consumption (B) and desulfurization curves (2-HBP production) (C) of *G. alkanivorans* strain 1B in the shake-flask cultures with different culture medium formulations (F#1 to F#4) containing decreasing concentrations of N-source (NH_4Cl). F#1 corresponds to the control assay with the original SFM medium (default medium). All formulations contain 10 g L^{-1} total sugars (as carbon source) and $250\text{ }\mu\text{M}$ DBT as sulfur source. Fru: fructose; Glu: glucose. Standard deviation ($n = 3$) is represented as error bars.



A.2. Magnesium-source. Magnesium (Mg) is an element essential for cell division and protoplasm formation, being especially important for Gram-positive bacteria, since it is incorporated in the structure of the peptidoglycan which is 90% of the cellular wall.²² The SFM medium usually used for the growth of *G. alkanivorans* strain 1B has 0.17 g L⁻¹ of MgCl₂·6H₂O (100% Mg). To select the minimal amount of Mg-source that supports bacterial growth and desulfurization ability, several formulations with decreasing concentrations of Mg-source were tested (75% (F#5) to 25% (F#7) of the original amount, as described in Table 1). The results obtained were compared with those of 100% Mg (F#1, control assay with default SFM medium).

Fig. 2A and B shows the time course profiles of cellular growth (OD_{600nm}) and sugar consumption for the tested medium formulations with the different Mg-source concentrations: F#1 (100% Mg = control with original medium), F#5 (75% Mg), F#6 (50% Mg) and F#7 (25% Mg). These results show that strain 1B growth and sugar consumption profiles for F#5 and F#6 were very similar to those observed for F#1. A total

consumption of the 10 g L⁻¹ total sugars, within 120 h, was observed for these three formulations achieving an OD_{600nm} around 10. However, when F#7 (25% Mg) was used as culture medium, the strain 1B only attained a maximum OD_{600nm} of 8.66 with an overall growth profile slightly below the others, despite having a similar maximal growth rate ($\mu_{\text{Max}} \sim 0.06 \text{ h}^{-1}$ to all). Moreover, strain 1B in F#7 medium consumed all the C-source (Fru + Glu) with the same sugar consumption profile as the other formulations. This fact contrasts with what was observed when N-source was limiting, where accumulation of sugars was detected.

Fig. 2C presents the desulfurization curves (2-HBP production) by the strain 1B grown in the different Mg-source medium formulations. As already described in the N-source minimization assays, for the control formulation (F#1) a $q_{2\text{-HBP}}$ of 2.45 $\mu\text{mol g}^{-1}$ (DCW) h⁻¹ and a maximum 2-HBP of 227.2 μM were achieved. For the other test formulations, the respective $q_{2\text{-HBP}}$ and maximum 2-HBP obtained were: 2.67 $\mu\text{mol g}^{-1}$ (DCW) h⁻¹ and 244.0 μM (F#5), 3.01 $\mu\text{mol g}^{-1}$ (DCW) h⁻¹ and 244.8 μM (F#6), and 3.37 $\mu\text{mol g}^{-1}$ (DCW) h⁻¹ and 250.0 μM (F#7). In all these formulations, strain 1B was capable of fully desulfurize the amount of DBT provided (250 μM).

These results show an evident enhancement of the maximum specific desulfurization rate ($q_{2\text{-HBP}}$) with the decrease of the Mg-source/C-source ratio, from 2.45 $\mu\text{mol g}^{-1}$ (DCW) h⁻¹ (ratio 100% Mg per 10 g L⁻¹ C in F#1) to 3.37 $\mu\text{mol g}^{-1}$ (DCW) h⁻¹ (ratio 25% Mg per 10 g L⁻¹ C in F#7). This fact is in agreement with results previously reported in studies of DBT desulfurization by strain 1B. Alves and Paixão⁹ reported a $q_{2\text{-HBP}}$ of 2.12 $\mu\text{mol g}^{-1}$ (DCW) h⁻¹ by strain 1B when SFM medium with a ratio 100% Mg per 10 g L⁻¹ Fru was used and Silva (23) reported a $q_{2\text{-HBP}}$ of 6.57 $\mu\text{mol g}^{-1}$ (DCW) h⁻¹ by strain 1B when SFM medium with a ratio 100% Mg per 25 g L⁻¹ Fru was used. These results also pointed out for the stimulatory effect on desulfurization by the decrease of Mg-source/C-source ratio.

Hence, F#7 (25% Mg) allowed a similar maximum growth rate, despite the lower value for maximum OD_{600nm}, and the enhancement of desulfurization. Therefore, it was stipulated as an appropriate amount to be further tested as minimal Mg-source within a SFM minimized medium.

A.3. Trace element solution (TES). For the SFM medium used to grow *G. alkanivorans* strain 1B, a sulfur-free TES stock solution⁹ is usually added in a concentration of 0.5 mL L⁻¹. Therefore, as in prior assays performed for N and Mg sources minimization, different formulations with decreasing concentrations of TES (F#8 to F#13, described in Table 1) were tested and compared with control (100% TES, F#1) in order to evaluate the minimum concentration of the TES needed by the bacterium to grow and desulfurize DBT. Fig. 3A and B presents the cellular growth (OD_{600nm}) and sugar consumption profiles for strain 1B grown on the different culture medium formulations containing decreasing concentrations of TES (100% to 0%). In Fig. 3B, only the sugar consumption profiles for the formulations that significantly differ from control (F#1) are represented.

These results show that the media formulations F#1 (100% TES), F#8 (75% TES), F#9 (50% TES) and F#10 (25% TES) induced a similar behavior on strain 1B and consequently the

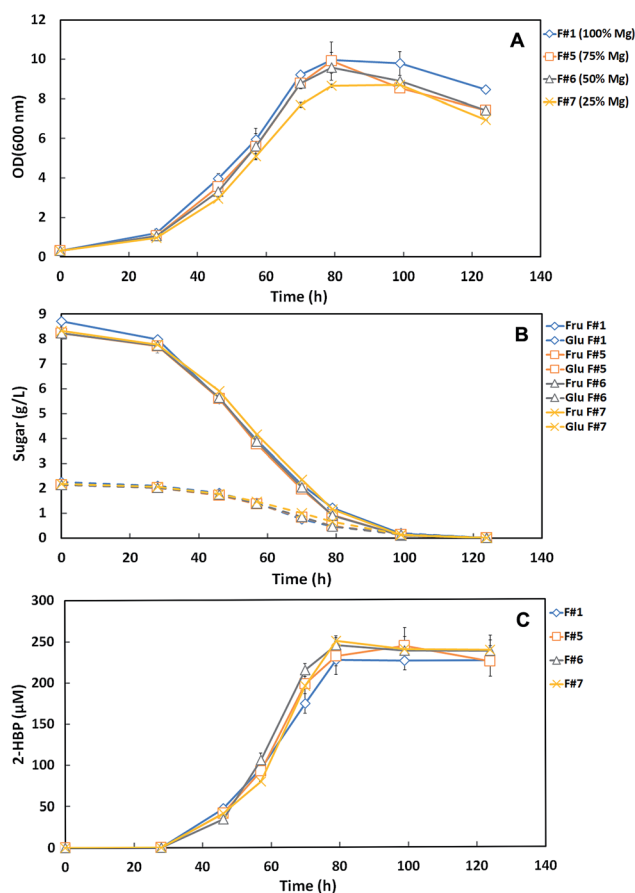


Fig. 2 Time course profiles for cellular growth (A), sugar consumption (B) and desulfurization curves (2-HBP production) (C) of *G. alkanivorans* strain 1B in the shake-flask cultures with different medium formulations (F#1, F#5 to F#7) containing decreasing concentrations of Mg-source (MgCl₂·6H₂O). F#1 corresponds to the control assay (100% Mg in SFM default medium). All formulations contain 10 g L⁻¹ total sugars (as carbon source) and 250 μM DBT as sulfur source. Fru: fructose; Glu: glucose. Standard deviation ($n = 3$) is indicated by the error bars.



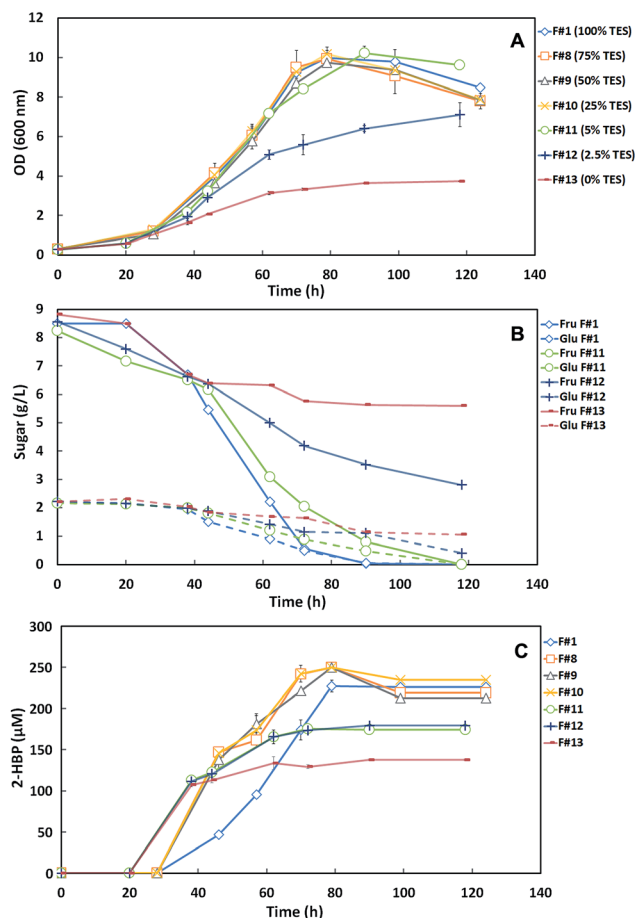


Fig. 3 Time course profiles for cellular growth (A) sugar consumption (B) and desulfurization curves (2-HBP production) (C) of *G. alkanivorans* strain 1B in the shake-flask cultures with different medium formulations (F#1, F#8 to F#13) containing decreasing concentrations of TES. In graphic (B) only F#1 and F#11 to F#13 are represented. F#1 corresponds to the control assay with the default medium. All formulations contain 10 g L⁻¹ total sugars (as carbon source) and 250 μM DBT as sulfur source. Fru: fructose; Glu: glucose. Standard deviation ($n = 3$) is indicated by the error bars.

growth curves and the sugar consumption profiles were identical. Indeed, for the control (F#1) and for F#8 to F#10 formulations a maximum OD_{600nm} ~ 10 was attained, with total sugar consumption (10 g L⁻¹) within 124 h. In the absence of the TES (F#13) the maximum OD_{600nm} achieved by strain 1B was only 3.73 with an accumulation of 6.6 g L⁻¹ of sugars. In F#12 (2.5% TES), the bacterium was able to attain an OD_{600nm} of 7.11 but still accumulated 3.2 g L⁻¹ of total sugars. However, in F#11 (5% TES), a maximum OD_{600nm} of 10.22 with a total sugar consumption was achieved, similarly to that observed for medium formulations with ≥25% TES (F#1, F#8 to F#10). These results indicate that a TES concentration ≥5% is required for maximum growth of strain 1B and full C-source consumption. However, when 5% TES was used the sugar consumption rate was somewhat slower than for ≥25% TES, prolonging the fermentation from 90–100 h to a total of 118 h.

Fig. 3C presents the desulfurization results as the time course profiles of 2-HBP production by strain 1B cultivated in

the different formulations with decreasing TES concentrations. These results show a desulfurization enhancement with the decrease of TES concentration up to 25% in the culture medium. However, for concentrations <25% TES, namely in F#11 (5%), F#12 (2.5%) and F#13 (0%), the reduction of trace elements was limiting and the desulfurization stopped within about 60 h, attaining lower levels of 2-HBP production. At 0% TES the strain 1B desulfurization was probably mediated by trace elements available from culture medium used to grow the bacterium. Table 2 summarizes the main metabolic parameters (maximum $q_{2\text{-HBP}}$ and maximum 2-HBP produced) for the DBT desulfurization by *G. alkanivorans* strain 1B grown in the formulations with decreasing TES concentrations. In agreement with the results presented in Fig. 3C, the $q_{2\text{-HBP}}$ values for F#8 (2.80 μmol g⁻¹ (DCW) h⁻¹), F#9 (3.19 μmol g⁻¹ (DCW) h⁻¹) and F#10 (3.02 μmol g⁻¹ (DCW) h⁻¹) were higher than the value of the control (F#1, 2.45 μmol g⁻¹ (DCW) h⁻¹). In overall, these results seem to indicate that there is some inhibitory effect caused by the TES, since a sequential decrease of TES concentration up to 25% (F#10) produces cells with higher desulfurization ability than the control. In fact, F#10 with 25% TES induces an enhancement of about 23% of the specific desulfurization rate in comparison with the control (100% TES). Nevertheless, most of the trace elements are essential for strain 1B growth. Therefore, an excessive reduction of these micro-nutrients may result in a poor microbial growth and consequently cause the incomplete desulfurization of the DBT present in culture medium. This can be due to an alteration on essential enzymes structure, which need the nutrients supplied in the TES to function properly.^{24,25}

Since 25% TES (F#10) was the lowest concentration tested that allowed maximum cell growth and in addition it has stimulated desulfurization, this was the chosen amount as the most appropriated to be further tested as minimal TES within a SFM minimized medium.

B. Minimized medium: commercial versus alternative C-source

Using the N-source, Mg-source and TES minimal concentrations, determined in the prior assays, a SFM minimum (SFMM) medium containing 85% N, 25% Mg and 25% TES was designed (Fig. S1†). Further, two formulations (F#14 vs. F#15, Table 1) were tested in comparison with the correspondent original medium (F#1 vs. F#1') to evaluate the whole effect of the collective nutrients reduction and to validate the minimized culture medium towards a cost-effective DBT desulfurization process by *G. alkanivorans* strain 1B.

Indeed, as already referred, it is also crucial the replacement of the expensive commercial C-sources (glucose/fructose) by alternative and more sustainable C-sources when the goal is to reduce the overall production costs. In this context, based on the information previously reported by Silva *et al.*¹² highlighting the great potential of JAJ as a sustainable alternative C-source that enhances the DBT desulfurization ability of strain 1B, the validation assays were performed by testing the SFMM medium into two formulations using a different C-source: F#14 with Fru



Table 2 Main metabolic parameters for the DBT desulfurization by *G. alkanivorans* strain 1B in the formulations with decreasing TES concentrations (100% = default medium up to 0%)

Medium formulations	F#1 (100% TES)	F#8 (75% TES)	F#9 (50% TES)	F#10 (25% TES)	F#11 (5% TES)	F#12 (2.5% TES)	F#13 (0% TES)
$q_{2\text{-HBP}}$ ($\mu\text{mol g}^{-1}$ (DCW) h^{-1})	2.45	2.80	3.19	3.02	1.32	1.26	0.68
2-HBP produced (μM)	227.2	250.0	250.0	250.0	179.5	173.7	133.5

+ Glu versus F#15 with JAJ, simultaneously with their respective control (F#1 vs. F#1', Table 1). In these assays, 10 g L^{-1} of total sugars (Fru + Glu or JAJ) were used as C-source and $250 \mu\text{M}$ DBT as S-source.

Fig. 4A–C presents the cellular growth ($\text{OD}_{600\text{nm}}$), the sugar consumption and biodesulfurization profiles for strain 1B grown in both minimized medium formulations (F#14 and F#15) and respective controls (F#1 and F#1'). Comparing the growth profiles (Fig. 4A), it can be observed that cells grown using the commercial C-source (Fru + Glu) attained higher $\text{OD}_{600\text{nm}}$ values and higher growth rates than those obtained with JAJ. Using the commercial sugars as C-source, strain 1B

attained a maximum $\text{OD}_{600\text{nm}}$ of ~ 11 with both formulations, while cells grown with JAJ only attained a value of ~ 10 . The highest $\text{OD}_{600\text{nm}}$ value was observed on F#14 (11.61) after 70 hours. Growth rates were higher for both media with Fru + Glu as C-source (F#1 and F#14), however in all formulations the sugar consumption presented similar profiles, with total consumption within 98 h (Fig. 4B). These results were further validated by a single factor ANOVA test, which confirmed that the differences in maximum $\text{OD}_{600\text{nm}}$ obtained between the two carbon sources were statistically significant, with a level of $p < 0.005$.

Based on the results from Fig. 4, the main metabolic parameters associated to the growth and desulfurization by strain 1B in these four tested formulations (F#1 vs. F#14; F#1' vs. F#15) are summarized in Table 3. These results demonstrate that independently of the C-source used, the minimized medium for N-source, Mg-source and TES (SFMM medium) enhanced the desulfurization by strain 1B in comparison with the original culture medium (SFM medium), namely from $2.15 \mu\text{mol g}^{-1}$ (DCW) h^{-1} (F#1) to $4.39 \mu\text{mol g}^{-1}$ (DCW) h^{-1} (F#14) for Fru + Glu and from $1.91 \mu\text{mol g}^{-1}$ (DCW) h^{-1} (F#1') to $3.58 \mu\text{mol g}^{-1}$ (DCW) h^{-1} (F#15) for JAJ. So, a biodesulfurization increase of about 2-fold was attained when the minimized medium was used instead the more expensive SFM medium. The difference observed in desulfurization rates between the SFM medium (F#1, F#1') and the SFMM medium (F#14, F#15) was statistically validated by a single factor ANOVA test, with a level of $p < 0.05$.

In addition, also based on Table 3 results, comparing the results by C-source (*i.e.* F#1 vs. F#1'; F#14 vs. F#15), it can be observed that for both culture media (SFM and SFMM) the growth rates (μ_{Max}) were lower when JAJ was used as C-source. This behavior can be due to the presence of an inhibitory compound, which can be formed during the JAJ acidic hydrolysis with hydrochloric acid, since the same behavior is not observed if enzymatic hydrolysis is used instead;^{23,26} and/or due to the use of the nutrients available on JAJ (a complex C-source) not only to cell growth, but also towards secondary metabolism, such as pigment production.^{19,27} But, despite of the slower growth rates in JAJ, only slight differences were observed in the maximum OD values attained when compared with those for Fru + Glu, and moreover the net biomass produced by *G. alkanivorans* strain 1B was even higher in JAJ, and this fact influences the respective specific 2-HBP production rate ($q_{2\text{-HBP}}$) values, which are lower than for Fru + Glu (Table 3: F#1 vs. F#1'; F#14 vs. F#15). Indeed, the maximum value of 2-HBP produced by strain 1B in the four formulations (F#1, F#1', F#14 and F#15) was quite similar ($169.49\text{--}173.42 \mu\text{M}$), as well as the overall 2-

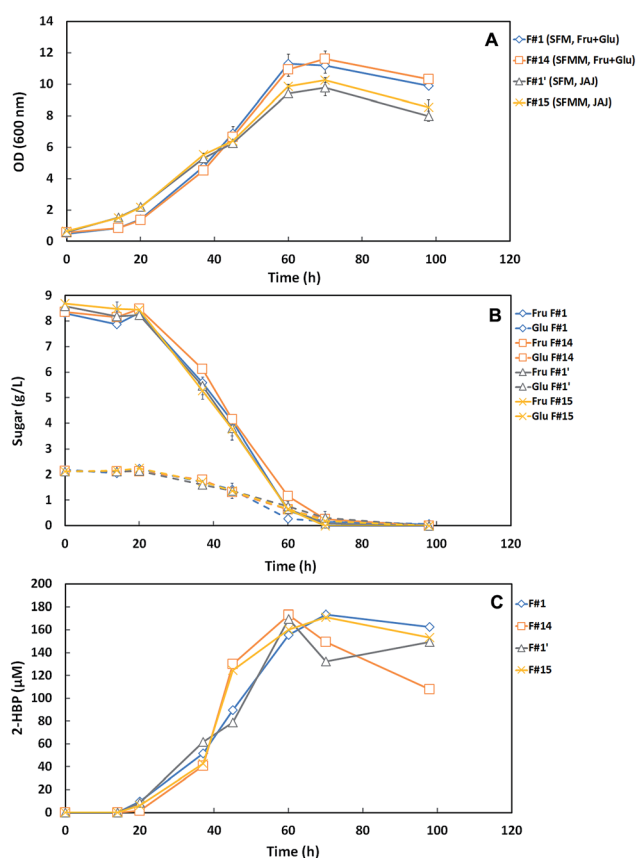


Fig. 4 Time course profiles for cellular growth (A) sugar consumption (B) and desulfurization curves (2-HBP production) (C) of *G. alkanivorans* strain 1B in the shake-flask cultures with the two SFMM medium formulations containing 10 g L^{-1} total sugars as C-source, Fru + Glu (F#14) or JAJ (F#15), and their respective controls with the default medium and the same C-sources (F#1, F#1'). These four formulations contained $\sim 250 \mu\text{M}$ as sulfur source. Fru: fructose; Glu: glucose. Standard deviation ($n = 2$) is indicated by the error bars.



Table 3 Main metabolic parameters for the DBT desulfurization by *G. alkanivorans* strain 1B in the minimized medium formulations with different C-source versus their respective controls: F#1 – SFM, Fru + Glu vs. F#14 – SFMM, Fru + Glu; and F#1' – SFM, JAJ vs. F#15 – SFMM, JAJ

Medium formulations	Fru + Glu		JAJ	
	F#1 ^b – SFM	F#14 – SFMM	F#1' – SFM	F#15 – SFMM
Max ^a OD _{600nm}	11.32	11.61	9.8	10.27
Max biomass (g L ⁻¹)	4.17	4.18	4.68	4.94
μ _{Max} (h ⁻¹)	0.072	0.071	0.054	0.056
Max <i>q</i> _{2-HBP} (μmol g ⁻¹ (DCW) h ⁻¹)	2.15	4.39	1.91	3.58
Max 2-HBP produced (μM)	173.42	173.21	169.49	171.17
2-HBP produced/C-source consumed (μmol g ⁻¹ sugar)	17.34	17.32	16.95	17.12

^a Max – maximum. ^b This was a new F#1 control that was repeated in simultaneously with the assays for F#14; F#1' and F#15.

HBP produced per g of consumed sugar (16.95–17.34 μmol g⁻¹), but the 2-HBP was produced faster in SFMM with Fru + Glu (60 h) due to the higher net production rate. In SFMM with JAJ, the maximum 2-HBP only was attained after 70 h, ~16.7% more time (Fig. 4C).

In overall, these results point out for the benefit of using a cheaper culture medium that enhances the bioprocess effectiveness. In fact, per each liter of the miniaturized BDS culture medium (SFMM medium), there is a net saving of about 15% N-source; 75% Mg-source and 75% TES in comparison with these nutrients in SFM medium. Based on data from Alves *et al.*³ and considering a process scale-up scenario for the production of 200 000 L of biocatalysts/bioreactor, a saving in expenses of about 700€ in N-source, 200€ in Mg-source and 75€ TES (metals solution) is attained if SFMM medium is used instead SFM medium.

When working on a larger scale, besides the costs associated strictly with the production of microbial biocatalysts, it is also necessary to account for possible left overs, which can represent a burden to environment (*e.g.* nutrient-rich wastewaters; biomass wastes). Using this SFMM medium and foreseeing a full C-source (sugars) consumption, most of the salts present in the water after recovering the biocatalyst should be at residual concentrations, except for the phosphate buffer, which is not consumed by the bacteria, and would thus require further removal. So, the production of less contaminated wastewaters can translate into a reduction of treatment costs, increasing the overall efficiency of the industrial process.

Moreover, if a suitable cheaper alternative C-source is also addressed this may turn the overall bioprocess more sustainable and cost-effective, since the C-source is usually one of the most important and expensive nutrients into the culture medium.¹⁶ Several alternative C-sources were already tested for BDS studies with strain 1B, but JAJ was the one with the most promising results,^{9–12,17} and for that reason JAJ (>80% fructose) should be better exploited as the optimal alternative C-source for the fructophilic bacterium, specially using an SSF approach.²⁶

In addition to the cost reduction within the biocatalysts production step, high added-value byproducts, such as the carotenoids from strain 1B,^{19,27} may be also exploited for their

potential benefits to turn BDS process economically viable.^{16,27–29} Indeed, if successfully implemented, the eco-friendly BDS technology could result in less environmental issues than the current industry's solution (HDS) to accomplish the stringent European sulfur limits for transportation fuels (<10 ppm, ultra-low sulfur program). BDS option involves a CO₂ emission reduction of 70 to 80%, smaller residue production and a lower net energy consumption, which in turn is translated in lower operational cost a in reduced capital cost, about two thirds of that for HDS.^{3,13,30}

IV. Conclusions

A microbial approach for petroleum desulfurization could be beneficial for a deep desulfurization in which the classical hydroprocessing methods are costly and non-selective. In this context, the utilization of desulfurizing microorganisms that can grow in low nutrient culture media without vitamins and other growth promoters (*e.g.* yeast extract, peptone, *etc*) is an advantage for BDS upgrade since it may reduce the biocatalyst production costs significantly. Herein, a minimized culture medium (SFMM medium), accounting a C-source of 10 g L⁻¹ (total sugars), was proposed for a cost-effective BDS by *G. alkanivorans* strain 1B. This SFMM medium, which allowed an improved desulfurization (2-fold higher *q*_{2-HBP}), contains less 25% of N-source; 75% of Mg-source and 75% of TES than the original default medium (SFM medium), which implies net savings of about 975€ if a process scale-up scenario for the production of 200 000 L of biocatalysts/bioreactor is considered. Significant additional savings may be still achieved within biocatalysts production process if cheaper alternative C-sources (*e.g.* JAJ) are further exploited.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was financed by FEDER funds through POFC-COMPETE and by National funds through FCT (Fundação para a Ciência e a Tecnologia) in the scope of the projects



Carbon4Desulf (FCOMP-01-0124-FEDER-013932) and Green-Fuel (PTDC/EAM-AMB/30975/2017). Tiago P. Silva also acknowledges to FCT for his Ph.D. financial support (SFRH/BD/104977/2014).

References

- 1 OPEC, *World Oil Outlook 2014*, OPEC Secretariat, Vienna, Austria, 2014.
- 2 OPEC, *World Oil Outlook 2017*, OPEC Secretariat, Vienna, Austria, 2017.
- 3 L. Alves, S. M. Paixão, R. Pacheco, A. F. Ferreira and C. M. Silva, *RSC Adv.*, 2015, **5**, 34047–34057.
- 4 M. Soleimani, A. Bassi and A. Margaritis, *Biotechnol. Adv.*, 2007, **25**, 570–596.
- 5 G. Mohebbi and A. S. Ball, *Microbiology*, 2008, **154**, 2169–2183.
- 6 L. Alves, R. Salgueiro, C. Rodrigues, E. Mesquita, J. Matos and F. M. Gírio, *Appl. Biochem. Biotechnol.*, 2005, **120**, 199–208.
- 7 L. Alves, M. Melo, D. Mendonça, F. Simões, J. Matos, R. Tenreiro and F. M. Gírio, *Enzyme Microb. Technol.*, 2007, **40**, 1598–1603.
- 8 L. Alves and S. M. Paixão, *Bioresour. Technol.*, 2011, **102**, 9162–9166.
- 9 L. Alves and S. M. Paixão, *New Biotechnol.*, 2014, **31**, 73–79.
- 10 L. Alves and S. M. Paixão, *Appl. Biochem. Biotechnol.*, 2014, **172**, 3297–3305.
- 11 T. P. Silva, S. M. Paixão, A. V. Teixeira, J. C. Roseiro and L. Alves, *J. Chem. Technol. Biotechnol.*, 2013, **88**, 919–923.
- 12 T. P. Silva, S. M. Paixão, J. C. Roseiro and L. Alves, *J. Appl. Microbiol.*, 2015, **118**, 609–618.
- 13 R. Vazquez-Duhalt, E. Torres, B. Valderrama and S. Le Borgne, *Energy Fuels*, 2002, **16**, 1239–1250.
- 14 J. J. Kilbane, *Curr. Opin. Biotechnol.*, 2006, **17**, 305–314.
- 15 M. A. Dinamarca, A. Rojas, P. Baeza, G. Espinoza, C. Ibacache-Quiroga and J. Ojeda, *Fuel*, 2014, **116**, 237–241.
- 16 S. M. Paixão, T. P. Silva, B. F. Arez and L. Alves, in *Applying Nanotechnology to the Desulfurization Process in Petroleum Engineering*, ed. T. Saleh, IGI Global, Hershey PA, 2016, vol. 13, pp. 390–425.
- 17 L. Alves, S. Marques, J. Matos, R. Tenreiro and F. M. Gírio, *Chemosphere*, 2008, **70**, 967–973.
- 18 S. M. Paixão, L. Alves, R. Pacheco and C. M. Silva, *Energy*, 2018, **150**, 468–481.
- 19 A. S. Fernandes, S. M. Paixão, T. P. Silva, J. C. Roseiro and L. Alves, *Bioprocess Biosyst. Eng.*, 2018, **41**, 143–155.
- 20 R. M. Singh, *Appl. Microbiol.*, 1971, **22**, 131–132.
- 21 J. P. Schimel and M. N. Weintraub, *Soil Biol. Biochem.*, 2003, **35**, 549–563.
- 22 M. Webb, *J. Gen. Microbiol.*, 1949, **5**, 485–495.
- 23 T. P. Silva, MSc thesis, Faculty of Sciences of the University of Lisbon, 2012.
- 24 T. Ohshiro, T. Kojima, K. Torii, H. Kawasoe and Y. Izumi, *J. Biosci. Bioeng.*, 1999, **88**, 610–616.
- 25 L. Alves, J. Matos, R. Tenreiro and F. M. Gírio, *J. Ind. Microbiol. Biotechnol.*, 2008, **35**, 69–73.
- 26 S. M. Paixão, B. F. Arez, J. C. Roseiro and L. Alves, *J. Environ. Manage.*, 2016, **182**, 397–405.
- 27 T. P. Silva, S. M. Paixão and L. Alves, *RSC Adv.*, 2016, **6**, 58055–58063.
- 28 S. Bandyopadhyay, R. Chowdhury, C. Bhattacharjee and S. Pan, *Fuel*, 2013, **113**, 107–112.
- 29 S. Bandyopadhyay, R. Chowdhury and C. Bhattacharjee, *Green Chem. Lett. Rev.*, 2014, **7**, 288–295.
- 30 J. M. Campos-Martin, M. C. Capel-Sanchez, P. Perez-Presas and J. L. G. Fierro, *J. Chem. Technol. Biotechnol.*, 2010, **85**, 879–890.

