



Cite this: *RSC Adv.*, 2019, 9, 26559

Photolysis of carboxymethylflavin in aqueous and organic solvent: a kinetic study†

Iqbal Ahmad,^a Tania Mirza,^a Syed Ghulam Musharraf,^b Zubair Anwar,^{id}*^a Muhammad Ali Sheraz,^{id}^a Sofia Ahmed,^{id}^a Muhammad Ahsan Ejaz^a and Adeela Khurshid^a

This is the first study on the photolysis of carboxymethylflavin (CMF), an intermediate in the photolysis of riboflavin (RF). CMF is photodegraded by removal of side-chain to lumichrome (LC) in acid solution and to LC and lumiflavin (LF) in alkaline solution. It also undergoes alkaline hydrolysis to 1,2-dihydro-1-methyl-2-keto-3-quinoxaline carboxylic acid (KA) and 1,2,3,4-tetrahydro-1-methyl-2,3-dioxoquinoxaline (DQ) by cleavage of isoalloxazine ring. CMF degrades to LC in organic solvents. The formation of LC in acid solution and organic solvents takes place by second-order reaction and those of LC, LF, KA and DQ in alkaline solution by first-order reactions. The values of second-order rate constants for the photolysis of CMF at pH 2.0 to 7.0 are in the range of 1.13 to 2.45 M⁻¹ s⁻¹ and those of first-order rate constants (*k*_{obs}) at pH 8.0–12.0 from 1.53 to 4.18 × 10⁻⁴ s⁻¹ and for the formation of photoproducts from 0.37 to 16.6 × 10⁻⁵ s⁻¹. The photolysis of CMF is enhanced, with pH, in the alkaline region since the excited state is sensitive to alkaline hydrolysis. The photolysis and fluorescence quantum yields of CMF in aqueous and organic solvents have been reported. CMF and photoproducts have been assayed spectrofluorimetrically. The mode of CMF photolysis is discussed.

Received 14th April 2019

Accepted 24th July 2019

DOI: 10.1039/c9ra02818h

rsc.li/rsc-advances

Introduction

Riboflavin (vitamin B₂) RF (1) (Fig. 1) has long been studied due to its biochemical importance, photosensitivity and pharmaceutical applications.^{1–8} It is photodegraded to several products in aqueous and organic solvents to yield formylmethylflavin (FMF) (2) as a major intermediate in the reaction.^{9–16} FMF undergoes hydrolysis^{17–19} and photolysis to form lumichrome (LC) (3) and lumiflavin (LF) (4) in aqueous solution^{20,21} and LC in organic solvents.²⁰ It is photooxidized to carboxymethylflavin (CMF) (5) in aqueous and organic solvents.^{12,13,16,21,22} CMF has also been suggested to act as an intermediate in the formation of LC and LF during the photolysis of RF in aqueous solution.²³ It is an isoalloxazine derivative like RF, FMF and LF^{1,24,25} and is susceptible to hydrolytic degradation in alkaline solution by the cleavage of the isoalloxazine ring to give 1,2-dihydro-1-methyl-2-keto-3-quinoxaline carboxylic acid (KA) (6) and 1,2,3,4-tetrahydro-1-methyl-2,3-dioxoquinoxaline (DQ) (7).^{26–29} So far no chemical or photodegradation studies of CMF have been reported since its isolation by Fukumachi and Sakurai in 1955.²²

An important consideration in degradation of chemical and pharmaceutical compounds is the effect of solvent on the rates of reaction.^{30–35} Solvent polarity plays an important role in the stabilization of drug formulations to prolong their shelf-lives.^{36–40} The effect of solvent on the photolysis of RF,^{5,16,41–44} FMF,^{20,21,45} LC⁴⁵ and other flavins^{46–48} has been studied. The photo-dynamics aspects of RF have also been investigated.⁴⁹ The photolysis of CMF in organic solvents needs to be investigated to determine the effect of solvent polarity on its degradation.

The present work involves a detailed kinetic study of photolysis of CMF in aqueous and organic solvents, characterize the photoproducts and evaluate the kinetics of its degradation reactions. The rate–pH profiles could provide information about the regions of minimum and maximum degradation of the compound. Attempts are made to correlate the kinetic parameters with solvent polarity and viscosity. The photochemical and fluorescence quantum yields of CMF in aqueous and organic solvents are determined. CMF and its photoproducts are assayed by a recently developed multicomponent spectrofluorimetric method.²⁹ The mode of photodegradation reactions of CMF is discussed.

Experimental section

Materials

LC and LF were procured from Sigma. CMF was prepared by the method of Fukumachi and Sakurai.²² KA and DQ were

^aBaqai Institute of Pharmaceutical Sciences, Baqai Medical University, Super Highway, Gadap Road, Near Toll Plaza, Karachi, Pakistan. E-mail: zubair_ana@hotmail.com

^bHEJ Research Institute of Chemistry, University of Karachi, Karachi-75270, Pakistan

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c9ra02818h



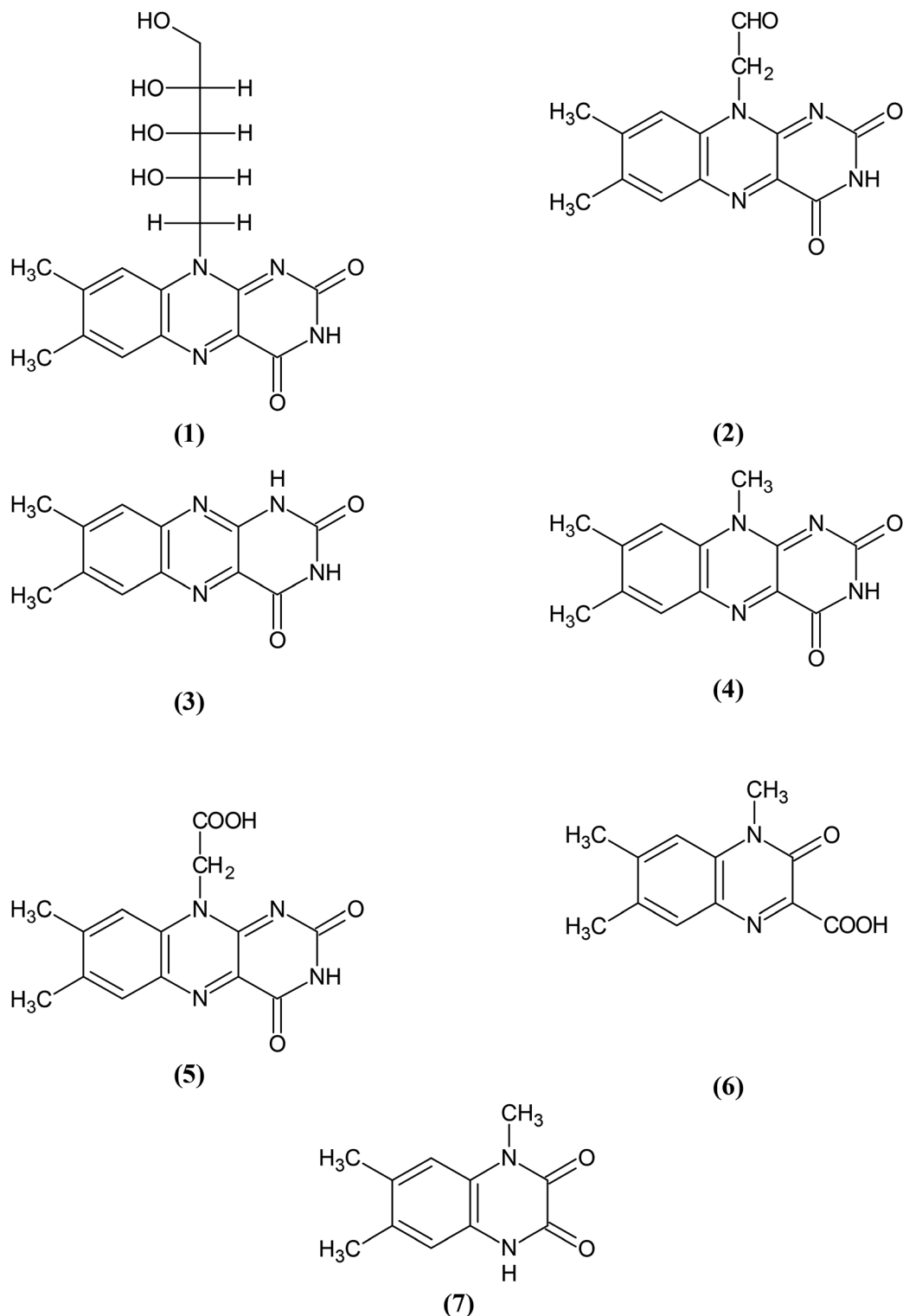


Fig. 1 Chemical structures of riboflavin (RF) and photoproducts.

prepared from RF by the methods of Surrey and Nachod⁵⁰ and Miles *et al.*,⁵¹ respectively. The side-chain of KA and DQ was cleaved by periodic acid⁵¹ oxidation. The compounds were purified by cellulose column chromatography using 1-butanol-1 propanol-acetic acid-water (50 : 30 : 2 : 18, v/v) as

solvent system.¹⁸ All the reagents and solvents of the highest purity were procured from Merck & Co. The buffers used were citric acid- Na_2HPO_4 (pH 2.5–8.0), H_3BO_3 -KCl-NaOH (pH 9.0–10.0) and Na_2HPO_4 -NaOH (pH 11.0–12.0). The ionic strength was 0.001 M in each case.



Precautions

The photolysis of CMF, thin-layer chromatography and fluorimetric assay have been carried out in a dark chamber.

Measurement of pH

All pH measurements were carried out with a digital pH meter (Model CP-501; sensitivity ± 0.01 units, Poland) using a combination pH electrode. The calibration of the electrode was performed using buffer solutions of pH 4.0, 7.0 and 9.0.

Measurement of UV-visible spectra

The spectral measurements were carried out with Thermo-Scientific UV-visible spectrophotometer (Evolution 201) using quartz glass cells of 10 mm path length.

Measurement of fluorescence

The measurement of fluorescence of CMF and photoproducts was performed out at 25 ± 1 °C using Jasco FP-8300 spectrofluorimeter (Japan). The wavelengths used for the determination of CMF and photoproducts are given in Table 1.

The fluorescence intensity was measured in relative units. Pure 0.01 mM solutions (pH 6.5) of these compounds were used as standards.

Light intensity measurement

The intensity of the radiation source, a Philips HPLN 125 W high pressure mercury vapor lamp, was determined by ferrioxalate actinometry⁵² and a value of $1.12 \pm 0.08 \times 10^{17}$ quanta per s was obtained.

Determination of quantum yields of photolysis

The quantum yields (Φ_{PD}) for the photochemical formation of LC in acid solution and organic solvents by second-order reaction and the photodegradation of CMF in alkaline solution by first-order reaction have been determined by the method of Zirak *et al.*^{53,54} and Tyagi and Penzkofer⁵⁵ as follows.

Formation of LC

Reaction description. The reactions involved in the formation of LC by CMF are described by eqn (8)–(11) and eqn (12)–(21) in Scheme 2 (see section on Mode of photolysis).

Experimental procedure. The amount of formed LC and the amount of absorbed excitation light by CMF during a certain

period is determined and the quantum yield of LC is calculated using the equation.

$$\Phi_{PL} = \frac{N_{LC}}{n_{ph\ abs}} \quad (1)$$

where, N_{LC} = number density of generated LC

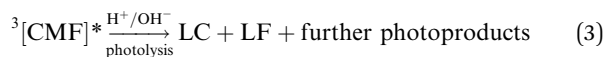
$$n_{ph\ abs} = \frac{I_{exc} t_{exc}}{h\nu_{exc}} \alpha_{exc} = \frac{I_{exc} t_{exc}}{h\nu_{exc}} \sigma_{exc}^1 [CMF] \quad (2)$$

$n_{ph\ abs}$ [cm^{-3}] = number density of CMF absorbed photons per 1 cm of sample length, I_{exc} [W cm^{-2}] = excitation light intensity in watt per cm^2 , t_{exc} [s] = time duration of light exposure in seconds, h [J s] = Planck constant, ν_{exc} [s^{-1}] = excitation light frequency, $h\nu_{exc}$ [J] energy of an excitation photon, α_{exc} [cm^{-1}] = absorption coefficient of CMF at excitation wavelength, σ_{exc} = absorption cross-section of CMF at excitation wavelength.

HPLC-Tof-MS system. Samples were recorded on HPLC-Tof-MS system (Agilent 6230). RP-C8 column was used for separation (1.0×50 mm, 3.5 μm , Agilent ZORBAX Eclipse XDB columns – C18 column). The mass spectrometer with ESI source was set in the positive ion mode. An isocratic mobile phase that is 0.1% formic acid in both water (eluent A) and methanol (eluent B) was used at a flow rate of 0.5 ml min^{-1} for 20 min. Injection volume was 2 μL .

Photodegradation of CMF

Reaction description. The reactions involved in the photolysis of CMF are described by eqn (12)–(16) in Scheme 2 (see section on Mode of photolysis), followed by.



Experimental procedure. The amount of photodegraded CMF and the amount of absorbed excitation light by CMF during a certain period is determined. The quantum yield of CMF is calculated using the equation.

$$\Phi_{PL} = \frac{N_{CMF}}{n_{ph\ abs}} \quad (4)$$

where, N_{CMF} = number density of degraded CMF, $n_{ph\ abs}$ = as given in eqn (23) in Scheme 2

Fluorescence quantum yield. The fluorescence quantum yield (Φ_F) is the ratio of photons emitted through fluorescence to photons absorbed.

$$\Phi_F = \frac{\text{number of photons emitted through fluorescence}}{\text{number of photons absorbed}}$$

It can be determined of a comparative method⁵⁶ using standard samples of a compound whose Φ_F is known. The standard and the test samples having identical absorbance at the same excitation wavelength are assumed to absorb same number of photons. Therefore, the ratio of integrated fluorescence intensities of the two solutes determined under identical conditions gives the ratio of values of the quantum yields. Since

Table 1 Excitation and emission wavelengths for the determination of CMF and photoproducts

Compound	Excitation wavelength (λ_{ex}) nm	Emission wavelength (λ_{em}) nm
CMF	445	530
LC	356	478
LF	445	530
KA	364	443
DQ	332	420



Φ_F of the standard is known, the Φ_F of test sample can be calculated.

The method involves the measurement of absorbance and the integrated fluorescence intensity (*i.e.* area of the fluorescence spectra) of the sample of the standard and the test samples. The integrated fluorescence intensity of the standard and the test samples are plotted against the absorbance of each sample. The gradients of the two plots are proportional to the quantum yields of the two compounds. The fluorescence quantum yield of the unknown is calculated from the product of the quantum yield of the standard and the quotient of the two gradients according to the following equation.

$$\begin{aligned}\Phi_F &= \Phi_s \left(\frac{A_s}{A_u} \right) \left(\frac{E_u}{E_s} \right) \left(\frac{\eta_u}{\eta_s} \right)^2 \\ &= \Phi_s \left(\frac{m_u}{m_s} \right) \left(\frac{\eta_u}{\eta_s} \right)^2\end{aligned}$$

where, Φ_s = fluorescence quantum yield of the standard, m = gradient of the plot of integrated fluorescence intensity against absorbance, η = refractive index of the solvent, A = absorbance of the solution at the fluorescence excitation wavelength (445 nm), E = integrated fluorescence intensity, subscripts “s” and “u” refer to the standard and unknown samples, respectively, RF with a Φ_F value of 0.24 (ref. 57) has been used as a standard in this case.

Thin-layer chromatography (TLC)

CMF and its side-chain cleavage products (LC and LF) were detected using 250 μm cellulose plates (Whatman CC41) and the solvent systems: (a) 1-butanol–1-propanol–acetic acid–water (50 : 30 : 2 : 18, v/v) and (b) 1-butanol–acetic acid–water (40 : 10 : 50, v/v, organic phase).¹²

The isoalloxazine ring cleavage products of CMF, *i.e.* KA and DQ, were detected using 250 μm silica gel G plates and the solvent system (c) chloroform–acetic acid–pyridine (50 : 10 : 10, v/v).²⁷ The spots of all these products were located by their characteristic fluorescence on excitation at 366 nm.

Photodegradation of CMF

A CMF solution (5×10^{-5} M) was prepared at pH 2.0–12.0, using appropriate buffer, in a 100 ml volumetric flask and placed in a thermostat bath maintained at 25 ± 1 °C in the radiation chamber. It was irradiated with the Philips HPLN 125 W high pressure mercury vapor lamp (emission at 405 and 435 nm, the later wavelength corresponding to the absorption maximum of CMF at 445 nm),^{19,22,29} fixed horizontally at a distance of 25 cm from the center of the flask. The same procedure was used for photolysis of CMF in organic solvents. Samples of degraded solutions were subjected to TLC and assay at appropriate intervals.

Assay of CMF and photoproducts

Aqueous solution. The assay of CMF and photoproducts has been carried out by a recently developed and validated spectrofluorimetric method.²⁹ Since the photoproducts of CMF are

the same as those of its hydrolytic degradation (LC, LF, KA, DQ), the method can be conveniently used for determination of these compounds. It involves extraction of the photodegraded solutions of CMF with chloroform at pH 2.0 to remove side-chain products, LC and LF, and their assay at 478 and 530 nm, respectively. This is followed by adjustment of the pH of aqueous phase to 6.5 and assay of CMF and the isoalloxazine ring cleavage products, KA and DQ, at 530, 443 and 420 nm, respectively.

Organic solvents. A 1 ml aliquot of degraded solution of CMF was evaporated to dryness under reduced pressure and the residue dissolved in 10 ml of pH 6.5 citro-phosphate buffer. The solution was used for fluorimetric assay of CMF and LC at 530 and 478 nm.

Results and discussion

Absorption characteristics of CMF

The UV-visible and IR spectra of CMF are shown in Fig. 2a and b, respectively. CMF exhibits absorption maxima at 223 nm (molar decadic extinction coefficient, $\epsilon = 24\,700\text{ M}^{-1}\text{ cm}^{-1}$), 266 nm ($\epsilon = 27\,800\text{ M}^{-1}\text{ cm}^{-1}$), 376 nm ($\epsilon = 9000\text{ M}^{-1}\text{ cm}^{-1}$) and 445 nm ($\epsilon = 10\,200\text{ M}^{-1}\text{ cm}^{-1}$) at pH 7.0 (Fig. 2a). These values are similar to those of RF and FMF having the same nucleus and different side-chains.⁵⁸ The FTIR spectrum of CMF shows peaks at 3250 cm^{-1} (NH), 1650 cm^{-1} (C=O), 1580 cm^{-1}

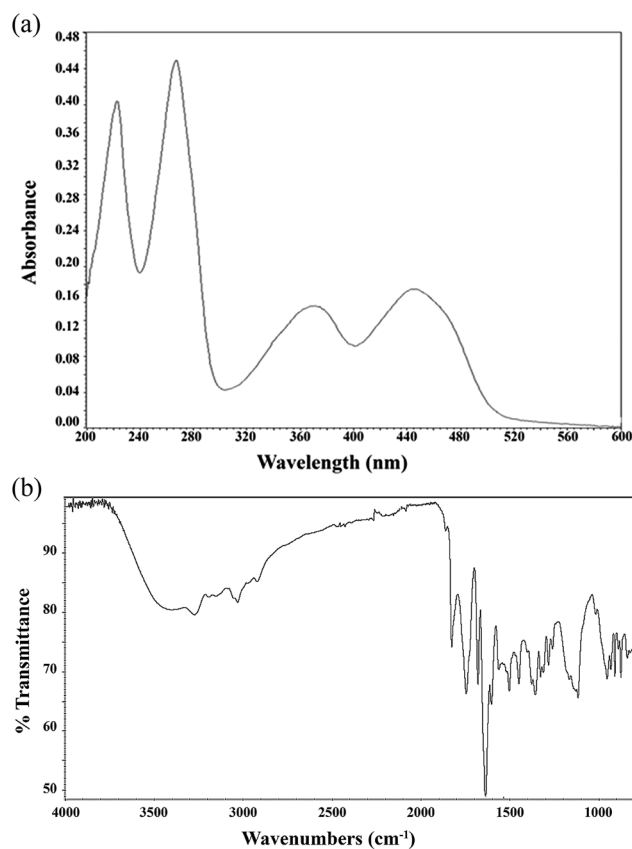


Fig. 2 (a) UV and visible absorption spectrum of CMF at pH 7.0. (b) FTIR spectrum of CMF.



(C=C) and 1540 cm^{-1} (C=N) (Fig. 2b). The broad absorption band in the $3300\text{--}2800\text{ cm}^{-1}$ region indicates hydrogen bonded O–H stretching of the carboxylic group. This is further supported by the peaks at 1700 cm^{-1} and 1300 cm^{-1} .

Nature of photoproducts

CMF was originally reported as a photoproduct in the photolysis of RF in alkaline solution in the presence of H_2O_2 , along with LC and LF.^{22,23} It has also been detected in the photolysis of RF^{12,13,51} and FMF and may be formed directly from RF²³ or by the oxidation of FMF.²¹ LC and LF are formed on the photolysis^{9,11,20,21} or hydrolysis of FMF^{17–19} and CMF²⁹ in the alkaline solution.

It has been found that the formation of LC and LF takes place on the photolysis of CMF. Thus, a second route of the formation of LC and LF, in addition to that of FMF, in the photolysis of RF is through CMF. The other two products formed by isalloxazine ring cleavage of CMF in alkaline solution are quinoxaline derivatives, KA and DQ. The formation of both of these products is enhanced with an increase in pH due to the hydrolysis of the isalloxazine ring,²⁹ as has previously been observed in the case of the hydrolysis of 9-methylisalloxazine.^{27,28} The only product detected on the photolysis of CMF in organic solvents is LC. All these products have been identified on comparison of their characteristics fluorescence under UV light (CMF and LF, yellow green; LC, sky blue; KA and DQ, blue) and retention (R_f) values with those of the reference compounds using the solvent systems reported in the experimental section. The identification of photoproducts of CMF was necessary to confirm their presence in degraded solutions prior to the application of the spectrofluorimetric method for their assay during the photolysis reactions.

Product composition

The photolysis of CMF leads to the formation of LC, LF, KA and DQ in aqueous solution as described in the above section. The composition of these products at 50% degradation of CMF at pH 8.0–12.0 is given in Table 2. The formation of LC in acid solution amounts to 50% and in the alkaline solution from 20 to 35% while the formation of LF in alkaline solution takes place to the extent of 8 to 13% of the degraded CMF in the pH range studied. The ring cleavage products, KA and DQ, occur to the extent of 5 to

10% and 2 to 6%, respectively. These results indicate that the formation of these photoproducts is gradually increased with pH due to the hydrolytic degradation of CMF which is accelerated in the presence of light. The decrease in the concentration of LC, with pH, is due to its formation by an acid-catalyzed reaction.¹⁷

Spectral characteristics of photolyzed solutions

CMF has been found to undergo spectral changes during photolysis in acid and alkaline solutions (Fig. 3). These changes have been found to vary with pH and the loss of the molecule on degradation. A typical set of absorption spectra of CMF on degradation at pH 4.0 is presented in Fig. 3a. It indicates the loss of peak at 445 nm and a shift towards 356 nm with concomitant increase in absorbance showing the formation of LC in acid solution. Another set of absorption spectra of CMF during photolysis at pH 11.0 is shown in Fig. 3b. The loss of absorbance at 445 nm indicates the degradation of CMF and a prominent shift of the 374 nm peak towards 356 nm showing the formation of LC. The absorption spectrum of LF is similar to that of CMF and, therefore, it cannot be distinguished from CMF to indicate any spectral change. The disappearance of 445 nm peak indicates the cleavage of the isalloxazine ring to form KA and DQ absorbing in the 300–400 nm region.^{27,28,59} Thus, the spectral changes on the photolysis of CMF at pH 11.0 indicate that the molecule is being transformed into its side-chain and ring cleavage products.

Assay of CMF and photoproducts

The photoproducts of CMF in acid (LC) and alkaline solution (LC, LF, KA and DQ) are same as those of the hydrolytic degradation reaction. However, the formation of LC and LF is increases in presence of light. Therefore, a single method can be applied to the determination of the products of hydrolytic and photolytic degradation. Recently, a spectrofluorimetric method has been developed and validated for the determination of CMF and its side-chain cleavage (LC and LF) and ring cleavage products (KA and DQ).²⁹ The accuracy of the method has been reported to be within 1%.²⁹ This is used for the determination of CMF and photoproducts during photolysis. The results of the assay of these compounds on the photolysis of CMF for a typical reaction (pH 10) are given in Table 3. These indicate an almost constant molar balance at various intervals showing the accuracy of the method. The assay data for the reactions carried out at pH 2.0–12.0 have been used to evaluate the kinetics of degradation of CMF.

Kinetics of photolysis of CMF

CMF undergoes photolysis to form LC in acid solution and LC, LF, KA and DQ in alkaline solution (Scheme 1).

The kinetics of these reactions has not so far been studied. A recent study²⁹ has shown that CMF is degraded by hydrolysis to form the above mentioned compounds in aqueous solution. Considering the photolysis of CMF in acid solution to form LC by a second-order reaction (k_1) and to form LC, LF, KA and DQ

Table 2 Product composition at 50% photolysis of 5.0×10^{-5} M CMF solutions at pH 8.0–12.0

pH	Time (min)	LC (%)	LF (%)	KA (%)	DQ (%)
8	75.3	34.8	8.4	4.8	2.0
9	36.5	30.8	10.4	6.0	3.0
10	30.8	26.2	11.0	7.8	4.8
11	28.7	23.2	12.4	9.2	6.0
12	27.6	20.2	13.0	10.4	6.4



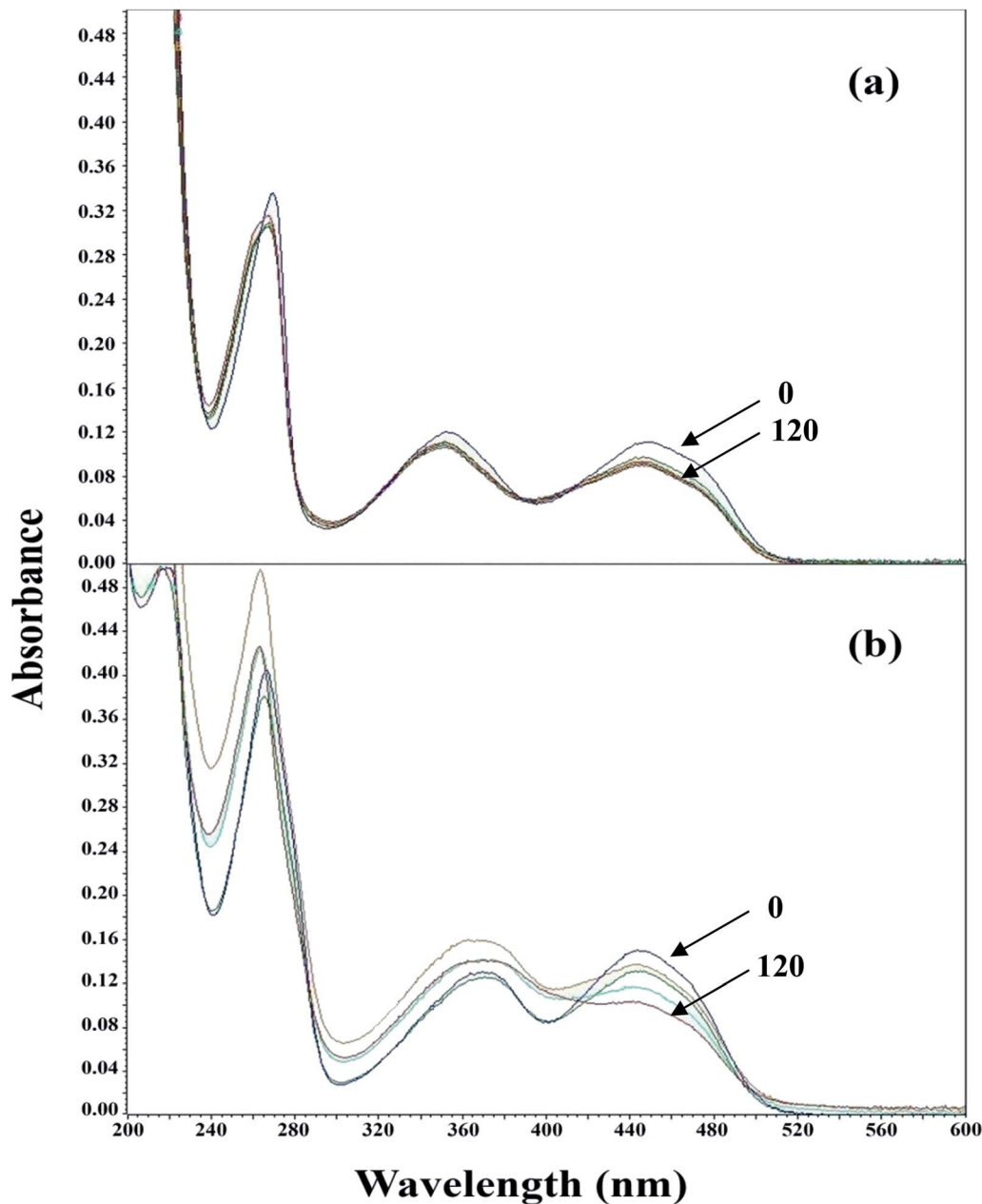


Fig. 3 Absorption spectra of CMF on photolysis at pH 4.0 (a), and at pH 11.0 (b). Times indicated are in min.

in alkaline solution by parallel first-order reactions, the rate constants, k_2 , k_3 , k_4 and k_5 , can be calculated by the method of Frost and Pearson⁶⁰ an applied previously to the photolysis of RF.⁶¹

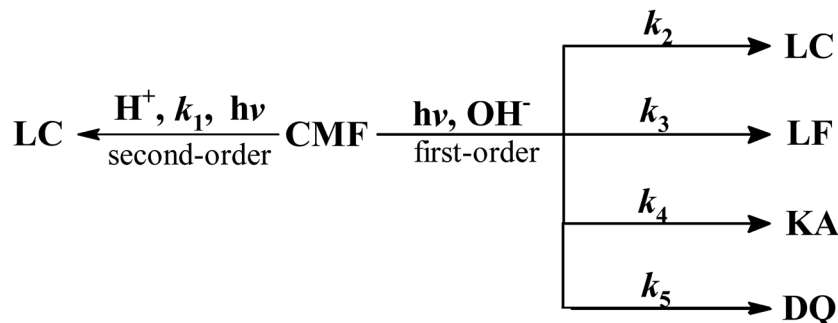
Photolysis of CMF in acid solution

$$\frac{-d[\text{CMF}]}{dt} = k_1[\text{CMF}]^2 \quad (5)$$

Table 3 Photolysis of CMF at pH 10.0. Concentrations of CMF and photoproducts

Time (min)	CMF ($\text{M} \times 10^5$)	LC ($\text{M} \times 10^5$)	LF ($\text{M} \times 10^5$)	KA ($\text{M} \times 10^5$)	DQ ($\text{M} \times 10^5$)	Total ($\text{M} \times 10^5$)
0	5.00	—	—	—	—	5.00
15	2.38	0.91	0.85	0.46	0.42	5.03
30	1.22	1.31	1.15	0.85	0.51	5.05
45	0.59	1.43	1.28	1.10	0.56	4.98
60	0.31	1.52	1.39	1.18	0.62	5.02





Scheme 1 Photolysis of CMF in acid and alkaline solution.

The rate constants, k_1 , for second-order photolysis reactions of CMF in acid solution determined from slopes of inverse concentration ($1/c$) against time plots are given in Table 4.

Photolysis of CMF in alkaline solution. If A , U , V , W and X are the corresponding concentrations of CMF, LC, LF, KA and DQ during degradation and A_0 is initial concentration of CMF, the overall rate constant (k_{obs}) can be written as:

$$\frac{-dA}{dt} = k_2A + k_3A + k_4A + k_5A = (k_2 + k_3 + k_4 + k_5)A = k_{\text{obs}}A$$

Therefore

$$k_{\text{obs}} = k_2 + k_3 + k_4 + k_5 \quad (6)$$

and

$$\ln \frac{(A_0)}{A} = k_{\text{obs}}t$$

or

$$A = A_0 e^{-kt} \quad (7)$$

Considering the degradation of A as a first-order reaction

$$\frac{dU}{dt} = k_2A = k_2A_0 e^{-kt}$$

and

$$U = \frac{k_2A_0}{k_{\text{obs}}} e^{-kt} + \text{constant}$$

Table 4 Second-order rate constants (k_1) and half-lives for the photolysis of CMF in acid solution

pH	k_1 ($\text{M}^{-1} \text{s}^{-1}$) \pm SD	$t_{1/2}$ (min)	ϕ^a
2.0	1.13 ± 0.01	295.0	0.16
3.0	4.20 ± 0.05	79.3	0.20
3.5	8.05 ± 0.09	41.4	0.23
4.0	10.85 ± 0.11	30.7	0.29
4.5	6.04 ± 0.07	55.2	0.06
5.0	2.75 ± 0.03	121.2	0.05
6.0	1.99 ± 0.02	167.5	0.05
7.0	2.45 ± 0.02	136.1	0.12

^a It represents the quantum yields for the formation of LC, the major side-chain photoproduct.

or

$$\begin{aligned} U &= U_0 + (k_2A_0/k_{\text{obs}})(1 - e^{-kt}) \\ V &= V_0 + (k_3A_0/k_{\text{obs}})(1 - e^{-kt}) \\ W &= W_0 + (k_4A_0/k_{\text{obs}})(1 - e^{-kt}) \\ X &= X_0 + (k_5A_0/k_{\text{obs}})(1 - e^{-kt}) \end{aligned} \quad (8)$$

If $U_0 = V_0 = W_0 = X_0 = 0$, the equations can be expressed as

$$V/U = k_3/k_2$$

$$W/U = k_4/k_2$$

$$X/U = k_5/k_2$$

or

$$U : V : W : X = k_2 : k_3 : k_4 : k_5 \quad (9)$$

The concentrations of the photoproducts are in a constant ratio to each other. These are independent of time and the initial concentration of CMF and can be used to determine the rate constants for the individual reactions. The values of k_{obs} for disappearance of CMF and k_2 , k_3 , k_4 and k_5 for formation of LC, LF, KA and DQ, respectively, at pH 8–12 are reported in Table 5. These rate constants vary with pH as discussed in a later section. The sum of the individual rate constants for the formation of the photoproducts is almost equal to the values of the k_{obs} for the photolysis of CMF at a particular pH (Table 5).

Photolysis of CMF in organic solvents. The photolysis of CMF in organic solvents has been found to follow second-order kinetics to form LC as observed in the case of FMF.²⁰



It is similar to that of CMF in acid solution as described above. However, the rates of the reaction are lower than those observed in the acid solution. This is due to a change in the polarity of the medium. The second-order rate constants, k_6 , for these reactions are reported in Table 6. The values increase on increasing the polarity of solvent.



Table 5 Apparent first-order rate constants (k_{obs}) for the photolysis of CMF at pH 8.0–12.0, and the rate constants for the formation of lumichrome (LC) (k_2), lumiflavin (LF) (k_3), keto acid (KA) (k_4) and dioxo-quinoxaline (DQ) (k_5), second-order rate constants (k') for OH^- ion catalyzed reaction and quantum yields of photolysis (Φ^a)

pH	$k_{\text{obs}} \times 10^4 \text{ (s}^{-1}\text{)} \pm \text{SD}$	$t_{1/2} \text{ (min)}$	$k_2 \times 10^4 \text{ (s}^{-1}\text{)}$	$k_3 \times 10^4 \text{ (s}^{-1}\text{)}$	$k_4 \times 10^4 \text{ (s}^{-1}\text{)}$	$k_5 \times 10^4 \text{ (s}^{-1}\text{)}$	$k' \text{ (M}^{-1} \text{ s}^{-1}\text{)}$	Φ^a
8	1.53 ± 0.06	75.3	0.76	0.36	0.23	0.18	153.0	0.32
9	3.07 ± 0.12	36.5	1.08	0.91	0.66	0.42	31.7	0.34
10	3.75 ± 0.15	30.8	1.47	1.10	0.70	0.48	3.75	0.36
11	4.01 ± 0.14	28.7	1.63	1.13	0.75	0.50	0.40	0.39
12	4.15 ± 0.16	27.6	1.67	1.15	0.80	0.53	0.04	0.41

^a It represents the overall quantum yields for the photolysis of CMF.

pH effect

CMF is sensitive to pH in the presence or absence of light and undergoes degradation to give LC in acid solution and LC, LF, KA and DQ in alkaline solution. The kinetic results (Table 2) indicate that CMF is more labile to side-chain cleavage than the ring cleavage. The k -pH profiles for photolysis of CMF in acid and alkaline regions have been determined. A plot of k_1 for the photolysis of CMF in acid region *versus* pH is shown in Fig. 4. The rate of the reaction is slow in the pH range 2 to 3 due to protonation of the molecule (N-10) as observed in the case of RF ($\text{p}K_{\text{a}} 1.7$)^{13,62} and is then increased up to pH 4. Above this value a rapid decrease in the rate up to pH 6 is observed. This is similar to that of the photolysis of FMF ($\text{p}K_{\text{a}} 3.5$)⁶³ which also have the highest rate around pH 4 in the acid region.²¹ The decrease in the rate above pH 4 can be explained on the basis of redox potentials of flavins which have lowest values in the pH range of 5 to 6 (*e.g.* RF -0.117 V at pH 5.0, -0.208 V at pH 7.0).³⁹ This would slow down the photoreduction of CMF and lead to a decrease in rate in this region. The values of k_1 for the photolysis of CMF in acid solution are in the range of 1.13 to $2.45 \text{ M}^{-1} \text{ s}^{-1}$ at pH 2 to 7 (Table 4). Flavins are known to degrade by photoreduction process.^{4,24,25}

The k -pH profile for photolysis of CMF in alkaline region is shown in Fig. 5. It represents a steep curve in the pH range 8–10 followed by a decline in rate upto pH 12 as a result of hydrolytic degradation. The lowering of rate above pH 10 appears to be due to the ionization of the $\text{N}_3\text{-H}$ group ($\text{p}K_{\text{a}} 10.2$)⁶² of the isoalloxazine nucleus. This is in accordance with the photolysis behavior of RF which is also less susceptible to degradation in the pH range 10.0–12.0.¹³ The increase in the rate in alkaline region is probably due to the fact that the flavin triplet state exists in a bent, diradical form which is more sensitive to

alkaline hydrolysis.¹¹ At pH 8 and above CMF undergoes isoalloxazine ring cleavage to form KA and DQ and, therefore, the values of k_{obs} are further increased with pH in this region as observed in the case of FMF¹⁹ and 9-methylisoalloxazine.^{27,28} All the products of alkaline photolysis of CMF (LC, LF, KA, DQ) are formed by parallel first-order kinetics and the rates of these reactions are in the range of 0.18 – $1.67 \times 10^{-4} \text{ s}^{-1}$ at pH 8.0–12.0 (Table 5). The rate-pH profiles for formation of LC, LF, KA and DQ in the alkaline region are similar to that of the degradation of CMF (Fig. 6). However, the individual rates of formation of these products are lower than that of the overall loss of CMF. The ratio of the rate constants for the formation of side-chain cleavage products, LC and LF (k_2 and k_3) and those of the formation of ring cleavage products, KA and DQ (k_4 and k_5) on the photolysis of CMF at pH 12 is 6.8 : 3.2.

Hydroxyl ion catalysis

In alkaline solution the photolysis of CMF is catalyzed by OH^- ions. In order to evaluate the effect of OH^- ions on the rate of photolysis of CMF, the $\log k_{\text{obs}}$ values were plotted against pH which showed a biphasic linear relation (Fig. 7). The values of second-order rate constants (k') for the OH^- ion catalyzed reactions at various pH values are reported in Table 5. The slight deviation in rate constants at 10.5 to 12.0 is due to the involvement of the $\text{N}_3\text{-anion}$ of CMF with a $\text{p}K_{\text{a}}$ value similar to that of RF (10.2)⁶² and being less sensitive to photolysis.

Effect of solvent

Dielectric constant. The photolysis of CMF in organic solvents follows second-order kinetics and the values of the rate constants

Table 6 Second-order rate constants (k_6) for the photolysis of CMF in organic solvents and water

Solvent	Acceptor number	Dielectric constant (ϵ)	Inverse viscosity (mPa s^{-1}) (25 °C)	$k_6 \text{ (M}^{-1} \text{ s}^{-1}\text{)}$
Ethyl acetate	17.1	6.02	2.268	0.98
1-Butanol	36.8	17.8	0.387	1.15
1-Propanol	37.3	20.1	0.514	1.20
Acetone	12.5	20.7	0.330	1.23
Ethanol	37.1	24.3	0.931	1.26
Methanol	41.3	32.6	1.828	1.42
Acetonitrile	18.9	38.5	2.898	1.56
Water	54.8	78.5	1.123	2.45



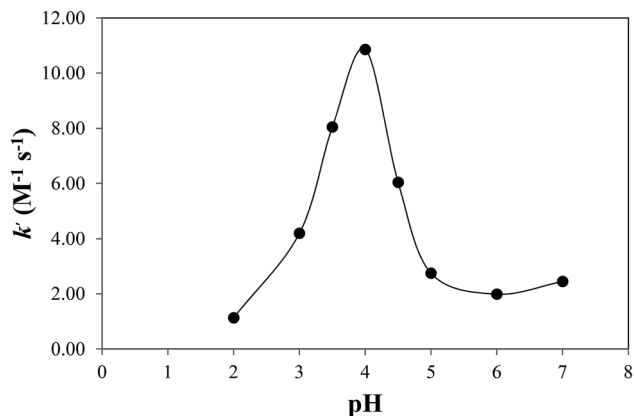


Fig. 4 k_1 -pH profile for photolysis of CMF at pH 2.0–7.0.

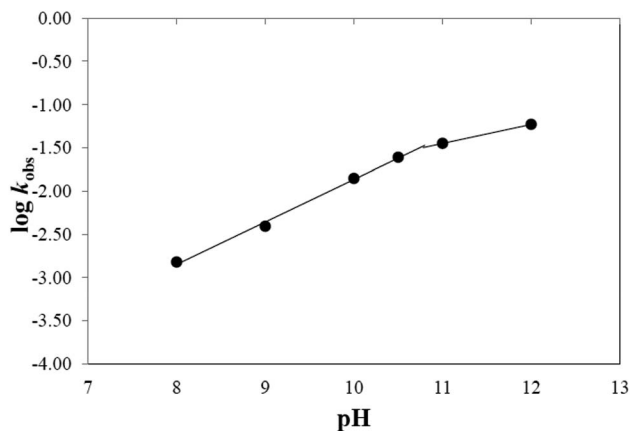


Fig. 7 $\log k_{obs}$ against pH plot for the photolysis of CMF in alkaline solution.

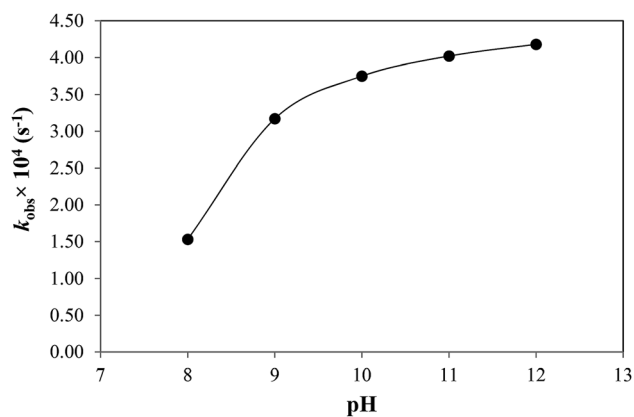


Fig. 5 k_{obs} -pH profile for the photolysis of CMF at pH 8.0–12.0.

(k_6) are given in Table 6. A plot of k_6 versus relative dielectric constant (ϵ) of the solvents is shown in Fig. 8. The increase in rate with ϵ indicates that the rate of reaction is controlled by the polarity of the solvent. This is probably due to the existence of a dipolar species in

photoreduction of flavins in aqueous and organic solvents as proposed by Ahmad and Tollin.⁴⁸ The reactivity of this intermediate depends on the extent of solvent interaction. This implies that if the polarity of the intermediate state is greater than that of the reactant it would lead to an increase in rate with an increase in relative dielectric constant of the solvent as reported in the case of RF¹⁶ and FMF.²⁰

The effect of solvent on the rate of a reaction can also be considered in terms of the acceptor number (AN) which determines the ability of a solvent to take shares in electron pairs from suitable donors.⁶⁴ When the $\ln(k_6)$ values for the photolysis of CMF are plotted against the AN of highly structured solvents (*i.e.* above 25), a linear relation is observed (Fig. 9). The AN is considered as a measure of the reactivity of a species in different solvents.⁶⁵ The solvent dielectric constant is considered as a good indicator for prediction of the extent of solute-solvent interaction.

Viscosity. The rate of a reaction may be influenced by the viscosity of the solvent as observed in the case of RF¹⁶ and

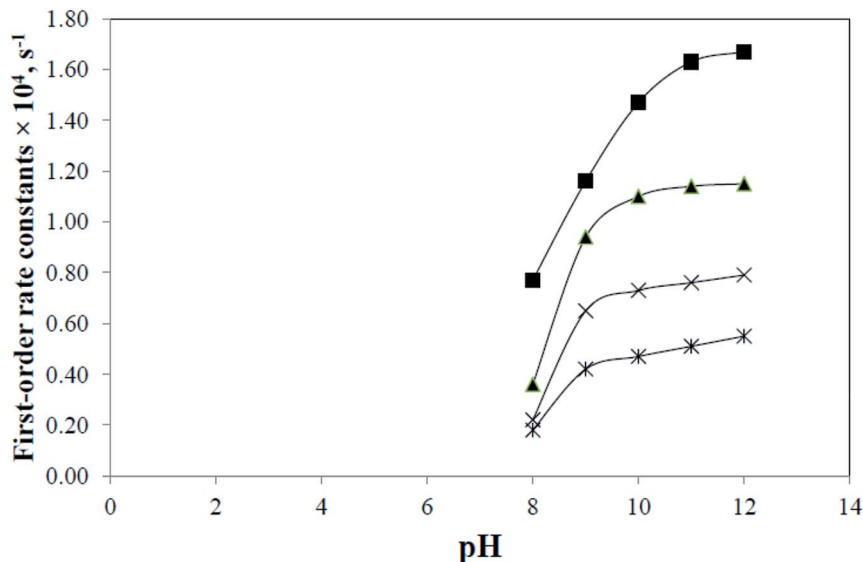


Fig. 6 k -pH profiles for the formation of photoproducts of CMF, (■) LC, (▲) LF, (×) KA, (*) DQ.



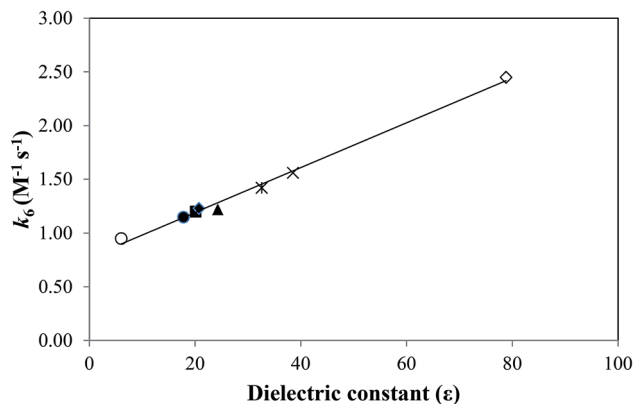


Fig. 8 Plot of k_{obs} for photolysis of RF against dielectric constant: (○) ethyl acetate, (●) 1-butanol, (■) 1-propanol, (◆) acetone, (▲) ethanol, (*) methanol, (×) acetonitrile, (◇) water.

FMF.²¹ A plot of k_6 versus inverse of solvent viscosity is shown in Fig. 10. It follows a linear relation indicating that the rate of reaction is slowed down by an increase in viscosity. This is in accordance with the quenching of flavin triplet state which is inhibited with an increase in solvent viscosity.⁴⁸ The rate of the reaction is suppressed by the viscosity of the solvent probably due to the involvement of a diffusion-controlled process.³⁵ The degradation of CMF in ethyl acetate does not comply with its behaviour in other solvents. This may be due to a difference in the orientation of the side-chain in that solvent⁴² that would determine the rate of reaction.

Effect of fluorescence

CMF possesses an isoalloxazine nucleus similar to that of other flavins (e.g. RF, FMF and LF) and emits a yellow green fluorescence at 530 nm when excited at 445 nm.^{19,29} Flavins are dipolar molecules and their fluorescence is destroyed by acid or alkali as a result of the cation or anion formation.⁶⁶ To observe the effect of pH on the ionization behavior of the molecule, the

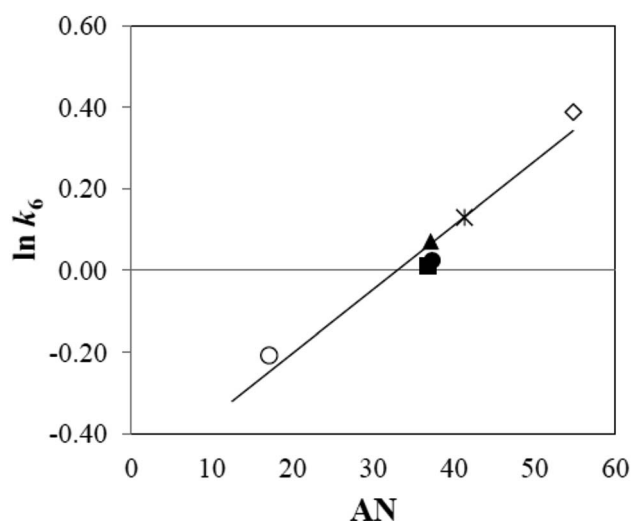


Fig. 9 Relation between $\ln k_6$ for CMF photodegradation and solvent acceptor number (AN) (symbols are as in Fig. 7).

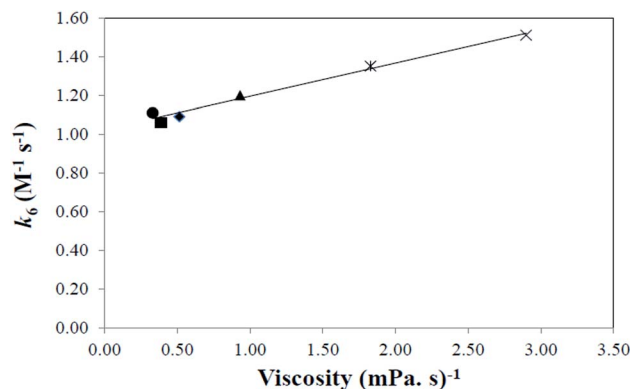


Fig. 10 Plot of k_6 for photolysis of CMF against inverse of viscosity: (●) 1-butanol, (■) 1-propanol, (◆) acetone, (▲) ethanol, (*) methanol, (×) acetonitrile.

fluorescence intensity of CMF was plotted against pH (Fig. 11). The curve indicates a slow increase in fluorescence intensity, with pH, in acid region due to gradual deprotonation of CMF. The maximum fluorescence is exhibited in the pH range 7 to 8 followed by a decline due to anion formation of the molecule in alkaline region. The ionized forms of flavins are less susceptible to photolysis in aqueous solution.¹³ The fluorescence of CMF has also been measured in organic solvents and the relative values of fluorescence intensity and quantum yields (0.09–0.27) of CMF in aqueous and organic solvents are given in Table 7. The deactivation of the excited singlet state (S_1) is enhanced with an increase in the polarity of the solvent and hence an increase in k_6 values with a loss in fluorescence intensity of CMF in the medium has been observed.

The quantum yield of fluorescence (Φ_F) of flavins depends on the solvent and is decreased with an increase in the polarity of the solvent.⁴¹ This is evident from the values of Φ_F and indicates a higher photostability of flavins in less polar solvents.⁶⁷ The fluorescence quantum yields may be affected by the degree of hydrogen bonding between the flavin and the solvent.⁶⁸

Dissociation constants of CMF

CMF tends to ionize in acid and alkaline medium due to protonation (N_{10}^+H) and anion formation ($N_{10}CH_2COO^-$ and N_3^-) of the molecule. The pK_a values of these groups have not been reported.

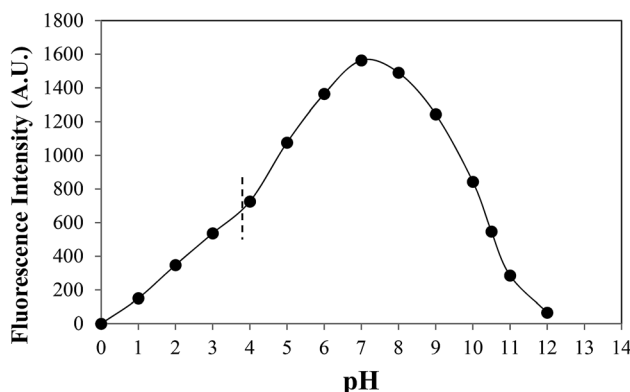


Fig. 11 A plot of fluorescence intensity of CMF against pH.



Table 7 Relative values of fluorescence intensity (% *F*) and quantum yields (Φ_F) in CMF (1.0×10^{-6} M) and organic solvents

Solvent	% <i>F</i>	Φ_F
Water (pH 7.0)	48.5	0.13
Acetonitrile	68.6	0.18
Methanol	87.2	0.23
Ethanol	100.0	0.27
1-Propanol	79.8	0.21
1-Butanol	36.1	0.10
Acetone	76.3	0.20
Ethyl acetate	33.7	0.09

Since CMF possesses the nucleus same as that present in RF, the pK_a s of N_{10}^+H and N_3^- group would be similar to those of RF (1.7 and 10.2, respectively).⁶² However, since the fluorescence of flavins is quenched by acid and alkali⁶⁶ due to ionization, the pK_a of the $N_{10}CH_2COOH$ group in the side-chain can be determined from the plot of fluorescence intensity *versus* pH (Fig. 11) according to the Henderson–Hasselbalch equation:

$$pH = pK_a + \log \frac{[\text{salt}]}{[\text{acid}]} \quad (11)$$

Considering the fluorescence intensity–pH curve, there appears to be an inflection point around pH 4.1 (with a fluorescence intensity corresponding to 50% ionization of the molecule). Therefore, a value of 4.1 can be considered as the pK_a of the $N_{10}CH_2COOH$ group of CMF. *N*-heterocyclic carboxylic acids have pK_a s in the range of 2.9 to 4.4.^{69,70} Fluorimetry has been used for the determination of the pK_a values of *N*-heterocyclic bases such as 8-methylquinoline and acridine.⁷¹

Quantum yields of photolysis

The quantum yields (Φ) of the photolysis of CMF at pH 2.0–12.0 have been determined and are given in Tables 3 and 4. The values of Φ in this pH range vary from 0.05 to 0.41 and are affected by the pH and reactivity of the excited triplet state of CMF. In the acid and neutral range (2.0–7.0), the Φ of formation of LC are low. In alkaline range (8.0–12.0), the Φ gradually increases since the triplet state has high reactivity in alkaline solution.¹¹ These values show a slight decrease in the pH range 10–12 due to anion formation of the molecule which has low susceptibility to photolysis like those of RF¹³ and FMF.²¹ The values of Φ of CMF are higher than those of RF photolysis in presence of divalent anions.⁷²

LC-ESI-MS analysis. Both, standard and CMF were analyzed by LC-ESI-MS using Tof-MS spectrometry. Riboflavin has shown an $[M + H]^+$ ion at m/z 377.1440 (calcd 377.1456) corresponding to $C_{17}H_{20}N_4O_6$, CMF has shown $[M + H]^+$ ion at m/z 301.0925 (calcd 301.0936) corresponding to $C_{14}H_{12}N_4O_4$. LC has shown $[M + H]^+$ ion at m/z 243.0872 (calcd 243.0882) corresponding to $C_{12}H_{10}N_4O_2$, similarly LF has shown $[M + H]^+$ ion at m/z 257.1033 (calcd 257.1033) corresponding to $C_{13}H_{12}N_4O_2$. While DQ has shown $[M + H]^+$ ion at m/z 205.0971 (calcd 205.0972) corresponding to $C_{16}H_{19}N_2O_2$. The peak for β -KA was not found using positive ionization mode which could probably appear in

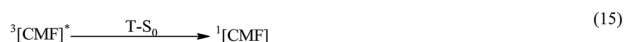
negative ionization mode of MS. Extracted ion chromatograms of all the mentioned compounds along with their high resolution masses (Fig. 1–5) are given in ESI.†

Mode of photolysis

CMF undergoes photolysis to form LC in acid solution. The kinetic data indicate that the transformation of CMF to LC takes place by a second-order reaction. This is in accordance with the photolysis of FMF which also yields LC in acid solution.²¹ The formation of LC and LF in alkaline solution has been found to take place by parallel first-order reactions as observed in the case of FMF.^{17–19,21} Therefore, an analogous scheme for the photolysis of CMF is proposed (Scheme 2).

In acid solution CMF is promoted to the excited singlet state, $^1[CMF]^*$, by the absorption of a photon of light (eqn (12)) which may be deactivated to the ground state, $^1[CMF]$ (eqn (13)), or is converted to triplet state, $^3[CMF]^*$ (eqn (14)) by intersystem crossing (ISC). $^3[CMF]^*$ may return to the ground state (eqn (15)) or on interaction with a ground state molecule is photolyzed to LC and CH_3COOH (eqn (16)). Alternatively it may give rise to a cationic $[CMF^{+\cdot}]$ and an anionic $[CMF^{-\cdot}]$ radical (eqn (17)), which on losing or accepting a proton form a neutral radical $[CMF^{\cdot}]$ (eqn (18)) and a semiquinone radical $[CMFH^{\cdot}]$ (eqn (19)), respectively, as described previously in the case of other flavins.⁴⁸ Two semiquinone radicals

Acid Solution



Alkaline Solution



Scheme 2



disproportionate⁷³ to give a neutral molecule and a dihydro intermediate product [CMFH₂] (eqn (20)) similar to that predicted by Berdicka⁷⁴ in the photolysis of RF and suggested by Ahmad *et al.* eqn (21) in the photolysis of FMF. The neutral radical reacts with a neutral molecule and they are degraded to a semiquinone radical, LC and CH₃COOH (eqn (22)).

In the alkaline solution the photolysis of CMF takes place by hydrolytic degradation as reported in the case of FMF.^{17,18,21} An increase in rate at pH 8.0–10.0 could be attributed to configurational changes and greater reactivity of the flavin triplet state to alkaline hydrolysis.¹¹ This leads to the formation of LF and formic acid (eqn (22)) and LC and acetic acid (eqn (23)). CMF is also degraded by hydrolytic cleavage of isoalloxazine ring in alkaline solution to yield KA (eqn (24)) and DQ (eqn (25)).²⁹

Conclusions

CMF is an intermediate product in the photolysis of RF. On visible irradiation the side-chain of CMF is removed to form LC in acid solution and LC and LF in alkaline solution. The isoalloxazine ring of the molecule is also cleaved in alkaline solution to yield quinoxaline derivatives. In organic solvents CMF is photodegraded to LC only. CMF is degraded in acid solution and organic solvents by second-order kinetics and in alkaline solution by simultaneous first-order kinetics. The rate constants for the formation of LC and LF in alkaline solution are higher than those of the quinoxaline derivatives. CMF degradation is increased in alkaline solutions due to the enhanced reactivity of the excited triplet state. The photostability of CMF is greater in organic solvents than in the aqueous solution due to a decrease polarity of the medium. The photoproducts of CMF have been identified chromatographically and determined spectrofluorimetrically in degraded solutions. The quantum yields of fluorescence and photolysis of CMF have been reported. A scheme for photodegradation reactions of CMF in aqueous and organic solvent is presented.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors gratefully acknowledge Higher Education Commission of Pakistan for financial support to conduct this study through a grant (Research Project # 20-3968) to Iqbal Ahmad. They Thank Professor Alfons Penzkofer University of Regensburg, Germany and Professor Marek Sikorski, Adam Mickiewicz University of Poznan, Poland for helpful discussions.

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