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Riboflavin-protected ultrasmall silver nanoclusters with enhanced antibacterial activity and the mechanisms†

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Developing silver nanomaterials with efficient antimicrobial properties is of importance for combating bacteria. Here, we report ultrasmall riboflavin-protected silver nanoclusters (RF@AgNCs) that can effectively kill or suppress the growth of Gram-positive *S. aureus*, Gram-negative *E. coli*, and fungi *C. albicans*. Riboflavin (RF) with intrinsic biocompatibility was used as a surface ligand to synthesize silver nanoclusters. TEM revealed that the synthesized RF@AgNCs were ultrasmall (2.4 ± 1.2 nm), spherical and well-dispersed. Antibacterial activity tests showed that RF@AgNCs possessed superior antibacterial efficacy in comparison with RF, AgNPs and mixed RF and AgNPs (RF + AgNPs). The mechanisms of antibacterial activity of RF@AgNCs were studied by fluorescence microscopy-based Live/Dead cell staining assays and ROS measurement. And the results illustrated that the integrity of the bacteria membrane was disrupted and intracellular high level ROS generation was induced by RF@AgNCs. The cytotoxic activities were also assessed and RF@AgNCs were found to be non-toxic to human red blood cells and mammalian cells. With the highly efficient antibacterial activity and acceptable biocompatibility, RF@AgNCs hold great promise in biomedical applications as well as in water sterilization and the textile industry.

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1. Introduction

Silver (Ag) and its compounds have been accepted as antimicrobial agents in medicine and public health for centuries. In ancient times, people used silver containers to store water or food to keep it fresh. Then silver foil was used to cover wounds for the prevention of ulceration. Silver nitrate (AgNO₃) was introduced to prevent newborn gonorrhoeal eye infections in 1884. In recent years, a new generation of antimicrobials, silver nanoparticles (AgNPs),^{1–4} has drawn significant attention owing to the intrinsic broad-spectrum antibacterial properties, non-drug resistance, good biocompatibility,⁵ and low dosage. They show more advantages than bulk Ag. It is reported that a lower concentration of AgNPs can inhibit or kill a wide range of bacteria in a few minutes.⁶ Recent studies show that AgNPs are able to improve wound healing.^{7–9} With the emergence of antibiotic-resistant bacteria, it is highly desirable to develop new efficacious broad-spectrum antibacterial agents.^{10,11} All these make AgNPs or their complexes one of the most promising antibacterial agents.¹² Now they have been extensively used in medical applications including wound dressings and

treatment, food packaging, the textile industry, water treatment, and environmental management.

Some studies have reported that the antibacterial activities of AgNPs depend on their size,^{13,14} shape,¹⁵ surface charge,¹⁶ and surface coating.¹⁷ AgNPs with different sizes exerted different antibacterial properties.^{1,18} AgNPs below 10 nm exhibited surface-dependent ion release and toxicity. They can effectively penetrate the bacterial wall and affect the integrity of membrane and the viability of bacteria, exhibiting higher potency against bacteria cells.^{19,20} Xie J. P. *et al.* have reported that silver nanoclusters (AgNCs) with 2–3 nm possess higher activities than AgNPs.¹⁹ In addition, surface coating (surface ligand) plays a prominent role in toxicity of nanoparticles (NPs). AgNPs are typically synthesized through surface coating which reduces AgNO₃ as well as stabilizes AgNPs in suspension. Hinton D. E. *et al.* have demonstrated that dissolution of AgNPs was largely dependent on their particle size, but their aggregation behavior and toxicity were more dependent on coating materials.^{21,22} Doktycz M. J. *et al.* have indicated that bacteria growth inhibition depended on the surface coating after the comparison of protein-protected AgNPs, colloidal AgNPs, and oleate-protected AgNPs.²³ Zhou Q. F. synthesized AgNPs with four different coatings and illustrated that surface coating controlled silver ion release and their antibacterial activity.²⁴ Moreover, surface coating determines potential cytotoxicity of AgNPs to human cells or the effect on environmental health.²⁵ Therefore, it is fundamentally important to optimize surface coating to

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develop small or ultrasmall AgNPs with high antibacterial properties but a proper human biocompatibility which is essential for daily life application.

Riboflavin (RF, vitamin B₂), an important component of cofactors FAD (flavin adenine dinucleotide) and FMN (flavin mononucleotide), is crucial for biological oxidation metabolism. RF can act as growth inhibitor and fungicide because riboflavin molecule can intercalate into DNA or RNA, making pathogens unable to replicate. Some studies have reported that RF effectively enhanced resistance against virus and fungi in plant.^{26,27} Others have indicated that RF can promote the disappearance of inflammation and can be used to treat burns or stomach ulcers and prevent upper oral ulcers. RF/light combination or their complex with other compounds are often used as antimicrobial or antiviral agents.^{28,29} Moreover, there is a conjugated double bond of isoalloxazine ring in RF molecular structure which can be involved in redox reactions. RF can be used for the reduction of AgNO₃ because the reduction potential is lower than Ag.^{30,31} Therefore, using riboflavin as surface coating to synthesize AgNPs may integrate the characteristics of RF and antimicrobial effects of silver nanoparticles which may show more efficient antimicrobial than RF or AgNPs alone. Furthermore, RF has intrinsic biocompatibility and RF as surface ligand may have no toxicity to human or environment.

In this study, riboflavin-protected silver nanoclusters (RF@AgNCs) were prepared by RF reducing AgNO₃. RF has certain antibacterial properties and serving as surface coating can reduce the cytotoxicity of silver nanoclusters. The antimicrobial activity against Gram-positive bacteria (*S. aureus*), Gram-negative bacteria (*E. coli*), and fungus (*C. albicans*) were evaluated. Fluorescence images showed that RF@AgNCs could disrupt bacteria cell membranes. The antibacterial mechanism was discussed by intracellular reactive oxygen species (ROS)

level assay. The schematic diagram of RF@AgNCs against bacteria is shown in Fig. 1. *In vitro* biocompatibility of AgNCs was evaluated by hemolysis assay and MTT test. The results show that the RF@AgNCs have superior antimicrobial and antifungal effects and low toxicity to normal human cells.

2. Materials and methods

2.1. Reagents and materials

Riboflavin and AgNO₃ were purchased from Aladdin Bio-Chem Technology Co., LTD (Shanghai, China). Phosphate-buffered saline (PBS, pH 7.2) was bought from Hyclone, Thermo Scientific (Massachusetts, USA). Hoechst 33342 and propidium iodide (PI) were purchased from Beyotime Biotechnology (Shanghai, China). 3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) and 2',7'-dichlorofluorescein diacetate (DCFH-DA) were bought from Sigma-Aldrich (St. Louis, USA). The Gram-positive *Staphylococcus aureus* (*S. aureus*) (CGMCC 1.0089), Gram-negative *Escherichia coli* (*E. coli*) (CMCC (B) 44102), and fungi *Candida albicans* (*C. albicans*) (CGMCC 2.2086) were bought from the Microbiology Institute of Shaanxi Province (Xi'an, China). Human Umbilical Vein Endothelial (HUVE) cells (KCB 2012087YJ) and Human Kidney (HK-2) cells (KCB 200815YJ) were purchased from the Conservation Genetics, the Chinese Academy of Sciences Kunming Cell Bank (Kunming, China). The human blood sample was supplied by Xi'an No. 3 Hospital. All experiments performed were in compliance with relevant laws, as well as the guidelines of Biomedical Research Ethics Committee of the School of Life Science & Technology, Xi'an Jiaotong University. And the experiments were approved by all the above mentioned institutes. The volunteer had provided his informed consent for the experiments. Ultrapure water (18.2 MΩ) was used throughout

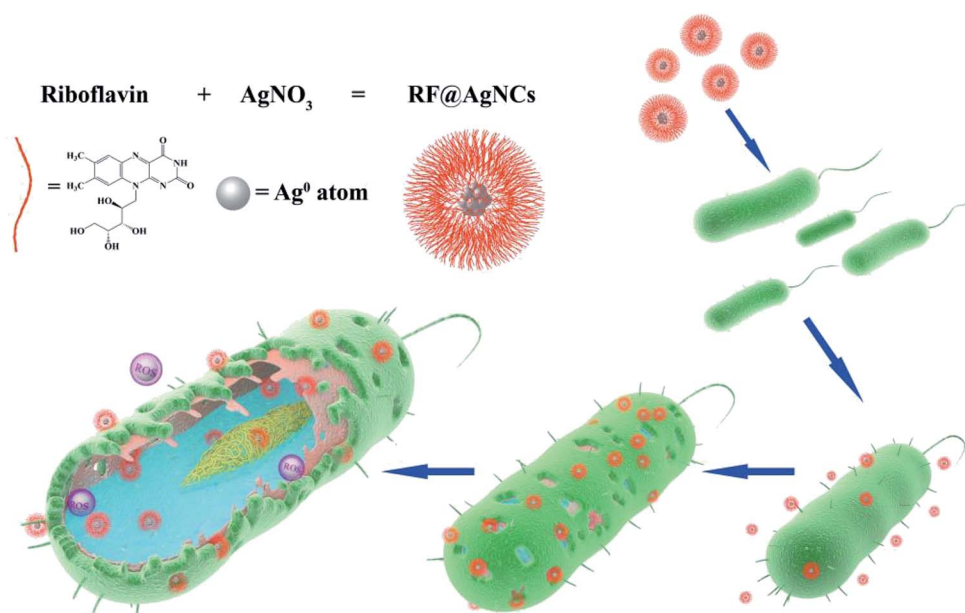


Fig. 1 Schematic diagram of the synthesis and antibacterial activity of RF@AgNCs.



the experiment. All glassware and magnetic stirring bars were soaked in *aqua regia*, rinsed with ultrapure water, and dried in an oven before use.

2.2. Preparation of and characterization of RF@AgNCs

Riboflavin-protected silver nanoclusters (RF@AgNCs) were synthesized according to the previously reported method³⁰ with some modification. In brief, 1 mL 1 mM homogeneous riboflavin solution was added into 8 mL water under stirring at room temperature. Then, 1 mL AgNO₃ solution (0.1 mM) was introduced dropwise to the above solution. The mixture was stirred for 3 h at 4 °C. The color of the mixture changed from bright yellow to reddish orange, indicating the formation of AgNCs. The morphologies of RF@AgNCs were observed with transmission electron microscopy (TEM, JEM-2100F, JEOL). The measurement of UV-vis absorption (Tecan Infinite M200 Pro, Switzerland) and zeta potential (Mastersizer Nano ZS90, Malvern) were performed to characterize the surface modification of RF@AgNCs.

Silver nanoparticles (AgNPs) were synthesized based on the previous method.³² Briefly, 10 mM Tollen reagent was firstly prepared by AgNO₃ and ammonium hydroxide at a certain proportion. Then, 0.5 g β -cyclodextrin and 2 mL 50 mM glucose were added into 1 mL 10 mM Tollen reagent and ultrapure water was added to the total volume of 50 mL. When the mixture was heated to be boiled, it was continuously stirred and refluxed for 15 min until the solution turned to light yellow. The synthesized AgNPs was analyzed by the Microplate Reader and the Malvern Mastersizer.

2.3. Bacteria growth and antimicrobial activity test

S. aureus, *E. coli*, and *C. albicans* were recovered from frozen glycerol stocks on Luria–Bertani (LB) agar plates or potato dextrose (PD) agar plates at 37 °C. Single colony of bacteria were then incubated for 24 h to exponential phase. Then, the bacteria suspension was diluted by PBS to achieve the concentration of 1×10^7 colony forming unit (CFU) per mL (OD₆₀₀ = 0.05) prior to antibacterial test.

Antibacterial activities of RF@AgNCs against each kind of bacteria were determined by the colony counting method. PBS, RF (0.28 $\mu\text{g mL}^{-1}$), AgNPs (0.125 $\mu\text{g mL}^{-1}$, on the basis of Ag atoms), mixed RF and AgNPs (RF + AgNPs, 0.125 $\mu\text{g mL}^{-1}$ Ag atoms and 0.28 $\mu\text{g mL}^{-1}$ RF) were used as control groups. Each bacterial group was incubated separately with control groups and RF@AgNCs (0.125 $\mu\text{g mL}^{-1}$, on the basis of Ag atoms) under continuous orbital shaking at 200 rpm at 37 °C for 3 h. Then the treated bacteria suspension of each group was diluted by 10 000 folds, and 50 μL of the dilutions were added into the agar plates to let the bacteria grow at 37 °C. Finally, the inactivation of bacterial cells was evaluated by counting the number of colony forming units (CFU) on the agar plate after 48 h incubation. Each assay was carried out in triplicate.

2.4. Fluorescent microscopy study

The bacterial cells (1×10^7 CFU mL⁻¹) were incubated separately with RF@AgNCs and control groups at 37 °C for 3 h. Then

1 $\mu\text{g mL}^{-1}$ Hoechst 33342 and 5 $\mu\text{g mL}^{-1}$ PI were added to the treated bacteria solutions respectively and the solutions were kept in dark for 15 min. After rinsed with PBS for 3 times, the new suspension (5 μL) was dropped on a glass slide for fluorescent microscopy imaging (Nikon ECLIPSE Ti, Japan).

2.5. TEM images of bacteria cells

The bacteria solutions (1×10^7 CFU mL⁻¹, 500 μL) were mixed with RF@AgNCs solution (2 $\mu\text{g mL}^{-1}$, 500 μL) at room temperature, respectively. After incubation for 2 h, 2 μL of the mixture were dropped on a carbon-coated Cu grid (400 Mesh) for TEM observation.

2.6. ROS measurement

DCFH-DA fluorescent probe was used for the evaluation of the level of intracellular reactive oxygen species (ROS) when exposed to NPs. The bacteria cells were incubated with RF@AgNCs and control groups for 2 h respectively. The treated bacteria solution was incubated with 5 μM DCFH-DA in an orbital shaker (200 rpm) at 37 °C for 30 min. Then the bacteria suspension was centrifuged, washed and re-suspended in ultrapure water to remove the redundant DCFH-DA outside cells. The fluorescent signal ($\lambda_{\text{ex}}/\lambda_{\text{em}} = 484/525$ nm) was measured on a fluorescence microscope (Nikon ECLIPSE Ti, Tokyo, Japan) and Microplate Reader (Tecan Infinite M200 Pro) to evaluate ROS production in bacteria cells. Each sample was taken for three replicates.

2.7. Hemolysis assay

Hemolysis assay was performed with red blood cells (RBCs). The erythrocytes were collected from fresh human blood (5 mL) by centrifugation at 2000 rpm for 5 min, and washed three times with PBS. Then 0.5 mL of RBC solution was mixed with 0.5 mL of RF@AgNCs. After incubation at 37 °C (150 rpm) for 30 min, the mixture was centrifuged at 4000 rpm for 5 min. At last, the absorbance of the supernatant was measured at 570 nm. RBCs incubated with PBS and water were treated as negative and positive control, respectively. All samples were prepared in triplicate at one test. The percent hemolysis was calculated by the following equation:

$$\% \text{ hemolysis} = \frac{\text{sample absorbance} - \text{negative control absorbance}}{\text{positive control absorbance} - \text{negative control absorbance}} \times 100$$

2.8. MTT assay

Two types of cells (HK-2 cells and HUVE cells) were used in MTT assay to evaluate the effects of RF@AgNCs treatment on cell proliferation behavior. In brief, cells were maintained in low-glucose Dulbecco's Modified Eagle's Medium with 10% fetal bovine serum (FBS) and 1% antibiotic (100 U mL⁻¹ penicillin and 100 $\mu\text{g mL}^{-1}$ streptomycin). Cells were cultured at 37 °C under a humidified atmosphere of 5% CO₂ (96-well plate, 4000



cells per well). Media was removed and cells were washed with PBS. Cells were treated with RF@AgNC at various concentrations ($0.125\text{--}5\ \mu\text{g mL}^{-1}$) for 24 and 48 h, respectively. Then cell samples were incubated with MTT solution for another 4 h to determine living cells. The culture medium containing MTT was removed and $150\ \mu\text{L}$ dimethyl sulfoxide (DMSO) was added into each well to dissolve the formazan crystals. The $\text{OD}_{570\ \text{nm}}$ value was measured by a Microplate Reader. Cells without any treatment were normalized as 100%. Each experiment was performed in triplicate.

3. Results and discussion

3.1. Characterization of RF@AgNCs

Riboflavin (vitamin B_2) has two forms of oxidation and reduction types because there is a conjugated double bond of isoalloxazine ring in its structure. In this study, riboflavin (RF) was used as reducing agent to synthesize RF-protected silver nanoclusters (RF@AgNCs). Synthesized RF@AgNCs were characterized by UV-vis absorption spectrum, TEM, and zeta potential measurements. In UV-vis absorption spectrum, the peaks decrease of RF and the appearance of the peak at $535\ \text{nm}$ indicates the formation of RF@AgNCs (Fig. 2a). TEM images show that synthesized RF@AgNCs are well-dispersed and quasi-spherical (Fig. 2b). The lattice fringes of the RF@AgNCs are consistent with metallic Ag which exhibits a discerned lattice spacing of $0.207\ \text{\AA}$ (inset in Fig. 2b). The size distribution of RF@AgNCs was in concordance with normal distribution curve and the average size was $2.4 \pm 1.2\ \text{nm}$ (Fig. S1†). The zeta potential of RF@AgNCs was $+0.283\ \text{mV}$, indicating a neutral surface of RF@AgNCs. Moreover, AgNPs as control group were also characterized by UV-vis absorption spectrum and zeta potential measurements (Fig. S2†).

3.2. *In vitro* antibacterial activity of RF@AgNCs

The plate counting method was used to evaluate the antimicrobial activity of RF@AgNCs against Gram-positive *S. aureus*,

Gram-negative *E. coli*, and fungi *C. albicans*. Fig. 3a shows the antibacterial photographs after the bacteria were treated with RF@AgNCs ($0.125\ \mu\text{g mL}^{-1}$, on the basis of Ag atoms) and the control groups including PBS, RF ($0.28\ \mu\text{g mL}^{-1}$), AgNPs ($0.125\ \mu\text{g mL}^{-1}$), and the mixed RF & AgNPs (RF + AgNPs) for 2 h, respectively. Apparently, RF@AgNCs efficiently inhibited the growth of three kinds of bacteria because the colonies were hardly observed on the Petri dish. Although AgNPs and mixed RF & AgNPs showed antibacterial effects against *E. coli*, the colony number was a bit more than that observed in the sample treated by RF@AgNCs. The relative viabilities of *S. aureus*, *E. coli* and *C. albicans* after treated with RF@AgNCs were 0.83, 2.08, 3.24%, respectively (Fig. 3b). The results indicate that RF@AgNCs are more effective in inhibiting the proliferation of the bacteria and fungi than control groups (RF, AgNPs, and RF + AgNPs), exhibiting superior antibacterial efficacy.

3.3. Study on the mechanisms about antibacterial activity of RF@AgNCs

3.3.1. Fluorescence imaging and TEM observation on bacterial membrane integrity. The toxicities of NPs to bacterial cells are based on direct damage of bacterial cell membranes.³³ We employed fluorescence microscopy-based Live/Dead cell staining assays to observe the integrity of bacterial cell membranes after incubated with RF@AgNCs. The fluorescent dye of PI cannot penetrate cell membrane of viable cells, but can permeate dying or dead cells to stain their DNA with red fluorescence. Hoechst 33342 is another fluorescent dye which is often employed to stain DNA of all live bacterial population with blue fluorescence. So intracellular staining of PI can identify dead cells, while Hoechst 33342 staining assay can identify live cells. After *S. aureus* treated with RF@AgNCs or control groups for 2 h and stained with PI or Hoechst respectively, the fluorescence images are shown in Fig. 4. From Fig. 4, it can be seen that almost no *S. aureus* cells can be stained by PI in the PBS group and RF group. The AgNPs group and mixed (RF + AgNPs) show lower intensities of red fluorescence, indicating a few of

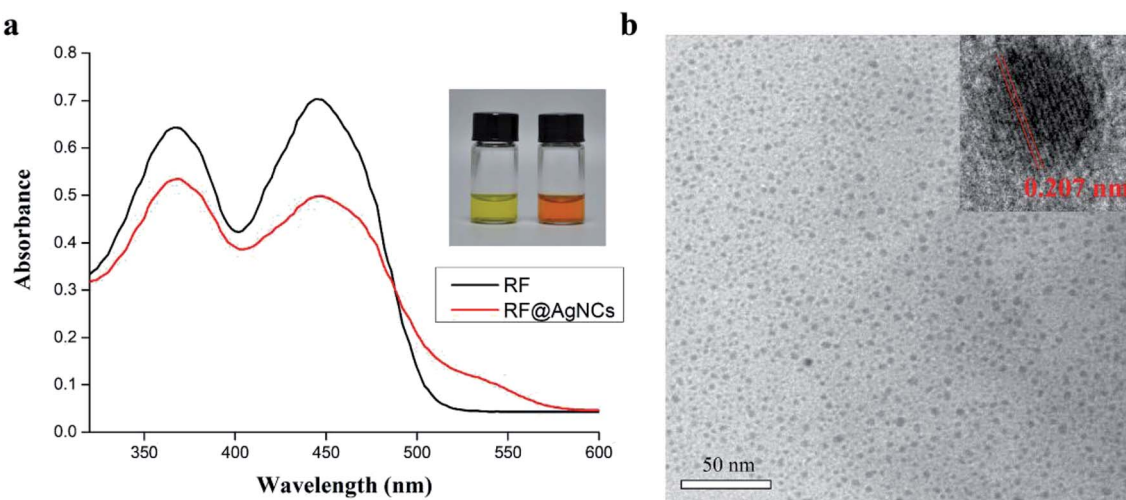


Fig. 2 Characteristics of synthesized RF@AgNCs. (a) UV-vis absorption spectrum of RF and RF@AgNCs. (b) TEM image of RF@AgNCs.



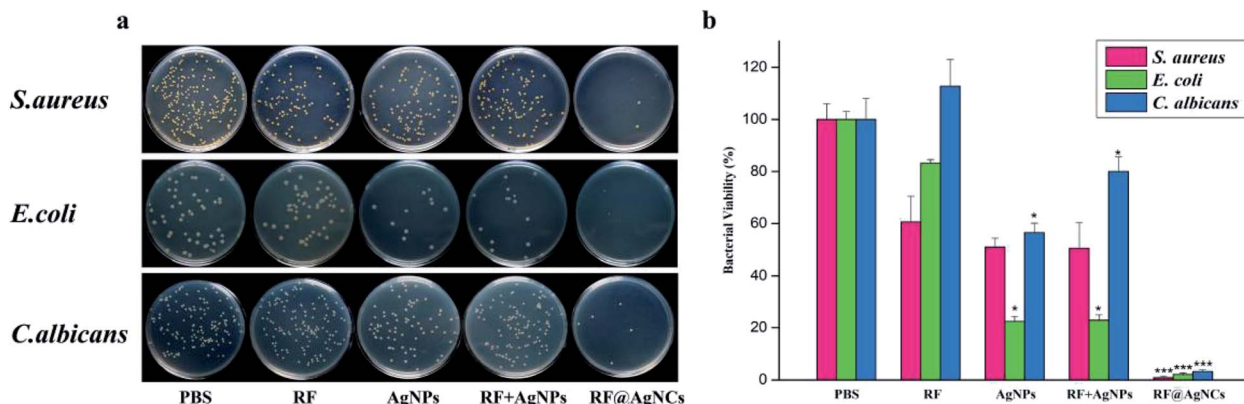


Fig. 3 Antibacterial assay *in vitro*. (a) Photographic images of bacterial colonies formed by *S. aureus*, *E. coli* and *C. albicans* after the treatment with RF@AgNCs and control groups (PBS, RF, AgNPs, RF + AgNPs), respectively. (b) The corresponding cell viability of *S. aureus*, *E. coli* and *C. albicans* incubated with various material suspensions. * $p < 0.05$, *** $p < 0.001$.

dying or dead bacteria cells. However, almost all *S. aureus* cells incubated with RF@AgNCs were stained to be red fluorescence with PI. Similarly, almost no *S. aureus* cells were stained to be blue fluorescence by Hoechst, which further confirms that few of live cells exist after treated with RF@AgNCs. The fluorescence microscopy images in Fig. S3 and S4† show RF@AgNCs-induced membrane damage in both *E. coli* and *C. albicans* cells. Therefore, strong red fluorescence and weak blue fluorescence indicate that the permeability of bacteria increase and the integrity of bacterial cell membranes were indeed disrupted by RF@AgNCs.

We also used TEM to directly observe the change of bacterial cell membranes integrity. Morones J. R. reported that AgNPs (1–10 nm) could enter into bacterial cells and cause further damage by possibly interacting with sulfur- and phosphorus containing compounds.²⁰ Fig. 5 shows TEM images of *S. aureus*, *E. coli*, and *C. albicans* untreated and treated with RF@AgNCs for 2 h. In the absence of RF@AgNCs, three bacterial cells exhibit the typical smooth structure, while the morphology and membrane integrity were strongly changed after the treatment with RF@AgNCs. We can see that RF@AgNCs penetrate bacteria cell easily and are aggregated or accumulated inside cells,

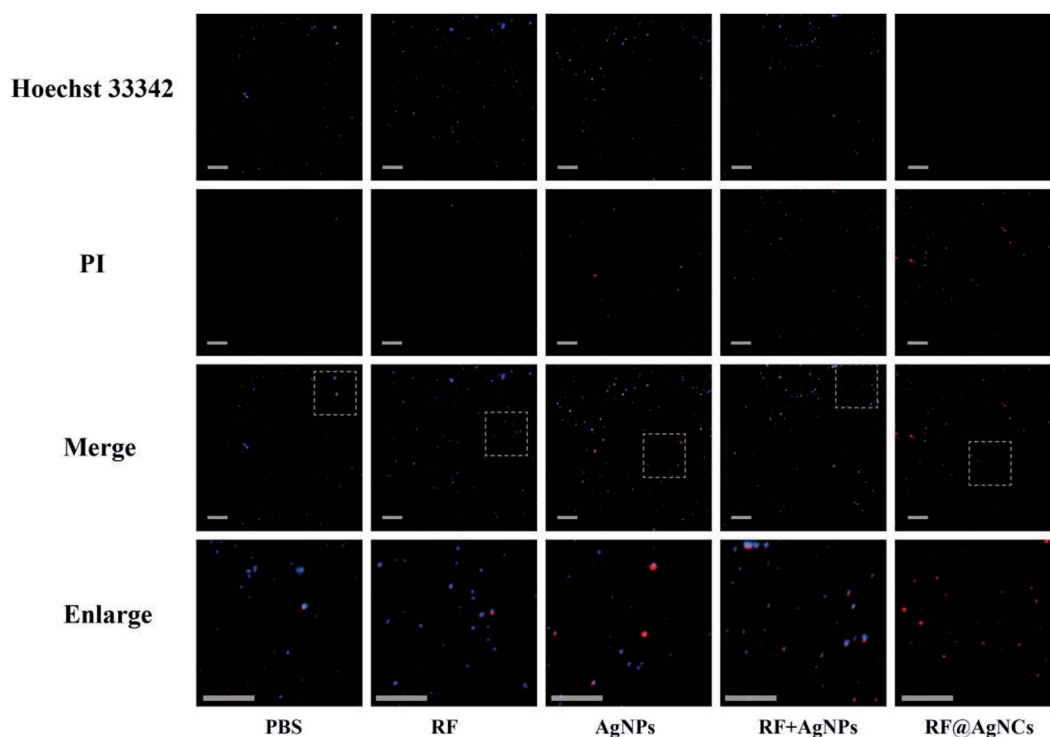


Fig. 4 Representative fluorescence microscopy images of the *S. aureus* cells incubated with RF@AgNCs and control groups for 2 h, and stained with PI and Hoechst respectively. The dead cells were visualized by PI staining (red), while living cells were observed by the Hoechst 33342 staining (blue). Scale bar is at 25 μm .



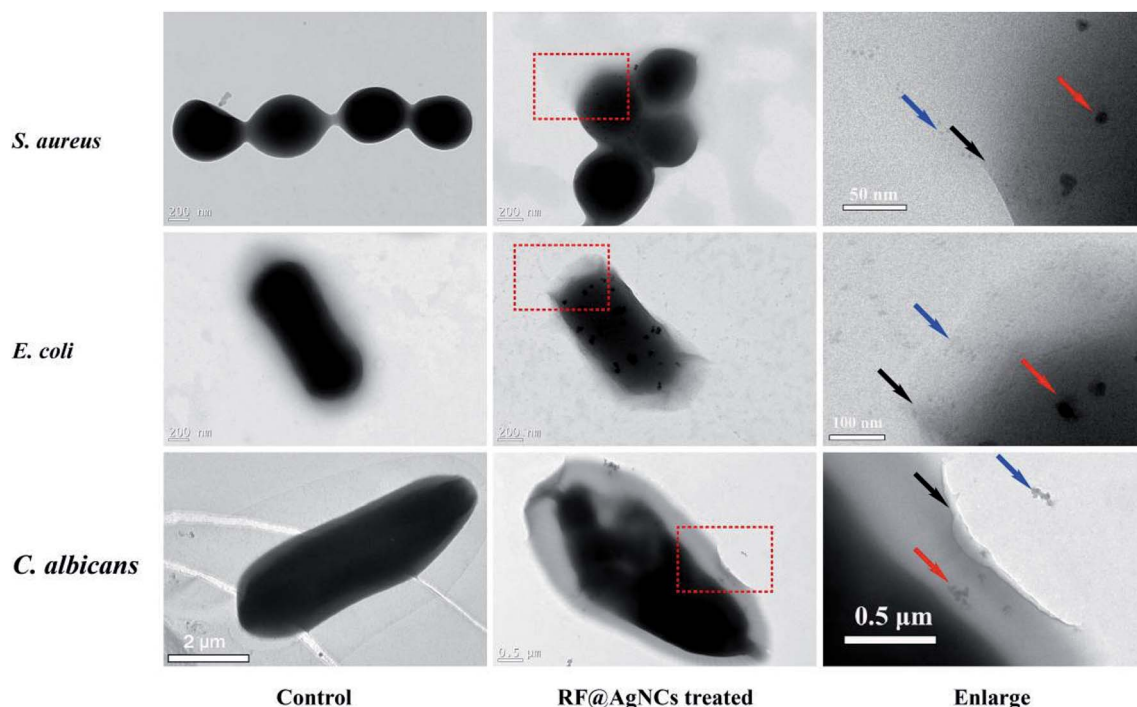


Fig. 5 TEM images of *S. aureus*, *E. coli*, and *C. albicans* untreated and treated with RF@AgNCs for 2 h. The blue arrow directs the RF-Ag clusters. Black arrow demonstrates the broken bacterial cells membrane. Red arrow indicates the aggregation of RF@AgNCs in the bacterial cells.

causing structural changes in bacterial cell walls or membranes. It can even be observed that some bacterial cells were broken (Fig. S5[†]). These morphological changes demonstrate the beneficial antibacterial activities of RF@AgNCs against Gram-positive *S. aureus*, Gram-negative *E. coli*, and fungi *C. albicans*.

3.3.2. Antibacterial mechanism by ROS assay. Many reports have shown that the toxicity of NPs is likely caused by the release of Ag ions and the generation of intracellular reactive oxygen species (ROS), which might impair the integrity of bacteria cell membranes, leading to bacteria death.³⁴ It is also reported that enhanced ROS would be generated by the RF upon

irradiation in the presence of AgNPs.^{35,36} In order to measure the intracellular levels of ROS after exposure to RF@AgNCs, an active oxygen highly specific fluorescent probe (DCFH-DA) is used. Non-fluorescent probe DCFH-DA penetrates into cells and hydrolyzed by enzymes to produce DCFH. In the presence of ROS, DCFH is oxidized to yield the highly fluorescent 2',7'-dichlorofluorescein (DCF) with green fluorescence. After the bacteria cells were incubated with RF@AgNCs or control groups for 2 h, the fluorescence images of three bacteria strains were shown in Fig. 6, S6 and S7.[†] From Fig. 6, it can be seen that the ROS level of *S. aureus* cells incubated with RF@AgNCs are

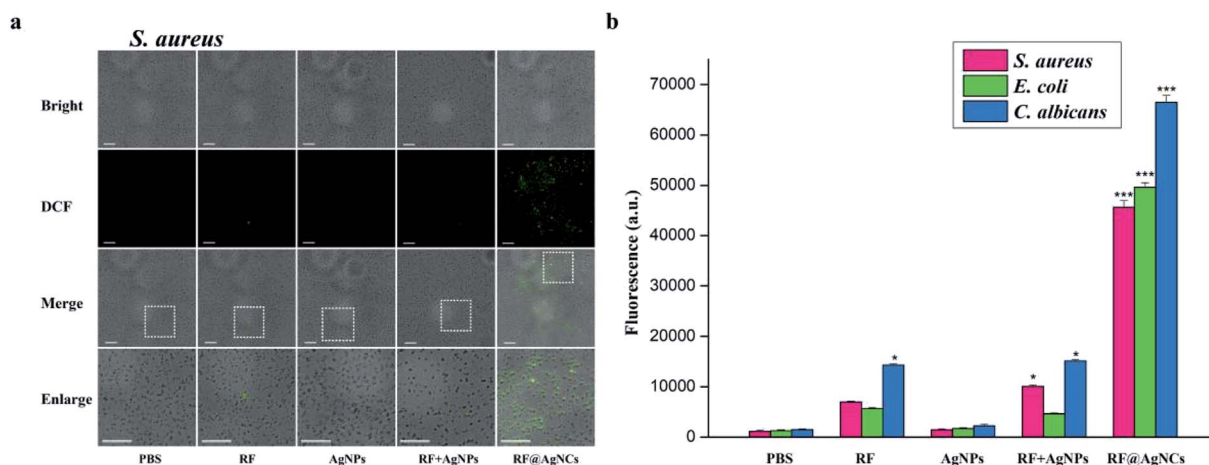


Fig. 6 Intracellular ROS production. (a) Fluorescent microscopic images of the *S. aureus* cells after incubated with RF@AgNCs and control groups for 2 h respectively, followed by staining with DCFH-DA. Scale bar is at 25 μ m. (b) The ROS levels of the *S. aureus*, *E. coli* cells, and *C. albicans* represented by the fluorescence intensity of DCF, detected by a microplate reader. * $p < 0.05$, *** $p < 0.001$.



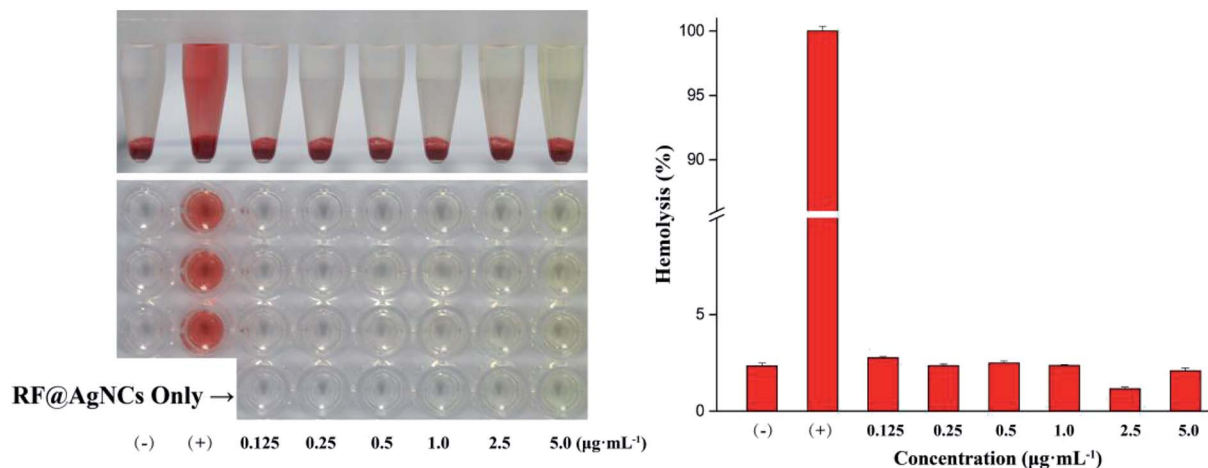


Fig. 7 Hemolysis assay of RF@AgNCs at different concentrations on RBCs.

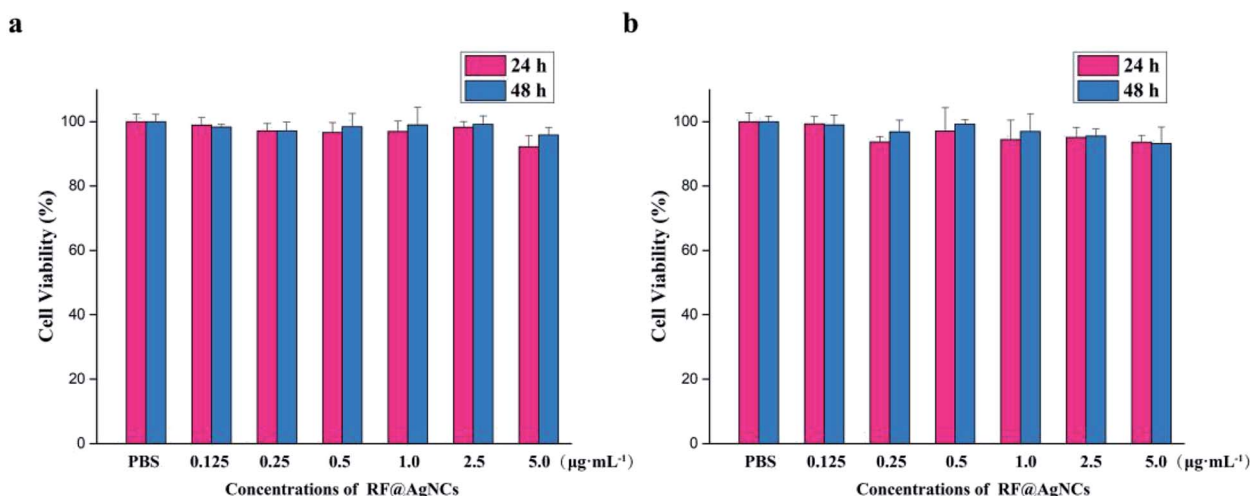


Fig. 8 Cell viability assay of RF@AgNCs at different concentrations. (a) HUVE cells and (b) HK-2 cells.

significantly enhanced in comparison with the control groups. Similarly, strong green fluorescence signal in *E. coli* cells and *C. albicans* cells was also observed after the treatment with RF@AgNCs in Fig. S6 and S7.† The results illustrate that RF@AgNCs significantly induced higher levels of intracellular ROS produced than those produced by other groups such as RF, AgNPs or mixed RF & AgNPs.

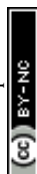
3.4. Hemolysis and cytotoxicity assays

To evaluate the biocompatibility or cytotoxicity of RF@AgNCs, hemolysis assay on human red blood cells (RBCs) and MTT assay on mammalian cells of HUVE cells and HK-2 cells were carried out. As shown in Fig. 7, the treatment of RBCs with RF@AgNCs at various concentrations ($0.125\text{--}5\ \mu\text{g}\cdot\text{mL}^{-1}$) show no observable hemolytic activity, indicating the biocompatibility of RF ligands with human blood cells. MTT assay were used to assess the proliferation of HUVE cells and HK-2 cells after exposed to RF@AgNCs at various concentrations ($0.125\text{--}5\ \mu\text{g}\cdot\text{mL}^{-1}$). As shown in Fig. 8, RF@AgNCs do not show obvious

influence on the proliferation of either HUVE cells or HK-2 cells after 24 h and 48 h incubations, respectively. Therefore, the results demonstrate that RF@AgNCs have good biocompatibility with human blood cells and mammalian cells.

4. Conclusions

In summary, we used riboflavin as biocompatible surface ligand to synthesize ultrasmall silver nanoclusters (RF@AgNCs). The RF@AgNCs exhibited very good antibacterial properties against *S. aureus*, *E. coli*, and *C. albicans*. The antibacterial mechanism studies showed that the integrity of bacteria cell membranes was disrupted and a large amount of intracellular ROS was produced after exposure to RF@AgNCs. Significantly, RF@AgNCs maintained intrinsic good biocompatibility with red blood cells and mammalian cells, making these systems promising for antimicrobial therapeutic use. With the efficient antimicrobial effect and good or acceptable biocompatibility,



RF@AgNCs would have great potential in biomedical applications as well as other fields such as textile industry.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 M. Rai, A. Yadav and A. Gade, *Biotechnol. Adv.*, 2009, **27**, 76–83.
- 2 W. R. Li, X. B. Xie, Q. S. Shi, H. Y. Zeng, Y. S. Ou-Yang and Y. B. Chen, *Appl. Microbiol. Biotechnol.*, 2010, **85**, 1115–1122.
- 3 V. K. Sharma, R. A. Yngard and Y. Lin, *Adv. Colloid Interface Sci.*, 2009, **145**, 83–96.
- 4 G. Franci, A. Falanga, S. Galdiero, L. Palomba, M. Rai, G. Morelli and M. Galdiero, *Molecules*, 2015, **20**, 8856–8874.
- 5 I. V. Vreck, I. Zuntar, R. Petlevski, I. Pavicic, M. D. Sikiric, M. Curlin and W. Goessler, *Environ. Toxicol.*, 2016, **31**, 679–692.
- 6 S. Chernousova and M. Eppe, *Angew. Chem., Int. Ed.*, 2013, **52**, 1636–1653.
- 7 M. Ovais, I. Ahmad, A. T. Khalil, S. Mukherjee, R. Javed, M. Ayaz, A. Raza and Z. K. Shinwari, *Appl. Microbiol. Biotechnol.*, 2018, **102**, 4305–4318.
- 8 S. B. Seo, S. H. S. Dananjaya, C. Nikapitiya, B. K. Park, R. Gooneratne, T. Y. Kim, J. Lee, C. H. Kim and M. De Zoysa, *Fish Shellfish Immunol.*, 2017, **68**, 536–545.
- 9 J. Tian, K. K. Y. Wong, C. M. Ho, C. N. Lok, W. Y. Yu, C. M. Che, J. F. Chiu and P. K. H. Tam, *ChemMedChem*, 2007, **2**, 129–136.
- 10 M. K. Rai, S. D. Deshmukh, A. P. Ingle and A. K. Gade, *J. Appl. Microbiol.*, 2012, **112**, 841–852.
- 11 R. K. Kunkalekar, M. M. Naik, S. K. Dubey and A. V. Salker, *J. Chem. Technol. Biotechnol.*, 2013, **88**, 873–877.
- 12 T. C. Dakal, A. Kumar, R. S. Majumdar and V. Yadav, *Front. Microbiol.*, 2016, **7**, 17.
- 13 A. Panacek, L. Kvitek, R. Prucek, M. Kolar, R. Vecerova, N. Pizurova, V. K. Sharma, T. Nevecna and R. Zboril, *J. Phys. Chem. B*, 2006, **110**, 16248–16253.
- 14 C. Carlson, S. M. Hussain, A. M. Schrand, L. K. Braydich-Stolle, K. L. Hess, R. L. Jones and J. J. Schlager, *J. Phys. Chem. B*, 2008, **112**, 13608–13619.
- 15 S. Pal, Y. K. Tak and J. M. Song, *Appl. Environ. Microbiol.*, 2007, **73**, 1712–1720.
- 16 A. M. El Badawy, R. G. Silva, B. Morris, K. G. Scheckel, M. T. Suidan and T. M. Tolaymat, *Environ. Sci. Technol.*, 2011, **45**, 283–287.
- 17 S. H. Tang and J. Zheng, *Adv. Healthcare Mater.*, 2018, **7**, 10.
- 18 G. A. Martinez-Castanon, N. Nino-Martinez, F. Martinez-Gutierrez, J. R. Martinez-Mendoza and F. Ruiz, *J. Nanopart. Res.*, 2008, **10**, 1343–1348.
- 19 K. Y. Zheng, M. I. Setyawati, T. P. Lim, D. T. Leong and J. P. Xie, *ACS Nano*, 2016, **10**, 7934–7942.
- 20 J. C. Jin, X. J. Wu, J. Xu, B. B. Wang, F. L. Jiang and Y. Liu, *Biomater. Sci.*, 2017, **5**, 247–257.
- 21 K. W. H. Kwok, W. Dong, S. M. Marinakos, J. Liu, A. Chilkoti, M. R. Wiesner, M. Chernick and D. E. Hinton, *Nanotoxicology*, 2016, **10**, 1306–1317.
- 22 X. Y. Yang, A. P. Gondikas, S. M. Marinakos, M. Auffan, J. Liu, H. Hsu-Kim and J. N. Meyer, *Environ. Sci. Technol.*, 2012, **46**, 1119–1127.
- 23 A. K. Suresh, D. A. Pelletier, W. Wang, J. W. Moon, B. H. Gu, N. P. Mortensen, D. P. Allison, D. C. Joy, T. J. Phelps and M. J. Doktycz, *Environ. Sci. Technol.*, 2010, **44**, 5210–5215.
- 24 Y. M. Long, L. G. Hu, X. T. Yan, X. C. Zhao, Q. F. Zhou, Y. Cai and G. B. Jiang, *Int. J. Nanomed.*, 2017, **12**, 3193–3206.
- 25 W. T. Lu, D. Senapati, S. G. Wang, O. Tovmachenko, A. K. Singh, H. T. Yu and P. C. Ray, *Chem. Phys. Lett.*, 2010, **487**, 92–96.
- 26 F. Liu, F. F. Wei, L. Wang, H. Liu, X. P. Zhu and Y. C. Liang, *Physiol. Mol. Plant Pathol.*, 2010, **74**, 330–336.
- 27 H. Boubakri, J. Chong, A. Poutaraud, C. Schmitt, C. Bertsch, A. Mliki, J. E. Masson and I. Soustre-Gacougnolle, *Eur. J. Plant Pathol.*, 2013, **136**, 837–855.
- 28 R. P. Goodrich, *Vox Sang.*, 2000, **78**, 211–215.
- 29 K. Makdoui, A. Backman, J. Mortensen and S. Crafoord, *Graefes Arch. Clin. Exp. Ophthalmol.*, 2010, **248**, 207–212.
- 30 B. Roy, P. Bairi and A. K. Nandi, *Analyst*, 2011, **136**, 3605–3607.
- 31 M. Svecova, P. Ulbrich, M. Dendisova and P. Matejka, *Spectrochim. Acta, Part A*, 2018, **195**, 236–245.
- 32 Y. Q. Li, Y. X. Zhu and Y. Zhang, *Chem. J. Chin. Univ.*, 2008, **29**, 669–672.
- 33 O. Choi, K. K. Deng, N. J. Kim, L. Ross, R. Y. Surampalli and Z. Q. Hu, *Water Res.*, 2008, **42**, 3066–3074.
- 34 G. Applerot, A. Lipovsky, R. Dror, N. Perkash, Y. Nitzan, R. Lubart and A. Gedanken, *Adv. Funct. Mater.*, 2009, **19**, 842–852.
- 35 L. S. A. de Melo, A. S. L. Gomes and S. Saska, *J. Fluoresc.*, 2012, **22**, 1633–1638.
- 36 R. Aiello, D. Castrogiovanni and J. Parisi, *Photochem. Photobiol.*, 2018, **94**, 1159–1166.

