

Cite this: *RSC Adv.*, 2019, 9, 17457

Gut microbiota: a new angle for traditional herbal medicine research

Longfei Lin,^{ID}†^a Liyu Luo,†^c Ming Zhong,^d Tanggui Xie,^d Yuling Liu,^{ID}^a Hui Li*^a and Jian Ni*^b

Traditional Herbal Medicine (THM) has been used for thousands of years, and is popular worldwide due to its effectiveness in a variety of diseases. THM has also formed the basis of the discovery of modern drugs like artemisinin and paclitaxel. However, at present, studies that focus on development in the field of THM are stagnant because currently, the effective ingredients in the herbal formulations and the ambiguity of the underlying mechanisms of action are unknown. In this review, we have investigated the studies available that focused on the efficacy, active ingredients and bioavailability of THM, and the function of gut microbiota in THM-mediated treatment of disease. We hypothesized that most THMs treat diseases via three mechanisms: (1) metabolizing into active metabolites by the action of gut microbiota, (2) regulation of gut microbiota balance, and (3) regulating the fermentation products of the gut microbes. Therefore, focusing on these aspects can help elucidate the pharmacodynamic constituents of THM preparations, and their therapeutic mechanisms of action.

Received 11th March 2019

Accepted 21st May 2019

DOI: 10.1039/c9ra01838g

rsc.li/rsc-advances

1. The therapeutic efficacy and spectrum of THM

In China, herbs have been used for thousands of years to treat various disorders, and form the basis of a theoretical and practical system for the treatment and prevention of diseases. Traditional Herbal Medicine (THM) is a fully institutionalized part of Chinese health care that is extensively used along with Western medicine. An estimated 40% of health care in China is based on THM, and 75% of health centers have a THM department.^{1,2} According to the China Food and Drug Administration (CFDA), 8760 pharmaceutical manufacturers currently market THM in China, and thereby use a total of 59 027 types of herbs. THM is also widely used in other countries, and by 20% to 25% of the population in Europe.³ THM is categorized in European countries along with Asiatic medicines, and mainly includes depurative plants used for the treatment of dermatological illnesses or hay baths for cancer treatment.⁴ European countries spend almost \$5 billion on over-the-counter herbal medicines; Germany accounted for 40% of the expenses.⁵ In the United States (US), alternative medicine, including herbs

among others, has also gained popularity in routine health care.⁶ In retail pharmacy in the US, the sale of herbal medicines has grown by approximately 20% annually, with nearly 20% of the population using herbal preparations.^{7,8} THM has also been widely used to treat diseases, such as malaria, diabetes mellitus, and hypertensive in Africa,^{9,10} as well as in an Indian traditional medicinal system, such as Ayurveda, which is a comprehensive scientific medicinal system, including *Charaka Samhita*, *Susruta Samhita*, *Madhava Nidana*, *et al.*¹¹

In general, the therapeutic efficiency of THM is not supported by clinical evidence. Due to the increasing usage and popularity of THM, several studies have been undertaken in recent years to clinically evaluate THM.¹² In 1986, it was showed that ginseng could improve certain psychomotor functions in healthy subjects.¹³ A randomized, double-blind, placebo-controlled clinical study was performed to indicate that daily consumption of *Lycium barbarum* for 14 days could improve neurological performance and gastrointestinal function.¹⁴ The biminne formulation is composed of 11 traditional Chinese herbs, and after 1 year of treatment was demonstrated to be safe and effective in relieving the symptoms of perennial allergic rhinitis.¹⁵ A Chinese herbal formulation improved the symptoms of irritable bowel syndrome in some patients, as proven by a randomized controlled trial in which 116 patients were enrolled.¹⁶ The efficiency of this formula was also verified by another randomized placebo-controlled trial.¹⁷ “Ba Wei Di Huang Wan” is a Japanese concoction of eight herbs that has successfully been used as a remedy for the elderly who present with cognitive decline and disability, and was shown to improve cognitive and physical functioning in a randomized trial in

^aInstitute Chinese Materia Medica, China Academy of Chinese Medical Sciences, Beijing, China

^bSchool of Chinese Material Medica, Beijing University of Chinese Medicine, Beijing, China

^cSchool of Pharmaceutical Science and Technology, Tianjin University, Tianjin, China

^dGuangxi Key Laboratory of Traditional Chinese Medicine Quality Standards, Guangxi Institute of Chinese Medicine and Pharmaceutical Science, Nanning 530022, China

† These two authors contributed equally.



patients with dementia.¹⁸ Many studies have been performed that focus on the clinical efficacy of THM, and all of them cannot be enumerated here.

Taken together, THM is used worldwide, especially in China, and is used as an alternative medicine in the US and Europe. Although some THM preparations have shown clinical evidence, the treatment mechanism is still unclear.

2. Active ingredients or inactive ingredients?

There are hundreds of ingredients in a THM preparation, which can be classified as either active or inactive. The World Health Organization (WHO) defines active ingredients of THM as those “with therapeutic activity.” For herbal medicines with known active ingredients, their preparation should be standardized to allow for a defined amount of these ingredients, if adequate analytical methods are available. In any case it is not possible to identify active ingredients, the complete preparation may be considered as an active ingredient.¹⁹ Inactive ingredients are those without detectable physiological activity and clinical effect. Pharmaceutically active ingredients, either one or more compounds, are used to evaluate THM, including Chinese herb medicine, Chinese patent medicine, and Traditional Chinese Medicine (TCM), regarding their therapeutic activity.²⁰ Although few active ingredients do not accurately represent the quality of TCM, the CFDA uses a single or multiple components as the quality control indicator(s) for most TCM formulations. Several techniques, including chemical fingerprinting, gas chromatography-flame ionization detection (GC-FID), and high-performance liquid chromatography-mass spectrometry, (HPLC-MS)^{21,22} can be used to analyze TCM composition, and help in quality control. However, due to the complex composition of THM not every component is isolated, therefore the

biological activity cannot be studied. The *Chinese Pharmacopoeia* part one 2015 edition includes information of 2598 species used in Chinese herb medicine and Chinese patent medicine, with information on less than 1% quality indicator components (Table 1). The question arises as to whether these few indicators are fully representative of the efficacy of the herbs, and if perhaps unknown components may in fact represent the inactive ingredients. For example, the active ingredient of one of the listed herbs, *Cacumen platycladi*, which is used for treating epistaxis, hematemesis, hemoptysis, hematochezia, abnormal uterine bleeding, hair loss, and premature greying, is quercetin. Quercetin is a flavonoid that inhibits the ferrous ion-dependent lipid peroxidation of lecithin liposomes,²³ and shows antioxidant and anti-cancer effects by inhibiting protein kinases.^{24,25} However, to our knowledge, no association has been reported between quercetin and epistaxis, hematemesis, hemoptysis, and hematochezia. Furthermore, the recommended dose of *C. platycladi* is 12 g, with an active dose of quercetin 12 mg (0.1%), which may or may not be the clinically effective dose.

Patent herbal medicine is produced after extraction, concentration (refinement), and drying of THM, with higher daily dosage requirement compared to Western medicine. Ganmao Qingre granules, also described in the *Chinese Pharmacopoeia*, are used to treat colds, fever, cough, and pharyngalgia. Its dosage is described as “oral or boiled in water, 12 g twice a day”, and its active ingredient is puerarin, which is present at only one thousandth (~10 mg) of the final formulation (Table 2). The pharmacological properties of puerarin include vasodilation, cardio-protection, neuroprotection, anti-cancer, anti-inflammation, analgesia, bone formation, inhibition of alcohol intake, inhibition of oxidative stress and apoptosis, and attenuation of insulin resistance.^{26,27} Several studies on puerarin have indicated that alleviating colds is not its main pharmacological effect. Taken together, most of the

Table 1 Chemical markers shared by various herbal medicines in the *Chinese Pharmacopoeia* 2015 edition

Traditional herbal medicine	Dosage per day	Quality indicator component	Content limit \geq
Radix ginseng	3–9 g	Ginsenoside Rb1 Ginsenoside Rg1, Ginsenoside Re	0.18% Sum 0.27%
<i>Schisandra chinensis</i> fructus	2–6 g	Schisandrin	0.40%
Plantaginis semen	9–15 g	Geniposide acid Verbascoside	0.40% 0.30%
Radix scrophulariae	9–15 g	Harpagide, harpagoside	Sum 0.45%
Cassiae semen	9–15 g	Chrysophanol Aurantio-obtusin	0.12% 0.080%
Radix saposchnikovia	5–10 g	Prim- <i>o</i> -glucosylcimifugin, 5- <i>o</i> -methyl Weiss Amie alcohol glycoside	Sum 0.24%
Forsythiae fructus	6–15 g	Forsythin Forsythoside A	0.15% 0.25%
<i>Lonicerae japonicae</i> caulis	9–30 g	Chlorogenic acid Loganin	0.070% 0.10%
Cacumen platycladi	6–12 g	Quercetin	0.10%
Radix rubiae	6–10 g	Rubimaillin Purpurin	0.20% 0.080%
<i>Curcuma longae</i> rhizoma	3–10 g	Curcumin	0.90%
Radix astragali	9–30 g	Astragaloside IV Genistein glucose	0.040% 0.020%



Table 2 Chemical markers shared by various patent herbal medicine in the *Chinese Pharmacopoeia* 2015 edition

Name	Indication	Specification	Dosage	Quality indicator component	Content limit \geq
Ganmao Qingre granules	Colds, fever, cough, and pharyngalgia	12 g per pouch	Once pouch and twice a day	Puerarin	10 mg per pouch
Huoxiang Zhengqi water	Heatstroke, gastrointestinal cold	10 mL per bottle	5–10 mL per time and twice a day	Hesperidin	0.18 mg mL
Niuhuang Shangqing tablets	Headache, dizziness, sore throats, aphtha, swollen gums, constipation	0.265 g per tablets	4 Tablets per time, two times a day	Baicalin Jasminoidin	1.1 mg per tablet 0.45 mg per tablet
Weikang Ling capsules	Acute and chronic gastritis, duodenal ulcer, gastrophelcosis, gastrorrhagia	0.4 g per capsule	4 Capsules and three times a day	Paeoniflorin	1.0 mg per capsule
Hugan tablets	Chronic hepatitis and early cirrhosis, reduced aminotransferase	0.36 g per tablet	4 Tablets per time, 3 times a day	Schisandrin	0.28 mg per tablet
Lidan Paishi granules	Cholecystitis, gallstone	3 g per pouch	Removing urinary calculus: two pouchs per time, twice a day; diminish inflammation: one pouch per time, twice a day	Emodin	3.0 mg per pouch
Fufang Danshen tablets	Coronary disease, angina pectoris	(1) 0.32 g per tablet (2) Sugar-coated tablet (3) 0.8 g per tablet	3 Tablets (1, 2) or 1 tablet (3) per time, 3 times a day	Ginsenoside Rg ₁ , Rb ₁ , Re, and notoginsenoside R ₁	(1, 2): Sum 6.0 mg per tablet (3): Sum 18.0 mg per tablet

components in a patent medicine are still unknown, and therefore the main therapeutic substances and their pharmacological effects remain unclear.

3. Low bioavailability of THM

THM formulations are complex, often composed of hundreds or even thousands of ingredients, such as flavonoids, alkaloids, saponins, anthraquinones, tannins, lignin, and terpenoids. The active ingredients in THM have low absorption and bioavailability *via* the oral route, which is significant since the oral route is the usual mode of THM administration. The main reasons for their low bioavailability include the following:²⁸ (1) low levels of the active ingredient due to the relatively crude extraction and purification process. (2) Poor solubility, lipid solubility, permeability, and gastrointestinal stability, and improper molecular size of the active substances. For example, water soluble phenolics are unable to cross the lipid membranes of intestinal cells,²⁹ and molecules larger than 0.4 nm in diameter have difficulty in passing through the aqueous channels in the intestinal epithelium.²⁹ (3) Drug–drug interactions (DDIs) of individual THM components, which are essential for its therapeutic effects.³⁰ These components may be dual or multi substrates of drug efflux transporters, such as P-glycoprotein (P-gp), multidrug resistance associated protein (MRP), cytochrome P450 (CYP450) and others.^{31–33} Thus, these specificity multi substrates of P-gp, MRP and CYP450 might be results of the low bioavailability of THM.

Given its successful use worldwide, evaluation of the pharmacokinetics and bioavailability of THM is critical for assessing

their safety and efficacy.³⁴ Sophisticated clinical pharmacokinetic and bioavailability evaluations in recent years (Table 3).^{35–42} indicated that most THM are not absorbed *via* the circulation. With increasing evidence implicating gut microbiota in human health and disease, it is highly likely that the therapeutic effects of THM are related to the gut microbiota.

4. Gut microbiota in human health and disease

The gut microbiome is very important for human health and is often considered a “hidden organ”. Recent estimates have suggested that the microbes residing in or on human body slightly exceed the number of human cells, which is approximately 1.3 : 1.^{43–45} The total number of genes encoded by the gut bacteria is at least 150 times that of the human genome.^{46,47} The ensemble of genetic material retrieved from the gut known as the “microbial group”, is called the “gut meta-genome”.⁴⁸ The human body can be considered a “super-organism” made of both eukaryotic cells and symbiotic microbes.⁴⁹ Under normal circumstances, human tissues and bacteria interact and maintain a functional balance, in addition to forming biological, chemical, mechanical, and immune barriers against invading pathogens.⁵⁰ The gut meta-genome and the host genome influence pathophysiological conditions and the drug metabolism.^{51,52} In 2007, the National Microbiology Program was launched by the National Institutes of Health (NIH) for human health assessment and surveillance, and early diagnosis and treatment of chronic diseases.^{46,48} Previous studies have shown that the gut microbiota closely interact with the gastrointestinal



Table 3 Pharmacokinetic research of THM in the clinic^a

Medicinal	Dosage	Samples	Target compound	Contents in extract or dosage	T_{max} (h)	C_{max} (ng mL ⁻¹)	$T_{1/2}$ (h)	AUC (ng h mL ⁻¹)
Rhei undulati rhizoma	100 mg kg ⁻¹	12 Healthy volunteers (9 men/3 women)	Rhein	12.308 mg g ⁻¹	59.69 ± 28.63 min	2.79 ± 1.02 µg mL	3.3759 ± 0.335	9746.689 ± 1011.72
Rhubarb	50 mg kg ⁻¹	Eight male volunteers	Rhein	22.8%	3.7 ± 0.6 h	37 ± 7.8	199.66 ± 63.50 min	18.64 ± 5.67 µg
Milk thistle (<i>Silybum marianum</i>) thistle extracts	600 mg milk	3 Healthy volunteers	Silychristin	8.93%	2.8 ± 1.3	21 ± 3.1	7.9 ± 1.7	401 ± 93
			Silydianin	22.0%	1.0 ± 0.4	23 ± 7.9	3.3 ± 0.6	140 ± 47
			Silybin A	34.0%	1.3 ± 0.3	131 ± 28	1.3 ± 0.5	63 ± 38
			Silybin B	8.59%	1.7 ± 0.3	113 ± 30	3.4 ± 0.8	647 ± 234
			Isosilybin A	3.67%	1.5 ± 0.0	59 ± 16	4.1 ± 1.3	799 ± 403
			Isosilybin B				4.0 ± 1.1	383 ± 195
Artichoke leaf extracts	2.4 g	14 Healthy volunteers	Caffeic acid	107.0 mg	0.83 ± 0.3	6.51 ± 1.89	3.08 ± 1.53	23.02 ± 7.15
Geriaforce™	0.625 g	14 Healthy volunteers	Caffeic acid	153.8 mg	0.94 ± 0.47	8.04 ± 2.64	2.69 ± 1.80	26.32 ± 10.69
Geriaforce™ tincture	90 drops, 2.73 mL	6 Healthy volunteers	Bilobalide	646.93 µg	1.09 ± 0.46	6.17 ± 5.67	2.33 ± 1.04	26.53 ± 26
			Ginkgolide A	298.14 µg	0.92 ± 0.38	3.81 ± 0.96	2.31 ± 0.66	12.42 ± 5.27
			Ginkgolide B	147.45 µg	0.88 ± 0.38	1.45 ± 0.98	2.34 ± 0.81	6.17 ± 3.77
Ginkgo fresh plant extract tablets	4 Tablets	6 Healthy volunteers	Bilobalide	1974.96 µg	0.88 ± 0.3	11.6 ± 3.77	2.52 ± 1.7	41.56 ± 31.94
			Ginkgolide A	881.52 µg	0.80 ± 0.34	7.27 ± 3.83	2.66 ± 2.05	29.75 ± 29.64
			Ginkgolide B	524.56 µg	2.35 ± 2.27	4.61 ± 2.87	4.01 ± 2.58	43.48 ± 27.64
EGB 761™	3 Tablets	6 Healthy volunteers	Bilobalide	3672.39 µg	1.06 ± 0.4	29.87 ± 18.42	2.08 ± 0.23	100.57 ± 38.91
			Ginkgolide A	1571.37 µg	0.93 ± 0.43	16.97 ± 5.79	3.6 ± 2.96	80.83 ± 57.47
			Ginkgolide B	836.46 µg	2.0 ± 1.63	10.9 ± 5.59	3.94 ± 1.42	71.73 ± 26.98
Cardiotonic pills	30 Pills	6 Male healthy volunteers	Tanshinol	160 µg per pill	1.33 ± 0.52	19.3 ± 5.1 ng	1.16 ± 0.42	48.5 ± 3.4
		6 Female healthy volunteers	Tanshinol	160 µg per pill	1.17 ± 0.68	31.3 ± 10.6	1.27 ± 0.21	77.6 ± 31.6
Safflower oral solution	Single dose	12 Healthy volunteers	Hydroxysafflor yellow A	140 mg	1 h	73.5 ± 8.7	2.6–3.5	
American ginseng powder	10 g	6 Healthy volunteers	Ginsenoside Rb ₁	31.7 mg g ⁻¹	4 h	19.90 ± 5.43		155.0 ± 19.5

^a T_{max} : peak time of component concentration *in vivo*; C_{max} : maximum concentration; $T_{1/2}$: eliminate half-time; "AUC": area under the curve.

tract, liver, skin, and central nervous system among other organs, and aid in physiological functions, such as digestion and absorption of nutrients, neurodevelopment and transmission, fat metabolism, and immune responses.^{53–56} Therefore, any disturbance in the gut microbiota may lead to various disorders, including metabolic diseases (obesity, diabetes), cardiovascular disease (hypertension), and mental disorders (depression, anxiety, cognitive decline, Parkinson's Disease).^{57–62}

The gut microbiota is thought to be comprised of more than 1000 various microbial species. Due to various reasons, the microbial composition can alter throughout their lifespan, therefore, its composition is not static and changes to its structure are dynamic, and overall gut enterotypes may vary from person to person.^{46,63} The Meta-genomics of Human Intestinal Tract (MetaHIT)⁶⁴ study identified nine bacteriophyta in the human intestine, including Firmicutes, Bacteroidetes, Proteobacteria, Actinobacteria, Fusobacteria, Verrucomicrobia, Cyanobacteria, Spirochaetes, and VadinBE97.^{65–67} Based on their relationship with the host, gut microbiota can be divided into the following three categories: (1) physiological bacteria that are beneficial to the host, including *Bifidobacterium* and *Lactobacillus*, (2) conditional pathogen or neutral bacteria, including *Enterococcus* and *Escherichia coli*, and (3) pathogenic bacteria, including *Proteus*, *Pseudomonas*, and *Staphylococcus aureus*.⁶⁸ Gut microbiota disorders can lead to metabolic disorders of carbohydrates, lipids, and other substances in the host, and thereby enhance inflammatory reactions and oxidative stress in the body, resulting in promoting the occurrence or deterioration of cardiovascular diseases.^{69–71} Gut microbiota can participate in bidirectional regulation of intestinal and central nervous system (CNS) through neurotransmitter, endocrine, immune and metabolic pathways. When the gut microbiota is disordered, it can affect the development of CNS diseases, including cerebral ischemia, Parkinson's disease, Alzheimer's disease, disseminated sclerosis, hepatic encephalopathy, and psychogenia.^{72–74} The intestinal epithelial system is the natural barrier of intestinal exogenous pathogens. In addition, gut microbiota disorders can increase intestinal permeability, and promote intestinal pathogens and intestinal toxins to enter the liver through the portal system, then will cause a series of pathophysiological reactions. The mechanism of action involved alterations in gut epithelial permeability, choline metabolism, endogenous alcohol production, release of inflammatory cytokines, regulation of hepatic Toll-like receptor (TLR), and bile acid metabolism.^{75,76} In addition, gut microbiota is also associated with a variety of diseases, such as diabetes, obesity, rheumatoid arthritis, inflammatory bowel disease (Fig. 1 and Table 4).

5. Therapeutic effects of THM through gut microbiota

In the circulation after oral administration, the concentration of active THM components is usually low. This opens the

possibility of THM acting *via* intestinal absorption, with considerable influence on the gut microbiota.

5.1 Metabolic effect of gut microbiota on THM

5.1.1 The main metabolic enzyme and bacterial strains in gut microbiota. Gut microbiota can produce many enzymes, such as β -D-glucuronidase, β -D-glucosidase, β -galactosidase, nitroreductase, azoreductase, α -L-rhamnosidase, and β -xylosidase. These enzymes are produced by different bacterial strains and metabolize the chemical components of THM.

β -D-glucosidase mainly plays a role in cleavage of β -glycoside bonds (β -1,1; β -1,2; β -1,3; β -1,6), and due to its poor specificity, the C–O glycoside bond, C–S bond, C–N bond, and C–F bond can also be cleaved by β -D-glucosidase. β -D-glucosidase is involved in the glycolysis pathway and is highly expressed in *Bifidobacteria* and *Bacteroides thetaiotaomicron*.^{106,107} β -D-glucuronidase, an important phase II metabolic enzyme, cleaved the β -1,4-glucuronide bond, and its activity was only observed in Firmicutes, such as *Lactobacillus*, *Enterococcus*, *Clostridium paraputrificum*, *B. fragilis*, and *B. uniformis*.¹⁰⁸ β -Xylosidase is an exonuclease that mainly hydrolyzes xyloside, which could be hydrolyzed to xylooligosaccharide from the non-reducing end in an exo-cutting manner, and the hydrolyzed product is xylose. β -Xylosidase exists in *Bifidobacterium*, *Clostridium stercorarium*, and *Thermoanaerobacterium saccharolyticum*.^{109–111} β -Galactosidase can catalyze and hydrolyze lactose to produce glucose and galactose, and catalyzes the galactosyl transfer reaction to form galactooligosaccharides. The activity of β -galactosidase is closely related to the composition of the gut microbiota, and the higher the activity of β -galactosidase when probiotics were dominantly present in intestinal flora.¹¹² β -Galactosidase is mainly produced by probiotics, such as *Bifidobacterium*, *Lactobacillus acidophilus*, and *Streptococcus thermophiles*.^{112,113} Nitroreductase is an NAD(P)H-dependent flavoenzyme, which can reduce the nitrogroup into amine or amino, and catalyze the reduction reaction of nitroaromatic compounds into aromatic amine. The metabolites can be used as a nitrogen source for bacterial growth. The activity of nitroreductase can be found in various intestinal bacteria, such as *Lactobacillus*, *Bifidobacterium*, and *Escherichia coli*.^{114–116}

In addition to the above-mentioned enzymes, many other metabolic enzymes are present in the gut, including α -L-rhamnosidase, azoreductase, and mannosidase. Most of these are produced by intestinal probiotics, such as *Bifidobacterium*, and produce α / β -galactosidase, α / β -glucosidase, β -fructofuranosidase, mannosidase, D-xylanase, D-xylose enzyme.¹¹³ *Bifidobacterium* has been shown to adjust intestinal dysfunction, improve human immunity, inhibit pathogen growth, and have anti-inflammatory, and anti-aging effects.^{117,118} *Bacteroides thetaiotaomicron* is one of the largest bacteria in the gut that can express β -glucosidase, β -galactosidase, β -fructofuranosidase, β -mannosidase, α -mannanase, and arabinase.¹¹⁹

Chemical constituents in THM are degraded and metabolized in the gut by the above-mentioned enzymes. Rutin has two metabolic pathways, which mainly work through *Bifidobacterium* in the gut; one was first hydrolyzed into isoquercetin by the



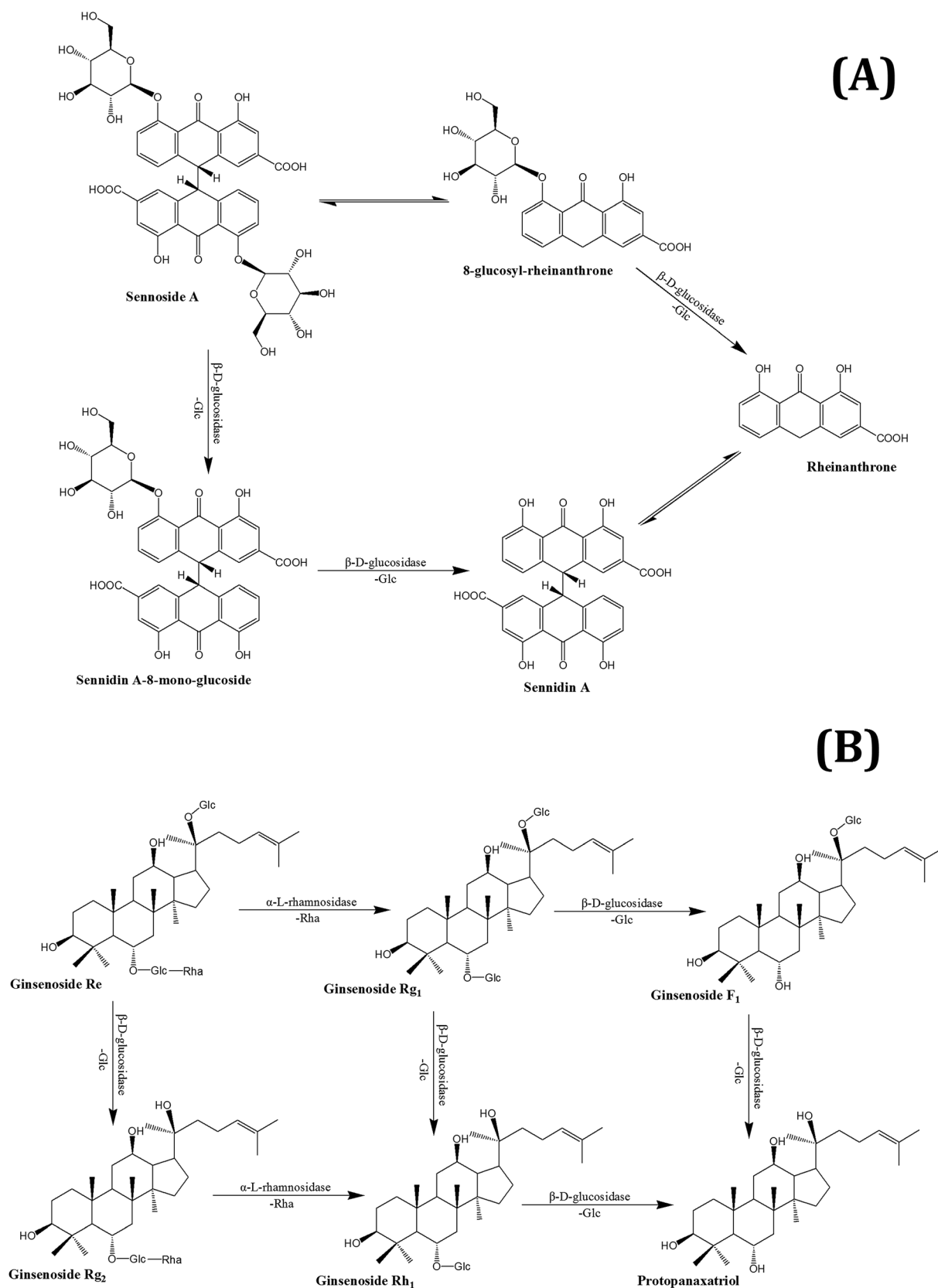


Fig. 1 The metabolism pathways of sennoside A (A) and ginsenoside Re (B) by gut microbiota.



Table 4 Changes in gut microbiota associated with disease

Body system	Disease	Related gut microbiota	References
Metabolic system	Hepatocirrhosis	<i>Veillonella atypica</i> , <i>Streptococcus salivarius</i>	77
	Liver cancer	<i>Helicobacter hepaticus</i>	78
	Type 2 diabetes	<i>Akkermansia muciniphila</i>	79
	NAFLD	Lachnospiraceae bacterium 609, <i>Barnesiella intestinihominis</i>	80
Alimentary system	Obesity	<i>Enterobacter cloacae</i> B29	81
	Diarrhea	<i>Escherichia/Shigella</i> , <i>Granulicatella</i>	82
	Irritable bowel syndrome	<i>Veillonella</i> , <i>Lactobacillus</i> , <i>Prevotella</i> , <i>Bacteroides</i>	83 and 84
	Inflammatory bowel disease	<i>Roseburia</i> , Enterobacteriaceae, Ruminococcaceae, <i>Campylobacter</i> , <i>Salmonella</i>	83 and 85
Immune system	Colorectal cancer	<i>Bacteroides fragilis</i> , <i>Streptococcus gallolyticus</i>	86 and 87
	Paradentitis	<i>Porphyromonas gingivalis</i>	88 and 89
	Rheumatic arthritis	<i>Prevotella copri</i> , <i>Lactobacillus</i> , <i>Tropheryma whippelii</i>	90 and 91
Respiratory system	Systemic lupus erythematosus	Enterobacteriaceae, Firmicutes/Bacteroidetes	92 and 93
	Asthma	<i>Aspergillus fumigatus</i>	94
Circulatory system	Chronic bronchitis	<i>Clostridium perfringens</i> , <i>Enterobacter</i> , <i>Enterococcus</i>	95
	Atherosclerosis	<i>Collinsella</i> , <i>Eubacterium</i> , <i>Roseburia</i>	96
	Hyperlipemia	<i>Lactobacillus</i>	97
Nervous system	Hypertension	<i>Bacteroides acidifaciens</i>	98
	Depression	Spore-forming bacteria	99
	Anxiety neurosis	<i>Campylobacter jejuni</i> , <i>Citrobacter rodentium</i>	100 and 101
	Autism	<i>Bacteroides fragilis</i>	102
	Parkinson's disease	Enterobacteriaceae, <i>Escherichia coli</i>	103
	Multiple sclerosis	<i>Methanobrevibacter</i> , <i>Akkermansia</i>	104
	Alcohol dependence	<i>Ruminococcus</i> , <i>Faecalibacterium</i> , <i>Subdoligranulum</i> , <i>Oscillibacter</i> , <i>Anaerofilum</i> , <i>Dorea</i> , <i>Blautia</i>	105

enzyme α -L-rhamnosidase, and further hydrolyzed to quercetin by β -D-glucosidase. The other was converted to leucocyanidin under the action of α -L-rhamnosidase and β -D-glucosidase.^{120,121} Under the action of α -L-rhamnosidase and β -D-glucosidase that are expressed by gut microbiota, ginsenoside Re can be converted into Rh1, F1, and a small amount of protopanaxadiol. There are also two metabolic pathways of sennoside in the intestinal flora. The first involves the hydrolysis of senna into senna glycoside by β -D-glucosidase, which is then reduced to form rheinanthrone, the other involves reduction of sennoside to 8-glucosyl-rheinanthrone, which is then hydrolyzed by β -D-glucosidase to rheinanthrone.¹²² Geniposide is hydrolyzed by β -D-glucuronidase of intestinal bacteria to genipin.¹²³ Paeoniflorin produced deglucose glucosides, and 7S- and 7R-paeonimetalines I and II under the action of β -D-glucosidase, β -D-glucocoeesterase and other enzymes, which are produced by *Bacteroides fragilis*, *Peptostreptococcus*, and *Lactobacillus brevis*.^{124,125} The metabolic pathways of several THM compounds in the gut are shown in Fig. 1.

5.1.2 The better bioactivity metabolites of THM in gut. During growth and reproduction, gut microbiota produce several enzymes, including glycosidases, nitroreductase, azo reductase, and various carbohydrate hydrolyzing enzymes,^{113,126,127} which can degrade THM components. The active components of THM are transformed into new active metabolites by the action of specific microbial enzymes, which exert different biological effects.

Saponins: Saponins have antipyretic, sedative, anti-cancer and other pharmacological effects, and are commonly present in *ginseng*, *Panax notoginseng* and *Bupleurum*. Ginsenosides are

the main active constituents of *P. ginseng*, and are metabolized to compound K (20-O-(β -D-glucopyranosyl)-20(S)-protopanaxadiol) by gut microbiota before absorption into the circulation. Compound K has hepato-protective, anti-cancer, anti-wrinkling, and anti-aging properties, and is more potent than the parental ginsenosides.¹²⁸⁻¹³⁰ Ginsenosides can be metabolized by *Bacteroides* JY-6, *Bifidobacterium* K506, *Eubacterium* A-44, *Prevotella oris*, and *Fusobacterium* K-60.^{131,132} Ginsenoside Re can be converted into Rh1, which exerts a strong estrogenic effect. Ginsenoside Rh1 increased the proliferation of MCF-7 cells 2.1-fold at a concentration of 1 mM, however levels of ginsenoside Re were not significantly increased when compared with the blank control group.¹³³ Furthermore, Ardipusilloside-I is a triterpenoid saponin isolated from *Ardisia pusilla* DC, and has potent anti-tumor activity. Ardipusilloside-I is de-glycosylated into four compounds by the action of bacterial enzymes, two of them have the same anti-tumor capacity as Ardipusilloside-I.¹³⁴ Glycyrrhizin is difficult to be absorbed in the intestine. It interacts with intestinal bacteria to produce glycyrrhetic acid, which was absorbed by the body and showed stronger anti-hepatotoxicity activity when compared with glycyrrhizin.¹³⁵

Flavonoids: Flavonoids are the active ingredients of various THM formulations. Baicalin, a flavone glycoside, can be de-glycosylated into baicalein or methylated into aglycon orohylin A by gut microbes. Baicalein and orohylin A have been shown to improve antihistamine-induced pruritus responses in mice when compared to baicalin, and compared to baicalin inhibited the contraction of guinea pig ileum induced by histamine (with 50% inhibitory concentration of 0.28 mmol



L^{-1}).¹³⁶ Poncirin is the active constituent of *Poncirus trifoliata*, and is metabolized by gut microbiota to ponciretin both *in vitro* and *in vivo*. The anti-inflammatory effect of ponciretin is superior to that of poncirin, and can attenuate colitis by inhibiting lipopolysaccharide (LPS) binding to TLR4 on macrophages, and restore the Th17/Treg balance.¹³⁷ The anti-inflammatory and anti-tumor effects of lancemaside A (also has β -glycosidic bonds and α -rhamnoside bond in the structure), the active constituent of *Codonopsis pilosula*, might be due to echinocystic acid, which is a metabolite produced by the action of gut microbiota.¹³⁸ Codonolaside II, echinocystic acid 28-O-*b*-D-xylopyranosyl-(1,4)-*a*-L-rhamnopyranosyl-(1,2)-*a*-L-arabinopyranosyl ester, and echinocystic acid 28-O-*a*-L-rhamnopyranosyl-(1,2)-*a*-L-arabinopyranosyl ester are other products that were produced by glycosidic bond hydrolysis.¹³⁹ In a previous study, it was shown that echinocystic acid potently inhibited acetylcholinesterase activity, and significantly reversed scopolamine-induced memory and learning deficits on passive avoidance tasks. Echinocystic acid more potently shortened the escape latencies prolonged by treatment with scopolamine when compared to lancemaside A in a Morris water maze task.¹⁴⁰

Sennoside: Sennoside is the main active ingredient in rhubarb, and has poor diarrhea effect due to its poor intestinal absorption. Sennoside glycoside (rheinanthrone), which is produced by hydrolysis of sennoside by the *Bifidobacteria* secreted β -D-glucosidase, has diarrheal effects.¹²² Rheinanthrone reflects the laxative effect by stimulating the activation of macrophages and reducing the expression of colonic aquaporin-3.¹⁴¹ Similarly, barbaloin can be hydrolyzed to aloe-rheinanthrone by the action of *Eubacterium* spp. BAR, which has a pronounced diarrheal effect.¹⁴²

Others: Rhapontin is metabolized into rhapontigenin by intestinal bacteria, which has stronger antithrombotic and anti-allergic effects when compared to its parental compound.¹⁴³ Geniposide is hydrolyzed into genipin by intestinal bacteria, which have cholagogue and anti-hepatitis effects, however an intravenous injection of geniposide does not have a cholagogue effect.¹²³

Gut microbiota not only increase the pharmacological activity of active THM ingredients, but also reduce the toxicity of certain ingredients. For example, aconitine is decarboxylated, methylated, hydroxylated, and esterified by intestinal bacteria

into mono-, di-, and lipid alkaloids, and metabolites, which have significantly less toxicity but the same pharmacological effect when compared to the parental compound.¹⁴⁴ In addition, some active THM ingredients, including ginsenoside Rb₁, crocin, amygdalin, geniposide, puerarin, ginsenoside Re, poncirin, hesperidin, and baicalin are metabolized by the gut microbiota to compound K, crocetin, benzaldehyde, genipin, daidzein, ginsenoside Rh₁, ponciretin, hesperetin, and baicalein, which have a more potent anti-tumor effect compared to the parental compounds.¹⁴⁵ The pharmacodynamics of part of the prototype compounds in THM and their metabolites in the gut are presented in Table 5.

5.1.3 Un-confirmed bioactivity metabolites of THM in gut.

As described in section "5.1.1", active substances of THM are metabolized by gut microbial enzymes. Although it is not clear whether the metabolites have strong transient pharmacological activity, several studies have shown that gut microbiota does change the behavior of these compounds *in vivo*.

O-glycoside bond compounds: Oxyglucosides are one of the most common compounds present in THM, and include oxyglucosides, glucosinolates, and azaglycosides, which are characterized by glycosidic bonds. Glycosides are highly susceptible to hydrolysis, and glycosidic bond cleavage results in the formation of aglycones. Water-soluble psoralen and isopsoralen glycosides can be hydrolyzed into psoralen and isopsoralen by the gut microbiota.¹⁴⁶ Hattori *et al.* showed that paeoniflorin is primarily metabolized into paeonian I, followed by paeoniins II, III, and IV by *Peptostreptococcus anaerobius* and *Lactobacillus*.^{124,125,147} Arctiin is the major lignan glycoside of *Fructus arctii*, and is hydrolyzed by *Blautia* spp. AUH-JLD56 into 3'-desmethylarctigenin.¹⁴⁸ Loganin is an important constituent of *Fructus corni*, and is metabolized by intestinal bacteria into log 1 and log 2, which have immuno-modulatory, anti-shock, and anti-arrhythmia functions.¹⁴⁹ Astilbin, the active ingredient of *Smilax glabra* Roxb, exhibits a variety of pharmacological activities, including anti-inflammation, anti-cancer properties. It is metabolized by intestinal bacteria into two main bioactive metabolites that are identified as eriodictyol and quercetin.¹⁵⁰ Myricitrin is an important botanical flavonol glycoside with significant anti-inflammatory and antioxidant properties. It is de-glycosylated and de-hydroxylated into quercetin-3-O-rhamnoside and quercetin, respectively.¹⁵¹ Ziyuglycoside I is one of

Table 5 Pharmacodynamics of prototype compounds in THM and its metabolites in the gut

Prototype compounds in THM	Metabolites in gut	The efficacy of metabolites superior to prototypes
Ginsenosides/ginsenoside Re	Compound K/ginsenoside Rh1	Hepato-protective, anti-cancer, anti-wrinkling, and anti-aging properties/estrogenic effect
Glycyrrhizin	Glycyrrhetic acid	Anti-hepatotoxicity
Lancemaside A	Echinocystic acid	Anti-inflammatory, anti-tumor and anti-Alzheimer's disease
Baicalin	Baicalein, orohylin A	Anti-histamine-induced pruritus response
Poncirin	Ponciretin	Anti-inflammatory
Sennosides	Rheinanthrone	Diarrheal and laxative effect
Barbaloin	Aloe-rheinanthrone	Diarrheal effect
Rhapontin	Rhapontigenin	Antithrombotic and anti-allergic effects
Geniposide	Genipin	Cholagogue effects



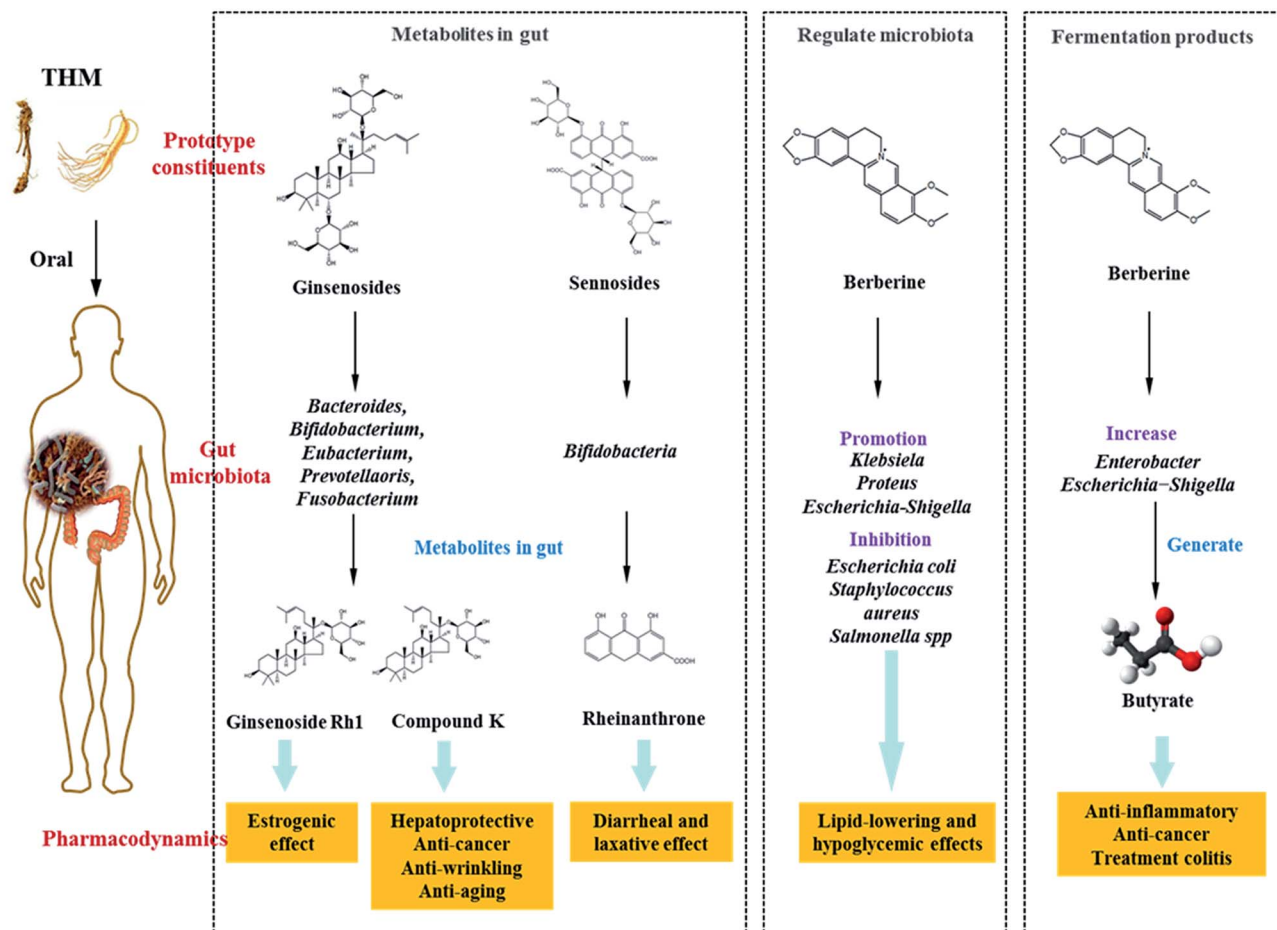


Fig. 2 The treating diseases mechanisms of THM acting via gut microbiota: (1) metabolizing into active metabolites by the action of gut microbiota; (2) regulation of gut microbiota balance; (3) regulating the fermentation products of the gut microbes.

the major active ingredients in *Sanguisorba officinalis* with free radical scavenging and elastase inhibitory activities. The gut microbiota degrade ziyuglycoside I into M0-Glu-Ara+O, M0-Ara, M0-Glu-COOH, M0-Glu, M0-Glu-Ara+O and M0-Ara+H₂O.¹⁵² Rutin, the active ingredient of *Flos sophorae*, is metabolized by *Eubacterium ramulus* into quercetin. Reduction and hydrogenation of quercetin into alphitonin, followed by oxidation cleaved generated 3,4-dihydroxyphenylacetic acid and phloroglucinol.^{153,154} Moreover, studies have shown that α -L-rhamnosidase, a rhamno-glycoside metabolizing enzyme that is secreted by *Bifidobacterium dentium*, and can hydrolyze rutin, poncirin, naringin, and ginsenoside Re. It is more effective in hydrolyzing (1 → 6) bonds than (1 → 2) bonds of rhamnoglycosides.¹⁵⁵

C-glycoside bond compounds: The C-glycoside bonds of pueraria isoflavones (active ingredient of Kudzu root) are difficult to hydrolyze. The CG19-1 strain of *Corynebacterium* isolated from human intestine specifically hydrolyzes the C-glycoside bonds of puerarin, and other aromatic compounds.¹⁵⁶

Others: Procyanidins are common polyphenols present in many THM, and exhibit antibacterial, antioxidant, antiviral and anti-inflammatory properties. A-type and B-type procyanidins are metabolized to benzoic acid, 2-phenylacetic acid, and 3-phenylpropionic acid, respectively, by gut microbiota.¹⁵⁷

Ellagitannin, a polyphenol isolated from *Polygonum capitatum* with potential antitumor activity, is metabolized into thirteen metabolites by the intestinal bacteria *in vitro*.¹⁵⁸ In addition, *Eubacterium ramulus* SY8519 dehydrodaidze into *O*-demethylandrostatin.¹⁵⁹

Taken together, although the activities of the above-mentioned metabolites have not yet been confirmed, gut microbiota can certainly alter the composition of THM in the gut.

5.2 THM regulate gut microbiota balance

Under normal circumstances, the intestinal microflora maintains a symbiotic or antagonistic relationship with the host, and constitutes a micro-ecological balance in the human body. If this balance is disturbed during pathophysiological conditions, dysbiosis may occur. Therefore, the physiological relationship between normal flora and host is transformed into a pathological relationship. Intestinal dysbiosis is largely the result of the overuse of antibiotics, and is associated with various diseases of the digestive system, allergies, and immune disorders among others.

In previous studies, it has been shown that several THM and its active compounds have prebiotic-like effects that promote



the proliferation of intestinal bacteria. Berberine, the active ingredient of *Rhizoma coptidis*, prevents high-fat diet-induced obesity and diabetes. Berberine also exerts a significant antimicrobial activity by inhibiting the assembly function of FtsZ and halting bacterial division. Berberine improves gut microbiota composition in rats, promotes the growth of probiotics and production of short-chain fatty acids, lowers the intestinal pH and the accumulation of exogenous antigens, increases the levels of lipopolysaccharide-binding protein and leptin, reduces serum adiponectin, and exerts lipid-lowering and hypoglycemic effects.^{160,161} Furthermore, Berberine significantly enriches the genera of *Klebsiella*, *Proteus*, *Escherichia*, and *Shigella* of the Enterobacteriaceae family in gut microbiota, and inhibits the growth of *Lactobacillus in vitro*.^{162,163} Astragali Radix is a THM, which is known for its immunogenic effect. Its active ingredient is calycosin-7-*O*-glucoside, which is hydrolyzed to aglycone calycosin by gut microbiota *in vitro*. Calycosin has a higher permeability when compared with calycosin-7-*O*-glucoside, although the parental compound stimulates the growth of *Lactobacillus* and *Bifidobacterium*.¹⁶⁴

Extracts of THM or its active compounds also have regulatory effects on the gut microbiota. THM polysaccharides can restore the homeostasis of gut microbiota and promote the absorption of small molecules. Ginseng polysaccharides in the Du-Shen-Tang formulation have shown to improve intestinal metabolism and absorption of certain ginsenosides and restore perturbed gut microbiota, and enhanced the growth of *Lactobacillus* spp. and *Bacteroides* spp.¹⁶⁵ Saponins and polysaccharides of *Polygonatum kingianum* prevented the increase in fasting blood glucose levels and enhanced the amount of fasting insulin, which may improve intestinal micro-ecology by decreasing the abundance of Bacteroidetes and Proteobacteria, thereby increasing that of Firmicutes.¹⁶⁶ *Ganoderma lucidum* extracts added to a high-fat mouse diet not only reduced the ratio of Firmicutes/Bacteroidetes and endotoxin-producing bacteria induced by the high-fat diet, but also maintained the integrity of the intestinal barrier, and reduced endotoxemia.^{167,168} In addition, extracts of ginseng and *Semen coicis* have shown to improve intestinal dysbiosis and symptoms of colonic inflammation, as demonstrated in an ulcerative colitis model induced by trinitrobenzene sulfonic acid. Furthermore, these extracts also promoted the growth of probiotics including *Lactobacillus* and *Bifidobacterium*, and inhibited the growth of pathogens, such as *E. coli*, *S. aureus* and *Salmonella* spp., which was verified *in vitro*.¹⁶⁹ Chinese herbs and herbal extracts, such as polysaccharides from the seed of *Plantago asiatica* L. and *Dendrobium candidum* could treat diabetes mellitus by regulating the gut microbiota.¹⁷⁰ Furthermore, *Ulmus rubra* and triphala significantly increased the relative abundance of butyrate-producing bacteria, and *Glycyrrhiza glabra* was increased in propionate-producing species.¹⁷¹

5.3 Gut microbiota fermentation products are influenced by THM

The major gut fermentation products in healthy adults involve gases and organic acids, in particular the three short-chain fatty

acids (SCFAs) acetate, propionate, and butyrate, which have anti-inflammatory and anti-cancer effects.¹⁷² Acetate is the main product of colonic fermentation, most of which is produced by the fermentation of undigested carbohydrates in the intestine by colonic anaerobic bacteria.¹⁷³ Propionate is the main product of *Bacteroides* fermentation, whereas butyrate is the main metabolite of Firmicutes, which can be absorbed and utilized by colonic epithelial cells, and are the preferred source of energy for both the colon and cecum.¹⁷⁴ Bacteria that produce SCFAs include *Bacteroides*, *Clostridium*, *Bifidobacterium*, *Eubacterium*, *Streptococcus*, and *Peptostreptococcus*.

Immune regulation and anti-inflammatory: SCFAs participated in intestinal immune regulation by affecting the release of inflammatory factors, immunochemokines, and inhibition of epidemic effector cell proliferation, and play an important role in the intestinal defense against pathogenic bacteria.¹⁷⁵ Recent data have shown that SCFAs also regulate the size and function of the colonic Treg cell pool, and protect against colitis in a Ffar2-dependent manner.¹⁷⁶ SCFAs could inhibit histone deacetylase, reduce NF- κ B activity, and inhibit IL-8 and TNF- α , which are released from neutrophils and macrophages, thereby reducing the inflammatory response in the intestine.¹⁷⁷ Butyrate could also inhibit intestinal inflammation through inhibiting the IFN- γ /STAT1 signaling pathway, tyrosine and serine phosphorylation, nuclear translocation, and DNA binding activity.¹⁷⁸ A total of 30 mmol L⁻¹ acetate, propionate and butyrate reduced the release of TNF- α without affecting the release of IL- κ protein, thereby indicating that SCFAs have a good therapeutic effect on colitis.¹⁷⁹

Anti-tumor: physiological doses of acetate, propionate, and butyrate inhibited the growth and proliferation of rectal tumor cells, and induced tumor cell differentiation, and apoptosis.¹⁸⁰ SLC5A8, which is predominantly expressed in the colon, is a Na(+)-coupled transporter for short-chain fatty acids that inhibited tumor growth by regulating the bacterial fermentation product butyrate.¹⁸¹ SCFAs also protected the colonic mucosa from tumorigenesis by regulating colon cell phenotype, DNA synthesis and methylation, and c-myc levels.^{182,183}

Energy metabolism: in the context of energy metabolism, SCFAs activate the GPR43, thereby suppressing insulin signaling in adipocytes, which inhibits fat accumulation and promotes the metabolism of unincorporated lipids and glucose in other tissues.¹⁸⁴ SCFAs may also be the key factors involved in improving the pathophysiology of diabetes.¹⁸⁵

Others: SCFAs function in maintaining the balance of water and electrolytes in the gut. Perfusion of SCFAs can significantly reduce the secretion of water in the colon, and subsequently, butyrate significantly reduced the secretion of Na⁺, K⁺, and Cl⁻, while propionate reduced the secretion of HCO₃⁻, and acetate inhibited the secretion of Na⁺ and Cl⁻.¹⁸⁶ Intravenous SCFAs effectively maintained the height, width, crypt depth and mucosal thickness of intestinal villi, increased the proliferation capacity of intestinal mucosal cells, reduce intestinal mucosal atrophy caused by triphosphopyridine nucleotides, maintained the morphology of intestinal mucosa and maintained the integrity of intestinal epithelial cells.¹⁸⁷ SCFAs also promoted glucagon-like peptide-2 secretion in intestinal cells, which is an



intestinal epithelial-specific growth factor that promotes intestinal mucosal growth and repairs intestinal epithelial damage.¹⁸⁸

In recent studies, it was confirmed that THM affected the fermentation products of gut microbiota, for example berberine lowered blood lipid and glucose levels *via* multi-target mechanisms, and affected the SCFAs. Oral administration of berberine modified intestinal bacterial composition by increasing the abundance of butyrate-producing bacteria, which then entered the circulation and reduced blood lipid and glucose levels.¹⁸⁹ Polysaccharides from seeds of *Plantago asiatica* L. increased SCFAs and fecal moisture along with lowering the pH in the colon, which might be beneficial for colon health.¹⁹⁰ Polysaccharides from *Polygonatum kingianum* decreased total SCFAs, acetate, propionate, and butyrate in high-fat diet-induced type 2 diabetic rat gut.¹⁶⁶ Moreover, the leaves of *Passiflora edulis* decreased acetate and butyrate levels in normal male Wistar rat gut.¹⁹¹ Polysaccharides from *Chrysanthemum morifolium*, Xiexin Tang, polysaccharides S-3 from Sijunzi decoction increased acetate, propionate and butyrate levels in rat gut.¹⁹²

6. Perspectives

THM is the foundation of modern medicine, and many successful THM-based pharmaceutical drugs have been formulated. Prominent examples include morphine, which was isolated from opium in 1806, strychnine (1818), ephedrine (1887), paclitaxel (1971), and more recently artemisinin, which was the subject of the 2015 Nobel Prize in Physiology or Medicine. These drugs have made significant contributions to human health, and have indicated the importance of THM in modern medical research and development. However, to date very few active ingredients of THM have been identified by pharmacodynamical and biological assays either *in vitro* or *in vivo*. Furthermore, the content of these substances in THM preparations is very low, and most of them have poor bioavailability. Nevertheless, thousands of years of observing the practical benefits of THM and modern clinical research has proven the efficacy of THM in several diseases. Further research and development is still required to elucidate the underlying mechanism of action.

Disturbances in gut microbiota composition are related to the occurrence, development, and treatment of various diseases. Several studies have shown that active components of THM or its extracts could interact with gut microbiota, thereby resulting in positive effects (Fig. 2). These findings suggested that the role of THM in treating diseases may be related to gut microbiota, and there may be three likely modes of interaction: (1) metabolism of THM by microbes into active and even more potent metabolites, (2) regulation of gut microbiota by THM components, and (3) promoting the synthesis of therapeutic fermentation products, such as SCFAs by the gut microbiota. In addition, gut microbiota can increase the toxicity of several THM, such as amygdalin, hydrolyze this to HCN under the anaerobic Bacteroidetes phylum in the gut and cause toxicity.¹⁹³

In summary, both historic and modern science validate the efficacy of THM, however research and development in this field

has staggered due to the complexity of ingredients and the unknown mechanism of action. Thus, in recent years, research on gut microbiota has brought a new perspective into the study of THM.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This study was financially supported by the Fundamental Research Funds for the Central public welfare research institutes (project no. ZXKT17013 and ZZ11-092) and Young Elite Scientists Sponsorship Program by CACM (2018QNRC2-C01).

References

- 1 T. Hesketh and W. X. Zhu, *BMJ*, 1997, **315**, 115–117.
- 2 J. Xu and Y. Yang, *Health Policy*, 2009, **90**, 133–139.
- 3 P. Fisher and A. Ward, *BMJ*, 1994, **309**, 107–111.
- 4 F. Firenzuoli and L. Gori, *Evid. Based Complement Altern. Med.*, 2007, **4**, 37–40.
- 5 P. A. De Smet, *N. Engl. J. Med.*, 2005, **352**, 1176–1178.
- 6 D. M. Eisenberg, R. C. Kessler, C. Foster, F. E. Norlock, D. R. Calkins and T. L. Delbanco, *N. Engl. J. Med.*, 1993, **328**, 246–252.
- 7 R. C. Kessler, R. B. Davis, D. F. Foster, M. I. Van Rompay, E. E. Walters, S. A. Wilkey, T. J. Kaptchuk and D. M. Eisenberg, *Ann. Intern. Med.*, 2001, **135**, 262–268.
- 8 C. H. Wu, C. C. Wang and J. Kennedy, *Clin. Ther.*, 2011, **33**, 1749–1758.
- 9 O. S. Olorunnisola, A. Adetutu, E. A. Balogun and A. J. Afolayan, *J. Ethnopharmacol.*, 2013, **150**, 71–78.
- 10 A. C. Liwa, L. R. Smart, A. Frumkin, H. A. Epstein, D. W. Fitzgerald and R. N. Peck, *Curr. Hypertens. Rep.*, 2014, **16**, 437.
- 11 S. Sen and R. Chakraborty, *J. Tradit. Complement. Med.*, 2017, **7**, 234–244.
- 12 J. L. Tang and T. W. Wong, *Hong Kong Med. J.*, 1998, **4**, 208–210.
- 13 L. D'Angelo, R. Grimaldi, M. Caravaggi, M. Marcoli, E. Perucca, S. Lecchini, G. M. Frigo and A. Crema, *J. Ethnopharmacol.*, 1986, **16**, 15–22.
- 14 H. Amagase and D. M. Nance, *J. Altern. Complement. Med.*, 2008, **14**, 403–412.
- 15 G. Hu, R. S. Walls, D. Bass, B. Ramon, D. Grayson, M. Jones and V. Gebiski, *Ann. Allergy, Asthma, Immunol.*, 2002, **88**, 478–487.
- 16 A. Bensoussan, N. J. Talley, M. Hing, R. Menzies, A. Guo and M. Ngu, *JAMA*, 1998, **280**, 1585–1589.
- 17 W. K. Leung, J. C. Wu, S. M. Liang, L. S. Chan, F. K. Chan, H. Xie, S. S. Fung, A. J. Hui, V. W. Wong, C. T. Che and J. J. Sung, *Am. J. Gastroenterol.*, 2006, **101**, 1574–1580.
- 18 K. Iwasaki, S. Kobayashi, Y. Chimura, M. Taguchi, K. Inoue, S. Cho, T. Akiba, H. Arai, J. C. Cyong and H. Sasaki, *J. Am. Geriatr. Soc.*, 2004, **52**, 1518–1521.



- 19 World Health Organization, *General guidelines for methodologies on research and evaluation of traditional medicine*, World Health Organization, Geneva, 2000.
- 20 S. Li, Q. Han, C. Qiao, J. Song, C. Lung Cheng and H. Xu, *Chin. Med.*, 2008, **3**, 7.
- 21 X. M. Liang, Y. Jin, Y. P. Wang, G. W. Jin, Q. Fu and Y. S. Xiao, *J. Chromatogr. A*, 2009, **1216**, 2033–2044.
- 22 Y.-Z. Liang, P. Xie and K. Chan, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2004, **812**, 53–70.
- 23 I. B. Afanas'ev, A. I. Dorozhko, A. V. Brodskii, V. A. Kostyuk and A. I. Potapovitch, *Biochem. Pharmacol.*, 1989, **38**, 1763–1769.
- 24 G. L. Russo, M. Russo, C. Spagnuolo, I. Tedesco, S. Bilotto, R. Iannitti and R. Palumbo, *Cancer Treat. Res.*, 2014, **159**, 185–205.
- 25 D. W. Lamson and M. S. Brignall, *Alternative Medicine Review*, 2000, **5**, 196–208.
- 26 S. Y. Wei, Y. Chen and X. Y. Xu, *Chin. J. Nat. Med.*, 2014, **12**, 407–414.
- 27 Y. X. Zhou, H. Zhang and C. Peng, *Phytother. Res.*, 2014, **28**, 961–975.
- 28 K. Kesarwani and R. Gupta, *Asian Pac. J. Trop. Biomed.*, 2013, **3**, 253–266.
- 29 F. Veiga, C. Fernandes and F. Teixeira, *Int. J. Pharm.*, 2000, **202**, 165–171.
- 30 T. H. Xue and R. Roy, *Science*, 2003, **300**, 740–741.
- 31 D. Pal, D. Kwatra, M. Minocha, D. K. Paturi, B. Budda and A. K. Mitra, *Life Sci.*, 2011, **88**, 959–971.
- 32 X. K. Huo, Q. Liu, C. Y. Wang, Q. Meng, H. J. Sun, J. Y. Peng, X. C. Ma and K. X. Liu, *Eur. J. Pharm. Sci.*, 2013, **50**, 420–428.
- 33 C. Mohn, H. G. Hacker, R. A. Hilger, M. Gutschow and U. Jaehde, *Pharmazie*, 2013, **68**, 622–627.
- 34 P. Goldman, *Ann. Intern. Med.*, 2001, **135**, 594–600.
- 35 J. H. Lee, J. M. Kim and C. Kim, *J. Ethnopharmacol.*, 2003, **84**, 5–9.
- 36 W. Zhu, X. M. Wang, L. Zhang, X. Y. Li and B. X. Wang, *Am. J. Chin. Med.*, 2005, **33**, 839–850.
- 37 Z. Wen, T. E. Dumas, S. J. Schrieber, R. L. Hawke, M. W. Fried and P. C. Smith, *Drug Metab. Dispos.*, 2008, **36**, 65–72.
- 38 S. M. Wittemer, M. Ploch, T. Windeck, S. C. Muller, B. Drewelow, H. Derendorf and M. Veit, *Phytomedicine*, 2005, **12**, 28–38.
- 39 K. Woelkart, E. Feizlmayr, P. Dittrich, E. Beubler, F. Pinl, A. Suter and R. Bauer, *Phytother. Res.*, 2010, **24**, 445–450.
- 40 T. Lu, J. Yang, X. Gao, P. Chen, F. Du, Y. Sun, F. Wang, F. Xu, H. Shang, Y. Huang, Y. Wang, R. Wan, C. Liu, B. Zhang and C. Li, *Drug Metab. Dispos.*, 2008, **36**, 1578–1586.
- 41 A. Wen, J. Yang, Y. Jia, Z. Yang, Y. Tian, Y. Wu, Z. Wang and Z. He, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2008, **876**, 41–46.
- 42 C. Z. Wang, K. E. Kim, G. J. Du, L. W. Qi, X. D. Wen, P. Li, B. A. Bauer, M. B. Bissonnette, M. W. Musch, E. B. Chang and C. S. Yuan, *Am. J. Chin. Med.*, 2011, **39**, 1161–1171.
- 43 R. Sender, S. Fuchs and R. Milo, *Cell*, 2016, **164**, 337–340.
- 44 R. Sender, S. Fuchs and R. Milo, *PLoS Biol.*, 2016, **14**, e1002533.
- 45 T. S. Postler and S. Ghosh, *Cell Metab.*, 2017, **26**, 110–130.
- 46 J. Qin, R. Li, J. Raes, M. Arumugam, K. S. Burgdorf, C. Manichanh, T. Nielsen, N. Pons, F. Levenez, T. Yamada, D. R. Mende, J. Li, J. Xu, S. Li, D. Li, J. Cao, B. Wang, H. Liang, H. Zheng, Y. Xie, J. Tap, P. Lepage, M. Bertalan, J. M. Batto, T. Hansen, D. Le Paslier, A. Linneberg, H. B. Nielsen, E. Pelletier, P. Renault, T. Sicheritz-Ponten, K. Turner, H. Zhu, C. Yu, S. Li, M. Jian, Y. Zhou, Y. Li, X. Zhang, S. Li, N. Qin, H. Yang, J. Wang, S. Brunak, J. Dore, F. Guarner, K. Kristiansen, O. Pedersen, J. Parkhill, J. Weissenbach, H. I. T. C. Meta, P. Bork, S. D. Ehrlich and J. Wang, *Nature*, 2010, **464**, 59–65.
- 47 S. R. Gill, M. Pop, R. T. Deboy, P. B. Eckburg, P. J. Turnbaugh, B. S. Samuel, J. I. Gordon, D. A. Relman, C. M. Fraser-Liggett and K. E. Nelson, *Science*, 2006, **312**, 1355–1359.
- 48 C. Human, Microbiome Project, *Nature*, 2012, **486**, 215–221.
- 49 J. Lederberg, *Science*, 2000, **288**, 287–293.
- 50 N. Arpaia, C. Campbell, X. Fan, S. Dikiy, J. van der Veeken, P. deRoos, H. Liu, J. R. Cross, K. Pfeffer, P. J. Coffey and A. Y. Rudensky, *Nature*, 2013, **504**, 451–455.
- 51 W. Jia, H. Li, L. Zhao and J. K. Nicholson, *Nat. Rev. Drug Discovery*, 2008, **7**, 123–129.
- 52 M. G. Rooks and W. S. Garrett, *Nat. Rev. Immunol.*, 2016, **16**, 341–352.
- 53 P. Bercik, S. M. Collins and E. F. Verdu, *Neurogastroenterol. Motil.*, 2012, **24**, 405–413.
- 54 A. L. Kau, P. P. Ahern, N. W. Griffin, A. L. Goodman and J. I. Gordon, *Nature*, 2011, **474**, 327–336.
- 55 G. Sharon, T. R. Sampson, D. H. Geschwind and S. K. Mazmanian, *Cell*, 2016, **167**, 915–932.
- 56 D. Kelly, S. Conway and R. Aminov, *Trends Immunol.*, 2005, **26**, 326–333.
- 57 M. Bajzer and R. J. Seeley, *Nature*, 2006, **444**, 1009–1010.
- 58 L. M. Bustos Fernandez, J. S. Lasa and F. Man, *J. Clin. Gastroenterol.*, 2014, **48**, 657–666.
- 59 J. L. Pluznick, R. J. Protzko, H. Gevorgyan, Z. Peterlin, A. Sipos, J. Han, I. Brunet, L. X. Wan, F. Rey, T. Wang, S. J. Firestein, M. Yanagisawa, J. I. Gordon, A. Eichmann, J. Peti-Peterdi and M. J. Caplan, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 4410–4415.
- 60 H. S. Ejtahed, J. Mohtadi-Nia, A. Homayouni-Rad, M. Niafar, M. Asghari-Jafarabadi and V. Mofid, *Nutrition*, 2012, **28**, 539–543.
- 61 T. R. Sampson, J. W. Debelius, T. Thron, S. Janssen, G. G. Shastri, Z. E. Ilhan, C. Challis, C. E. Schretter, S. Rocha, V. Gradinaru, M. F. Chesselet, A. Keshavarzian, K. M. Shannon, R. Krajmalnik-Brown, P. Wittung-Stafshede, R. Knight and S. K. Mazmanian, *Cell*, 2016, **167**, 1469–1480.
- 62 J. Qin, Y. Li, Z. Cai, S. Li, J. Zhu, F. Zhang, S. Liang, W. Zhang, Y. Guan, D. Shen, Y. Peng, D. Zhang, Z. Jie, W. Wu, Y. Qin, W. Xue, J. Li, L. Han, D. Lu, P. Wu, Y. Dai, X. Sun, Z. Li, A. Tang, S. Zhong, X. Li, W. Chen, R. Xu, M. Wang, Q. Feng, M. Gong, J. Yu, Y. Zhang, M. Zhang, T. Hansen, G. Sanchez, J. Raes, G. Falony, S. Okuda, M. Almeida, E. LeChatelier, P. Renault, N. Pons,



- J. M. Batto, Z. Zhang, H. Chen, R. Yang, W. Zheng, S. Li, H. Yang, J. Wang, S. D. Ehrlich, R. Nielsen, O. Pedersen, K. Kristiansen and J. Wang, *Nature*, 2012, **490**, 55–60.
- 63 E. S. Bliss and E. Whiteside, *Front. Physiol.*, 2018, **9**, 900.
- 64 M. Arumugam, J. Raes, E. Pelletier, D. Le Paslier, T. Yamada, D. R. Mende, G. R. Fernandes, J. Tap, T. Bruls, J. M. Batto, M. Bertalan, N. Borruel, F. Casellas, L. Fernandez, L. Gautier, T. Hansen, M. Hattori, T. Hayashi, M. Kleerebezem, K. Kurokawa, M. Leclerc, F. Levenez, C. Manichanh, H. B. Nielsen, T. Nielsen, N. Pons, J. Poulain, J. Qin, T. Sicheritz-Ponten, S. Tims, D. Torrents, E. Ugarte, E. G. Zoetendal, J. Wang, F. Guarner, O. Pedersen, W. M. de Vos, S. Brunak, J. Dore, H. I. T. C. Meta, M. Antolin, F. Artiguenave, H. M. Blottiere, M. Almeida, C. Brechot, C. Cara, C. Chervaux, A. Cultrone, C. Delorme, G. Denariatz, R. Dervyn, K. U. Foerstner, C. Friss, M. van de Guchte, E. Guedon, F. Haimet, W. Huber, J. van Hylckama-Vlieg, A. Jamet, C. Juste, G. Kaci, J. Knol, O. Lakhdari, S. Layec, K. Le Roux, E. Maguin, A. Merieux, R. Melo Minardi, C. M'Rini, J. Muller, R. Oozeer, J. Parkhill, P. Renault, M. Rescigno, N. Sanchez, S. Sunagawa, A. Torrejon, K. Turner, G. Vandemeulebrouck, E. Varela, Y. Winogradsky, G. Zeller, J. Weissenbach, S. D. Ehrlich and P. Bork, *Nature*, 2011, **473**, 174–180.
- 65 P. B. Eckburg, E. M. Bik, C. N. Bernstein, E. Purdom, L. Dethlefsen, M. Sargent, S. R. Gill, K. E. Nelson and D. A. Relman, *Science*, 2005, **308**, 1635–1638.
- 66 F. Backhed, R. E. Ley, J. L. Sonnenburg, D. A. Peterson and J. I. Gordon, *Science*, 2005, **307**, 1915–1920.
- 67 R. E. Ley, D. A. Peterson and J. I. Gordon, *Cell*, 2006, **124**, 837–848.
- 68 F. Backhed, J. K. Manchester, C. F. Semenkovich and J. I. Gordon, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 979–984.
- 69 G. Rogler and G. Rosano, *Eur. Heart J.*, 2014, **35**, 426–430.
- 70 A. F. Ahmad, N. C. Ward and G. Dwivedi, *Curr. Opin. Cardiol.*, 2018, **34**, 1.
- 71 Y. Nagatomo and W. H. Tang, *J. Card. Fail.*, 2015, **21**, 973–980.
- 72 J. F. Cryan and S. M. O'Mahony, *Neurogastroenterol. Motil.*, 2011, **23**, 187–192.
- 73 Y. Wang and L. H. Kasper, *Brain Behav. Immun.*, 2014, **38**, 1–12.
- 74 E. A. Mayer, R. Knight, S. K. Mazmanian, J. F. Cryan and K. Tillisch, *J. Neurosci.*, 2014, **34**, 15490–15496.
- 75 H. Chu, B. Williams and B. Schnabl, *Liver Res.*, 2018, **2**, 43–51.
- 76 L. Bird, *Nat. Rev. Immunol.*, 2012, **12**, 153.
- 77 N. Qin, F. Yang, A. Li, E. Prifti, Y. Chen, L. Shao, J. Guo, E. Le Chatelier, J. Yao, L. Wu, J. Zhou, S. Ni, L. Liu, N. Pons, J. M. Batto, S. P. Kennedy, P. Leonard, C. Yuan, W. Ding, Y. Chen, X. Hu, B. Zheng, G. Qian, W. Xu, S. D. Ehrlich, S. Zheng and L. Li, *Nature*, 2014, **513**, 59–64.
- 78 J. G. Fox, Y. Feng, E. J. Theve, A. R. Raczynski, J. L. Fiala, A. L. Doernte, M. Williams, J. L. McFaline, J. M. Essigmann, D. B. Schauer, S. R. Tannenbaum, P. C. Dedon, S. A. Weinman, S. M. Lemon, R. C. Fry and A. B. Rogers, *Gut*, 2010, **59**, 88–97.
- 79 A. Everard, C. Belzer, L. Geurts, J. P. Ouwerkerk, C. Druart, L. B. Bindels, Y. Guiot, M. Derrien, G. G. Muccioli, N. M. Delzenne, W. M. de Vos and P. D. Cani, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 9066–9071.
- 80 T. Le Roy, M. Llopis, P. Lepage, A. Bruneau, S. Rabot, C. Bevilacqua, P. Martin, C. Philippe, F. Walker, A. Bado, G. Perlemuter, A. M. Cassard-Doulier and P. Gerard, *Gut*, 2013, **62**, 1787–1794.
- 81 N. Fei and L. Zhao, *ISME J.*, 2013, **7**, 880–884.
- 82 M. Pop, A. W. Walker, J. Paulson, B. Lindsay, M. Antonio, M. A. Hossain, J. Oundo, B. Tamboura, V. Mai, I. Astrovskaya, H. Corrada Bravo, R. Rance, M. Stares, M. M. Levine, S. Panchalingam, K. Kotloff, U. N. Ikumapayi, C. Ebruke, M. Adeyemi, D. Ahmed, F. Ahmed, M. T. Alam, R. Amin, S. Siddiqui, J. B. Ochieng, E. Ouma, J. Juma, E. Mailu, R. Omoro, J. G. Morris, R. F. Breiman, D. Saha, J. Parkhill, J. P. Nataro and O. C. Stine, *Genome Biol.*, 2014, **15**, R76.
- 83 A. W. DuPont and H. L. DuPont, *Nat. Rev. Gastroenterol. Hepatol.*, 2011, **8**, 523–531.
- 84 A. Swidsinski, J. Weber, V. Loening-Baucke, L. P. Hale and H. Lochs, *J. Clin. Microbiol.*, 2005, **43**, 3380–3389.
- 85 X. C. Morgan, T. L. Tickle, H. Sokol, D. Gevers, K. L. Devaney, D. V. Ward, J. A. Reyes, S. A. Shah, N. LeLeiko, S. B. Snapper, A. Bousvaros, J. Korzenik, B. E. Sands, R. J. Xavier and C. Huttenhower, *Genome Biol.*, 2012, **13**, R79.
- 86 T. Wang, G. Cai, Y. Qiu, N. Fei, M. Zhang, X. Pang, W. Jia, S. Cai and L. Zhao, *ISME J.*, 2012, **6**, 320–329.
- 87 L. Aymeric, F. Donnadieu, C. Mulet, L. du Merle, G. Nigro, A. Saffarian, M. Berard, C. Poyart, S. Robine, B. Regnault, P. Trieu-Cuot, P. J. Sansonetti and S. Dramsi, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**, E283–E291.
- 88 G. Hajishengallis, R. P. Darveau and M. A. Curtis, *Nat. Rev. Microbiol.*, 2012, **10**, 717–725.
- 89 K. Honda, *Cell Host Microbe*, 2011, **10**, 423–425.
- 90 J. U. Scher and S. B. Abramson, *Nat. Rev. Rheumatol.*, 2011, **7**, 569–578.
- 91 V. Moos and T. Schneider, *Eur. J. Clin. Microbiol. Infect. Dis.*, 2011, **30**, 1151–1158.
- 92 H. Z. Apperloo-Renkema, H. Bootsma, B. I. Mulder, C. G. Kallenberg and D. van der Waaij, *Epidemiol. Infect.*, 1994, **112**, 367–373.
- 93 A. Hevia, C. Milani, P. Lopez, A. Cuervo, S. Arbolea, S. Duranti, F. Turrone, S. Gonzalez, A. Suarez, M. Gueimonde, M. Ventura, B. Sanchez and A. Margolles, *mBio*, 2014, **5**, e01548–01514.
- 94 M. R. Edwards, N. W. Bartlett, T. Hussell, P. Openshaw and S. L. Johnston, *Nat. Rev. Microbiol.*, 2012, **10**, 459–471.
- 95 X. L. Zheng, Y. Yang, B. J. Wang, J. Wang and H. Q. Tang, *Chin. J. Integr. Med.*, 2017, **23**, 196–200.
- 96 F. H. Karlsson, F. Fak, I. Nookaew, V. Tremaroli, B. Fagerberg, D. Petranovic, F. Backhed and J. Nielsen, *Nat. Commun.*, 2012, **3**, 1245.



- 97 W. C. Huang, Y. M. Chen, N. W. Kan, C. S. Ho, L. Wei, C. H. Chan, H. Y. Huang and C. C. Huang, *Nutrients*, 2015, **7**, 3767–3782.
- 98 F. Z. Marques, E. Nelson, P. Y. Chu, D. Horlock, A. Fiedler, M. Ziemann, J. K. Tan, S. Kuruppu, N. W. Rajapakse, A. El-Osta, C. R. Mackay and D. M. Kaye, *Circulation*, 2017, **135**, 964–977.
- 99 J. M. Yano, K. Yu, G. P. Donaldson, G. G. Shastri, P. Ann, L. Ma, C. R. Nagler, R. F. Ismagilov, S. K. Mazmanian and E. Y. Hsiao, *Cell*, 2015, **161**, 264–276.
- 100 R. P. Gaykema, L. E. Goehler and M. Lyte, *Brain Behav. Immun.*, 2004, **18**, 238–245.
- 101 P. Bercik, E. F. Verdu, J. A. Foster, J. Macri, M. Potter, X. Huang, P. Malinowski, W. Jackson, P. Blennerhassett, K. A. Neufeld, J. Lu, W. I. Khan, I. Cortesey-Theulaz, C. Cherbut, G. E. Bergonzelli and S. M. Collins, *Gastroenterology*, 2010, **139**, 2102–2112.
- 102 E. Y. Hsiao, S. W. McBride, S. Hsien, G. Sharon, E. R. Hyde, T. McCue, J. A. Codelli, J. Chow, S. E. Reisman, J. F. Petrosino, P. H. Patterson and S. K. Mazmanian, *Cell*, 2013, **155**, 1451–1463.
- 103 F. Scheperjans, V. Aho, P. A. Pereira, K. Koskinen, L. Paulin, E. Pekkonen, E. Haapaniemi, S. Kaakkola, J. Eerola-Rautio, M. Pohja, E. Kinnunen, K. Murros and P. Auvinen, *Mov. Disord.*, 2015, **30**, 350–358.
- 104 S. Jangi, R. Gandhi, L. M. Cox, N. Li, F. von Glehn, R. Yan, B. Patel, M. A. Mazzola, S. Liu, B. L. Glanz, S. Cook, S. Tankou, F. Stuart, K. Melo, P. Nejad, K. Smith, B. D. Topcuolu, J. Holden, P. Kivisakk, T. Chitnis, P. L. De Jager, F. J. Quintana, G. K. Gerber, L. Bry and H. L. Weiner, *Nat. Commun.*, 2016, **7**, 12015.
- 105 S. Leclercq, S. Matamoros, P. D. Cani, A. M. Neyrinck, F. Jamar, P. Starkel, K. Windey, V. Tremaroli, F. Backhed, K. Verbeke, P. de Timary and N. M. Delzenne, *Proc. Natl. Acad. Sci. U. S. A.*, 2014, **111**, E4485–E4493.
- 106 Y. Kim, J. A. Hollenbaugh, D. H. Kim and B. Kim, *PLoS One*, 2011, **6**, e21781.
- 107 R. F. Thornton, E. C. Murphy, T. F. Kagawa, P. W. O'Toole and J. C. Cooney, *BMC Microbiol.*, 2012, **12**, 190.
- 108 M. Mroczynska and Z. Libudzisz, *Pol. J. Microbiol.*, 2010, **59**, 265–269.
- 109 Y. J. Hyun, B. Kim and D. H. Kim, *J. Microbiol. Biotechnol.*, 2012, **22**, 535–540.
- 110 Suryani, T. Kimura, K. Sakka and K. Ohmiya, *Biosci., Biotechnol., Biochem.*, 2004, **68**, 609–614.
- 111 J. K. Yang, H. J. Yoon, H. J. Ahn, B. I. Lee, J. D. Pedelacq, E. C. Liang, J. Berendzen, M. Laivenieks, C. Vieille, G. J. Zeikus, D. J. Voadlo, S. G. Withers and S. W. Suh, *J. Mol. Biol.*, 2004, **335**, 155–165.
- 112 P. Brigidi, B. Vitali, E. Swennen, G. Bazzocchi and D. Matteuzzi, *Res. Microbiol.*, 2001, **152**, 735–741.
- 113 M. Miwa, T. Horimoto, M. Kiyohara, T. Katayama, M. Kitaoka, H. Ashida and K. Yamamoto, *Glycobiology*, 2010, **20**, 1402–1409.
- 114 V. Kadiyala, L. J. Nadeau and J. C. Spain, *Appl. Environ. Microbiol.*, 2003, **69**, 6520–6526.
- 115 A. de Moreno de LeBlanc and G. Perdigon, *Biocell*, 2005, **29**, 15–24.
- 116 S. Punj and G. H. John, *Curr. Issues Mol. Biol.*, 2009, **11**, 59–65.
- 117 E. Isolauri, S. Rautava and S. Salminen, *Gastroenterol. Clin. North Am.*, 2012, **41**, 747–762.
- 118 S. Rautava and E. Isolauri, *Curr. Issues Mol. Biol.*, 2002, **3**, 15–22.
- 119 A. Cartmell, L. S. McKee, M. J. Pena, J. Larsbrink, H. Brumer, S. Kaneko, H. Ichinose, R. J. Lewis, A. Vikso-Nielsen, H. J. Gilbert and J. Marles-Wright, *J. Biol. Chem.*, 2011, **286**, 15483–15495.
- 120 A. Amaretti, S. Raimondi, A. Leonardi, A. Quartieri and M. Rossi, *Nutrients*, 2015, **7**, 2788–2800.
- 121 P. F. Zhou, J. Z. Wang, X. X. Pang and H. C. Lv, *J. Guangdong Pharm. Coll.*, 2011, **27**, 582–586.
- 122 M. Matsumoto, A. Ishige, Y. Yazawa, M. Kondo, K. Muramatsu and K. Watanabe, *PLoS One*, 2012, **7**, e31700.
- 123 T. Akao, K. Kobashi and M. Aburada, *Biol. Pharm. Bull.*, 1994, **17**, 1573–1576.
- 124 T. Akao, Y. Z. Shu, Y. Matsuda, M. Hattori, T. Namba and K. Kobashi, *Chem. Pharm. Bull.*, 1988, **36**, 3043–3048.
- 125 Y. Z. Shu, M. Hattori, T. Akao, K. Kobashi, K. Kagei, K. Fukuyama, T. Tsukihara and T. Namba, *Chem. Pharm. Bull.*, 1987, **35**, 3726–3733.
- 126 P. J. Turnbaugh, C. Quince, J. J. Faith, A. C. McHardy, T. Yatsunenko, F. Niazzi, J. Affourtit, M. Egholm, B. Henrissat, R. Knight and J. I. Gordon, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 7503–7508.
- 127 I. M. de Oliveira, J. A. Henriques and D. Bonatto, *Biochem. Biophys. Res. Commun.*, 2007, **355**, 919–925.
- 128 K. A. Kim, I. H. Jung, S. H. Park, Y. T. Ahn, C. S. Huh and D. H. Kim, *PLoS One*, 2013, **8**, e62409.
- 129 A. D. Kim, K. A. Kang, H. S. Kim, D. H. Kim, Y. H. Choi, S. J. Lee, H. S. Kim and J. W. Hyun, *Cell Death Dis.*, 2013, **4**, e750.
- 130 L. Chen, Y. Meng, Q. Sun, Z. Zhang, X. Guo, X. Sheng, G. Tai, H. Cheng and Y. Zhou, *Cell Death Dis.*, 2016, **7**, e2334.
- 131 E. A. Bae, M. J. Han, E. J. Kim and D. H. Kim, *Arch. Pharmacol. Res.*, 2004, **27**, 61–67.
- 132 I. H. Jung, J. H. Lee, Y. J. Hyun and D. H. Kim, *Biol. Pharm. Bull.*, 2012, **35**, 573–581.
- 133 E. A. Bae, J. E. Shin and D. H. Kim, *Biol. Pharm. Bull.*, 2005, **28**, 1903–1908.
- 134 W. Y. Cao, Y. N. Wang, P. Y. Wang, W. Lei, B. Feng and X. J. Wang, *Molecules*, 2015, **20**, 20569–20581.
- 135 M. Nose, M. Ito, K. Kamimura, M. Shimizu and Y. Ogihara, *Planta Med.*, 1994, **60**, 136–139.
- 136 H. T. Trinh, E. H. Joh, H. Y. Kwak, N. I. Baek and D. H. Kim, *Acta Pharmacol. Sin.*, 2010, **31**, 718–724.
- 137 G. D. Kang and D. H. Kim, *J. Ethnopharmacol.*, 2016, **189**, 175–185.
- 138 E. H. Joh and D. H. Kim, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2010, **878**, 1875–1880.



- 139 N. Komoto, M. Ichikawa, S. Ohta, D. Nakano, T. Nishihama, M. Ushijima, Y. Kodera, M. Hayama, O. Shiota, S. Sekita and M. Kuroyanagi, *J. Nat. Med.*, 2010, **64**, 321–329.
- 140 I. H. Jung, S. E. Jang, E. H. Joh, J. Chung, M. J. Han and D. H. Kim, *Phytomedicine*, 2012, **20**, 84–88.
- 141 R. Kon, N. Ikarashi, C. Nagoya, T. Takayama, Y. Kusunoki, M. Ishii, H. Ueda, W. Ochiai, Y. Machida, K. Sugita and K. Sugiyama, *J. Ethnopharmacol.*, 2014, **152**, 190–200.
- 142 T. Akao, Q. M. Che, K. Kobashi, M. Hattori and T. Namba, *Biol. Pharm. Bull.*, 1996, **19**, 136–138.
- 143 E. K. Park, M. K. Choo, H. K. Yoon and D. H. Kim, *Arch. Pharmacol. Res.*, 2002, **25**, 528–533.
- 144 Y. Kawata, C. M. Ma, M. R. MeSelhy, N. Nakamura, H. Wang, M. Hattori, T. Namba, K. Satoh and Y. Kuraishi, *J. Tradit. Med.*, 1999, **16**, 15–23.
- 145 Y. S. Kim, J. J. Kim, K. H. Cho, W. S. Jung, S. K. Moon, E. K. Park and D. H. Kim, *J. Microbiol. Biotechnol.*, 2008, **18**, 1109–1114.
- 146 Y. F. Wang, Y. N. Liu, W. Xiong, D. M. Yan, Y. Zhu, X. M. Gao, Y. T. Xu and A. D. Qi, *J. Ethnopharmacol.*, 2014, **151**, 609–617.
- 147 M. Hattori, Y. Z. Shu, M. Shimizu, T. Hayashi, N. Morita, K. Kobashi, G. J. Xu and T. Namba, *Chem. Pharm. Bull.*, 1985, **33**, 3838–3846.
- 148 M. Y. Liu, M. Li, X. L. Wang, P. Liu, Q. H. Hao and X. M. Yu, *J. Agric. Food Chem.*, 2013, **61**, 12060–12065.
- 149 X. N. Li, C. H. Huo, Q. Wang, X. W. Zhang, X. N. Sheng and L. T. Zhang, *Biomed. Chromatogr.*, 2008, **22**, 367–373.
- 150 M. Zhao, J. Xu, D. W. Qian, J. M. Guo, S. Jiang, E. X. Shang and J. A. Duan, *Biomed. Chromatogr.*, 2014, **28**, 1024–1029.
- 151 L. Y. Du, M. Zhao, J. Xu, D. W. Qian, S. Jiang, E. X. Shang, J. M. Guo, P. Liu, S. L. Su, J. A. Duan and X. J. Leng, *Expert Opin. Drug Metab. Toxicol.*, 2014, **10**, 921–931.
- 152 G. J. Wang, H. X. Fu, W. Ye, X. Zheng, J. C. Xiao, D. Kang, T. Rao, Y. H. Shao, L. Xie and Y. Liang, *J. Pharm. Biomed. Anal.*, 2016, **128**, 191–200.
- 153 K. Keppler, E. M. Hein and H. U. Humpff, *Mol. Nutr. Food Res.*, 2006, **50**, 686–695.
- 154 S. Tranchimand, P. Brouant and G. Iacazio, *Biodegradation*, 2010, **21**, 833–859.
- 155 S. H. Bang, Y. J. Hyun, J. Shim, S. W. Hong and D. H. Kim, *J. Microbiol. Biotechnol.*, 2015, **25**, 18–25.
- 156 A. Braune and M. Blaut, *Environ. Microbiol.*, 2011, **13**, 482–494.
- 157 K. Ou, P. Sarnoski, K. R. Schneider, K. Song, C. Khoo and L. Gu, *Mol. Nutr. Food Res.*, 2014, **58**, 2196–2205.
- 158 J. Fu, J. Y. Ma, X. F. Zhang, Y. Wang, R. Feng, Y. C. Chen, X. S. Tan, Y. Y. Zhang, Y. P. Sun, Y. Zhou, C. Ma, C. Y. He, Z. X. Zhao and X. W. Du, *J. Pharm. Biomed. Anal.*, 2012, **71**, 162–167.
- 159 S. Yokoyama, T. Niwa, T. Osawa and T. Suzuki, *Arch. Microbiol.*, 2010, **192**, 15–22.
- 160 X. Zhang, Y. Zhao, M. Zhang, X. Pang, J. Xu, C. Kang, M. Li, C. Zhang, Z. Zhang, Y. Zhang, X. Li, G. Ning and L. Zhao, *PLoS One*, 2012, **7**, e42529.
- 161 J. Han, H. Lin and W. Huang, *Med. Sci. Monit.*, 2011, **17**, RA164–167.
- 162 W. Xie, D. Gu, J. Li, K. Cui and Y. Zhang, *PLoS One*, 2011, **6**, e24520.
- 163 C. Y. Kang, X. Zhang, M. Zhao, M. Li, L. P. Zhao and Y. F. Zhao, *Chin. J. Microecol.*, 2013, **25**, 1117–1122.
- 164 J. Q. Ruan, S. Li, Y. P. Li, W. J. Wu, S. M. Y. Lee and R. Yan, *Drug Metab. Dispos.*, 2015, **43**, 1601–1611.
- 165 S. S. Zhou, J. Xu, H. Zhu, J. Wu, J. D. Xu, R. Yan, X. Y. Li, H. H. Liu, S. M. Duan, Z. Wang, H. B. Chen, H. Shen and S. L. Li, *Sci. Rep.*, 2016, **6**, 22474.
- 166 H. Yan, J. Lu, Y. Wang, W. Gu, X. Yang and J. Yu, *Phytomedicine*, 2017, **26**, 45–54.
- 167 C. J. Chang, C. S. Lin, C. C. Lu, J. Martel, Y. F. Ko, D. M. Ojcius, S. F. Tseng, T. R. Wu, Y. M. Chen, J. D. Young and H. C. Lai, *Nat. Commun.*, 2017, **8**, 16130.
- 168 C. J. Chang, C. S. Lin, C. C. Lu, J. Martel, Y. F. Ko, D. M. Ojcius, S. F. Tseng, T. R. Wu, Y. Y. Chen, J. D. Young and H. C. Lai, *Nat. Commun.*, 2015, **6**, 7489.
- 169 M. Guo, S. Ding, C. Zhao, X. Gu, X. He, K. Huang, Y. Luo, Z. Liang, H. Tian and W. Xu, *J. Ethnopharmacol.*, 2015, **162**, 7–13.
- 170 B. Zhang, R. Yue, Y. Chen, M. Yang, X. Huang, J. Shui, Y. Peng and J. Chin, *Evid. Based Complement Altern. Med.*, 2019, **2019**, 2634898.
- 171 C. T. Peterson, V. Sharma, S. Uchitel, K. Denniston, D. Chopra, P. J. Mills and S. N. Peterson, *J. Altern. Complement. Med.*, 2018, **24**, 656–665.
- 172 P. Louis, G. L. Hold and H. J. Flint, *Nat. Rev. Microbiol.*, 2014, **12**, 661–672.
- 173 O. Ezquerro, B. Pons and M. T. Tena, *J. Chromatogr. A*, 2002, **963**, 381–392.
- 174 S. E. Pryde, S. H. Duncan, G. L. Hold, C. S. Stewart and H. J. Flint, *FEMS Microbiol. Lett.*, 2002, **217**, 133–139.
- 175 A. M. Kabat, N. Srinivasan and K. J. Maloy, *Trends Immunol.*, 2014, **35**, 507–517.
- 176 P. M. Smith, M. R. Howitt, N. Panikov, M. Michaud, C. A. Gallini, Y. M. Bohlooly, J. N. Glickman and W. S. Garrett, *Science*, 2013, **341**, 569–573.
- 177 J. Tan, C. McKenzie, M. Potamitis, A. N. Thorburn, C. R. Mackay and L. Macia, *Adv. Immunol.*, 2014, **121**, 91–119.
- 178 L. Klampfer, J. Huang, T. Sasazuki, S. Shirasawa and L. Augenlicht, *Mol. Cancer Res.*, 2003, **1**, 855–862.
- 179 S. Tedelind, F. Westberg, M. Kjerrulf and A. Vidal, *World J. Gastroenterol.*, 2007, **13**, 2826–2832.
- 180 A. Hague, D. J. Elder, D. J. Hicks and C. Paraskeva, *Int. J. Cancer*, 1995, **60**, 400–406.
- 181 M. Thangaraju, G. Cresci, S. Itagaki, J. Mellinger, D. D. Browning, F. G. Berger, P. D. Prasad and V. Ganapathy, *J. Gastrointest. Surg.*, 2008, **12**, 1773–1781; discussion 1781–1772.
- 182 N. J. Emenaker and M. D. Basson, *Dig. Dis. Sci.*, 2001, **46**, 96–105.
- 183 D. L. Worthley, V. L. Whitehall, R. K. Le Leu, N. Irahara, R. L. Buttenshaw, K. A. Mallitt, S. A. Greco, I. Ramsnes, J. Winter, Y. Hu, S. Ogino, G. P. Young and B. A. Leggett, *Dig. Dis. Sci.*, 2011, **56**, 387–396.



- 184 I. Kimura, K. Ozawa, D. Inoue, T. Imamura, K. Kimura, T. Maeda, K. Terasawa, D. Kashiara, K. Hirano, T. Tani, T. Takahashi, S. Miyauchi, G. Shioi, H. Inoue and G. Tsujimoto, *Nat. Commun.*, 2013, **4**, 1829.
- 185 A. Puddu, R. Sanguineti, F. Montecucco and G. L. Viviani, *Mediators Inflammation*, 2014, **2014**, 162021.
- 186 G. H. Rabbani, M. J. Albert, H. Rahman and A. K. Chowdhury, *Dig. Dis. Sci.*, 1999, **44**, 1547–1553.
- 187 D. R. Donohoe, N. Garge, X. Zhang, W. Sun, T. M. O'Connell, M. K. Bunger and S. J. Bultman, *Cell Metab.*, 2011, **13**, 517–526.
- 188 H. F. Mangian and K. A. Tappenden, *JPEN, J. Parenter. Enteral Nutr.*, 2009, **33**, 607–617; discussion 617.
- 189 Y. Wang, J. W. Shou, X. Y. Li, Z. X. Zhao, J. Fu, C. Y. He, R. Feng, C. Ma, B. Y. Wen, F. Guo, X. Y. Yang, Y. X. Han, L. L. Wang, Q. Tong, X. F. You, Y. Lin, W. J. Kong, S. Y. Si and J. D. Jiang, *Metabolism*, 2017, **70**, 72–84.
- 190 J. L. Hu, S. P. Nie, F. F. Min and M. Y. Xie, *J. Agric. Food Chem.*, 2012, **60**, 11525–11532.
- 191 B. Thomas, B. S. M. Vithiya, T. A. A. Prasad, S. B. Mohamed, C. M. Magdalane, K. Kaviarasu and M. Maaza, *J. Nanosci. Nanotechnol.*, 2019, **19**, 2640–2648.
- 192 W. Feng, H. Ao and C. Peng, *Front. Pharmacol.*, 2018, **9**, 1354.
- 193 V. Jaswal, J. Palanivelu and C. Ramalingam, *Biochem. Biophys. Rep.*, 2018, **14**, 125–132.

