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Synthesis of a novel resorcin[4]arene–glucose conjugate and its catalysis of the CuAAC reaction for the synthesis of 1,4-disubstituted 1,2,3-triazoles in water†

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The Cu(I)-catalyzed azide–alkyne cycloaddition (CuAAC) in aqueous media using resorcin[4]arene glycoconjugate (RG) is reported. The eight β-D-glucopyranoside moieties constructed on the resorcin[4]arene upper rim provide a pseudo-saccharide cavity that offers a suitable host environment for water-insoluble hydrophobic azido and/or alkyne substrates in water. The utility of RG was established as an efficient inverse phase transfer catalyst for the CuAAC in water as a green approach for the synthesis of 1,4-disubstituted 1,2,3-triazole species. The catalytic utility of RG (1 mol%) was demonstrated in a multicomponent one-pot CuAAC for various azido/alkyne substrates. The RG acts as a molecular host and a micro-reactor resulting in the 1,4-disubstituted 1,2,3-triazoles in excellent yield.

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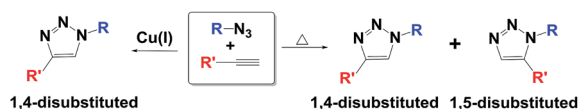
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Introduction

The classical Huisgen¹ cycloaddition reaction for the synthesis of 1,2,3-triazole involves thermal 1,3-dipolar cycloaddition of organic azides with alkynes, though in low yield and mixed regioselectivity. Sharpless² and Meldal's³ research groups later independently developed the improved procedure involving the copper(I)-catalyzed Huisgen 1,3-dipolar cycloaddition reaction, which is the widely studied 'click' reaction (Scheme 1). The impact of the copper catalyzed azide–alkyne click reaction in various branches of science is increasing exponentially as evidenced from numerous recent reviews available in the literature since 2010.⁴

The three most common facile protocols for CuAAC include (i) use of copper(I) salts (mostly in organic solvents), (ii) the reduction of a copper(II to I), and (iii) oxidation of Cu(0 to I). Of the three protocols described above, the method employing *in situ* reduction of copper(II) salts is known to be more practical



Scheme 1 Azide–alkyne cycloaddition (AAC) reaction under different reaction conditions.

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and can be carried out in aqueous conditions. From review of literature, it is easy to conclude that water is an appropriate choice as a solvent for the CuSO₄/sodium ascorbate catalyzed click protocol, which results in the formation of the triazole in high yields and with excellent regioselectivity. However, despite the efficiency of the CuAAC reaction, there are limitations to using the procedure especially when the substrates are not water-soluble. The protocol in essence requires deoxygenated conditions in the presence of mixed aprotic organic solvents such as THF, CH₃CN, CH₂Cl₂, toluene, *etc.* and due to the oxidative tendency of the copper(I), a higher catalyst concentration throughout the reaction is needed. To stabilize the catalyst, several phosphine-based complexes and amine-based (bound with different heterocyclic donors) ligands have been used for rate acceleration.⁵ Additionally, a number of heterogeneous Cu catalysts,⁶ including amberlyst resin-supported,⁷ polymer-supported,⁸ and zeolite-supported⁹ have been explored to catalyze the triazole formation. To speed up the azide–alkyne reaction, use of surfactants and phase transfer catalyst,¹⁰ the microwave¹¹ and ultrasound irradiations¹² have also been reported.¹³

Recently, our research group reported resorcin[4]arene cavity glycoconjugates (RCGs)¹⁴ as inverse phase transfer catalysts with abilities to catalyze organic reactions in aqueous media. We also reported on the RCGs ability to catalyze the formation of 1,4-disubstituted 1,2,3-triazoles in water without the addition of any co-organic solvents.¹⁴ It is noteworthy to mention that we were the first to establish the concept of the spatial directionality of β-D-glucopyranoside units on the resorcin[4]arene rigid structure “cavitand”. The RCGs possesses a unique molecular host system “pseudo-saccharide bucket”



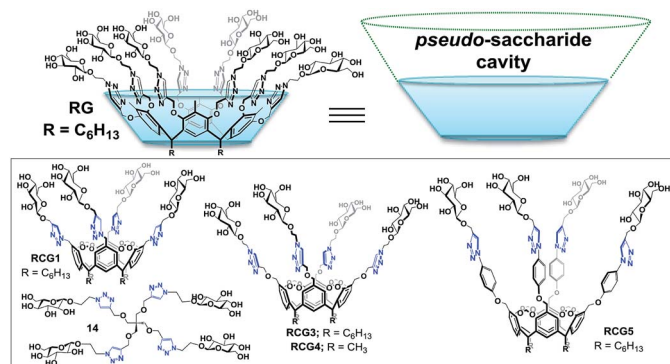


Fig. 1 Resorcin[4]arene glycoconjugate (RG & RCGs¹⁴).

which can encapsulate organic substrates and catalyze chemical reactions in water.¹⁴

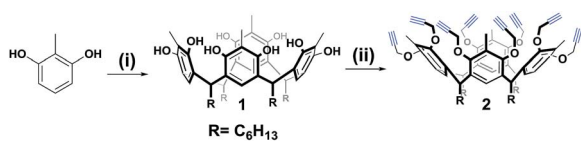
In this manuscript, we describe the synthesis of a resorcin[4]arene glycoconjugate (**RG**) (Fig. 1) and its application as a micro-reactor for the synthesis of 1,4-disubstituted 1,2,3-triazole species in aqueous media *via* the CuAAC reaction. **RG** structure consists of eight β -D-glucopyranoside moieties constructed on the phenolic parts on the resorcin[4]arene upper rim *via* multiple 1,4-disubstituted 1,2,3-triazole linkages. The eight arm resorcin[4]arene glycoconjugate offers an enlarged flexible pseudo-saccharide cavity that can act as a molecular vessel for water-insoluble azido and/or alkyne substrates in aqueous environment.

Results and discussion

Synthesis of eight arm resorcin[4]arene glycoconjugate (RG)

For the synthesis of the novel **RG**, resorcin[4]arene **1** (ref. 14 and 15) was first synthesized upon the acid-catalyzed cycl-condensation reaction of methyl resorcinol with heptanal. Compound **1** was then treated with propargyl bromide in the presence of potassium carbonate in refluxing acetone to achieve the octa-propargyl resorcin[4]arene intermediate **2** (Scheme 2).

Resorcin[4]arene **2** was characterized conclusively from its NMR and spectral data. In its ¹H-NMR spectrum, the benzylic protons ($-\text{CH}_3$, **H_a**) were found as a singlet at 2.25 ppm. The alkyne protons ($-\text{C}\equiv\text{CH}$, **H_f**) were observed as a triplet at 2.50 ppm ($J = 2.4$ Hz) and the propargyl methylene protons ($-\text{OCH}_2\text{C}\equiv$, **H_{c,c'}**) showed as two set of double doublets with $J = 15.3$ and 2.4 Hz at 4.20 and 4.36 ppm. Its ¹³C-NMR spectrum had resonances for the alkyne carbons **C_e** and **C_f** at 74.8 and 79.5 ppm, respectively, and the propargyl methylene carbons ($-\text{OCH}_2\text{C}\equiv$, **C_c**) were at 60.3 ppm (Fig. 2). The molecular formula ($\text{C}_{80}\text{H}_{96}\text{O}_8$) of compound **2** was confirmed from its *m/z*



(i) heptaldehyde, HCl (37%), ethanol, reflux, overnight; (ii) propargyl bromide, K_2CO_3 , acetone, reflux

Scheme 2 Synthesis of octa-alkyne resorcin[4]arene **2**.

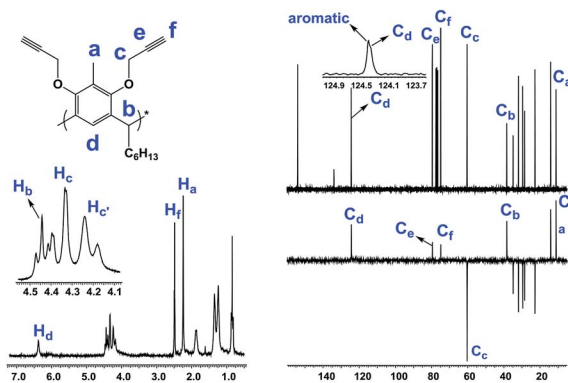
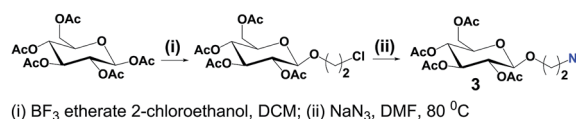
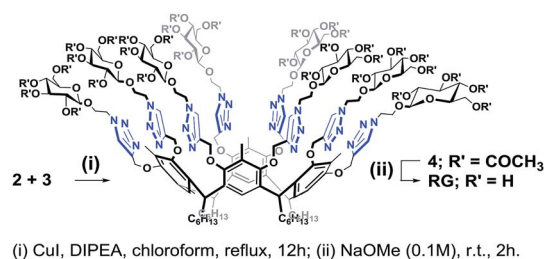


Fig. 2 Partial ¹H-, ¹³C- and DEPT NMR spectra (250 and 62.5 MHz, CDCl_3) of **2**.



Scheme 3 Synthesis of **3**.



Scheme 4 Synthesis of **RG**.

measurement using ESI-Q-TOF HRMS: observed 1207.7034 ($\text{M} + \text{Na}$)⁺, calculated 1207.7003 ($\text{M} + \text{Na}$)⁺.

2-Azidoethyl β -D-glucopyranoside tetraacetate (**3**) was prepared following the typical procedures found in literature (Scheme 3) and unambiguously characterized.^{14,16}

Octa-propargyl resorcin[4]arene **2** and azido glucopyranoside **3** were then coupled together *via* the CuAAC reaction. The reaction was carried out in refluxing chloroform in presence of CuI (10 mol%) and DIPEA (6.0 equiv.) to yield the octa-acetoxy-**RG** (**4**) in excellent yield. Global deacetylation using NaOMe solution (0.1 M) resulted in **RG** in gram quantity (Scheme 4).

The structures of the **RG** and its octa-acetoxy precursor **4** were established from the respective ¹H- and ¹³C-NMR data. Namely, the absence of the alkyne protons (**H_f**), the emergence of the triazole protons (**1H**, at 7.69 ppm) and the shift of the propargyl protons (**H_c**, from 4.11 ppm to 4.50 ppm) confirmed the structure of **4** (Fig. 3). In the ¹H-NMR spectra of **RG** (Fig. 3b), the disappearance of the acetate protons ($-\text{OCOCH}_3$, **OAc**) from the region of 1.70–2.10 ppm was confirmatory of its structure. The molecular weight of the products {for **4** *m/z* (ESI-Q-TOF): observed 2283.8955 ($\text{M} + 2\text{Na}$)²⁺, calculated 2283.8984 ($\text{M} +$



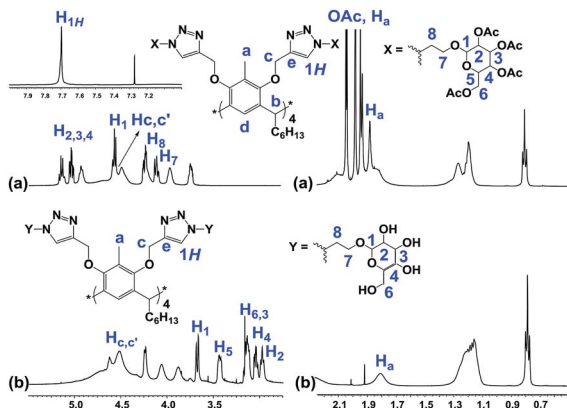


Fig. 3 Partial $^1\text{H-NMR}$ spectra of (a) **4** (500 MHz, CDCl_3) and (b) **RG** (500 MHz, DMSO-d_6).

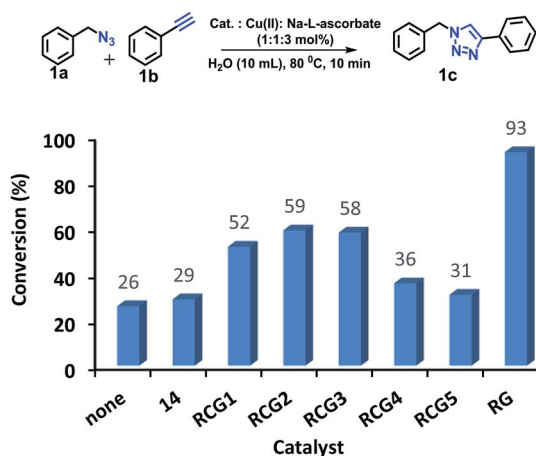


Fig. 4 CuAAC of benzyl azide and phenyl acetylene at $80\text{ }^\circ\text{C}$. RCG catalyst structures in Fig. 1.

$2\text{Na})^{2+}$; for **RG** m/z (ESI-Q-TOF): observed 1611.7374 ($\text{M} + 2\text{Na})^{2+}$, calculated 1611.7294 ($\text{M} + 2\text{Na})^{2+}$) confirmed the respective molecular formula.

Optimizing CuAAC in water catalyzed by RG

The CuAAC reactions were investigated to examine the catalytic activity in aqueous environment. For optimization, the CuAAC reaction of benzyl azide (1.0 mmol) with phenyl acetylene (1.05 equiv.) with/without the addition of a catalyst (1 mol%) was carried out (Fig. 4). The reactions were performed in the presence of copper sulfate (1 mol%) and sodium L-ascorbate (3 mol%) in 10 mL of distilled deionized water at $80\text{ }^\circ\text{C}$ (Fig. 4). The chemical structures and synthetic procedure for RCG catalysts evaluated in Fig. 4 have been previously reported by our research group.¹⁴

The coupling reaction was much slower in absence of added RCG catalyst and in presence of compound **14** (Fig. 1),¹⁴ which lacks the spatial directionality of the RCGs. While the RCGs catalyzed the CuAAC reaction between benzyl azide and phenyl acetylene, there were significant differences in substrate conversion to suggest a dependence of catalytic activity on

Table 1 CuAAC of substituted benzyl azides **1a–4a** and alkyne substrates **1b–4b** in the presence of **RG**^a

	1a–4a	1b–4b	1c–16c	
Substrates (c log <i>P</i> value)				
Product (time, yield ^b)	1a (3.18)	2a (3.26)	3a (3.71)	4a (4.13)
1b (0.344)	12c (15 minutes, 94%)	4c (15 minutes, 95%)	8c (15 minutes, 95%)	16c (20 minutes, 91%)
	10c (25 minutes, 94%)	2c (20 minutes, 98%)	6c (25 minutes, 98%)	14c (45 minutes, 92%)
	9c (20 minutes, 98%)	1c (20 minutes, 98%)	5c (25 minutes, 99%)	13c (30 minutes, 99%)
4b (2.51)	11c (45 minutes, 91%)	3c (45 minutes, 99%)	7c (45 minutes, 92%)	15c (65 minutes, 94%)

^a Reaction condition: benzyl azide derivative (1 mmol) and alkyne substrates (1.05 mmol), Cu(II) (1 mol%), Na-L-ascorbate (3 mol%), **RG** (1 mol%), water (10 mL), $80\text{ }^\circ\text{C}$. ^b Isolated yield.

chemical structure (Fig. 4). Remarkably, the CuAAC reaction was almost completed with more than 93% conversion only in 10 minutes when **RG** (1 mol%) was added but only 26% conversion was observed in its absence (no catalyst). Obviously, the fast CuAAC in the presence of **RG** indicates that it provides a unique molecular environment that is capable of catalyzing the CuAAC reaction efficiently.

Scoping the CuAAC in water using RG

To scope the CuAAC reactions catalyzed by **RG** in aqueous media, we have investigated coupling of substituted benzyl azides **1a–4a** with aromatic and aliphatic alkynes **1b–4b** (Table 1).

As recorded in Table 1, the **RG** catalyzed CuAAC reactions of a variety of substituted benzyl azides in water and all reactions led to the desired 1,4-disubstituted 1,2,3-triazole products (**2c–16c**) in high yields (>90% isolated). Not surprisingly, it was determined that the reaction took longer to reach completion as the hydrophobicity of the substrate pair increased, as evidenced by their clog *P* values (Table 1). For example, the coupling between the 4-bromo benzyl azide (**4a**, clog *P* = 4.13) and 1-hexyne (**4b**, clog *P* = 2.51) took nearly 65 minutes to completion while the reactions between propargyl alcohol (**1b**, clog *P* = -0.34) and 3-methoxy benzyl azide (**1a**, clog *P* = 3.18) was completed in 15 minutes.

To further evaluate the effectiveness of the **RG** as a catalyst in CuAAC of hydrophobic substrates, we carried out the coupling of phenyl acetylene (**3b**) with alkylated *ortho*-azido phenols (**5a–9a**) of increasing steric bulk and clog *P* values from 2.55–4.95. A



Table 2 CuAAC of *o*-azido phenol derivatives (**5a–9a**) with phenyl acetylene **1b** with/without **RG**^a

Entry	Azide (clog <i>P</i> value)	Product	Time (min)	Catalyst (% yield) ^b	
				None	RG
1	5a R = OH (2.55)	17c	30	64	96
2	6a R = OMe (3.18)	18c	30	22	97
3	7a R = OEt (3.71)	19c	30	16	98
4	8a R = O <i>n</i> Bu (4.77)	20c	30	12	98
5	9a R = O <i>n</i> Bn (4.95)	21c	30	<10	99

^a Reaction condition: azidophenol derivative (1 mmol) and phenyl acetylene **1b** (1.05 mmol), Cu(II) (1 mol%), Na-L-ascorbate (3 mol%), cat. (1 mol%), water (10 mL), 80 °C. ^b Isolated yield.

comparative study of the CuAAC reaction in absence and presence of the **RG** is shown in Table 2.

As expected, the coupling of *o*-azido phenols (**5a**, clog *P* = 2.44) with phenyl acetylene **1b** catalyzed by **RG** resulted in triazole **17c** in 96% yield and even without added catalyst the yield was 64% due to its higher hydrophilicity (entry 1). However, replacing the hydroxyl functionality with alkoxy groups, *i.e.* –OMe, –OEt, –O*n*Bu, –OPh of progressively higher bulk, hydrophobicity, and clog *P* value led to much slower reactions in absence of the **RG**. The reactions catalyzed by the **RG** resulted in near quantitative conversion in about 30 minutes irrespective of the bulk or hydrophobicity of the azide substrates, attesting to its effectiveness as a catalyst in the CuAAC reactions in water even for the much bulkier and hydrophobic substrates.

Di-CuAAC reactions in water using **RG**

Simultaneous multiple CuAAC reactions have found interest in synthesis of the polymers and dendrimers and the need for efficient catalyst that can catalyze reactions in water has never been greater.¹⁷ We have therefore investigated **RG** for catalyzing the di-CuAAC reactions in water of di-propargyl benzene derivatives **5b–7b** with substituted benzyl azides **1a–4a** (Table 3). Interestingly, the di-CuAAC reaction were completed within 45 min resulting in the desired bis-1,2,3-triazole products **22c–33c** in gram quantities.

Multicomponent one-pot CuAAC reactions in water

Organic azides are not always considered safe for handling because of their toxic and shock sensitive nature and there have

Table 3 Di-CuAAC of di-alkynes **5b–7b** with benzyl azide derivatives **1a–4a** in water in the presence of **RG**^{a,b}

Product	Time (min)	Yield (%)
22c	45min	95%
23c	45min	91%
24c	45min	94%
25c	60min	92%
26c	45min	94%
27c	45min	92%
28c	45min	91%
29c	60min	93%
30c	45min	96%
31c	45min	96%
32c	45min	92%
33c	60min	92%

^a Reaction condition: di-alkyne derivative (1 mmol) and azides (2.1 mmol), Cu(II) (1 mol%), Na-L-ascorbate (3 mol%), **RG** (1 mol%), water (10 mL), 45 min, 80 °C. ^b Isolated yields.

been alternative methods employed for their *in situ* synthesis. The azides can be prepared from their corresponding halides upon the addition of sodium azide. Hence, a multicomponent one-pot CuAAC reaction between *in situ* generated azide from its corresponding precursor and alkyne is highly desirable.¹⁸

We have investigated the multicomponent one-pot CuAAC reactions of phenyl acetylene **3b** with the aryl bromides and sodium azide catalyzed by the **RG**. Complete conversion were accomplished within 25–55 minutes even for a bulkier aryl system (naphthyl) to achieve the 1,2,3-triazoles in excellent isolated yield (90–96%) (Table 4, entries 1–5). In addition, reaction with α -bromo acetophenones (entries 6 and 7), α -bromoesters (entry 8) and ally bromide (entry 9) led to quantitative conversion within 25–30 min to the desired triazoles **35c–37c** in 91–95% isolated yield. However, the multi component CuAAC failed when saturated aliphatic bromides were used as substrates, *i.e.*, butyl, heptyl and dodecyl bromides (entries 10–12). We suspect the π – π interactions play an important role in the encapsulation, in water, of the substrates molecule in the hydrophobic cavity of the **RG** and the lack of these interactions in the aliphatic alkyl bromide does not allow their encapsulation in the **RG** cavity and hence no observed reaction.

Inclusion complex of **RG** with benzyl azide (**2a**) and phenyl acetylene (**3b**) in D₂O

We set out to probe the encapsulation of the substrates by the **RG**. Specifically, we have studied the encapsulation of benzyl azide and phenyl acetylene by **RG** using NMR. The ¹H-NMR spectra of the guest (**2a** & **3b**) were recorded in the presence of the host (**RG**) in a 1 : 1 molar ratio for 2 mM concentrated solution in D₂O at 25 °C (Fig. 5). In the ¹H-NMR, the aromatic protons (**H**₂, **H**₃, **H**₄) in **2a** were shifted up field from 7.35 ppm to 6.90 ppm upon addition of the **RG** (Fig. 5a). Similarly, the aromatic protons (**H**₆, **H**₇, **H**₈) in **3b** were shifted upfield from 7.35 and 7.45 ppm to 6.98



Table 4 Multicomponent one-pot CuAAC of phenyl acetylene, aryl/alkyl bromides and sodium azides in the presence of **RG**^a

Entry	Product	Time (min)	Yield ^b (%)
1		25	92
2		30	94
3		30	96
4		50	90
5		55	91
6		30	92
7		30	95
8		25	93
9		25	91
10		200	<10
11		200	N/A
12		200	N/A

^a Reaction condition: phenyl acetylene (1 mmol), bromide derivative (1.05 mmol) and sodium azide (1.1 mmol), **RG** (1 mol%), water (10 mL), 80 °C. ^b Isolated yields, N/A = not isolated.

and 7.15 ppm, respectively upon addition of **RG** (Fig. 5b). The shielding of the guest ¹H NMR resonances upon addition of **RG** indicated their encapsulation in the **RG** cavity. The encapsulation of the substrates in the **RG** cavity may explain the catalytic activity of the **RG**. In addition, the observation that the CuAAC reactions of the aromatic and π bond containing azides were accelerated in the presence of **RG** (Table 4), the encapsulation of the guest most likely involves π - π host-guest interactions.

Proposed mechanism for the CuAAC using **RG**

Based upon the reaction catalysed and the encapsulation observed, the proposed mechanism for the CuAAC reaction catalyzed by **RG** may proceed *via* step shown in Fig. 6. Starting with inclusion of the alkyne and azide substrates in the pseudo- β -D-glucopyranoside cavity of **RG**. The closed proximity of the alkyne and azide in the presence of Cu(I) catalyst accelerate the cycloaddition process resulting in the copper-triazole complex followed by protonation and dissociation of the desired 1,4-di substituted 1,2,3-triazole (Fig. 6).⁴⁹ The binding of the Cu(I) to the multiple triazoles in the **RG** may also facilitate the coupling reaction inside the **RG** cavity.

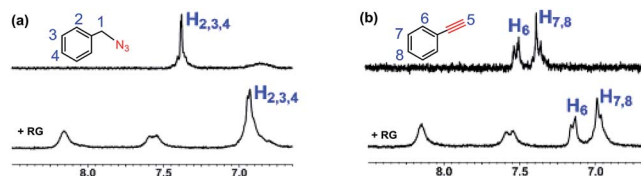


Fig. 5 Partial ¹H-NMR spectra (250 MHz, D₂O, 2 mM) at 25 °C of (a) benzyl azide and benzyl azide + **RG** (1 : 1); (b) phenylacetylene and phenylacetylene + **RG** (1 : 1).

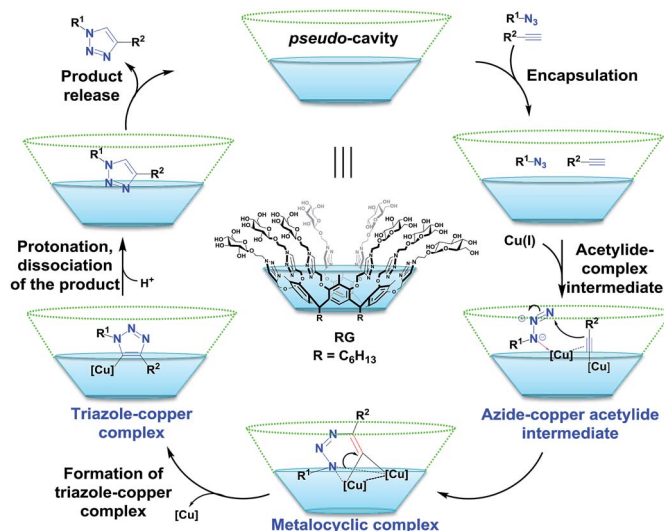


Fig. 6 Proposed mechanism for the CuAAC in aqueous media catalyzed by **RG**.

Conclusion

In conclusion, a new resorcinarene sugar conjugate (**RG**) is reported. It was determined that **RG** is an efficient catalyst for the CuAAC reaction in water at only 1 mol%. Based upon the reaction catalysis and the encapsulation observed, **RG** catalytic mechanism, we believe, begins with inclusion of the alkyne and azide substrates resulting in the copper-triazole complex followed by protonation and dissociation to the desired 1,4-di substituted 1,2,3-triazole. **RG** was found to catalyze the coupling of the alkyne/azide pair and also of the multicomponent alkyl bromide/sodium azide/alkyne to the 1,4-disubstituted 1,2,3-triazole products in excellent yield within short period of time in water.

Experimental section

General

¹H- and ¹³C-NMR spectra were recorded on a Bruker DRX-250, a Inova-400 and a DD-500 spectrometers. Sample concentrations were about 10% (w/v) in CDCl₃ or DMSO-d₆ and the *J* values are given in Hz. The mass spectral analyses were performed on an Aligent Technologies 6540 UHD Accurate-Mass Q-TOF LC/MS. The clog *P* values were calculated using ChemDraw Professional, Version 15.1.0.144.



Materials and reagents

All reagents were used with no further purification unless otherwise specified. 2-Methyl resorcinol (98%) was purchased from Acros Organics Chemical Company. Octa-hydroxy resorcin [4]arene (**1**) and 2-azidoethyl β -D-glucopyranoside tetraacetate (**3**) were synthesized following synthetic procedures reported previously.^{14–16}

Octa-propargyl resorcin[4]arene (2). Compound **1** (5 g, 5.7 mmol) was dissolved in 110 mL acetone. Potassium carbonate (6.26 g, 45 mmol) was added into the solution and allowed to stir for 10 min at room temperature. Propargyl bromide (8.1 mL, 90 mmol) was then added and the reaction mixture was refluxed overnight. After completion, the reaction mixture was cooled at room temperature and the salt was filtered out followed by the concentration of acetone. The product was purified by column chromatography as a pale yellow solid using 10% EA/hexane yielding 6.04 g (90%) yield. ¹H NMR (250 MHz, CDCl₃) δ = 0.85 (t, J = 6.5 Hz, 12H), 1.25–1.36 (m, 16H), 1.36 (m, 16H), 1.82–1.94 (m, 8H), 2.25 (s, 12H), 2.50 (t, J = 2.4 Hz, 4H), 4.11 (d, J = 15.3 Hz, 8H), 4.25 (d, J = 15.3 Hz, 8H), 4.45 (t, J = 7.1 Hz, 4H), 6.39 (s, 4H); ¹³C NMR (62.5 MHz, CDCl₃) δ = 11.1, 14.0, 22.7, 28.5, 29.5, 31.9, 34.9, 38.3, 60.3, 74.8, 79.5, 124.3, 124.3, 133.9, 154.0; HRMS [M + Na]⁺ calcd for C₈₀H₉₆O₈Na 1207.7003, found 1207.7034.

Octa-sugar acetate resorcin[4]arene (6). Compound **2** (1 g, 0.84 mmol) was dissolved in 30 mL chloroform. CuI (32 mg, 0.17 mmol) and DIPEA (0.88 mL, 5 mmol) were then added to the solution. Sugar azide **3** (5.6 g, 13.5 mmol) was then added to the solution. The reaction mixture was then refluxed overnight. After completion, the reaction was worked up using ammonium hydroxide solution and the organic phase was extracted and dried using Na₂SO₄. Chloroform was evaporated and the product was purified by column chromatography using 5% MeOH/DCM in order to separate the product from the excess sugar azide. The product was collected as yellow oil that solidified slowly to pale yellow solid in 3.48 g (91%) yield. ¹H NMR (500 MHz, CDCl₃) δ = 0.80 (t, J = 6.6 Hz, 12H), 1.15–1.35 (m, 32H), 1.78–1.85 (bs, 12H), 1.88 (s, 12H), 1.93 (s, 12H), 1.98 (s, 24H), 2.04 (s, 12H), 2.04 (s, 12H), 2.07–2.16 (bs, 12H), 2.37 (bm, 8H), 3.71–3.77 (m, 8H), 3.91–4.00 (m, 8H), 4.06–4.14 (m, 8H), 4.22 (t, J = 2.9 Hz, 8H), 4.25 (dd, J = 2.9, 4.4 Hz, 4H), 4.49 (bs, 16H), 4.55 (d, J = 8.1 Hz, 8H), 4.58 (t, J = 8.1 Hz, 4H), 4.93 (t, J = 7.7 Hz, 8H), 5.03 (ddd, J = 3.3, 9.5, 13.2 Hz, 8H), 5.13 (q, J = 9.9, 19.1 Hz, 8H), 7.69 (s, 8H); ¹³C NMR (125 MHz, CDCl₃) δ = 10.7, 14.0, 20.3, 20.4, 20.4, 20.6, 22.7, 28.6, 29.6, 31.8, 35.2, 38.0, 49.5, 61.6, 65.9, 67.5, 68.1, 70.7, 71.7, 72.4, 100.4, 123.7, 124.1, 124.2, 144.3, 144.5, 169.3, 170.0, 170.5; HRMS [M + 2Na]²⁺ calcd for C₂₀₈H₂₈₀N₂₄O₈₈Na₂ 2283.8984, found 2283.8955.

Resorcin[4]arene glycoconjugate (RG). Acetylated sugar resorcin[4]arene **6** (1 g) was dissolved in (0.1 M) sodium methoxide solution. The reaction mixture was allowed to stir at room temperature for 4 h. After completion, the reaction mixture was neutralized using Dowex 50W-X8. This was followed by filtration and concentration of methanol with no further purification to afford **RG** as a pale yellow solid in 92%;

¹H NMR (250 MHz, DMSO-d₆) δ = 0.79 (t, J = 6.6 Hz, 12H), 1.14–1.36 (m, 32H), 2.27 (bm, 8H), 2.27 (bs, 12H), 2.98 (t, J = 8.1 Hz, 8H), 3.05 (ddd, J = 3.3, 9.2, 18.3 Hz, 8H), 3.11–3.19 (m, 16H), 3.41–3.47 (m, 8H), 3.67 (d, J = 10.6 Hz, 8H), 3.89 (bs, 8H), 4.07 (bs, 8H), 4.24 (d, J = 7.0 Hz, 8H), 4.53 (bs, 24H), 4.63 (bs, 16H), 7.93–8.39 (bs, 12H); ¹³C NMR (62.5 MHz, DMSO-d₆) δ = 11.1, 14.0, 22.2, 28.1, 29.1, 31.4, 34.8, 37.8, 49.6, 61.1, 65.4, 67.3, 70.0, 73.3, 76.6, 77.0, 102.9, 123.8, 124.3, 124.7, 142.9, 143.1; HRMS [M + 2Na]²⁺ calcd for C₁₄₄H₂₁₆N₂₄O₅₆Na₂ 1611.7294, found 1611.7374.

Typical procedure for screening and scoping the CuAAC in water catalyzed by RG

Aryl azide (1 mmol) and aryl/alkyl alkyne (1.05 equiv.) were added to a solution of copper(II) sulfate pentahydrates, Na-ascorbate and **RG** (1 : 3 : 1 mol%) in 10 mL de-ionized distilled water. The reaction mixture was then heated to 80 °C for 15–65 min. After completion, the solid reaction product was filtered off, dried, and weighed to calculate the isolated yield. When the product was not solid (**10c** and **11c**), the reaction mixture was extracted with DCM (2 × 5 mL). The combined organic layer was collected and dried over MgSO₄. DCM was then removed using a rotary evaporator and ¹H-NMR was taken in CDCl₃ or DMSO-d₆.

Typical procedure for three component one-pot CuAAC catalyzed by RG

Phenyl acetylene **1b** (1 mmol) and aryl/alkyl bromides (1.05 equiv.) with sodium azide (1.1 equiv) were added to a solution of copper(II) sulfate pentahydrates, Na-ascorbate and **RG** (1 : 3 : 1 mol%) in 10 mL de-ionized distilled water. The reaction mixture was then heated to 80 °C for 20–55 min. After completion, the solid reaction product was filtered off, dried, and weighed to calculate the isolated yield. When the conversion was low (entries 10–12, Table 4), the reaction mixture was extracted with DCM (2 × 5 mL). The combined organic layer was collected and dried over MgSO₄. The solvent was then removed using a rotary evaporator and ¹H-NMR was taken in CDCl₃.

4-((Benzyloxy)methyl)-1-(2-methylbenzyl)-1H-1,2,3-triazole (6c). White solid (98%); ¹H NMR (400 MHz, CDCl₃) δ 2.25 (s, 3H), 4.55 (s, 2H), 4.62 (s, 2H), 5.48 (s, 2H), 7.11–7.13 (m, 1H), 7.15–7.21 (m, 2H), 7.24 (d, J = 7.0 Hz, 2H), 7.29 (m, 4H), 7.33 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 18.9, 52.2, 63.6, 72.4, 126.5, 127.7, 127.8, 128.3, 129.0, 129.4, 130.9, 132.3, 136.8, 137.6.

4-Butyl-1-(2-methylbenzyl)-1H-1,2,3-triazole (7c). White solid (90%); ¹H NMR (400 MHz, CDCl₃) δ 0.87 (t, J = 7 Hz, 3H), 1.27–1.36 (m, 2H), 1.54–1.61 (m, 2H), 2.23 (s, 3H), 2.64 (t, J = 7.8 Hz, 2H), 5.45 (s, 2H), 7.15–7.18 (m, 2H) 7.22–7.26 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 13.7, 18.9, 22.2, 25.3, 31.4, 51.0, 120.2, 126.5, 128.8, 129.1, 130.8, 132.7, 136.8.

4-((Benzyloxy)methyl)-1-(3-methoxybenzyl)-1H-1,2,3-triazole (10c). Brown oil (94%); ¹H NMR (400 MHz, CDCl₃) δ 3.71 (s, 2H), 4.54 (s, 2H), 4.62 (s, 2H), 5.40 (s, 2H), 6.77 (s, 1H), 6.79–6.85 (m, 2H), 7.20–7.26 (m, 2H), 7.28–7.30 (m, 3H), 7.47 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 53.6, 54.9, 63.3, 72.1, 113.4, 113.7, 119.9, 122.3, 127.4, 127.5, 128.0, 135.8, 137.5, 145.1, 159.7.



4-Butyl-1-(3-methoxybenzyl)-1H-1,2,3-triazole (11c). Brown oil (91%) yield; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 0.78 (t, $J = 7.3$ Hz, 3H), 1.19–1.28 (m, 2H), 1.46–1.54 (m, 2H), 2.56 (t, $J = 7.7$ Hz, 2H), 3.61 (s, 3H), 5.31 (s, 2H), 6.66 (s, 1H), 6.71 (t, $J = 9.0$ Hz, 2H), 7.12 (t, $J = 7.9$ Hz, 1H), 7.19 (s, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 13.3, 21.8, 24.9, 31.0, 53.2, 54.7, 113.0, 113.4, 119.6, 120.3, 129.5, 136.2, 148.2, 159.5.

1-(2-Ethoxyphenyl)-4-phenyl-1H-1,2,3-triazole (19c). White solid (98%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 1.37 (t, $J = 6.9$ Hz, 3H), 4.08 (q, $J = 6.9$ Hz, 2H), 7.04 (m, 2H), 7.34 (m, 2H), 7.43 (t, $J = 7.5$ Hz, 2H), 7.81 (d, $J = 7.8$ Hz, 1H), 7.89 (d, $J = 7.6$ Hz, 2H), 8.38 (s, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 14.5, 64.6, 113.2, 121.0, 121.6, 125.0, 125.6, 126.3, 127.9, 128.7, 129.8, 130.7, 147.0, 150.1.

1-(2-Butoxyphenyl)-4-phenyl-1H-1,2,3-triazole (20c). White solid (98%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 0.93 (t, $J = 6.8$ Hz, 3H), 1.40–1.47 (m, 2H), 1.72–1.78 (m, 2H), 4.05 (t, $J = 6.3$ Hz, 2H), 7.08 (t, $J = 8.9$ Hz, 2H), 7.32–7.39 (m, 2H), 7.44 (t, $J = 8.0$ Hz, 2H), 7.83 (d, $J = 7.8$ Hz, 1H), 7.88 (d, $J = 8.1$ Hz, 2H), 8.36 (s, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 13.7, 19.2, 31.0, 68.7, 113.2, 121.0, 121.7, 125.2, 125.7, 126.4, 128.0, 128.8, 129.9, 130.7, 147.0, 150.4.

1,2-Bis((1-benzyl-1H-1,2,3-triazol-4-yl)methoxy) benzene (22c). White solid (95%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 5.15 (s, 4H), 5.44 (s, 4H), 6.85–6.90 (m, 2H), 6.95–7.00 (m, 2H), 7.19 (d, $J = 1.9$ Hz, 2H), 7.20 (d, $J = 3.8$ Hz, 2H), 7.30 (d, $J = 1.9$ Hz, 3H), 7.31 (d, $J = 1.7$ Hz, 2H), 7.58 (s, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 54.1, 63.5, 115.7, 122.2, 128.0, 128.7, 129.0, 134.5, 148.4.

1,2-Bis((1-(2-methylbenzyl)-1H-1,2,3-triazol-4-yl)methoxy) benzene (23c). White solid (94%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 2.21 (s, 6H), 5.12 (s, 4H), 5.43 (s, 4H), 6.83–6.87 (m, 2H), 6.93–6.98 (m, 2H), 7.03 (d, $J = 7.6$ Hz, 2H), 7.11–7.16 (m, 4H), 7.20–7.24 (m, 2H), 7.47 (bs, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 18.8, 63.4, 115.6, 122.0, 126.4, 128.9, 129.1, 130.8, 132.4, 136.6, 148.3.

1,2-Bis((1-(3-methoxybenzyl)-1H-1,2,3-triazol-4-yl)methoxy) benzene (24c). White solid (94%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 3.72 (s, 6H), 5.12 (s, 4H), 5.41 (s, 4H), 6.74 (s, 2H), 6.78 (m, 4H), 6.88 (bs, 2H), 6.98 (bs, 4H), 7.22 (t, $J = 8.4$ Hz, 2H), 7.71 (bs, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 54.3, 55.1, 63.5, 113.5, 113.9, 115.4, 120.1, 121.9, 129.9, 135.7, 148.2, 159.7.

1,2-Bis((1-(4-bromobenzyl)-1H-1,2,3-triazol-4-yl)methoxy) benzene (25c). Yellow solid (96%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 5.16 (s, 4H), 5.45 (s, 4H), 6.90–6.93 (m, 2H), 6.99–7.02 (m, 2H), 7.09 (d, $J = 8.7$ Hz, 4H), 7.44 (d, $J = 7.7$ Hz, 4H), 7.70 (bs, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 53.5, 115.6, 122.2, 122.7, 129.6, 132.1, 133.5, 148.4.

1,3-Bis((1-benzyl-1H-1,2,3-triazol-4-yl)methoxy) benzene (26c). Yellow solid (94%) yield; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 5.11 (s, 4H), 5.61 (s, 4H), 6.61 (d, $J = 2.2$ Hz, 1H), 6.63 (d, $J = 2.2$ Hz, 1H), 6.71 (t, $J = 2.3$ Hz, 1H), 7.18 (t, $J = 8.2$ Hz, 1H), 7.30–7.40 (m, 10H), 8.28 (s, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 52.8, 61.1, 101.6, 107.3, 124.6, 127.9, 128.1, 128.7, 135.9, 159.2.

1,3-Bis((1-(2-methylbenzyl)-1H-1,2,3-triazol-4-yl)methoxy) benzene (27c). White solid (92%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 2.24 (s, 6H), 5.08 (s, 4H), 5.49 (4H), 6.52–6.55 (m, 3H), 7.09–7.13 (m, 3H), 7.16–7.20 (m, 4H), 7.23–7.27 (m, 2H), 7.40 (s, 2H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 18.9, 52.3, 62.0, 102.1, 107.5, 126.6, 129.1, 129.4, 129.9, 130.9, 132.3, 136.8, 159.3.

1,3-Bis((1-(3-methoxybenzyl)-1H-1,2,3-triazol-4-yl)methoxy) benzene (28c). White solid (91%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 3.73 (s, 6H), 5.11 (s, 4H), 5.46 (s, 4H), 6.54 (m, 3H), 6.76 (s, 2H), 6.83 (t, $J = 7.5$ Hz, 1H), 7.25 (m, 2H), 7.58 (bs, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 54.1, 55.2, 61.8, 101.9, 107.5, 113.6, 114.1, 120.1, 129.9, 130.1, 135.8, 159.3, 159.9.

1,3-Bis((1-(4-bromobenzyl)-1H-1,2,3-triazol-4-yl)methoxy) benzene (29c). Yellow solid (96%); $^1\text{H NMR}$ (400 MHz, DMSO-d_6) δ 5.07 (s, 4H), 5.56 (s, 4H), 6.57 (d, $J = 2.3$ Hz, 1H), 6.59 (d, $J = 2.3$ Hz, 1H), 6.67 (t, $J = 2.5$ Hz, 1H), 7.14 (t, $J = 7.9$ Hz, 1H), 7.22–7.26 (m, 4H), 7.51–7.55 (m, 4H), 8.28 (s, 2H); $^{13}\text{C NMR}$ (100 MHz, DMSO-d_6) δ 52.5, 61.5, 102.1, 107.8, 121.9, 125.2, 130.6, 132.1, 135.8, 143.5, 159.7.

1,4-Bis((1-benzyl-1H-1,2,3-triazol-4-yl)methoxy) benzene (30c). Yellow solid (92%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 5.06 (s, 4H), 5.60 (s, 4H), 6.95 (s, 4H), 7.30 (m, 2H), 7.32–7.33 (m, 3H), 7.34 (d, $J = 1.8$ Hz, 1H), 7.35–7.36 (m, 2H), 7.37–7.40 (m, 2H), 8.25 (s, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 52.8, 61.6, 115.6, 124.5, 127.9, 128.1, 128.73, 135.9, 143.2, 152.3.

1,4-Bis((1-(2-methylbenzyl)-1H-1,2,3-triazol-4-yl)methoxy) benzene (31c). White solid (93%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 2.24 (s, 6H), 5.06 (s, 4H), 5.49 (s, 4H), 6.83 (s, 4H), 7.10–7.11 (m, 2H), 7.16–7.19 (m, 4H), 7.23–7.27 (m, 2H), 7.41 (bs, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 18.9, 52.4, 62.6, 115.8, 126.6, 129.1, 129.3, 130.9, 132.3, 136.8, 152.7.

1,4-Bis((1-(3-methoxybenzyl)-1H-1,2,3-triazol-4-yl)methoxy) benzene (32c). White solid (92%); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 3.74 (s, 6H), 5.09 (bs, 4H), 5.46 (s, 4H), 6.75 (s, 2H), 6.81–6.86 (m, 8H), 7.24 (m, 2H), 7.67 (bs, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 54.3, 55.2, 62.4, 113.6, 114.1, 115.7, 120.1, 130.0, 135.7, 152.6, 159.9.

1,4-Bis((1-(4-bromobenzyl)-1H-1,2,3-triazol-4-yl)methoxy) benzene (33c). Yellow solid (92%); $^1\text{H NMR}$ (400 MHz, DMSO-d_6) δ 5.07 (s, 4H), 5.60 (s, 4H), 6.95 (s, 4H), 7.26 (d, $J = 8.6$ Hz, 4H), 7.57 (d, $J = 8.6$ Hz, 4H), 8.26 (s, 2H); $^{13}\text{C NMR}$ (100 MHz, DMSO-d_6) δ 52.1, 61.5, 115.6, 121.4, 124.6, 130.2, 131.7, 135.4, 143.3, 152.3.

1-Phenyl-2-(4-phenyl-1H-1,2,3-triazol-1-yl)ethan-1-one (35c). Pale yellow solid (92%) yield; $^1\text{H NMR}$ (400 MHz, DMSO-d_6) δ 6.26 (s, 2H), 7.35 (m, 1H), 7.47 (t, $J = 7.5$ Hz, 2H), 7.63 (t, $J = 7.5$ Hz, 2H), 7.75 (m, 1H), 7.87 (d, $J = 7.6$ Hz, 2H), 8.09 (d, $J = 7.8$ Hz, 2H), 8.52 (s, 1H); $^{13}\text{C NMR}$ (100 MHz, DMSO-d_6) δ 56.0, 123.0, 125.2, 128.2, 127.9, 128.2, 128.9, 130.7, 134.3, 146.3, 192.2.

1-(4-Methoxyphenyl)-2-(4-phenyl-1H-1,2,3-triazol-1-yl)ethan-1-one (36c). Pale yellow solid (95%) yield; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 3.84 (s, 3H), 5.77 (s, 2H), 6.93 (d, $J = 6.9$ Hz, 2H), 7.28 (t, $J = 7.3$ Hz, 1H), 7.37 (t, $J = 7.8$ Hz, 2H), 7.90 (s, 1H), 7.92 (d, $J = 7.8$ Hz, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 55.2, 114.4, 125.8, 126.9, 128.8, 130.6, 164.6, 188.6.

Conflicts of interest

There are no conflicts to declare.



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