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Fluorescence-tunable copper nanoclusters and their application in hexavalent chromium sensing†

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Generally, metal nanoclusters are synthesized using only a single ligand. Thus, the properties and applications of these nanomaterials are limited by the nature of the ligand used. In this study, we have developed a new synthetic strategy to prepare bi-ligand copper nanoclusters (Cu NCs). These bi-ligand Cu NCs are synthesized from copper ions, thiosalicylic acid, and cysteamine by a simple one-pot method, and they exhibit high quantum yields (>18.9%) and good photostability. Most interestingly, the fluorescence intensities and surface properties of the Cu NCs can be tailored by changing the ratio of the two ligands. Consequently, the bi-ligand Cu NCs show great promise as fluorescent probes. Accordingly, the Cu NCs were applied to the inner-filter-effect-based detection of hexavalent chromium in water. A wide linear range of 0.1–1000 μM and a low detection limit (signal-to-noise ratio = 3) of 0.03 μM was obtained. The recoveries for the real sample analysis were between 98.3 and 105.0% and the relative standard deviations were below 4.54%, demonstrating the repeatability and practical utility of this assay.

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1. Introduction

Fluorescent metal nanoclusters (NCs), which consist of numerous of metal atoms, have drawn increasing research interest in recent years. Unlike bulk materials or larger metal nanomaterials that have continuous band structures, metal NCs have discontinuous band structures and exhibit molecule-like properties.^{1–3} These new fluorescent nanomaterials are known to be water soluble, non-toxic, biocompatible, and stable, as well as exhibiting high quantum yields and outstanding catalytic properties.^{4–7} Thiolate-protected Au NCs and Ag NCs, which are considered to be promising candidates for application in chemical sensing and bioimaging, have drawn particular attention.^{8–11} However, Au and Ag are scarce and expensive. Conversely, copper is an easily obtained and cheap metal, and is thus widely used in daily life. Cu NCs have been investigated for their application to a range of fields, including as sensors, drug carriers, catalysts, and bioimaging.^{12–14}

The relationship between the ligands employed to prepare the NCs and the light-emitting mechanism of the resultant material is an important issue in this research field. The interactions between the ligand and the metal core strongly affect the fluorescence properties of the metal NCs. Specifically, during ligand to metal charge transfer (LMCT), the positive charge on the metal and the electron-donating ability of the

surface ligands strongly affect the light-emitting properties of the metal NCs.¹⁵

Many different ligands have been used to synthesize Cu NCs in recent years. For example, proteins that contain amine or thiol groups such as bovine serum albumin, human serum albumin, and soy protein have been used to engender the resulting Cu NCs with water solubility, and the Cu NCs produced were also highly fluorescent.^{6,16–26} However, these proteins are expensive and unstable in acidic environments, restricting the application of this type of Cu NCs. Cu NCs protected by small thiol molecules such as glutathione, cysteine, or penicillamine are very easy to prepare.^{4,27–36} However, most of these types of Cu NCs have low quantum yields (QYs), limiting their application.

All the above approaches employ a single ligand species to direct and cap the nascent NCs. Thus, we have found that Cu NCs capped using two different ligands are rarely studied and the effects of two ligands on Cu NC surfaces and fluorescence properties are relatively unexplored.

Chromium (Cr(VI)) is widely used in industrial processes such as metal smelting, leather tanning, electroplating, and dyeing. All these processes result in the release of large amounts of Cr(VI) into the environment, potentially contaminating drinking water and/or entering the food chain. Cr(VI) contamination is highly associated with allergic dermatitis, carcinogenesis, and mutagenesis in animals and humans. In addition to traditional instrumental analysis methods, several strategies have been developed for quantitative determination of Cr(VI) using nanomaterials based on phenomena such as the inner filter effect (IFE),^{37–42} electrochemical detection,^{43,44} and absorbance detection.^{45–48} Of these, the IFE mechanism is the most

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widely exploited owing to its low detection limit and comparative freedom from matrix effects.⁴⁹

Herein, we propose a new synthetic strategy to synthesis bi-ligand Cu NCs and a discussion of the relationship between the two ligands and the properties of the Cu NCs. We found that the fluorescence intensities and surface properties of the bi-ligand Cu NCs can be easily adjusted by changing the ratio of the two ligands. Furthermore, the blue emission of the bi-ligand Cu NCs is strongly quenched in the presence of Cr(vi) through the IFE mechanism, and this property allows our Cu NCs to be applied as a nanosensor for detecting Cr(vi).

2. Experimental

2.1 Materials

Cu(NO₃)₂·5H₂O, cysteamine (CysA), and metal ions were received from Sigma-Aldrich (St. Louis, MO, USA). Thiosalicylic acid (TA) was received from Acros (Morris Plains, NJ, USA). Monobasic, dibasic, tribasic sodium salt of phosphate, phosphoric acid, and Tris were obtained from J.T. Baker (Phillipsburg, NJ, USA). Ultrapure water (18.25 MΩ cm⁻¹) from a Milli-Q system of Millipore (Billerica, MA, USA) was used to prepare all aqueous and organic-aqueous solutions. All chemicals and solvents were analytical grade and used without further purification.

2.2 Instrumentation

UV-vis absorption spectra were performed on a Lambda EZ210 UV-vis spectrophotometer (Perkin Elmer, USA). Fluorescent spectra were performed on an F-7000 Fluorescence Spectrophotometer (Hitachi, Japan). Both of the excitation and emission slits were maintained at 5.0 nm. The morphology and mean diameter of bi-ligand Cu NCs were performed on a transmission electron microscope (TEM, JEM-2100, Hitachi, Japan). X-ray photoelectron spectra were measured on an X-ray photoelectron spectroscopy (XPS, K-Alpha, Thermo, USA). Binding energy calibration was based on C 1s at 284.6 eV. The stabilities and selectivity of bi-ligand Cu NCs were analyzed using an Infinite 200 PRO microplate reader (Tecan, Switzerland). The QY of bi-ligand Cu NCs were measured according to an established procedure. Quinine sulfate in 0.1 M H₂SO₄ (literature QY = 0.54) was chosen as a standard. Absolute values were calculated using the standard reference sample that has a fixed and known QY value. In order to minimize reabsorption effects, absorbencies in the 10 mm FL cuvette were kept under 0.1 at the excitation wavelength (340 nm).

2.3 Preparation of bi-ligand Cu NCs

The bi-ligand Cu NCs were prepared by the reported protocol with a new strategy.⁵⁰ Aqueous 2 mL of 0.2 M Cu(NO₃)₂ was adding in to 8 mL of bi-ligand solutions prepared by 50 mM TA (in EtOH) and 50 mM CysA (see the detailed proportions in Table 1). The brown copper/thiol complexes were formed in the vial. A 10 mL of EtOH solutions were added in to the complexes solution to make the final concentration of EtOH was control at 40%. Then the solutions were carried out by ultrasound for

10 min. Finally, the mixture was allowed to 40 min at 75 °C. The Cu NCs were purified by centrifugation (3000g, 10 min) in order to remove the precipitate from the reaction solution.

The final solution was stored at -20 °C refrigerator until it was used. For simplicity, the concentrations of as-prepared bi-ligand Cu NCs were denoted as 1×. The as-prepared bi-ligand Cu NCs were denoted as Cu NC-0 to Cu NC-4 according to CysA amount were 0, 100, 200, 250 and 300 μmol, respectively.

2.4 Determination of Cr(vi)

The Cu NC-2 prepared by TA-to-CysA molar ratio of 1 : 1 were used for the detection of Cr(vi). A Cr(vi) stock solution (10 mM) and phosphate buffer solutions (100 mM, pH 5.0) were prepared. 1400 μL ultrapure water, phosphate buffer (200 μL, 100 mM, pH 5.0), 200 μL of 0.1× Cu NC-2 and 200 μL of the Cr(vi) solution were added in to a quartz cuvette, sequentially. The final concentrations of Cu NC-2 in the mixtures were 0.01×, while that for Cr(vi) were 0.1–1000 μM. The fluorescence spectra were recorded after the preparation. The excitation wavelength was setting at 355 nm, and the PMT voltage was 600 V.

2.5 Detecting Cr(vi) in mineral water

Mineral water samples received from local supermarket were centrifuged at 12 000 rcf for 30 min and filtrated through 0.22 μm membranes. 400 μL of ultrapure water, 400 μL of mineral water, phosphate buffer (200 μL, 100 mM, pH 5.0), 200 μL of 0.1× Cu NC and 1000 μL of the Cr(vi) solution were added in to a quartz cuvette, sequentially. The fluorescence spectra were recorded after the preparation. The excitation wavelength was setting at 355 nm, and the PMT voltage was 600 V.

3. Results and discussion

3.1 Synthesis and characterization of bi-ligand Cu NCs

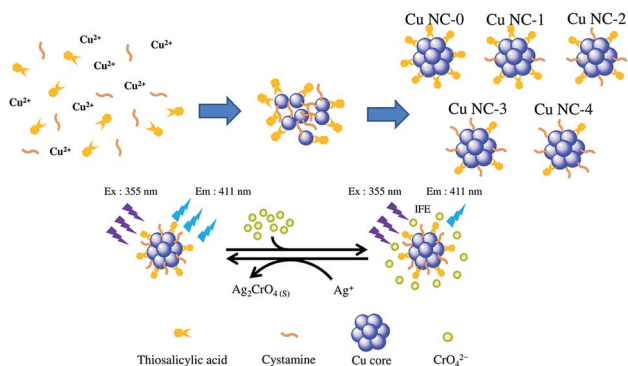
Thiol molecules are usually used as etching reagents for the formation of Cu NCs.^{51–55} However, this strategy requires long reaction times (>4 h) and a large amount of thiol (molar ratio: R-SH/Cu > 5). The synthesis of our bi-ligand Cu NCs is illustrated in Scheme 1. In this study, the bi-ligand Cu NCs were synthesized by heating copper ions in the presence of thiosalicylic acid (TA) and cysteamine (CysA) at different ratios. Our bi-ligand Cu NCs can be prepared using less reagent (molar ratio: R-SH/Cu = 1) and a shorter reaction time (<2 h) compared to those required

Table 1 The relationship between TA/CysA ratio and optical properties of bi-ligand Cu NCs^a

Cu NCs	TA amount (μmol)	CysA amount (μmol)	Abs ₃₄₀	Quantum yield (%)
Cu NC-0	400	0	0.089	18.9
Cu NC-1	300	100	0.071	25.4
Cu NC-2	200	200	0.057	30.5
Cu NC-3	150	250	0.042	31.5
Cu NC-4	100	300	0.034	34.0

^a The TA amount, CysA amount, absorbance at 340 nm and quantum yield of bi-ligand Cu NCs.





Scheme 1 Schematic illustration of the formation of bi-ligand Cu NC with different surface properties and IFE fluorescence determination of Cr(vi) using bi-ligand Cu NC. The bi-ligand Cu NCs were synthesized from copper ions, thiosalicylic acid (TA) and cysteamine (CysA). The fluorescent of bi-ligand Cu NCs can be quenched by Cr(vi) through an IFE then recovered by adding Ag⁺.

for the conventional method. This is due to the fact that both ligands act as reducing agents and protective agents during the reaction.

Upon fluorescence spectral measurement, the emission peaks of the different bi-ligand Cu NCs samples are observed at similar wavelengths. Fig. 1A shows the fluorescence excitation and emission spectrum of Cu NC-2. The maximum emission peak is observed at 411 nm upon excitation at 330 nm, which is similar to that previously reported for TA-Cu NCs.⁵⁰ The shape and size of the bi-ligand Cu NCs were examined using transmission electron microscopy (TEM), revealing that the morphologies of these bi-ligand Cu NCs are similar. From the TEM images and size distribution profiles shown in Fig. 1B, it can be seen that Cu NC-3 has an average particulate size of *ca.* 3.71 ± 1.01 nm. A small fraction of the Cu NC-3 has aggregated to form larger aggregates through hydrogen bonding and π - π

stacking.⁵⁰ X-ray photoelectron spectroscopy analysis was conducted to determine the composition of the Cu NCs and the oxidation state of copper therein. It can be seen from Fig. 1C that signals from C, O, N, S, and Cu are detected. These results directly confirm the elemental composition of the bi-ligand Cu NCs. The two distinct peaks at 932.3 and 952.1 eV in the Cu 2p spectrum are attributed to the 2p_{3/2} and 2p_{1/2} electrons of Cu(0), respectively (Fig. 1D). There is a small characteristic satellite peak around 942.0 eV for Cu 2p_{3/2}, which indicates the presence of some unreacted copper ions adsorbed on the surface of the bi-ligand Cu NCs.

Stability is a very important criterion for a nanosensor. Consequently, we investigated the salt tolerance and photo-stability of Cu NC-2 by measuring its fluorescence intensity. As shown in Fig. S1†, the fluorescence intensity changes little during 20 days storage at −20 °C. Furthermore, Fig. S2A† shows that the fluorescence intensity of Cu NC-2 remains almost the same for NaCl concentrations up to 1 M. In Fig. S2B†, it can be seen that the fluorescence intensity does not change after 1 h of irradiation. These results indicate that Cu NC-2 has very good salt tolerance and light stability.

3.2 Surface and fluorescence properties of bi-ligand Cu NCs

The fluorescent of Cu NCs capped by TA can be attributed to inter-band and/or intra-band electronic transitions through LMCT. The formulas and properties of the five bi-ligand Cu NCs samples synthesized are shown in Table 1. The absorbance of these bi-ligand Cu NCs increases gradually with increasing TA content, and their QY values increase with increasing CysA content (18.9–34.0%).

To confirm the relationship between the spectral properties and synthetic formula, two plots have been drawn. As shown in Fig. 2A, the absorbance at 340 nm is linearly related to TA content, and Fig. 2B shows that QY is linearly related to CysA content. A higher absorbance means more energy is absorbed by the bi-ligand Cu NCs through LMCT (S-Cu), and a higher QY

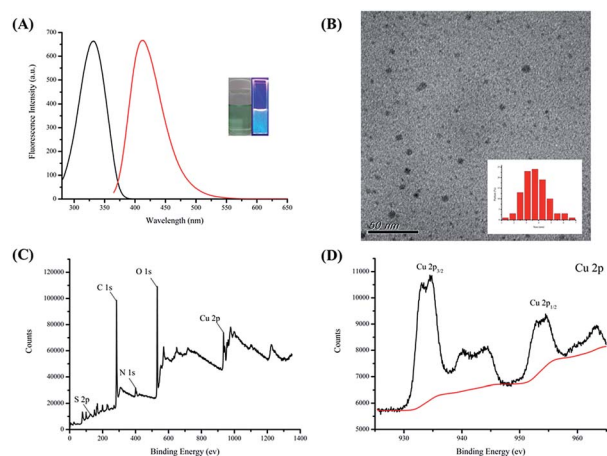


Fig. 1 Characterization of Cu NCs. (A) Excitation (black line) and emission spectra (red line) of Cu NC-2. The inset photograph is Cu NC-2 shown under daylight (left) and UV illumination (right). The excitation and emission wavelengths were 330 nm and 411 nm, respectively. (B) The TEM images of the Cu NC-3. Inset (B): particle size distribution of Cu NC-3. XPS survey (C) and Cu 2p region (D) spectrum of the Cu NC-2.

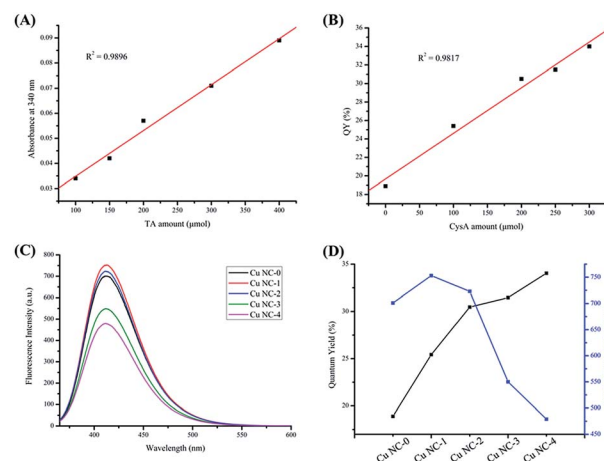


Fig. 2 Optical properties of bi-ligand Cu NCs. (A) Linear plot of the absorbance against TA amount in bi-ligand Cu NC. (B) Linear plot of the quantum yield against CysA amount in bi-ligand Cu NC. (C) Emission spectra of bi-ligand Cu NC. (D) The fluorescence intensity and quantum yield of bi-ligand Cu NC.



means more energy is released through the fluorescence process. Therefore, we can speculate that the TA absorbs excitation light, while CysA promotes electron transition between the ligand and metal. Specifically, the aromatic ring on TA absorbs energy from the excitation light and the CysA helps the excited electrons transfer to the copper through the inductive effect of the terminal amine, thereby increasing the QY. These results also support the conclusions drawn in a previous study.¹⁵ The emission spectra of the bi-ligand Cu NCs are shown in Fig. 2C. Hence, the ratio of these two ligands can be used to effectively regulate the fluorescence intensity of bi-ligand Cu NCs but also tune their surface properties. To develop a good fluorescence sensor, QY was plotted against the fluorescence intensity. As shown in Fig. 2D, the addition of CysA increases the QY, but reduces fluorescence intensity. Since Cu NC-2 exhibits both high QY and fluorescence intensity, Cu NC-2 was selected as a fluorescence sensor for all subsequent experiments.

The two ligands TA and CysA have amine and carboxyl groups, respectively. In a previous study, Au NPs prepared with a carboxyl protective agent were applied to the detection of melamine through hydrogen-bonding recognition interactions,³⁶ which resulted in the aggregation of the Au NPs.

To prove that there are different amounts of amine groups on the surfaces of the different bi-ligand Cu NCs, various volumes of bi-ligand Cu NCs were mixed with 2 mL Au NPs (1.97 mg mL^{-1}). The results are shown in Fig. 3. The Au NPs begin aggregating when 17 μL Cu NC-4 is added. Furthermore, a significant color change is observed for the Au NPs when 25 μL Cu NC-4 is added. Color changes are also observed when 30 μL of Cu NC-3 is added, 40 μL of Cu NC-2 is added, and 80 μL of Cu NC-1 is added. However, no color change is observed upon the addition of Cu NC-0, even when 80 μL is added.

Cu NC-4 has the highest CysA content, and thus less of it is required to cause Au NP aggregation. Cu NC-0, which is synthesized using only TA, does not cause Au NP aggregation. The strong interaction between the Au NPs and the amine group of CysA on the surface of the bi-ligand Cu NCs plays an important role in Au NP aggregation. This is strong evidence to indicate that the bi-ligand Cu NCs samples have different amounts of amine groups on their surfaces. Thus, the surface

properties can be easily tuned by controlling the ratio of TA and CysA. The fluorescent of Cu NCs capped by TA can be attributed to inter-band and/or intra-band electronic transitions through LMCT. The formulas and properties of the five bi-ligand Cu NCs samples synthesized are shown in Table 1. The absorbance of these bi-ligand Cu NCs increases gradually with increasing TA content, and their QY values increase with increasing CysA content (18.9–34.0%).

3.3 IFE determination of Cr(vi) using bi-ligand Cu NCs

The excitation spectrum of Cu NC-2 and the absorption spectrum of Cr(vi) are shown in Fig. S3†. There is a large overlap between the two spectra, which indicates that the fluorescence of Cu NC-2 will be quenched owing to absorption by the Cr(vi) through the IFE mechanism. To confirm the IFE mechanism, silver ions were added into this system, as illustrated in Fig. 4. Upon the addition of 1 mM Cr(vi), the fluorescence intensity of Cu NC-2 is initially quenched. However, the fluorescence is recovered by adding 0.2 M silver ions to the solution. The fluorescence intensity of the solution increases after centrifugation to remove the precipitate. From these results, we could conclude that the fluorescence intensity of Cu NC-2 is quenched by Cr(vi) due to the IFE (red line) and recovered by silver ions owing to the IFE being turned off. It is thought that the fluorescence intensity does not recover completely because some of the Cu NC-2 is co-precipitated with the silver chromate. This specific fluorescence response of Cu NC-2 offers an attractive opportunity to develop a fluorescent sensor for Cr(vi) detection.

3.4 Optimization for Cr(vi) sensing

Several experiments were conducted with the aim of optimizing the sensor efficacy. Fig. S4A† shows the fluorescence intensity changes for Cu NC-2 upon adding 1 mM Cr(vi) at different pH. The maximum fluorescence changes at pH 5, so pH 5 was chosen for the assay conditions. Because the IFE is turned on by absorbing incident light, the excitation wavelength is very important for analytical performance. As shown in Fig. S4B†,

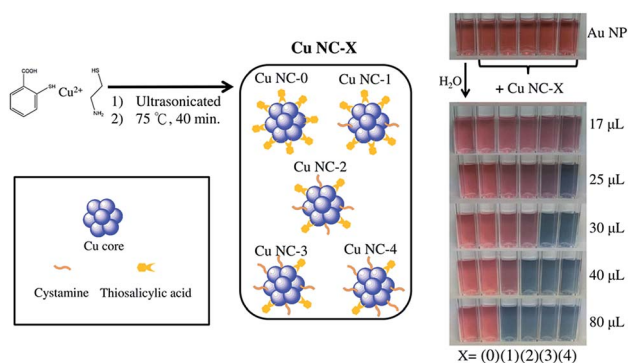


Fig. 3 Schematic illustration of the surface-tunable bi-ligand Cu NCs. The bi-ligand Cu NCs synthesized by different ratio of TA and CysA show different affinity to Au NPs.

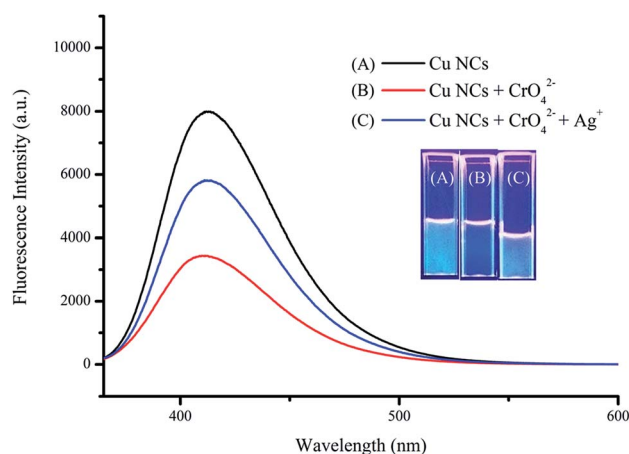


Fig. 4 Fluorescence of bi-ligand Cu NC-2 switches off and on. Fluorescence spectra of Cu NC-2 (A), Cu NC-2 containing 0.1 mM of Cr(vi) (B) and Cu NC-2 containing 0.1 mM of Cr(vi) and 20 mM of Ag⁺ after centrifugation (C).



the fluorescence intensity differences were investigated under different excitation wavelengths from 330 to 370 nm. To avoid the fluorescence intensity being too low to observe (thus negatively affecting the detection limit) an excitation wavelength of 355 nm was chosen for subsequent experiments.

Reaction time is also an important factor for the analytical assay. The time scan fluorescence spectra of Cu NC-2 with and without the addition of 1 mM Cr(VI) are shown in Fig. S5†. The fluorescence intensities of these two samples are very stable for 30 min. This phenomenon indicates that our assay requires only a very short reaction time and the assay results are stable for at least 30 min. Thus, the results confirm that our assay provides quick-response and stable sensing.

3.5 Detection performance of Cu NC-2 in Cr(VI) sensing

The performance of Cu NC-2 for detecting Cr(VI) based on fluorescence quenching through the IFE mechanism was carefully evaluated. Fig. 5A shows that the fluorescence intensity at 411 nm gradually decreases with increasing Cr(VI) concentration from 0.1 to 1000 μM . These results indicate that this fluorescence assay is very sensitive to changes in the Cr(VI) concentration. The experimental results presented in Fig. 5B demonstrate good linearity between the fluorescence intensity and the Cr(VI) concentration in the range 0.1–1000 μM with

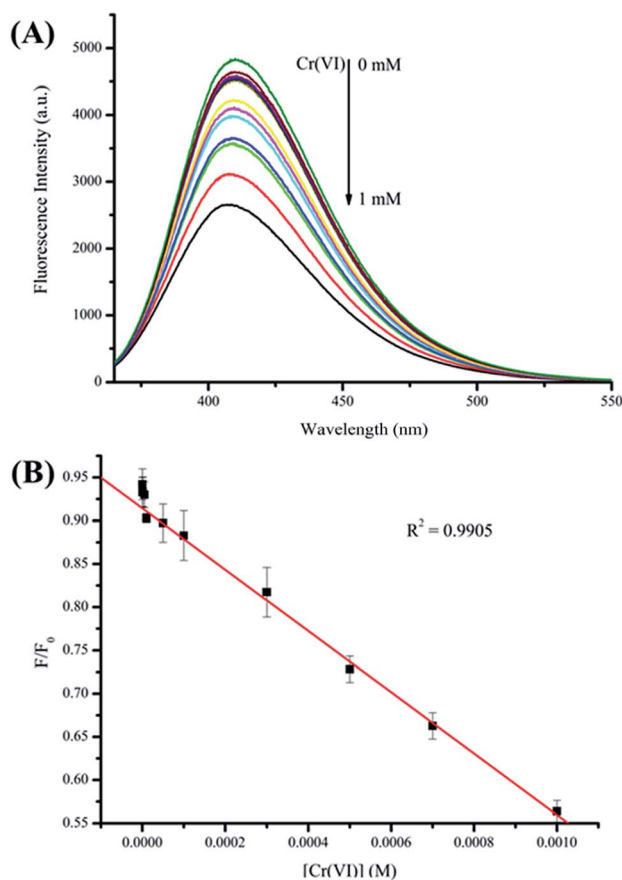


Fig. 5 Linear range of IFE detection of Cr(VI). (A) Detection of Cr(VI) in standard solutions using Cu NC-2. (B) Linear plot of relative fluorescence intensities of Cu NC-2 against Cr(VI) concentration.

a low limit of detection of 0.03 μM at a signal-to-noise ratio of 3. The detection limit is substantially lower than the guideline limit stipulated by the World Health Organization (0.96 μM).⁵⁷

Selectivity is another important criterion for sensors. Accordingly, interference and selectivity tests were performed. As shown in Fig. 6, it can be seen that most common metal ions that exist in the environment do not significantly interfere with our method. Only Cr(VI) causes an obvious change in fluorescence, and other metal ions like potassium, cobalt, calcium, magnesium, barium, mercury, copper, lithium, zinc, and nickel ions do not cause this change (<7.5%). Thus, this method has excellent selectivity for the detection of Cr(VI).

To demonstrate that our assay can be used to detect Cr(VI) in real samples. The developed sensor, Cu NC-2, was applied to the determination of Cr(VI) in mineral water samples obtained from a local supermarket. The water samples were analyzed after spiking with Cr(VI) at 10, 14, and 20 μM . The recoveries obtained are from 98.3 to 105.0% and the relative standard deviations are less than 4.54% ($n = 3$). These data are shown in Table S1†. Compared with other sensors reported in the literature (Table S2†), our sensor is more sensitive than most

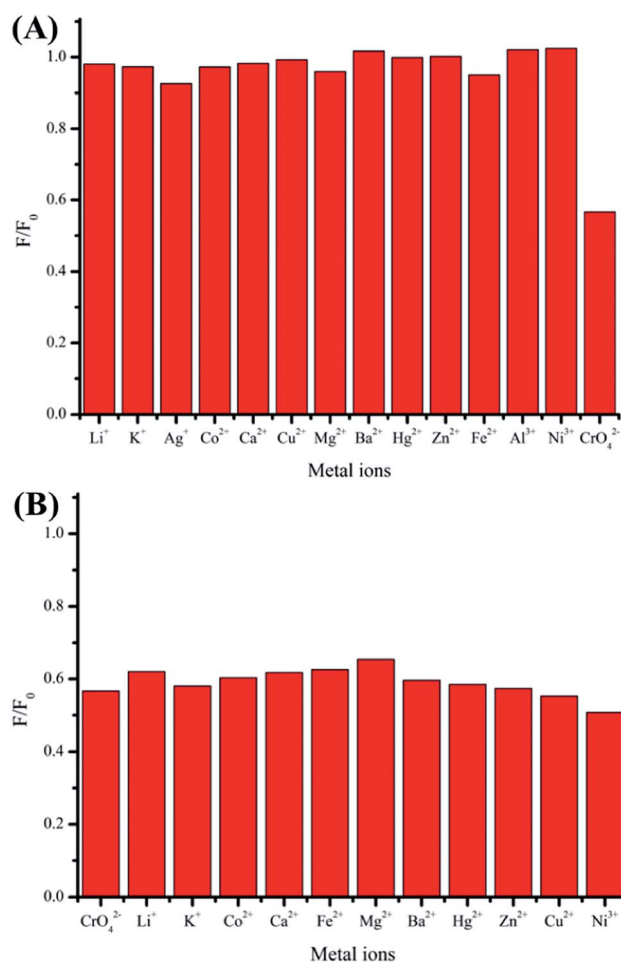


Fig. 6 Selectivity and interference of IFE detection of Cr(VI). Relative emission intensities (F/F_0) of as-prepared Cu NC-2 after the addition of 10^{-4} M of different metal ions (A) in the presence of 10^{-4} M of Cr(VI) (B). Mixtures were prepared in phosphate buffer (0.1 M, pH 5.0).



nanosensors reported. Thus, as well as presenting a high QY and simple preparation, our assay has a short response time and can be used for real water sample sensing.

4. Conclusions

In summary, a novel bi-ligand Cu NC nanosensor was prepared and its fluorescence properties were investigated, leading to the development of a simple, stable, selective, and sensitive method for detecting Cr(VI) in water samples. Using this bi-ligand strategy, the synthesized Cu NCs not only exhibited high QYs but also tunable surface and fluorescence properties. After optimization, the QY of the bi-ligand Cu NCs reached 34.0%, and they were used for IFE-based Cr(VI) detection. We anticipate that more customized Cu NCs may be developed by this multiple-ligand synthetic strategy, and our research provides a simple and effective example.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 S. Choi, R. M. Dickson and J. Yu, *Chem. Soc. Rev.*, 2012, **41**, 1867–1891.
- 2 R. W. Murray, *Chem. Rev.*, 2008, **108**, 2688–2720.
- 3 X. Yuan, Z. T. Luo, Y. Yu, Q. F. Yao and J. P. Xie, *Chem.-Asian J.*, 2013, **8**, 858–871.
- 4 T. Zhou, Q. Yao, T. Zhao and X. Chen, *Talanta*, 2015, **141**, 80–85.
- 5 X. Yang, Y. Feng, S. Zhu, Y. Luo, Y. Zhuo and Y. Dou, *Anal. Chim. Acta*, 2014, **847**, 49–54.
- 6 P. Ni, Y. Sun, H. Dai, J. Hu, S. Jiang, Y. Wang and Z. Li, *Biosens. Bioelectron.*, 2015, **63**, 47–52.
- 7 Y. Zhong, Q. Wang, Y. He, Y. Ge and G. Song, *Sens. Actuators, B*, 2015, **209**, 147–153.
- 8 X. Le Guével, C. Spies, N. Daum, G. Jung and M. Schneider, *Nano Res.*, 2012, **5**, 379–387.
- 9 J. Wang, J. Ye, H. Jiang, S. Gao, W. Ge, Y. Chen, C. Liu, C. Amatore and X. Wang, *RSC Adv.*, 2014, **4**, 37790–37795.
- 10 J. Wang, G. Zhang, Q. Li, H. Jiang, C. Liu, C. Amatore and X. Wang, *Sci. Rep.*, 2013, **3**, 1157.
- 11 Y. Wang, J. T. Chen and X. P. Yan, *Anal. Chem.*, 2013, **85**, 2529–2535.
- 12 M. De, P. S. Ghosh and V. M. Rotello, *Adv. Mater.*, 2008, **20**, 4225–4241.
- 13 J. R. Bhamore, B. Deshmukh, V. Haran, S. Jha, R. K. Singhal, N. Lenka, K. Kumar and Z. V. K. Murthy, *New J. Chem.*, 2018, **42**, 1510–1520.
- 14 J. R. Bhamore, S. Jha, A. I. Mungara, R. K. Singhal, D. Sonkeshariya and S. K. Kailasa, *Biosens. Bioelectron.*, 2016, **80**, 243–248.
- 15 Z. Wu and R. Jin, *Nano Lett.*, 2010, **10**, 2568–2573.
- 16 H. Chen, L. Lin, H. Li, J. Li and J. M. Lin, *ACS Nano*, 2015, **9**, 2173–2183.
- 17 D. Q. Feng, W. Zhu, G. Liu and W. Wang, *RSC Adv.*, 2016, **6**, 96729–96734.
- 18 C. Zhang, L. Zeng, X. Zhu, C. Yu, X. Zuo, X. Xiao and J. Nan, *Anal. Methods*, 2013, **5**, 2203–2208.
- 19 L. Jin, Z. Zhang, A. Tang, C. Li and Y. Shen, *Biosens. Bioelectron.*, 2016, **79**, 108–113.
- 20 H. Li, C. Wang, P. Gai, T. Hou, L. Ge and F. Li, *RSC Adv.*, 2016, **6**, 76679–76683.
- 21 Y. Qiao, T. Xu, Y. Zhang, C. Zhang, L. Shi, G. Zhang, S. Shuang and C. Dong, *Sens. Actuators, B*, 2015, **220**, 1064–1069.
- 22 A. Singh, T. Rai and D. Panda, *RSC Adv.*, 2016, **6**, 55539–55545.
- 23 X. Q. Liao, R. Y. Li, Z. J. Li, X. L. Sun, Z. P. Wang and J. K. Liu, *New J. Chem.*, 2015, **39**, 5240–5248.
- 24 S. Xu, M. Deng, Y. Sui, Y. Zhang and F. Chen, *RSC Adv.*, 2014, **4**, 44644–44649.
- 25 J. K. Yang, H. Kang, H. Lee, A. Jo, S. Jeong, S. J. Jeon, H. I. Kim, H. Y. Lee, D. H. Jeong, J. H. Kim and Y. S. Lee, *ACS Appl. Mater. Interfaces*, 2014, **6**, 12541–12549.
- 26 G. Zhang, T. Xu, H. Du, Y. Qiao, X. Guo, L. Shi, Y. Zhang, S. Shuang, C. Dong and H. Ma, *J. Mater. Chem. C*, 2016, **4**, 3540–3545.
- 27 P. C. Chen, Y. C. Li, J. Y. Ma, J. Y. Huang, C. F. Chen and H. T. Chang, *Sci. Rep.*, 2016, **6**, 24882.
- 28 H. Huang, H. Li, J. J. Feng, H. Feng, A. J. Wang and Z. Qian, *Sens. Actuators, B*, 2017, **241**, 292–297.
- 29 Y. Huang, H. Feng, W. Liu, Y. Zhou, C. Tang, H. Ao, M. Zhao, G. Chen, J. Chen and Z. Qian, *Anal. Chem.*, 2016, **88**, 11575–11583.
- 30 Y. Huang, W. Liu, H. Feng, Y. Ye, C. Tang, H. Ao, M. Zhao, G. Chen, J. Chen and Z. Qian, *Anal. Chem.*, 2016, **88**, 7429–7434.
- 31 X. Jia, X. Yang, J. Li, D. Li and E. Wang, *Chem. Commun.*, 2014, **50**, 237–239.
- 32 J. Y. Ma, P. C. Chen and H. T. Chang, *Nanotechnology*, 2014, **25**, 195502.
- 33 L. Kong, X. Chu, W. Liu, Y. Yao, P. Zhu and X. Ling, *New J. Chem.*, 2016, **40**, 4744–4750.
- 34 D. Li, Z. Chen, Z. Wan, T. Yang, H. Wang and X. Mei, *RSC Adv.*, 2016, **6**, 34090–34095.
- 35 S. M. Lin, S. Geng, N. Li, N. B. Li and H. Q. Luo, *Talanta*, 2016, **151**, 106–113.
- 36 R. Y. Li, H. Y. Wang, X. Y. Zhou, X. Q. Liao, X. L. Sun and Z. J. Li, *New J. Chem.*, 2016, **40**, 732–739.
- 37 F. Cai, X. Liu, S. Liu, H. Liu and Y. Huang, *RSC Adv.*, 2014, **4**, 52016–52022.
- 38 M. Cui, G. Song, C. Wang and Q. Song, *Microchim. Acta*, 2015, **182**, 1371–1377.
- 39 H. L. Tao, M. Z. Xu, Q. J. Zhang, X. H. Zhang, S. H. Li, F. X. Zhong, C. Sun and X. F. Liao, *Anal. Methods*, 2013, **5**, 5961–5968.



- 40 M. Rong, L. Lin, X. Song, Y. Wang, Y. Zhong, J. Yan, Y. Feng, X. Zeng and X. Chen, *Biosens. Bioelectron.*, 2015, **68**, 210–217.
- 41 Z. Zhang, C. Sha, A. Liu, Z. Zhang and D. Xu, *J. Fluoresc.*, 2015, **25**, 335–340.
- 42 A. Zheng, J. Chen, G. Wu, G. Wu, Y. G. Zhang and H. Wei, *Spectrochim. Acta, Part A*, 2009, **74**, 265–270.
- 43 W. Jin, G. Wu and A. Chen, *Analyst*, 2014, **139**, 235–241.
- 44 R. Ouyang, S. A. Bragg, J. Q. Chambers and Z. L. Xue, *Anal. Chim. Acta*, 2012, **722**, 1–7.
- 45 W. Chen, F. Cao, W. Zheng, Y. Tian, Y. Xianyu, P. Xu, W. Zhang, Z. Wang, K. Deng and X. Jiang, *Nanoscale*, 2015, **7**, 2042–2049.
- 46 J. F. Guo, D. Q. Huo, M. Yang, C. J. Hou, J. J. Li, H. B. Fa, H. B. Luo and P. Yang, *Talanta*, 2016, **161**, 819–825.
- 47 F. M. Li, J. M. Liu, X. X. Wang, L. P. Lin, W. L. Cai, X. Lin, Y. N. Zeng, Z. M. Li and S. Q. Lin, *Sens. Actuators, B*, 2011, **155**, 817–822.
- 48 X. Wu, Y. Xu, Y. Dong, X. Jiang and N. Zhu, *Anal. Methods*, 2013, **5**, 560–565.
- 49 J. Li, X. Li, X. Shi, X. He, W. Wei, N. Ma and H. Chen, *ACS Appl. Mater. Interfaces*, 2013, **5**, 9798–9802.
- 50 Y. J. Lin, P. C. Chen, Z. Yuan, J. Y. Ma and H. T. Chang, *Chem. Commun.*, 2015, **51**, 11983–11986.
- 51 Y. S. Borghei, M. Hosseini, M. R. Ganjali and S. Hosseinkhani, *Sens. Actuators, B*, 2017, **248**, 133–139.
- 52 R. Ghosh, U. Goswami, S. S. Ghosh and A. Paul, *ACS Appl. Mater. Interfaces*, 2015, **7**, 209–222.
- 53 R. Gui, J. Sun, X. Cao, Y. Wang and H. Jin, *RSC Adv.*, 2014, **4**, 29083–29088.
- 54 C. Wang, H. Cheng, Y. Huang, Z. Xu, H. Lin and C. Zhang, *Analyst*, 2015, **140**, 5634–5639.
- 55 S. Yang, X. Sun and Y. Chen, *Mater. Lett.*, 2017, **194**, 5–8.
- 56 K. Ai, Y. Liu and L. Lu, *J. Am. Chem. Soc.*, 2009, **131**, 9496–9497.
- 57 S. H. Frisbie, E. J. Mitchell, H. Dustin, D. M. Maynard and B. Sarkar, *Environ. Health Perspect.*, 2012, **120**, 775–778.

