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Celecoxib attenuates hepatocellular proliferative capacity during hepatocarcinogenesis by modulating a PTEN/NF- κ B/PRL-3 pathway[†]

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Although the efficacy of celecoxib on various cancer cell behaviors, including aberrant proliferation, in cultured hepatocellular carcinoma (HCC) cells has been demonstrated, whether celecoxib regulates cell proliferation by targeting PRL-3-associated signaling transduction during hepatocarcinogenesis *in vivo* has been incompletely studied. Here, we investigate the anti-proliferative efficacy of celecoxib in a rapid HCC mouse model established by hydrodynamic transfection of activated AKT and c-Met proto-oncogenes. The results show that celecoxib is effective at delaying the malignant transformation of hepatocytes by reducing the protein expression of Ki67, Cyclin D1 and c-Myc in the AKT/c-Met HCC-bearing mice. Mechanistically, celecoxib increases the protein expression of PTEN and suppresses the protein expression of NF- κ B and PRL-3 in the liver of the HCC mice. Using PTEN-silenced and LPS-stimulated approaches *in vitro*, a mechanism by which celecoxib regulates a PTEN/NF- κ B/PRL-3 pathway in HCC cells was illuminated. Altogether, our study demonstrates that celecoxib attenuates the hepatocellular proliferative capacity during hepatocarcinogenesis, which is probably attributable to its regulation of the PTEN/NF- κ B/PRL-3 pathway.

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Introduction

Hepatocellular carcinoma (HCC) is the 6th most prevalent cancer and the 3rd most frequent malignancy causing cancer-related death worldwide.¹ Although therapeutic strategies against HCC, including surgical resection and systemic chemotherapy, have been used to improve the survival of HCC patients, the survival rate of these patients is still less than 14%.² Excessive proliferation of hepatocytes is the primary indication of both hepatocarcinogenesis and poor prognosis in HCC patients.³ Thus, the discovery of therapeutic agents capable of inhibiting crucial oncogenic pathways fueling hepatocellular proliferation and malignant transformation is urgently needed for the management of HCC.

Phosphatase and tensin homologue (PTEN) is a tumor suppressor that controls the intracellular levels of phosphatidylinositol-3,4,5-triphosphate and has an essential role in

manipulating the phosphoinositide 3-kinase (PI3K) network to control cellular homeostasis.⁴ Mutations and loss of PTEN may occur during tumor initiation and progression that interferes with its enzymatic activity, subsequently leading to the activation of its downstream signaling pathways, and ultimately promoting aberrant proliferation, invasion and metastasis in human cancers.⁵ Hepatocytes lacking PTEN fails to harmonize the PI3K-linked program, which triggers a variety of cellular phenotypes that favor hepatocarcinogenesis.⁶ Furthermore, upon activation by systemic suppression of PTEN, AKT moves to the cytoplasm and nucleus, where it phosphorylates multiple downstream oncogenic targets,^{7,8} and further stimulates a pivotal transcriptional factor nuclear factor κ B (NF- κ B) DNA-binding activity for enhancing the transcription of pro-oncogenes, such as Cyclin D1 (ref. 9) and c-Myc.¹⁰

Mounting evidence shows that overexpression of phosphatases of regenerating liver (PRLs) accelerates tumorigenic and metastatic processes.¹¹ Moreover, PRLs plays an important role in controlling cell cycles and accelerating tumor cell proliferation.¹² Notably, PRL-3, one member of protein tyrosine phosphatase superfamily serving as a metastasis-associated phosphatase,¹³ is a potential molecular target for inhibition of proliferation through the AKT/mTORC1 signaling in glioblastoma.¹⁴ Furthermore, it has been reported that NF- κ B is capable of binding the PRL-3 promoter and enhances gene transcription and expression of PRL-3 in breast cancer.¹⁵ However, whether chemicals could suppress hepatocellular proliferation during hepatocarcinogenesis by modulating the NF- κ B/PRL-3 cascade remains unclear.

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Recent studies have shown that selective cyclooxygenase type 2 (COX-2) inhibitors possess anti-tumor activity in various tumor types, including HCC.¹⁶ Celecoxib (Fig. 1), a kind of nonsteroidal anti-inflammatory drugs (NSAIDs), is effective at decelerating cell growth in multiple preclinical cancer models through suppression of proliferation.¹⁷ Although previous literatures showed the inhibitory efficacy of celecoxib in cultured HCC cells, the molecular mechanism by which celecoxib delays hepatocellular malignant transformation and tumorigenesis has not been well demonstrated. In this study, the anti-proliferative activity of celecoxib was investigated in an AKT/c-Met-driven HCC mouse model *via* hydrodynamic transfection that led to downregulation of PTEN expression and elevation of NF- κ B and PRL-3 expression. Our data indicate that celecoxib impairs excessive proliferation during carcinogenesis of hepatocytes, potentially resulting from its regulation of a PTEN/NF- κ B/PRL-3 pathway.

Methods and materials

Chemicals and reagents

Celecoxib powder and capsules were obtained from Sigma-Aldrich (St. Louis, MO, USA) and Pfizer Pharmaceuticals LLC (Celebrex, New York, USA), respectively. Lipopolysaccharide (LPS) was purchased from Invitrogen (San Diego, CA, USA). A hydrosoluble tetrazolium salt WST-8 (2-(2-methoxy-4-nitrophenyl)-3-(4-nitrophenyl)-5-(2,4-disulfophenyl)-2H-tetrazolium, monosodium salt) was obtained from Dojindo Laboratories (Kumamoto, Japan) for the CCK-8 assay. The E.Z.N.A.® Plasmid Maxi Kit (Omega Bio-tek, Inc. Doraville, GA, USA) was used for plasmid extractions and purifications before hydrodynamic and transient transfection into mice and *in vitro* HCC cells, respectively.

Hydrodynamic injection and animal administration

Female wild-type FVB/N mice were purchased from Charles River (Beijing, China) and housed under controlled temperature (23 °C) and lighting (12 h light/12 h dark cycle) with free access

to water and standard mouse chow. FVB/N mice were divided into four groups ($n = 6$): (1) wild-type control mice (WT); (2) AKT/c-Met HCC mice (AKT/c-Met, co-expression of AKT/c-Met *via* hydrodynamic transfection); (3) AKT/c-Met HCC mice treated with low dose of celecoxib (AKT/c-Met + CELE-L, celecoxib 125 mg per kg per day); AKT/c-Met HCC mice treated with high dose of celecoxib (AKT/c-Met + CELE-H, celecoxib 250 mg per kg per day). Hydrodynamic transfection was performed as previously described.¹⁸ A volume of 2 mL normal saline solution containing 20 μ g pT3-EF1 α -HA-myr-AKT, 20 μ g pT3-EF1 α -V5-c-Met and 1.6 μ g pCMV-sleeping beauty transposase (SB) was injected into the caudal vein of the FVB/N mice within 7 seconds. Since the AKT/c-Met-injected mice developed a lethal burden of liver tumor within 6 to 8 weeks post-injection,¹⁸ mice were sacrificed by decapitation 6 weeks after injection in the present study. This study was performed in accordance with the Animal Management Rules of the Ministry of Health of the People's Republic of China and were approved by the Animal Care and Use Committee of the Hubei University of Chinese Medicine (Wuhan, China).

Cell culture, transient transfection and treatment

The human HCC cell lines (Huh-7 and SMMC-7721) were purchased from the American Type Culture Collection and cultured in DMEM supplemented with 10% fetal bovine serum, penicillin (1 mM) and streptomycin (1 mM) at 37 °C in a humidified 5% CO₂ atmosphere. The transient transfection of HCC cells with pT3-EF1 α -HA-myr-AKT/c-Met or sgPTEN, a sgRNA-plasmid vector targeting human PTEN (Santa Cruz Biotechnology), was performed according to the manufacturer's protocol using Lipofectamine™ 2000 (Invitrogen, Carlsbad, CA, USA). The transfected cells were then incubated with celecoxib (0–50 μ M) for 24 h.

Quantitative real-time polymerase chain reaction (qPCR)

The total cellular RNA was extracted using the TRIzol reagent (Invitrogen, USA). Oligo (dT) 18 primer was used to bind the poly-A tails of mRNA for generating complementary DNA (cDNA). The qPCR assay was performed on a Mini-Opticon (Bio-Rad, USA) using Fast Start Universal SYBR Green Master Mix (Roche, USA). The housekeeping gene β -actin was employed as an internal control. Primers for qPCR are listed in the ESI Table 1.†

Western blotting and immunohistochemistry

The protein preparation and immunoblotting procedures for hepatic tissues and cultured cells were performed as previously described.¹⁹ The specific primary antibodies used in the immunoblotting assay are listed in ESI Table 2.†

The nuclear antigen Ki67, NF- κ B and E-cadherin were detected by immunohistochemistry in the liver tissues of mice. After deparaffinization of the liver tissue sections, antibody incubation and visualization of immunoreactivity were conducted as previously described.²⁰ Specific primary antibodies used for immunohistochemistry are listed in ESI Table 2.†

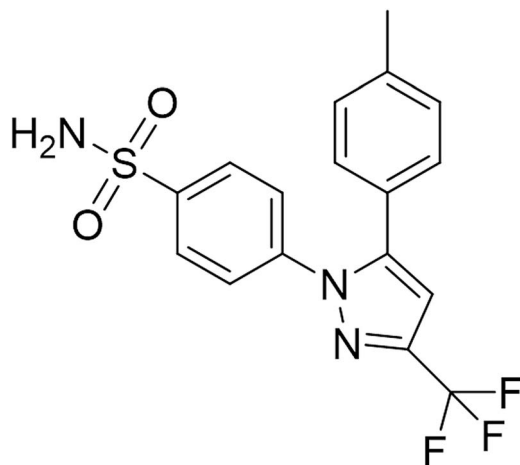


Fig. 1 The chemical structure of celecoxib.



Statistical analysis

Data analysis was performed with Prism 6 (GraphPad, USA). Data are shown as the mean \pm S.D. from at least three independent experiments. Statistical significance was measured using a two-tailed unpaired *t*-test (between two groups) or ANOVA (between three or more groups) and a statistical cutoff of $P < 0.05$.

Results

Celecoxib suppresses aberrant hepatocellular proliferation in AKT/c-Met-induced HCC-bearing mice

Immunohistochemical assessment of the proportion of cells positive for Ki67 is the most common approach for determining the proliferative index and evaluating the prognosis of cancer patients with HCC and other neoplasms.²¹ As expected, we found that hepatocellular tumors consisting mainly of lipid-rich cells (ESI Fig. 1†) and highly proliferative Ki67-positive liver tumor cells (Fig. 2A) were eradicated upon celecoxib administration in the CELE-H group. Moreover, compared to the control group, celecoxib treatment downregulated both the protein and mRNA expression of two kinds of G1-phase regulatory proteins, c-Myc and Cyclin D1 in the CELE-H group (Fig. 2B–D). Thus, the results indicate that the inhibition of cell proliferation might be a potential mechanism by which celecoxib alleviates AKT/c-Met-triggered hepatocarcinogenesis.

Celecoxib elevates the protein expression of PTEN and inhibits the protein expression of NF- κ B and PRL-3 in HCC-bearing mice

Using Western blotting, immunohistochemistry and qPCR analyses as described in the Methods, we investigated the molecular mechanisms underlying the inhibitory efficacy of celecoxib on hepatocellular proliferation in mice. The results revealed that celecoxib treatment elevated the level of PTEN expression in liver tissues relative to that in the AKT/c-Met group (Fig. 3A and B). Moreover, celecoxib effectively reduced the protein expression of NF- κ B in mice of the CELE-H group (Fig. 3A–D). PRL-3 is commonly overexpressed to promote cell proliferation in tumor malignant transformation.²² Of note, our results indicated that the mRNA and protein expression of PRL-3 were suppressed upon celecoxib administration in mice of the CELE-L and CELE-H groups (Fig. 3A–C). Furthermore, celecoxib enhanced the immunohistochemical staining of E-cadherin, a known downstream negative effector of PRL-3, in mice of the CELE-L and CELE-H groups (Fig. 3D). Therefore, these data suggested that celecoxib obstructs hepatocellular proliferation in the AKT/c-Met-transfected mice, which is probably due to its regulation of PTEN, NF- κ B and PRL-3 expression.

Celecoxib upregulates the protein expression of PTEN and inhibits the protein expression of NF- κ B and PRL-3 in HCC cells

To confirm whether celecoxib is efficacious in HCC-bearing mice by regulating a PTEN/NF- κ B/PRL-3 pathway, we further evaluated the protein expression of the aforementioned targets

at different concentrations (10–50 μ M) of celecoxib in cultured HCC cells. The results revealed that celecoxib enhanced the expression of PTEN and repressed the expression of NF- κ B and PRL-3 in a concentration-dependent manner in SMMC-7721 (Fig. 4A and B) and Huh-7 (Fig. 4C and D) cells. Similarly, celecoxib prevented the enhancing effect of AKT/c-Met transfection on the protein expression of NF- κ B and PRL-3 in a dose-dependent manner (Fig. 4E and F). Furthermore, consistent with *in vivo* findings, the protein expression of Cyclin D1 and c-Myc was suppressed by celecoxib incubation in the AKT/c-Met overexpressing cells (Fig. 4E and F).

Celecoxib regulates a PTEN/NF- κ B/PRL-3 pathway in HCC cells

Next, genetic and LPS-induced approaches were used to expound the mechanism by which celecoxib alleviates the proliferative capacity of hepatocytes. To do so, SMMC-7721 cells were transiently transfected with the sgRNA-plasmid targeting PTEN were further incubated with celecoxib (10–50 μ M) for 24 hours. The data suggested that PTEN silence led to the elevated expression of NF- κ B and PRL-3, indicating that PTEN may act upstream of NF- κ B and PRL-3 (Fig. 5A and B). Notably, celecoxib treatment restored the protein expression of PTEN, and repressed the sgPTEN-mediated elevated protein expression of NF- κ B and PRL-3 (Fig. 5A and B). To determine whether celecoxib could regulate the NF- κ B/PRL-3 signaling transduction in hepatocytes, we assessed the effect of celecoxib on PRL-3 expression in the presence of LPS, well known to trigger innate immune responses and enhance the expression and activity of NF- κ B.²³ As shown in Fig. 5C and D, the protein expression of NF- κ B and PRL-3 was elevated by LPS stimulation in SMMC-7721 HCC cells. Further, the LPS-induced HCC cells in the presence of celecoxib (10–50 μ M) for 24 h showed a dose-dependent reduction of NF- κ B and PRL-3 expression at the protein level (Fig. 5C and D). Overall, our data indicate that celecoxib is capable of regulating the PTEN/NF- κ B/PRL-3 pathway in HCC cells.

Discussion

Although previous studies have reported that COX inhibitors suppress cell proliferation and tumor formation, whether COX-2 inhibitors can interfere with this process during hepatocarcinogenesis remains poorly defined. In the present study, we sought to determine the anti-proliferative efficacy of a selective COX-2 inhibitor, celecoxib, in a rapid HCC mouse model established by hydrodynamic injection. The present data suggest that celecoxib diminished aberrant hepatocellular proliferation during hepatocarcinogenesis *via* a PTEN/NF- κ B/PRL-3 pathway.

In this study, celecoxib could repress the expression of proliferation-associated regulators (Ki67, c-Myc and Cyclin D1) in AKT/c-Met-driven hepatocarcinogenesis, indicating that its efficacy delaying hepatic tumorigenesis is mainly due to the inhibition of proliferation. Mechanistically, the protein expression of PTEN was elevated by celecoxib treatment in the AKT/c-Met-transfected mice,



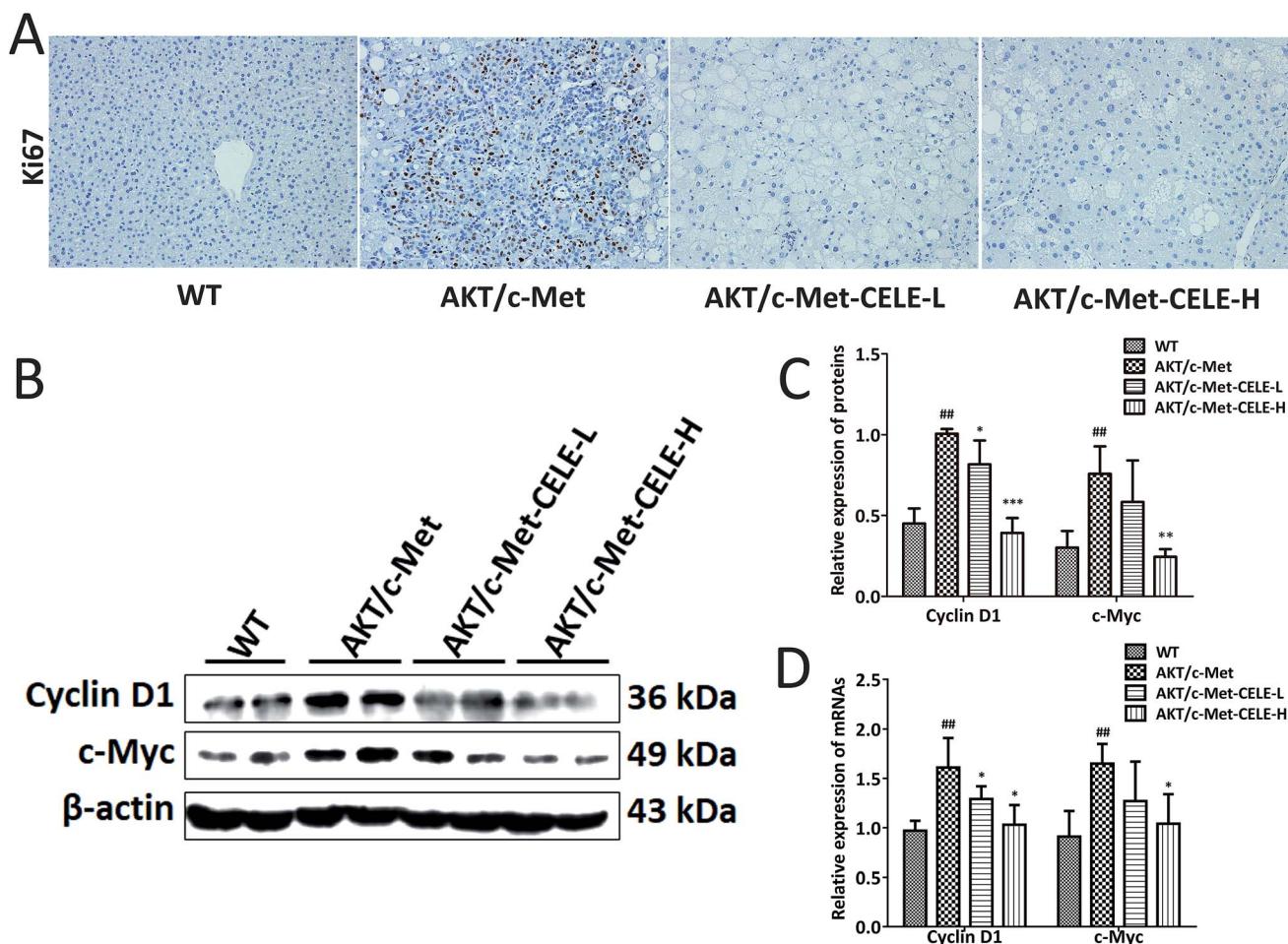


Fig. 2 Celecoxib inhibits AKT/c-Met-driven hepatocellular proliferation during hepatocarcinogenesis. (A) Observation of nuclear Ki67 staining in the liver tissues. AKT/c-Met-induced high expression of Ki67 was suppressed by celecoxib treatment. Ki67 positive cells are recognized by their yellow-brown nuclear staining. Original magnification: 200 \times . (B) Celecoxib suppressed the protein expression of Cyclin D1 and c-Myc in the livers of AKT/c-Met HCC-bearing mice. Liver samples from each group were subjected to Western blot assay, and representative bands are shown. (C and D) Histograms show the relative protein and mRNA expression levels of Cyclin D1 and c-Myc quantified by densitometric analysis of Western blot and qPCR assay, respectively. The housekeeping gene β -actin was used as an internal control. Data are expressed as the mean \pm S.D., $n = 3$ per group. $##P < 0.01$ vs. the WT group. $*P < 0.05$, $**P < 0.01$, $***P < 0.001$ vs. the AKT/c-Met group. CELE-L and CELE-H represent intragastric administration of celecoxib at low (125 mg) and high (250 mg) doses, respectively.

supporting the important tumor suppressor role of PTEN for obstructing malignant transformation of hepatocytes in HCC, as observed in other chemotherapeutic processes in HCC cells.^{24,25} In the cytoplasm, activated/phosphorylated extracellular-signal-regulated kinases (ERKs) phosphorylate a separate set of substrates including kinases and cytoskeletal proteins in cellular signaling responses.²⁶ Aberrant ERK1/2 signaling is in part responsible for oncogenesis by targeting PTEN. Indeed, blockade of ERK activation by the MAP kinase/ERK kinase (MEK) inhibitor rescues PTEN suppression at both the transcriptional and post-translated levels.²⁷ Of note, our group²⁸ and others²⁹ have previously reported that celecoxib was effective at precluding the phosphorylation of ERK1/2 at Thr202/Tyr204 residues in hepatocytes. Thus, we believe that celecoxib-induced elevation of PTEN expression is highly relevant to its effect on ERK phosphorylation, especially in the context of hepatocarcinogenesis with overexpression of c-Met that serves as an upstream modulator of ERK/MEK signaling for stimulating ERK

activation. In addition, COX-2 is the inducible isoform of cyclooxygenases (COXs), the rate-limiting enzymes converting arachidonic acid to prostaglandins (PGEs), expressed constitutively during hepatocarcinogenesis, potentially fueling the excessive inflammatory response required for malignant transformation.³⁰ Selective COX-2 inhibitors (including celecoxib) bind the active center of COX-2 and further block its enzymatic function, thereby alleviating the production of PGEs (especially PGE₂). Indeed, previous reports suggest that induction of PTEN expression is a typical feature for the efficacy of celecoxib and another selective COX-2 inhibitor NS398 on cancer progression,^{31,32} indicating that suppression of PGE synthesis by celecoxib treatment contributes to the elevation of PTEN expression in the present investigation. Furthermore, accumulating evidence demonstrates that microRNAs (miRNAs) with pro-oncogenic properties may represent available targets for the anti-proliferative actions of NSAIDs in a wide variety of cancer subtypes.³³ For instance, aberrant expression of miR-21 is



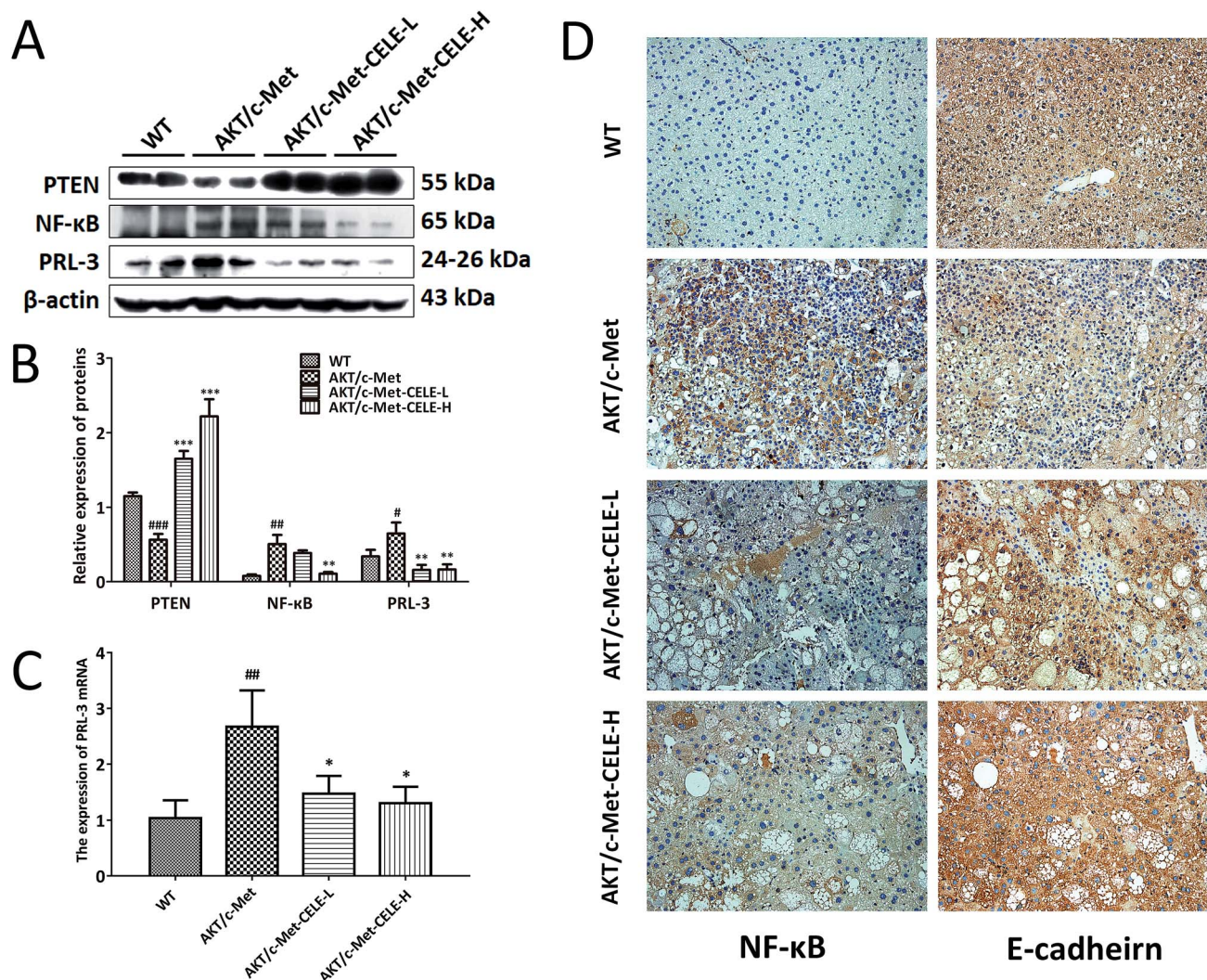


Fig. 3 Celecoxib elevates the expression of PTEN and E-cadherin, and suppresses the expression of NF- κ B and PRL-3. (A) Celecoxib enhances the protein expression of PTEN and downregulates the protein expression of NF- κ B and PRL-3 in livers of AKT/c-Met HCC-bearing mice. Liver samples from each group were quantified by Western blot assay, and representative bands are shown. (B) Histograms represent the protein expression of PTEN, NF- κ B and PRL-3 quantified by densitometric analysis of Western blots. (C) Histograms show the mRNA expression of PRL-3 quantified by qPCR. (D) The IHC staining of NF- κ B and E-cadherin shows that the AKT/c-Met-induced high expression of NF- κ B and low expression of E-cadherin were improved by celecoxib treatment. Original magnification: 200 \times . The housekeeping gene β -actin was used as an internal control. Data are expressed as the mean \pm S.D., $n = 3$ per group. # $P < 0.05$, ## $P < 0.01$, ### $P < 0.001$ vs. the WT group. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs. the AKT/c-Met group. CELE-L and CELE-H represent intragastric administration of celecoxib at low (125 mg) and high (250 mg) doses, respectively.

involved in both the initiation and progression of human cancers and plays its oncogenic roles in tumor malignant transformation by modulating PTEN expression directly at the 3' untranslated regions of PTEN.³⁴ Notably, upon administration of aspirin and NS398 repressed the expression of miR-21 in colon gastric carcinoma cells.^{31,33} Thus, it would be important to further test whether celecoxib influences a potential COX-2/PGEs/miR-21 axis that sequentially regulates the expression of PTEN in HCC initiation.

The deficiency or loss of PTEN contributes to the high proliferation index in HCC by acting through its downstream AKT/mTOR pathway involved in modulating phenotypic characteristics of cancer cells, including cell proliferation.³⁴ Inversely, elevation of PTEN expression inhibits colon cancer

progression by repressing AKT phosphorylation.³⁵ Although the previous evidence demonstrates that celecoxib induced apoptosis through its regulation of PDK1/PTEN/AKT pathway in HCC cells *in vitro*,³⁶ the PTEN-associated mechanism by which celecoxib inhibits cell proliferation in hepatocarcinogenesis remains unclear. In fact, NF- κ B is a dimer of Rel proteins that is sequestered in the cytoplasm as an inactive form through interaction with an inhibitory κ B ($\text{I}\kappa\text{B}$) protein. The PTEN/AKT/mTORC1 cascade activates NF- κ B predominantly through mechanisms that regulate nuclear translocation or trans-activation potential. Moreover, the degradation of $\text{I}\kappa\text{B}$ is responsible for constitutive activation of the NF- κ B signaling in HCC.³⁷ Furthermore, ubiquitination regulates the degradation



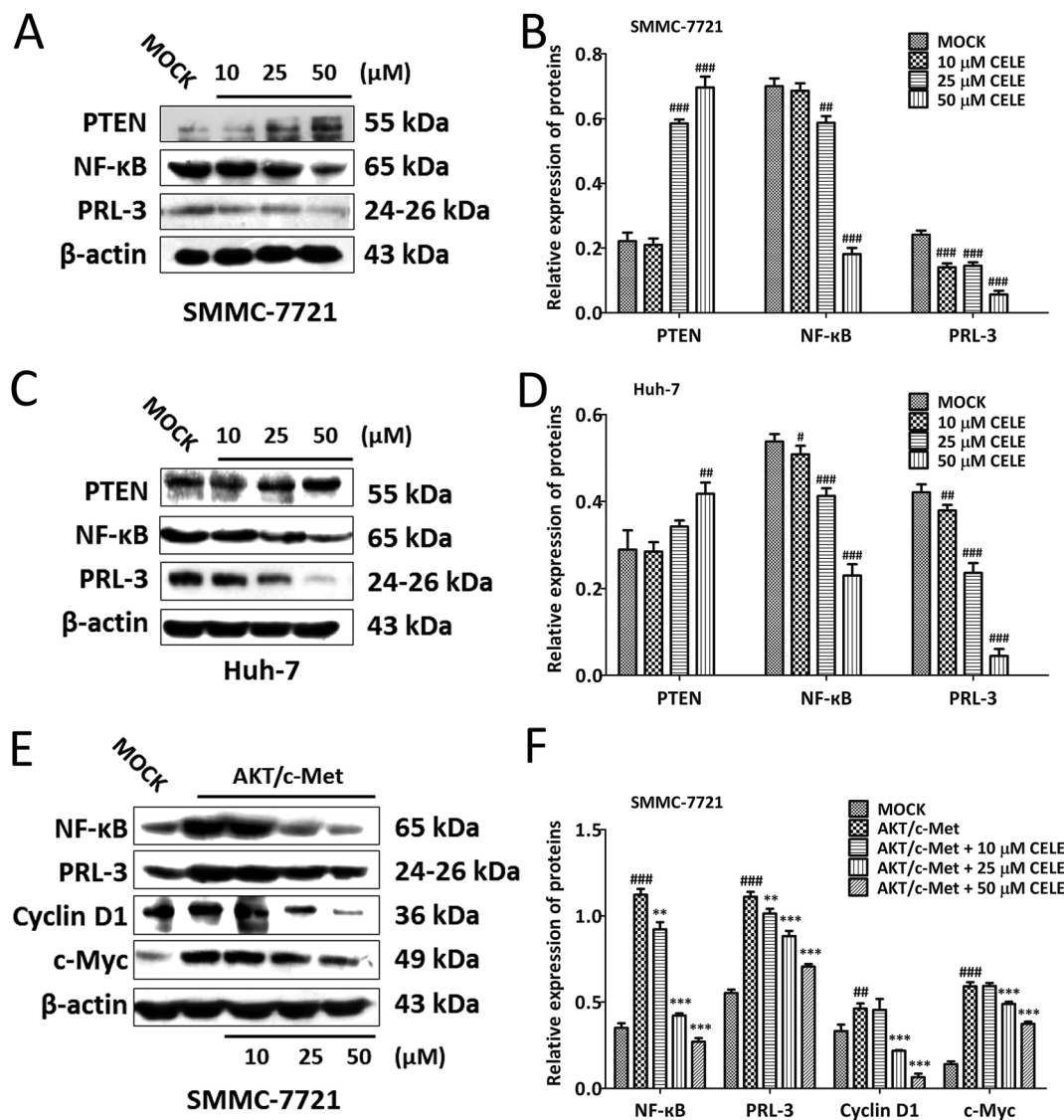


Fig. 4 Celecoxib enhances the protein expression of PTEN and inhibits the protein expression of NF- κ B and PRL-3 in HCC cells. Western blot analysis of the protein expression of PTEN, NF- κ B and PRL-3 in (A) SMMC-7721 cells and (C) Huh-7 cells. Histograms in (B) and (D) represent the intensity of the immunoblot bands in (A) and (C), respectively. (E) Western blot analysis for the protein expression of PTEN, NF- κ B and PRL-3 in AKT/c-Met-transfected SMMC-7721 cells. Histograms in (F) represent the intensity of the immunoblot bands in (E) quantified by densitometry. The housekeeping gene β -actin was used as an internal control. Data are expressed as the mean \pm S.D., $n = 3$ per group. # $P < 0.05$, ## $P < 0.01$, ### $P < 0.001$ vs. the MOCK group. ** $P < 0.01$, *** $P < 0.001$ vs. the AKT/c-Met group.

of I κ B, processing of NF- κ B precursors, and activation of I κ B kinase (IKK) in the NF- κ B pathway.³⁸ Inhibition of NF- κ B by proteasome inhibitors or prevention of I κ B phosphorylation induces apoptosis in myeloma cell lines.³⁹ In accordance with previous data in human gastric cancer cell lines,⁴⁰ our results revealed that the efficacy of celecoxib in AKT/c-Met HCC-bearing mice was associated with a marked reduction in the protein expression of NF- κ B subunit p65. Furthermore, our *in vitro* genetic studies demonstrated that PTEN is required for celecoxib-induced repression of NF- κ B expression. Therefore, it is possible that celecoxib decreases the elevated protein expression of NF- κ B during AKT/c-Met-driven hepatocarcinogenesis due to its suppressing action on I κ B ubiquitination and

degradation, which is associated with the celecoxib-induced elevation of PTEN expression. Obviously, it would be important to further test whether celecoxib influences the post-translational modification of I κ B that regulates NF- κ B activity in HCC.

Accumulating evidence indicates that PRL-3 could promote tumor cell proliferation and survival in various kinds of neoplasms, including lung, breast, colorectal and gastric carcinoma.⁴¹⁻⁴⁴ The aberrant PRL-3 expression fuels cell cycle progression by elevating the expression of Cyclin D1 and cyclin A-cyclin-dependent kinase 2 (CDK2), activating the signal transducer and activator of transcription 5 (STAT5)/AKT pathways and suppressing the expression of p21 WAF1/CIP1, a CDK



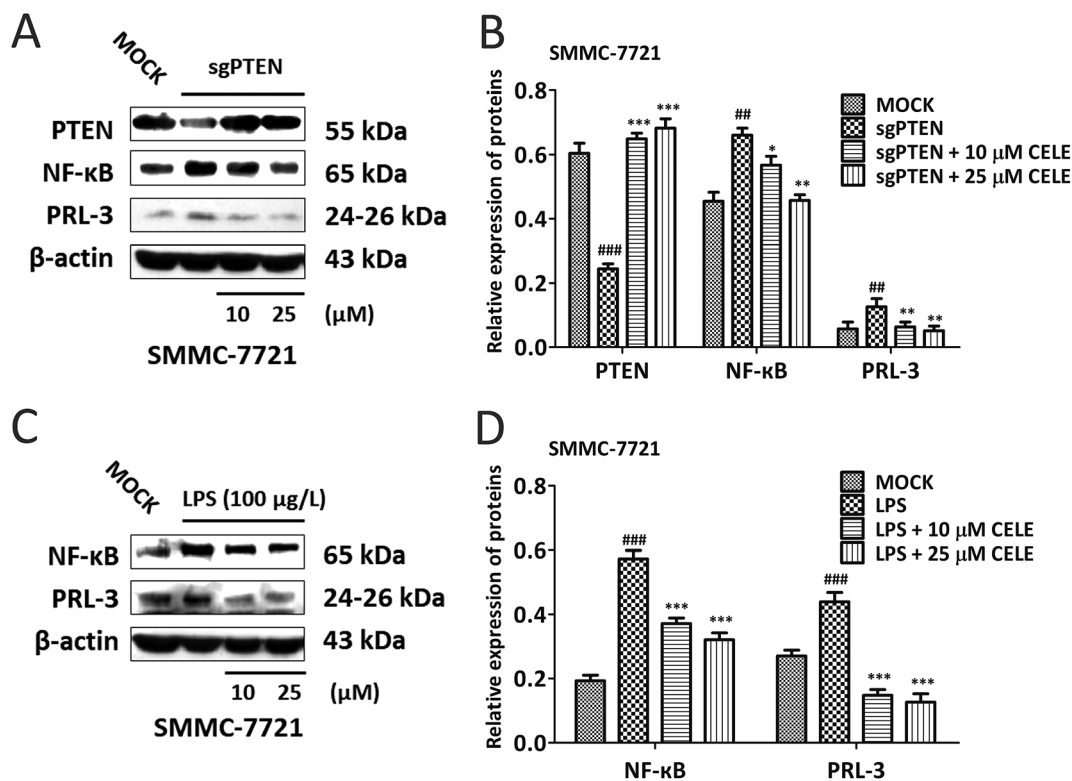


Fig. 5 Celecoxib regulates a PTEN/NF- κ B/PRL-3 pathway in HCC Cells. (A) Western blot analysis of the protein expression of PTEN, NF- κ B and PRL-3 in PTEN-silenced SMMC-7721 cells. Histograms in (B) represent the intensity of the immunoblot bands in (A) quantified by densitometry. (C) Western blotting analysis for the protein expression of NF- κ B and PRL-3 in LPS-induced SMMC-7721 cells. Histograms in (D) represent the intensity of the immunoblot bands in (C) quantified by densitometry. The housekeeping gene β -actin was used as an internal control. Data are expressed as the mean \pm S.D., $n = 3$ per group. $##P < 0.01$, $###P < 0.001$ vs. the MOCK group. $*P < 0.05$, $**P < 0.01$, $***P < 0.001$ vs. the sgPTEN-transfected group or the LPS-simulated group.

inhibitory protein that serves as an effector of the p53-dependent G1 arrest.⁴⁵ However, whether PRL-3 is a potential target for the anti-proliferative efficacy of celecoxib during HCC initiation is still unclear. For the first time, we found that celecoxib suppressed the expression of PRL-3 at the transcriptional and post-transcriptional levels during hepatocarcinogenesis. Importantly, NF- κ B, the ubiquitous transcription factor which binds DNA as a heterodimeric complex composed of members of the Rel/NF- κ B family of polypeptides, is responsible for transcriptionally activating the PRL-3 expression by binding the PRL-3 promoter.¹⁵ Our data suggests that celecoxib is capable of downregulating the expression of NF- κ B, which might impair its nuclear translocation and obstruct transcriptional activity, subsequently resulting in the suppression of PRL-3 mRNA transcription. Intriguingly, PRLs can also enhance the phosphorylation of ERK1/2, which could promote tumor cell proliferation by transcriptionally controlling the expression of c-Myc and Cyclin D1 in multiple types of cancer.^{46–48} Furthermore, as a metastasis associated tyrosine phosphatase, PRL-3 post-transcriptionally decreases the expression of PTEN by affecting multiple effectors (*e.g.*, vinculin and RhoA).⁴⁹ Thus, further investigations need to be performed to elucidate whether celecoxib could affect the PRL-3/ERK1/2 pathway or a potential PRL-3/PTEN feedback loop in excessively

proliferating hepatocytes during hepatocarcinogenesis with AKT and c-Met co-activation.

Conclusion

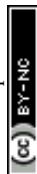
Altogether, our *in vitro* and *in vivo* data demonstrate that celecoxib could suppress hepatocellular proliferation by interfering with a PTEN/NF- κ B/PRL-3 pathway in hepatocarcinogenesis. Importantly, our study introduces PRL-3 as a novel therapeutic target of celecoxib in hepatocarcinogenesis. The present study also warrants further investigation of celecoxib and other COX-2 inhibitors for their anti-proliferative effect against HCC initiation. Furthermore, combined therapy of celecoxib with other drugs such as multikinase inhibitors should be evaluated to establish the clinical utility of NSAIDs for the prevention and treatment of HCCs.

Conflicts of interest

The authors declare that they have no competing interests.

Abbreviations

cDNA complementary DNA



CDK2	cyclin-dependent kinase 2
COX-2	cyclooxygenase-2
COXs	cyclooxygenases
ERK1/2	extracellular signal-regulated kinases 1/2
FASN	fatty acid synthase
HCC	hepatocellular carcinoma
IκB	inhibitory κB
IKK	IκB kinase
Ki-67	antigen Ki-67
LPS	lipopolysaccharide
MEK	MAPK/ERK kinase
mTORC1	mammalian target of rapamycin complex 1
NF-κB	nuclear factor κB
NSAIDs	nonsteroidal anti-inflammatory drugs
PGs	prostaglandins
PI3K	phosphoinositide 3-kinase
PRL	phosphatase of regenerating liver
PTP4A	class IVA of protein tyrosine phosphatase
PTEN	phosphatase and tensin homologue
qPCR	quantitative real-time polymerase chain reaction
STAT5	signal transducer and activator of transcription 5

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References

- 1 J. Wang, T. Yang, H. Chen, H. Li and S. Zheng, *Biochem. Biophys. Res. Commun.*, 2018, **498**, 424–430.
- 2 G. Deng, S. Zeng, Y. Qu, Q. Luo, C. Guo, L. Yin, Y. Han, Y. Li, C. Cai, Y. Fu and H. Shen, *J. Exp. Clin. Cancer Res.*, 2018, **37**, 156.
- 3 Y. Xiong, S. Wu, H. Yu, J. Wu, Y. Wang, H. Li, H. Huang and H. Zhang, *Exp. Cell Res.*, 2018, **371**, 185–195.
- 4 M. L. Sulis and R. Parsons, *Trends Cell Biol.*, 2003, **13**, 478–483.
- 5 L. Liu, H. Long, Y. Wu, H. Li, L. Dong, J. L. Zhong, Z. Liu, X. Yang, X. Dai, L. Shi, M. Ren and Z. Lin, *Cell. Signalling*, 2018, **50**, 90–99.
- 6 J. J. Li, H. K. Yin, D. X. Guan, J. S. Zhao, Y. X. Feng, Y. Z. Deng, X. Wang, N. Li, X. F. Wang, S. Q. Cheng, Y. Bao and D. Xie, *Br. J. Cancer*, 2018, **118**, 1337–1348.
- 7 C. Y. Chen, J. Chen, L. He and B. L. Stiles, *Front. Endocrinol.*, 2018, **9**, 338.
- 8 R. L. Dillon, D. E. White and W. J. Muller, *Oncogene*, 2007, **26**, 1338–1345.
- 9 A. S. Baldwin, *J. Clin. Invest.*, 2001, **107**, 241–246.
- 10 F. A. La Rosa, J. W. Pierce and G. E. Sonenshein, *Mol. Cell. Biol.*, 1994, **14**, 1039–1044.
- 11 M. D. Lin, H. T. Lee, S. C. Wang, H. R. Li, H. L. Hsien, K. W. Cheng, Y. D. Chang, M. L. Huang, J. K. Yu and Y. H. Chen, *BMC Dev. Biol.*, 2013, **13**, 18.
- 12 J. M. Salamoun, K. E. McQueeney, K. Patil, S. J. Geib, E. R. Sharlow, J. S. Lazo and P. Wipf, *Org. Biomol. Chem.*, 2016, **14**, 6398–6402.
- 13 W. B. Zhao, Y. Li, X. Liu, L. Y. Zhang and X. Wang, *Int. J. Mol. Med.*, 2008, **22**, 187–192.
- 14 L. Wang, J. Liu, Z. Zhong, X. Gong, W. Liu, L. Shi and X. Li, *Brain Res.*, 2016, **1646**, 441–450.
- 15 H. H. Gari, G. D. DeGala, M. S. Lucia and J. R. Lambert, *Oncogenesis*, 2016, **5**, e255.
- 16 C. Y. Wu, C. J. Wang, C. C. Tseng, H. P. Chen, M. S. Wu, J. T. Lin, H. Inoue and G. H. Chen, *World J. Gastroenterol.*, 2005, **11**, 3197–3203.
- 17 B. Vijayakumar, V. Kannappan and V. Sathyanarayananmoorthi, *J. Mol. Struct.*, 2016, **1121**, 16–25.
- 18 J. J. Hu, L. Che, L. Li, M. G. Pilo, A. Cigliano, S. Ribback, X. L. Li, G. Latte, M. Mela, M. Evert, F. Dombrowski, G. H. Zheng, X. Chen and D. F. Calvisi, *Sci. Rep.*, 2016, **6**, 20484.
- 19 J. J. Hu, X. Li, J. X. Zhou, C. Zhang, G. H. Zheng and Z. P. Qiu, *RSC Adv.*, 2018, **8**, 13976–13983.
- 20 S. S. Zhang, X. H. Song, D. Cao, Z. Xu, B. A. Fan, L. Che, J. J. Hu, B. Chen, M. J. Dong, M. G. Pilo, A. Cigliano, K. Evert, S. Ribback, F. Dombrowski, R. M. Pascale, A. Cossu, G. Vidili, A. Porcu, M. M. Simile, G. M. Pes, G. Giannelli, J. Gordan, L. X. Wei, M. Evert, W. M. Cong, D. F. Calvisi and X. Chen, *J. Hepatol.*, 2017, **67**, 1194–1203.
- 21 C. Stroescu, A. Dregnea, B. Ivanov, C. Pechianu, V. Herlea, O. Sgarbura, A. Popescu and I. Popescu, *J. Gastrointest. Liver Dis.*, 2008, **17**, 411–417.
- 22 F. Liang, J. Liang, W. Q. Wang, J. P. Sun, E. Udho and Z. Y. Zhang, *J. Biol. Chem.*, 2007, **282**, 5413–5419.
- 23 T. Luedde and R. F. Schwabe, *Nat. Rev. Gastroenterol. Hepatol.*, 2011, **8**, 108–118.
- 24 C. He, X. Dong, B. Zhai, X. Jiang, D. Dong, B. Li, H. Jiang, S. Xu and X. Sun, *Oncotarget*, 2015, **6**, 28867–28881.
- 25 X. Zhang, S. Jia, S. Yang, Y. Yang, T. Yang and Y. Yang, *J. Cell. Biochem.*, 2012, **113**, 3528–3535.
- 26 J. W. Ramos, *Int. J. Biochem. Cell Biol.*, 2008, **40**, 2707–2719.
- 27 S. E. Beck and J. M. Carethers, *Cancer Biol. Ther.*, 2007, **6**, 1313–1317.
- 28 Z. P. Qiu, C. Zhang, J. X. Zhou, J. J. Hu, L. Sheng, X. Li, L. Chen, X. Li, X. K. Deng and G. H. Zheng, *Mol. Carcinog.*, 2019, **58**, 31–41.
- 29 Y. H. Paik, J. K. Kim, J. I. Lee, S. H. Kang, D. Y. Kim, S. H. An, S. J. Lee, D. K. Lee, K. H. Han, C. Y. Chon, S. I. Lee, K. S. Lee and D. A. Brenner, *Gut*, 2009, **58**, 1517–1527.
- 30 H. Koga, S. Sakisaka, M. Ohishi, T. Kawaguchi, E. Taniguchi, K. Sasatomi, M. Harada, T. Kusaba, M. Tanaka, R. Kimura, Y. Nakashima, O. Nakashima, M. Kojiro, T. Kurohiji and M. Sata, *Hepatology*, 1999, **29**, 688–696.
- 31 H. Li, J. Cheng, Y. Mao, M. Jiang and X. Fan, *OncoTargets Ther.*, 2015, **8**, 3245–3253.
- 32 T. D. Kabir, R. J. Leigh, H. Tasena, M. Mellone, R. D. Coletta, E. K. Parkinson, S. S. Prime, G. J. Thomas, I. C. Paterson, D. Zhou, J. McCall, P. M. Speight and D. W. Lambert, *Aging*, 2016, **8**, 1608–1635.
- 33 E. Yiannakopoulou, *Cell. Oncol.*, 2014, **37**, 167–178.



- 34 F. Meng, R. Henson, H. Wehbe-Janek, K. Ghoshal, S. T. Jacob and T. Patel, *Gastroenterology*, 2007, **133**, 647–658.
- 35 L. L. Zhang, G. G. Mu, Q. S. Ding, Y. X. Li, Y. B. Shi, J. F. Dai and H. G. Yu, *J. Biol. Chem.*, 2015, **290**, 15018–15029.
- 36 J. Leng, C. Han, A. J. Demetris, G. K. Michalopoulos and T. Wu, *Hepatology*, 2003, **38**, 756–768.
- 37 G. He and M. Karin, *Cell Res.*, 2011, **21**, 159–168.
- 38 Z. J. Chen, *Nat. Cell Biol.*, 2005, **7**, 758–765.
- 39 H. Ni, M. Ergin, Q. Huang, J. Z. Qin, H. M. Amin, R. L. Martinez, S. Saeed, K. Barton and S. Alkan, *Br. J. Haematol.*, 2001, **115**, 279–286.
- 40 Z. Chen, M. Liu, X. Liu, S. Huang, L. Li, B. Song, H. Li, Q. Ren, Z. Hu, Y. Zhou and L. Qiao, *Int. J. Mol. Med.*, 2013, **32**, 93–100.
- 41 Y. X. Lian, R. Chen, Y. H. Xu, C. L. Peng and H. C. Hu, *Gene*, 2012, **511**, 169–176.
- 42 P. den Hollander, K. Rawls, A. Tsimelzon, J. Shepherd, A. Mazumdar, J. Hill, S. A. Fuqua, J. C. Chang, C. K. Osborne, S. G. Hilsenbeck, G. B. Mills and P. H. Brown, *Cancer Res.*, 2016, **76**, 1942–1953.
- 43 J. Zhang, Z. Xiao, D. Lai, J. Sun, C. He, Z. Chu, H. Ye, S. Chen and J. Wang, *Br. J. Cancer*, 2012, **107**, 352–359.
- 44 E. R. Sharlow, P. Wipf, K. E. McQueeney, A. Bakan and J. S. Lazo, *Expert Opin. Invest. Drugs*, 2014, **23**, 661–673.
- 45 S. Qu, B. Liu, X. L. Guo, H. S. Shi, M. F. Zhou, L. Li, S. L. Yang, X. Z. Tong and H. H. Wang, *Cancer*, 2014, **120**, 2130–2141.
- 46 G. Pintus, B. Tadolini, A. M. Posadino, B. Sanna, M. Debidda, F. Bennardini, G. Sava and C. Ventura, *Eur. J. Biochem.*, 2002, **269**, 5861–5870.
- 47 E. Halilovic, Q. B. She, Q. Ye, R. Pagliarini, W. R. Sellers, D. B. Solit and N. Rosen, *Cancer Res.*, 2010, **70**, 6804–6814.
- 48 Y. Cao, Y. Tu, J. Mei, Z. Li, Z. Jie, S. Xu, L. Xu, S. Wang and Y. Xiong, *Mol. Med. Rep.*, 2013, **7**, 1805–1811.
- 49 H. Wang, S. Y. Quah, J. M. Dong, E. Manser, J. P. Tang and Q. Zeng, *Cancer Res.*, 2007, **67**, 2922–2926.

