


Cite this: *RSC Adv.*, 2019, 9, 4325

In situ polymerization and FT-IR characterization of poly-glycine on pencil graphite electrode for sensitive determination of anti-emetic drug, granisetron in injections and human plasma†

Marwa F. B. Ali‡ and Fatma A. M. Abdel-aal *

In situ polymerization is a simple and efficient technique for modification and fabrication of modified electrodes in voltammetry. An efficient and highly sensitive square wave voltammetric (SWV) method was developed for analysis of a 5-HT₃ antagonist granisetron (GRN) using *in situ* polymerized glycine on pencil graphite electrode surface. It was found that the fabricated polymer enhanced the sensitivity by more than two times and enhanced the surface activity by more than three times. Surface area measurements showed that poly-Gly/PGE have large surface area of 44.3 mm², when compared to that of bare PEG (12.1 mm²). Several methods as cyclic voltammetry (CV), electrochemical impedance spectroscopy (EIS) and scanning electron microscopy (SEM) techniques were used to characterize the modified surface. Further, FT-IR spectroscopic study was used to predict the way of glycine polymerization on electrode surface and the possible interaction mechanism with GRN. After optimization, the proposed method showed a linear response of GRN concentrations in the range from 0.08 to 3.00 μmol L⁻¹ with a limit of detection (LOD) of 26.2 nmol L⁻¹ (9.14 ng mL⁻¹). The method was utilized for GRN determination in ampoules and in real human plasma samples.

Received 8th January 2019

Accepted 28th January 2019

DOI: 10.1039/c9ra00179d

rsc.li/rsc-advances

1. Introduction

Electro-polymerization is an innovative approach to fabricate polymer film modified electrodes (PMEs), this process takes place *in situ* for immobilization of organic molecules to form highly sensitive conducting polymer over electrode surface. PMEs have attracted much attention due to their high stability, selectivity, sensitivity, reproducibility, good homogeneity and strong adherence to electrode surface.^{1–3} In the last two decades the use of poly(amino acid) for electrode modification was applied in the electroanalysis field, especially poly-glycine (poly-Gly), that have been used for determination of many pharmaceutical compounds.^{4–11} Although poly-Gly polymer was used for modification of carbon electrodes previously, but the utility of poly-Gly as a conducting polymer at surface of graphite electrode was not reported before. In addition, the proposed method investigated the mechanism of glycine polymerization and the structure of the formed polymer over electrode surface using FT-IR technique.

Granisetron (GRN), is a 5-HT₃ antagonists used in treatment of nausea and vomiting especially that caused by cancer chemotherapy.¹² Serotonin (5-HT) receptors are distributed throughout the mucosal enterochromaffin cells in the gut and the central nervous system.¹³ Serotonin is released by the enterochromaffin cells in response to chemotherapeutic agents that may stimulate vagal afferents (*via* 5-HT₃ receptors) to initiate the vomiting reflex. GRN suppresses vomiting and nausea by inhibiting serotonin binding to 5-HT₃ receptors.

Various analytical methods were reported for quantitative determination of GRN in bulk, different pharmaceutical and biological samples, as spectrophotometric,^{14,15} spectrofluorimetric,¹⁶ HPTLC,^{17,18} and HPLC^{19–22} methods. Electrochemical techniques are widely used nowadays for their high sensitivity, selectivity, cost effective and high sample throughput within short analysis time. From literature survey, there is only two electroanalytical methods were reported for determination of GRN.^{23,24} The 1st one²³ used hanging mercury drop electrode (HMDE) which suffer from several drawbacks such as toxicity of mercury, surface area of a drop of mercury is never constant, applied voltage produces changes in surface tension and hence change in drop size with limitation for analysis of more positive potential range. The 2nd one²⁴ utilized platinum nanoparticles modified glassy carbon electrode (GCE). Although the reported sensor showed higher sensitivity,²⁴ but the authors have not provided a justification about

Department of Pharmaceutical Analytical Chemistry, Faculty of Pharmacy, Assiut University, 71526 Assiut, Egypt. E-mail: famo207@yahoo.com; fatmamoustafa@aun.edu.eg; Fax: +20-88-2080774

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c9ra00179d

‡ These authors contributed equally to this work.



the number of electrons involved in the suggested oxidation mechanism of GRN. The authors suggested that the oxidation of GRN is attributed to the loss of two electrons in two steps in acidic medium (pH 2.0) while there is only one peak developed as shown in their results. In addition, the reported method²⁴ was not applied in biological fluids and the authors didn't prove its liability for determination of GRN in human plasma.

The several advantages of pencil graphite electrode (PEG) over other carbon electrodes, such as the low cost, availability, low background current, wide potential window and ease of modification, made it favorable for many workers for analysis of different compounds in pharmaceutical and biological samples.^{25–27} Therefore, this work aimed to fabricate poly-Gly modified graphite electrode for the 1st time for determination of GRN at nanoscale level in pharmaceutical formulations and in real human plasma. Full characterization study of the formed poly-Gly polymer was performed using scanning electron microscopy (SEM), electrochemical impedance spectroscopy (EIS), cyclic voltammetric (CV) and square wave voltammetric (SWV) techniques. Furthermore, FT-IR spectroscopic study was carried out to investigate the mechanism of glycine polymerization on graphite electrode surface and its chemical reaction with the studied drug GRN.

2. Experimental

Materials, chemicals, apparatus and all preparation procedures used during the whole study are mentioned in the ESI.† Full fabrication process of poly-Gly/PGE electrode is shown in Fig. 1.

3. Results and discussion

3.1. Preliminary investigations of GRN on different carbon electrodes

SWV technique was utilized to investigate the electrochemical behavior of GRN at various carbon electrodes such as glassy carbon electrode (GCE) and carbon paste electrode (CPE). Fig. 2 showed the SW voltammograms of 1.0×10^{-5} M GRN in 0.04 M of BR (pH 7.0) at GCE (curve a), CPE (curve b), PGE bare (curve c). GRN has a broad weak peak at GCE at 0.77 V and a slightly

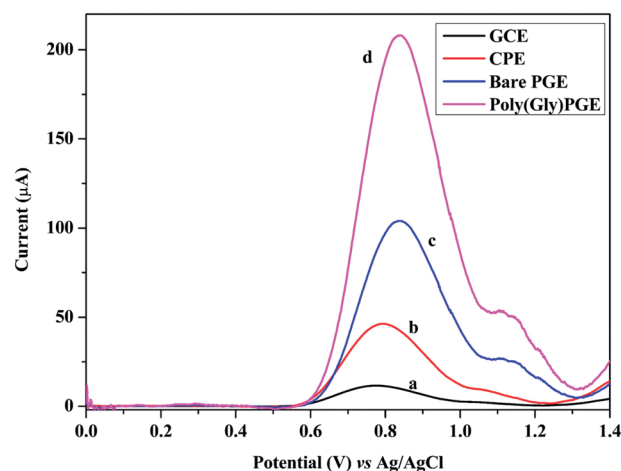


Fig. 2 Square wave voltammograms for 1.0×10^{-5} M GRN solution at (a) GCE, (b) CPE, (c) bare PGE and (d) poly-Gly/PGE. Parameters: 0.04 M BR buffer (pH = 7.0), frequency = 150 Hz, pulse height = 5 mV, step height = 5 mV, accumulation time = 60 s.

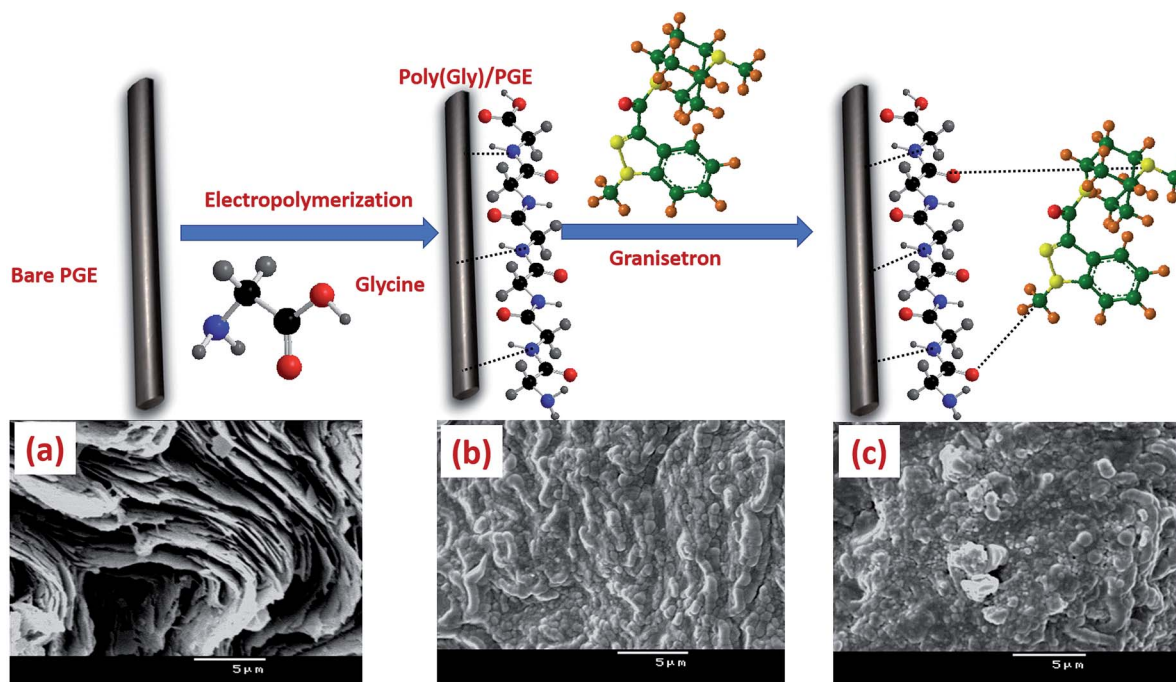


Fig. 1 Preparation process of poly-Gly/PGE with SEM images of (a) bare PGE, (b) poly-Gly/PGE and (c) poly-Gly/PGE with an adsorbed layer of GRN.



higher peak at CPE at 0.8 V, while a sharp and high peak was produced at PGE bare at 0.85 V. From the results shown in Fig. 2, PGE was selected for subsequent measurements.

The next step was the electro-polymerization of glycine on the surface of PGE after its activation in 0.2 M phosphate buffer at pH 7.0. After modification of the PGE surface with poly-Gly polymer, the anodic current of GRN significantly increased to the double and becomes sharper when compared to that of the bare PGE electrode. These results indicated that poly-Gly polymer can enhance the rate of electron transfer for GRN oxidation and act as a promoter. The *in situ* formed poly-Gly layer had an excellent catalytic activity on the electrochemical oxidation of GRN on PGE electrode as shown in Fig. 2 (curve d).

3.2. Electro-polymerization parameters of glycine on PGE

Various polymerization conditions such as potential range, number of scan cycles, conc. of glycine and pH of the medium which may affect the catalytic oxidation of GRN were investigated. A solution of 1.0×10^{-5} M GRN in 0.04 M BR at pH 7.0 was used to evaluate the catalytic results throughout the experiments.

Polymerization of glycine on PGE surface is achieved *via* CV technique. A wide potential range for polymerization of glycine was set from negative value -0.5 V to 2.0 V as reported before.²⁸ Number of cyclic scans of the electrode in glycine solution was also optimized as it is one of the significant factors that affect the electrocatalytic activity of the formed polymer. As indicated in Fig. 3a, increasing the no. of scanning cycles increases the thickness of the deposited polymer layers and consequently an enhancement in the oxidation peak current of GRN was observed. The peak current increases by increasing the number of scans until 7 cycles, however higher number of scans more

than 7 cycles showed a decrease in peak current value. This can be attributed to the amount of the modifier attached to the surface of the electrode that reached a saturation point after 7 cycles.

The concentration of glycine monomer also affects the catalytic activity of the electrode. As shown in Fig. 3b, by increasing the concentration of glycine the oxidation current is significantly increased. However, if the concentration of glycine was higher than 1.0 mM, the resultant film blocks the surface of the electrode and the sensitivity of the poly-Gly/PGE electrode was decreased.

Different pH values of phosphate buffer were tested from pH 3.0 to 7.0 for electro-polymerization of glycine as indicated in Fig. 3c. Glycine has different forms (acidic, basic or zwitterion), hence the electrolyte pH has a great influence on the polymerization process.²⁹

Glycine has 2 pK_a values (2.2 and 9.6) and it was found that the best pH for its polymerization is the pH 5.0. This pH is near its isoelectric point and considered the optimum for formation of poly-Gly polymer at the surface of the electrode. Above pH 5.0 the peak of GRN is distorted and disappeared. Hence, phosphate buffer at pH 5.0 was chosen for polymerization of glycine on electrode surface throughout the SWV measurements.

3.3. Characterization of poly-Gly modified PGE

3.3.1. SEM characterization. The surface morphology studies of cross sections of (a) bare PGE and (b) modified poly-Gly/PGE, fabricated by electro-polymerization with 7 electrochemical cycles, and (c) modified poly-Gly/PGE after adsorption of GRN oxidation products on the surface of the electrode, SEM images are shown in Fig. 1. In Fig. 1a, bare PGE appears as thin fragile layers of graphite with large pores in between whereas

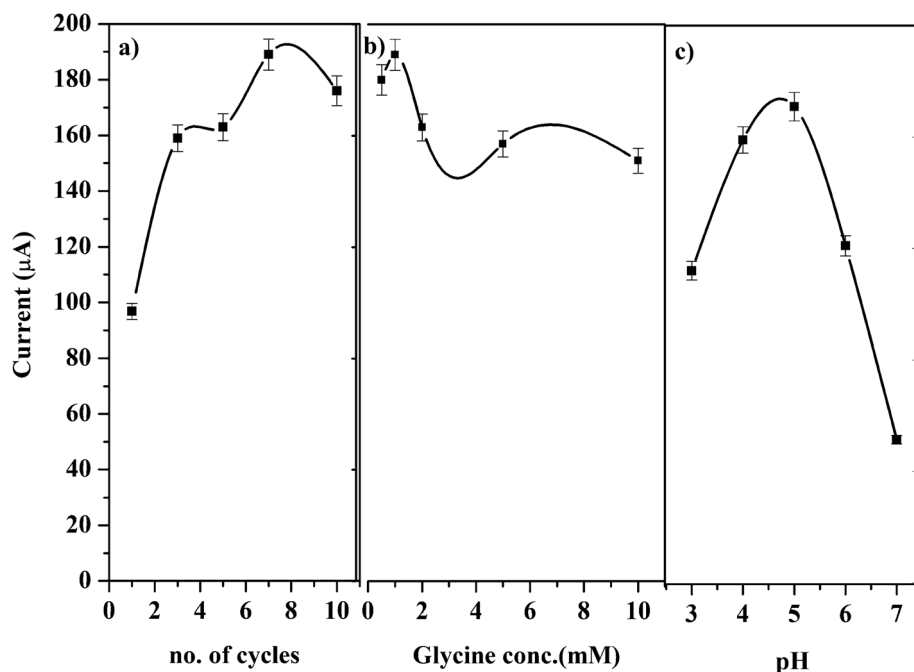


Fig. 3 Effects of (a) scanning cycle, (b) concentration of glycine, and (c) pH of the medium on the peak current of 1.0×10^{-5} M GRN solution.

after polymerization of glycine on the surface of the electrode as indicated in Fig. 1b, the electrode surface is covered with a compact layer of poly-Gly polymer dispersed all over the surface as chains of small particles. After polymerization of glycine on the electrode surface, the modified electrode is used as a working electrode for determination of GRN. The presence of poly-Gly polymer promotes the oxidation of GRN and the oxidation products of GRN are adsorbed on the surface of the electrode extensively as appeared in Fig. 1c. The adsorbed oxidation products appear as a white layer spread on the surface of the modified electrode that assures the occurrence of adsorption at the surface of the electrode.

3.3.2. FT-IR spectral characterization of the formed polymer. The formation of poly-Gly polymer on PGE surface is examined by FT-IR spectroscopy. Fig. 4 presents the FT-IR spectra of glycine monomer (Fig. 4a) and poly-Gly polymer (Fig. 4b). The spectra show fewer distinct absorption bands, by conversion of glycine peptide to poly-Gly polymer. There is a well-recognized band at 1400 cm^{-1} in the spectra of glycine as shown in (Fig. 4a). This band becomes less intense and appears only as a hump in the spectrum of poly-Gly polymer (Fig. 4b). This band is probably due to ionized carboxyl groups from dipolar or zwitter ion forms, and it would be expected that contributions from each molecule decreased with increasing the number of molecules present in the polymer. Regarding to the spectral region around 3000 cm^{-1} , the spectra in this region are characterized by absorption bands of bonded N-H stretching, symmetrical and non-symmetrical C-H stretching of CH_2 . It can be seen in (Fig. 4a) that there is a small band at 3160 cm^{-1} and it is slightly shifted to a higher wavenumber with the increase in band intensity that appeared at 3300 cm^{-1} as the molecular weight of the glycine peptides is increased. The band at 3300 cm^{-1} is a hydrogen bonded N-H as has been suggested to be the intermolecular hydrogen bonding between the molecules forming the polymer. This explanation agrees with the results published before by Blout *et al.*³⁰ and Fischer *et al.*³¹

3.4. Electrochemical activity of the modified poly-Gly/PGE

The effective electrode area was examined by the CV method using $5.0\text{ mM K}_3\text{Fe}(\text{CN})_6$ as a standard at various scan rates.

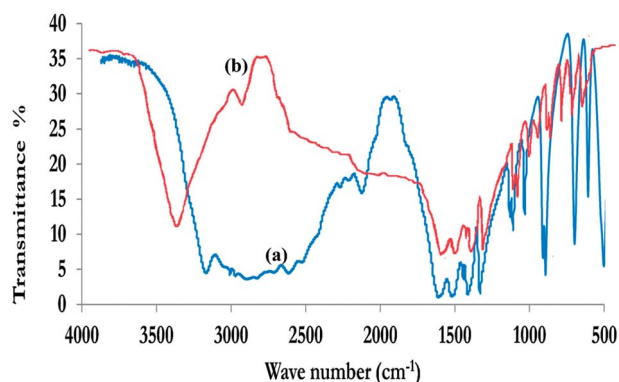


Fig. 4 FT-IR spectra of (a) glycine monomer and (b) poly-glycine polymer.

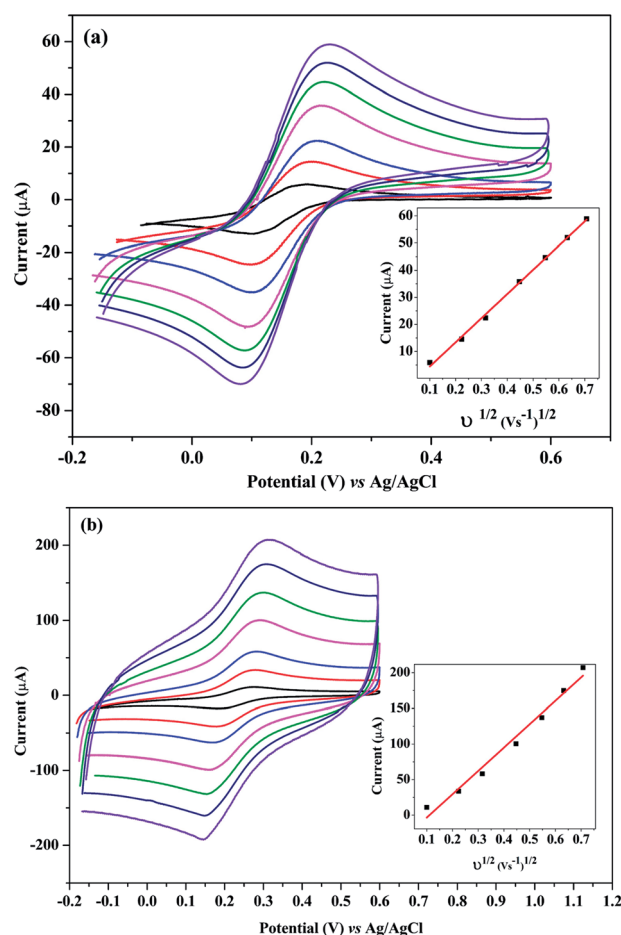


Fig. 5 Cyclic voltammograms for the electrochemical responses of $\text{K}_3[\text{Fe}(\text{CN})_6]$ at (a) bare PGE and (b) poly-Gly/PGE in 0.5 M KCl containing $1.0\text{ mM K}_3[\text{Fe}(\text{CN})_6]$ at different scan rates (0.01 to 0.4 V s^{-1}).

Randles-Sevcik equation was used³² for a reversible process as following:

$$I_p = (2.69 \times 10^5) A n^{3/2} D_R^{1/2} C_0 \nu^{1/2}$$

where, I_p refers to anodic peak current, A is the surface area of the electrode, n is the number of electrons transferred, D_R is diffusion coefficient, ν is the scan rate and C_0 is the concentration of $\text{K}_3\text{Fe}(\text{CN})_6$, respectively.

From the slope of the plot of I_p vs. $\nu^{1/2}$ at different scan rates (0.01 to 0.4 V s^{-1}) as shown in Fig. 5(a and b), the surface areas of the bare and poly-Gly/PGE were calculated to be 12.1 and 44.3 mm^2 , respectively. These results indicated that after PGE surface modification by Gly-polymerization, the effective surface area was increased by more than three times. This improvement is reflected by the increment in peak current of $1.0\text{ mM K}_3\text{Fe}(\text{CN})_6$ on poly-Gly/PGE (Fig. 5b) when compared with bare PGE (Fig. 5a). This confirms the change in surface property of the modified electrode. Hence, the fabricated poly-Gly/PGE has stable and sensitive electrochemical behavior.



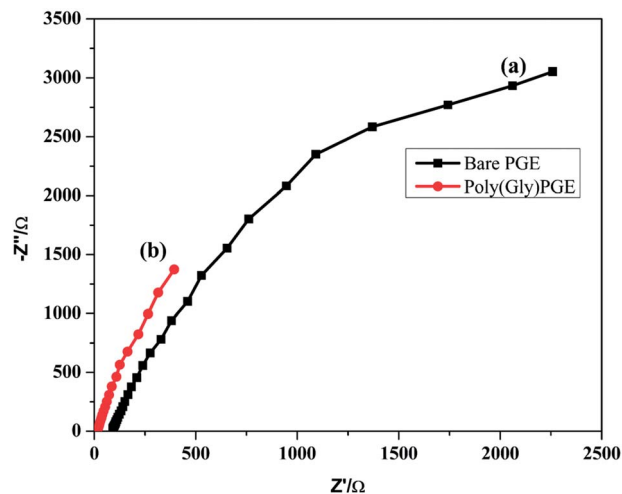


Fig. 6 Nyquist plot of ESI of (a) bare PGE and (b) poly-Gly/PGE using 0.5 M KCl containing 1.0 mM $K_3[Fe(CN)_6]$ at scan rate 0.1 V s^{-1} .

3.5. Electrochemical impedance spectroscopy (EIS)

EIS was examined to calculate the interfacial properties of the electrode and solution. Charge transfer resistance (R_{ct}) is an important character of electron transfer across the electrode interface. EIS of bare PGE and poly-Gly/PGE in 1.0 mM $K_3[Fe(CN)_6]$ in 0.5 M KCl were examined at the open circuit potential over the frequency range of 1 Hz to 100 kHz. The data were fitted according to the Randles equation, and the resulting Nyquist plots are shown in Fig. 6. The Nyquist plot of bare PGE shows a semi-circular model, whereas that of poly-Gly/PGE shows linear curve. The charge transfer resistance values (R_{ct}) were calculated by z-view software and found to be 108.3 and 18.47 for bare PGE and poly-Gly/PGE, respectively. This result indicates that the poly-Gly polymer coating on PGE surface enhanced electron transfer across the electrode–solution interface.

3.6. Effect of scan rate

Effect of scan rate on GRN oxidation was studied at the poly-Gly/PGE by CV. The cyclic voltammograms for $1.0 \times 10^{-5} \text{ M}$ GRN in 0.04 M BR (pH 7.0) at the poly-Gly/PGE at various scan rates from 0.01 to 0.4 V s^{-1} were recorded (Fig. 7a). In order to know whether the reaction on the surface of the proposed electrode was under diffusion or adsorption control, scan rate studies were carried out. The relationship between the oxidation peak current (I_{P_1} or I_{P_2}) and the scan rate (ν) showed a linear response. Further, the relationship between logarithm scan rate ($\log \nu$) on the logarithm peak current ($\log I_P$) was also investigated (Fig. 7b and c), giving a straight line with a slope of 0.5653 and 0.4805 for 1st and 2nd peak, respectively, according to the following equations:

$$\log I_{P_1}(\mu\text{A}) = 0.5653 \log \nu + 2.6231 \text{ (V s}^{-1}\text{)} \quad (r = 0.9800)$$

$$\log I_{P_2}(\mu\text{A}) = 0.4805 \log \nu + 2.6067 \text{ (V s}^{-1}\text{)} \quad (r = 0.9983).$$

Consequently, the oxidative stripping of GRN was found to be adsorption–diffusion controlled process.

By increasing of the scan rate, the peak potential of GRN was moved to more positive values that ensure the irreversibility of GRN oxidation process. A linear response between $E_{P_{1,2}}$ and $\log \nu$ in the range from 0.01 to 0.4 V s^{-1} because after this scan rate the 2nd peak disappears. Fig. 7b and c indicated both relations that represented as:

$$E_{P_1}(\text{V}) = 0.1737 \log \nu \text{ (V s}^{-1}\text{)} + 0.9175 \quad (r = 0.9608)$$

$$E_{P_2}(\text{V}) = 0.1345 \log \nu \text{ (V s}^{-1}\text{)} + 1.1977 \quad (r = 0.9682).$$

In the CV reverse scan, no cathodic peak was observed, hence the reaction would be represented according to Laviron theory for totally irreversible electrode process,³³ by the following equation:

$$\text{Slope} = 2.303RT/\alpha nF \quad (3)$$

where T is the absolute temperature (298 K), n the number of electrons transferred in the rate determining step, α is the transfer coefficient, R is the universal gas constant ($8.314 \text{ J mol}^{-1} \text{ K}^{-1}$) and F is the Faraday constant ($96.480 \text{ C mol}^{-1}$). The value of αn can be calculated from the slope of E_P vs. $\log \nu$ plot for the 1st and 2nd peak, which were found to be 0.1737 and 0.1345, respectively. The values of αn were calculated as 0.34 and 0.44 for the 1st and 2nd peak, respectively. Assuming that (α) equals 0.5, the number of electrons transferred in the two steps of the oxidation process will be equal to 0.68 and 0.88 which is close to one electron in each step. These calculations agree with the proposed oxidation mechanism of GRN in Fig. 8.

3.7. Mechanism of the electro-oxidation of GRN using poly-Gly/PGE

GRN metabolized *in vivo* assuming that oxidation occurs first on the nitrogen atom of the *N*-methyl in the indazole ring,³⁴ which is electroactive resulting in hydroxylation of the benzene ring with formation of 7-hydroxy granisetron. In order to investigate the oxidation behavior of GRN, Douliche M. *et al.*²⁴ perform a comparative study between indole and GRN using cyclic voltammetry at bare GCE as a function of pH. It was found that the cyclic voltammogram of indole is closely related to that of GRN in phosphate buffer at pH 7.0. Also, the behavior of GRN oxidation is closely related to that of omeprazole as reported by Jorge. S. M. A. *et al.* in 2009.³⁵ At the indazole moiety, the first oxidation step occurs on N_1 by loss of one electron (giving the 1st oxidation peak P_1) forming radical cation that is stabilized by transfer of the positive charge to the ortho position. On addition of water, 7-hydroxygranisetron is produced as indicated in Fig. 8 (the same as in hydroxylation pathway that occurs inside the body³⁶). The oxidation is favored on N_1 more than other nitrogen atoms present in the compound due to the high availability of its electrons (charge on each nitrogen is calculated *via* Chem Draw program, and it was found as the following: -0.6432 , $+0.2732$, $+0.2325$ and -0.1209 for N_1 , N_2 ,



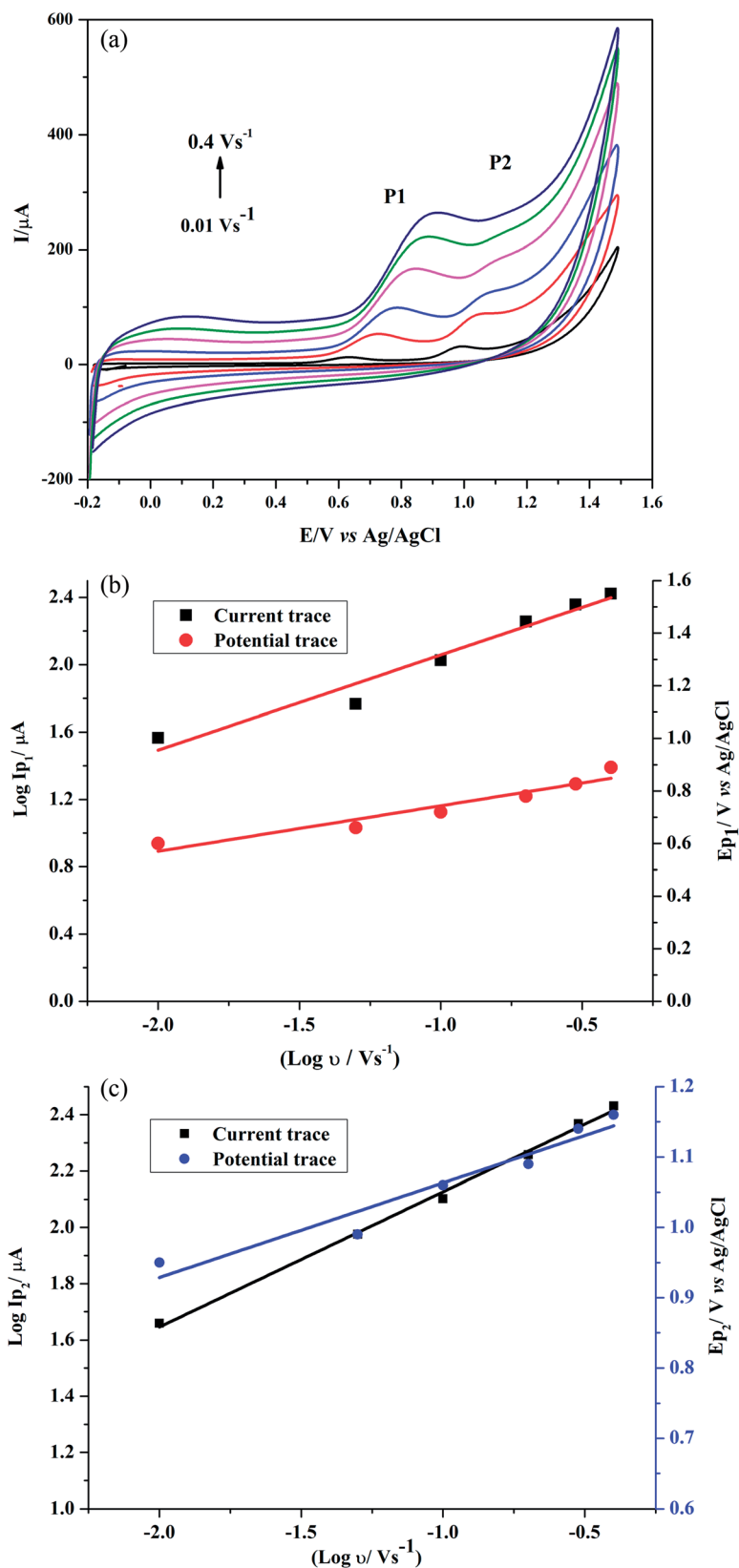


Fig. 7 (a) Cyclic voltammograms to study the effect of different scan rates (0.01 to 0.4 V s^{-1}) on the electrochemical response of $1.0 \times 10^{-5} \text{ M}$ GRN solution. Relationship between \log scan rate ($\log \nu$) and \log peak current ($\log I_p$) or potential (V) for the 1st peak (b) and the 2nd peak (c).



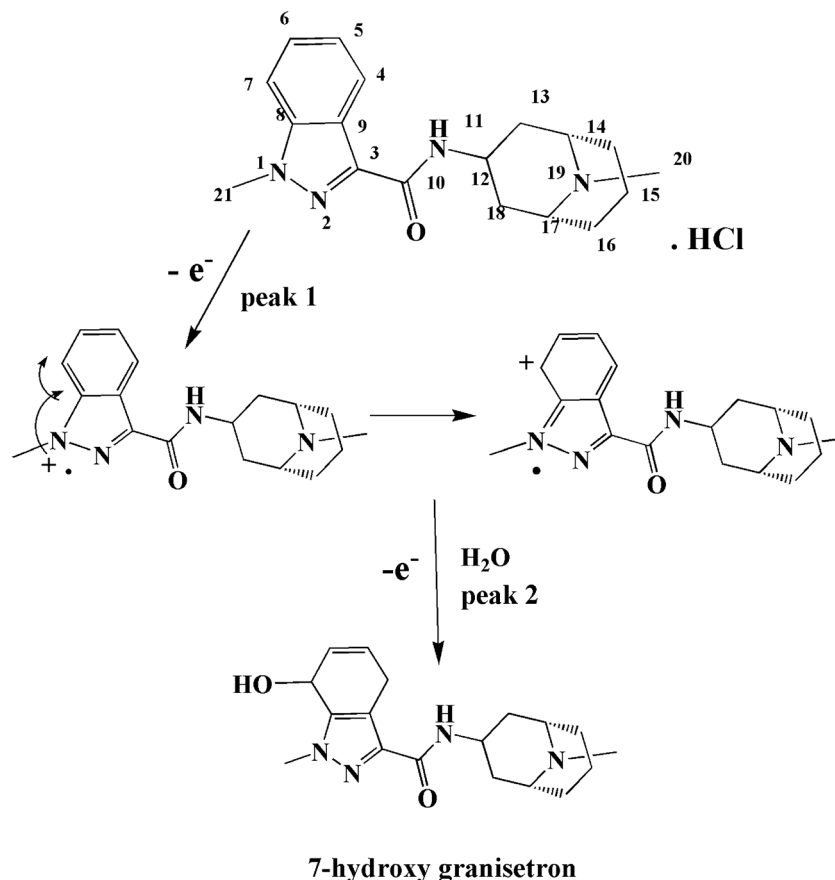


Fig. 8 The proposed oxidation mechanism of granisetron.

N_{11} and N_{19} , respectively). In the 2nd oxidation step peak, there is a loss of another electron leading to formation of 7-hydroxy granisetron. As indicated before in Fig. 7a, the two oxidation steps appear at slow scan rates while on increasing the scan rate the 2nd oxidation peak began to decrease till disappearance or overlapping with the 1st one. So, the 1st peak was chosen for further quantitative determination of GRN.

3.8. Influence of supporting electrolyte and pH

The type of supporting electrolytes and pH value of buffer solution has a great influence on the voltammetric response of GRN. The current responses of 1.0×10^{-6} M GRN were investigated in various supporting electrolytes such as BR, citrate and phosphate buffer solutions. As observed from Fig. 9, the oxidation peaks of GRN showed relatively high sensitivity and better shape in BR buffer solution. Further, the effect of pH on the oxidation peak current (I_p) of 1.0×10^{-6} M GRN was examined in 0.04 M of BR buffer at various pH ranged from 4.0 to 8.0 and it was found that it increases with increasing pH value until it reaches 7.0 and then decreases again. Hence, BR buffer at pH 7.0 has been chosen for quantitative analysis of GRN for further experiments. While the second oxidation peak P_2 that only appears after pH 7.0 as a small hump as shown in Fig. 9a, wasn't used for quantitation. The plot between E_{P_1} and pH (Fig. 9b) showed that E_p decreased gradually by increasing the electrolyte pH according to the following equation:

$$E_p = -0.09 \text{ pH} + 1.58 \quad (r = 0.9879).$$

The slope of the equation was 90 mV which is away from the theoretical value (59 mV) indicating that this step is not pH dependent.

The effect of anions in different supporting electrolytes such as KNO_3 , KCl and KClO_4 on GRN peak current was also examined. It was found that a well-defined peak of GRN was observed in BR buffer containing KNO_3 salt. Different concentrations of KNO_3 solutions were tested from 0.001 to 0.1 M, and it was found that the best result was obtained with 0.01 M KNO_3 solution.

3.9. Influence of voltammetric parameters

Various voltammetric parameters were examined over the range of 3–25 mV for pulse height, 10–250 Hz for frequency and 3–25 mV for step height. It was observed that pulse height of 10 mV, frequency of 250 Hz and step height of 10 mV were the optimum one. These parameters greatly affect the voltammograms for higher sensitivity and excellent peak shape. The effect of accumulation potential and time on the oxidation peak current of GRN was examined also by SWV. The accumulation potential, E_{acc} was investigated between -0.5 V and 0.5 V. The E_{acc} was increased from -0.5 V to -0.2 V and then the oxidation peak current of GRN became stable then gradually decreased. Hence, E_{acc} of -0.2 V was selected for further analysis.



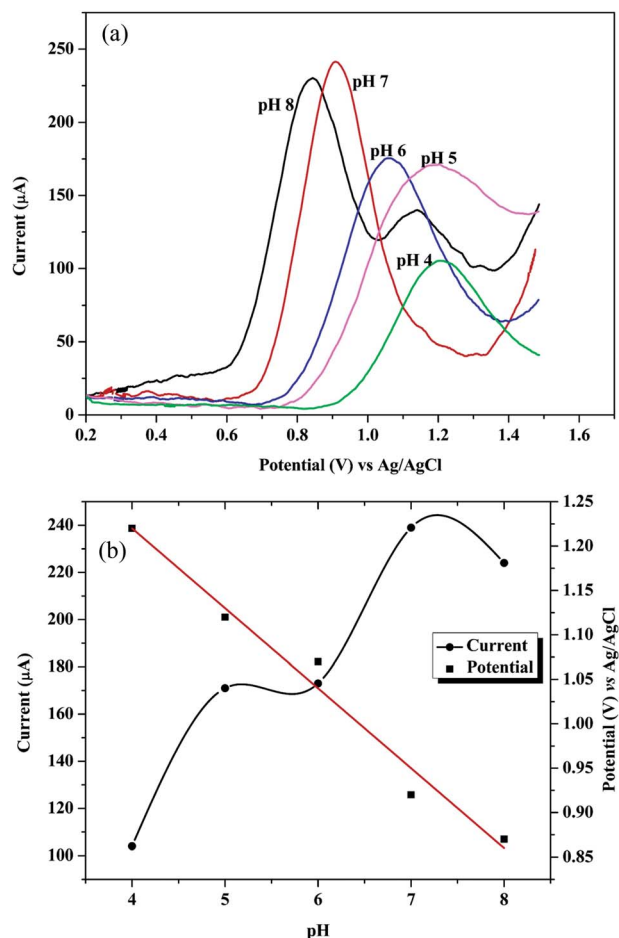


Fig. 9 (a) Square wave voltammograms for the effect of different pHs on the oxidation peak of 1.0×10^{-5} M GRN solution at poly-Gly/PGE; (b) effect of pH of the supporting electrolyte on the peak oxidation current and potential of 1.0×10^{-5} M GRN solution.

Moreover, the effect of accumulation time (t_{acc}) had a great influence on GRN peak current. The oxidation peak current of GRN at poly-Gly/PGE increased with t_{acc} up to 120 s at the E_{acc} of -0.2 V, suggesting that accumulation improves method sensitivity. When t_{acc} was further increased from 120 to 180 s, the oxidation peak current decreased revealing that the amount of GRN at poly-Gly/PGE reached a saturation state at electrode surface.

3.10. Validation results

Under the optimum conditions, the proposed method was validated according to International Conference on Harmonization (ICH) guidelines.³⁷

3.10.1. Linearity range. The proposed modified electrode showed a linear response oxidative stripping of GRN using the 1st peak in the concentration range from 0.08 to $3.0 \mu\text{mol L}^{-1}$ with good correlation coefficient of 0.9998. While the 2nd peak is weak and appeared as a hump at moderately high scan rate, so it can't be used for quantitation. Detection and quantification limits were statistically calculated as $3.3\sigma/b$ and $10\sigma/b$, respectively, where b is the slope and σ is the standard deviation of intercept. The proposed method showed detection and

quantification limits of 26.2 nmol L^{-1} (9.14 ng mL^{-1}) and 79.4 nmol L^{-1} (27.7 ng mL^{-1}), respectively. Fig. 10 shows the typical SWV of different concentrations of GRN at the modified electrode. It was found that the oxidation peak current increased linearly with increasing the concentration of GRN in the range from 0.08 to $3.0 \mu\text{mol L}^{-1}$. Inset figure shows the linear calibration plot over the concentration range under the optimum conditions. Table 1 shows the limits of detection (LOD) and quantification (LOQ). The low values of LOD and LOQ reflect high sensitivity of the proposed modified electrode towards electro-oxidation of GRN.

3.10.2. Precision. The precision of the presented method was determined by intra-day and inter-day precision by analyzing nine samples (for intra-day) and 45 samples at 5 days (for inter-day) for the concentration of $1.0 \mu\text{mol L}^{-1}$. The % RSD values of the peak current of GRN were 2.67 and 3.12 for intra- and inter-day precision, respectively. The results indicate that the method is precise, and confidence as shown in Table 1.

3.11. Interferences study

Co-existing compounds capable of competing with GRN in the adsorption site on the electrode surface may result in a decrease or an increase in peak current. Therefore, the effects of common excipients, biologically active compounds and divalent metals were examined to evaluate the selectivity of the developed procedure. The tolerance limit for interfering species was defined as the maximum concentration of foreign substances that made a relative error less than 5% for determination of $1.0 \mu\text{mol L}^{-1}$ GRN under the optimum conditions. For this investigation, the results show that over 200-fold excess of hydroxypropyl methylcellulose, lactose, magnesium stearate, 100-fold of polyethylene glycol, starch, and ascorbic acid, 50-fold of uric acid, glucose, sucrose, citric acid and some metals such as Cu^{2+} and Zn^{2+} did not affect GRN response. The recovery results were

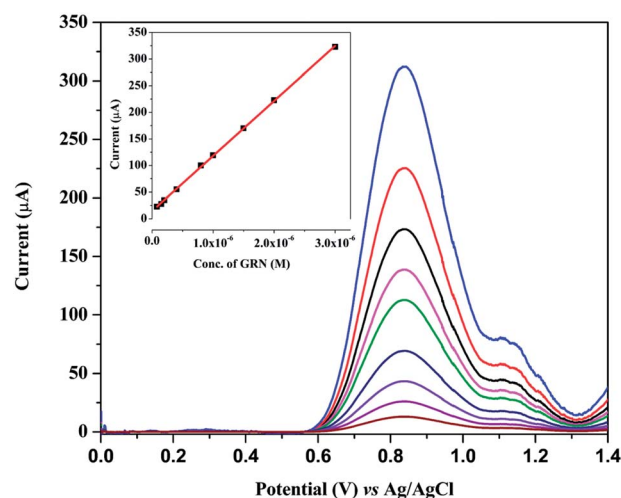


Fig. 10 Square wave voltammograms for different concentrations of GRN (0.08 to $3.0 \mu\text{M}$). Optimum parameters: 0.04 M BR buffer (pH = 7.0), $E_{acc} = -0.2$ V, frequency = 250 Hz, pulse height = 10 mV, step height = 10 mV and $t_{acc} = 120$ s. Inset: calibration curve between concentration and peak current.



Table 1 Regression data of the calibration line for quantitative determination of GRN in pure form and in human plasma

Parameter	Pure form	Spiked human plasma
Linearity range ($\mu\text{mol L}^{-1}$)	0.08–3.00	0.25–3.40
LOD (nmol L^{-1}) ^a	26.2	47.21
LOQ (nmol L^{-1}) ^b	79.4	143.07
Intercept, a ($\mu\text{mol L}^{-1}$) \pm SD ^c	14.65 \pm 0.534	75.27 \pm 1.054
Slope, b ($\mu\text{A } \mu\text{mol}^{-1} \text{ L}$) \pm SD ^d	103.41 \pm 0.413	21.88 \pm 0.104
Correlation coefficient (r)	0.9998	0.9997
Determination coefficient (r^2)	0.9997	0.9994
Intra-day precision ($n = 9$) ^e (% RSD)	Not more than 2.67	Not more than 3.04
Inter-day precision, 5 days, ($n = 45$) ^e (% RSD)	Not more than 3.12	Not more than 3.45

^a Limit of detection. ^b Limit of quantitation. ^c Standard deviation of the intercept. ^d Standard deviation of the slope. ^e Number of experiments.

ranged from 96.1 to 103.2% for 1.0 $\mu\text{mol L}^{-1}$ GRN solution, indicating that there was no matrix interference with GRN by the proposed SWV method. This indicates the selectivity of the proposed method and hence its suitability for the determination of GRN in pharmaceuticals and in complex matrices.

3.12. Application of the proposed method to pharmaceutical and biological samples

The proposed electrode was successfully utilized for determination of GRN in pharmaceutical formulation; the results of analysis of GRN are recorded in Table 2. Standard addition method was used, where known quantities of pure GRN were mixed with definite amounts of the studied formulation and the mixtures were analyzed. The total amount of the drug was then measured, and the amount of the added drug was determined by the difference. Good recoveries were obtained (96.5–98.0%) with good agreement with the label claim as shown in Table 2. Further, the results were statistically compared to those obtained by a previously reported method.³⁸ From the values of the calculated t and F at 95% confidence level, it can be revealed that there is no significant difference between the proposed method and the reported one.³⁸

The applicability of the proposed electrode for detection of GRN in human plasma was attempted. GRN is a potent and highly selective 5-HT₃ receptor and it is detectable in human plasma at less than 30 min following IV injection.^{39,40} The proposed sensor shows a linear response obtained by spiking plasma samples with GRN in the concentration range from 0.25 to 3.40 $\mu\text{mol L}^{-1}$.

The linear relationships can be expressed according to the following equation:

$$I_P (\mu\text{A}) = 75.27 C_{\text{GRN}} (\mu\text{mol L}^{-1}) + 21.88 (r = 0.9997, n = 8).$$

The proposed method shows detection and quantification limits in plasma of 47.21 and 143.07 nmol L^{-1} , respectively as shown in Table 1. Good recoveries of GRN achieved from plasma matrix, confirming the suitability of the proposed method for analysis of GRN in biological fluid. Further, real human plasma samples obtained from humans receiving GRN ampoule were collected after 20 min from IV injection, analyzed by the proposed SWV method and the results were summarized in Table 2. Concentration of GRN in real human plasma was calculated from calibration equation.

The good recoveries results of GRN obtained from plasma matrices, confirms the suitability, feasibility and cost effective of the proposed method for analysis of GRN in biological fluid when compared with previously reported electrochemical and HPLC methods.^{23,41–44}

The developed method is characterized by its simplicity and speed and although its sensitivity is not higher than other reported polarographic and chromatographic methods,^{24,41–44} but it has many advantages over them. The developed method prevents the toxicity hazards of mercury used in polarographic technique and it offers many advantages over HPLC such as speed and low consumption of organic solvents. Added to that, from the economic point of view, it does not need sophisticated and expensive instrumentation and it can be easily utilized for

Table 2 Recovery and standard addition method results of GRN in Em–Ex ampoule and human plasma using poly(Gly)/PGE

Em-Ex ampoule				Human plasma			
Labeled claim	3 mg			Conc. claimed after 20 min	0.25 μmol L ⁻¹		
Amount found	2.96 mg			Conc. found after 20 min ^b	0.24 μmol L ⁻¹		
% Recovery ± SD ^a	98.5 ± 1.05			% Recovery ± SD ^a	96.0 ± 3.35		
Student <i>t</i> - and <i>F</i> -test ^c	<i>t</i> _{calc.} = 1.54 <i>F</i> _{calc.} = 1.76						
Standard addition method							
Added (mg)	0.10	0.15	0.20	Added (μg)	10	20	30
Found (mg)	0.098	0.145	0.193	Found (μg)	9.63	19.34	29.05
% Recovery ± SD ^a	98.0 ± 1.37	96.6 ± 1.15	96.5 ± 0.98	% Recovery ± SD ^a	96.3 ± 2.45	96.70 ± 2.84	96.8 ± 2.52

^a Average of 5 determinations. ^b Average of 5 volunteers. ^c At confidence level 95%, theoretical t and F -values equal 3.182 and 2.36, respectively.



analysis of a lot of biological samples in short time. Its sensitivity is not far from previously reported methods, but it is highly sensitive enough to detect it in plasma without the need for more complicated extraction techniques that used in other methods and highly expensive detectors (like mass detectors).

3.13. Stability, repeatability and reproducibility of poly-Gly/PGE

The SWV of 1.0 μM of GRN was recorded daily to evaluate the stability of the newly modified electrode after preservation in a closed container. The fabricated sensor showed stable results over 10 days and the peak retained was calculated to be $97.3 \pm 2.03\%$ of its initial value. This confirms the high stability of the fabricated poly-Gly/PGE.

4. Conclusion

In the presented work, a highly sensitive poly-Gly/PGE was prepared and utilized for voltammetric determination of GRN. All electrochemical parameters were studied and optimized. The poly-Gly polymer coating PGE surface enhanced the electron transfer of GRN across the electrode–solution interface. The SWV method was validated according to ICH guidelines and showed higher sensitivity ($\text{LOD} = 26.2 \text{ nmol L}^{-1}$) with high precision value ($\% \text{ RSD} < 3.12$). The modified electrode was characterized via several methods as cyclic voltammetry (CV), electrochemical impedance spectroscopy (EIS) and scanning electron microscopy (SEM) techniques. Further, FT-IR characterization of the formed poly-Gly layer was investigated and the mechanism of the formed electrode was further discussed. The proposed method was successfully applied to determine GRN in its dosage forms and in human plasma. No interference from matrix of real samples was observed and the proposed method was highly selective for analysis of GRN.

Conflicts of interest

The authors declare that there is no conflict of interest.

Acknowledgements

The authors gratefully acknowledge Prof. Abdel Maaboud Ismail Mohamed (Pharmaceutical Analytical Chemistry Department, Faculty of Pharmacy, Assiut University, aimragerh@yahoo.com) for his valuable help in this research article.

References

- Y. Ohnuki, T. Ohsaka, H. Matsuda and N. Oyama, *J. Electroanal. Chem. Interfacial Electrochem.*, 1983, **158**, 55–67.
- L. Yang, F. Zhao and B. Zeng, *Electrochim. Acta*, 2016, **210**, 293–300.
- S. A. Kumar, C. F. Tang and S. M. Chen, *Talanta*, 2008, **74**, 860–866.
- J. G. G. Manjunatha, *J. Food Drug Anal.*, 2017, 1–8, DOI: 10.1016/j.jfda.2017.05.002.
- B. R. L. Ferraz, F. R. F. Leite and A. R. Malagutti, *J. Solid State Electrochem.*, 2015, **20**, 2509–2516.
- A. M. Yu, H. L. Zhang and H. Y. Chen, *Electroanalysis*, 1997, **9**, 788–790.
- C. M. Kuskur, K. B. E. Kumara Swamy and H. Jayadevappa, *J. Anal. Bioanal. Tech.*, 2015, **6**, 1–6.
- T. Thomas, R. J. Mascarenhas, B. E. Swamy, P. Martis, Z. Mekhalif and B. S. Sherigara, *Colloids Surf., B*, 2013, **110**, 458–465.
- G. Zhang, P. He, W. Fenga, S. Ding, J. Chena, L. Li, H. He, S. Zhang and F. Dong, *J. Electroanal. Chem.*, 2016, **760**, 24–31.
- Y. Li, X. Liu and W. Wei, *Electroanalysis*, 2011, **23**, 2832–2838.
- V. N. Palakollu, N. Thapliyal, T. E. Chiwunze, R. Karpoomath, S. Karunanidhi and S. Cherukupalli, *Mater. Sci. Eng., C*, 2017, **77**, 394–404.
- R. de Wit, M. Aapro and P. Blower, *Cancer Chemother. Pharmacol.*, 2005, **56**, 231–238.
- M. A. Kamm, *Aliment. Pharmacol. Ther.*, 2002, **16**, 343–351.
- P. Panzade, P. Puranik, V. Mogal and M. Patnni, *Asian J. Res. Chem.*, 2010, **3**, 634–636.
- I. I. Hewala, M. M. Bedair and S. M. Shousha, *Drug Test. Anal.*, 2013, **5**, 234–241.
- S. Lakshmana Prabu, A. Joseph, A. A. Shirwaikar and R. S. Jeyaprakash, *Indian Drugs*, 2008, **45**, 563–566.
- P. S. Prabu, P. Selvamani and S. Latha, *Lat. Am. J. Pharm.*, 2010, **29**, 1455–1458.
- A. V. Halerao and S. V. Shirolkar, *J. Pharm. Res.*, 2012, **5**, 3208–3212.
- F. C. Chen, L. H. Wang, J. Guo, X. Y. Shi and B. X. Fang, *J. Anal. Methods Chem.*, 2017, **2017**, 6749087.
- Y. Zhou, J. Jiang, P. Hu and H. Wang, *Biomed. Chromatogr.*, 2014, **28**, 1597–1600.
- I. Hewala, H. El-Fataty, E. Emam and M. Mabrouk, *J. AOAC Int.*, 2001, **94**, 1447–1460.
- I. Hewala, H. El-Fataty, E. Emam and M. Mubrouk, *Talanta*, 2010, **82**, 184–195.
- R. Jain and R. Sharma, *J. Pharm. Anal.*, 2012, **2**, 443–449.
- M. Douliche, N. K. Bakirhan, B. Uslu, B. Saidat, M. Trari and S. A. Ozkan, *Sens. Actuators, B*, 2017, **251**, 572–582.
- M. I. Said, A. H. Rageh and F. A. M. Abdel-aal, *RSC Adv.*, 2018, **8**, 18698–18713.
- F. A. M. Abdel-aal, A. H. Rageh, M. I. Said and G. A. Saleh, *Anal. Chim. Acta*, 2018, **1038**, 29–40.
- F. A. M. Abdel-aal and M. F. B. Ali, *J. Electrochem. Soc.*, 2017, **164**, H1053–H1063.
- A. M. Yu, H. L. Zhang and H. Y. Chen, *Electroanalysis*, 1997, **9**, 788–790.
- G. Herlem and T. Gharbi, in *Biosensors – Emerging Materials and Applications*, ed. P. A. Serra, Institute FEMTO-ST/Department of Optic, UMR CNRS 6174, University of Franche-Comte, France, 2011, ch. 10, pp. 179–208.
- E. R. Blout and S. G. Linsley, *J. Am. Chem. Soc.*, 1952, **74**, 1946–1951.
- G. Fischer, X. Cao, N. Cox and M. Francis, *Chem. Phys.*, 2005, **313**, 39–49.
- A. J. Bard and L. R. Faulkner, *Electrochemical Methods, Fundamentals and Applications*, Wiley, New York, 1980.



- 33 E. Laviron, *J. Electroanal. Chem.*, 1979, **101**, 19–28.
- 34 <https://www.drugbank.ca/drugs/DB00889>, (accessed 17/2/2018).
- 35 S. M. A. Jorge, A. D. R. Pontinha and A. M. Oliveira-Bretta, *Electroanalysis*, 2010, **22**, 625–631.
- 36 H. Nakamura, N. Ariyoshi, K. Okada, H. Nakasa, K. Nakazawa and M. Kitada, *Curr. Drug Metab.*, 2005, **6**, 469–480.
- 37 *International Conference on Harmonization (ICH) Topic Q2 (R1): Validation of Analytical Procedure: Text and Methodology*, Nov 2005.
- 38 S. A. Kanchana, A. Aruna, V. Niraimathi and A. J. Suresh, *Int. J. Chem. Sci.*, 2010, **8**, 1169–1173.
- 39 A. Gurpide, B. Sadaba, S. Martin-Algarra, J. R. Azanza, J. M. Lopez-Picazo, M. A. Campanero, J. P. Cabello, I. Gil-Aldea, S. de la Cruz, V. Fernandez Gallego, C. Reyna, C. Olier Garate, M. J. Blanco-Prieto, J. Ceballos, J. Garcia-Foncillas and J. L. Perez-Gracia, *Oncologist*, 2007, **12**, 1151–1155.
- 40 I. Wada, T. Takeda, M. Sato, H. Saitoh, T. Nakabayashi, K. Mino, T. Honma, M. Takada and K. Hirano, *Biol. Pharm. Bull.*, 2001, **24**, 432–435.
- 41 V. K. Boppana, *J. Chromatogr. A*, 1995, **692**, 195–202.
- 42 F. Pinguet, F. Bressolle, P. Martel, D. Salabert and C. Astre, *J. Chromatogr. B: Biomed. Sci. Appl.*, 1996, **675**, 99–105.
- 43 S. Xiang, W. Wang, B. Xiang, H. Deng and S. Xie, *Int. J. Mass Spectrom.*, 2007, **262**, 174–179.
- 44 R. Jin-min, L. Yan-rong, J. Ye, D. Xiang-yu and H. Fu, *Chin. J. Mod. Appl. Pharm.*, 2008, **4**.

