PAPER

Concise total syntheses of phelligridins A, C, and D†

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We have established a concise and scalable synthetic pathway for phelligridins A (**1**), C (**2**) and D (**3**). The synthetic highlights were Suzuki–Miyaura coupling and aldol-type condensation of α-pyrone. Phelligridin A was synthesized in four steps, while phelligridins C and D were each synthesized in six steps. Furthermore, we have revealed that the whole structure is essential for the cytotoxicity of phelligridins.

Phelligridin A (**1**) is a polyphenol isolated from the fruiting body of *Phellinus igniarius*, a fungus collected in Liaoning Province, China in 2003 (Fig. 1).[1] A year later, analogs of **1**, phelligridins C (**2**) and D (**3**), were also isolated from the fungus.[2] Also in 2004, meshimakobnols A and B were independently isolated from the Japanese mushroom of the same genus (*Phellinus linteus*). Meshimakobnols A and B were found to be identical to **3** and **2**, respectively.[3] Structurally, they possess a tricyclic fused ring system including two adjacent α-pyrone rings. In addition, they inhibit the cell growth of a variety of cancer cells.[2,4] However, the reported values of cytotoxicity against lung cancer A549 cells differed greatly. Furthermore, hispidin (**4**), a similar pyrone compound to phelligridin D (**3**), inhibits in a dose-dependent manner the activity of BACE 1, which produces amyloid β, a causative biomolecule of Alzheimer's disease.[5] Recently, Shigemori and co-workers reported on the inhibitory effect of

phelligridin D (**3**) on the aggregation of 42-mer amyloid β.[6] However, the biological studies were limited to those of the natural products or their congeners. Because a structure–activity relationship study on phelligridins/meshimakobnols is expected to provide lead compounds for cancer and Alzheimer's disease, we decided to establish an adaptable synthetic route for their structure–activity relationships. Herein, we described a concise and scalable total synthesis of phelligridins A (**1**), C (**2**), and D (**3**) and their biological evaluation.

The retrosynthetic pathway of phelligridins is shown in Scheme 1. Phelligridins C (**2**) and D (**3**) would be derived from dimethylphelligridin A (**6**) by aldol-type condensation with the corresponding benzaldehyde. Dimethylphelligridin A (**6**) would be synthesized from boronate ester **7** and bromopyrone **8** by Suzuki–Miyaura coupling. In this synthetic plan, several analogs of the phelligridins can be synthesized by changing the two coupling partners of the aldol-type condensation and Suzuki–Miyaura coupling.

Our synthesis of phelligridins started from the commercially available α-pyrone **9** (Scheme 2). Bromination[7] and protection of **9** gave the bromopyrone **8**, a precursor of Suzuki–Miyaura coupling. The coupling reaction between **8** and boronate ester **7** (ref. 8) followed by removal of the MOM group and concomitant
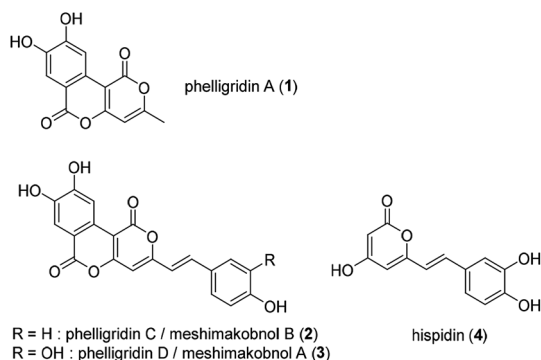

Fig. 1 Structure of phelligridins A (**1**), C (**2**), D (**3**), and hispidin (**4**).

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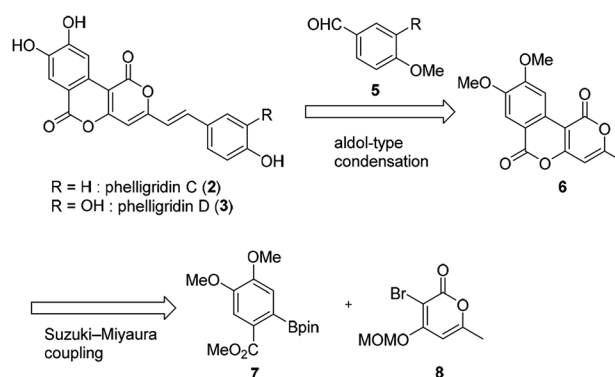

Scheme 1 Retrosynthetic pathway of phelligridins.
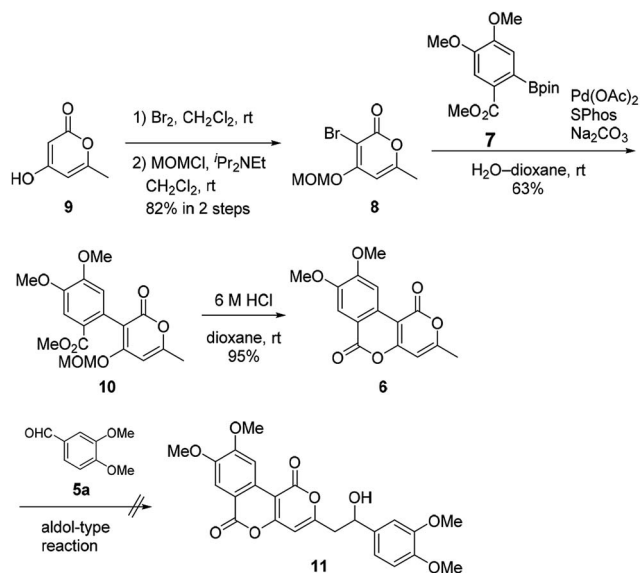
Scheme 2 Synthesis of dimethylphelligridin A (**6**) and aldol-type reaction.

lactonization gave dimethylphelligridin A (**6**) as a precursor of the aldol-type condensation. Next, we examined the synthesis of phelligridin D (**3**). The aldol-type reaction of dimethylphelligridin A (**6**) and aldehyde **5a** was carried out under reported conditions, LHMDS/THF[9] or ^{t}BuOK/DMF.[10] However, under these conditions, **6** was insoluble in solvent, and the reaction did not proceed.

Therefore, we then tried aldol-type reaction with **10** (Scheme 3). The aldol reaction of **10** and **5a** gave the aldol **12** in moderate yield. Subsequently, the aldol **12** was dehydrated *via* mesylation, and the MOM group was removed to obtain tetramethylphelligridin D (**14**). Finally, removal of the four methyl groups in **14** under HCl·py gave phelligridin D (**3**). Although we had achieved the total synthesis of **3**, there are three problems needed to be solved to adapt the synthetic route to a scalable preparation.
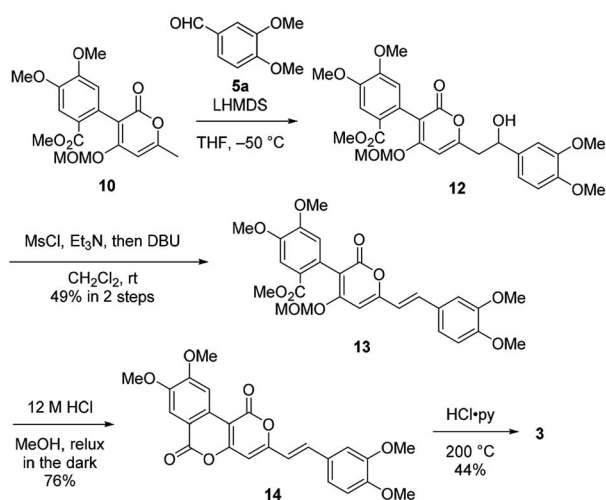
Firstly, the solubility of phelligridin D, as well as that of the synthetic intermediates from compound **10**, was remarkably low in a variety of solvents. Therefore, there were restrictions on reaction conditions and purification methods. Secondly, isomerization of the trans olefin in **3** and **14** occurred easily by light. Finally, the demethylation of **14** required severe reaction conditions. All the aforementioned drawbacks resulted in a low overall yield for phelligridin D (**3**). In order to solve these three problems, we decided to protect the phenolic hydroxy groups as MOM ethers.

Suzuki–Miyaura coupling of bromopyrone **8** and di-MOM boronate ester **15** gave the coupling compound **16** (Scheme 4). The aldol-type reaction with di-MOM aldehyde **17** and dehydration afforded **18**. In this case, the aldol-type reaction of **16** proceeded in high yield presumably due to improved solubility of **16**. Finally, phelligridin D (**3**) was synthesized by deprotection of five MOM groups under mild acidic conditions.

Having established a synthetic route for phelligridin D (**3**), the total synthesis of phelligridin A (**1**) and C (**2**) was carried out using synthetic intermediate **16** (Scheme 5). Removal of the three MOM groups in **16** gave phelligridin A (**1**). In addition, phelligridin C (**2**) was synthesized in three steps from **16** by changing the aldehyde of the aldol-type reaction to **19**.

With the synthetic samples in hand, the cytotoxicity against A549 cells and HeLa S3 cells of phelligridins were evaluated (Table 1). Phelligridin C (**2**) and D (**3**) showed cytotoxicity against A549 cells with IC_{50} of 1.6 μM and 1.1 μM, respectively. These values were closer to those reported by Nagatsu than by Shi.[11] On the other hand, phelligridin A (**1**), a left segment of phelligridin C (**2**) and D (**3**), and hispidin (**4**), a right segment of phelligridin D (**3**), show no cytotoxicity. These results showed that the combination of left and right segments is essential for the cytotoxicity of the phelligridins. In addition, we evaluated cytotoxicity against human cervical cancer cells. In cytotoxicity

Scheme 3 Total synthesis of phelligridin D (**3**).
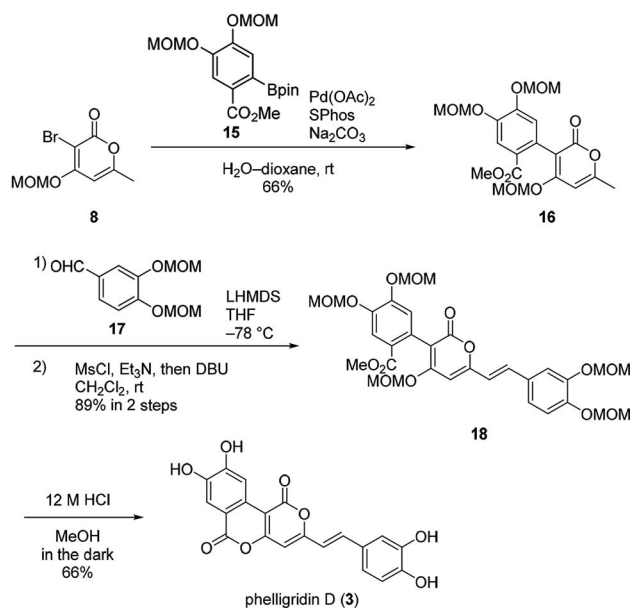
Scheme 4 Concise total synthesis of phelligridin D (**3**).
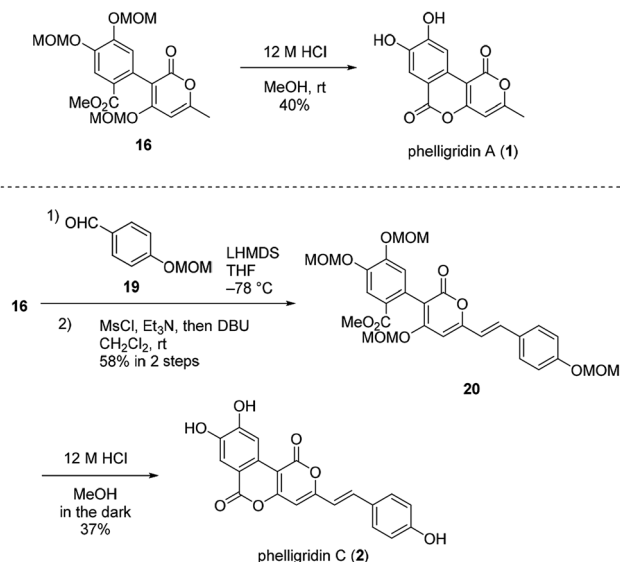
Scheme 5 Total syntheses of phelligridins A (**1**), C (**2**).

Table 1 Cytotoxicity of phelligridins A, C, D, and hispidin

| Cpd | A549 cells[a] (μM) | | | HeLa S3 cells (μM) |
| | Syn. | Nat. | | Syn. |
	IC$_{50}$	IC$_{50}$	GI$_{50}$	IC$_{50}$
1	>100	>0.192[b]	—	>100
2	1.6	0.012[b]	15.0[c]	3.0
3	1.1	0.016[b]	22.6[c]	2.1
4	>100	—	—	>100

[a] See ESI. [b] Ref. 2. [c] Ref. 4.

against HeLa S3 cells, the same tendencies were observed as well.

Conclusions

We have established a concise and scalable synthetic pathway for phelligridins A (**1**), C (**2**), and D (**3**). The synthetic highlights were Suzuki–Miyaura coupling and aldol-type condensation of α-pyrone. Phelligridin A was synthesized in four steps, while phelligridins C and D were each synthesized in six steps. Furthermore, we have investigated their cytotoxicity and revealed that the whole structure is essential for the cytotoxicity of phelligridins. A structure–activity relationships study based on this synthetic route is ongoing in our laboratory.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

Notes and references

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11 Very recently, we got the personal communication from Shi that the unit of cytotoxicity was wrong in ref. 2 and the values of cytotoxicity should be given in μg mL^{-1}. The revised values of cytotoxicity against A549 cells are as follows: **2**, 4.4 μM (0.012 μg mL^{-1}); **3**, 6.1 μM (0.016 μg mL^{-1}). Our results correspond to the revised values.