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Seven new cytotoxic phenylspirodrimane derivatives from the endophytic fungus *Stachybotrys chartarum*[†]

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Seven undescribed phenylspirodrimane derivatives, stachybochartins A–G (1–7), and four known analogues (8–11) were isolated from the endophytic fungus *Stachybotrys chartarum* obtained from *Pinellia ternata*. Stachybochartins A–D are four rare C–C-coupled dimeric derivatives and stachybochartin G features a seco-bisabosqual skeleton. Their structures and configurations were elucidated *via* spectroscopic analysis, electronic circular dichroism (ECD) calculations, the ECD exciton chirality method and the modified Mosher's method. Stachybochartins A–D and G displayed cytotoxic activities against MDA-MB-231 breast cancer cells and U-2OS osteosarcoma cells, with IC₅₀ values ranging from 4.5 to 21.7 μM. Stachybochartins C and G exerted strong anti-proliferative activities against U-2OS cells in concentration- and time-dependent manners and induced apoptosis.

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1. Introduction

Phenylspirodrimanes are a group of meroterpenoids in which a benzene ring attaches to a drimane sesquiterpene, forming a spirofuran structure. They are particular metabolites that have only been discovered from two fungal genera, *Stachybotrys* and *Memnoniella*.¹ Phenylspirodrimane members have been reported to have diverse biological activities, such as anti-osteoporosis activity,² the ability to inhibit immune-complex disease^{3,4} and tyrosine kinase receptors,⁵ and anti-hyperlipidemic effects.⁶ Due to their unique structures and potential medical applications, chemists are also interested in the synthesis of phenylspirodrimanes and have developed different processes to totally synthesize several bioactive examples, like K-76, L-671, and corallidictyals A–D.^{7–9} It is worth noting that a few derivatives of phenylspirodrimanes have been found as dimers. The two monomers in dimers such as stachybochartins A–C^{10,11} are normally linked through different alkyl diamine chains, which are presumably derived from different amino acids. Chartarlactam L was the first dimer reported with

a C_{8'}–C_{8'''} linkage^{6,12} and, thereafter, two dimers, stachartone A and stachartarin A,^{13,14} were reported to unusually dimerize through the formation of a 2-phenyl-1-indanone fragment. The dimers were also found to possess potential biological activities that were different to those of the monomers, such as antibacterial activities, HIV-1 protease inhibitory effects, and potential ET receptor antagonists.^{10,11,15}

Recently, during metabolite research into endophytic fungi from *Pinellia ternata*,^{16–19} chemical investigation of the strain *Stachybotrys chartarum* PT2-12 led to the discovery of four phenylspirodrimane C–C-coupled dimers (1–4) and three new monomers (5–7), along with four known examples (8–11) (Fig. 1). Differently to the C_{8'}–C_{8'''} linkage in chartarlactam L,⁶ the two monomers in 1 and 2 were dimerized through the unusual C_{7'}–C_{7'''} linkage. The isolates were assayed for their cytotoxic activities against three human cancer cell lines (MDA-MB-231, U-2OS and MCF-7). Stachybochartins A–D and G showed potent cytotoxicity against MDA-MB-231 and U-2OS cells and stachybochartins C and G were found to exert significant anti-proliferative activities against U-2OS cells through the activation of caspase-dependent apoptosis. Herein, the isolation, structural elucidation, and cytotoxic activities of these stachybochartins are described.

2. Results and discussion

The molecular formula of stachybochartin A (1) was determined as C₄₆H₆₂O₁₀, from the positive HRESIMS ion peak at *m/z*: 797.4232 [M + Na]⁺ (calcd for C₄₆H₆₂NaO₁₀: 797.4235), revealing 16 degrees of unsaturation. In the ¹H NMR spectrum, the obviously recognized signals were one aromatic

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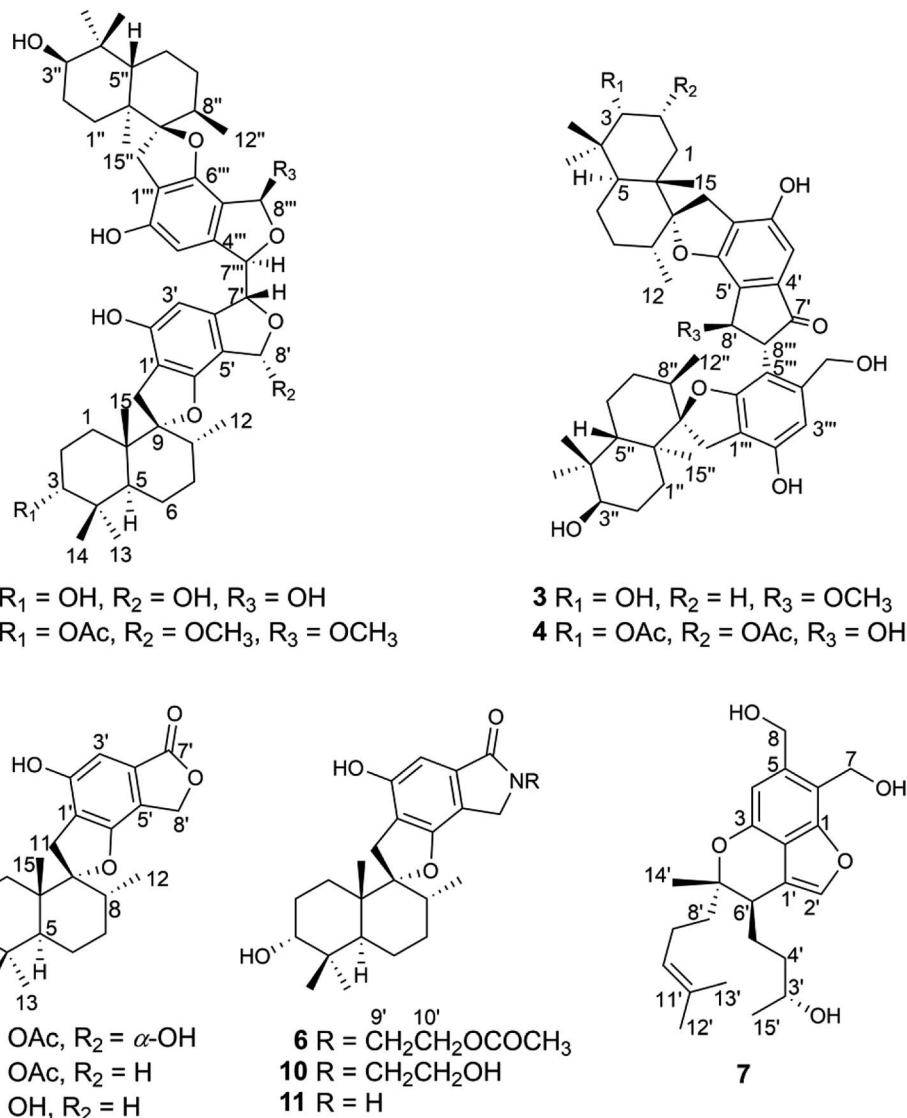


Fig. 1 The structures of compounds 1–11.

singlet (δ_{H} 6.31, s), three oxymethine signals (δ_{H} 5.70, s; δ_{H} 4.72, s; δ_{H} 3.33, m), and four methyl signals (δ_{H} 1.03, 3H, s; δ_{H} 0.97, 3H, s; δ_{H} 0.88, 3H, s; δ_{H} 0.76, 3H, d, $J = 6.4$ Hz), while the remaining signals were from twelve protons in the field range from δ_{H} 2.00 to 3.00. Analysis of ^{13}C NMR and DEPT spectra revealed the presence of 23 carbon signals from four methyl (δ_{C} 29.0, 23.0, 16.6, and 16.0), five methylene (δ_{C} 32.6, 32.5, 26.1, 25.4 and 22.3), six methine (δ_{C} 108.7, 97.3, 76.5, 64.1, 41.5 and 38.5), five nonprotonated aromatic (δ_{C} 160.7, 154.8, 134.2, 114.7, and 109.6), and three quaternary (δ_{C} 98.7, 43.2 and 37.4) carbons. The above data resembled that from stachybotrylactones, yet only showed half the total mass revealed by MS studies.²⁰ Therefore, **1** was deduced to be a dimeric phenylspirodrimane with a symmetrical structure. HMBC results further confirmed the phenylspirodrimane structure (Fig. 2). The correlations from $\text{H}_3\text{-15}$ to H-1/H-5/H-9/H-10, from $\text{H}_3\text{-14}$ to H-3/H-4, and from H-3 to H-1/H-5 verified the drimane sesquiterpene moiety. Meanwhile, the spirobenzofuran moiety

was revealed through correlations from $\text{H}_2\text{-11}$ to C-1', C-2', C-6', C-9, and C-10. It was further noticed that the carbonyl C-7' and the oxygenated methylene C-8' seen in stachybotrylactones²⁰ were replaced by the hemiacetal C-8' at δ_{H} 5.70/ δ_{C} 97.3 and the oxygenated methine C-7' at δ_{H} 4.72/ δ_{C} 64.1 in **1**. The methine nature of C-7' revealed the C-7'–C-7''' bond between the two symmetrical units. This was also evidenced by the HMBC correlations from H-7' (δ_{H} 4.72) to C-7', C-3', C-4', and C-8', as well as from H-8' (δ_{H} 5.70) to C-4', C-5', C-6', and C-7'. The monomeric core in **1** was deduced to possess the same relative configuration as those of stachybotrylactones from ROESY analysis (Fig. 2). The key ROESY correlations of H-3/ $\text{H}_3\text{-14}$ / $\text{H}_3\text{-15}$, $\text{H}_3\text{-15}$ /H-11a/H-8, H-11b/ $\text{H}_3\text{-12}$, and H-5/ $\text{H}_3\text{-13}$ confirmed the β -orientations of H-3, $\text{H}_3\text{-14}$, $\text{H}_3\text{-15}$, H₂-11 and H-8, while H-5, $\text{H}_3\text{-12}$, and $\text{H}_3\text{-13}$ were α -oriented. The ROESY correlations from H-8' to $\text{H}_3\text{-12}$ and H-7' further revealed that H-7' and H-8' were on the same side of the molecular plane. Based on the ECD exciton chirality method,^{21,22} the (R)-



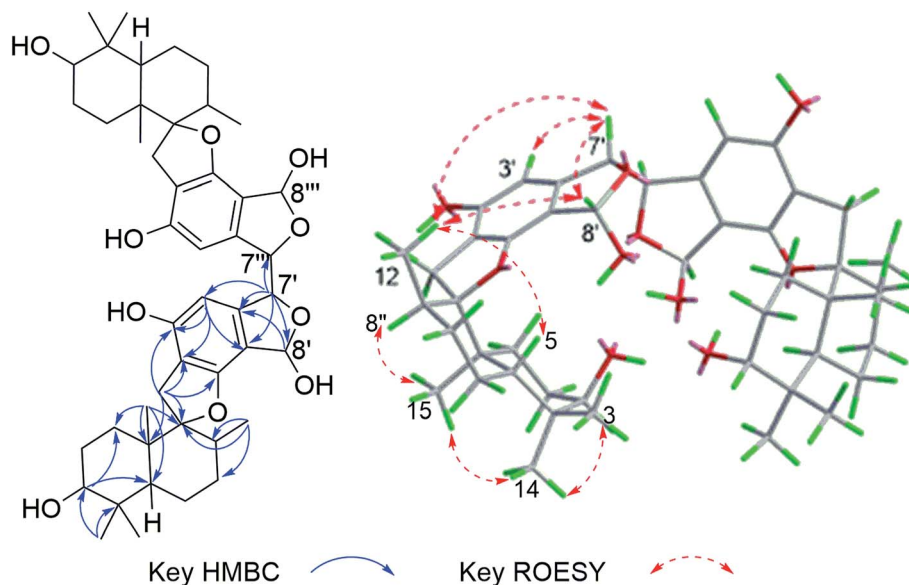


Fig. 2 The key HMBC and ROESY correlations for stachybochartin A (1).

configuration of C-7'/C-7''' was revealed through the negative couplet of the benzene moiety at around 210 nm (Fig. 3a), which was further confirmed *via* ECD calculations (Fig. 3b).

Stachybochartin B (2) was determined to have the molecular formula $C_{50}H_{68}O_{11}$, from the HRESIMS ion peak at m/z : 867.4658 $[M + Na]^+$. The 1D NMR spectra signals of 2 were apparently double those of 1, which indicated that 2 was an analogue of 1 with different substituents. The methylation of the hydroxyl groups at C-8' and C-8''' and the acetylation of the hydroxyl group at C-3 in 2 were confirmed *via* the HMBC correlations from δ_H 3.66 ($-OCH_3$) to C-8', from δ_H 3.67 ($-OCH_3$) to C-8''', from H-8' to δ_C 56.2 ($-OCH_3$), from H-8''' to δ_C 56.4 ($-OCH_3$), and from H-3 (δ_H 4.58)/ δ_H 2.00 ($-OCOCH_3$) to δ_C 172.5 ($-OCOCH_3$). The relative configurations of the two monomeric units were determined to be identical to those in 1 *via* ROESY correlations (Fig. S1[†]). In addition, the well matched ECD curves of 1 and 2 (Fig. S2[†]) established that

their absolute configurations are the same. The dimeric compounds of 1 and 2 were speculated to be derived from the monomer stachybotrylactone (9), where two monomers formed a C-C bond *via* a free radical coupling reaction at C-7' and C-7'''.

Stachybochartin C (3) was obtained as a yellow amorphous solid. Its molecular formula $C_{47}H_{64}O_9$ with 16 degrees of unsaturation was determined *via* the positive HRESIMS ion peak at m/z : 795.4447 $[M + Na]^+$ (calcd for $C_{47}H_{64}NaO_9$: 795.4443). The 1D and 2D NMR data indicated that 3 was another dimer but with a different dimeric mode. Careful analysis of the 1H - 1H COSY correlations revealed two spin systems (C-1-C-3 and C-5-C-12), which further assisted the elucidation of the drimane moieties through the HMBC correlations from H₃-15 to H-1/H-5/H-9/H-10, H₃-14 to H-3/H-4, and H-3 to H-1/H-5. The correlations from H₂-11 to C-1', C-2', C-6', C-9, and C-10 further confirmed that the attached benzene ring

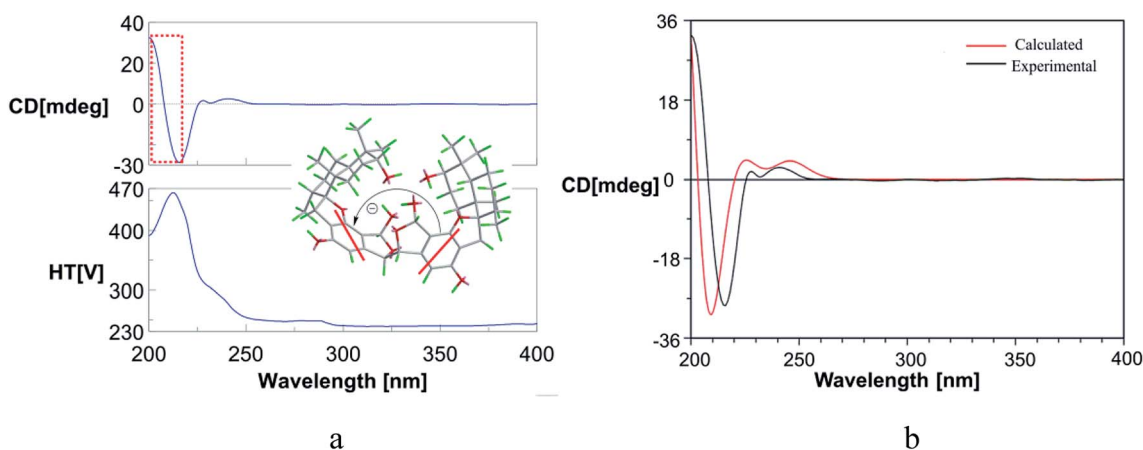


Fig. 3 (a) The ECD spectrum of stachybochartin A (1) in MeOH. The negative couplet at around 208 nm (a negative Cotton effect at 214 nm and a positive one at 202 nm) in the ECD spectrum indicated the 7'*R*, 7'''*R* configuration in 1; (b) a comparison of the experimental and calculated ECD spectra of 1.



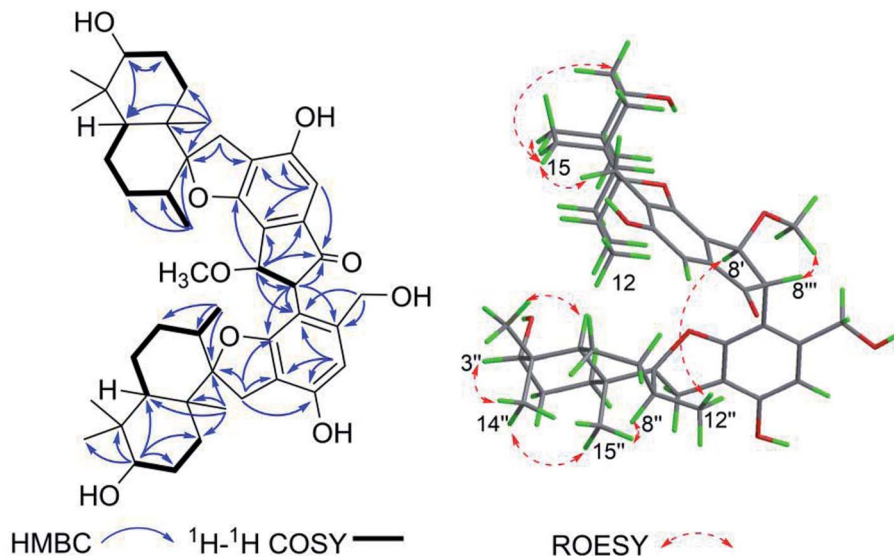


Fig. 4 The key HMBC and ROESY correlations for stachybochartin C (3).

helped form the spirofuran structure. Similar HMBC correlations also established the other monomeric unit. The 2-phenyl-1-indanone fragment, as in stachartone A,^{13,14} was mainly determined *via* HMBC correlations from H-8' (δ_{H} 4.89) to C-4', C-5', C-6', C-7' and C-8''' and H-8''' to C-8', C-7', C-4'', C-5''' and C-6'''. Meanwhile, an additional methoxyl signal was also observed at C-8' in 3 due to HMBC correlation from δ_{H} 3.72 (3H, s, -OCH₃) to C-8' (δ_{C} 85.3). Therefore, the planar structure of 3 was determined as shown. Though ROESY analysis (Fig. 4), the relative configuration of 3 was established to be the same as that of stachartone A.¹³ The ROESY correlations from H-3 to H₃-14/H₃-15 and from H₃-15 to H₂-11 and H-8 revealed that H-3, H₃-14, H₃-15, H₂-11 and H-8 had the same orientation, leaving H-5, H₃-12 and H₃-13 with the opposite orientation. The same NOEs were also observed for the other drimane moiety, which therefore possessed the same relative configuration. Moreover, the correlations between H-8''' and 8'-OCH₃ and between H-8' and H₃-12'' confirmed that H-8' and H-8''' were oriented oppositely and that the two drimane moieties were closely approximated in space. ECD calculations were further used to verify the configuration. As shown in Fig. 5, the 3a calculated spectrum showed overall agreement with the experimental ECD spectrum, which determined the 3*R*, 5*S*, 8*R*, 9*R*, 10*S*, 8'*R*, 3''*R*, 5''*S*, 8''*R*, 9''*R*, 10''*S* and 8'''*R* configuration of 3.

The molecular formula C₅₀H₆₆O₁₂ of stachybochartin D (4) was determined from HRESIMS data. 1D and 2D NMR data analysis revealed that 4 possessed the same dimeric phenylspirodrimane skeleton as 3, but with different substituents. Two additional acetoxy groups were placed at C-2 and C-3, as established *via* the HMBC correlations from δ_{H} 1.88 (3H, s) to δ_{C} 172.3 and from δ_{H} 2.09 (3H, s) to δ_{C} 173.2. Meanwhile, the lack of a methoxyl signal and the chemical shifts of H-8' (δ_{H} 5.18) and C-8' (δ_{C} 75.7) revealed a hydroxyl group at C-8'. The relative configuration of 4 was deduced to be the same as that of 3 due to their similar ROESY correlations (Fig. S3†), and H-2 in 4 had the same orientation as H-3 and H₃-15 according to their

observed mutual NOEs. The identical ECD curves of 3 and 4 (Fig. S4†) additionally indicated that their absolute configurations were the same.

The molecular formula of stachybochartin E (5), C₂₅H₃₂O₇, was determined from the HRESIMS ion peak at *m/z*: 467.2038 [M + Na]⁺ (calcd for C₂₅H₃₂NaO₇: 467.2040). Its structure was determined to be similar to that of stachybotrylactone acetate (8)²³ on the basis of their similar NMR data. The difference between them was the appearance of a hydroxyl group at C-2 (δ_{H} 4.07, ddd, *J* = 12.2, 4.3, 2.6 Hz; δ_{C} 65.7) in 5, which was also evidenced by the HMBC correlations from H-2 to C-1 and C-3. The small coupling constant of *J*_{H-3/H-2} (*J* = 2.6 Hz) indicated

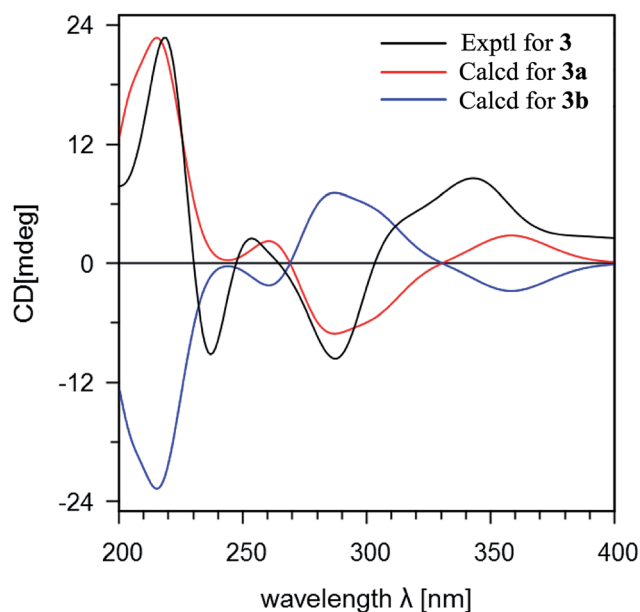


Fig. 5 A comparison of the experimental and calculated ECD spectra of stachybochartin C (3). (3a: 3*R*, 5*S*, 8*R*, 9*R*, 10*S*, 8'*R*, 3''*R*, 5''*S*, 8''*R*, 9''*R*, 10''*S*, 8'''*R*; 3b: 3*S*, 5*R*, 8*S*, 9*S*, 10*R*, 8'*S*, 3''*S*, 5''*R*, 8''*S*, 9''*S*, 10''*R*, 8'''*S*).



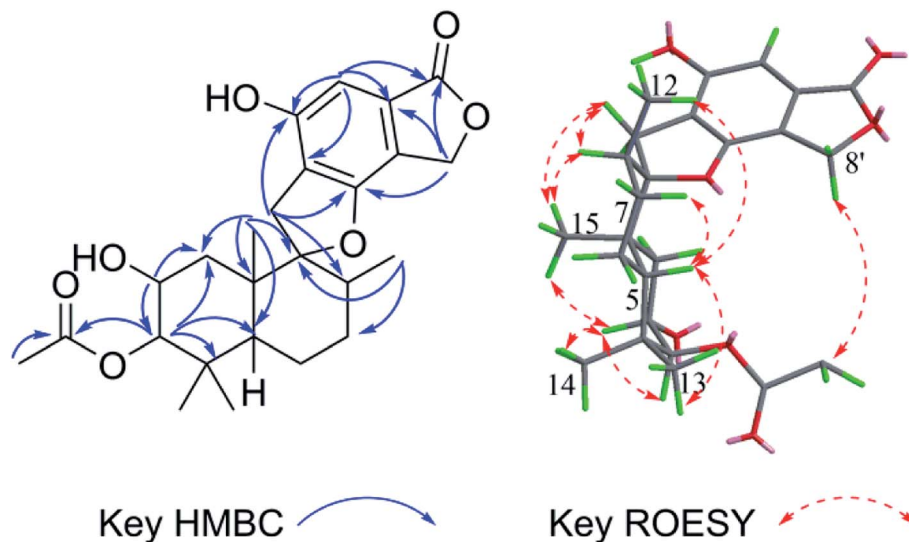


Fig. 6 The key HMBC and ROESY correlations for stachybochartin E (5).

that the orientation of H-2 and H-3 was the same, and the ROESY correlations of H-2/H₃-14/H₃-15/H-3, H₃-15/H-8 and H-5/H₃-13 further suggested the β -orientation of H₃-15, H-8, H₂-11, H-3 and H-2 and the α -orientation of H-5 and H₃-12 (Fig. 6). In the ECD spectrum, the distinctive positive Cotton effect (CE) at 219 nm and negative CEs at 268 and 339 nm (Fig. S5[†]) provided evidence for an absolute configuration of 2*R*, 3*S*, 5*S*, 8*R*, 9*R* and 10*S*.²⁴

Stachybochartin F (6) was assigned the molecular formula of C₂₇H₃₈NO₆ from positive HRESIMS data. The ¹H and ¹³C NMR spectra of 6 were almost identical to those of stachybotramide (10).²³ The differences were the additional acetyl group and the upfield-shifted H₂-10' of 6, suggesting that 6 was the 10'-OH acetylated derivative of stachybotramide. This deduction was proved through the HMBC correlation from H₂-10' (δ_{H} 4.32) to the carbonyl carbon at δ_{C} 172.7. Moreover, the ROESY correlations (Fig. S6[†]) suggested that it had the same relative configuration as that of 5, and their similar CEs (Fig. S7[†]) confirmed their identical absolute configurations.

Stachybochartin G (7) had the molecular formula C₂₃H₃₂O₅. Analysis of the NMR data clearly indicated that 7 was closely related to stachybisbin B,²⁵ and the absence of an acetyl group indicated the deacetylation of 7-OH in 7. The planar structure of 7 was confirmed *via* HMBC correlations (Fig. S8[†]). The ROESY correlations of H-6'/H₂-8' and H₃-14'/H₂-5' indicated that H₃-14' and H-6' had opposite orientation, and the identical ECD curve (Fig. S9[†]) to that of stachybisbin B determined the same 6'*R* and 7'*S* configurations in 7. In addition, the 3'*S* configuration was determined *via* the modified Mosher's method (Fig. 7).¹⁸ Therefore, the absolute configuration of 7 was determined to be 3'*S*, 6'*R*, and 7'*S*.

All the isolates were evaluated for their cytotoxicity against three human cancer cell lines: MDA-MB-231, MCF-7, and U-2OS. Stachybochartins A–D (1–4) and G (7) displayed activity against the human cancer cell lines MDA-MB-231 and U-2OS, with IC₅₀ values ranging from 4.5 to 21.7 μM , but all the isolates were inactive (IC₅₀ > 50 μM) against the cell line MCF-7 (Table S1[†]).

Stachybochartins C (3) and G (7) showed a more remarkable effect on U-2OS cells than on other cell types, thus their cytotoxic effects were further evaluated on U-2OS cells with doxorubicin (DOX) as a positive control (Fig. 8A). As shown in Fig. 8B, both 3 and 7 significantly inhibited the cell viability in concentration- and time-dependent manners, which was also confirmed by the results of colony formation assays, as shown in Fig. 8C. Together, these data indicated that 3 and 7 inhibited the proliferation of U-2OS cells.

To investigate whether the cytotoxicity was associated with cell apoptosis, we respectively detected the apoptosis of U-2OS cells after treatment with stachybochartins C (3) and G (7). Compared with the control, the percentage of apoptotic cells increased in a dose-dependent manner for both the stachybochartin C and G groups (Fig. 9A), indicating

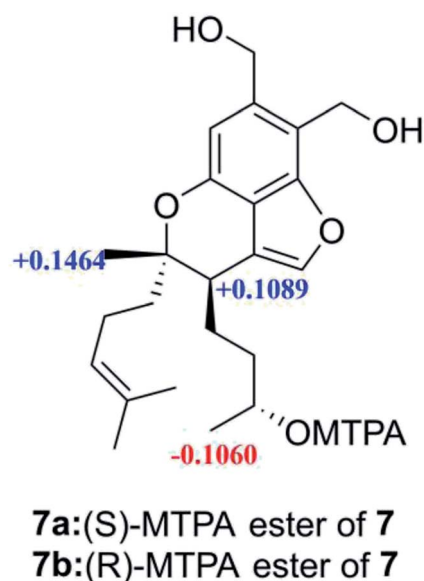


Fig. 7 $\Delta\delta$ values (in ppm) = $\delta_{\text{S}} - \delta_{\text{R}}$, obtained for the (S)- and (R)-MPTA esters.



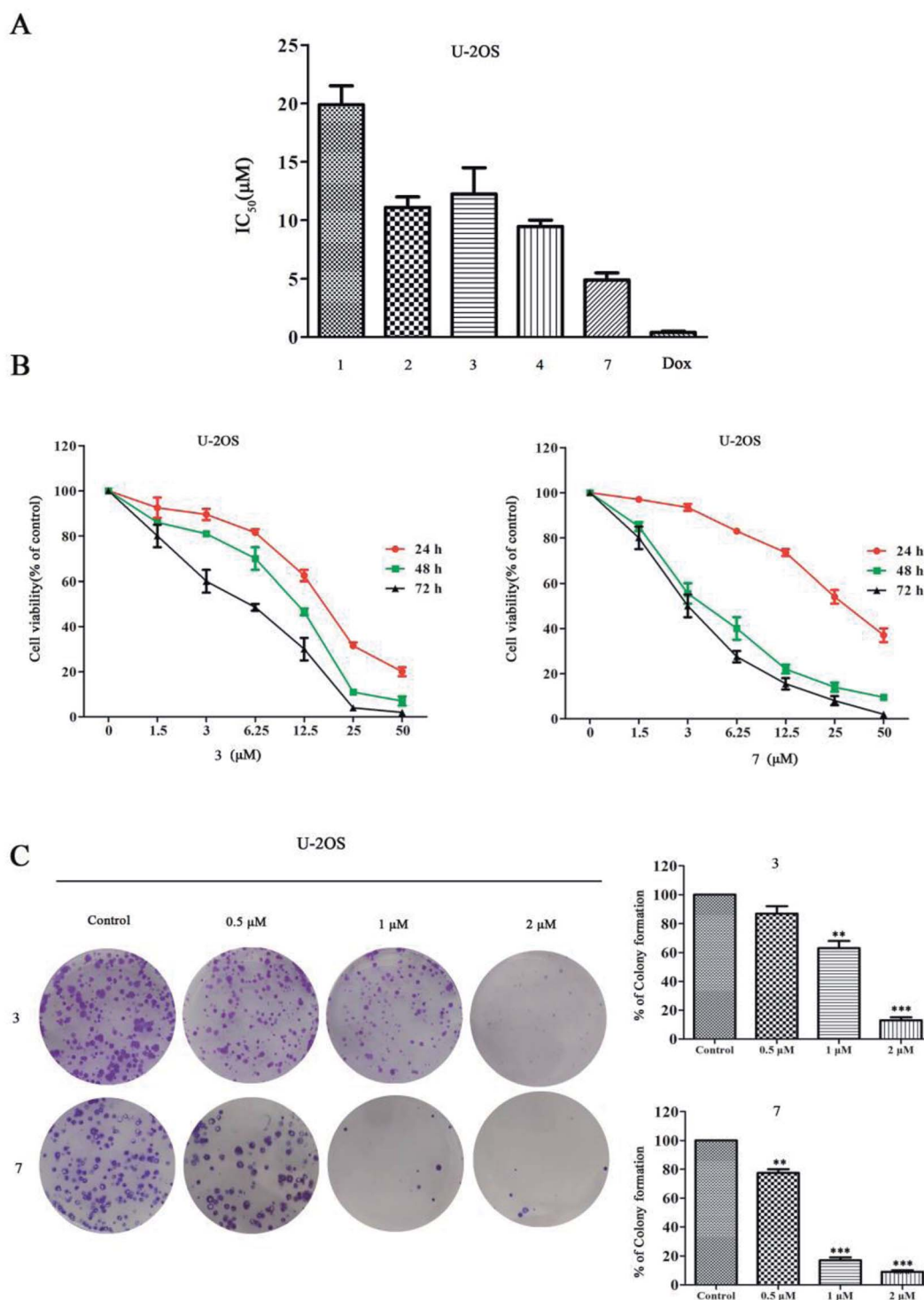


Fig. 8 The effects of stachybochartins A–D (1–4) and G (7) on cell viability in U-2OS cells and on inhibiting U-2OS cell proliferation. (A) U-2OS cells were treated with 1–4, 7 and DOX (positive control) for 48 h, and the IC₅₀ values were quantified using MTT assays. (B) U-2OS cells were treated with different concentrations of 3 and 7 (0, 1.5, 3, 6.25, 12.5, 25 or 50 μM) for the indicated times, and cell viability was determined via MTT assays. (C) The effects of 3 and 7 on the clonogenic ability of U-2OS cells treated for 48 h are shown. The data are presented as mean ± SEM from three independent experiments. *: $P < 0.05$, **: $P < 0.01$ and ***: $P < 0.001$, compared to the control.

apoptosis in the cells. The caspase-cascade system plays a crucial role in cell apoptosis, and caspases are the executors of apoptosis.²⁶ Therefore, we examined the expression of caspase-related proteins in stachybochartin G-treated U-2OS

cells. Significant caspase-8, caspase-9 and PARP processing were observed in the total cell lysates, and their cleavages markedly increased in a dose-dependent manner (Fig. 9B). Thus, these data suggest that the cytotoxic effects of



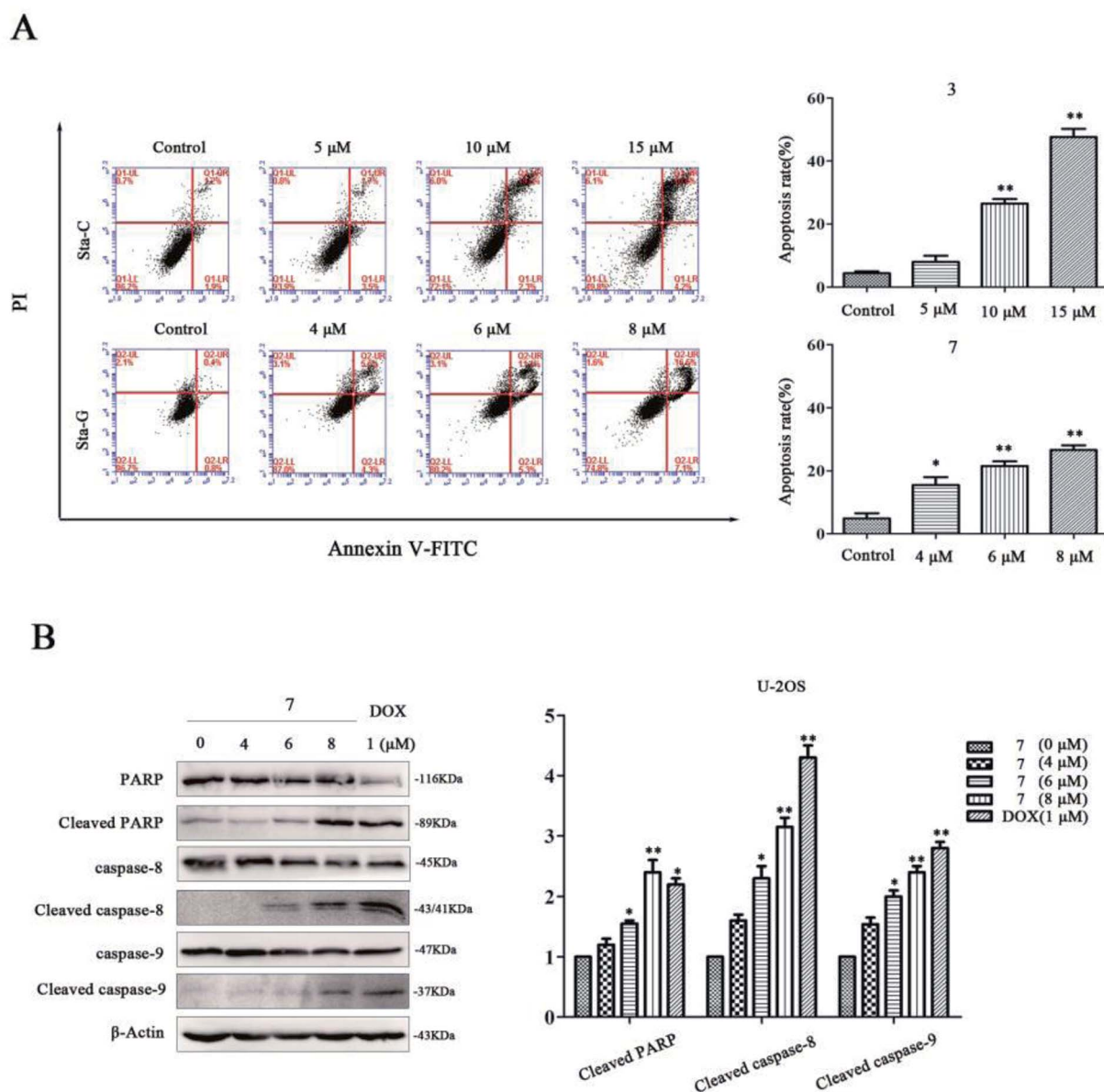


Fig. 9 Stachybochartins C (3) and G (7) induced caspase-dependent apoptosis in U-2OS cells. (A) U-2OS cells were pretreated with 3 (0–15 μM) or 7 (0–8 μM) for 48 h and apoptosis rates were analyzed *via* flow cytometry. (B) After treatment with 7 (0–8 μM) and DOX (1 μM, as a positive control) for 48 h, the expression levels of PARP and cleaved PARP, pro-caspase-8, cleaved caspase-8, pro-caspase-9, and cleaved caspase-9, were analyzed *via* western blotting. β-Actin was used as a protein loading control. The data are presented as mean ± SEM from three independent experiments. *: $P < 0.05$ and **: $P < 0.01$, compared to the control.

stachybochartin G on U-2OS cells were caused by the activation of caspase-dependent apoptosis.

3. Conclusions

In conclusion, seven phenylspirodrimane derivatives, stachybochartins A–G (1–7), together with four known analogues were isolated from the endophytic fungus *Stachybotrys chartarum*. The four dimers are structurally infrequent, especially stachybochartins A and B with their C7'–C7''' linkages. The dimers, along with stachybochartin G, displayed cytotoxic activities against human MDA-MB-231 breast cancer cells and U-2OS osteosarcoma

cancer cells, and the activation of caspase-dependent apoptosis was the possible pathway for their anticancer activities. In addition, stachybochartin G, which features a seco-bisabosqual skeleton, showed stronger activity than normal phenylspirodrimanes. These findings may provide evidence for further research into the anticancer activities of these compounds.

4. Experimental section

4.1. General experimental procedures

Optical rotations were measured with a Jasco P-1020 polarimeter (Jasco, Tokyo, Japan). UV spectra were acquired with



a Shimadzu UV-2450 spectrophotometer (Shimadzu, Tokyo, Japan). CD spectra were recorded using a Jasco J-810 spectrometer (Jasco, Tokyo, Japan). IR spectra (KBr disks, in cm^{-1}) were obtained using a Bruker Tensor 27 spectrometer (Bruker, Karlsruhe, Germany). NMR spectra (^1H : 500 MHz; ^{13}C : 125 MHz) were recorded at 300 K with a Bruker Avance III NMR instrument (Bruker, Karlsruhe, Germany), with TMS as the internal standard. CD_3OD ($\delta_{\text{H}} = 3.31$ and $\delta_{\text{C}} = 49.15$) was used as the solvent. High-resolution electrospray ionization mass spectra (HRESIMS) data were obtained using an Agilent 6520B UPLC-Q-TOF instrument (Agilent Technologies, Santa Clara, CA, USA). Column chromatography (CC) was carried out using silica gel (Qingdao Marine Chemical Co. Ltd., Qingdao, China), Sephadex LH-20 (Pharmacia, Stockholm, Sweden), and RP-C18 silica (40–63 μm , Fuji, Japan). The fractions obtained from column chromatography were studied *via* TLC with pre-coated silica gel GF254 (Qingdao Haiyang Chemical Co., Ltd., China) plates. Analytical HPLC was performed using an Agilent 1200 Series instrument with a DAD detector, using a Shim-pack VPODS column (5 μm , 4.6×250 mm; Agilent Technologies, Palo Alto, CA, USA). Preparative HPLC was carried out using Shimadzu LC-10A apparatus equipped with a Shim-pack RP-C18 column (10 μm , 20×200 mm, i.d., Shimadzu, Tokyo, Japan) and Shimadzu SPD-20A detector.

4.2. Fungal materials and fermentation

The fungal strain *Stachybotrys chartarum* PT2-12 was isolated from rhizomes of *Pinellia ternata*, which were collected from the suburb of Nanjing, Jiangsu Province, People's Republic of China, in October 2014. It was identified according to morphological studies, which were reinforced by 18S rDNA and internal transcribed spacer (ITS) sequences with 100% identity matching to known *S. chartarum* (GenBank accession No. KC787692.1).

The fungus *S. chartarum* was cultured on potato dextrose agar (PDA) at 28 °C. Seven days later, agar plugs were cut into small pieces (about $0.3 \times 0.3 \times 0.3$ cm^3) under aseptic conditions, 3–4 pieces of which were inoculated in an Erlenmeyer flask (500 mL) containing 200 mL of potato dextrose liquid medium. Then the flasks were incubated on a rotary shaker at 28 °C and 160 rpm for 7 days to prepare the seed culture. Fermentation was carried out in 30 Erlenmeyer flasks (2 L), each containing 320 g of rice and 400 mL of distilled H_2O ; these were cultivated at 28 °C for 45 days. The solid culture medium was soaked overnight and sterilized at 121 °C for 20 min before use.

4.3. Extraction and isolation

The solid rice material was extracted with an equal volume of ethyl acetate (EtOAc) three times at room temperature, and the

Table 1 ^1H (500 MHz) and ^{13}C (125 MHz) NMR data for **1** and **2** in CD_3OD

1			2 unit A			2 unit B		
Position	δ_{H} , mult. (<i>J</i> in Hz)	δ_{C}	Position	δ_{H} , mult. (<i>J</i> in Hz)	δ_{C}	Position	δ_{H} , mult. (<i>J</i> in Hz)	δ_{C}
1/1''	1.82, m	25.4	1	1.93, m	25.9	1''	1.82, m	25.4
	1.05, m			1.12, m			1.09, m	
2/2''	1.96, m	26.1	2	1.90, m	23.3	2''	1.92, m	26.1
	1.52, m			1.53, m			1.52, m	
3/3''	3.33, m	76.5	3	4.58, t (2.4)	79.7	3''	3.32, m	76.4
4/4''		38.5	4		37.8	4''		38.5
5/5''	2.18, dd (12.4, 2.0)	41.5	5	2.18, dd (11.4, 2.3)	42.5	5''	2.23, dd (11.9, 1.7)	41.5
6/6''	1.58, m	22.3	6	1.53, m	22.0	6''	1.53, m	22.0
	1.50, m			1.60, m			1.60, m	
7/7''	1.58, m	32.6	7	1.56, m	32.6	7''	1.56, m	32.6
	1.49, m			1.62, m			1.62, m	
8/8''	1.82, m	38.5	8	1.82, m	38.1	8''	1.83, m	38.8
9/9''		98.7	9		98.6	9''		98.6
10/10''		43.5	10		43.5	10''		98.8
11/11''	3.17, d (16.3)	32.5	11	3.17, d (16.3)	32.4	11''	3.15, d (16.4)	32.6
	2.78, d (16.3)			2.83, d (16.3)			2.79, d (16.4)	
12/12''	0.76, d (6.5)	16.0	12	0.80, d (6.5)	15.9	12''	0.76, d (6.5)	16.0
13/13''	0.97, s	29.0	13	0.96, s	28.5	13''	0.97, s	29.0
14/14''	0.88, s	23.0	14	0.91, s	22.2	14''	0.88, s	23.0
15/15''	1.03, s	16.6	15	1.06, s	16.3	15''	1.03, s	16.7
1'/1'''		114.7	1'		114.6	1'''		114.7
2'/2'''		154.8	2'		155.2	2'''		154.9
3'/3'''	6.31, s	108.7	3'	6.35, s	108.8	3'''	6.32, s	108.7
4'/4'''		134.2	4'		133.9	4'''		134.3
5'/5'''		109.6	5'		109.2	5'''		109.5
6'/6'''		160.7	6'		160.7	6'''		160.5
7'/7'''	4.72, s	64.1	7'	4.75, d (2.3)	64.1	7'''	4.76, d (2.3)	64.4
8'/8'''	5.7, s	97.3	8'	5.71, s	97.4	8'''	5.57, s	97.8
			8'-OCH ₃	3.66, s	56.2	8'''-OCH ₃	3.67, s	56.4
			3-OAc	2.00, s	21.3			
					172.5			



organic solvent was evaporated to dryness under vacuum to afford the crude extract (213.0 g). The EtOAc extract was fractionated *via* silica gel CC using PE (petroleum ether)–EtOAc (v/v: 80 : 1, 40 : 1, 20 : 1, 10 : 1, 5 : 1, 2 : 1, 1 : 1 and 0 : 1) gradient elution to give eight fractions (Fr.1–Fr.8). Fr.7 (1 : 1, 789 mg) was subjected to Sephadex LH-20 CC with CH₂Cl₂–MeOH (1 : 1) as the eluent, giving nine subfractions (Fr.7.1–Fr.7.9) after combination according to TLC. Fr.7.7 (64.7 mg) was separated *via* RP-HPLC, eluted with CH₃CN–H₂O (42 : 58) at a flow rate of 10.0 mL min⁻¹, to afford **5** (1.7 mg; *t*_R = 23.6 min), **8** (3.2 mg; *t*_R = 33.5 min), and **9** (45.4 mg; *t*_R = 44.8 min). Fr.8 (0 : 1; 21.4 g) was repeatedly subjected to chromatography using an ODS column, eluting with gradient MeOH–H₂O (v/v: 20 : 80, 30 : 70, 40 : 60, 50 : 50, 60 : 40, 80 : 20 and 100 : 0), giving seven subfractions (Fr.8.1–Fr.8.7) after combination according to TLC. The subfraction Fr.8.4 was purified *via* preparative HPLC with CH₃CN–H₂O (40 : 60), to give **7** (5.8 mg; *t*_R = 32.3 min). Fr.8.5 was purified *via* preparative HPLC with CH₃CN–H₂O (50 : 50), to give **6** (4.8 mg; *t*_R = 18.8 min), **10** (17.6 mg, *t*_R = 26.5 min), and **11** (31.0 mg; *t*_R = 39.9 min). Fr.8.6 (1.0 g) was further separated

via medium pressure preparative HPLC (ODS MPLC) with MeOH–H₂O (85 : 15), to give eight fractions (Fr.8.6.1–Fr.8.6.8). Fr.8.6.3 (143.1 mg) was purified *via* preparative HPLC with CH₃CN–H₂O (70 : 30) to give **3** (39.1 mg; *t*_R = 13.1 min) and **2** (9.6 mg; *t*_R = 34.1 min). Fr.8.6.4 (116.3 mg) was purified *via* preparative HPLC with CH₃CN–H₂O (70 : 30) to give **1** (15.9 mg; *t*_R = 22.8 min). Fr.8.6.7 (89.9 mg) was purified *via* preparative HPLC with CH₃CN–H₂O (85 : 15) to give **4** (17.2 mg, *t*_R = 12.9 min).

Stachybochartin A (1). Yellow powder; [α]_D²⁵: –33.5 (*c* 0.12, MeOH); UV (MeOH) λ_{\max} (log ϵ): 213 (4.27), 288 (3.25) nm; IR (KBr) ν_{\max} : 3429, 2928, 1629, 1458, 1358, 1261, 1123, 1083, 1038, 960 cm⁻¹; CD (MeOH) λ_{\max} ($\Delta\epsilon$): 202 (+31.9), 214 (–48.1), and 242 (+3.2) nm; ¹H and ¹³C NMR data: see Table 1; HRESIMS *m/z*: 797.4232 [M + Na]⁺ (calcd for C₄₆H₆₂NaO₁₀: 797.4235).

Stachybochartin B (2). Yellow powder; [α]_D²⁵: –63.6 (*c* 0.14, MeOH); UV (MeOH) λ_{\max} (log ϵ): 213 (4.82), 288 (3.66) nm; IR (KBr) ν_{\max} : 3431, 2925, 1713, 1630, 1457, 1382, 1263, 1124, 1083, 1038, 962 cm⁻¹; CD (MeOH) λ_{\max} ($\Delta\epsilon$): 202 (+21.3), 214 (–33.6), and 241 (+2.8) nm; ¹H and ¹³C NMR data: see Table 1; HRESIMS *m/z*: 867.4658 [M + Na]⁺ (calcd for C₅₀H₆₈NaO₁₁: 867.4654).

Table 2 ¹H (500 MHz) and ¹³C (125 MHz) NMR data for **3** and **4** in CD₃OD

Position	3 unit A		3 unit B		4 unit A		4 unit B				
	δ_{H} , mult. (<i>J</i> in Hz)	δ_{C}	δ_{H} , mult. (<i>J</i> in Hz)	δ_{C}	δ_{H} , mult. (<i>J</i> in Hz)	δ_{C}	δ_{H} , mult. (<i>J</i> in Hz)	δ_{C}			
1	1.97, m 1.14, m	25.5	1''	1.45, m 1.05, m	25.0	1	2.14, t (12.2) 1.43, m	31.7	1''	1.06, m 1.47, m	25.0
2	1.98, m 1.57, m	25.9	2''	2.00, m 1.58, m	25.7	2	5.26, ddd (12.2, 4.0, 2.1)	69.9	2''	1.99, m 1.57, m	25.7
3	3.34, m	76.4	3''	3.22, t (2.4)	77.8	3	4.94, d (2.1)	78.2	3''	3.23, t (2.6)	77.9
4		38.6	4''		38.8	4		39.3	4''		38.8
5	2.19, dd (11.7, 2.9)	41.6	5''	1.17, m	41.2	5	2.26, dd (11.4, 1.8)	41.8	5''	1.21, m	41.3
6	1.58, m 1.52, m	22.1	6''	1.51, m 1.13, m	21.7	6	1.51, m 1.57, m	21.7	6''	1.08, m 1.48, m	21.5
7	1.62, m 1.53, m	32.6	7''	1.58, m 1.20, m	32.6	7	1.63, m 1.55, m	32.0	7''	1.20, m 1.60, m	32.5
8	1.87, m	38.5	8''	1.63, m	37.7	8	1.89, m	38.2	8''	1.63, m	37.8
9		99.9	9''		98.9	9		99.0	9''		98.9
10		43.5	10''		43.4	10		44.8	10''		43.4
11	3.28, d (17.4) 2.88, d (17.4)	33.2	11''	3.08, d (16.5) 2.75, d (16.5)	32.6	11	3.27, d (17.7) 2.92, d (17.7)	33.4	11''	3.08, d (16.7) 2.75, d (16.7)	32.6
12	0.75, d (6.4)	16.4	12''	0.65, d (6.4)	16.2	12	0.78, d (6.5)	16.1	12''	0.63, d (6.4)	16.1
13	0.94, s	29.0	13''	0.84, s	29.2	13	0.93, s	28.3	13''	0.86, s	29.2
14	0.88, s	22.8	14''	0.79, s	23.2	14	1.05, s	22.0	14''	0.80, s	23.2
15	1.07, s	16.6	15''	0.90, s	16.1	15	1.17, s	17.1	15''	0.91, s	16.1
1'		123.0	1'''		113.0	1'		122.7	1'''		113.1
2'		156.4	2'''		153.6	2'		156.2	2'''		153.5
3'	6.76, s	102.4	3'''	6.41, s	108.7	3'	6.78, s	102.2	3'''	6.39, s	108.8
4'		139.6	4'''		142.0	4'		139.5	4'''		142.0
5'		125.0	5'''		110.2	5'		127.6	5'''		110.4
6'		160.0	6'''		161.0	6'		160.1	6'''		161.0
7'		206.4	7'''	4.64, d (12.5) 4.57, d (12.5)	63.5	7'		207.1	7'''	4.61, s	63.7
8'	4.89, d (3.5)	85.3	8'''	4.02, d (3.5)	55.7	8'	5.18, d (3.2)	75.7	8'''	3.92, d (3.2)	59.1
2-OAc						2-OAc	1.88, s	20.9			
								172.3			
3-OAc						3-OAc	2.09, s	20.9			
								173.2			
8'-OCH ₃	3.72, s	59.0									



Stachybochartin C (3). White powder; $[\alpha]_{\text{D}}^{25}$: -38.3 (c 0.13, MeOH); UV (MeOH) λ_{max} ($\log \epsilon$): 214 (4.63), 287 (3.89) nm; IR (KBr) ν_{max} : 3429, 2932, 2877, 1705, 1623, 1459, 1386, 1350, 1255, 1112 cm^{-1} ; CD (MeOH) λ_{max} ($\Delta\epsilon$): 202 (+1.0), 219 (+16.5), 237 (-10.1), 251 (-0.3), 288 (-9.0), and 345 (+3.3) nm; ^1H and ^{13}C NMR data: see Table 2; HRESIMS m/z : 795.4447 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{47}\text{H}_{64}\text{NaO}_9$: 795.4443).

Stachybochartin D (4). White powder; $[\alpha]_{\text{D}}^{25}$: -29.2 (c 0.12, MeOH); UV (MeOH) λ_{max} ($\log \epsilon$): 213 (4.15), 287 (3.43) nm; IR (KBr) ν_{max} : 3431, 2921, 2851, 1713, 1629, 1464, 1385, 1260, 1125, 621 cm^{-1} ; CD (MeOH) λ_{max} ($\Delta\epsilon$): 205 (+1.0), 220 (+21.9), 236 (-19.0), 250 (-1.2), 288 (-15.4), and 344 (+5.4) nm; ^1H and ^{13}C NMR data: see Table 2; HRESIMS m/z : 857.4478 $[\text{M} - \text{H}]^-$ (calcd for $\text{C}_{50}\text{H}_{65}\text{O}_{12}$: 857.4482).

Stachybochartin E (5). Pale yellow powder; $[\alpha]_{\text{D}}^{25}$: -8.2 (c 0.10, MeOH); UV (MeOH) λ_{max} ($\log \epsilon$): 219 (4.09), 268 (3.34), 309

(3.08) nm; IR (KBr) ν_{max} : 3419, 2922, 2851, 1744, 1628, 1469, 1384, 1334, 1261, 1126, 1088, 957, 619 cm^{-1} ; CD (MeOH) λ_{max} ($\Delta\epsilon$): 219 (+3.77), 268 (-1.04), and 339 (-0.28) nm; ^1H and ^{13}C NMR data: see Table 3; HRESIMS m/z : 467.2038 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{25}\text{H}_{32}\text{NaO}_7$: 467.2040).

Stachybochartin F (6). Yellow powder; $[\alpha]_{\text{D}}^{25}$: -27.7 (c 0.14, MeOH); UV (MeOH) λ_{max} ($\log \epsilon$): 218 (4.16), 264 (3.52), 300 (3.14) nm; IR (KBr) ν_{max} : 3422, 2924, 2875, 1742, 1670, 1468, 1387, 1349, 1239, 1142, 1079, 622 cm^{-1} ; CD (MeOH) λ_{max} ($\Delta\epsilon$): 215 (+2.29), 264 (-2.41), and 338 (-0.17) nm; ^1H and ^{13}C NMR data: see Table 3; HRESIMS m/z : 472.2693 $[\text{M} + \text{H}]^+$ (calcd for $\text{C}_{27}\text{H}_{38}\text{NO}_6$: 472.2694).

Stachybochartin G (7). Yellow gum; $[\alpha]_{\text{D}}^{25}$: -27.8 (c 0.10, MeOH); UV (MeOH) λ_{max} ($\log \epsilon$): 220 (4.00), 258 (3.41), 262 (3.42), 282 (3.00) nm; IR (KBr) ν_{max} : 3423, 2921, 2851, 1648, 1384, 1126, 619 cm^{-1} ; CD (MeOH) λ_{max} ($\Delta\epsilon$): 219 (+18.1), 259

Table 3 ^1H (500 MHz) and ^{13}C (125 MHz) NMR data for compounds 5–7 in CD_3OD

Position	5		6		Position	7	
	δ_{H} (multi, J in Hz)	δ_{C}	δ_{H} (multi, J in Hz)	δ_{C}		δ_{H} (multi, J in Hz)	δ_{C}
1	1.73, t (12.2)	34.7	1.83, m	25.4	1		154.2
2	1.40, dd (12.2, 4.3)	65.7	1.09, m	26.0	2		117.7
	4.07, ddd (12.2, 4.3, 2.6)		1.97, m				
3	4.87, d (2.6)	81.4	3.34, t (3.4)	76.4	3		150.2
4		39.2		38.7	4	6.70, s	109.2
5	2.04, m	41.7	2.13, dd (11.6, 2.6)	41.4	5		140.4
6	1.60, m	21.5	1.54, m	22.1	6		116.4
	1.50, m		1.60, m				
7	1.50, m	32.1	1.59, m	32.3	7	4.91, s	56.0
	1.65, m		1.53, m				
8	1.91, m	37.8	1.86, m	38.4	8	4.79, d (1.1)	63.4
9		100.1		99.8	1'		117.8
10		44.8		43.5	2'	7.51, s	140.0
11	3.29, d (17.5)	32.9	3.23, d (17.0)	33.0	3'	3.69, sext (6.1)	68.8
	2.96, d (17.5)		2.85, d (17.0)				
12	0.79, d (6.5)	15.9	0.73, d (6.5)	15.9	4'	1.59, m	38.4
13	0.92, s	28.4	0.98, s	28.9	5'	1.47, m	27.3
						1.99, m	
14	1.00, s	22.1	0.89, s	23.0	6'	1.33, m	42.4
						2.96, dd (10.6, 3.0)	
15	1.13, s	17.3	1.05, s	16.5	7'		85.3
1'		121.6		119.0	8'	1.64, m	39.8
2'		156.7		155.2	9'	1.55, m	23.3
						2.10, m	
3'	6.74, s	103.7	6.67, s	102.2	10'	4.97, t (7.1)	125.0
4'		119.4		114.5	11'		132.6
5'		128.2		134.7	12'	1.60, s	25.8
6'		156.5		157.5	13'	1.51, s	17.6
7'		173.5		171.4	14'	1.37, s	22.0
8'	5.31, d (14.8)	68.8	4.55, d (17.0)	48.9	15'	1.13, d (6.2)	23.5
	5.16, d (14.8)		4.38, d (17.0)				
9'			3.84, m	42.9			
10'			4.32, m	63.2			
3-OAc		172.9					
10'-OAc	2.09, s	21.1		172.7			



(−7.1) nm; ^1H and ^{13}C NMR data: see Table 3; HRESIMS m/z : 411.2141 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{23}\text{H}_{32}\text{NaO}_5$: 411.2142).

4.4. Cell cultures

MDA-MB-231 (human triple-negative breast cancer), MCF-7 (human breast cancer) and U-2OS (human osteosarcoma) cells were obtained from the Cell Bank of the Shanghai Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences (Shanghai, China). MDA-MB-231 was cultured in DMEM medium. MCF-7 cells and U-2OS cells were cultured in RPMI-1640 medium. All cell lines were supplemented with 10% FBS containing $50\ \mu\text{g mL}^{-1}$ penicillin and $50\ \mu\text{g mL}^{-1}$ streptomycin, and were grown in an incubator at $37\ ^\circ\text{C}$ with 5.0% CO_2 under a humidified environment.

4.5. Cell viability assays and colony formation assays

Cell viability was measured *via* a standard colorimetric test using MTT. Cells were plated in standard 96-well culture plates (5×10^3 cells per well). After a 24 h period of incubation, cells were treated with various concentrations of stachybochartin C (3) or stachybochartin G (7) for 24, 48 and 72 h. DMSO (0.1%) was used as a vehicle, the final concentration of which was less than 0.1% (v/v). $20\ \mu\text{L}$ of MTT ($5\ \text{mg mL}^{-1}$) solution was added to each well and the plates were incubated for about four hours. The formed formazan crystals were dissolved in DMSO ($150\ \mu\text{L}$ per well) under constant shaking for 10 min. The absorbance of formazan production was measured using an ELISA reader (SpectraMax Plus384, Molecular Devices, Sunnyvale, CA) at 570 nm (test wavelength) and 650 nm (reference wavelength). Cell viability was calculated *via* the following formula:

$$\% \text{ cell viability} = (A_t/A_s) \times 100\%$$

where A_t and A_s denote the absorbances of the test substance and solvent control, respectively.

For colony formation, 1000 cells were plated in 6-well culture plates. After treatment with various concentrations of stachybochartin C, stachybochartin G, and 0.1% DMSO for 14 days, the cells were stained with crystal violet solution (Beyotime, Jiangsu, China). The cell viability assays and colony formation assays were performed as previously described.²⁷

4.6. Annexin V-Alexa Fluor 647/PI staining assay for cell apoptosis

U-2OS cells were treated with various concentrations of stachybochartin C or stachybochartin G for 48 h. For flow cytometry, 1×10^6 cells in $100\ \mu\text{L}$ of binding buffer were stained with $5\ \mu\text{L}$ of Annexin V-Alexa Fluor 647 (BD Biosciences, San Jose, CA, USA) and $5\ \mu\text{L}$ of PI (Miltenyi Biotec, Shanghai, China). After incubation at room temperature and protection from light for 30 min, the cells were then analyzed *via* flow cytometry (488 nm excitation and 647 nm emission filters) using a BD Accuri C6 flow cytometer.

4.7. Western blot analysis

U-2OS cells were incubated with various concentrations of stachybochartin G or 0.1% DMSO for 24 h. After harvesting, the

cells were treated with cell lysis buffer (Beyotime, Jiangsu, China) to extract the total proteins and centrifuged at 12 000 rpm for 10 min. Proteins were extracted for western blot analysis, as previously described.²⁸ Bound immuno-complexes were detected using the ChemiDOC™ XRS+ system (BioRad Laboratories, Hercules, CA).

Conflicts of interest

The authors declare that they have no conflicts of interest.

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