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Application of dehydroalanine as a building block for the synthesis of selenocysteine-containing peptides†

 Kishorkumar M. Reddy and Govindasamy Mugesha *

Selenocysteine (Sec), the 21st proteinogenic amino acid, is inserted co-translationally into number of natural proteins. It is coded by a dual function stop codon UGA (opal). It is a redox active amino acid found at the active sites of several enzymes that are involved in oxidation–reduction reactions. These enzymes include the three major mammalian selenoproteins glutathione peroxidase (GPx), thioredoxin reductase (TrxR), and iodothyronine deiodinase (Dio). Although Sec is structurally similar to its sulfur analogue cysteine (Cys), the lower pK_a of the selenol group in Sec as compared to that of Cys and the interesting redox properties of the selenium atom in peptides and proteins play crucial roles in redox catalysis. However, the chemical synthesis of Sec-containing peptides has been a difficult task. In this paper, we report on a new method for the synthesis of Sec and Sec-containing peptides using dehydroalanine (Dha) as a building block.

Introduction

The genetically encoded selenocysteine (Sec, U)^{1,2} is incorporated into proteins by suppressing the stop codon UGA.³ The redox active Sec in proteins plays various roles in biological systems, ranging from maintaining the redox homeostasis to hormone activation and protein folding.⁴ Glutathione peroxidase (GPx), iodothyronine deiodinase (Dio) and thioredoxin reductase (TrxR) are some of the important selenoenzymes that play key roles in mammalian systems.⁴ The lower pK_a of the selenol group in Sec as compared to the thiol group in cysteine (Cys) and the relatively lower reduction potential of the selenium atom in the proteins make Sec a unique moiety for catalysis.⁵ However, the synthesis of selenoproteins follows a unique pathway. The biosynthesis of Sec involves the use of special mRNA sequence known as selenocysteine inserting sequence (SECIS) adjacent to the UGA codon in the prokaryotes and at the 3'-untranslation region in the eukaryotes. In prokaryotes, it requires a selenophosphate synthetase for the synthesis of selenophosphate, selenocysteine synthase to convert serine to Sec on tRNA, a specialised tRNA and Sec-binding protein to bind with tRNA and SECIS. On the other hand, in eukaryotes, the incorporation of Sec into proteins requires selenophosphate synthetase-2 for the synthesis of selenophosphate, SepSec synthase that can convert the phosphoserine to Sec on tRNA, a specialised tRNA, a Sec-binding protein and an elongation factor to bind with tRNA and SECIS.⁶

Interestingly, the key enzyme SepSec synthase in eukaryotes and selenocysteine synthase in prokaryotes are known to convert phosphoserine and serine, respectively, to selenocysteine on the tRNA using pyridoxal pyrophosphate as a co-factor. Recent studies showed that the mechanism of this pathway involves the formation of a dehydroalanine (Dha) intermediate, which is further converted to Sec by a nucleophilic attack of selenophosphate (Fig. 1).⁷ The formation of Dha through a spontaneous non-enzymatic elimination of phosphoserine is detected during protein aging in human cells.⁸ Furthermore, Dha is also produced during lantipeptide natural product biosynthesis in prokaryotes.⁹

Dehydroalanine (Dha) is a widely used precursor for the post-translational modifications in proteins. The amino acid residues serine (Ser), Cys or Sec are first inserted at the position of interest in proteins. These residues are then activated to form good leaving groups, which upon elimination, produces Dha. Synthetically, Dha can be produced by the oxidative elimination of an alkylated derivatives of Cys and Sec. Recently, biosynthetic incorporation of Sec-derivatives followed by an oxidative elimination of the aryl moiety with hydrogen peroxide has been shown to produce Dha moiety on peptides and proteins.¹⁰ However, only a few reports are available on the incorporation of selenium into proteins or peptides by chemical modifications. It has been shown that the activation of a serine residue with phenylmethanesulfonyl fluoride (PMSF), followed by displacement of the sulfonate with hydrogen selenide at pH 5.5 can give Sec-containing proteins.¹¹ Another method involving Native Chemical Ligation (NCL) of polypeptides between C-terminal thioester and N-terminal Sec residue has also been reported.¹² The solid-phase peptide synthesis (SPPS) of Sec-containing peptides using Fmoc-Sec(*p*mob)-OH is widely used,

Department of Inorganic and Physical Chemistry, Indian Institute of Science, Bangalore 560 012, India. E-mail: mugesha@iisc.ac.in

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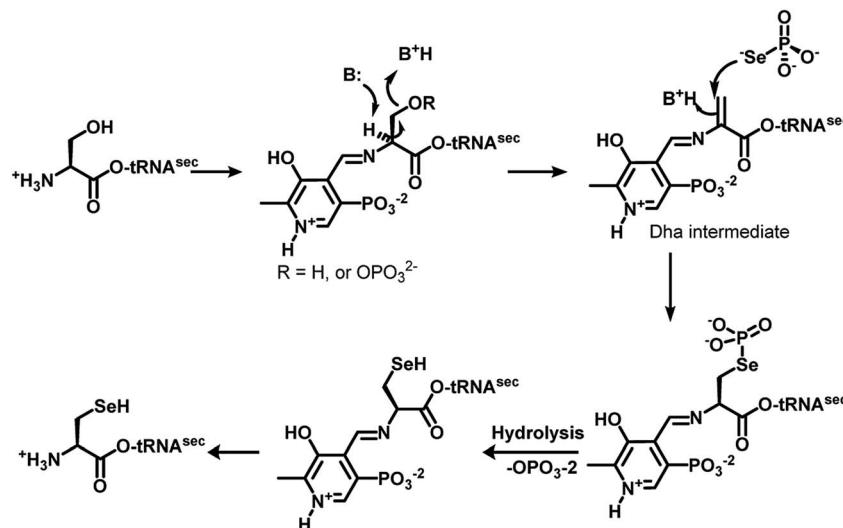


Fig. 1 Mechanism of SepSec synthase/selenocysteine synthase, which shows the formation of dehydroalanine before the attack of selenophosphate.⁷

but the protected amino acid is known to form Dha *via* α -hydrogen atom abstraction mechanism during the elongation step. The Dha residue thus formed can be attacked by piperidine to form β -piperidine adducts.¹³ As Dha plays a key role in protein modification and SPPS, we have developed a new method to insert Sec into peptides using a synthetic Dha as a building block.

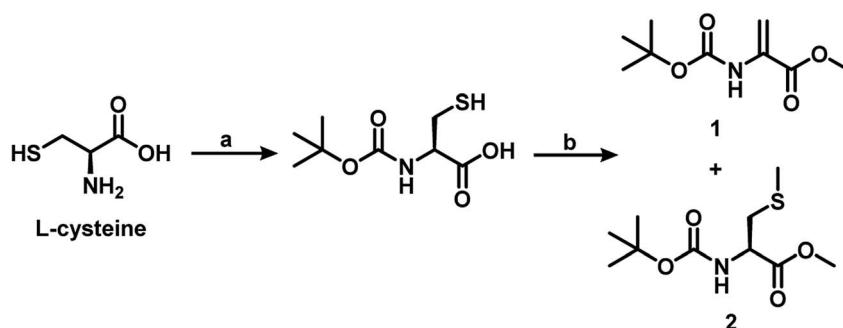
Results and discussion

Synthesis of peptides having Dha moiety

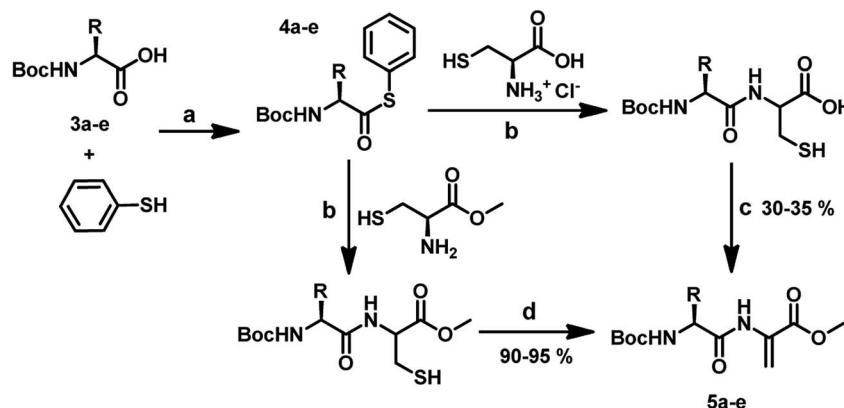
The early attempts to generate Dha on proteins and peptides relied on sulfonylation of Ser, followed by elimination under experimental conditions that were found to be too harsh for most proteins.¹⁴ Recent studies indicate that dehydroalanine (Dha) can be generated under mild conditions *via* post-translational modifications of Ser or Cys. When these two amino acids are part of peptides or proteins, the elimination of water and hydrogen sulfide, respectively, from these amino acid moieties produce the Dha. In our study, the Dha derivatives **1** and **5a–e** were synthesized following the literature procedures

with minor modifications.¹⁵ The Dha derivative **1** was synthesized starting from L-cysteine. The amine group of cysteine was protected with acid labile Boc and the resulting Boc-L-cysteine was treated with 5 equiv. of K_2CO_3 in DMF to afford compound **1** in 30% yield, along with the methylated cysteine derivative (**2**) in about 65% yield (Scheme 1). This observation indicates that it is important to protect the carboxylic group, before treating with methyl iodide for an alkylation–elimination reaction. Davis and co-workers showed that the reaction of BocCysOMe with a series of alkylating agents can produce Dha and the yield of Dha depends on the nature of alkylating agent. They showed that 1,4-dibromo- and 1,4-diiodobutanes are the most efficient of the reagents for the generation of Dha. 1,4-Dichlorobutane, on the other hand, was found to be slow to alkylate, and in this case, they recovered mostly the unreacted BocCysOMe. Other reagents such as 1,5-diiodopentane, 1,4-bis(methansulfonyl)-butandiol and α,α' -dibromo-*o*-xylene produced BocDhaOMe in only 11–12% yield.¹⁶

For the synthesis of Dha-based dipeptides **5a–e**, we employed the Boc-protected amino acids **3a–e**. To achieve this, compounds **3a–e** were treated with *N*-methylmorpholine, ethyl



Scheme 1 Synthetic route to dehydroalanine **1** from the L-cysteine. (a) 1 M aq. NaOH, Boc anhydride, 1,4 dioxane, 27 °C, 6 h. (b) K_2CO_3 , MeI, DMF, 27 °C, 20 h.



3a, 4a, 5a, R = -H, 3b, 4b, 5b, R = -CH₃, 3c, 4c, 5c, R = -CH₂CH(CH₃)₂, 3d, 4d, 5d, R = -CH(CH₃)CH₂CH₃, 3e, 4e, 5e, R = -CH₂C₆H₅

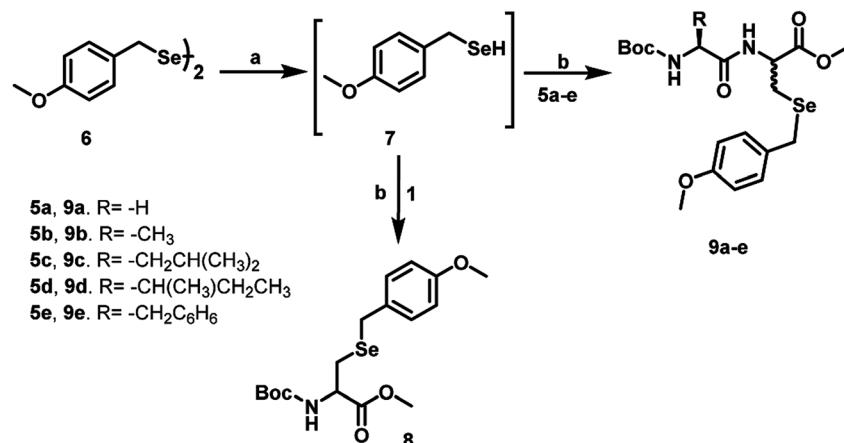
Scheme 2 Synthetic route to the dipeptides **5a–e** from the protected amino acids **3a–e** respectively via the formation of the corresponding thioesters **4a–e**. (a) Ethyl chloroformate, *N*-methylmorpholine, dry THF, molecular sieves, 27 °C, 6 h (b) NaBH₄, MeOH, 5 h. (c) K₂CO₃, MeI, DMF, 27 °C, 20 h. (d) K₂CO₃, 1,4-diiodobutane, DMF, 27 °C, 4 h.

chloroformate and thiophenol in dry THF to form the C-terminal thioesters **4a–e**. Peptide C-terminal thioesters are generally used as key intermediates in a variety of applications. Particularly, they have been used recently for the native chemical ligation methods for the total chemical synthesis of proteins.¹⁷ Therefore, it was thought worthwhile to explore the use of C-terminal thioesters for the synthesis of compounds **5a–e**. The native chemical ligation was performed between compounds **4a–e** and L-cysteine to produce the dipeptides bearing free thiol and carboxylic groups. When these dipeptides were treated with K₂CO₃ in DMF for 24 h, the dipeptides bearing C-terminal dehydroalanine centre (**5a–e**) were obtained (Scheme 2). Similar to the reaction of Boc-Cys with MeI, the formation of *S*-methylated peptides was observed as side products in these reactions as well. When we used L-cysteine methyl ester instead of L-cysteine in the native chemical ligation, we obtained dipeptides with a free thiol group. These

dipeptides when treated with 1,4-diiodobutane and K₂CO₃ in DMF for 4 h, afforded C-terminal Dha dipeptides in 90–95% without any other side products.

Reaction of Dha with *p*Mob selenol

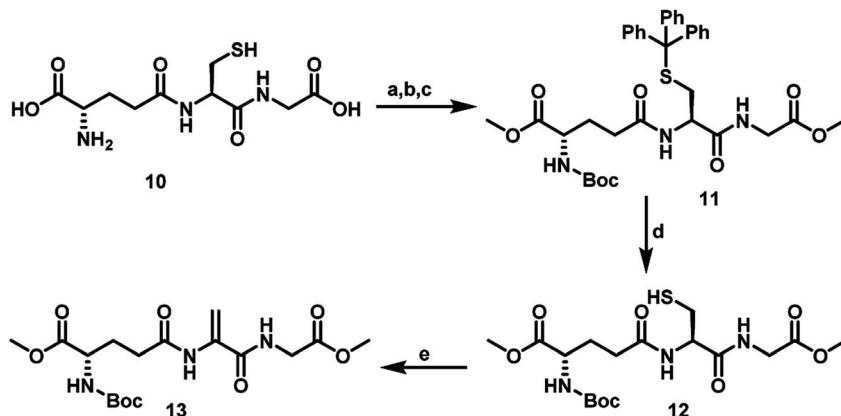
Synthetically, Dha is a versatile precursor for various natural and unnatural amino acid derivatives.¹⁸ The α,β -unsaturated carbonyl moiety present in Dha can undergo Michael-type conjugate addition reactions with various nucleophiles. These reactions are generally carried out in aqueous media at neutral or slightly basic pH and temperatures below 40 °C, the conditions that are compatible with proteins. Given the higher nucleophilicity of the selenolate as compared to that of a thiolate, it is expected that the Michael addition of aryl selenolates to Dha amides (selena-Michael addition) should be a highly favoured process. It has been shown that Dha is suitable for chemically generating Sec in proteins.¹⁹ However, only one



**5a, 9a, R = -H
5b, 9b, R = -CH₃
5c, 9c, R = -CH₂CH(CH₃)₂
5d, 9d, R = -CH(CH₃)CH₂CH₃
5e, 9e, R = -CH₂C₆H₅**

Scheme 3 Synthetic route to selenocysteine derivative **8** and dipeptides **9a–e** starting from the dipeptides **5a–e**. (a) NaBH₄, MeOH, 0 °C, 10 min, (b) 50 mM sodium phosphate buffer pH-8, MeOH, 27 °C, 6–8 h.

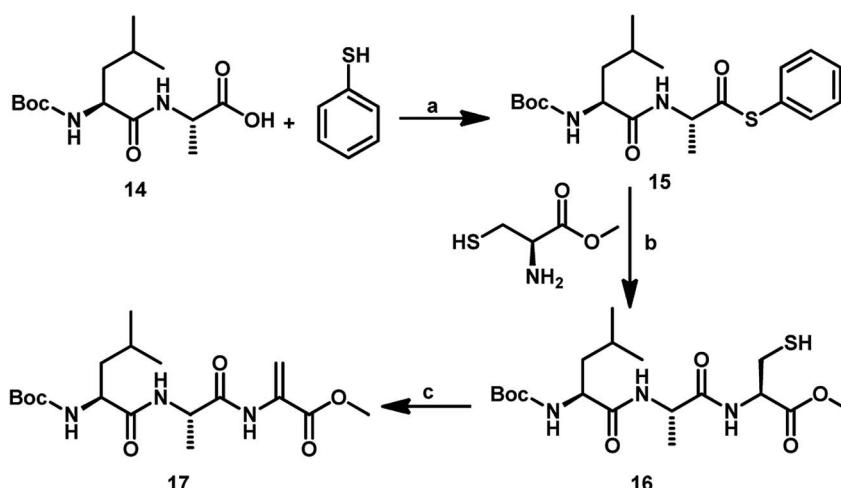




Scheme 4 Synthetic route to glutathione analogue **13** starting from the glutathione **10**. (a) Trityl alcohol, TFA. (b) Triethylamine, water, Boc anhydride. (c) K₂CO₃, MeI, DMF, 27 °C, 20 h. (d) Triethylsilane, 4% TFA, DCM, 27 °C, 1 h. (e) K₂CO₃, 1,4-diiodobutane in DMF, 27 °C, 4 h.

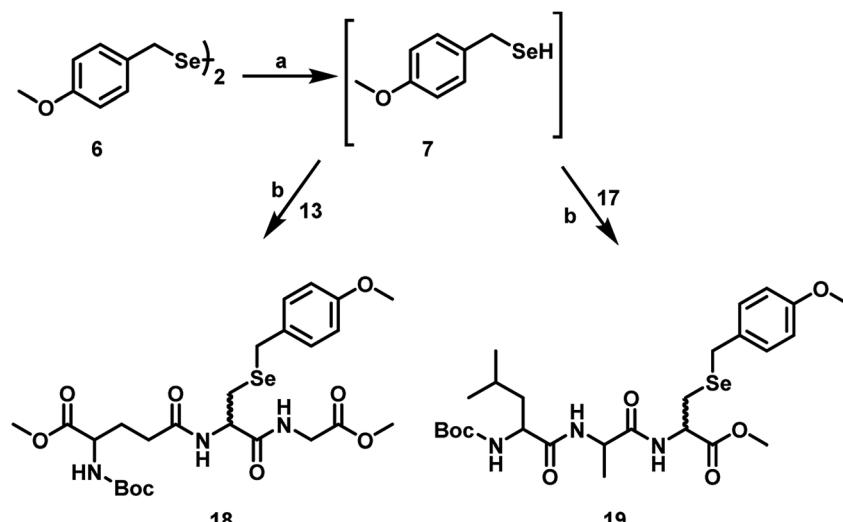
example of selena-Michael addition to Dha has been reported in the literature. This involved the formation of a Se-allyl-selenocysteine from a selenium nucleophile derived *in situ* from the precursor allylselenocyanate.¹⁹ However, this methodology may be useful for broader applications, when suitable selenium nucleophiles are available for the Michael addition. For our study, we selected *p*-methoxybenzyl (*p*Mob) as the protecting group as it is a well-known protecting group for the selenium in Sec.²⁰ For the reactions with Dha, we synthesized *p*Mob diselenide (**6**), which upon reduction with NaBH₄, *in situ* can generate *p*Mob selenol (**7**).²¹ The reduction of compound **6** by NaBH₄ was carried out in MeOH, in 50 mM sodium phosphate buffer pH 8.0 to obtain the corresponding selenol **7**. When the *in situ* generated *p*Mob selenol **7** was treated with the Dha derivative **1**, the *p*Mob-protected selenocysteine derivative **8** was obtained in good yield. Similarly, the reaction of **7** with the Dha-based dipeptides **5a–e** in 50 mM sodium phosphate buffer at pH 8.0 produced the Sec dipeptides **9a–e** in quantitative yields (Scheme 3).

We have extended this methodology to tripeptides bearing Dha at the C-terminal and central positions. For this purpose, we selected glutathione (GSH, **10**) as a model peptide to introduce Dha between two amino acid residues. It is known that GSH is an important antioxidant that prevents damage to cellular components caused by reactive oxygen species (ROS).²² It is a unique tripeptide with a γ -peptide linkage between the carboxyl group of the glutamate side chain and the amine group of cysteine. The carboxyl group of cysteine is attached by a normal peptide linkage to glycine. Therefore, the Dha methodology may be useful for the synthesis of the selenium analogue of GSH (GSeSeG). Previously, Tamura *et al.* synthesized the selenium analogue of glutathione disulphide, γ -L-glutamyl-L-selenocysteinylglycine, by a liquid phase method. In this method, the selenol group of Sec was protected by the *p*Mob group. The overall yield of the final product was ~9% based on the starting compound, Se-(*p*Mob)-L-selenocysteine.²³ Later, Iwaoka and co-workers reported the synthesis and antioxidant activity of GSeSeG. They synthesized GSeSeG from the Sec



Scheme 5 Synthetic route to the tripeptide **17** starting from dipeptide **14**. (a) Ethyl chloroformate, *N*-methylmorpholine, dry THF, molecular sieves, 27 °C, 6 h. (b) NaBH₄, MeOH, 5 h. (c) K₂CO₃, 1,4-diiodobutane, DMF, 27 °C, 4 h.





Scheme 6 Synthetic route to seleno-tripeptides **18** and **19** starting from the Dha-peptides **13** and **17** respectively. (a) NaBH_4 , MeOH , 0°C , 10 min, (b) 50 mM sodium phosphate buffer pH 8, MeOH , 27°C , 6–8 h.

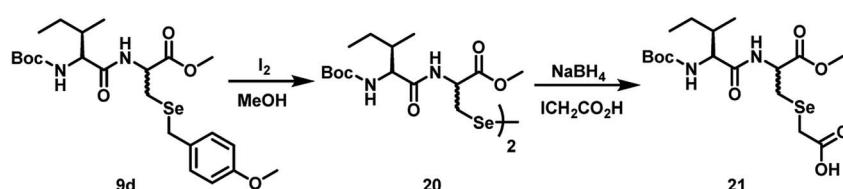
derivative, Fmoc-Sec(*p*Mob)-OH, using a solid-phase peptide synthesis method.²⁴

To synthesize the GSeSeG *via* the Dha route, it was necessary to protect the free amino and carboxylic groups. The protected GSH derivative **12** was synthesized by following the literature procedure.²⁵ Briefly, the thiol group of GSH was protected with trityl group by treating the thiol with trityl alcohol in TFA. The resulting trityl-protected GSH was treated with Boc anhydride under basic conditions to afford *N*-Boc-*S*-trityl-protected GSH. Esterification of this derivative with MeI in DMF produced *N*-Boc-*S*-trityl-glutathione methyl ester **11**. The deprotection of the trityl group by treating with triethylsilane in TFA afforded *N*-Boc-glutathione methyl ester **12**. The Dha analogue of glutathione **13** was obtained by treating **12** with 1,4-diiodobutane in DMF (Scheme 4).

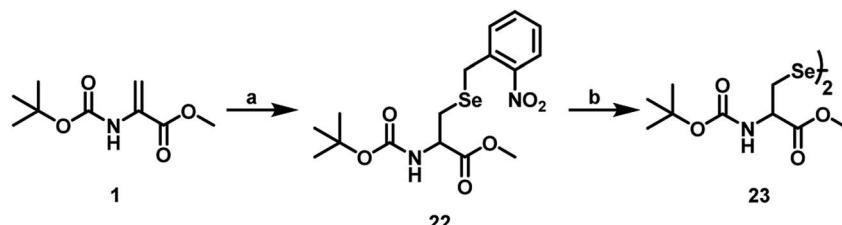
In addition to GSeSeG, several other selenium-containing peptides have been studied as glutathione peroxidase mimetics. Iwaoka and co-workers showed that the change in the amino acid sequence in a series of selenium-containing peptides alters the antioxidant activity.²⁴ For example, they showed that the replacement of the γ -Glu moiety in GSeSeG with a Leu residue significantly decreased the antioxidant activity. Therefore, we aimed at the synthesis of Sec-containing peptides having other amino acids such as Leu and Ala residues. For this, the tripeptide with C-terminal Dha centre (**17**)

was synthesized by following a conventional solution-phase peptide synthesis method. The Boc-protected dipeptide amino acid (Boc-NH-Leu-Ala-COOH, **14**) was treated with thiophenol in the presence of *N*-methylmorpholine and ethyl chloroformate in dry THF to give the dipeptide bearing C-terminal thioester **15**, which was used for the native chemical ligation with *L*-cysteine methyl ester to give the tripeptide **16**. Treatment of compound **16** with 1,4 diiodobutane and K_2CO_3 in DMF for 4 h afforded the tripeptide bearing Dha at C-terminal **17** (Scheme 5). We employed *L*-cysteine in the chemical ligation and followed by treatment with MeI and K_2CO_3 in DMF for 24 h afforded the Dha peptide in 30% yield.

Although the addition of functionalized thiols to Dha under mild conditions is generally a facile process, the nature of amino acid moiety attached to the Dha appears to modulate its reactivity. In this study, we observed that the nature of solvents plays an important role in the conversions as well as the reaction time. When the coupling of tripeptides bearing Dha (**13** and **17**) with *p*Mob selenol were performed in MeOH and 50 mM sodium phosphate buffer at pH 8, the selenium analogue of glutathione **18** and selenopeptide **19** were obtained in 80–90% yield. This agrees with an earlier report that the rate of Michael addition of amines and thiols to Dha amides was greatly accelerated in the presence of water (Scheme 6).²⁶



Scheme 7 Reaction of the selenocysteine dipeptide **9d** with iodine in methanol and a subsequent reaction with sodium borohydride and iodoacetic acid to produce the alkylated derivative **21**.



Scheme 8 Coupling of *o*-nitrobenzyl selenol with the Dha derivative **1** to afford the selenocysteine derivative **22**. (a) $\text{o-NO}_2\text{-C}_6\text{H}_4\text{-CH}_2\text{SeCN}$, NaBH_4 , MeOH , $0\text{ }^\circ\text{C}$ for 30 min; $27\text{ }^\circ\text{C}$, 8 h (b) 366 nm UV light, ACN and phosphate buffer pH 7 (1 : 1), 2 h.

Formation of epimers

It has been shown earlier that the addition of thiols to Dha lead to epimeric products, and the formation of epimeric (D-/L-) mixtures in these reactions can be viewed as a limitation of the synthetic functionalization of Dha.²⁷ This methodology may not be useful when enantiomerically pure selenium compounds are required. However, for the selenocysteine derivatives, the chirality may not play any key role in certain biological activities such as peroxynitrite scavenging. To understand the number of isomers produced in the selenocysteine derivatives, we carried out ^{77}Se NMR studies in CD_3OD at $27\text{ }^\circ\text{C}$ (Fig. S1, ESI†). For compound **9d**, two signals at 230.4 and 231.4 ppm were observed, indicating the formation of two epimers. As the amino acid attached to the selenocysteine moiety was enantiomerically pure (*R*-configuration), the two epimers formed in the reactions are expected to have *R,R* and *R,S* configurations (Scheme 7).

When compound **9d** was treated with I_2 in MeOH , it produced the corresponding diselenide **20**. The ^{77}Se NMR spectrum obtained for **20** showed four peaks at 295.6, 298.1, 298.6, 298.9 ppm, indicating the formation four epimers with possible configurations of *R,R,R,S*; *R,S,R,R*; *R,R,R,R* and *R,S,S,R*. The reduction of the diselenide **20** by sodium borohydride produced the corresponding selenol, which was trapped by treating the reaction mixture with iodoacetic acid. The alkylated compound **21** also showed two peaks in the ^{77}Se NMR spectrum at 190.4 and 191.1 ppm, indicating the formation of two epimers with *R,R* and *R,S* configurations.

Selenocysteine derivatives with a photo-cleavable group

In addition to the use of *p*Mob group, which can be removed from the peptide by treating with iodine in MeOH , it is known that a photo labile group such as *o*-nitrobenzyl group can also be used as a protecting group for selenium in selenocysteine.²⁸ For the introduction of *o*-nitrobenzyl group, we synthesized *o*-nitrobenzyl selenocyanide ($\text{o-NO}_2\text{-C}_6\text{H}_4\text{-CH}_2\text{SeCN}$) as precursor by following the literature procedure.²⁹ Briefly, the procedure involves treatment of *o*-nitrobenzyl bromide or chloride with KSeCN in acetone at reflux condition. The product must be recrystallized using EtOH to remove the impurities. Subsequently, *o*-nitrobenzyl selenocyanide was treated with NaBH_4 in MeOH to obtain *o*-nitrobenzyl selenol, which was added carefully to the Dha derivative **1** in MeOH to obtain compound **22** in around 30% yield (Scheme 8). The irradiation of compound **22**

at 366 nm UV light for around 2 h, produced the diselenide **23**, which was characterized by ESI-MS (Fig. S2B and S2C, ESI†). The UV-visible spectra of compound **23** exhibited a band (λ_{max}) at around 254, which disappeared upon irradiation (Fig. S2A, ESI†).

Conclusions

In this paper, we reported the incorporation of selenocysteine into peptides *via* dehydroalanine intermediate (Dha). Using this methodology, several di-, tri- and penta-peptides having Dha moiety were synthesized. A selenium moiety has been introduced into these peptides using *p*-methoxybenzyl (*p*Mob) selenol, derived from the corresponding diselenide. The *p*Mob group can be cleaved easily by treating the peptides with iodine in methanol. The Dha methodology was found to be useful, particularly, for the synthesis of the selenium analogue of glutathione. The Dha route can also be used to synthesize selenocysteine derivatives having photo-cleavable *o*-nitrobenzyl group, which can be readily cleaved under UV irradiation. Although the Dha methodology yields the selenium compounds as mixture of epimers, the compounds produced by this method should be useful for studies that do not depend on the chirality of the selenocysteine moiety.

Experimental section

General procedure

All the amino acids were purchased from GL Biochem (Shanghai) Ltd. Methanol was obtained from Merck. All other chemicals were of the highest purity available. Most reactions were carried out in a well-ventilated fume hood to avoid the unpleasant odour and toxic nature of the reaction mixtures involved. THF was dried over sodium metal with benzophenone. Thin-layer chromatography analyses were carried out on pre-coated silica gel plates (Merck), and spots were visualized under UV radiation. Column chromatography was performed on glass columns loaded with silica gel. ^1H (400 MHz), ^{13}C (100.56 MHz), and ^{77}Se (76.29 MHz) NMR spectra were obtained on a Bruker 400 MHz NMR spectrometer. Chemical shift values are cited with respect to SiMe_4 as internal (^1H and ^{13}C) and Me_2Se as external (^{77}Se) standard. Mass spectral studies were carried out on a Bruker Daltonics Esquire 6000 plus mass spectrometer with ESI-MS mode analysis.



Synthesis of 1

Step 1. In a 250 mL round bottom flask, L-cysteine (1.0 g, 8.2 mmol) was dissolved in 1 M aq. NaOH (25 mL). To the solution, Boc anhydride (2.16 g, 9.9 mmol) in 1,4-dioxane (8 mL) was added and the reaction mixture was stirred for 6 h at 27 °C. The reaction mixture was acidified with aq. KHSO₄ (50 mL) and extracted with ethyl acetate (3 × 50 mL). The organic layer was washed with water (3 × 50 mL) followed by brine (3 × 50 mL). The combined organic layer was dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure to give thick oil as product Boc-L-cysteine in good yield (1.7 g, 95%). The product was used in the next time without purification.

Step 2. To a solution of Boc-L-cysteine (1.5 g, 6.8 mmol) in DMF (30 mL), 5 equiv. of K₂CO₃ (4.69 g, 33.9 mmol) was added at 0 °C and the suspension was stirred for 15 min. To this stirred solution, methyl iodide (5 equiv., 425 µL, 6.8 mmol) was added and the stirring was continued for 24 h at 27 °C. The completion of reaction was followed by TLC. The reaction mixture was neutralized with aq. KHSO₄ and extracted with ethyl acetate (3 × 50 mL). The organic layer was washed with water (3 × 20 mL) and brine (5 × 20 mL) to remove the DMF. The combined organic layer was dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The crude residue was purified by column chromatography using 10% Pet. ether and ethylacetate to afford **1** (30%, 408 mg) as colorless oil. ¹H NMR (CDCl₃) δ (ppm) 6.97 (br, 1H), 6.08 (s, 1H), 5.63–5.64 (d, 1H), 3.74 (s, 3H), 1.40 (s, 9H); ¹³C NMR (CDCl₃) δ (ppm) 28.15, 52.76, 80.54, 105.05, 131.30, 152.46, 164.35; ESI-MS: *m/z* calcd for C₉H₁₅NO₄Na [M + Na]⁺: 224.0899, found 224.1136.

General procedure for the synthesis of thioesters **4a–e, 15**

The Boc-protected amino acid (**3a–e, 14**) (1.0 equiv., 5.71 mmol), *N*-methylmorpholine (1.2 equiv., 6.85 mmol) and dry THF (25 mL) were mixed in a dry flask containing 4 Å molecular sieves and the solution was cooled to 0 °C. To this, ethyl chloroformate (1.5 equiv., 8.56 mmol) was added and the reaction mixture was stirred for 15 min. Thiophenol (1.2 equiv., 6.85 mmol) and *N*-methylmorpholine (1.2 equiv., 6.85 mmol) were added and the reaction mixture was slowly warmed to 27 °C. The stirring was continued for 12 h at this temperature. The completion of the reaction was monitored by TLC. The reaction mixture was filtered to remove molecular sieves, and the solvent was removed under reduced pressure. The resulting residue was dissolved in dichloromethane (100 mL) and washed sequentially with NaHCO₃ (3 × 50 mL), H₂O (3 × 50 mL), and brine (3 × 50 mL). The organic phase was dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The product was purified by column chromatography to afford the products (**4a–e, 15**) (80–90%) as white solids. The spectral values obtained were matching with the literature values.^{15,30}

General procedure for the synthesis of Dha peptides **5a–e, 17**

Cysteine·HCl (1.0 equiv., 3.74 mmol) was added to MeOH (15 mL). NaBH₄ (2.0 equiv., 7.48 mmol) was added slowly, and the mixture was stirred for 1 h under N₂ atm. Thioesters (**4a–e, 15**)

(1.0 equiv., 3.74 mmol) was added in MeOH (10 mL), and the reaction mixture was stirred till the starting materials were fully consumed (~4 h, TLC analysis). After the removal of MeOH under reduced pressure, the residue was treated with water (20 mL). The reaction mixture was acidified with 1 N HCl, extracted with ethylacetate (3 × 50 mL) and washed with water (2 × 25 mL), followed by brine (3 × 25 mL). The organic layer was dried over anhydrous Na₂SO₄. After reducing the volume of the solvent to ~10 mL, TECP (0.5 equiv., 1.87 mmol) was added to reduce any disulphide bonds formed during the progress of the reaction. The reaction mixture was diluted with water (50 mL) and the aqueous layer was extracted once again with ethylacetate (3 × 50 mL) and washed with water (2 × 25 mL), followed by brine (3 × 25 mL). The organic layer was dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The intermediates formed in these reactions were used directly for the next step without further purification. These intermediate products were dissolved in DMF (25 mL) and added MeI (5 equiv., 18.7 mmol) and K₂CO₃ (5 equiv., 18.7 mmol) at 0 °C. After 15 min the ice bath was removed, and the reaction mixture was stirred at 27 °C for about 12 h. After completion of the reaction, which was followed by TLC, dil. HCl was added to neutralize the reaction mixture and the compound was extracted with ethylacetate (3 × 50 mL). The organic layer was washed with brine solution (5 × 25 mL) to remove DMF. It was dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The product was purified by column chromatography using 100–200 mesh silica and 20% petroleum ether and ethylacetate as eluent to afford the Dha-containing peptide **5a–e, 17** as viscous liquids which becomes solids on standing for long time.

Compound 5a. Thick viscous liquid, yield: 40% (386 mg, 1.49 mmol). ¹H NMR (D₄-CD₃OD), δ (ppm): 6.43 (s, 1H), 5.88 (s, 1H), 3.88 (s, 3H), 3.79 (s, 3H), 1.47 (s, 9H); ¹³C NMR (D₄-CD₃OD) δ (ppm) 28.64, 45.43, 49.00, 53.35, 81.00, 110.04, 132.84, 158.51, 165.38, 171.18; ESI-MS: *m/z* calcd for C₁₁H₁₈N₂O₅Na [M + Na]⁺: 281.1113, found: 281.0737.

Compound 5b. Thick viscous liquid, yield: 30% (305 mg, 1.12 mmol). ¹H NMR (CDCl₃), δ (ppm): 8.45 (br, 1H), 6.55 (s, 1H), 5.86 (s, 1H), 5.16–5.27 (br, 1H), 4.23–4.24 (br, 1H), 3.79 (s, 3H), 1.44 (s, 9H), 1.36–1.37 (d, 3H); ¹³C NMR (CDCl₃) δ 17.92, 28.26, 50.98, 52.93, 80.37, 109.26, 130.91, 155.53, 164.25, 171.78; ESI-MS: *m/z* calcd for C₁₂H₂₀N₂O₅Na [M + Na]⁺: 295.1270, found: 295.1051.

Compound 5c. Thick viscous liquid, yield: 40% (465 mg, 1.48 mmol). ¹H NMR (CDCl₃), δ (ppm): 8.41 (br, 1H), 6.57 (s, 1H), 5.86 (s, 1H), 5.05 (br, 1H), 4.19 (br, 1H), 3.79 (s, 3H), 1.66–1.69 (m, 2H), 1.47–1.51 (m, 1H), 1.41 (s, 9H), 0.90–0.92 (t, 6H); ¹³C NMR (CDCl₃), δ (ppm): 21.78, 23.01, 24.81, 28.28, 41.04, 52.93, 53.96, 80.27, 109.28, 130.91, 155.74, 164.28, 171.78; ESI-MS: *m/z* calcd for C₁₅H₂₆N₂O₅Na [M + Na]⁺: 337.1739, found: 337.1586.

Compound 5d. Thick viscous liquid, yield: 35% (408 mg, 1.30 mmol). ¹H NMR (CDCl₃), δ (ppm): 8.22 (br, 1H), 6.59 (s, 1H), 5.88 (s, 1H), 5.09 (br, 1H), 4.07 (brs, 1H), 3.81 (s, 3H), 1.92–1.93 (m, 1H), 1.47–1.51 (m, 1H), 1.44–1.51 (m, 1H), 1.42 (s, 9H), 1.09–1.16 (m, 1H), 0.87–0.94 (m, 6H); ¹³C NMR (CDCl₃), δ (ppm): 11.60, 15.73, 24.79, 28.36, 37.40, 53.06, 60.15, 80.27, 109.35,



130.71, 155.82, 164.41, 170.78; ESI-MS: *m/z* calcd for $C_{15}H_{26}N_2O_5Na$ [M + Na]⁺: 337.1739, found: 337.2340.

Compound 5e. White solid, yield: 35% (457 mg, 1.31 mmol). ¹H NMR (CDCl₃), δ (ppm): 8.15 (br, 1H), 7.18–7.31 (m, 5H), 6.61 (s, 1H), 5.89–5.89 (d, 1H), 4.99 (s, 1H), 4.43 (s, 1H), 3.78 (s, 3H), 3.11–3.16 (m, 2H), 1.40 (s, 9H); ¹³C NMR (CDCl₃), δ (ppm): 28.32, 38.30, 53.00, 56.64, 80.64, 109.48, 127.19, 128.88, 129.32, 130.72, 136.39, 155.50, 164.13, 170.38; ESI-MS: *m/z* calcd for $C_{18}H_{24}N_2O_5Na$ [M + Na]⁺: 371.1583, found: 371.3443.

Compound 17. Pale yellow solid, yield: 30% (430 mg, 1.11 mmol). ¹H NMR (CDCl₃), δ (ppm): 0.83–0.89 (m, 6H), 1.06–1.13 (m, 1H), 1.35–1.37 (d, 3H), 1.39 (s, 9H), 1.49–1.54 (m, 1H), 1.82 (br, 1H), 3.80 (s, 3H), 3.98–4.0 (d, 1H), 4.62–4.65 (dd, 1H), 5.34–5.36 (d, 1H), 5.87 (s, 1H), 6.52 (s, 1H), 7.0–7.01 (d, 1H), 8.54 (s, 1H); ¹³C NMR (CDCl₃), δ (ppm): 11.2, 15.5, 17.7, 24.8, 28.3, 37.3, 49.6, 52.9, 59.1, 79.9, 109.7, 131.2, 156.0, 164.2, 171.1, 172.3; ESI-MS: *m/z* calcd for $C_{18}H_{31}N_3O_6Na$ [M + Na]⁺: 408.2111; found: 408.2048.

Compound 13. Compound 12 was synthesized by following the reported procedure.²⁷ To a solution of 12 (500 mg, 1.15 mmol, 1 equiv.) in DMF (15 mL) was added 5 equiv. of K₂CO₃ (794 mg, 5.74 mmol) at 0 °C and stirred for 15 min, then added 5 equiv. of 1,4 diiodobutane (152 μ L, 5.75 mmol) and stirred for 6 h at 27 °C. The completion of reaction was monitored by TLC. After completion of the reaction, 1 N HCl was added to neutralize the reaction mixture and extracted with ethylacetate (3 \times 25 mL), washed with brine (5 \times 15 mL) to remove the DMF. The organic layer was dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The product was purified by column chromatography using 100–200 mesh silica and 5% dichloromethane and methanol as eluent to afford the product 13 as light yellow oil. Yield: 90% (415 mg, 1.03 mmol) ¹H NMR (CDCl₃), δ (ppm): 1.37 (s, 9H), 1.93–2.14 (m, 2H), 2.39 (brs, 2H), 3.68 (s, 3H), 3.72 (s, 3H), 4.06 (s, 2H), 4.25 (s, 1H), 5.38 (s, 1H), 6.37 (s, 1H), 7.35 (s, 1H), 8.16 (s, 1H); ¹³C NMR (CDCl₃), δ (ppm): 28.8, 30.2, 33.8, 42.1, 52.9, 53.0, 53.4, 80.5, 103.8, 134.3, 156.1, 164.9, 170.7, 171.5, 173.3; ESI-MS: *m/z* calcd for $C_{17}H_{27}N_3O_8Na$ [M + Na]⁺: 424.1696; found: 424.4928.

General procedure for the synthesis of Sec derivatives

NaBH₄ (4 equiv., 6.0 mmol) was added portion-wise to a solution of *p*-methoxy benzylselenide or *o*-NO₂-C₆H₄-CH₂SeCN (2 equiv., 3.0 mmol) in degassed MeOH (1 mL per 0.03 mmol) at 0 °C under N₂ atmosphere. An immediate evolution of hydrogen gas was observed. After 5 min, this solution was added dropwise to Dha containing aminoacid (1)/dipeptides (5a–e)/tripeptides (13, 17) (1 equiv., 1.50 mmol) in 0.5 M sodium phosphate (pH-8) (1 mL per 0.003 mmol) and stirred at 27 °C for 4–6 h. The reaction mixture was treated with EtOAc and washed subsequently with 1 M aq. HCl, saturated NaHCO₃, water and brine. The organic layers were combined and dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The crude residue was purified by column chromatography (100–200 mesh silica gel for compounds 8, 9a–9e, and 230–400 mesh silica gel for compound 18, 19 & 22) to afford the protected

selenocysteine derivatives, seleno dipeptides 8, 9a–e, 18, 19, in 80–90% yields, and 22 in 35% yield.

Compound 8. Eluent for column chromatography: 20% Pet. ether/ethylacetate. Slight yellow solid, yield: 95% (570 mg, 1.41 mmol). ¹H NMR (CDCl₃), δ (ppm): 1.45 (s, 9H), 2.88 (s, 2H), 3.74 (s, 3H), 3.76 (s, 2H), 3.78 (s, 3H), 4.56–4.61 (t, 1H), 5.28–5.30 (d, 1H), 6.81–6.83 (d, 2H), 7.19–7.21 (d, 2H); ¹³C NMR (CDCl₃), δ (ppm): 25.9, 27.5, 28.4, 52.6, 53.5, 55.4, 80.2, 114.1, 130.1, 130.7, 155.2, 158.7, 171.8; ⁷⁷Se NMR (CDCl₃), δ (ppm): 215.3; ESI-MS: *m/z* calcd for $C_{17}H_{25}NO_5SeNa$ [M + Na]⁺: 426.0796; found: 426.0714.

Compound 9a. Eluent for column chromatography: 20% Pet. ether/ethylacetate. Thick viscous oil, yield: 90% (621 mg, 1.35 mmol). ¹H NMR (CDCl₃), δ (ppm): 1.41 (s, 9H), 2.85–2.86 (m, 2H), 3.69 (s, 3H), 3.71 (s, 2H), 3.73 (s, 3H), 4.79–4.84 (m, 1H), 5.43–5.46 (t, 1H), 6.77–6.79 (m, 2H), 7.14 (br, 1H), 7.16–7.17 (d, 2H); ¹³C NMR (CDCl₃), δ (ppm): 25.1, 27.4, 28.3, 44.2, 52.0, 52.6, 55.2, 80.1, 114.0, 130.0, 130.5, 156.1, 169.6, 171.2; ⁷⁷Se NMR (CDCl₃), δ (ppm): 216.4; ESI-MS: *m/z* calcd for $C_{19}H_{28}N_2O_6SeNa$ [M + Na]⁺: 483.1010; found: 483.2105.

Compound 9b. Eluent for column chromatography: 20% Pet. ether/ethylacetate. Yield: 90% (639 mg, 1.34 mmol) ¹H NMR (CDCl₃), δ (ppm): 1.33–1.36 (dd, 3H), 1.43 (d, 9H), 2.83–2.94 (m, 2H), 3.72 (d, 3H), 3.73 (s, 2H), 3.77 (s, 3H), 4.19–4.20 (br, 1H), 4.80–4.84 (m, 1H), 5.09–5.11 (br, 1H), 6.79–6.82 (dd, 2H), 6.85–7.0 (br, 1H), 7.17–7.19 (d, 2H); ¹³C NMR (CDCl₃), δ (ppm): 17.96, 18.04, 24.53, 26.73, 26.78, 27.91, 49.59, 49.65, 51.75, 52.02, 52.05, 54.71, 77.35, 79.23, 113.53, 129.61, 130.13, 130.24, 155.04, 155.11, 158.08, 170.81, 170.93, 172.73; ⁷⁷Se NMR (CDCl₃), δ (ppm): 214.9, 215.7; ESI-MS: *m/z* calcd for $C_{20}H_{30}N_2O_6SeNa$ [M + Na]⁺: 497.1167; found: 497.4734.

Compound 9c. Eluent for column chromatography: 20% Pet. ether/ethylacetate. Yield: 85% (658 mg, 1.27 mmol) ¹H NMR (CDCl₃), δ (ppm): 0.2–0.93 (t, 3H), 1.42–1.43 (d, 9H), 1.45–1.47 (m, 1H), 1.65–1.70 (m, 1H), 2.86–2.91 (m, 2H), 3.72–3.74 (d, 3H), 3.77 (s, 2H), 3.79 (s, 3H), 4.08–4.15 (br, 1H), 4.81–4.85 (m, 1H), 4.90–4.94 (t, 1H), 6.80–6.86 (dd, 2H), 6.88–6.97 (br, 1H), 7.18–7.20 (d, 2H); ¹³C NMR (CDCl₃), δ (ppm): 21.67, 21.75, 22.64, 22.73, 24.41, 24.49, 24.72, 26.90, 26.97, 28.06, 40.96, 40.99, 51.88, 52.15, 52.19, 52.77, 54.88, 77.35, 79.40, 113.68, 129.76, 130.25, 130.40, 155.39, 155.48, 158.25, 170.94, 171.02, 172.63; ⁷⁷Se NMR (CDCl₃), δ (ppm): 213.5, 214.0; ESI-MS: *m/z* calcd for $C_{23}H_{36}N_2O_6SeNa$ [M + Na]⁺: 539.1636; found: 539.1557.

Compound 9d. Eluent for column chromatography: 20% Pet. ether/ethylacetate. Yield: 80% (619 mg, 1.20 mmol) ¹H NMR (CDCl₃), δ (ppm): 0.89–0.95 (m, 6H), 1.08–1.18 (m, 1H), 1.43–1.44 (d, 9H), 1.46–1.50 (m, 1H), 1.84–1.97 (m, 1H), 2.87–2.95 (m, 2H), 3.73 (s, 3H), 3.74 (s, 2H), 3.78 (s, 3H), 3.97–4.05 (m, 1H), 4.82–4.87 (m, 1H), 5.01 (br, 1H), 6.55–6.72 (m, 1H), 6.81–6.23 (dd, 2H), 7.18–7.20 (d, 2H); ¹³C NMR (CDCl₃), δ (ppm): 11.44, 11.51, 15.46, 15.66, 24.57, 24.74, 25.11, 27.25, 27.37, 28.31, 37.33, 37.39, 51.95, 51.99, 52.49, 52.53, 55.22, 59.16, 79.81, 114.00, 130.01, 130.44, 130.54, 155.64, 155.72, 158.59, 171.08, 171.17, 171.48, 171.53; ⁷⁷Se NMR (CDCl₃), δ (ppm): 213.7, 214.0; ESI-MS: *m/z* calcd for $C_{23}H_{37}N_2O_6SeNa$ [M + Na + H]⁺: 540.1715; found: 540.4047.





Compound 9e. Eluent for column chromatography: 20% Pet. ether/ethylacetate. Yield: 85% (696 mg, 1.26 mmol) ^1H NMR (CDCl_3), δ (ppm): 1.40 (s, 9H), 2.73–2.87 (m, 2H), 3.07–3.11 (m, 2H), 3.68–3.72 (m, 5H), 3.78 (s, 3H), 4.39 (br, 1H), 4.77–4.83 (m, 1H), 5.00 (br, 1H), 6.61–6.66 (m, 1H), 6.81–6.83 (d, 2H), 7.17–7.29 (m, 5H), 7.30 (d, 2H); ^{13}C NMR (CDCl_3), δ (ppm): 24.77, 24.84, 27.02, 27.08, 28.07, 38.23, 51.86, 52.04, 52.27, 52.30, 54.97, 55.37, 77.36, 79.71, 113.78, 126.58, 128.31, 129.16, 129.82, 130.28, 130.40, 136.52, 136.62, 155.20, 155.30, 158.35, 170.78, 170.96, 171.25; ^{77}Se NMR (CDCl_3), δ (ppm): 217.9, 218.6; ESI-MS: m/z calcd for $\text{C}_{26}\text{H}_{34}\text{N}_2\text{O}_6\text{SeNa} [\text{M} + \text{Na}]^+$: 573.1480; found: 573.3649.

Compound 18. Eluent for column chromatography: 10% dichloromethane/methanol. Yield: 80% (718 mg, 1.19 mmol) ^1H NMR (CD_3OD), δ (ppm): 1.42 (s, 9H), 1.85–1.96 (m, 1H), 2.04–2.22 (t, 2H), 2.69–2.75 (m, 1H), 2.89–2.97 (m, 1H), 3.69 (s, 3H), 3.71 (s, 2H), 3.75 (s, 3H), 3.79 (s, 2H), 3.95–3.96 (br, 2H), 4.15–4.18 (q, 1H), 6.81–6.83 (d, 2H), 7.21–7.24 (d, 2H), 8.28–8.31 (t, 1H); ^{13}C NMR (CD_3OD), δ (ppm): 24.77, 24.84, 27.02, 27.08, 28.07, 38.23, 51.86, 52.04, 52.27, 52.30, 54.97, 55.37, 77.36, 79.71, 113.78, 126.58, 128.31, 129.16, 129.82, 130.28, 130.40, 136.52, 136.62, 155.20, 155.30, 158.35, 170.78, 170.96, 171.25; ^{77}Se NMR (CD_3OD), δ (ppm): 217.9, 218.6; ESI-MS: m/z calcd for $\text{C}_{25}\text{H}_{37}\text{N}_3\text{O}_9\text{SeNa} [\text{M} + \text{Na}]^+$: 626.1593; found: 626.9387.

Compound 19. Eluent for column chromatography: 10% dichloromethane/methanol. Yield: 85% (743 mg, 1.26 mmol) ^1H NMR (CDCl_3), δ (ppm): 0.86–0.93 (m, 6H), 1.08–1.16 (m, 1H), 1.24 (s, 1H), 1.36–1.39 (dd, 3H), 1.42–1.43 (d, 9H), 1.48 (br, 1H), 1.73 (br, 1H), 1.86–1.90 (m, 1H), 2.83–2.94 (m, 2H), 3.72 (s, 3H), 3.73–3.74 (d, 2H), 3.78–3.79 (d, 3H), 3.94–3.97 (t, 1H), 4.49–4.54 (m, 1H), 4.78–4.80 (m, 1H), 5.04–5.06 (d, 1H), 6.53–6.94 (m, 1H), 6.81–6.84 (dd, 2H), 7.18–7.20 (dd, 2H); ^{13}C NMR (CDCl_3), δ (ppm): 11.26, 15.48, 15.53, 18.48, 24.63, 24.79, 24.97, 27.12, 27.23, 28.27, 29.54, 37.40, 48.61, 52.13, 52.32, 52.39, 55.11, 59.01, 79.47, 113.89, 129.92, 130.49, 130.54, 155.82, 158.46, 171.07, 171.13, 171.73, 171.80, 172.09, 172.18; ^{77}Se NMR (CDCl_3), δ (ppm): 221.8, 222.8; ESI-MS: m/z calcd for $\text{C}_{26}\text{H}_{41}\text{N}_3\text{O}_7\text{SeNa} [\text{M} + \text{Na}]^+$: 610.2007; found: 610.1074.

Compound 23. Eluent for column chromatography: 20% Pet. ether/ethylacetate. Yield: 35% (217 mg, 0.52 mmol) ^1H NMR (CDCl_3), δ (ppm): 1.44 (s, 9H), 2.90–3.01 (m, 2H), 3.73 (s, 3H), 4.06–4.15 (q, 2H), 4.59–4.61 (d, 1H), 5.32–5.34 (d, 1H), 7.36–7.41 (m, 2H), 7.51–7.55 (m, 1H), 8.01–8.03 (dd, 1H); ^{13}C NMR (CDCl_3), δ (ppm): 25.4, 27.1, 28.8, 53.1, 54.0, 80.7, 126.3, 128.6, 132.4, 133.8, 135.9, 148.3, 155.5, 171.9; ^{77}Se NMR (CDCl_3), δ (ppm): 233; ESI-MS: m/z calcd for $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_6\text{SeNa} [\text{M} + \text{Na}]^+$: 441.0541; found: 441.3427.

Conflicts of interest

There are no conflicts of interest to declare.

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Notes and references

- (a) T. C. Stadtman, *Science*, 1974, **183**, 915–922; (b) J. E. Cone, R. M. Del Río, J. N. Davis and T. C. Stadtman, *Proc. Natl. Acad. Sci. U. S. A.*, 1976, **73**, 2659–2663.
- (a) A. Böck, K. Forchhammer, J. Heider and C. Baron, *Trends Biochem. Sci.*, 1991, **16**, 463–467; (b) A. Böck, K. Forchhammer, J. Heider, W. Leinfelder, G. Sawers, B. Veprek and F. Zinoni, *Mol. Microbiol.*, 1991, **5**, 515–520.
- (a) F. Zinoni, A. Birkmann, T. C. Stadtman and A. Böck, *Proc. Natl. Acad. Sci. U. S. A.*, 1986, **83**, 4650–4654; (b) I. Chambers, J. Frampton, P. Goldfarb, N. Affara, W. McBain and P. R. Harrison, *EMBO J.*, 1986, **5**, 1221–1227; (c) B. J. Lee, P. J. Worland, J. N. Davis, T. C. Stadtman and D. L. Hatfield, *J. Biol. Chem.*, 1989, **264**, 9724–9727.
- (a) L. Flohé, E. A. Günzler and H. H. Schock, *FEBS Lett.*, 1973, **32**, 132–134; (b) J. T. Rotruck, A. L. Pope, H. E. Ganther, A. B. Swanson, D. G. Hafeman and W. G. Hoekstra, *Science*, 1973, **179**, 588–590; (c) H. Sies, *Angew. Chem., Int. Ed.*, 1986, **25**, 1058–1071; (d) D. Behne, A. Kyriakopoulos, H. Meinhold and J. Köhrle, *Biochem. Biophys. Res. Commun.*, 1990, **173**, 1143–1149; (e) T. Tamura and T. C. Stadtman, *Proc. Natl. Acad. Sci. U. S. A.*, 1996, **93**, 1006–1011; (f) G. Mugesha, W.-W. du Mont and H. Sies, *Chem. Rev.*, 2001, **101**, 2125–2180; (g) J. Köhrle, *Methods Enzymol.*, 2002, **347**, 125–167; (h) G. Roy, B. K. Sarma, P. P. Phadnis and G. Mugesha, *J. Chem. Sci.*, 2005, **117**, 287–303; (i) M. Conrad, M. Schneider, A. Seiler and G. W. Bornkamm, *Biol. Chem.*, 2007, **388**, 1019–1025; (j) K. P. Bhabak and G. Mugesha, *Acc. Chem. Res.*, 2010, **43**, 1408–1419; (k) T. Wirth, *Angew. Chem., Int. Ed.*, 2015, **55**, 10074–10076; (l) S. Mondal, K. Raja, U. Schweizer and G. Mugesha, *Angew. Chem., Int. Ed.*, 2016, **55**, 7606–7630.
- (a) A. P. Arnold, K. S. Tan and D. L. Rabenstein, *Inorg. Chem.*, 1986, **25**, 2433–2437; (b) B. J. Byun and Y. K. Kang, *Biopolymers*, 2011, **95**, 345–353.
- (a) W. Leinfelder, E. Zehelein, M. A. MandrandBerthelot and A. Böck, *Nature*, 1988, **331**, 723–725; (b) A. A. Turanov, X. M. Xu, B. A. Carlson, M. H. Yoo, V. N. Gladyshev and D. L. Hatfield, *Adv. Nutr.*, 2011, **2**, 122–128; (c) E. G. Varlamova, M. V. Goltyaev, S. V. Novoselov, V. I. Novoselov and E. E. Fesenko, *Mol. Biol.*, 2013, **47**, 558–567.
- (a) S. Palioura, R. L. Sherrer, T. A. Steitz, D. Söll and M. Simonovic, *Science*, 2009, **325**, 321–325; (b) K. Forchhammer and A. Böck, *J. Biol. Chem.*, 1991, **266**, 6324–6328.
- (a) P. M. Masters, *Calcif. Tissue Int.*, 1985, **37**, 236–241; (b) Z. Wang, B. Lyons, R. J. W. Truscott and K. L. Schey, *Aging Cell*, 2014, **13**, 226–234.
- (a) P. J. Knerr and W. A. van der Donk, *Annu. Rev. Biochem.*, 2012, **81**, 479–505; (b) Y. Shi, X. Yang, N. Garg and W. A. van der Donk, *J. Am. Chem. Soc.*, 2011, **133**, 2338–2341.

10 J. Wang, S. M. Schiller and P. G. Schultz, *Angew. Chem., Int. Ed.*, 2007, **46**, 6849–6851.

11 Z. P. Wu and D. Hilvert, *J. Am. Chem. Soc.*, 1989, **111**, 4513–4514.

12 (a) R. J. Hondal, B. L. Nilsson and R. T. Raines, *J. Am. Chem. Soc.*, 2001, **123**, 5140–5141; (b) R. Quaderer, A. Sewing and D. Hilvert, *Helv. Chim. Acta*, 2001, **84**, 1197–1206.

13 (a) D. Besse and L. Moroder, *J. Pept. Sci.*, 1997, **3**, 442–453; (b) M. Rooseboom, N. P. E. Vermeulen, L. Andreadou and J. N. M. Commandeur, *J. Pharmacol. Exp. Ther.*, 2000, **294**, 762–769.

14 D. H. Strumeyer, W. N. White and D. E. Koshland Jr, *Proc. Natl. Acad. Sci. U. S. A.*, 1963, **50**, 931–935.

15 L. Markey, S. Giordani and E. M. Scanlan, *J. Org. Chem.*, 2013, **78**, 4270–4277.

16 (a) D. H. Rich, J. Tam, P. Mathiaparanam, J. A. Grant and C. Mabuni, *J. Chem. Soc., Chem. Commun.*, 1974, 897–898; (b) J. M. Chalker, S. B. Gunnoo, O. Bouteureira, S. C. Gerstberger, M. Fernández-González, G. J. L. Bernardes, L. Griffin, H. Hailu, C. J. Schofield and B. G. Davis, *Chem. Sci.*, 2011, **2**, 1666–1676.

17 (a) P. E. Dawson and S. B. H. Kent, *Annu. Rev. Biochem.*, 2000, **69**, 923–960; (b) P. Thapa, R. Y. Zhang, V. Menon and J. P. Bingham, *Molecules*, 2014, **19**, 14461–14483.

18 (a) B. G. Davis, R. C. Lloyd and J. B. Jones, *Chem.-Asian J.*, 2009, **4**, 630–640; (b) J. Dadová, S. R. G. Galam and B. G. Davis, *Curr. Opin. Chem. Biol.*, 2018, **46**, 71–81.

19 Y. A. Lin, O. Bouteureira, L. Lercher, B. Bhushan, R. S. Paton and B. G. Davis, *J. Am. Chem. Soc.*, 2013, **135**, 12156–12159.

20 (a) M. D. Gieselman, L. Xie and V. A. Wilfred A, *Org. Lett.*, 2001, **3**, 1331–1334; (b) R. J. Hondal, *Protein Pept. Lett.*, 2005, **12**, 757–764.

21 O. Flögel, G. Casi, D. Hilvert and D. Seebach, *Helv. Chim. Acta*, 2007, **90**, 1651–1666.

22 A. Pompella, A. Visvikis, A. Paolicchi, V. De Tata and A. F. Casini, *Biochem. Pharmacol.*, 2003, **66**, 1499–1503.

23 T. Tamura, T. Oikawa, A. Ohtaka, N. Fujii, N. Esaki and K. Soda, *Anal. Biochem.*, 1993, **208**, 151–154.

24 S. Yoshida, F. Kumakura, I. Komatsu, K. Arai, Y. Onuma, H. Hojo, B. G. Singh, K. I. Priyadarsini and M. Iwaoka, *Angew. Chem., Int. Ed.*, 2011, **50**, 2125–2128.

25 D. Crich, V. Krishnamurthy and T. K. Hutton, *J. Am. Chem. Soc.*, 2006, **128**, 2544–2545.

26 B. N. Naidu, M. E. Sorenson, T. P. Connolly and Y. Ueda, *J. Org. Chem.*, 2003, **68**, 10098–10102.

27 C. D. Spicer and B. G. Davis, *Nat. Commun.*, 2014, **5**, 4740.

28 M. Abbas and L. A. Wessjohann, *Org. Biomol. Chem.*, 2012, **10**, 9330–9333.

29 D. Plano, Y. Baquedano, D. Moreno-Mateos, M. Font, A. Jiménez-Ruiz, J. A. Palop and C. Sanmartín, *Eur. J. Med. Chem.*, 2011, **46**, 3315–3323.

30 (a) X. Guan, M. Drake and Z. Tan, *Org. Lett.*, 2013, **15**, 6128–6131; (b) Q. Wan, J. Chen, Y. Yuan and S. J. Danishefsky, *J. Am. Chem. Soc.*, 2008, **130**, 15814–15816.

