



Cite this: *RSC Adv.*, 2019, 9, 1759

Total synthesis of diazaquinomycins H and J using double Knorr cyclization in the presence of triisopropylsilane†

Allan M. Prior and Dianqing Sun *

The first total synthesis of diazaquinomycins H (1) and J (2), which are promising anti-tuberculosis natural product leads, has been achieved *via* selective amidation of diamine 6 with Meldrum's acid derivatives, subsequent EDC coupling with 3-oxobutanoic acid, followed by double Knorr cyclization in the presence of triisopropylsilane (TIPS). We found that the addition of TIPS was crucial to obtain pure diazaquinomycins H and J, while preventing isomerization of the terminal iso-branched tail in sulfuric acid. Our developed synthesis provided diazaquinomycins H (1) and J (2) in 8 steps from commercially available starting materials in 25% and 21% overall yields, respectively. The spectroscopic data of synthetic diazaquinomycins H (1) and J (2) agreed very favorably with those of reported natural products.

Received 28th November 2018
Accepted 4th January 2019

DOI: 10.1039/c8ra09792e

rsc.li/rsc-advances

Introduction

Diazaquinomycins (DAQs) are secondary metabolites produced by terrestrial and aquatic *Actinobacteria*, predominantly from the genus *Streptomyces* sp.^{1–5} Spanning the last three and a half decades, nine DAQs have been reported, however, diazaquinomycins D, F, and G were not obtained in pure form (Fig. 1).^{1–5} Specifically, diazaquinomycin D was a trace component in a sample of diazaquinomycin C that was produced by *Streptomyces* sp. GW48/1497 when cultivated in soybean flour and mannitol.³ Its structure was assigned using (–)-ESI mass spectrometry.³ The isomeric diazaquinomycins F and G were isolated as an inseparable mixture;² their structures were established using a combination of NMR and UV spectroscopy, HRMS, and X-ray crystallography.²

Diazaquinomycins H (1) and J (2), bearing a long lipophilic side chain with a unique terminal isopropyl functionality, are the latest additions to this family and were isolated by Mallowney *et al.* from fresh water-derived *Actinobacteria* in Lake Michigan.¹ Notably, diazaquinomycins H and J along with

diazaquinomycin A showed excellent *in vitro* antibacterial activity against *Mycobacterium tuberculosis* (*M. tuberculosis*) with minimum inhibitory concentration (MIC) values of 0.04, 0.07, and 0.10 $\mu\text{g mL}^{-1}$, respectively.¹ In addition, these analogues did not display mammalian cytotoxicity against Vero cells when tested at 28 μM , however, the limited quantity (0.3 mg) of isolated diazaquinomycins H and J prevented further antimicrobial assessment.¹ More extensive evaluation and profiling using diazaquinomycin A revealed that it also retained potent anti-tuberculosis activity (MIC = 0.06–0.27 $\mu\text{g mL}^{-1}$) against drug resistant *M. tuberculosis* strains with known resistance to rifampicin, isoniazid, streptomycin, kanamycin, or cycloserine.¹ The specific mechanism of action of diazaquinomycin A against *M. tuberculosis* remains unclear, though it was previously reported to interfere with the folate pathway,⁶ acting as a competitive inhibitor of thymidylate synthase.^{7,8} However, *M. tuberculosis* does not encode human thymidylate synthase and *M. tuberculosis* thymidylate synthase enzymes (ThyA and ThyX) were not targeted by diazaquinomycin A.¹

Diazaquinomycins A and B were first synthesized in 1988 by Kelly *et al.* *via* a double Knorr cyclization of a symmetrical di- β -ketoanilide hydroquinone precursor.⁹ In 1990, Lee *et al.* reported the synthesis of a diazaanthraquinone scaffold using hetero Diels–Alder methodology by coupling a dienophile with a 1-azadiene, followed by dibromohydrin formation and oxidation.¹⁰ In 1998 and 2000, two alternative syntheses of diazaquinomycin A were reported by Perez *et al.* and similarly made use of hetero Diels–Alder chemistry to construct the tricyclic core.^{11,12} Subsequent aromatization, N-oxide formation, and tosyl chloride mediated N-oxide rearrangement afforded diazaquinomycin A.^{11,12} The Rinehart group reported the synthesis of deoxyxyboquinone (DNQ) possessing a 1,8-diazaanthraquinone scaffold *via* the oxidation of natural product

Department of Pharmaceutical Sciences, The Daniel K. Inouye College of Pharmacy, University of Hawaii at Hilo, 34 Rainbow Drive, Hilo, Hawaii, 96720, USA. E-mail: dianqing@hawaii.edu; Fax: +1 808 933 2974; Tel: +1 808 933 2960

† Electronic supplementary information (ESI) available: ¹H and ¹³C NMR, DEPTQ, HBMC, HSQC, COSY, HRMS, and HPLC of diazaquinomycins H (1) and J (2); ¹H NMR, DEPTQ spectra and HPLC chromatogram of isomeric mixture of 1 as well as 2; ¹H, ¹³C NMR spectra and HPLC chromatogram of 3a–c, 4a–b, 5a–b, and 7a–b; ¹H, ¹³C NMR spectra, HRMS, and HPLC chromatogram of 13; synthesis of 14 with a shorter branched side chain and its ¹H, ¹³C NMR spectra; ¹H, ¹³C NMR spectra comparison of 9-methyldecanoic acid and 4-methylpentanoic acid before and after being treated with conc. H₂SO₄; comparison of NMR spectra for pure diazaquinomycin H (1) with isomeric mixture of 1; and zoomed ¹H NMR regions of isomeric mixture of 2 with varying concentrations of CF₃CO₂D in CDCl₃. See DOI: 10.1039/c8ra09792e



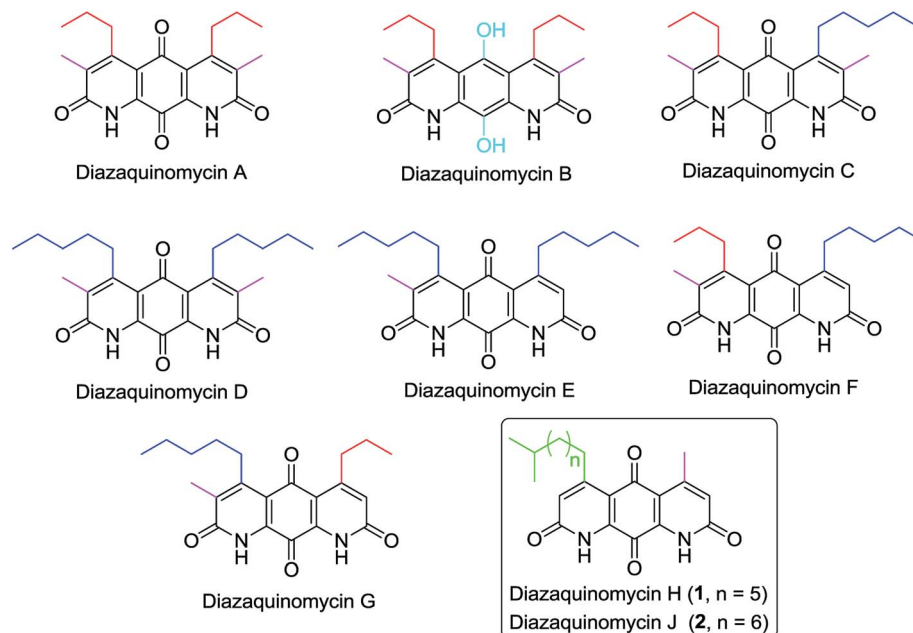
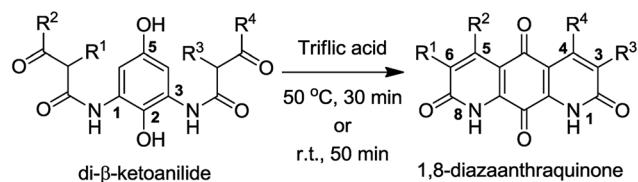


Fig. 1 Chemical structures of naturally occurring diazaquinomycins A–H and J.

deoxybomycin in hot nitric acid.¹³ Hergenrother and coworkers reported the synthesis of DNQ analogues starting from a monocyclic dipinacolborane.^{14,15} Their synthetic route used Suzuki–Miyaura cross coupling, followed by Buchwald–Hartwig amidation as key ring forming methodology. Final oxidation of the penultimate tricyclic methoxy intermediate to quinone was done using nitric acid¹⁵ or a sequence of hydrobromic acid, followed by salcomine in the presence of oxygen.¹⁴

We became interested in diazaquinomycins due to their unique 1,8-diazaanthraquinone scaffold and promising antitubercular and antibacterial activities.¹⁴ Recently, we expanded the scope of an underexplored double Knorr cyclization and demonstrated that this methodology can serve as a general and expedient route to symmetrical or unsymmetrical 1,8-diazaanthraquinone scaffolds (Scheme 1).¹⁶ We established that the 5-hydroxy substituent on the di- β -ketoanilide hydroquinone substrate is essential for successful double Knorr cyclization. On the other hand, sterically hindered di- β -ketoanilides with benzyl or isopropyl substitution on the side chains adjacent to β -carbonyl were poor substrates in this reaction.¹⁶ In this work, we extend this methodology to further develop an efficient and first chemical synthesis of diazaquinomycins H (1) and J (2) with a long iso-branched aliphatic side chain.



Scheme 1 Double Knorr cyclization of di- β -ketoanilides to 1,8-diazaanthraquinones.

Results and discussion

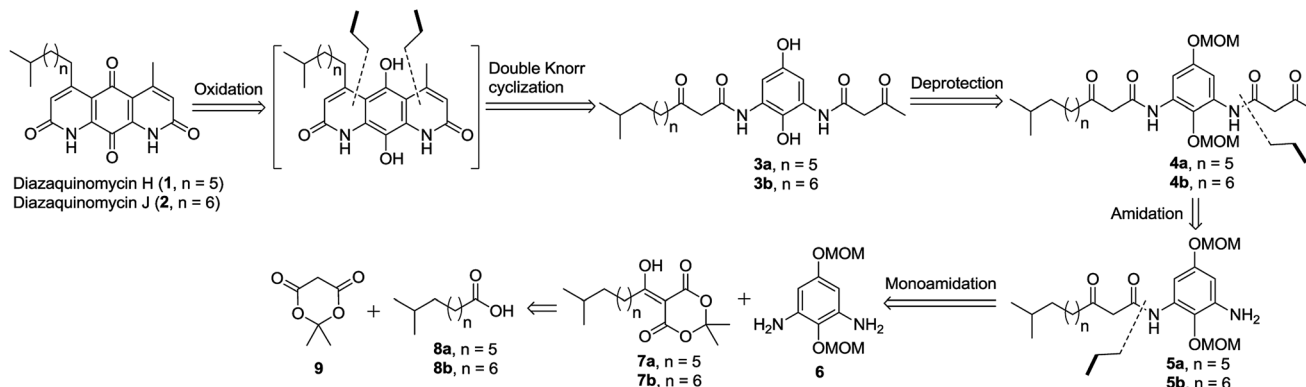
Retrosynthetic analysis

The retrosynthetic approach to diazaquinomycins H (1) and J (2) is outlined in Scheme 2. A key synthetic step would involve the double Knorr cyclization of di- β -ketoanilide hydroquinones **3a** and **3b** having their long iso-branched side chain preinstalled. Hydroquinones **3a** and **3b** will be obtained from **4a** and **4b** after removal of MOM protecting groups. The di- β -ketoanilides **4a** and **4b** will be obtained in two sequential steps, firstly amidation of diamine **6** with Meldrum's acid derivatives (**7a** or **7b**), then 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC) coupling with 3-oxobutanoic acid. The Meldrum's acid derivatives can be obtained through coupling commercially available iso-branched carboxylic acids **8a** and **8b** with Meldrum's acid (**9**). We envisioned that challenges may reside in the double Knorr cyclization of di- β -ketoanilide precursor **3a** or **3b** having a long, iso-branched aliphatic side chain, as well as the monoamidation of diamine intermediate **6**.¹⁶

Synthesis of key di- β -ketoanilide intermediates **3a–b** via selective amidation of **6** with Meldrum's acid derivatives

The synthesis of di- β -ketoanilide hydroquinone substrates **3a** and **3b** started from commercially available 1,4-diacetoxybenzene (**10**) and iso-branched 8-methylnonanoic acid (**8a**) or 9-methyldecanoic acid (**8b**) (Scheme 3). Diamine **6** was obtained in 4 synthetic steps from **10** following our reported synthesis.¹⁶ Meldrum's acid derivatives **7a** and **7b** were synthesized by coupling **8a** and **8b** with Meldrum's acid (**9**) using *N,N*-dicyclohexylcarbodiimide (DCC) and DMAP in DCM in 78% and 84% yields, respectively, following a modified literature protocol.¹⁷



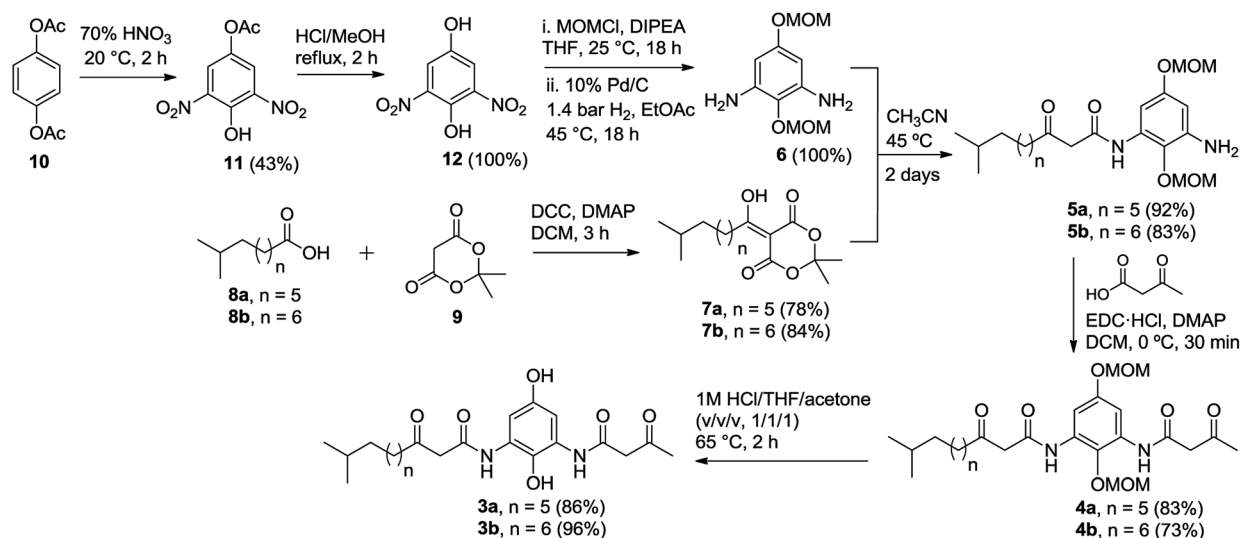


Scheme 2 Retrosynthetic analysis to diazaquinomycins H and J.

With diamine **6** and Meldrum's acid derivatives **7a** and **7b** in hand, we evaluated the synthesis towards mono- β -ketoanilides **5a** and **5b**. Our previous synthesis of a mono- β -ketoanilide analogue *via* EDC coupling of **6** with a β -ketoacid was successful, but resulted in poor isolated yield (25%).¹⁶ Therefore, an improved synthesis was developed by coupling diamine **6** with Meldrum's acid derivative **7a** or **7b** to form mono- β -ketoanilide **5a** or **5b**, respectively. Initially, **6** was treated with 0.7 molar equivalents of Meldrum's acid derivative at 45 °C in dry acetonitrile. After 2 days the Meldrum's acid derivative was consumed and HPLC analysis at 254 nm showed a mono- to di- β -ketoanilide ratio of 5 : 1, along with unreacted **6**. When 0.6 equivalents of Meldrum's acid derivative was used relative to **6**, the mono- to di- β -ketoanilide ratio increased to 9 : 1. This improvement greatly facilitated purification of mono- β -ketoanilides using flash column chromatography, affording **5a** and **5b** in 92% and 83% isolated yields, respectively. It is noteworthy to mention that the yields of **5a** and **5b** were calculated based on Meldrum's acid derivative **7a** or **7b**, which was used as the limiting reagent in order to favor the formation of desired mono- β -ketoanilide, as well as due to the valuable nature of carboxylic acid precursors **8a** and **8b** (approx. \$900 per gram).

Unreacted diamine **6** could also be recovered from column purification. To the best of our knowledge, this represents the first example reporting the synthesis of mono- β -ketoanilides directly from a diamine precursor using Meldrum's acid derivatives. In the literature, it was reported that β -ketoanilides or β -ketoamides can form when 5-substituted 2,2-dimethyl-1,3-dioxane-4,6-dione (acyl Meldrum's acid) derivatives were treated with anilines^{18,19} or amines,^{20,21} in solvent such as benzene^{18,19} or acetonitrile.^{21,22}

Once **5a–b** were synthesized, the second β -ketoanilide side chain was installed *via* EDC coupling of **5a** or **5b** with 3-oxobutanoic acid in dichloromethane at 0 °C. We found that a cold reaction temperature was necessary to slow the decomposition of 3-oxobutanoic acid (*via* decarboxylation), allowing for reaction completion. This afforded the unsymmetrical di- β -ketoanilides **4a** and **4b** in 83% and 73% yields, respectively, after flash column chromatography. Alternatively, crude product **5a** or **5b** containing unreacted **6** could be used directly in EDC coupling with excess 3-oxobutanoic acid since the resulting di- β -ketoanilide reaction mixture can be easily separated by flash column chromatography. Next, the MOM protecting groups were removed by treating **4a** or **4b** with 1 M HCl : THF : acetone

Scheme 3 Synthesis of unsymmetrical di- β -ketoanilide hydroquinones **3a–b**.

(1 : 1 : 1) at 65 °C for 2 h to afford **3a** or **3b** in 86% and 96% yields, respectively, after trituration with diethyl ether. The trituration step was introduced because in our hands the purification of **3a** or **3b** by flash column chromatography on silica gel led to poor isolated yields. This observation was in contrast to linear chain analogues, which could be isolated in good yields using flash column chromatography, suggesting the iso-branched functionality in **3a** or **3b** may play a major role in decomposition on silica gel.

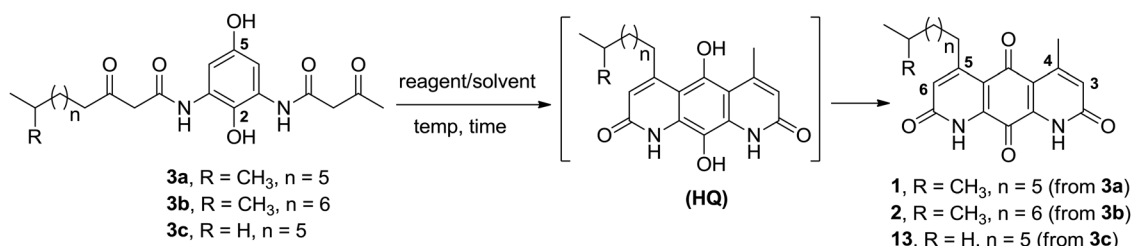
Double Knorr cyclization and optimization studies

Using our previously optimized conditions,¹⁶ **3a** or **3b** was subjected to double Knorr cyclization in an effort to synthesize diazaquinomycins H (**1**) and J (**2**). Very surprisingly, double Knorr cyclization of **3a** or **3b** in triflic acid at 50 °C resulted in a complex reaction mixture, and diazaquinomycins H (**1**) or J (**2**) could not be detected and isolated (Table 1, entries 1–2). This revealed that the longer ($n = 5, 6$) and/or iso-branched side chain significantly impacted this reaction outcome. Presumably, when cyclization rate is lowered, decomposition could occur through anilide hydrolysis,^{9,16} 4-pyrone formation or heterolytic cleavage of anilide.²³ Additionally, hydrocarbons have been shown to undergo isomerization^{24–26} under certain acidic conditions. Moreover, the length of aliphatic side chain can have an effect in these processes, with longer chains showing faster conversion rates.^{27,28} To probe the effect of branched functionality on reaction outcome as well as given the precious nature of **3a–b**, we next synthesized model substrate **3c**

with a linear side chain ($n = 5$) for comparison and further optimization studies (Table 1, entries 3–10). Interestingly, when the double Knorr cyclization was performed using **3c** in triflic acid, the reaction yielded a mixture of tricyclic hydroquinone (HQ) and **13** in 33% and 33% HPLC yields, respectively, along with unidentified decomposition byproducts (Table 1, entry 3). Nevertheless, the fact that linear chain analogue **3c** could be cyclized using triflic acid demonstrated that decomposition of **3a** and **3b** in triflic acid is probably accelerated by having an iso-branched side chain, as this is the only structural difference between these substrates. Furthermore, the observation of HQ intermediates in this study was striking because they were not previously detected in the double Knorr cyclization of shorter chain analogues.¹⁶ The slower rate of oxidation from HQ intermediates to desired quinone products was likely due to steric effect of the adjacent longer aliphatic chain at the 5 position.

Considering that sulfuric acid is much milder acid than triflic acid, we next investigated if sulfuric acid can perform better in the double Knorr cyclization of long-chain (*e.g.*, $n = 5$ or 6) di- β -ketoanilide substrates, while potentially minimizing the byproduct formation. In contrast to triflic acid, cyclization of **3c** using sulfuric acid at 50 °C for 30 min gave a clean reaction profile and afforded its corresponding HQ intermediate and **13** in 60% and 40% HPLC yields, respectively (Table 1, entry 4). Continued stirring at 50 °C for 3 h (Table 1, entry 5) and 18 h (Table 1, entry 6) could not complete the oxidation of HQ into **13**. At 110 °C in sulfuric acid, the oxidation of HQ into **13** was greatly accelerated, affording **13** in 91% isolated yield after 1 h

Table 1 Optimization of double Knorr cyclization of di- β -ketoanilides **3a–c**



Entry	Substrate	Reagent/solvent	Temp (°C)	Time (min)	HPLC yield (%)	
					HQ intermediate ^a	Product ^b
1	3a	TfOH	50	30	0	0
2	3b	TfOH	50	30	0	0
3	3c	TfOH	50	30	33	33
4	3c	H ₂ SO ₄	50	30	60	40
5	3c	H ₂ SO ₄	50	3 h	50	50
6	3c	H ₂ SO ₄	50	18 h	33	67
7	3c	H ₂ SO ₄	110	30	10	80
8	3c	H ₂ SO ₄	110	60	1	90 (91)
9	3c	H ₂ SO ₄	25	15	89	11
10 ^c	3c	H ₂ SO ₄	25	30	0	100 (99)
11 ^c	3a	H ₂ SO ₄	25	30	0	100 (99) ^d
12 ^c	3b	H ₂ SO ₄	25	30	0	100 (99) ^d

^a Hydroquinone (HQ) intermediates were observed using HPLC and MS and not isolated. ^b Isolated yield in parentheses. ^c The reaction was performed in sulfuric acid for 30 min, then diluted in methanol, and continued to stir for 2–3 h. ^d LC-MS and ¹³C NMR revealed that **1** or **2** contained its corresponding branched side chain isomer(s).



(Table 1, entries 7–8). The rate limiting step for this transformation was the oxidation of HQ intermediate to **13** since starting material **3c** was already consumed after 2 min based on HPLC monitoring. When the double Knorr cyclization of **3c** was performed at room temperature in sulfuric acid, an 8 : 1 mixture of HQ and **13** was formed after 15 min (Table 1, entry 9) and additional stirring at room temperature could not complete the oxidation into **13**. In the literature, structurally similar tricyclic hydroquinone scaffolds have been shown to spontaneously be oxidized to anthraquinone in the presence of oxygen.^{29–33} We re-analyzed some previous HPLC samples and found that the HQ intermediate can be oxidized into **13** within 2 h when diluted in methanol. Based on our finding, a milder and optimized protocol was developed, which involved stirring **3c** in sulfuric acid at room temperature for 30 min, followed by dilution in methanol and continued stirring open to air until the oxidation of HQ into **13** was complete (Table 1, entry 10). This provided **13** in 99% isolated yield as a red solid after removing methanol *in vacuo*, precipitation with water, and collecting solids by centrifugation.

Double Knorr cyclization of **3a** or **3b** in sulfuric acid leading to C-5 tail isomerization

Having established a milder double Knorr cyclization protocol, we proceeded to synthesize diazaquinomycins H (**1**) and J (**2**) from **3a** or **3b**, respectively. Di- β -ketoanilide **3a** or **3b** was treated in sulfuric acid at room temperature for 30 min, then diluted with methanol and stirred at room temperature to complete the oxidation of HQ intermediates to diazaquinomycins H (**1**) and J (**2**) (Table 1, entries 11 and 12). In both cases, reactions went to completion and red solids were obtained in corresponding to 99% isolated yields. Based on their analytical HPLC profiles and ¹H NMR spectra, the purity of **1** and **2** appeared to be excellent. However, additional carbon signals were observed in their ¹³C NMR spectra, especially in the C-5 aliphatic side chain region. Further LC-MS analysis revealed that both synthesized diazaquinomycins H (**1**) and J (**2**) samples also contained corresponding C-5 aliphatic side chain isomers. The representative HPLC chromatogram of the isomeric mixture of **1** showed a single peak (Fig. 2a). However, using a longer HPLC column under optimized mobile phase conditions, we were able to detect and partially separate **1** and its side chain isomers, which were confirmed by (+)-ESI mass spectrometry (Fig. 2b). To further investigate the isomerization rate during the double Knorr cyclization from **3a** to **1**, we closely monitored the reaction using LC-MS and the results are shown in Fig. 2c. For clarity and improving the peak resolution, the mass range on the MS detector was set to exclusively monitor $m/z = 383 \pm 2$. Our LC-MS reaction monitoring data showed the presence of the C-5 chain isomer even after 2 min, which continued to increase over the course of 60 min.

It was concluded that the C-5 distal iso-branched side chain of diazaquinomycins H (**1**) or J (**2**) must play a critical role in isomerization since our linear chain model compound **13** was obtained in pure form. In addition, we found that the length of the iso-branched alkyl chain also played an important role in

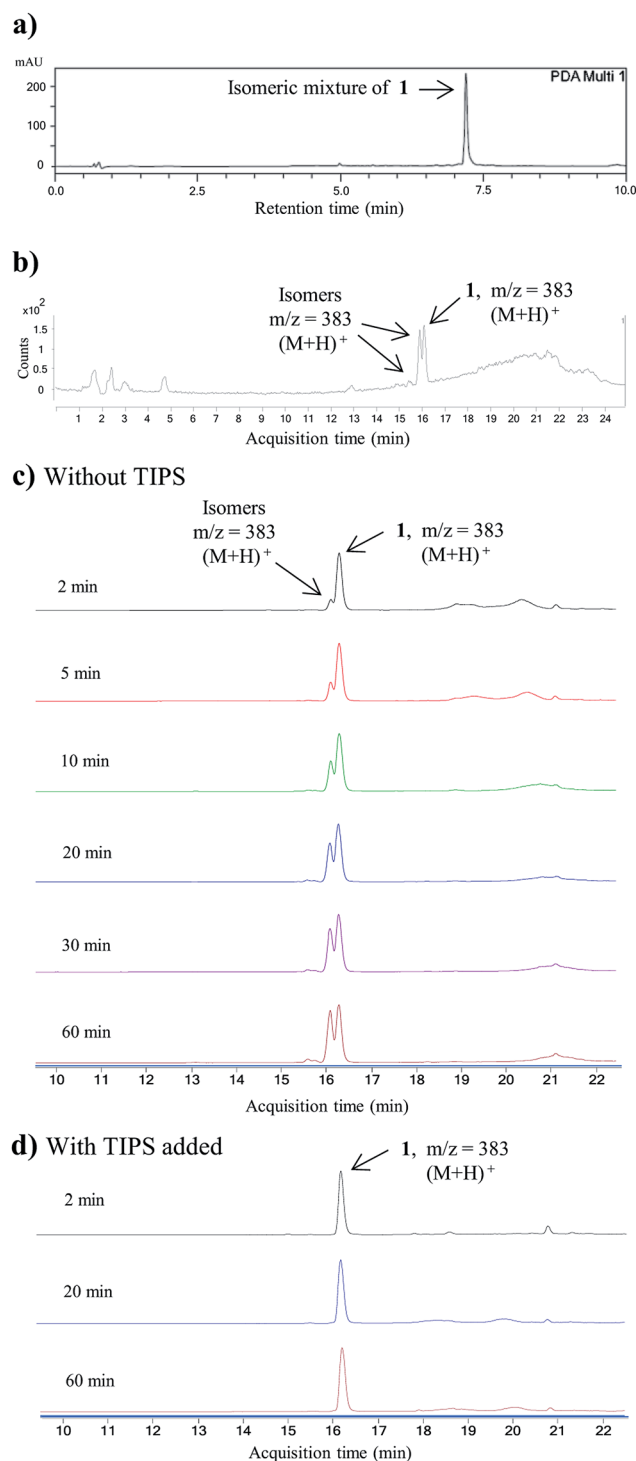
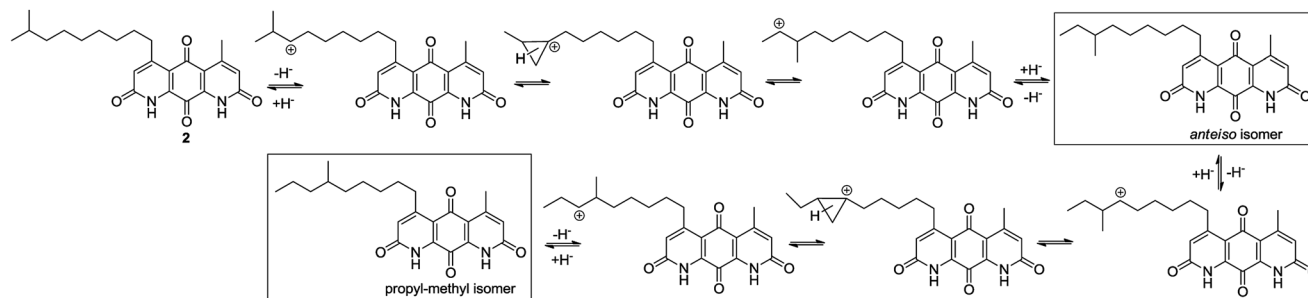


Fig. 2 (a) HPLC chromatogram for the isomeric mixture of **1**. (b) LC-MS chromatogram of the isomeric mixture of **1**. (c) LC-MS reaction monitoring of double Knorr cyclization of **3a** to **1**, which was performed in sulfuric acid only at room temperature. (d) LC-MS reaction monitoring of double Knorr cyclization of **3a** to **1** in sulfuric acid at room temperature with TIPS added. For clarity, LC-MS analyses shown in Fig. (c) and (d) were recorded with MS detector set to monitor $m/z = 383 \pm 2$.





Scheme 4 Proposed isomerization mechanism for diazaquinomycin J (2) in sulfuric acid.

this isomerization process, with a longer iso-branched alkyl chain being more prone to isomerization. For example, for comparison to **2**, we also synthesized an iso-branched four-carbon shorter alkyl chain analogue **14** from **3d** (Scheme S1, ESI[†]), which did not undergo isomerization during double Knorr cyclization in H₂SO₄. Moreover, in a side by side comparison, 9-methyldecanoic acid (**8b**) showed notable isomerization after being treated in conc. H₂SO₄ for 30 min at room temperature (Fig. S1, ESI[†]), whereas 4-methylpentanoic acid with a five-carbon shorter chain was stable under the same conditions even after 18 h (Fig. S2, ESI[†]).

The isomerization of iso-branched analogues may result from carbenium ion formation near the terminal end of the C-5 aliphatic chain in sulfuric acid, followed by hydride or methide migration.^{24,25} A proposed isomerization mechanism for representative diazaquinomycin J (**2**) is shown in Scheme 4. Unlike expected, the isomerization pathway probably proceeds *via* the initial formation of a secondary carbocation center, in which subsequent rearrangement results in anteiso and propyl-methyl isomers. In the case of initial formation of a tertiary carbocation center at the iso-branched tail, rearrangement may not take place due to its potential to generate a less favorable primary carbocation intermediate. Therefore, the secondary carbenium ion formation in the C-5 side chain is induced instead, upon exposure of **2** to concentrated sulfuric acid. Skeletal rearrangement of the carbenium ion then takes place *via* a non-classical protonated cyclopropane intermediate^{25,27,34,35} to form a rearranged carbenium ion. The newly formed carbenium ion further undergoes intermolecular hydride transfer with a reactant alkane to form an isomerized alkane (anteiso-isomer), while generating another carbenium ion and making the process catalytic. In a similar manner, the formed anteiso-isomer can also be rearranged to form additional isomers (*e.g.*, propyl-methyl isomer).

Adding TIPS effectively preventing isomerization in double Knorr cyclization of **3a** or **3b** in sulfuric acid

As shown in Fig. 2b, c and 3b, the double Knorr cyclization of **3a** and **3b** in sulfuric acid resulted in significant isomerization of the terminal C-5 iso-branched aliphatic chain. Attempts to prevent isomerization by lowering the reaction temperature or shortening the exposure time to sulfuric acid were unsuccessful. To overcome this problem, we envisioned that adding a carbocation scavenger such as triisopropylsilane (TIPS) in the

reaction may eliminate isomerization, since the isomerization process in acid presumably took place *via* carbenium ion formation.²⁵ This hypothesis led to an extremely successful outcome and adding TIPS in the reaction completely suppressed the isomerization of **1** or **2** in sulfuric acid, and no isomerization products were detected up to 60 min (Fig. 2d and 3c). In short, **3a** or **3b** was treated with H₂SO₄ and TIPS (5 : 1) at room temperature for 15 min, followed by stirring in methanol for 2 h to complete the oxidation to diazaquinomycin scaffolds (Scheme 5). This provided pure samples of diazaquinomycins H (**1**) and J (**2**) in 89% and 84% isolated yields, respectively.

A direct comparison of ¹H and ¹³C NMR spectra for isomeric mixture and pure diazaquinomycin J (**2**) was made and is shown in Fig. 3d–g. When only sulfuric acid was used in double Knorr cyclization, **2** and its isomers were produced. The doublet for the C-5 iso-branch (H-20/H-21) at 0.87 ppm was seen overlapping with a multiplet (Fig. 3d). In contrast, when TIPS was included in the double Knorr cyclization of **3b** in sulfuric acid, pure diazaquinomycin J (**2**) was obtained, and the multiplet was absent from the ¹H NMR spectrum with a clean doublet at 0.87 ppm observed (Fig. 3e). The ¹³C NMR spectrum for the isomeric mixture of **2** showed multiple sets of carbon signals, one of which corresponded to those of reported natural diazaquinomycin J (**2**)¹ (Fig. 3f). When the double Knorr cyclization was carried out in the presence of TIPS, the extra carbon peaks in the ¹³C NMR spectrum of isolated pure diazaquinomycin J (**2**) were absent (Fig. 3g). It should be noted, the additional carbon signals in the isomeric mixture (Fig. 3f) could not be completely assigned; however, characteristic chemical shift patterns in the up field region (<21 ppm) matched extremely well with those of anteiso and propyl-methyl isomers (Fig. 3a). The presence of carbon signals at 11 and 19 ppm strongly suggested the presence of an anteiso-type^{36–39} substitution pattern and the presence of 14, 19 and 20 ppm signals strongly suggested the presence of a propyl-methyl isomer form.^{36,39} In each case, the terminal methyl group migrated to the C-18 or C-17 position of the alkyl chain, respectively. The direct comparison of NMR spectra for the isomeric mixture of **1** and pure synthetic **1** is provided in ESI (Fig. S3[†]).

Comparison of ¹H and ¹³C NMR data of natural and synthetic diazaquinomycins H (**1**) and J (**2**)

Finally, the ¹H and ¹³C NMR spectroscopic data of synthesized diazaquinomycins H (**1**) and J (**2**) compared very favorably with



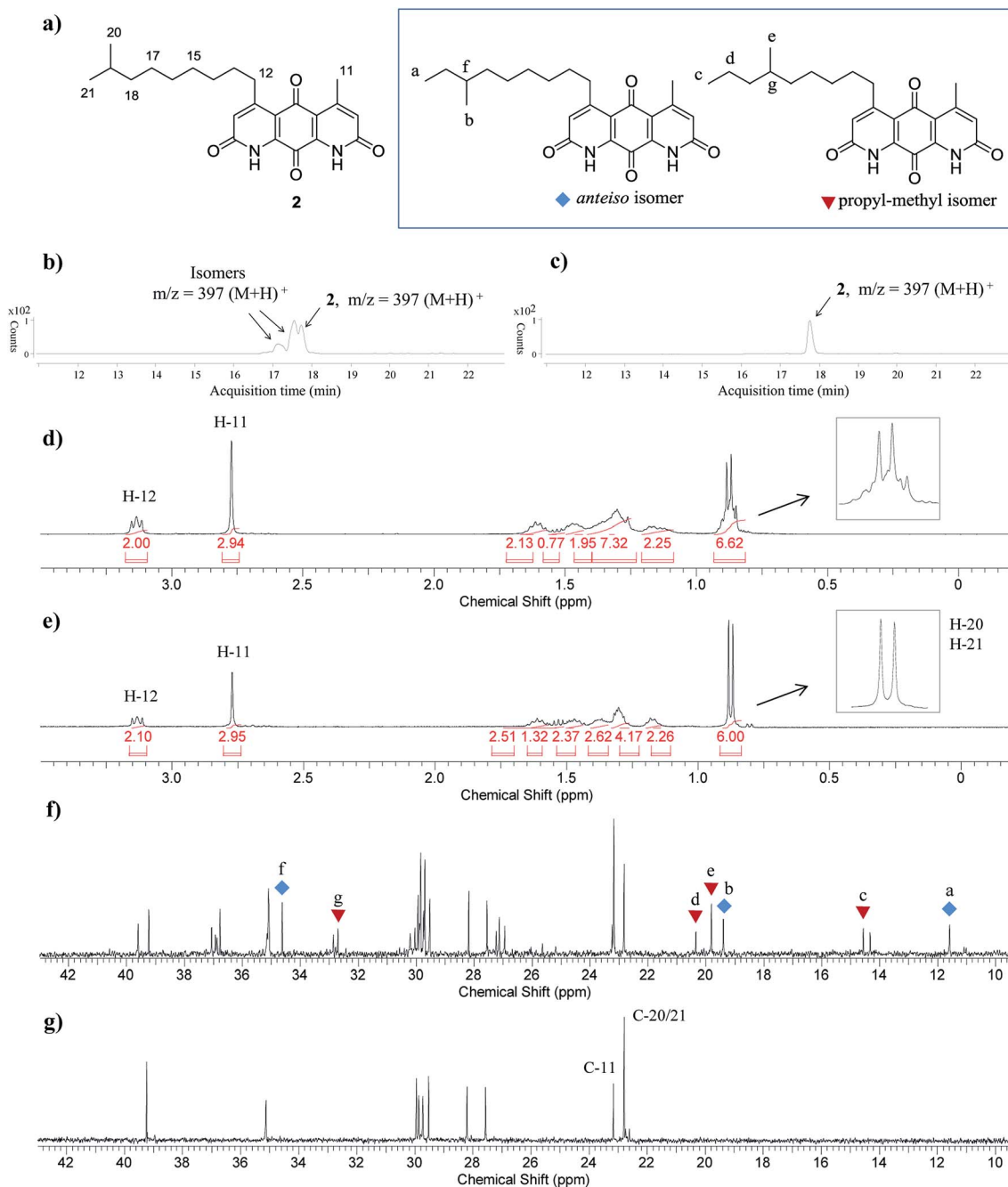


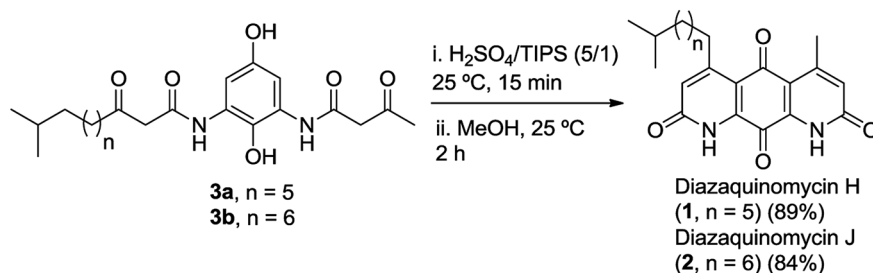
Fig. 3 (a) Structures of diazaquinomycin J (2) and proposed *anteiso* and propyl-methyl isomers. (b) LC-MS chromatogram of isomeric mixture of 2; (c) LC-MS chromatogram of pure synthetic 2; (d) zoomed ¹H NMR spectrum of isomeric mixture of 2; (e) zoomed ¹H NMR spectrum of pure synthetic 2; (f) zoomed ¹³C NMR spectrum of isomeric mixture of 2; (g) zoomed ¹³C NMR spectrum of pure synthetic 2. For clarity, LC-MS analyses shown in (b) and (c) were recorded with MS detector set to monitor $m/z = 397 \pm 2$.

reported data for the isolated natural products¹ (Tables 2 and 3). It should also be noted, NH resonance was not observed in our ¹H NMR spectra, presumably due to overlapping with CF₃CO₂D co-solvent or hydrogen/deuterium exchange (Table 2). In addition, for synthetic diazaquinomycin H (1), protons H-3 and H-6 resonate at 7.00 ppm, agreeing with reported value for 1¹. However, for the natural sample of diazaquinomycin J (2), protons H-3 and H-6 were reported at 6.93 ppm.¹ Given the volatile nature of added CF₃CO₂D co-solvent, we believe, this

slight up field shift is likely due to a difference in percentage of CF₃CO₂D in CDCl₃. In our hands, we observed notable variations to the chemical shifts of protons that reside close to the 1,8-anthraquinone core (H-3, H-6, H-11, and H-12) when the percentage of CF₃CO₂D varied from 0.4 to 2% (ESI, Fig. S4†).

As shown in Table 3, the ¹³C NMR data for synthetic diazaquinomycin H and J samples also compared favorably with reported data for the isolated natural products.¹ However, an inconsistency was noticed for the reported chemical shift of





Scheme 5 Optimized synthesis of diazaquinomycins H (1) and J (2).

amide carbons (C-2/C-7) for **2** that was up field shifted ($\Delta \approx 2.5$ and 2.6 ppm), when compared to both synthetic samples of **1** and **2** as well as natural sample of **1**. It is worthwhile mentioning, these reported chemical shift values overlap with the CF₃CO₂D co-solvent peaks, which could make their precise assignments challenging. In our hands, the signals for amide carbons (C-2/C-7) in **2** clearly resonate at 163.6 or 163.4 ppm and are consistent with those of other diazaquinomycin samples.¹⁶ It appears that the signal intensity for amide carbons (C-2/C-7) in diazaquinomycin J (**2**) is significantly lower in our DEPTQ spectrum relative to ¹³C NMR spectrum (ESI[†]). Notably, these signals were almost completely absent in our DEPTQ spectrum for diazaquinomycin J (**2**). In this light, it is possible that the amide carbons (C-2/C-7) were not observed in the reported DEPTQ spectrum of **2** and therefore were assumed to overlap with the CF₃CO₂D co-solvent. More extensive two dimensional HMBC, HSQC and COSY NMR studies (ESI[†]) of synthetic diazaquinomycins H (**1**) and J (**2**) further confirmed their structures and also matched well with reported spectra of isolated natural products.¹

Experimental section

General methods


Solvents and reagents were purchased from Sigma-Aldrich, Acros, or Fisher Scientific, and used without further purification. 8-Methylnonanoic acid (**8a**) and 9-methyldecanoic acid (**8b**) were purchased from BOC Sciences and Synnovator Inc, respectively. Reactions were monitored either by thin-layer chromatography (TLC) or by analytical high performance liquid chromatography (HPLC) employing a Shimadzu LC-20A series HPLC system. ¹H and ¹³C NMR spectra were recorded on a Bruker Avance III HD-400 (ultrashield) spectrometer (400 MHz and 100 MHz, respectively). All chemical shifts are given as δ value (ppm) and coupling constants, *J*, are reported in hertz (Hz). NMR solvent peaks were referenced as follows: (¹H NMR) CDCl₃: 7.27 ppm, DMSO-*d*₆: 2.50 ppm. (¹³C NMR) CDCl₃: 77.23 ppm, DMSO-*d*₆: 39.51 ppm. s = singlet; t = triplet; q = quartet; p = pentet; m = multiplet. High-resolution mass spectroscopy (HRMS) data was obtained using an Agilent 6530 Q-TOF LC/MS. Melting points were determined in open

Table 2 Comparison of ¹H NMR data of natural and synthetic diazaquinomycins H (1) and J (2)

Position	Natural ^a 1 δ (ppm)	Synthetic ^b 1 δ (ppm)	$\Delta\delta$ (ppm)	Natural ^a 2 δ (ppm)	Synthetic ^b 2 δ (ppm)	$\Delta\delta$ (ppm)
1, 8	8.05 (s)	H/D exc. ^c	—	8.03 (s)	H/D exc. ^c	—
3, 6	7.00 (s)	7.00 (s)	0.00	6.93 (s)	7.00 (s)	0.07
12	3.13 (t)	3.14 (t)	0.01	3.10 (t)	3.13 (t)	0.03
11	2.77 (s)	2.77 (s)	0.00	2.74 (s)	2.77 (s)	0.03
13	1.60 (p)	1.66–1.57 (m)	–0.01	1.59 (m)	1.66–1.57 (m)	0.02
18	1.53 (m)	1.57–1.50 (m)	0.00	1.17 (m)	1.21–1.13 (m)	0.00
14	1.47 (p)	1.50–1.43 (m)	0.00	1.45 (m)	1.50–1.42 (m)	0.01
15	1.35 (m)	1.40–1.27 (m)	–0.01	1.35 (m)	1.40–1.34 (m)	0.02
16	1.31 (m)	1.40–1.27 (m)	0.03	1.31 (m)	1.33–1.27 (m)	–0.01
17	1.19 (q)	1.22–1.15 (m)	0.00	1.28 (m)	1.33–1.27 (m)	0.02
19	0.87 (d)	0.88 (d)	0.01	1.52 (m)	1.57–1.50 (m)	0.01
20	0.87 (d)	0.88 (d)	0.01	0.86 (d)	0.87 (d)	0.01
21				0.86 (d)	0.87 (d)	0.01

^a 900 MHz (CDCl₃/1% CF₃CO₂D). ^b 400 MHz (CDCl₃/1% CF₃CO₂D). ^c Hydrogen/deuterium exchange.



Table 3 Comparison of ^{13}C NMR data of natural and synthetic diazaquinomycins H (1) and J (2)


Position	Natural ^a 1 δ (ppm)	Synthetic ^b 1 δ (ppm)	$\Delta\delta$ (ppm)	Natural ^a 2 δ (ppm)	Synthetic ^b 2 δ (ppm)	$\Delta\delta$ (ppm)
10	180.1	180.1	0.0	180.0	180.0	0.0
9	172.9	172.9	0.0	173.0	172.8	-0.2
2	163.2	163.7	0.5	161.0 ^c	163.6	2.6
7	163.0	163.5	0.5	160.9 ^c	163.4	2.5
5	160.1 ^c	160.8 ^c	0.7	160.2 ^c	161.2 ^c	1.0
4	155.6	156.6	1.0	154.6	156.6	2.0
9a	136.8	137.0	0.2	136.8	136.9	0.1
8a	136.3	136.5	0.2	136.2	136.4	0.2
3	128.6	128.3	-0.3	128.9	128.3	-0.6
6	127.6	127.3	-0.3	128.0	127.3	-0.7
4a	118.0	118.6	0.6	118.3	118.6	0.3
10a	117.5	118.1	0.6	117.8	118.1	0.3
17	39.1	39.2	0.1	27.5	27.6	0.1
12	35.0	35.1	0.1	35.0	35.1	0.1
13	29.7	29.9	0.2	29.9	29.9	0.0
14	29.7	29.8	0.1	29.8	29.9	0.1
15	29.7	29.8	0.1	29.7	29.7	0.0
18	28.1	28.2	0.1	39.1	39.2	0.1
16	27.4	27.4	0.0	29.6	29.5	-0.1
11	23.1	23.1	0.0	23.0	23.2	0.2
19	22.8	22.8	0.0	28.1	28.2	0.1
20	22.8	22.8	0.0	22.8	22.8	0.0
21				22.8	22.8	0.0

^a 226.2 MHz (CDCl₃/1% CF₃CO₂D). ^b 100 MHz (CDCl₃/2% CF₃CO₂D). ^c Overlapping with TFA co-solvent peak.

capillary tube using a Buchi B-540 melting point apparatus. Compounds were purified by flash column chromatography on silica gel using a Biotage Isolera One system. The purity of compounds was determined by analytical HPLC (Shimadzu LC-20A series) using a Gemini, 3 μm , C18, 110 \AA column (50 mm \times 4.6 mm, Phenomenex) and flow rate of 1 mL min⁻¹. Gradient conditions: solvent A (0.1% trifluoroacetic acid in water) and solvent B (acetonitrile): 0–2.0 min 100% A, 2.0–7.0 min 0–100% B (linear gradient), 7.0–8.0 min 100% B, UV detection at 254 nm and 220 nm. LC-MS data was obtained using an Agilent 1260 Infinity HPLC system coupled with an Agilent G6120A quadrupole mass spectrometer. HPLC separation was performed using a SunFire C18 column (Waters), 100 \AA , 3.5 μm , 2.1 mm \times 150 mm, and flow rate of 0.2 mL min⁻¹. Gradient conditions: solvent A (0.1% formic acid in water) and solvent B (0.1% formic acid in acetonitrile): 0–5.0 min 40% B, 5.0–15.0 min 40–100% B (linear gradient), 15.0–18.0 min 100% B. The mass spectrometer was run in (+)-ESI mode with a scan range (m/z) of 100–1700.

Double Knorr cyclization using H₂SO₄/TIPS; syntheses of diazaquinomycins H (1) and J (2)

4-Methyl-5-(7'-methyloctyl)-1H,8H-1,8-diazaanthracene-2,7,9,10-tetraone (diazaquinomycin H, 1). Sulfuric acid (0.60

mL) was added to a stirring suspension of 3a (12.0 mg, 0.0286 mmol) and triisopropylsilane (TIPS) (0.12 mL) at room temperature (rapid evolution of gas was observed). The dark brown solution was stirred at room temperature for 15 min, then diluted with water (5 mL) to precipitate a sticky beige solid (became hot). The hot suspension was stirred for 10 min, cooled to room temperature, and the water layer was removed *via* pipette. The beige solid was dissolved in MeOH (150 mL) to give an orange solution that was stirred at room temperature for 2 h open to air. Water (10 mL) was added and the solution was concentrated *in vacuo* using a rotary evaporator to precipitate a solid. The mixture was washed with diethyl ether (25 mL \times 3), and then centrifuged to collect the solid. The resulting solid was dried under vacuum to afford 1 (9.70 mg, 0.0254 mmol, 89%) as a red solid after trituration with EtOAc. R_f = 0.41 (DCM/MeOH, 9 : 1); mp > 200 °C (dec); ¹H NMR (400 MHz, CDCl₃/1% CF₃CO₂D): δ = 7.00 (s, 2H), 3.14 (t, J = 7.3 Hz, 2H), 2.77 (s, 3H), 1.66–1.57 (m, 2H), 1.57–1.50 (m, 1H), 1.50–1.43 (m, 2H), 1.40–1.27 (m, 4H), 1.22–1.15 (m, 2H), 0.88 (d, J = 6.7 Hz, 6H); ¹H NMR (400 MHz, CDCl₃/2% CF₃CO₂D): δ = 7.04 (s, 2H), 3.16 (t, J = 7.3 Hz, 2H), 2.79 (s, 3H), 1.66–1.57 (m, 2H), 1.57–1.50 (m, 1H), 1.50–1.44 (m, 2H), 1.40–1.27 (m, 4H), 1.22–1.15 (m, 2H), 0.88 (d, J = 6.7 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃/2% CF₃CO₂D): δ = 180.1, 172.9, 163.7, 163.5, 160.8, 156.6, 137.0, 136.5, 128.3,



127.3, 118.6, 118.1, 39.2, 35.1, 29.9, 29.8, 29.8, 28.2, 27.4, 23.1, 22.8; HRMS (ESI): m/z $[M + H]^+$ calcd for $C_{22}H_{27}N_2O_4^+$: 383.1965, found: 383.1969; HPLC purity: 100% (254 nm), 100% (220 nm), t_R : 7.18 min.

4-Methyl-5-(8'-methylonyl)-1H,8H-1,8-diazaanthracene-2,7,9,10-tetraone (diazaquinomycin J, 2). Following the optimized double Knorr cyclization procedure using H_2SO_4 /TIPS; compound **3b** (20.0 mg, 0.0461 mmol) afforded **2** (15.4 mg, 0.0389 mmol, 84%) as an orange-red solid. R_f = 0.45 (DCM/MeOH, 9 : 1); mp > 200 °C (dec); 1H NMR (400 MHz, $CDCl_3$ /1% CF_3CO_2D): δ = 7.00 (s, 2H), 3.13 (t, J = 7.3 Hz, 2H), 2.77 (s, 3H), 1.66–1.57 (m, 2H), 1.57–1.50 (m, 1H), 1.50–1.42 (m, 2H), 1.40–1.34 (m, 2H), 1.33–1.27 (m, 4H), 1.21–1.13 (m, 2H), 0.87 (d, J = 6.7 Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$ /2% CF_3CO_2D): δ = 7.02 (s, 2H), 3.15 (t, J = 7.3 Hz, 2H), 2.79 (s, 3H), 1.66–1.57 (m, 2H), 1.57–1.50 (m, 1H), 1.50–1.43 (m, 2H), 1.41–1.35 (m, 2H), 1.33–1.25 (m, 4H), 1.21–1.13 (m, 2H), 0.87 (d, J = 6.7 Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$ /2% CF_3CO_2D): δ = 180.0, 172.8, 163.6, 163.4, 161.2, 156.7, 136.9, 136.4, 128.3, 127.3, 118.6, 118.1, 39.2, 35.1, 30.0, 29.9, 29.7, 29.5, 28.2, 27.6, 23.2, 22.8; HRMS (ESI): m/z $[M + H]^+$ calcd for $C_{23}H_{29}N_2O_4^+$: 397.2122, found: 397.2126; HPLC purity: 100% (254 nm), 100% (220 nm), t_R : 7.40 min.

Typical procedure for removal of MOM protecting groups

***N*-(2,5-Dihydroxy-3-(3-oxobutanamido)phenyl)-10-methyl-3-oxoundecanamide (3a).** Compound **4a** (226 mg, 0.445 mmol) was dissolved in 1 M HCl/THF/acetone (1 : 1 : 1, 70 mL) and stirred at 65 °C for 2 h under nitrogen protection. The reaction was cooled, diluted with EtOAc (200 mL) and washed with water (100 mL \times 3). The organic layer was dried (anhydrous Na_2SO_4), filtered, and concentrated to give **3a** (160 mg, 0.381 mmol, 86%) as an off white solid after trituration with diethyl ether (5 mL). R_f = 0.11 (EtOAc/hexanes, 1 : 1); mp > 150 °C (dec); 1H NMR (400 MHz, DMSO- d_6): δ = 9.78 (s, 2H), 9.02 (s, 1H), 8.74 (s, 1H), 6.99 (d, J = 2.9 Hz, 1H), 6.97 (d, J = 2.9 Hz, 1H), 3.67 (s, 2H), 3.65 (s, 2H), 2.53 (t, J = 7.2 Hz, 2H), 2.19 (s, 3H), 1.53–1.43 (m, 3H), 1.26–1.20 (m, 6H), 1.17–1.09 (m, 2H), 0.84 (d, J = 6.6 Hz, 6H); ^{13}C NMR (100 MHz, DMSO- d_6): δ = 205.1, 203.1, 166.0, 165.9, 149.9, 131.1, 127.9, 104.8, 104.7, 51.5, 50.8, 42.2, 38.4, 30.2, 29.1, 28.5, 27.4, 26.6, 22.9, 22.5; HRMS (ESI): m/z $[M + H]^+$ calcd for $C_{22}H_{33}N_2O_6^+$: 421.2333, found: 421.2330; $[M + Na]^+$ calcd for $C_{22}H_{32}N_2NaO_6^+$: 443.2153, found: 443.2151; HPLC purity: 100% (254 nm), 100% (220 nm), t_R : 7.00 min.

***N*-(2,5-Dihydroxy-3-(3-oxobutanamido)phenyl)-11-methyl-3-oxododecanamide (3b).** Compound **4b** (88 mg, 0.168 mmol) provided **3b** (70.0 mg, 0.161 mmol, 96%) as an off white solid after trituration with diethyl ether (2 mL). R_f = 0.12 (EtOAc/hexanes, 1 : 1); mp > 150 °C (dec); 1H NMR (400 MHz, DMSO- d_6): δ = 9.77 (s, 2H), 9.00 (s, 1H), 8.75 (s, 1H), 6.99 (d, J = 2.8 Hz, 1H), 6.98 (d, J = 2.8 Hz, 1H), 3.67 (s, 2H), 3.65 (s, 2H), 2.53 (t, J = 7.2 Hz, 2H), 2.20 (s, 3H), 1.53–1.43 (m, 3H), 1.27–1.20 (m, 8H), 1.16–1.09 (m, 2H), 0.84 (d, J = 6.6 Hz, 6H); ^{13}C NMR (100 MHz, DMSO- d_6): δ = 205.1, 203.1, 166.00, 165.95, 149.9, 131.1, 127.9, 104.8, 104.7, 51.6, 50.8, 42.2, 38.5, 30.2, 29.2, 28.9, 28.5, 27.4, 26.8, 22.9, 22.5; HRMS (ESI): m/z $[M + H]^+$ calcd for $C_{23}H_{35}N_2O_6^+$: 435.2490, found: 435.2495; $[M + Na]^+$ calcd for $C_{23}H_{34}N_2NaO_6^+$:

457.2309, found: 457.2320; HPLC purity: 100% (254 nm), 100% (220 nm), t_R : 7.19 min.

***N*-(2,5-Dihydroxy-3-(3-oxobutanamido)phenyl)-3-oxoundecanamide (3c).** *N*-(2,5-Bis(methoxymethoxy)-3-(3-oxobutanamido)phenyl)-3-oxoundecanamide (90.0 mg, 0.182 mmol) afforded **3c** (30.0 mg, 0.0739 mmol, 41%) as an off white solid after flash column chromatography (silica gel, hexanes/EtOAc 9 : 1 to 1 : 2, v/v). R_f = 0.11 (EtOAc/hexanes, 1 : 1); mp 159–160 °C; 1H NMR (400 MHz, DMSO- d_6): δ = 9.77 (s, 2H), 9.01 (s, 1H), 8.74 (s, 1H), 6.99 (d, J = 2.9 Hz, 1H), 6.97 (d, J = 2.9 Hz, 1H), 3.67 (s, 2H), 3.65 (s, 2H), 2.53 (t, J = 7.2 Hz, 2H), 2.19 (s, 3H), 1.52–1.43 (m, 2H), 1.29–1.19 (m, 10H), 0.85 (t, J = 7.0 Hz, 3H); ^{13}C NMR (100 MHz, DMSO- d_6): δ = 205.1, 203.1, 166.0, 165.9, 149.9, 131.1, 127.87, 127.85, 104.77, 104.74, 51.5, 50.8, 42.2, 31.2, 30.2, 28.8, 28.6, 28.5, 22.9, 22.1, 13.9; HRMS (ESI): m/z $[M + H]^+$ calcd for $C_{21}H_{31}N_2O_6^+$: 407.2177, found: 407.2179; $[M + Na]^+$ calcd for $C_{21}H_{30}N_2NaO_6^+$: 429.1996, found: 429.1998; HPLC purity: 98.3% (254 nm), 98.5% (220 nm), t_R : 6.83 min.

Typical EDC coupling procedure for the synthesis of di- β -ketoanilides

***N*-(2,5-Bis(methoxymethoxy)-3-(3-oxobutanamido)phenyl)-10-methyl-3-oxoundecanamide (4a).** EDC·HCl (273 mg, 1.42 mmol) was added to a stirring solution of **5a** (200 mg, 0.472 mmol), 3-oxobutanoic acid (144 mg, 1.42 mmol) and DMAP (11.0 mg, 0.0902 mmol) in DCM (10 mL) at 0 °C. The solution was stirred at 0–5 °C for 30 min. The reaction was diluted with DCM (100 mL) and washed with water (50 mL \times 2). The organic layer was dried (anhyd. Na_2SO_4), filtered and concentrated to yield **4a** (200 mg, 0.394 mmol, 83%) as a sticky pale yellow oil after purification by flash column chromatography (silica gel, hexanes/EtOAc 9 : 1 to 3 : 2, v/v). R_f = 0.23 (EtOAc/hexanes, 1 : 1); 1H NMR (400 MHz, $CDCl_3$): δ = 9.42 (s, 1H), 9.37 (s, 1H), 7.79 (s, 2H), 5.15 (s, 2H), 5.07 (s, 2H), 3.65 (s, 3H), 3.58 (s, 2H), 3.55 (s, 2H), 3.47 (s, 3H), 2.58 (t, J = 7.5 Hz, 2H), 2.33 (s, 3H), 1.66–1.56 (m, 2H), 1.55–1.45 (m, 1H), 1.32–1.23 (m, 6H), 1.18–1.10 (m, 2H), 0.85 (d, J = 6.5 Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = 206.8, 204.1, 163.8, 163.7, 154.2, 132.4, 131.83, 131.78, 105.0, 104.9, 101.3, 94.9, 58.4, 56.4, 51.3, 50.4, 44.2, 39.1, 31.2, 29.8, 29.2, 28.1, 27.4, 23.6, 22.8; HRMS (ESI): m/z $[M + H]^+$ calcd for $C_{26}H_{41}N_2O_8^+$: 509.2857, found: 509.2846; $[M + Na]^+$ calcd for $C_{26}H_{40}N_2NaO_8^+$: 531.2677, found: 531.2673; HPLC purity: 99.6% (254 nm), 99.8% (220 nm), t_R : 7.40 min.

***N*-(2,5-Bis(methoxymethoxy)-3-(3-oxobutanamido)phenyl)-11-methyl-3-oxododecanamide(4b).** Compound **5b** (145 mg, 0.331 mmol) afforded **4b** (125 mg, 0.239 mmol, 73%) as a sticky pale yellow oil after flash column chromatography (silica gel, hexanes/EtOAc 9 : 1 to 3 : 2, v/v). R_f = 0.26 (EtOAc/hexanes, 1 : 1); 1H NMR (400 MHz, $CDCl_3$): δ = 9.44 (s, 1H), 9.39 (s, 1H), 7.80 (s, 2H), 5.15 (s, 2H), 5.07 (s, 2H), 3.66 (s, 3H), 3.59 (s, 2H), 3.56 (s, 2H), 3.47 (s, 3H), 2.59 (t, J = 7.5 Hz, 2H), 2.34 (s, 3H), 1.66–1.57 (m, 2H), 1.56–1.45 (m, 1H), 1.33–1.22 (m, 8H), 1.18–1.10 (m, 2H), 0.86 (d, J = 6.5 Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = 206.8, 204.2, 163.8, 163.7, 154.1, 132.4, 131.81, 131.77, 105.0, 104.9, 101.3, 94.9, 58.4, 56.4, 51.3, 50.4, 44.2, 39.2, 31.2, 29.9, 29.6, 29.2, 28.1, 27.5, 23.6, 22.8; HRMS (ESI): m/z $[M +$



H^+ calcd for $C_{27}H_{43}N_2O_8^+$: 523.3014, found: 523.3012; $[M + Na]^+$ calcd for $C_{27}H_{42}N_2NaO_8^+$: 545.2833, found: 545.2844; HPLC purity: 100% (254 nm), 100% (220 nm), t_R : 7.59 min.

Typical procedure for the synthesis of mono- β -ketoanilides

***N*-(3-Amino-2,5-bis(methoxymethoxy)phenyl)-10-methyl-3-oxoundecanamide (5a)**. To a solution of **6** (200 mg, 0.877 mmol) in dry CH_3CN (15 mL) was added **7a** (157 mg, 0.527 mmol) followed by stirring at 45 °C for 48 h under nitrogen protection. The solvent was removed *in vacuo* and the remaining residue was purified by flash column chromatography (silica gel, hexanes/EtOAc 9 : 1 to 4 : 1, v/v), affording **5a** (205 mg, 0.483 mmol, 92%) as a clear oil. R_f = 0.45 (EtOAc/Hexanes, 1 : 1); 1H NMR (400 MHz, $CDCl_3$): δ = 9.34 (s, 1H), 7.43 (d, J = 2.9 Hz, 1H), 6.23 (d, J = 2.9 Hz, 1H), 5.09 (s, 2H), 5.00 (s, 2H), 3.95 (br s, 2H), 3.62 (s, 3H), 3.53 (s, 2H), 3.45 (s, 3H), 2.57 (t, J = 7.5 Hz, 2H), 1.65–1.55 (m, 2H), 1.54–1.44 (m, 1H), 1.32–1.23 (m, 6H), 1.17–1.10 (m, 2H), 0.85 (d, J = 6.7 Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = 206.8, 163.6, 154.6, 140.4, 132.3, 130.2, 100.3, 99.9, 99.2, 94.8, 58.0, 56.2, 50.5, 44.2, 39.1, 29.7, 29.2, 28.1, 27.3, 23.6, 22.8; HRMS (ESI): m/z $[M + H]^+$ calcd for $C_{22}H_{37}N_2O_6^+$: 425.2646, found: 425.2645; $[M + Na]^+$ calcd for $C_{22}H_{36}N_2NaO_6^+$: 447.2466, found: 447.2460; HPLC purity: 100% (254 nm), 100% (220 nm), t_R : 7.30 min.

***N*-(3-Amino-2,5-bis(methoxymethoxy)phenyl)-11-methyl-3-oxododecanamide (5b)**. Used typical procedure; compound **6** (365 mg, 1.60 mmol) was coupled with **7b** (300 mg, 0.962 mmol) to provide **5b** (350 mg, 0.799 mmol, 83%) as a clear oil that solidified into an off white solid on standing. R_f = 0.46 (EtOAc/hexanes, 1 : 1); mp 64–65 °C; 1H NMR (400 MHz, $CDCl_3$): δ = 9.35 (s, 1H), 7.44 (d, J = 2.9 Hz, 1H), 6.24 (d, J = 2.9 Hz, 1H), 5.10 (s, 2H), 5.01 (s, 2H), 3.98 (br s, 2H), 3.63 (s, 3H), 3.54 (s, 2H), 3.46 (s, 3H), 2.58 (t, J = 7.5 Hz, 2H), 1.65–1.56 (m, 2H), 1.56–1.45 (m, 1H), 1.33–1.21 (m, 8H), 1.18–1.10 (m, 2H), 0.86 (d, J = 6.7 Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = 206.9, 163.6, 154.6, 140.4, 132.4, 130.3, 100.4, 100.0, 99.3, 94.9, 58.0, 56.2, 50.5, 44.2, 39.2, 29.9, 29.6, 29.2, 28.1, 27.5, 23.6, 22.8; HRMS (ESI): m/z $[M + H]^+$ calcd for $C_{23}H_{39}N_2O_6^+$: 439.2803, found: 439.2782; $[M + Na]^+$ calcd for $C_{23}H_{38}N_2NaO_6^+$: 461.2622, found: 461.2599; HPLC purity: 100% (254 nm), 100% (220 nm), t_R : 7.52 min.

5-(1-Hydroxy-8-methylonylidene)-2,2-dimethyl-1,3-dioxane-4,6-dione (7a). The following procedure was modified from a literature protocol.¹⁷ To a solution of 8-methylnonanoic acid (500 mg, 2.91 mmol), Meldrum's acid (502 mg, 3.49 mmol) and DMAP (426 mg, 3.49 mmol) in DCM (10 mL) at 5 °C was added DCC (719 mg, 3.49 mmol). The solution was allowed to warm to room temperature and stirred for 3 h under nitrogen atmosphere. The mixture was diluted with DCM (20 mL) and filtered. The filtrate was washed with 1 M HCl (20 mL \times 2), water (20 mL) and brine (20 mL). The organic layer was dried (Na_2SO_4), filtered and concentrated to afford **7a** (675 mg, 2.27 mmol, 78%) as a clear oil after flash column chromatography (silica gel, hexanes/EtOAc 9 : 1 to 6 : 1, v/v). R_f = 0.27 (EtOAc/hexanes, 1 : 4). 1H NMR (400 MHz, $CDCl_3$): δ = 15.29 (s, 1H), 3.09–3.03 (m, 2H), 1.73 (s, 6H), 1.72–1.65 (m, 2H), 1.57–1.46 (m, 1H), 1.45–1.36 (m, 2H), 1.33–1.24 (m, 4H), 1.18–1.11 (m, 2H), 0.86 (d, J =

6.6 Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = 198.5, 170.8, 160.4, 104.9, 91.4, 39.1, 35.9, 29.7, 29.6, 28.1, 27.3, 27.0, 26.4, 22.8; HRMS (ESI): m/z $[M - H]^-$ calcd for $C_{16}H_{25}O_5^-$: 297.1707, found: 297.1709; HPLC purity: 100% (254 nm), 100% (220 nm), t_R : 7.99 min.

5-(1-Hydroxy-9-methyldecylidene)-2,2-dimethyl-1,3-dioxane-4,6-dione (7b). The following procedure was modified from a literature protocol.¹⁷ To a solution of 9-methyldecanoic acid (300 mg, 1.61 mmol), Meldrum's acid (278 mg, 1.93 mmol) and DMAP (235 mg, 1.93 mmol) in DCM (8 mL) at 5 °C was added DCC (398 mg, 1.93 mmol). The solution was allowed to warm to room temperature and stirred for 3 h under nitrogen atmosphere. The mixture was diluted with DCM (20 mL) and filtered. The filtrate was washed with 1 M HCl (20 mL \times 2), water (20 mL) and brine (20 mL). The organic layer was dried (Na_2SO_4), filtered and concentrated to afford **7b** (420 mg, 1.35 mmol, 84%) as a clear oil after flash column chromatography (silica gel, hexanes/EtOAc 9 : 1 to 6 : 1, v/v). R_f = 0.28 (EtOAc/hexanes, 1 : 4). 1H NMR (400 MHz, $CDCl_3$): δ = 15.29 (s, 1H), 3.09–3.03 (m, 2H), 1.73 (s, 6H), 1.72–1.64 (m, 2H), 1.55–1.45 (m, 1H), 1.44–1.35 (m, 2H), 1.35–1.29 (m, 2H), 1.29–1.23 (m, 4H), 1.18–1.10 (m, 2H), 0.86 (d, J = 6.6 Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = 198.5, 170.8, 160.4, 104.9, 91.4, 39.2, 35.9, 29.9, 29.6, 29.5, 28.1, 27.5, 27.0, 26.4, 22.8; HRMS (ESI): m/z $[M - H]^-$ calcd for $C_{17}H_{27}O_5^-$: 311.1864, found: 311.1860; HPLC purity: 100% (254 nm), 100% (220 nm), t_R : 8.16 min.

4-Methyl-5-octyl-1H,8H-1,8-diazaanthracene-2,7,9,10-tetraone (13). A solution of **3c** (21.0 mg, 0.0517 mmol) in sulfuric acid (5 mL) was stirred at room temperature for 30 min. The reaction was chilled using an ice-water bath followed by careful addition of MeOH (20 mL) over a period of 10 min to keep internal temperature < 30 °C. The resulting solution was allowed to warm to room temperature and stirred for 3 h. Water (15 mL) was added and MeOH was removed *in vacuo* using a rotary evaporator (water bath temperature < 35 °C) to precipitate a solid. The solid was collected by centrifugation and washed with cold water (2 \times 5 mL) using the centrifuge, carefully removing the supernatant layer between each washing step. The solid was triturated with EtOAc (5 mL) and dried under vacuum to afford **13** (18.8 mg, 0.0510 mmol, 99%) as a red solid. R_f = 0.41 (DCM/MeOH, 9 : 1); mp > 200 °C (dec); 1H NMR (400 MHz, $CDCl_3$ /1% CF_3CO_2D): δ = 7.01 (s, 2H), 3.14 (t, J = 7.3 Hz, 2H), 2.78 (s, 3H), 1.65–1.57 (m, 2H), 1.51–1.43 (m, 2H), 1.41–1.28 (m, 8H), 0.90 (t, J = 7.0 Hz, 3H); 1H NMR (400 MHz, $CDCl_3$ /2% CF_3CO_2D): δ = 7.09 (s, 2H), 3.15 (t, J = 7.3 Hz, 2H), 2.79 (s, 3H), 1.66–1.57 (m, 2H), 1.52–1.43 (m, 2H), 1.41–1.28 (m, 8H), 0.90 (t, J = 7.0 Hz, 3H); ^{13}C NMR (100 MHz, $CDCl_3$ /2% CF_3CO_2D): δ = 180.0, 172.8, 164.0, 163.9, 162.1, 157.5, 137.0, 136.5, 128.2, 127.2, 119.0, 118.5, 35.3, 32.1, 30.0, 29.8, 29.5, 29.4, 23.1, 22.9, 14.1; HRMS (ESI): m/z $[M + H]^+$ calcd for $C_{21}H_{25}N_2O_4^+$: 369.1809, found: 369.1816; HPLC purity: 99.2% (254 nm), 99.2% (220 nm), t_R : 7.03 min.

Conclusions

In summary, the first total synthesis of anti-tuberculosis diazaquinomycins **H** (1) and **J** (2) has been accomplished *via*



selective amidation of diamine **6** with Meldrum's acid derivatives, subsequent EDC coupling with 3-oxobutanoic acid, and double Knorr cyclization in the presence of TIPS. Adding TIPS during the double Knorr cyclization was crucial in preventing isomerization of the C-5 iso-branched tail in sulfuric acid. Diazaquinomycins H (**1**) and J (**2**) were obtained in 8 steps from commercially available starting materials in 25% and 21% overall yields, respectively. In contrast to triflic acid, sulfuric acid gave much cleaner double Knorr cyclization product for long-chain ($n = 5$ or 6) di- β -ketoanilide substrates. In addition to a terminal isopropyl functionality, we found that the length of the iso-branched alkyl chain also played an important role in the isomerization process in sulfuric acid, with a longer iso-branched alkyl chain being more prone to isomerization. We observed tricyclic hydroquinone intermediates by HPLC and mass spectrometry, indicating that oxidation to diazaquinomycin scaffold was notably slower for long-chain analogues compared to short-chain analogues. Moreover, oxidation of the hydroquinone intermediates to diazaquinomycins was greatly accelerated by simply stirring in methanol open to air. The spectroscopic data for synthesized diazaquinomycins H (**1**) and J (**2**) compared very well with reported data for the natural products, with minor discrepancies noted.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by Hawaii Community Foundation LE'AHU FUND for Pulmonary Research (16ADVC-78728) and UHH DKICP RTRF fund. We thank Justin Reinicke for his expert support for developing LC-MS method and acquiring low resolution LC-MS and HRMS data. We also thank Skyla Lee for her assistance with the synthesis of intermediate compound **6**.

Notes and references

- M. W. Mullowney, C. H. Hwang, A. G. Newsome, X. Wei, U. Tanouye, B. Wan, S. Carlson, N. J. Barranis, E. Ó hAinmhire, W.-L. Chen, K. Krishnamoorthy, J. White, R. Blair, H. Lee, J. E. Burdette, P. K. Rathod, T. Parish, S. Cho, S. G. Franzblau and B. T. Murphy, *ACS Infect. Dis.*, 2015, **1**, 168–174.
- M. W. Mullowney, E. Ó hAinmhire, A. Shaikh, X. Wei, U. Tanouye, B. D. Santarsiero, J. E. Burdette and B. T. Murphy, *Mar. Drugs*, 2014, **12**, 3574–3586.
- R. P. Maskey, I. Grün-Wollny and H. Laatsch, *Nat. Prod. Res.*, 2005, **19**, 137–142.
- S. Ōmura, Y. Iwai, K. Hinotozawa, H. Tanaka, Y. Takahashi and A. Nakagawa, *J. Antibiot.*, 1982, **35**, 1425–1429.
- S. Ōmura, A. Nakagawa, H. Aoyama, K. Hinotozawa and H. Sano, *Tetrahedron Lett.*, 1983, **24**, 3643–3646.
- S. Ōmura, M. Murata, K. Kimura, S. Matsukura, T. Nishihara and H. Tanaka, *J. Antibiot.*, 1985, **38**, 1016–1024.
- M. Murata, T. Miyasaka, H. Tanaka and S. Ōmura, *J. Antibiot.*, 1985, **38**, 1025–1033.
- K. Tsuzuki, T. Yokozuka, M. Murata, H. Tanaka and S. Ōmura, *J. Antibiot.*, 1989, **42**, 727–737.
- T. R. Kelly, J. A. Field and Q. Li, *Tetrahedron Lett.*, 1988, **29**, 3545–3546.
- H. Lee and W. K. Anderson, *Tetrahedron Lett.*, 1990, **31**, 4405–4408.
- J. M. Pérez, P. López-Alvarado, C. Avendaño and J. C. Menéndez, *Tetrahedron Lett.*, 1998, **39**, 673–676.
- J. M. Pérez, P. López-Alvarado, E. Pascual-Alfonso, C. Avendaño and J. C. Menéndez, *Tetrahedron*, 2000, **56**, 4575–4583.
- K. L. Rinehart Jr and H. B. Renfroe, *J. Am. Chem. Soc.*, 1961, **83**, 3729–3731.
- E. I. Parkinson, J. S. Bair, M. Cismesia and P. J. Hergenrother, *ACS Chem. Biol.*, 2013, **8**, 2173–2183.
- J. S. Bair, R. Palchaudhuri and P. J. Hergenrother, *J. Am. Chem. Soc.*, 2010, **132**, 5469–5478.
- A. M. Prior and D. Sun, *Synthesis*, 2018, **50**, 859–871.
- T. Knoth, K. Warburg, C. Katzka, A. Rai, A. Wolf, A. Brockmeyer, P. Janning, T. F. Reubold, S. Eschenburg, D. J. Manstein, K. Hübel, M. Kaiser and H. Waldmann, *Angew. Chem., Int. Ed.*, 2009, **48**, 7240–7245.
- M. T. Huggins, P. S. Barber, D. Florian and W. Howton, *Synth. Commun.*, 2008, **38**, 4226–4239.
- C. S. Pak, H. C. Yang and E. B. Choi, *Synthesis*, 1992, **1992**, 1213–1214.
- H. Kikuchi, K. Sasaki, J. Sekiya, Y. Maeda, A. Amagai, Y. Kubohara and Y. Oshima, *Bioorg. Med. Chem.*, 2004, **12**, 3203–3214.
- M. Syrpas, E. Ruysbergh, L. Blommaert, B. Vanelslander, K. Sabbe, W. Vyverman, N. De Kimpe and S. Mangelinckx, *Mar. Drugs*, 2014, **12**, 352–367.
- C. Ma, X. Li, X. Liang, K. Jin, J. Cao and W. Xu, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 4948–4952.
- R. M. Forbis and K. L. Rinehart Jr, *J. Am. Chem. Soc.*, 1973, **95**, 5003–5013.
- D. J. am Ende and L. F. Albright, *Ind. Eng. Chem. Res.*, 1994, **33**, 840–848.
- Y. Ono, *Catal. Today*, 2003, **81**, 3–16.
- A. K. Roebuck and B. L. Evering, *J. Am. Chem. Soc.*, 1953, **75**, 1631–1635.
- S. T. Sie, *Ind. Eng. Chem. Res.*, 1993, **32**, 397–402.
- J. Weitkamp, ACS Symposium Series, in *Hydrocracking and hydrotreating*, ed. J. W. Ward and S. A. Qader, vol. 20, American Chemical Society, Washington, DC, 1975, pp. 1–27, DOI: 10.1021/bk-1975-0020.
- A. D. Broadbent and J. M. Stewart, *Can. J. Chem.*, 1983, **61**, 1965–1969.
- K. Krohn, G. Carlson, H. Köhle, H. Hussain and I. R. Green, *ARKIVOC*, 2008, **xiv**, 216–233.
- L. Kurc, V. Vopravil and L. Červený, *Res. Chem. Intermed.*, 2001, **27**, 249–258.
- Y. Ren, Z. Mi, S. Cheng, L. Wang and Y. Wang, *React. Kinet. Catal. Lett.*, 2004, **83**, 71–77.



- 33 P. Shaul, M. Frenkel, E. B. Goldstein, L. Mittelman, A. Grunwald, Y. Ebenstein, I. Tsarfaty and M. Fridman, *ACS Med. Chem. Lett.*, 2013, **4**, 323–328.
- 34 G. Sastre, A. Chica and A. Corma, *J. Catal.*, 2000, **195**, 227–236.
- 35 A. Corma and A. V. Orchillés, *Microporous Mesoporous Mater.*, 2000, **35–36**, 21–30.
- 36 E. Hedenström, B.-V. Nguyen and L. A. Silks III, *Tetrahedron: Asymmetry*, 2002, **13**, 835–844.
- 37 H. Chiba, H. Agematu, K. Dobashi and T. Yoshioka, *J. Antibiot.*, 1999, **52**, 700–709.
- 38 K. Yamada, N. Wada, H. Onaka, R. Matsubara, R. Isobe, M. Inagaki and R. Higuchi, *Chem. Pharm. Bull.*, 2005, **53**, 788–791.
- 39 F. D. Gunstone, *Chem. Phys. Lipids*, 1993, **65**, 155–163.

