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# Development of hollow ferrogadolinium nanonetworks for dual-modal MRI guided cancer chemotherapy†

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The development of hollow ferrogadolinium nanonetworks has not been reported for nanomedicine application until now. In this study, we developed a hollow and porous ferrogadolinium nanonetwork structure using the one-pot solvothermal method. This nanoparticle could be simultaneously used as a  $T_1$  and  $T_2$  dual-modal magnetic resonance imaging (MRI) contrast agent. In addition, the hollow lumen and abundant pores of the nanonetworks maximized the loading capacity and conferred the nanoplateforms for suitable anticancer drug loading capacity. Using these nanonetworks, MRI and anticancer experiments were conducted *in vitro* and satisfactory dual-modal MRI and cancer chemotherapy results were obtained. Therefore, the nanonetworks with dual-modal MRI and drug loading abilities effectively complement the ferrogadolinium composites' library and hold great promise in nanomedicine for simultaneous cancer diagnosis and chemotherapy.

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## 1. Introduction

Currently, magnetic resonance imaging (MRI) is one of the most powerful non-invasive imaging modalities used in the diagnosis of different cancers.<sup>1–3</sup> However, more than half of the diseased patients require the use of an MRI contrast agent (CA) to enhance the contrast of the targeted site within the surrounding tissues and improve the resolution of the MRI images.<sup>4,5</sup> There are two types of CAs commonly used: Gd-based compounds are often employed as  $T_1$  CAs and Fe-based magnetic nanomaterials are often utilized as  $T_2$  CAs.<sup>6–9</sup> However, the inherent limitations of single mode CAs are often criticized, thus rendering dual-modal MRI CAs as more acceptable because the synergistic combination of  $T_1$ - and  $T_2$ -weighted MRI can provide more comprehensive imaging information, increase the diagnostic accuracy, and improve the overall prognosis of patients.<sup>10,11</sup>

In the study of dual-modal MRI CAs, most attention has been paid to ferrogadolinium nanocomposites because Gd- and Fe-based compounds are commonly used as  $T_1$  and  $T_2$  CAs,

respectively.<sup>12,13</sup> It is worth noting that as  $T_1$  CAs are directly in contact with the magnetic core and located within the magnetic field generated by the magnetic core and opposite to the external magnetic field,  $T_2$  CAs can severely perturb  $T_1$  imaging.<sup>6,7,14</sup> Hence,  $T_1$  CAs should be on the inside or away from the outside of the magnetic core. However, many previous studies have failed to observe this.<sup>15–17</sup> Many interesting nanostructures of ferrogadolinium composites have been reported to reduce the interference of  $T_2$  CAs for  $T_1$  imaging, including yolk-like structures, barbell-like structures, and others.<sup>6,18</sup> However, their synthesis process is multi-step and very complicated. Furthermore, many of these nanostructures are limited in their ability to load drugs because of their non-porous surface structures.<sup>6,7,15–23</sup>

Up until now, a hollow and porous ferrogadolinium nanonetwork (HPFN) has not been reported. In this case,  $Gd^{3+}$  is located inside the HPFN and has a parallel spin ordering with the same direction as the magnetic field induced by the Fe-based material, which will not disturb the  $T_1$  imaging, thus providing effective  $T_1$  imaging.<sup>7,10,24</sup> Moreover, the porous and hollow structure is beneficial for the water molecules contacting  $T_1$  and  $T_2$  CAs as well as for efficient drug loading.

In this study, we developed a HPFN using the one-pot solvothermal method. This nanoparticle simultaneously and efficiently exerted  $T_1$  and  $T_2$  contrast ability. In addition, the hollow lumen and abundant mesopores conferred the nanonetworks a high specific surface area, which was critical for the abundant loading of anticancer drugs. Cisplatin, a commonly used anticancer drug,<sup>25,26</sup> was selected as the model drug and loaded into the nanonetworks. Using these nanonetworks, MRI and

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anticancer experiments were conducted *in vitro* and satisfactory dual-modal MRI and cancer chemotherapy results were obtained. Therefore, the easily synthesized nanonetworks with dual-modal MRI and drug loading abilities effectively complement the ferrogadolinium nanocomposites' library and holds great promise in nanomedicine for simultaneous cancer diagnosis and chemotherapy.

## 2. Materials and methods

### 2.1. Chemicals

All obtained chemicals were of analytical grade and used without further purification.  $\text{Fe}(\text{acac})_3$ ,  $\text{Gd}(\text{acac})_3$ , 1,6-hexanediamine,  $\text{CH}_3\text{COONa}$ , cisplatin, ethylene glycol, and fluorescein isothiocyanate (FITC) were purchased from Aladdin Co. (Shanghai, China). All other chemicals were provided by the Sinopharm Co. (Shanghai, China).

### 2.2. Preparation of amine-functionalized HPFN

$\text{Fe}(\text{acac})_3$  (0.5 g), and  $\text{Gd}(\text{acac})_3$  (0.5 g) were added into ethylene glycol (30 mL) and vigorously stirred at 80 °C for 1 h. Then, 1,6-hexanediamine (3.6 g) was added into the solution and stirred at 80 °C for an additional 0.5 h.  $\text{CH}_3\text{COONa}$  (4 g) was then added into the solution and reacted for 0.5 h. The mixture was sealed in a Teflon-lined stainless-steel autoclave and heated at 200 °C for 24 h. The resulting product was precipitated by centrifugation, briefly rinsed with water and ethanol three times to

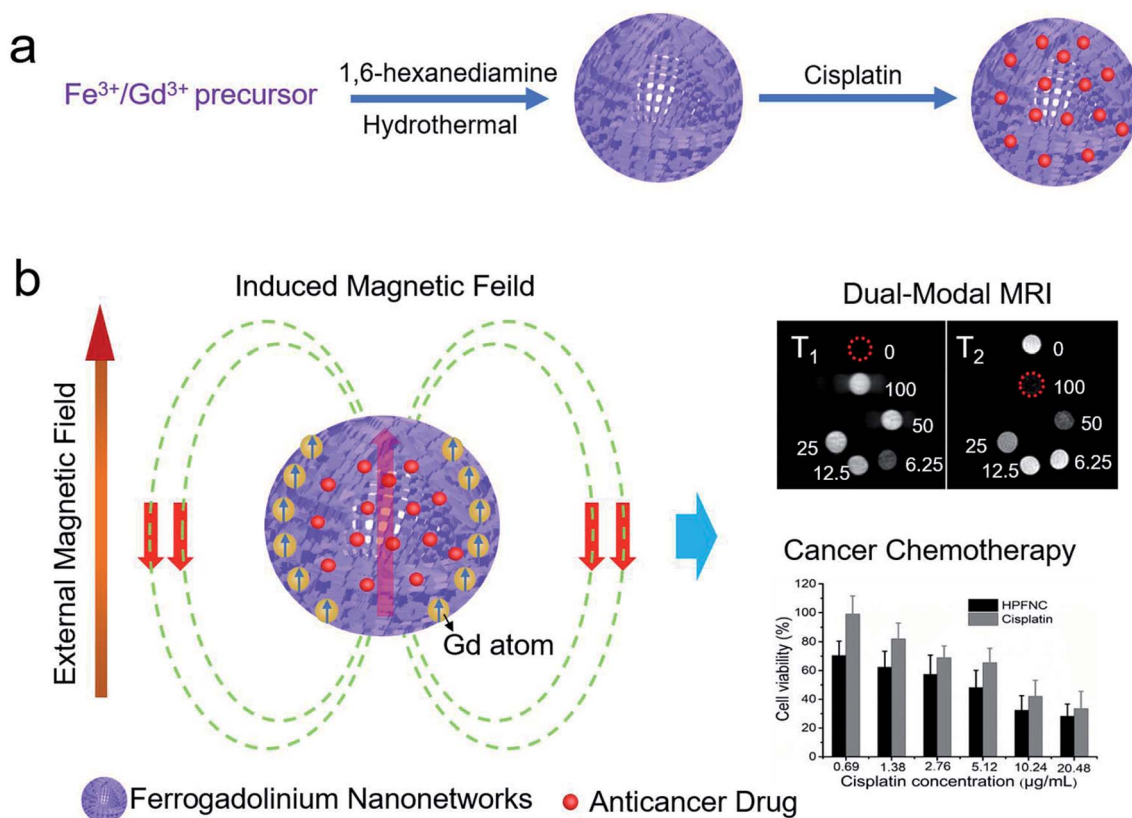
effectively remove the solvent and excess 1,6-hexanediamine, and then dried at 60 °C before characterization and application.

### 2.3. Loading cisplatin into HPFN

HPFN (30 mg) were dispersed into 10 mL of cisplatin solution (2 mg  $\text{mL}^{-1}$ ) under stirring overnight. Cisplatin-loaded HPFN nanonetworks (designated as HPFNC) were centrifuged and the supernatant was detected by inductively coupled plasma optical emission spectrometry (ICP-OES) to quantify the unloaded cisplatin. The loading capacity of cisplatin could be calculated using the equation  $\text{LC} (\%) = [2 \text{ mg mL}^{-1} - c(\text{cisplatin})_{\text{sup}}] \times V / m_{\text{nanoplatform}} \times 100\%$ . In the *in vitro* experiments, the HPFN and HPFNC were re-suspended in distilled water and filtered using a 0.8  $\mu\text{m}$ -membrane filter.

### 2.4. Observing the cellular uptake of HPFN *in vitro*

Confocal laser scanning microscopy (CLSM) was used to define the intracellular localization of fluorescein isothiocyanate (FITC)-labeled HPFN (designated as HPFN-FITC) in HeLa cells. To form HPFN-FITC conjugates, FITC was loaded into the HPFN. HeLa cells seeded in 60 mm-polystyrene plates ( $5 \times 10^5$  cells per dish) were treated with HPFN-FITC (24  $\mu\text{g mL}^{-1}$ ) for 1 h, 3 h, and 6 h, and then stained with NucRed Live 647 ReadyProbes Reagent (Invitrogen, USA) for 15 min at 37 °C in the dark. The cells were washed twice with PBS (pH 7.4) and fixed with 4% paraformaldehyde for 30 min. The cells were then



**Scheme 1** Schematic of (a) the synthesis for ferrogadolinium nanonetworks and (b) their applications in dual-modal MRI guided cancer chemotherapy.



visualized using a Leica SP8 confocal microscope with 100/63 $\times$  oil-immersion objective lens.

### 2.5. Measuring MRI *in vitro*

The longitudinal ( $T_1$ ) and transverse ( $T_2$ ) relaxation time of HPFN was detected using a 9.4 T MRI scanner. The  $T_1$  and  $T_2$  relaxation rates were measured using the inversion recovery method and a multi-echo spin echo sequence, respectively. The imaging parameters were as follows: TR = 6000 ms; TE = 5.6 ms; slice thickness = 1.0 mm; matrix = 96  $\times$  96. Relaxivity ( $r_1$  and  $r_2$ ) was determined using a linear fit of the relaxation time as a function of the amount of Gd or Fe ions determined by ICP-OES.

### 2.6. Cytotoxicity assays

HeLa cells at a density of  $5 \times 10^4$  cells per  $m^2$  were incubated in a humidified incubator at 37  $^\circ\text{C}$  for 24 h. The adherent HeLa cells were then treated with different concentrations of cisplatin and HPFNC in the growth medium at 37  $^\circ\text{C}$  in 96-well plates for 24 h and 48 h. Following this, MTT (5 mg  $mL^{-1}$ , 10  $\mu\text{L}$ ) was added to each well and incubated for an additional 4 h. The supernatant was removed, and 100  $\mu\text{L}$  of DMSO was added to completely dissolve the formazan crystals by vigorously shaking the cell culture plate for 10 min until no particulate matter was visible. The absorbance in each well was measured at 570 nm using a microplate reader (Biotek, USA). The biocompatibility of

the HPFN was also tested by following the above process, except that cisplatin was replaced with HPFN.

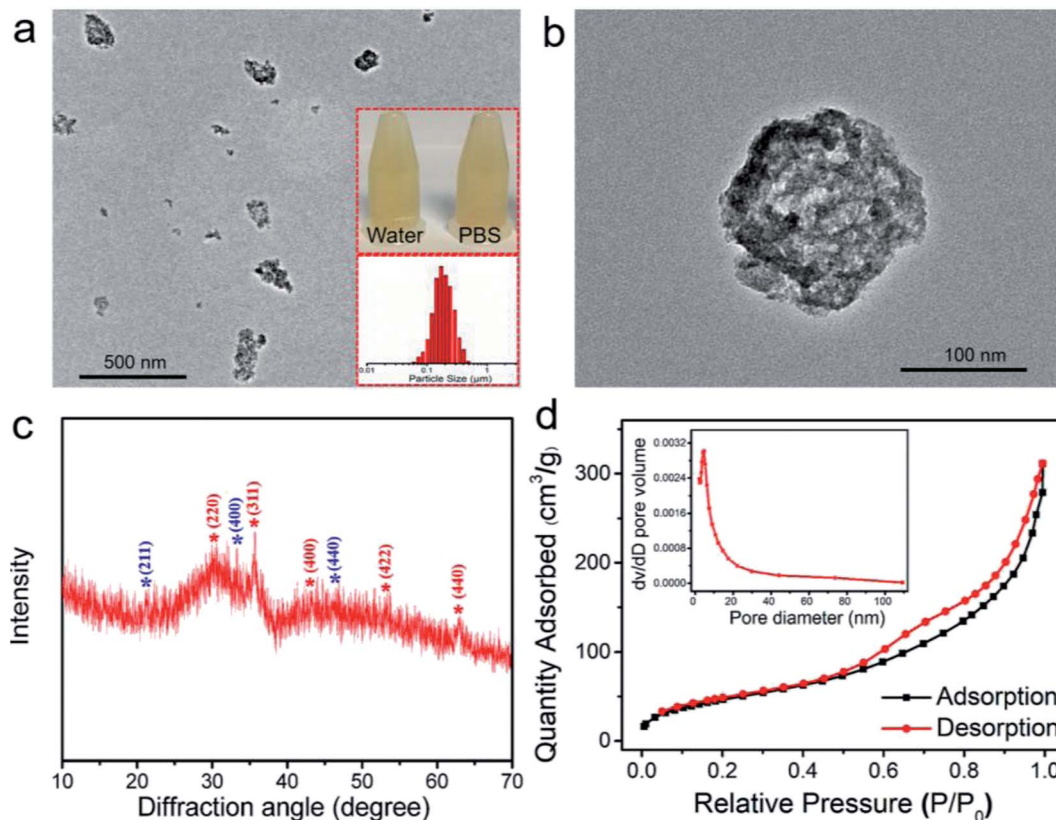
### 2.7. Characterization

The general morphology and microstructure of the sample were observed under a scanning electron microscope (SEM; Hitachi, Ltd., Tokyo, Japan) and a H-800 transmission electron microscope (TEM; Hitachi, Ltd., Tokyo, Japan). The nitrogen adsorption-desorption isotherm of the sample was measured using a porosimetry analyzer (Tristar II, 3020M, Micromeritics, USA). The structure and interaction of the sample was characterized using an X-ray diffractometer (XRD) (TTR-III, Rigaku Co., Japan) and an X-ray photoelectron spectroscope (XPS) (Thermo ESCALAB 250XI, USA), respectively. The composition of the sample was determined using a Thermo Nicolet iS10 Fourier transform infrared (FT-IR) spectrometer (Thermo Electron, Co., USA), and the spectral data were processed using the software of the spectrometer (OMNIC). The magnetic behavior of the sample was measured by a superconducting quantum interference device magnetometer (Bruker Co., Germany).

## 3. Results and discussion

### 3.1. Synthesis and characterization of HPFN

As shown in Scheme 1, we successfully synthesized a multi-functional HPFN nanomaterial using a simple solvothermal method.  $\text{Gd}^{3+}$  could localize anywhere within the



**Fig. 1** (a) Low magnification and (b) high magnification TEM images of HPFN. Inset: HPFN ( $0.5 \text{ mg mL}^{-1}$ ) dispersed in water and DMEM medium. Hydrodynamic size distribution of HPFN. (c) XRD pattern, and (d)  $\text{N}_2$  adsorption/desorption isotherms of HPFN.



nanonetworks, and it had a parallel spin ordering with the same direction as the magnetic field induced by the Fe-based material. At this point, the Fe-based material did not disturb  $\text{Gd}^{3+}$ ; instead, it further enhanced the  $T_1$  imaging. In addition, the porous and hollow structure with a large specific surface area was beneficial for the water molecules contacting  $T_1$  and  $T_2$  CAS as well as for sufficient loading of the drugs. In this case, HPFN could be used for simultaneous cancer diagnosis and chemotherapy.

As shown in Fig. 1a and b, the TEM micrographs display that the HPFN adopted a hollow and porous nanonetwork architecture with homogeneous dispersion. It is well known that good dispersibility of the nanoparticles is important for the nanomedical applications. The nanonetworks could be well dispersed both in water and PBS, and the hydrodynamic size analysis of the HPFN also showed that the nanonetworks possessed homogeneous size distribution. These results were consistent with the observation from TEM images. XRD peaks appearing in the HPFN display broad and low-intensity peaks, which are likely to be indexed to spinel  $\text{Fe}_3\text{O}_4$  (JCPDS no. 19-0629) and low-crystallinity  $\text{Gd}_2\text{O}_3$  (JCPDS no. 12-0797) (Fig. 1c), confirming that the nanomaterial was composed of ferrogadolinium oxide. As previously reported, many modifiers with active groups such as amino or carboxylate groups could control the formation of nanomaterials and improve the colloidal stability and biocompatibility of the prepared nanomaterials

during the reaction processes.<sup>27</sup> The mechanism could be attributed to the fact that these charged groups can coordinate with metal cations in solution and affect the nucleation and mass transfer process of the nanocrystals, thus resulting in the formation of controllable nanostructures. Similarly, in our case, two  $-\text{NH}_2$  groups exist in the head and tail of each 1,6-hexanediamine molecule with five C–C bonds in the middle. As shown in Fig. S1,† during the solvothermal process, in the first 1 h,  $\text{Fe}(\text{acac})_3$  and  $\text{Gd}(\text{acac})_3$  were continuously degraded to release iron and gadolinium cations, which could easily be coordinated by 1,6-hexanediamine, probably forming large molecular networks by the linking of positively charged iron and gadolinium cation-coordinated 1,6-hexanediamine molecules.<sup>27</sup> At the same time, the iron and gadolinium cations in the formed large molecular networks are slowly hydrated to form  $\text{Fe}(\text{OH})_3$  and  $\text{Gd}(\text{OH})_3$ , and reduced by ethylene glycol to form  $\text{Fe}_3\text{O}_4$  or undergo pyrolysis to form  $\text{Gd}_2\text{O}_3$  at high temperature and pressure condition.<sup>28</sup> During this process, the formed amorphous solid spheres grow continuously and start crystallizing at the surface of the spheres because the surface is in full contact with the solution.<sup>29</sup> Compared with the large and well-crystallized particles on the exteriors of the spheres, the inner amorphous crystallites have higher surface energy, which provides the driving force for the Ostwald ripening. Therefore, the inner crystallites tend to dissolve, resulting in interior void space and porous structure.<sup>29</sup>

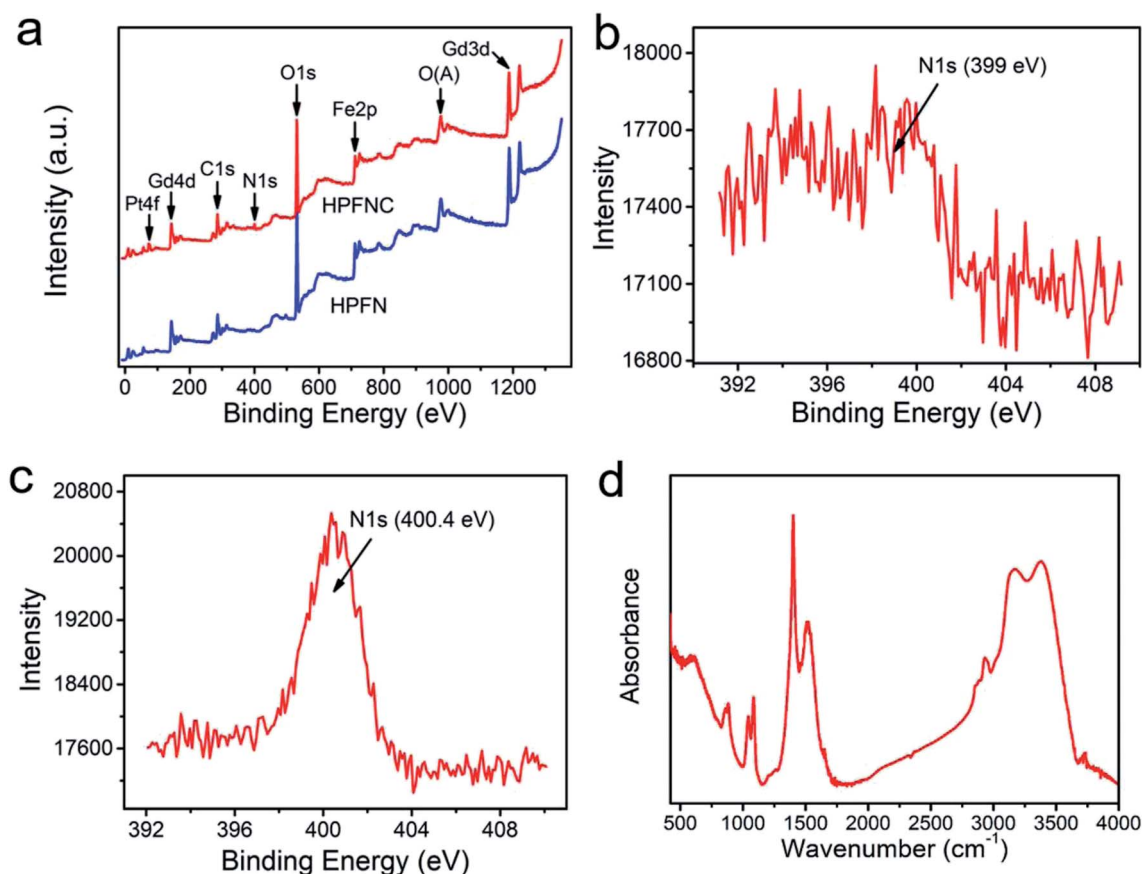


Fig. 2 (a) Full XPS spectra and N 1s peaks of (b) HPFN, and (c) HPFNC. (d) FT-IR spectrum of HPFN.





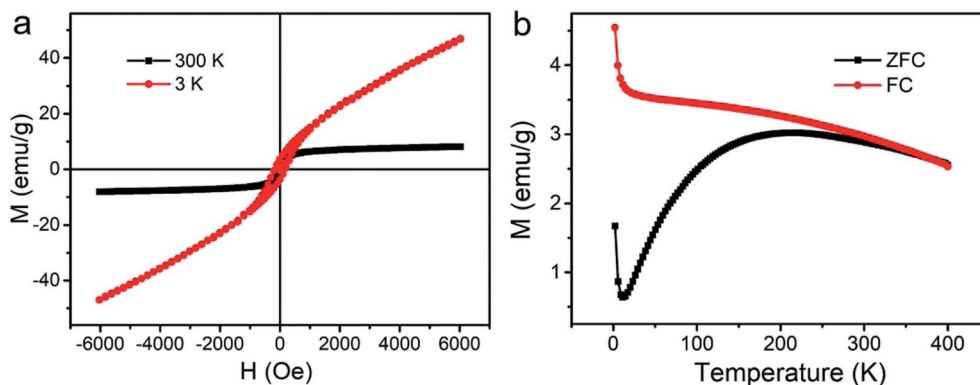


Fig. 3  $M-H$  and  $M-T$  curves of HPFN.

Subsequently, the specific surface area and porosity of HPFN were analyzed using nitrogen adsorption-desorption isotherm curves (Fig. 1d). The data showed that the HPFN had a high specific surface area ( $175.3 \text{ m}^2 \text{ g}^{-1}$ ) and a uniform pore size distribution (predominantly 4.6 nm). The isotherm curve showed that the typical hysteresis behavior at 0.5–0.8  $P/P_0$  was attributed to the delay in nitrogen evaporation from the hollow-space of lumen, which was blocked by the surrounding mesopores. This finding demonstrated that HPFN is an ideal drug carrier.

XPS analysis of the samples (Fig. 2a) confirmed the presence of Fe, Gd, O, C, and N elements, indicating the existence of ferro-gadolinium oxide and 1,6-hexanediamine in this nanocomposite. After loading cisplatin, a new peak at 73.3 eV (Pt 4f) appeared, indicating that cisplatin was successfully loaded into HPFN to form HPFNC. Moreover, the N 1s peak shifted from 399 eV (Fig. 2b) to 400.4 eV (Fig. 2c), confirming that cisplatin was loaded into HPFN through coordination between cisplatin and the  $\text{NH}_2$  group of HPFN. The drug loading amount was calculated as 7.9%.

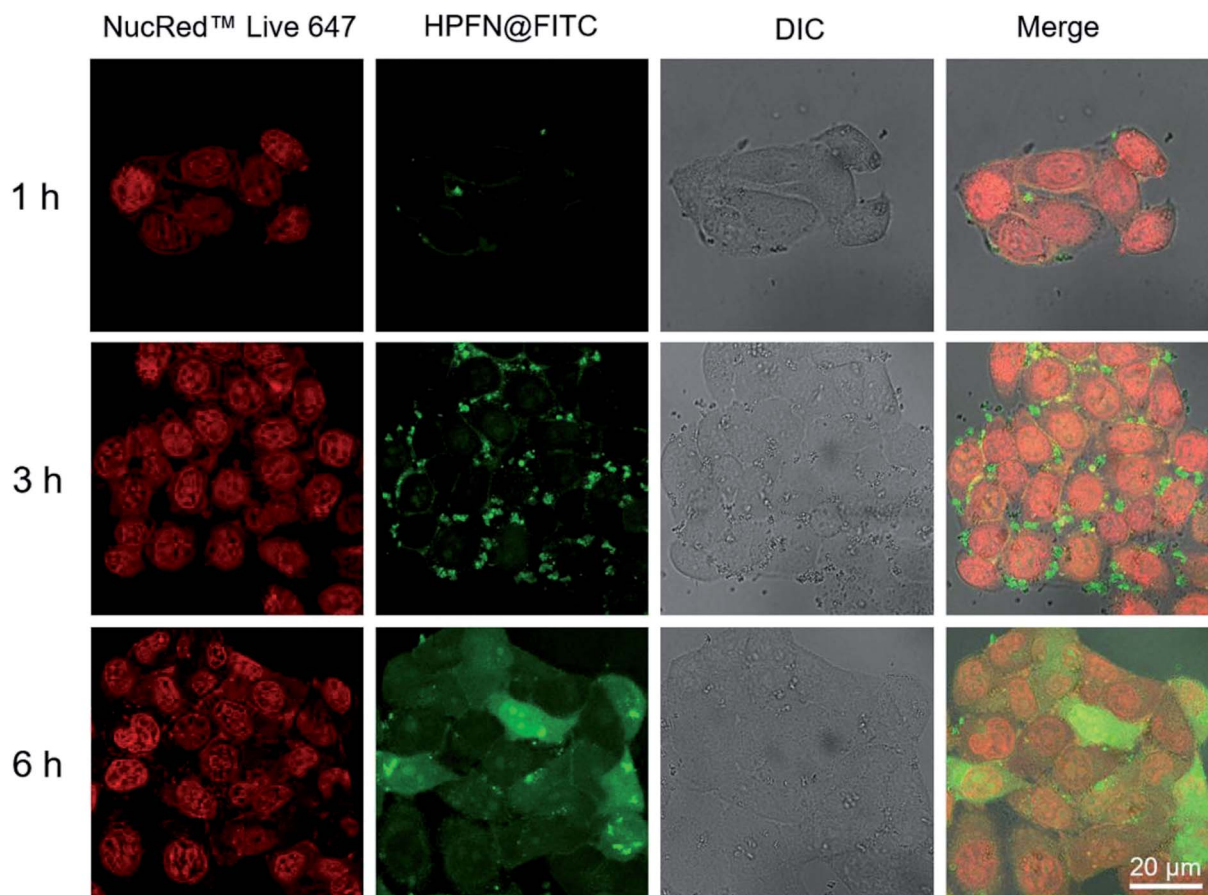


Fig. 4 (a) CLSM images of HeLa cells treated with HPFN-FITC for different indicated times. For each panel, the images from left to right show cell nuclei stained by NucRed Live 647 ReadyProbes Reagent (red), FITC fluorescence in cells (green), bright field, and a merged image of the three left-most images, respectively. All images had the same scale bar of 20  $\mu\text{m}$ .



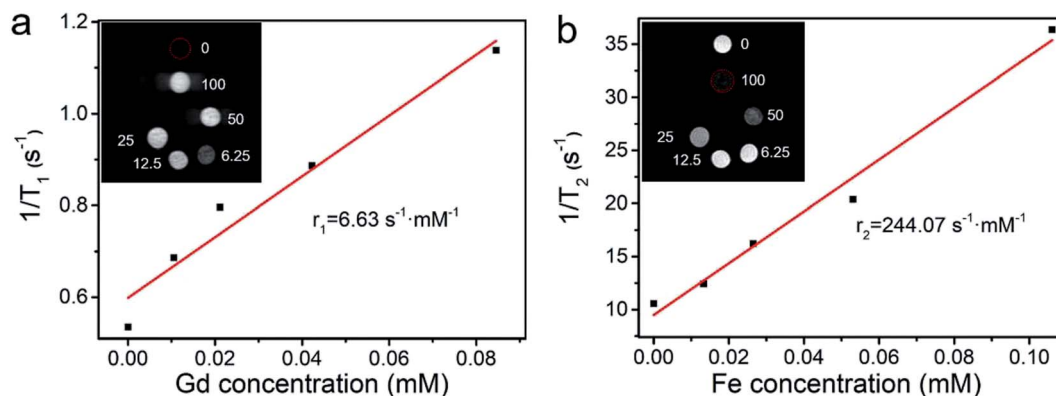


Fig. 5 (a)  $T_1$  and (b)  $T_2$  relaxivity and MR images of HPFN at different concentrations.

Subsequently, the composition of HPFN was confirmed using the FT-IR spectrum (Fig. 2d). The peaks at  $519\text{ cm}^{-1}$ ,  $588\text{ cm}^{-1}$ ,  $1512\text{ cm}^{-1}$ , and  $1080\text{ cm}^{-1}$  were respectively assigned to Gd–O stretching vibration, Fe–O stretching vibration, N–H bending vibration, and C–N stretching vibration (Fig. 2d), confirming the existence of  $\text{Gd}_2\text{O}_3$ ,  $\text{Fe}_3\text{O}_4$ , and 1,6-hexanediamine in HPFN.

Field-dependent magnetization curves showed that the saturated magnetization value of HPFN was  $9.6\text{ emu g}^{-1}$  at 300 K (Fig. 3a). In addition, the magnetization of HPFN continuously increased along with the increase in the applied magnetic field at 3 K, which may have been due to the enhanced spin canting effect of  $\text{Gd}^{3+}$  in HPFN.<sup>30,31</sup> In addition, the hysteresis at 3 K indicated that the nanonetworks possessed superparamagnetism. Standard zero-field cooling (ZFC) and field cooling (FC) measurements of HPFN displayed similar curves compared to those of paramagnetic  $\text{Gd}_2\text{O}_3$  at a low temperature, but gradually became similar to the curves of superparamagnetic  $\text{Fe}_3\text{O}_4$  with an increase in temperature (Fig. 3b). These magnetic properties of HPFN were similar to those previously reported for ferrogadolinium oxide composites,<sup>6</sup> indicating that the nanonetwork was composed of ferrogadolinium oxide composite.

### 3.2. Observation of uptake by HeLa cells

For the CLSM experiment, after the HeLa cells were incubated with HPFN-FITC for different times, the fluorescence of the cells displayed a time-dependent enhancement (Fig. 4). In the initial 1 h, weak green fluorescence was observed within the cytomembrane, indicating that only a few HPFN-FITC particles were localized within the cytomembrane of the HeLa cells. With the increase in incubation time, the fluorescence within the cytomembrane gradually increased, and weak fluorescence was also found in the cytoplasm. After 6 h of incubation, strong fluorescence distributed uniformly within the cells, suggesting that increasing number of HPFN-FITC particles crossed the cell membrane and entered the cytoplasm. These results confirmed that the HPFN could be effectively internalized by the HeLa cells, and can enhance the localized dose in HeLa cells.

### 3.3. $T_1$ – $T_2$ dual-modal MRI Performance

As shown in Fig. 5, with the increase in concentrations of HPFN,  $T_1$ - and  $T_2$ -weighted MR images gradually brightened and

darkened, respectively, indicating that the HPFN had a dose-dependent MRI contrast ability. The  $r_1$  value of the HPFN was  $6.63\text{ s}^{-1}\text{ mM}^{-1}$ , which was calculated as the ratio of the proton relaxation ( $1/T_1$ ) to the  $\text{Gd}^{3+}$  ion concentration. The  $r_2$  value of HPFN was  $244.07\text{ s}^{-1}\text{ mM}^{-1}$ , which was calculated as the ratio of the  $1/T_2$  to the  $\text{Fe}^{3+}$  ion concentration. These values were higher than the  $r_1$  and  $r_2$  values of the commercially available Gd or Fe-based contrast agents.<sup>32,33</sup> On one hand, the high  $r_1$  and  $r_2$  values may be attributed to the porous and hollow structure of HPFN, which was beneficial for the water freely contacting  $\text{Gd}^{3+}$  ions or being close to  $\text{Fe}_3\text{O}_4$  and thus, good  $T_1$  and  $T_2$  imaging was obtained. On the other hand, the Fe-based material inside the HPFN would not disturb the  $\text{Gd}^{3+}$  ions.

### 3.4. Investigation of biocompatibility and antitumor effect of the nanosystem

The release behavior of the drugs from HPFN under different pH conditions was explored before it was used for the antitumor experiment (Fig. S2†). The result indicated that the cisplatin release was pH-dependent. Cisplatin had low drug release rate at pH 7.4. However, the drug release rate could be promoted under acidic condition. This was because the coordination

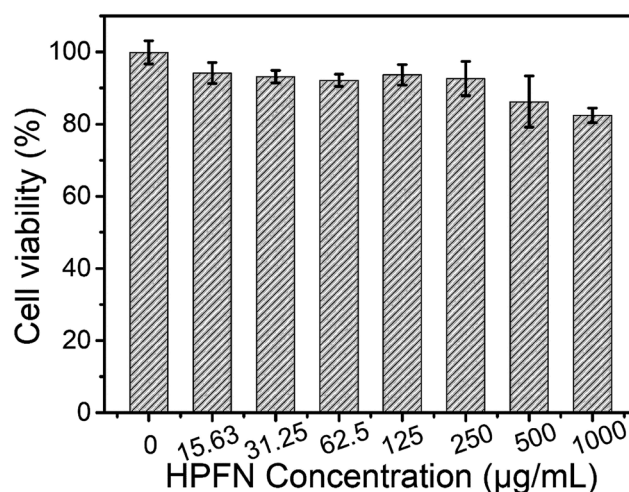


Fig. 6 Biocompatibility of HeLa cells treated with HPFN with increasing concentrations.



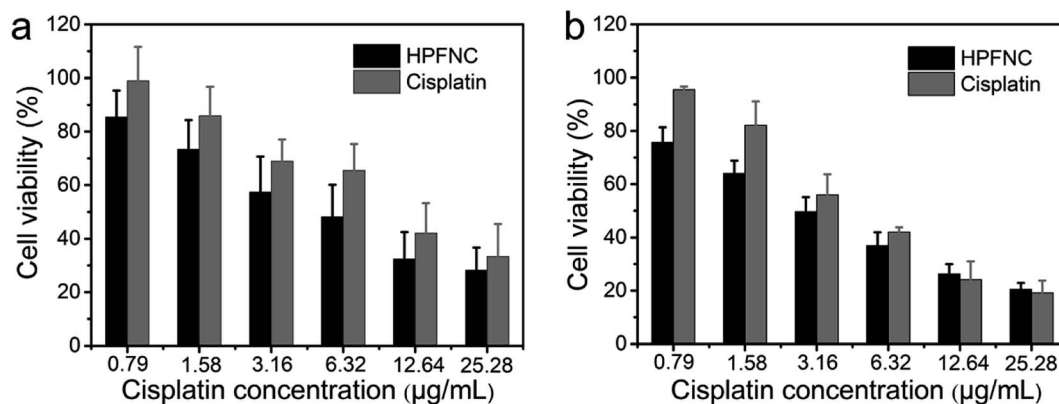


Fig. 7 Viability of HeLa cells treated with free cisplatin and HPFN-cisplatin with increasing cisplatin concentrations for (a) 24 h and 48 h (b).

effect between  $-NH_2$  and cisplatin could be easily destroyed under acidic conditions.<sup>28</sup> We know that good biocompatibility of nanomaterials is important for biomedical applications. As shown in Fig. 6, no significant decrease in the viability of the HeLa cells was observed after they were incubated with increasing HPFN concentrations for 48 h, indicating that HPFN had good biocompatibility. Following this, the viability of the HeLa cells incubated with free cisplatin and HPFNC was assessed. We found that cellular apoptosis induced by free cisplatin and HPFNC was strictly proportional to the dose of the anticancer drug (Fig. 7). Clearly, HPFNC displayed higher cancer cell inhibition than free cisplatin. This was because HPFN enhanced the localized drug dose in the HeLa cells. In addition, compared with 24 h of co-cultivation, 48 h of co-cultivation of the HeLa cells with either free cisplatin or HPFNC resulted in enhanced cellular apoptosis, indicating that the cellular apoptosis induced by free cisplatin and HPFNC was proportional to the co-cultivation time. This was because cisplatin needed enough time to affect DNA transcription. It is worth noting that at a high drug dose ( $>10.24 \mu\text{g mL}^{-1}$ ) after 48 h of co-cultivation, free cisplatin displayed a slightly higher cancer cell inhibition than HPFNC. This might be attributed to the fact that at a high-dose of HPFNC and a longer co-cultivation time, too many HPFNC particles were taken up by the cells, which inhibited the cells' uptake of more HPFNC. This was also confirmed when only a slight decrease in the viability of the HeLa cells was obtained on further increasing the dose of HPFNC.

## 4. Conclusions

In this study, we successfully developed a hollow and porous ferrogadolinium oxide nanonetwork (HPFN) using a simple solvothermal method. This nanoparticle with high  $T_1$  and  $T_2$  relaxivity ( $r_1 = 6.63 \text{ s}^{-1} \text{ mM}^{-1}$ ,  $r_2 = 244.07 \text{ s}^{-1} \text{ mM}^{-1}$ ) could be simultaneously used as  $T_1$  and  $T_2$  dual-modal MRI CA. The model anticancer drug cisplatin was efficiently loaded into HPFN with a large specific surface area ( $175.3 \text{ m}^2 \text{ g}^{-1}$ ), and the loading amount of cisplatin was up to 7.9%. The HPFN displayed satisfactory dual-modal MRI contrast ability. In the *in vitro* experiments, the HPFNC can efficiently kill cancer cells in

a dose/time-dependent manner. Therefore, HPFN with dual-modal MRI and drug loading ability could be used as a potential nanomedicine for simultaneous cancer diagnosis and chemotherapy.

## Conflicts of interest

The authors declare no conflict of interest.

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## References

- P. Mi, D. Kokuryo, H. Cabral, H. L. Wu, Y. Terada, T. Saga, I. Aoki, N. Nishiyama and K. Kataoka, *Nat. Nanotechnol.*, 2016, **11**, 724–730.
- B. R. Smith and S. S. Gambhir, *Chem. Rev.*, 2017, **117**, 901–986.
- C. Sun, J. S. Lee and M. Q. Zhang, *Adv. Drug Delivery Rev.*, 2008, **60**, 1252–1265.
- Y. Chen, K. Ai, J. H. Liu, X. Y. Ren, C. H. Jiang and L. H. Lu, *Biomaterials*, 2016, **77**, 198–206.
- J. N. Liu, W. B. Bu and J. L. Shi, *Chem. Rev.*, 2017, **117**, 6160–6224.
- X. Sun, R. H. Du, L. Zhang, G. L. Zhang, J. C. Qian, X. H. Tian, J. W. Zhou, J. C. He, Y. Y. Wang, Y. Q. Wu, K. Zhong, D. Q. Cai, D. H. Zou and Z. Y. Wu, *ACS Nano*, 2017, **11**, 7049–7059.
- Z. J. Zhou, D. T. Huang, J. F. Bao, Q. L. Chen, G. Liu, Z. Chen, X. Y. Chen and J. H. Gao, *Adv. Mater.*, 2012, **24**, 6223–6228.
- D. Frascione, C. Diwoky, G. Almer, P. Opriessnig, C. Vonach, K. Gradauer, G. Leitinger, H. Mangge, R. Stollberger and R. Prassl, *Int. J. Nanomed.*, 2012, **7**, 2349–2359.
- A. Yadollahpour, S. A. Hosseini, S. Rashidi and F. Farhadi, *Int. J. Pharm. Res. Allied Sci.*, 2016, **5**, 251–257.



- 10 J. Estelrich, M. J. Sanchez-Martín and M. A. Busquets, *Int. J. Nanomed.*, 2015, **10**, 1727–1741.
- 11 A. Alipoura, Z. Soran-Erdema, M. Utkura, V. K. Sharma, O. Algina, E. U. Saritasa and H. V. Demir, *Magn. Reson. Imaging*, 2018, **49**, 16–24.
- 12 Z. J. Zhou, C. Q. Wu, H. Y. Liu, X. L. Zhu, Z. H. Zhao, L. R. Wang, Y. Xu, H. Ai and J. Gao, *ACS Nano*, 2015, **9**, 3012–3022.
- 13 S. Santra, S. D. Jativa, C. Kaittanis, G. Normand, J. Grimm and J. M. Perez, *ACS Nano*, 2012, **6**, 7281–7294.
- 14 J. Choi, J. Lee, T. Shin, H. Song, E. Kim and J. Cheon, *J. Am. Chem. Soc.*, 2010, **132**, 11015–11017.
- 15 K. H. Bae, Y. B. Kim, Y. Lee, J. Y. Hwang, H. W. Park and T. G. Park, *Bioconjugate Chem.*, 2010, **21**, 505–512.
- 16 F. F. Li, D. B. Zhi, Y. F. Luo, J. Q. Zhang, X. Nan, Y. J. Zhang, W. Zhou, B. S. Qiu, L. P. Wen and G. L. Liang, *Nanoscale*, 2016, **8**, 12826–12833.
- 17 H. D. Cai, X. An, S. H. Wen, J. C. Li, G. X. Zhang, X. Y. Shi and M. W. Shen, *Part. Part. Syst. Charact.*, 2015, **32**, 934–943.
- 18 K. Cheng, M. Yang, R. P. Zhang, C. X. Qin, X. H. Su and Z. Cheng, *ACS Nano*, 2014, **8**, 9884–9896.
- 19 X. Sun, G. L. Zhang and Z. Y. Wu, *Curr. Med. Chem.*, 2018, **25**, 3036–3057.
- 20 G. H. Im, S. M. Kim, D. G. Lee, W. J. Lee, J. H. Lee and I. S. Lee, *Biomaterials*, 2013, **34**, 2069–2076.
- 21 P. Verwilt, S. Park, B. Yoon and J. S. Kim, *Chem. Soc. Rev.*, 2015, **44**, 1791–1806.
- 22 X. L. Liu, C. T. Ng, P. Chandrasekharan, H. T. Yang, L. Y. Zhao, E. Peng, Y. B. Lv, W. Xiao, J. Fang, J. B. Yi, H. Zhang and K. H. Chuang, *Adv. Healthcare Mater.*, 2016, **5**, 2092–2104.
- 23 T. H. Shin, J. S. Choi, S. Yun, I. S. Kim, H. T. Song, Y. Kim, K. I. Park and J. Cheon, *ACS Nano*, 2014, **8**, 3393–3401.
- 24 G. L. Zhang, R. H. Du, L. L. Zhang, D. Q. Cai, X. Sun, Y. Zhou, J. Zhou, J. C. Qian, K. Zhong, K. Zheng, D. Kaigler, W. Q. Liu, X. Zhang, D. H. Zou and Z. Y. Wu, *Adv. Funct. Mater.*, 2015, **25**, 6101–6111.
- 25 F. Ries and J. Klastersky, *Am. J. Kidney Dis.*, 1986, **8**, 368–379.
- 26 A. Hosseini, S. Fanoudi, H. Mollazadeh, A. Aghaei and M. Boroushaki, *J. Pharm. BioAllied Sci.*, 2018, **10**, 66–71.
- 27 J. Gao, X. Ran, C. Shi, H. Cheng, T. Cheng and Y. Su, *Nanoscale*, 2013, **5**, 7026–7033.
- 28 X. Sun, C. Cai, Q. Wang, *et al.*, *Phys. Chem. Chem. Phys.*, 2016, **18**, 7820–7828.
- 29 W. Cheng, K. Tang, Y. Qi, J. Sheng and Z. Liu, *J. Mater. Chem.*, 2010, **20**, 1799–1805.
- 30 P. Du, J. H. Lim, S. H. Kim and J. S. Yu, *Opt. Mater. Express*, 2016, **6**, 1896–1904.
- 31 I. Sadiq, S. Naseem, M. Naeem Ashiq, M. Asif Iqbal, I. Ali, M. A. Khan, S. Niaz and M. U. Rana, *J. Magn. Magn. Mater.*, 2015, **395**, 159–165.
- 32 H. Chen, G. D. Wang, W. Tang, T. Todd, Z. P. Zhen, C. Tsang, K. Hekmatyar, T. Cowger, R. B. Hubbard, W. Z. Zhang, J. Stickney, B. Z. Shen and J. Xie, *Adv. Mater.*, 2014, **26**, 6761–6766.
- 33 H. Wei, O. T. Brunsa, M. G. Kaulb, E. C. Hansen, M. Barch, A. Wiśniowska, O. Chen, Y. Chen, N. Li, S. Okada, J. M. Cordero, M. Heine, C. T. Farrar, D. M. Montana, G. Adam, H. Ittrich, A. Jasanoff, P. Nielsen and M. G. Bawendi, *Proc. Natl. Acad. Sci. U. S. A.*, 2017, **114**, 2325–2330.

