

Cite this: *RSC Adv.*, 2019, **9**, 151

## miR-425 suppresses EMT and the development of TNBC (triple-negative breast cancer) by targeting the TGF- $\beta$ 1/SMAD 3 signaling pathway

Yingping Liu<sup>a</sup> and Jinglong Chen<sup>id, \*b</sup>

**Background:** EMT has a crucial effect on the progression and metastasis of tumors. This work will elucidate the role of miR-425 in EMT and the development of TNBC. **Methods:** The differential miRNA expression among non-tumor, para-tumor (adjacent tissue of tumor) and tumor tissues was analyzed. The luciferase activities of TGF- $\beta$ 1 3'UTR treated with miR-425 were determined. Then human breast cancer cell lines were treated with mimics or inhibitors of miR-425, and then the cell proliferation and migration, and invasion ability were assessed. The expression of TGF- $\beta$ 1 and markers of epithelial cells and mesenchymal cells were analyzed. The influences of miR-425 on the development of TNBC through inducing EMT by targeting the TGF- $\beta$ 1/SMAD3 signaling pathway in TNBC cell lines were investigated. Furthermore, xenograft mice were used to explore the potential roles of miR-425 on EMT and the development of TNBC *in vivo*. **Results:** Compared with non-tumor tissues, 9 miRNAs were upregulated and 3 miRNAs were down-regulated in tumor tissues. The relative expression of miR-425 in tumor tissues was obviously much lower than that in para-tumor and non-tumor tissues. MiR-425 suppressed TGF- $\beta$ 1 expression, and further inhibited expression of mesenchymal cell markers, while it exerted effects on cell proliferation and migration of TNBC cell lines. Moreover, the agomir of miR-425 could protect against the development process in a murine TNBC xenograft model. **Conclusions:** Our results demonstrated that miR-425 targets TGF- $\beta$ 1, and was a crucial suppressor on EMT and the development of TNBC through inhibiting the TGF- $\beta$ 1/SMAD3 signaling pathway. This suggests that aiming at the TGF- $\beta$ 1/SMAD3 signaling pathway by enhancing relative miR-425 expression, is a feasible therapy strategy for TNBC.

Received 25th October 2018  
 Accepted 17th December 2018

DOI: 10.1039/c8ra08872a  
[rsc.li/rsc-advances](http://rsc.li/rsc-advances)

### Introduction

The most usual cancer of women is BC (Breast Cancer) worldwide.<sup>1</sup> Moreover, the morbidity of BC is the seventh among all cancers, and its mortality rate is third in women's world.<sup>2</sup> According to different prognoses and options of treatment, BC is divided into several subtypes of disease associated with heterogeneity.<sup>3,4</sup> Based on the expression of ERBB2/HER2 (epidermal growth factor receptor 2), PR (progesterone receptor), and ER (estrogen receptor), patients with BC are classified in three subtypes of triple-negative, HER2+, and ER+/PR+ in the clinic. The overall prognosis of patients with Her2+ or ER+/PR+ subtypes has been dramatically improved by treatment with targeted therapy at HER2 with hormone. However, the aggressiveness of TNBC (triple-negative breast cancer) and the lack of effective targeted therapy cause the worst overall

survival rate of patients with TNBC.<sup>4</sup> About 20% patients with BC have TNBC, and they have high risk of visceral organ metastasis, such as brain, liver, and lung, *etc.*<sup>5</sup> Therefore, there is an urgent need to develop new therapeutic interventions for preventing and treating metastasis of TNBC.

EMT (epithelial to mesenchymal transition) plays an important role in metastasis and progression of cancer. During this process, the invasion and migration abilities increase, epithelial polarity vanishes away together with epithelial markers down-expression and mesenchymal markers over-expression.<sup>6-13</sup> Some transcription factors regulate the process of EMT, including Zeb1, Slug, Snail, TGF $\beta$ , and Twist.<sup>9,13</sup> The biological process of EMT is complex, including loss of epithelial characteristics and acquire mesenchymal phenotypes. Moreover, the expressions of mesenchymal cell markers, such as Smad 3,  $\alpha$ -SMA (alpha smooth muscle actin), *etc.*, enhanced.<sup>14,15</sup> The growth of tumor relies not only on tumor malignancy but also the relationship between stromal and tumor cells.<sup>14,15</sup> The system of tumor vessel includes pericytes, smooth muscle cells and, epithelial cells (ECs), which plays an important role in tumors progression and angiogenesis activation, such as BC.<sup>16,17</sup> The EMT process of ECs is conducted to the

<sup>a</sup>Department of Obstetrics and Gynecology, Beijing Obstetrics and Gynecology Hospital, Capital Medical University, 251# Yaojia Yuan Road, Chaoyang District, Beijing 100026, P. R. China

<sup>b</sup>Department of Oncology, Beijing Ditan Hospital, Capital Medical University, Beijing 100015, PR China. E-mail: [dtzlzx@sina.com](mailto:dtzlzx@sina.com)



progression of cancer, that has been identified by recent researches.<sup>18–20</sup> In this process, the properties of migration or invasion and markers of mesenchyma (e.g.,  $\alpha$ -SMA, FSP-1) are obtained, the epithelial markers (e.g., CD31, VE-cadherin) of ECs and junctions of cell–cell were losed.<sup>21</sup> These investigations confirm that all kinds of pathological processes occurs with, for example, cancer.<sup>18,19,22</sup> Therefore, inhibition of EMT process may be a great treatment strategy for tumor.

Lately, some scholars have revealed that TGF- $\beta$ 1 is a cytokine with multifunction, and plays a crucial role in pathophysiology of BC.<sup>23</sup> The canonical and noncanonical TGF- $\beta$ 1 signaling pathways gene mutations encoding TGF- $\beta$ 1 or the change of TGF- $\beta$ 1 receptors associates with BC oncogenic activity. The progression of epithelial cell cycle of BC is inhibited by TGF- $\beta$ 1 at early phase while apoptosis is promoted, which indicates its suppressive effects on tumor.<sup>24</sup> However, in the late phase, TGF- $\beta$ 1 promotes tumor progression, is associated with metastasis or invasiveness of tumor and higher motility of cancer cells. In addition, TGF- $\beta$ 1 induces EMT, and is related to modification of microenvironment in cancer.<sup>25</sup> The well known “TGF- $\beta$ 1 paradox” is called to describe this phenomenon, which suggests TGF- $\beta$ 1 can be used for the novel therapy approaches on BC with better understanding its functions.<sup>26</sup> By protecting against TGF- $\beta$ 1, several drugs for nonclinical and clinical investigation in early stages are developed based on these knowledge.

The current realization demonstrated that mesenchymal characteristics of TNBC cells with paclitaxel-resistance, and these cells has higher mesenchymal markers expression, including N-cadherin and vimentin. Thus, through inducing inhibition of TGF- $\beta$  signaling pathway can promote chemotherapeutic effect on anti-tumor and reduce metastasis after paclitaxel therapy with kinase inhibitor for TGF- $\beta$ 1 receptor.<sup>27</sup> Therefore, it indicates that the novel potential strategy for TNBC therapy through combination with kinase inhibitors of TGF- $\beta$  and its receptor.

Many data confirmed that microRNAs have crucial effect on activity of epithelial or EMT.<sup>28,29</sup> So, in our investigation, the relationship between down-expression of miR-425 and TNBC was identified by comparing miRNA expression profiles from breast cancer tissues of patients with TNBC. Over-expression of miR-425 played an important regulator in negative feedback for TGF- $\beta$  signaling and canonical pathway through suppressing expression of TGF- $\beta$ 1 and its changes associated with EMT in development of TNBC.

## Materials and methods

### Specimens of patients

The tissues of patients with TNBC were obtained from January 2014 to December 2017. All these patients were performed with mastectomy for BC, and did not receive any treatment before hospital. At the operation day, samples of tumor tissues, para-tumor tissues and non-tumor tissues were collected and handled immediately, respectively. The para-tumor is adjacent tissue of tumor.

This investigation and the utilization of human specimens in this work had been approved by the Ethics Committee of our

hospitals (Beijing Obstetrics and Gynecology Hospital, Capital Medical University, Beijing, China) on the basis of the Declaration of Helsinki. We clearly confirmed that the informed consents were obtained from all patients. Moreover, the recorded and documented of participant consents were kept in our hospital.

### Extraction of miRNAs and next-gener. sequencing

The miRNA-sequencing test was conducted after extraction of RNAs with TRIzol method and miRNeasy Mini Kit (Cat. no. 74104; Invitrogen Thermo Fisher Scientific, Inc., Waltham, MA, USA). Furthermore, total RNA was harvested from tissues for RT-qPCR assay with mirVana™ PARIS™ Kit (Cat. no. AM1556; Invitrogen Thermo Fisher Scientific, Inc, Waltham, MA, USA). Hybridization and ligation were conducted using Adaptor Mix. Moreover, microRNA sequencing RNAs using Illumina HiSeq 2000 platform was conducted after reversely transcribe of these RNAs.

### Data extraction and analysis

All the mature miRNAs sequences were identified through library preparation of reference sequences with miRBase (version 19.0) (<http://www.mirbase.org/>).<sup>30</sup> Given that the length of minimum read was 50 nucleotides, specific adapters were used to anneal to their 3' ends and extended the entire small RNAs during the library preparation. Thus, we removed the adapters and obtained the potential miRNAs with the length of 15–30 nucleotides for further analysis using cut adapt software *in silico*.<sup>31</sup> Subsequently, the sequences were perfectly matched with the prepared reference library with Bowtie (version 0.12.7) (<https://sourceforge.net/projects/bowtie-bio/files/bowtie/0.12.7/>).<sup>32</sup> Then the numbers of mapped reads were counted for every miRNA. Acquired data of all samples were normalized with RPM (reads per million).<sup>33</sup>

### RNA extraction

TRIzol method was used to extract total RNA from tissues based on the operation manual. Then, smaller RNA (<200 nt) was removed with mirVana RNA isolation kit (Cat. no. AM27828; Invitrogen Thermo Fisher Scientific, Inc., Waltham, MA, USA) according to the instructions for manufacturer.

### Culture of cell

The cell lines of human TNBC, including MDA-MB-231 (ATCC® HTB-26™) and Hs 578T (ATCC® HTB-126™) were purchased from ATCC (American Type Culture Collection), and then cultured in DMEM with 10% FBS accompanied by Streptomycin and Penicillin (bi-antibiotics) with atmosphere (5% CO<sub>2</sub>) at 37 °C.

### siRNA for TGF- $\beta$ 1

The well known siRNA of TGF- $\beta$ 1 supported by ref. 26 had been used in our study, which had been used as a positive in our study. The siRNA duplexes for TGF- $\beta$ 1 sequences are 5'-



GAAGCGGACUACUAUGCUAUU-3' (upstream) and 5'-UUCUUCGCCUGAUGAUACGAU-3' (downstream).

### qRT-PCR

The miR-425 expression level was assessed with TaqMan miRNA assays in tissues. The data analysis was performed with  $2^{-\Delta\Delta C_q}$  method (Livak 2011). The qRT-PCR assay was conducted using SYBR Green kit (Cat. no. QR0100-1KT; Sigma-Aldrich; Merck KGaA, Darmstadt, Germany). The thermocycling conditions for qPCR were as follows: pre-enaturation at 95 °C for 30 s, followed by 40 cycles of denaturation at 95 °C for 5 s and extension at 60 °C for 30 s.

The primer sequences of miR-425 were 5'-ACACTC-CAGCTGGAAATGACACGATCACTCC-3' (upstream) and 5'-TGGTGTGCGTGGAGTCG-3' (downstream), respectively.<sup>34</sup> Moreover, the primer sequences of U6 were 5'-TGACCTGAAACATACACGGGA-3' (upstream) and 5'-TATCGTTGTACTC-CACTCCTTGAC-3' (downstream), respectively.

The primer sequences of TGF-β1 were 5'-CAA-CACGATGCTTGAAGGTAACG-3' (upstream) and 5'-TCCAGAGA-GATGATTGCCGAGG-3' (downstream), respectively. The primer sequences of VE-cadherin were 5'-GGATGCAGAGGCTCACAGAG-3' (upstream) and 5'-CTGGCGGTTCACGTTGGACT-3' (downstream), respectively. The primer sequences of E-cadherin were 5'-TACACTGCCAGGAGCCAGA-3' (upstream) and 5'-TGGCACCACTGTCCGGATTA-3' (downstream), respectively. The primer sequences of N-cadherin were 5'-GGA-CAGTTCTGAGGGATCA-3' (upstream) and 5'-GGATTGCCTTC-CATGTCTGT-3' (downstream), respectively. The primer sequences of vimentin were 5'-GGCTCAGATTTCAGGAACAGC-3' (upstream) and 5'-GCTTCAACGGCAAAGTTCTC-3' (downstream), respectively. The primer sequences of α-SMA were 5'-TGTCCAGCCGTCCTTCAT-3' (upstream) and 5'-GGCGTAGTCTTCCTGATG-3' (downstream), respectively. The primer sequences of SMAD3 were 5'-AGCACACAA-TAACTTGGACC-3' (upstream) and 5'-TAAGACACACTGGAA-CAGCGGATG-3' (downstream), respectively. Moreover, the primer sequences of GAPDH (glyceraldehyde-3-phosphate dehydrogenase) were 5'-CTCATGACCACAGTCCATGCC-3' (upstream) and 5'-GGCATGGACTGTGGTCATGAG-3' (downstream), respectively.

### Reporter gene assays

The online software Targetscan (version 7.2) ([http://www.targetscan.org/vert\\_72/](http://www.targetscan.org/vert_72/)) was firstly used to predict the microRNA which could regulate the expression of TGF-β1. Therefore, miR-425 was acquired from the predict result. Furthermore, the 3'UTR mRNAs sequence of TGF-β1 was cloned into pmirGLO vector with luciferase reporter firefly and renilla. The control was used with reverse orientation of TGF-β1 3'UTRs mRNA. The complementary region sequence in TGF-β1 3'UTR was GGACUGCGGAUCUCUGUGUCAUU (Fig. 1), which located at position 151–157, and corresponding miR-425 seed sequence AGUUGCCUCACUAGCACAGUAA. Site-Mutation kit (Cat. no. E0554S; New England Biolabs Ltd, UK) was used to constructed TGF-β1 3'UTR mutated vector. Finally, Dual-Glo® Luciferase

Assay System (Cat. no. E2940; Promega Corporation, Madison, WI, USA) was utilized to analyze the activity of luciferase with analyzer VICTOR.

### The mimics of miR-425 and its inhibitors

The mimics of miR-425 and its inhibitors were transfected with Lipofectamine 3000. Moreover, the mimics sequences of miR-425 were 5'-AAUGACACGAUCACUCCGUUGA-3'.<sup>34</sup> Moreover, the control sequence of miR-425 mimics were 5'-UUCUCC-GAACGUGUCACGUTT-3'. The sequence of miR-425 agomir was 5'-asasugacacgaucacucccgusugsgas-3'. The sequence of NC (negative control) was 5'-ususcuccgaacgugucacgsussts-3', while it was not consistent with sequences of human genome.

### Western blot

A lysis buffer (1% NP-40, 0.1% SDS, 150 mM NaCl, 0.02%  $\text{NaN}_3$ , and 50 mM Tris pH 8.0) containing cocktail of inhibitor for protease and phenylmethylsulfonyl fluoride (1 mM) was used to lysate cells. Western blotting assays were performed with Bio-Rad Protean II minigel system. Then, 50  $\mu\text{g}$  protein sample was injected in every gel (12%) well, and then shifted to PVDF membrane after electrophoresis. 1× TBST (5% dry skimmed milk) was utilized to dispute the non-specific binding. Then these membranes were sequentially dealt with primary and secondary antibody, respectively. The membranes were incubated with primary antibodies at 4 °C overnight. The membranes were incubated with secondary antibodies with secondary antibodies goat anti-rabbit immunoglobulin G (IgG)-horseradish peroxidase (HRP) (sc-2004; 1 : 5000 dilution; Santa Cruz Biotechnology, Inc.) or goat anti-mouse IgG-HRP (sc-2005; 1 : 5000 dilution; Santa Cruz Biotechnology, Inc.) for 1 h at room temperature. Furthermore, these membranes were determined with enhanced ECL (chemiluminescence) reagent of Hyperfilm ECL kit (Cat. no. 28906835; Amersham GE Healthcare, Chicago, IL, USA), and exposed using X-ray film.

The primary antibodies included TGF-β1 (Santa Cruz, CA, USA; sc-130348, 1 : 1200), α-SMA (CST 19245, dilution 1 : 1200) and SMAD 3 (C67H9) (CST 9523, dilution 1 : 1500), p-SMAD 3 (CST-9520, dilution 1 : 1000), VE-cadherin (CST2158, dilution 1 : 1000), E-cadherin ( $24 \times 10^9$ ) (CST 3195, dilution 1 : 800), N-cadherin (D4R1H) (CST 13116, dilution 1 : 1200), vimentin (D21H3) (CST 5741, dilution 1 : 1000), β-actin ( $13 \times 10^5$ ) (CST 4970S, dilution 1 : 4000) (Cell Signaling, Danvers, MA, USA).

### Analysis of cell proliferation and migration

The proliferation ability of cells was assessed using MTS kit (Cat. no. G3580; Promega Corporation, Madison, WI, USA). Subculture the stock cultures of cells to  $2 \times 10^4$  cells per ml in medium containing 50  $\mu\text{M}$  2-mercaptoethanol (2-ME), when a density of  $2 \times 10^5$  cells per ml is reached, mimics or inhibitors of miR-425 and its control is added, respectively. Wash the cells twice with the above medium centrifugation at 300×g for 5 minutes. The number and viability of cells based on trypan blue stain, 50  $\mu\text{l}$  resuspend cells ( $1 \times 10^5$  cells per ml), was added into all wells of 96-well plate prepared, then incubate at 37 °C for 48 h. Furthermore, 20  $\mu\text{l}$  CellTiter 96® AQueous One



	Predicted consequential pairing of target region (top) and miRNA (bottom)	Site type	Context++ score
Position 151-157 of TGFB1 3' UTR hsa-miR-425-5p	5' ...GGACUGCGGAUCUCUGUGUCAUU... 3' AGUUGCCUCACAUAGCACAGUAA	7mer-m8	-0.19

**Fig. 1** Predicted consequential pairing of target region. The online software Targetscan (version 7.2) was firstly used to predict the microRNA which could regulate the expression of TGF- $\beta$ 1. Therefore, miR-425 was acquired from the predict result. The complementary region sequence in TGF- $\beta$ 1 3'UTR was GGACUGCGGAUCUCUGUGUCAUU, which located at position 151–157, and corresponding miR-425 seed sequence AGUUGCCUCACAUAGCACAGUAA.

Solution Reagent was added into per well, and then incubate at 37 °C for 2 h. Record the absorbance at 490 nm using a 96-well plate reader.

The ability of cells migration was detected using wound scratch assay using CytoSelect™ 24-Well Wound Healing Assay kit (Cat. no. CBA-120-5; Cell Biolabs, Inc., San Diego, CA, USA). At first, 24-well plate with CytoSelect™ Wound Healing Inserts was warm up for 10 minutes at room temperature, then sterile forceps was used to orient the inserts direction. Secondly, 500  $\mu$ l cell suspension ( $1 \times 10^6$  cells per ml) was added into each well. Then cultured in incubator overnight. The next day, the inserts were removed from wells, then slowly discarded the media. After washing, dead cells and debris in wells were further removed. Finally, hydrated cells were visualized with light microscope. The migration rate of the cells into the wound field were measured. Ten fields per experimental condition were randomly selected and micrographed with IX71 microscope (Olympus, Japan) to quantify cells of migration. Images represent at least three independent experiments.

#### Matrikel invasion assay

Matrikel was dissolved overnight at 4 °C and then diluted with serum-free DMEM (1 : 3). The diluted Matrikel was added to the upper chamber of the Transwell to form the membrane in three additions (15, 7.5 and 7.5  $\mu$ l), with 10 min between each addition. The Matrikel was evenly spread and covered all micropores at the bottom of the upper chamber. The two cell lines were collected at 48 h after transfection to prepare cell suspensions. Cell suspensions were seeded into the upper well, and 0.5 ml of DMEM containing 10% FBS was added to the lower chambers (24-well plates). The cells were cultured routinely for 48 h at 37 °C with 5% CO<sub>2</sub>. After using a cotton swab to gently wipe off the remaining free cells in the upper well, the well membrane was collected and fixed for 15–20 min in 95% ethanol. After rinsing with clean water, the membrane was stained for 10 min, and washed again with clean water. The membrane was then observed under a high-powered inverted microscope to count the number of cells on the membrane. The invasion ability of the cells was determined by counting the number of cells that passed through the Matrikel, and the average number from five high-magnification fields of view was used for each sample.

#### Animals

Nude mice (purchased from Laboratory Animal Sciences, Capital Medical University, Beijing, China) were used to

conduct our investigation at this work. A total of 12 nude mice, female, were randomly divided into 2 groups. The experiment was initiated with 6 week-old mice weighing 20–25 g. They were induced to generate xenograft model with injection of MDA-MB-231 cells, and then housed of Laboratory Animal Center, Capital Medical University, Beijing, China. Mice were maintained at 20–24 °C (temperature controlled) with 12 h light/dark cycle and standard laboratory diet in a pathogen-free environment. All experiments related to animals were performed based on Laboratory Animal Center, Capital Medical University, Beijing, China, Ethical Committee Acts. The experimental mice were dealt with according to the standards supported by the Animal Protection Committee of Capital Medical University. Moreover, the ethical approval was obtained for the animal experiments conducted in your study from Animal Protection Committee of Capital Medical University.

#### Experiment *in vivo*

The experiment was initiated with 6 week-old of mice, which weight was 20–25 g. Then, 1 ml suspension of MDA-MB-231 cells ( $1 \times 10^6$  cells per ml) was injected into subcutaneous tissue of nude mice at right flanks ( $n = 6$ /group). While tumor size of mice reached 3–8 mm, the experiment initiated as the 1st day. The agomir of miR-425 were formulated (1.0 mg ml<sup>-1</sup> in PBS). The nude mice were categorized into experiment and control groups at random, then injected with agomir of miR-425 (5  $\mu$ l) or its control for 6 times once every three days and terminated on the 18th day. Then, these mice were sacrificed at the 21st day, and tumor mass was dissected from each mouse. The methods of anesthesia and euthanasia conducted in our study was that 20% urethane anesthetized (at 1 g kg<sup>-1</sup>) was used to execute mice at three weeks after injection of agomir of miR-425 or its control. These tumors were cut into two sections, then the volume of tumors were determined every 3 days base on the formula width<sup>2</sup> × length × 0.5. Moreover, the expression of TGF- $\beta$ 1 were determined with western blotting and immunohistochemical staining assay, respectively.

The dose of urethane used for euthanasia of mice is (at 1 g kg<sup>-1</sup>), the following methods were used to evaluate consciousness or confirm death: lack of a heartbeat, lack of respiration, and lack of corneal reflex. The following conclusions can be drawn from this definition. A humane endpoint: (a) not necessarily means the humane killing of the animal, but could also result in interventions to alleviate the stressful/painful experimental procedure (e.g. performing surgery) or providing



analgesics. (b) It is not necessarily based on clinical signs but could also start from pre-clinical signs or from physiological or molecular biomarkers predictive of pain/distress later on the disease process. (c) Should be balanced against the scientific endpoints to be met. Thus, pain and distress might be intrinsic to a certain experimental model (*e.g.* arthritis). However, in this case the humane endpoint should never be beyond the scientific endpoint. (d) Should never be beyond the level of moral justification.

### Immunohistochemistry assay

Tissue samples were prepared using 10% neutral formalin, then embedded using paraffin after dehydration. Furthermore, it was cut into thick sections with 5  $\mu$ M (Marcela *et al.* 2018), dewaxed and hydrated. After antigen retrieval, incubation with primary antibody for these sections was performed. The sections were incubated with primary antibodies TGF- $\beta$ 1 (Santa Cruz, CA, USA; sc-130348, 1 : 200), or SMAD 3 (C67H9) (CST 9523, dilution 1 : 200) at 4 °C overnight, respectively. The sections were incubated with a DAB substrate kit (Cat. no. 34002; Thermo Scientific™ Pierce™, San Jose, CA, USA) after incubation with the secondary antibody. The membranes were incubated with secondary antibodies goat anti-rabbit immunoglobulin G (IgG)-horseradish peroxidase (HRP) (sc-2004; 1 : 1000 dilution; Santa Cruz Biotechnology, Inc.) or goat anti-mouse IgG-HRP, (sc-2005; 1 : 1000 dilution; Santa Cruz Biotechnology, Inc.) for 1 h at room temperature. These sections were assessed, and then photographed with microscope at 200 $\times$  (IX71; Olympus, Japan). Per sample was taken for 3–5 shots. The software Image-Proplus (version 6.0) was used measure proteins expression through detecting optical density in each photograph.

### Statistics

Welch *t*-test-paired was performed to compare isomiRs deregulation and selection of miRNAs between samples of patients and control. The errors were evaluated with false discovery rate (FDR) due to multiple comparisons. On the basis of the expression profiles, hierarchical clustering of selected miRNAs was conducted with Ward's agglomeration. Target Rank software (<http://genes.mit.edu/targetrank/>) was performed to

identify the significantly deregulated each seed sequence of miRNA in target genes between samples of patients and control.<sup>35</sup>

All data with normally distributed was represented as mean  $\pm$  SD (standard deviation). The differences between two groups were analyzed with Student's *t*-test. Kruskal-Wallis ANOVA method was used to analyze abnormally distributed data among groups. When the variances were equal, the least significant difference *post hoc* test was used, whereas when the variances were not uniform, Dunnett's *post hoc* test was used. SPSS (version 18.0; SPSS, Inc., Chicago, IL, USA) soft was performed for entire statistical analyses. While  $P < 0.05$  indicated statistically significant.

## Results

### Differential miRNA expression profiling

To assess the differential expression of miRNA profiles between tumorous and para-tumorous tissues, miRNA-sequencing experiments were conducted on the total RNA obtained from samples. Among the results of miRNAs-sequencing, 12 were differentially expressed between tumorous and para-tumorous tissues. Compared with para-tumorous tissues, 9 miRNAs were upregulated and 3 miRNAs were down-regulated in patients with TNBC (Table 1).

### The expression level of miR-425 in non-tumorous, para-tumorous, and tumorous tissues

Furthermore, the relative miR-425 expression levels in non-tumorous, para-tumorous, and tumor tissues ( $n = 60$ ) were determined using qRT-PCR method. Our work identified the relative miR-425 expression level was obviously less than that of non-tumorous or para-tumorous tissues. However, there was no obvious difference between para-tumorous and non-tumorous tissues (Fig. 2).

### Expression levels of miR-425 were determined in cell lines of human TNBC

Moreover, the relative miR-425 expression levels were investigated in two cell lines of human TNBC. Compared with basal

Table 1 Differentially expressed miRNAs in tumorous and tumorous tissues (fold-change  $>2.0$  and  $P$ -value  $< 0.01$ )

miRNA	Fold-change (tumorous/para-tumorous tissues)	<i>t</i> value	<i>P</i> value
miR-148b	5.73	8.381	$7.72 \times 10^{-4}$
miR-34a	4.08	6.935	$9.33 \times 10^{-3}$
miR-196a	4.01	7.220	$7.00 \times 10^{-3}$
miR-205	3.91	9.272	$5.46 \times 10^{-3}$
miR-328	3.18	2.702	$6.96 \times 10^{-3}$
miR-122	2.64	7.254	$1.77 \times 10^{-3}$
miR-155	2.45	3.823	$5.44 \times 10^{-3}$
miR-202	2.36	6.256	$7.68 \times 10^{-3}$
miR-let-7a	2.34	3.293	$4.42 \times 10^{-3}$
miR-7	0.35	-0.391	$5.49 \times 10^{-3}$
<b>miR-425</b>	0.18	-0.468	$7.53 \times 10^{-3}$
miR-31	0.14	-0.326	$2.07 \times 10^{-3}$



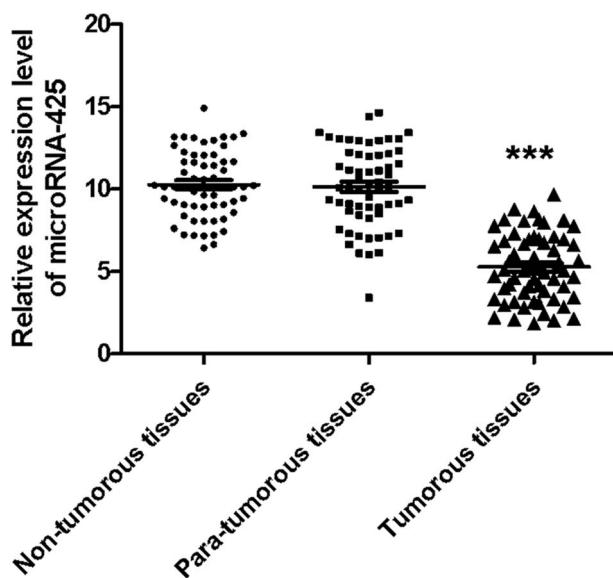


Fig. 2 The expression level of miR-425 in non-tumor, para-tumor, and tumor tissues. The relative miR-425 expression levels in non-tumor, para-tumor, and tumor tissues ( $n = 60$ ) were determined using qRT-PCR method. This result identified the relative miR-425 expression level in tumor tissues was obviously less than that of non-tumor or para-tumor tissues ( $P < 0.001$ ). However, there was no obvious difference between para-tumor and non-tumor tissues. \*\*\* $P < 0.001$ .

epithelial phenotype cell line Hs 578T, the relative miR-425 expression levels were significantly higher in mesenchymal phenotypic cell line MDA-MB-231 ( $P < 0.001$ ) (Fig. 3).

#### MiR-425 inhibited TGF- $\beta$ 1/SMAD3 pathway and mesenchymal markers expression in TNBC cell lines

At first, the target of miR-425 were predicted with Targetscan, an online software. Therefore, miR-425 was found that it is the possibly target of 3' UTRs untranslated region of TGF- $\beta$ 1 mRNA

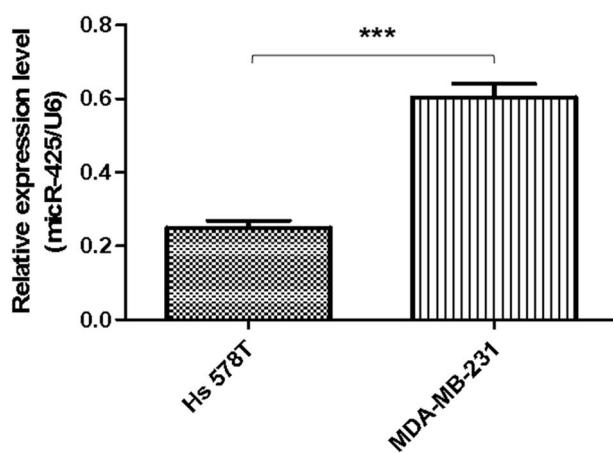


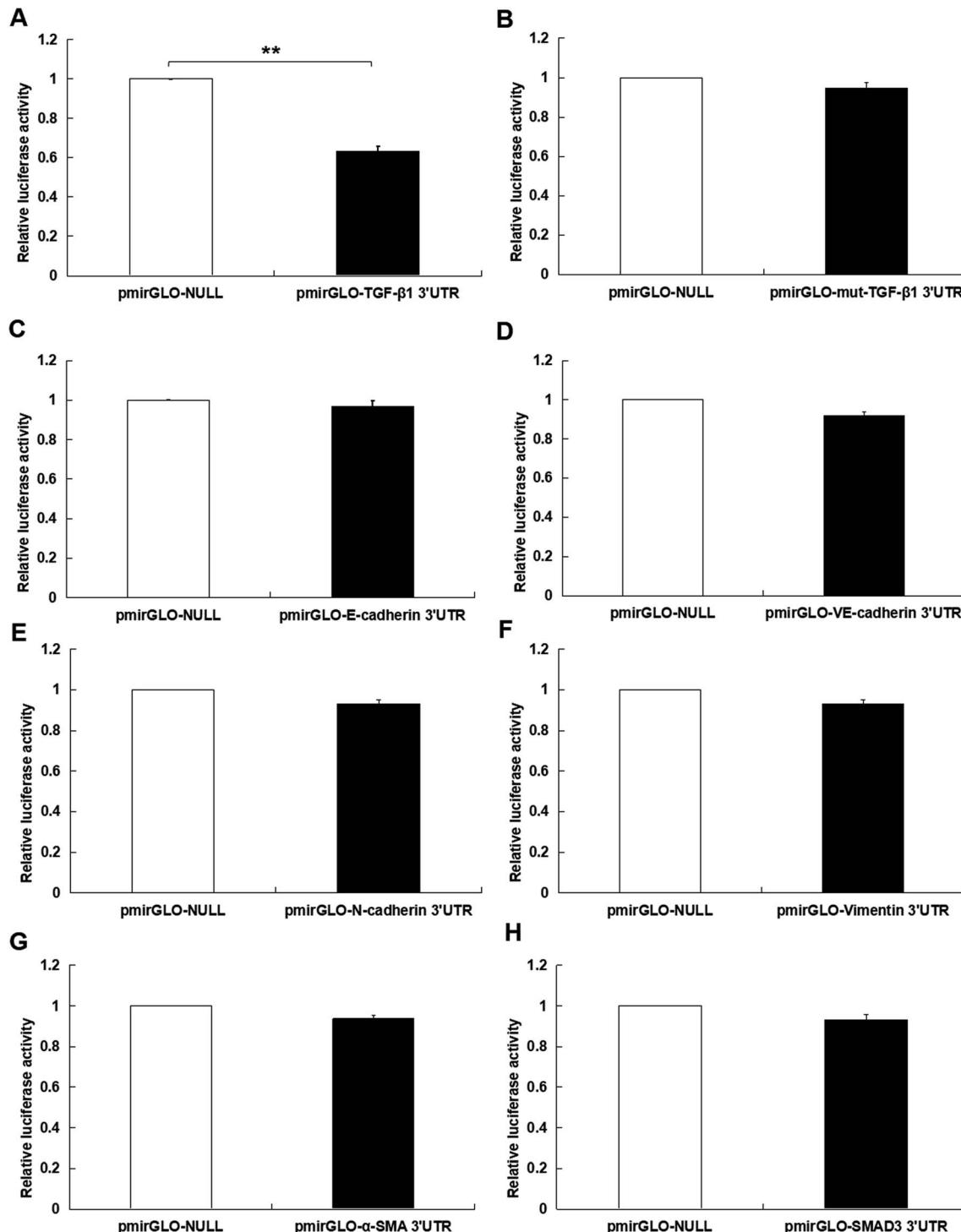
Fig. 3 Expression levels of miR-425 were determined in cell lines of human TNBC. The relative miR-425 expression levels were investigated in six cell lines of human TNBC. Compared with basal epithelial phenotype cell line Hs 578T, the relative miR-425 expression levels were significantly higher in mesenchymal phenotypic cell line MDA-MB-231 ( $P < 0.001$ ). The data are presented as means  $\pm$  SD from at least three independent experiments. \*\*\* $P < 0.001$ .

(Table 1). Further experiments were conducted using luciferase reporter gene system to identify whether miR-425 could straight bind with 3' UTRs of TGF- $\beta$ 1 mRNA. The data indicated 3' UTR luciferase activities of TGF- $\beta$ 1 significantly down-regulated in MDA-MB-231 cell line dealt with miR-425 (Fig. 4A). However, it did not show obviously change in the group of TGF- $\beta$ 1 mutation (Fig. 4B). Moreover, miR-425 did not bind directly to 3' UTRs of E-cadherin (Fig. 4C), VE-cadherin (Fig. 4D), N-cadherin (Fig. 4E), vimentin (Fig. 4F),  $\alpha$ -SMA (Fig. 4G), and SMAD3 (Fig. 4H), respectively. These data identified that miR-425 targeted to TGF- $\beta$ 1 mRNAs. The well known siRNA of TGF- $\beta$ 1 supported by ref. 26 had been used in our study, which had been used as a positive in our study. It could inhibit the TGF- $\beta$ 1/SMAD3 signaling pathway in TNBC cell lines (Fig. 5).

To explore the roles of miR-425 on mRNAs and proteins expression of TGF- $\beta$ 1,  $\alpha$ -SMA, vimentin, N-cadherin, E-cadherin, VE-cadherin, and SMAD3 in human BC cell lines, the qRT-PCR assay and western blotting trial were performed. These results demonstrated that miR-425 restrained mRNAs and proteins expressions of TGF- $\beta$ 1 in MDA-MB-231 cell line. However, miR-425 could not suppress the mRNAs expression of  $\alpha$ -SMA, vimentin, N-cadherin, VE-cadherin, E-cadherin, and SMAD3 (Fig. 6A). The western blotting result showed that miR-425 promoted proteins expression of E-cadherin and VE-cadherin, while suppressed proteins expression of  $\alpha$ -SMA, vimentin, N-cadherin, and SMAD3 (Fig. 6C). Moreover, miR-425 inhibitors could promote expression of TGF- $\beta$ 1 mRNA in MDA-MB-231, but did not affect mRNAs expression of  $\alpha$ -SMA, vimentin, N-cadherin, VE-cadherin, E-cadherin, and SMAD3 (Fig. 6A). The inhibitors of miR-425 could suppress the protein expression levels of N-cadherin, vimentin,  $\alpha$ -SMA, and SMAD3, but promote VE-cadherin and E-cadherin protein expression level (Fig. 6C). In a word, the result indicated that, by binding with TGF- $\beta$ 1 3'UTR, miR-425 could straight regulate mRNA and protein expression of TGF- $\beta$ 1, further affect protein expression of genes associated with EndMT process and TGF- $\beta$ 1/SMAD3 pathway. It was same as the results in cell line Hs 578T (Fig. 6B and D).

The mesenchymal markers expression of  $\alpha$ -SMA, vimentin, N-cadherin, SMAD3 and p-SMAD3 in TNBC cell lines were significantly inhibited, while endothelial markers expression of E-cadherin and VE-cadherin were obviously promoted by mimics of miR-425 *in vitro* compared with control group (Fig. 6C and D). But, the mesenchymal cell markers expression of TNBC cells were significantly promoted, while endothelial markers expression of E-cadherin and VE-cadherin were significantly suppressed by inhibitors of miR-425 in TNBC cells (Fig. 6C and D). The ratio of phosphorylated *versus* total protein did not show significant difference between each groups in MDA-MB-231 and Hs 578T cell lines (Fig. 6E and F). Our results identified that miR-425 suppressed mesenchymal markers expression and enhanced endothelial markers expression in TNBC cell lines, especially for the mesenchymal phenotypic cell line MDA-MB-231. The expressions of mesenchymal markers N-cadherin, vimentin,  $\alpha$ -SMA, SMAD3 and p-SMAD3 in the mesenchymal phenotypic cell line MDA-MB-231 were higher than that in the basal phenotypic epithelial cell line Hs 578T,





**Fig. 4** MiR-425 inhibited TGF- $\beta$ 1 expression in TNBC cell lines. The target of miR-425 were predicted with Targetscan, the online software. Therefore, miR-425 was found that possibly aimed to 3' UTRs untranslated region of TGF- $\beta$ 1 mRNA. Further experiments were conducted using luciferase reporter gene system to identify whether miR-425 could straight bind with 3' UTRs of TGF- $\beta$ 1 mRNA. The data indicated 3' UTR luciferase activities of TGF- $\beta$ 1 significantly down-regulated in MDA-MB-231 cell line dealt with miR-425 (A). However, it did not show obviously change in the group of TGF- $\beta$ 1 mutation (B). Moreover, miR-425 did not bind straight to 3' UTRs of E-cadherin (C), VE-cadherin (D), N-cadherin (E), vimentin (F),  $\alpha$ -SMA (G), and SMAD3 (H), respectively. These data identified that miR-425 targeted to TGF- $\beta$ 1 mRNAs. The data are presented as means  $\pm$  SD from three independent experiments. \* $P$  < 0.05, \*\* $P$  < 0.01.

and all could be suppressed obviously by mimics of miR-425 in the two cell lines. Moreover, the expressions of endothelial markers E-cadherin and VE-cadherin in the mesenchymal

phenotypic cell line MDA-MB-231 were less than that in the basal phenotypic epithelial cell line Hs 578T, and all two cell lines could be promoted significantly by mimics of miR-425.

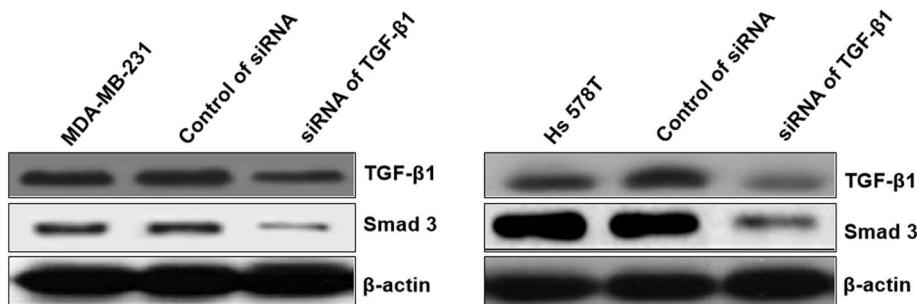


Fig. 5 The siRNA of TGF- $\beta$ 1 inhibited TGF- $\beta$ 1/SMAD3 signaling pathway in TNBC cell lines. The well known siRNA of TGF- $\beta$ 1 supported by ref. 26 had been used in our study, which had been used as a positive in our study. It could inhibit the TGF- $\beta$ 1/SMAD3 signaling pathway.

### MiR-425 suppressed cell proliferation, migration, and invasion of TNBC cell lines

To assess the biofunctions of miR-425, over-expressed and down-expressed miR-425 was conducted, respectively. Furthermore, several molecular functional tests were performed in TNBC cells. The proliferation ability of TNBC cells was suppressed by miR-425 compared to that of control, and suppressed by inhibitors of miR-425 (Fig. 7A). Moreover, the change of proliferation ability induced by mimics of miR-425 in mesenchymal phenotypic cell line MDA-MB-231 was significantly more than that of basal epithelial phenotype cell line Hs 578T, while the change between the two cell lines was statistically significant difference ( $P < 0.05$ ) (Fig. 7A).

Additionally, the inhibitors of miR-425 promoted migration ability of TNBC cell lines, but the mimics of miR-425 suppressed the migration ability of TNBC cell lines (Fig. 7B and C). Furthermore, the change of migration ability induced by inhibitors of miR-425 in mesenchymal phenotypic cell line MDA-MB-231 was higher than that of basal epithelial phenotype cell line Hs 578T, while the change between the two type cell lines was not statistically significant difference ( $P > 0.05$ ) (Fig. 7B and C). It identified that miR-425 suppressed the process of proliferation and migration in TNBC cell lines, especially for mesenchymal phenotypic cell line MDA-MB-231. The hypothesis to explain these results was that the inhibition roles induced by miR-425 in proliferation and migration of TNBC cell line, mesenchymal phenotypic cell line MDA-MB-231 were much more than the basal epithelial phenotype cell line Hs 578T (Fig. 7). Moreover, it maybe the mesenchymal characteristics of MDA-MB-231 cell line was obviously more than Hs 578T cell line.

Moreover, the inhibitors of miR-425 promoted invasion ability of TNBC cell lines, but the mimics of miR-425 suppressed the invasion ability of TNBC cell lines (Fig. 8).

### MiR-425 agomir suppressed TNBC xenografts

To detect anti-tumor activity induced by miR-425 agomir *in vivo*, human TNBC xenografts were established with MDA-MB-231 cells. Our work demonstrated that the size of tumor (Fig. 9A) and the average tumor weight (Fig. 9B) of control mice was much larger than that of experiment mice treated with agomir of miR-425, and the difference between two groups had

statistical significance. Moreover, the xenografts of control groups grew rapidly, but in the experiment mice treated with agomir of miR-425, the xenografts grew slowly *in vivo* (Fig. 9C). The maximum diameter exhibited by a single subcutaneous tumor of control mice in our study was 1.23 cm.

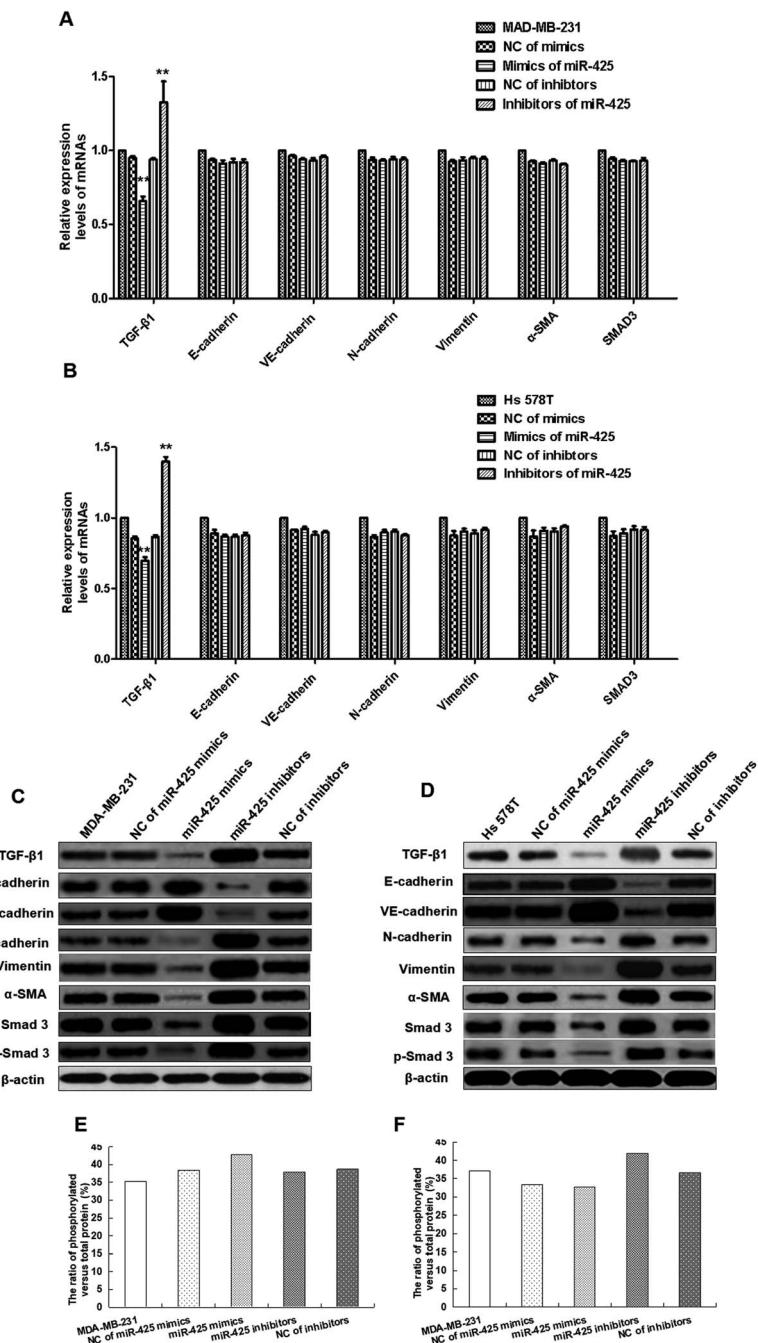
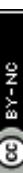
The expression proteins in xenografts were determined with immunohistochemical staining methods (Fig. 10A) and western blotting assay (Fig. 10B), respectively. The results revealed that when compare with control group, treatment with agomir of miR-425 could down-regulate the expression of SMAD3 and TGF- $\beta$ 1 with a statistically significant difference, and suppressed mesenchymal markers expression of xenograft. The mesenchymal markers expression of  $\alpha$ -SMA, vimentin, N-cadherin, SMAD3, p-SMAD3 and endothelial markers E-cadherin, VE-cadherin were determined in xenografts. The results indicated that compare with control group, treatment with agomir of miR-425 could obviously down-regulate the above mesenchymal markers expression in xenografts, while up-regulate expression of endothelial markers E-cadherin, VE-cadherin. To sum up, our data confirmed that the agomir of miR-425 could inhibit development of TNBC associated with EMT through aiming TGF- $\beta$ 1/SMAD3 signaling pathway *in vivo*.

## Discussion

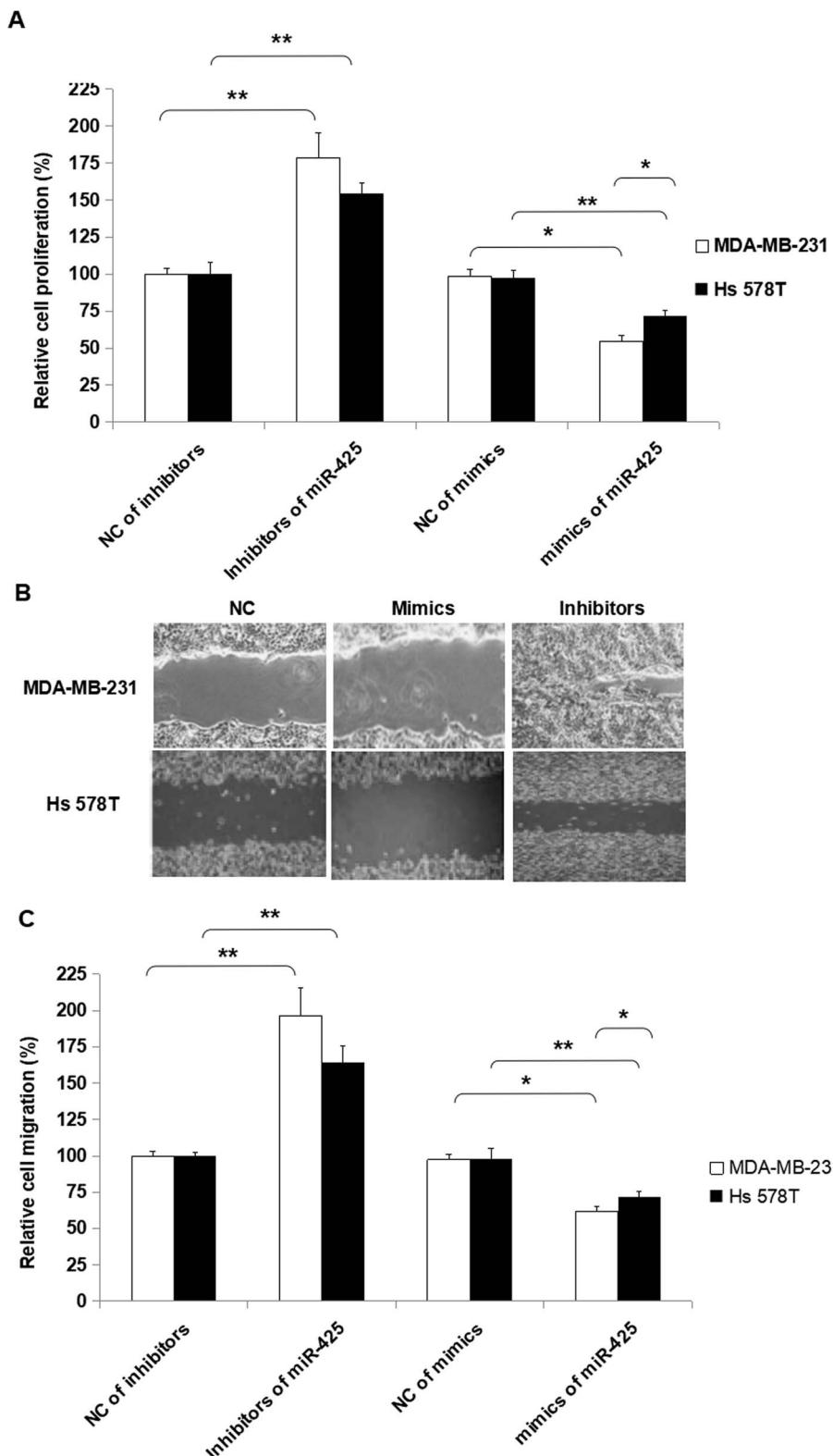
Through EMT (epithelial–mesenchymal transition), a mesenchymal phenotype in epithelial cells can be acquire, wherein the marker expression and acquired functions of mesenchymal cell, in addition to lost the epithelial functions and markers expression of epithelia. Besides, migration and delamination of epithelial cells can be caused by EMT, then mesenchymal cells derived from epithelial cell can migrate into the underlying tissue.<sup>36</sup> The ligands of TGF- $\beta$  (transforming growth factor  $\beta$ ) family often initiate the cellular process, EMT.<sup>37</sup> Many investigations aim at the markers of EMT from thirty-two cancer types<sup>38,39</sup> and the roles of TGF- $\beta$  pathway.<sup>38,40</sup> A number of elements are important factors in regulation of gene expression and EMT, while referred to development of TNBC.<sup>41–44</sup>

Moreover, the transcriptional regulation mechanism of EMT and how miRNA modulated EMT or development of TNBC was unclear yet. Recent research revealed that miR-34 might serve as inhibitor in nasopharyngeal carcinoma (NPC) by targeting





**Fig. 6** MiR-425 inhibited TGF- $\beta$ 1/SMAD3 pathway and mesenchymal markers expression in TNBC cell lines. To explore the roles of miR-425 on mRNAs and proteins expression of TGF- $\beta$ 1, E-cadherin, VE-cadherin, N-cadherin, vimentin,  $\alpha$ -SMA, total and phosphorylated SMAD3 (p-SMAD3) in human BC cell lines, the qRT-PCR assay and western blotting trial were performed. These results demonstrated miR-425 restrained TGF- $\beta$ 1 mRNAs and proteins expressions in MDA-MB-231 cell line. However, miR-425 could not suppress the expression of E-cadherin, VE-cadherin, N-cadherin, vimentin,  $\alpha$ -SMA, and SMAD3 mRNAs (A), but promote proteins expression of E-cadherin and VE-cadherin, while suppress proteins expression of N-cadherin, vimentin,  $\alpha$ -SMA, total and phosphorylated SMAD3 (p-SMAD3) (C). Moreover, miR-425 inhibitors could promote expression of TGF- $\beta$ 1 mRNA in MDA-MB-231, but did not affect mRNAs expression of E-cadherin, VE-cadherin, N-cadherin, vimentin,  $\alpha$ -SMA, total and phosphorylated SMAD3 (p-SMAD3) (A). The inhibitors of miR-425 could suppress the protein expression levels of N-cadherin, vimentin,  $\alpha$ -SMA, total and phosphorylated SMAD3 (p-SMAD3), but promote protein expression of E-cadherin and VE-cadherin (C). In a word, by binding with TGF- $\beta$ 1 3'UTR, the result indicated miR-425 could straight regulate its mRNA and protein expression, further affect protein expression of genes associated with EndMT process and TGF- $\beta$ 1/SMAD3 pathway. It was same as the results in other cell line Hs 578T (B and D). The mesenchymal markers expression of N-cadherin, vimentin,  $\alpha$ -SMA, SMAD3 and p-SMAD3 in TNBC cell lines were inhibited significantly, while endothelial markers expression of E-cadherin and VE-cadherin were obviously promoted by mimics of miR-425 *in vitro* compared with control group (C and D). But, the expression of mesenchymal cell markers in TNBC cell lines were significantly promoted, while endothelial markers expression of E-cadherin and VE-cadherin were significantly suppressed by mimics of miR-425 in TNBC cell lines (C and D). The ratio of phosphorylated versus total protein did not show significant difference between each groups in MDA-MB-231 and Hs 578T cell lines (E and F). The data are presented as means  $\pm$  SD from three independent experiments. \* $P$  < 0.05, \*\* $P$  < 0.01.



**Fig. 7** MiR-425 suppressed cell proliferation and migration of TNBC cell lines. To assess the biofunctions of miR-425, over-expressed and down-expressed miR-425 was conducted, respectively. Furthermore, several molecular functional tests were performed in TNBC cell lines. The proliferation ability of TNBC cell lines was suppressed by miR-425 compared to that of control. Moreover, the change of proliferation ability induced by mimics of miR-425 in mesenchymal phenotypic cell line MDA-MB-231 was significantly more than that of basal epithelial phenotype cell line Hs 578T, while the difference was statistically significant ( $P < 0.05$ ). Additionally, the inhibitors of miR-425 promoted proliferation ability of TNBC cell lines. Furthermore, the change of proliferation ability induced by inhibitors of miR-425 in mesenchymal phenotypic cell line MDA-MB-231 was higher than that of basal epithelial phenotype cell line Hs 578T, while the difference between the two type cell lines was not statistically significant ( $P > 0.05$ ). The data are presented as means  $\pm$  SD from three independent experiments. \* $P < 0.05$ , \*\* $P < 0.01$ .



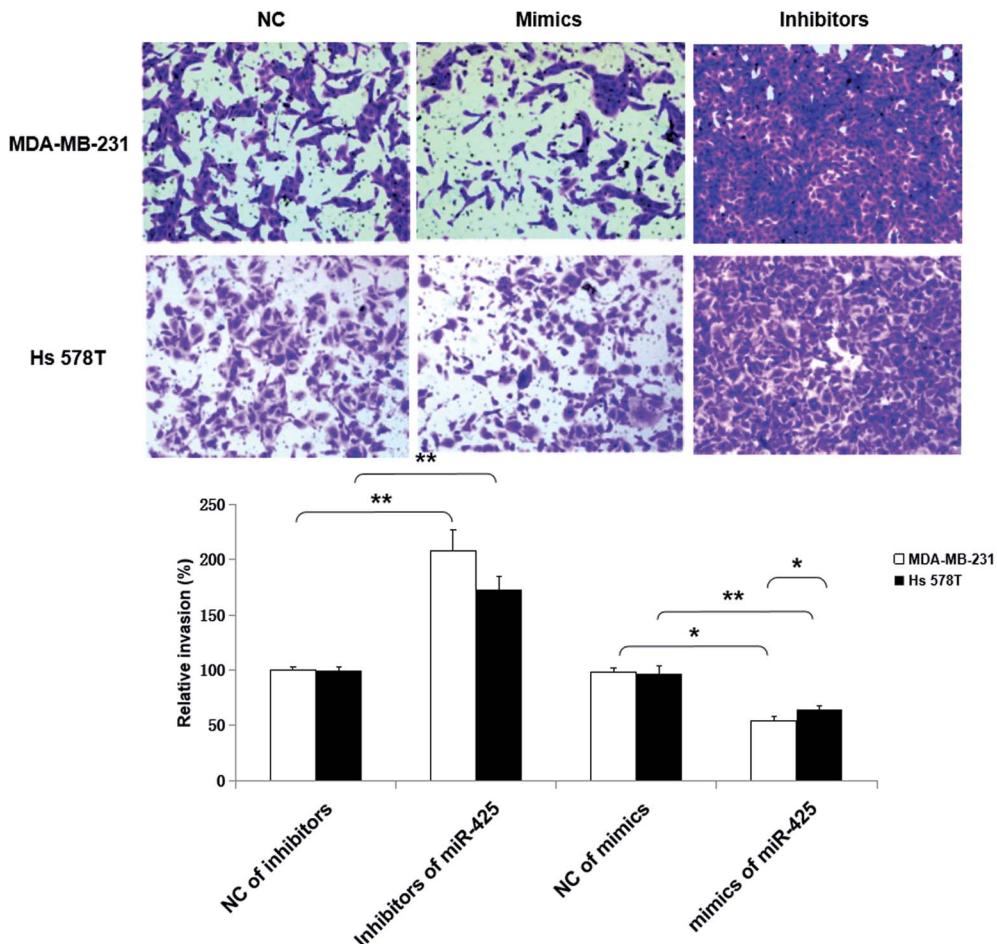


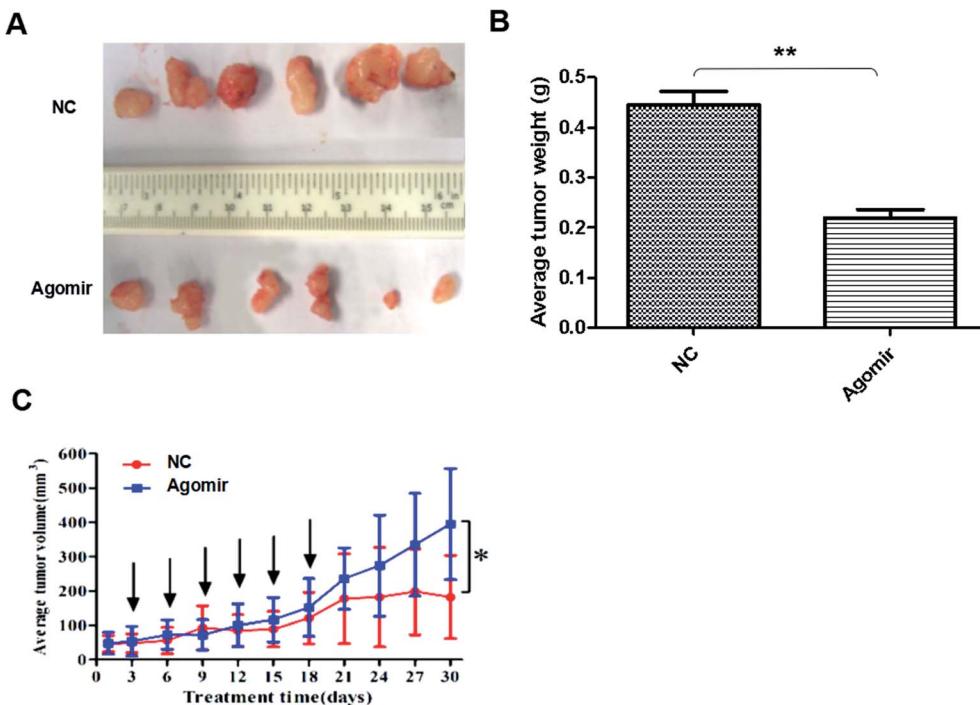
Fig. 8 MiR-425 suppressed cell invasion of TNBC cell lines. Moreover, the inhibitors of miR-425 promoted invasion ability of TNBC cell lines, but the mimics of miR-425 suppressed the invasion ability of TNBC cell lines.

TNBC-derived growth factor (HDGF). It indicated that miR-34 acted as a tumor suppressor in NPC, and may be a novel therapeutic target for the treatments of patients with NPC.<sup>45</sup> Moreover, over-expression of miR-425 is induced by IL-1 $\beta$ , which aim at 3'UTR of miR-425, could regulate homolog expression of tensin and phosphatase. The activation of NF- $\kappa$ B induced by IL-1 $\beta$  up-regulates miR-425 expression, while enhances transcription of miR-425 gene. Therefore, the proliferation of gastric cancer cell is promoted by miR-425 through suppressing tensin homolog and phosphatase. In summary, it confirm that over-expression of miR-425 dependent on NF- $\kappa$ B plays the critical role for the above process.<sup>46</sup> EMT could induce extracellular cytokine TNF- $\alpha$ , TGF- $\beta$ , PDGF, HGF or bFGF to bond with its receptor in BC, and then activate intracellular effectors, such as Ras, NF- $\kappa$ B, Smads, etc. Therefore, in this study, miRNAs were ascertained and searched after its effects on EMT and development process of TNBC.

At first, the differential miRNAs expression profiles between tumor and para-tumor tissues were assessed with miRNA-sequencing experiments. Then, we found that, compared with para-tumor tissues, 9 miRNAs were upregulated and 3 miRNAs (included miR-425) were down-regulated in tumor tissues of patients with TNBC. Furthermore, the relative expression level

of miR-425 in non-tumor, para-tumor, and tumor tissues were determined using qRT-PCR method. This work identified relative levels of miR-425 expression in tumor tissues were obviously less than that in para-tumor and non-tumor tissues. However, there was no obvious difference between para-tumor and non-tumor tissues. Consequently, the relative miR-425 expression levels were investigated in two cell lines of human TNBC. The results confirmed that miR-425 expression level was significantly decreased in the basal epithelial phenotype cell lines Ts 578T and mesenchymal phenotypic cell line MDA-MB-231, which indicated that miR-425 down-expression was related to phenotype of EMT.

Therefore, the targets of miR-425 were forecasted with Targetscan, an online software, and TGF- $\beta$ 1 was found that it is the possibly target of 3' UTRs of TGF- $\beta$ 1 mRNA. Then, further experiments were performed and identified that miR-425 could straight bind with 3' UTRs of TGF- $\beta$ 1 mRNA. In a word, miR-425 directly targeted to TGF- $\beta$ 1 mRNAs. Hence, miR-425 could straight control mRNA and the protein expression level of TGF- $\beta$ 1, further affect expression levels of E-cadherin, VE-cadherin, N-cadherin, vimentin,  $\alpha$ -SMA, and SMAD3 protein in TNBC cell lines. Taken together, our study maybe point out a novel aim at treatment for TNBC associated with EMT.



**Fig. 9** MiR-425 agomir suppressed TNBC xenografts. To detect anti-tumor activity induced by miR-425 agomir *in vivo*, human TNBC xenografts were established with MDA-MB-231 cells. Our work demonstrated that the size of tumor (A) and the average tumor weight (B) of control mice was much larger than that of experiment mice treated with agomir of miR-425, and the difference between two groups had statistical significance. Moreover, the xenografts of control groups grew rapidly, but in the experiment group, which was treated with agomir of miR-425, grew slowly *in vivo* (C). The data are presented as means  $\pm$  SD from three independent experiments. \*P < 0.05, \*\*P < 0.01.

Besides, to assess the biofunctions of miR-425, several molecular functional tests were performed in TNBC cell lines transfected with mimics or inhibitors of miR-425, respectively. It identified that miR-425 suppressed proliferation process and migration or invasion ability of TNBC cell lines, significantly promoted the expressions of epithelial markers E-cadherin and VE-cadherin, suppressed mesenchymal markers expression of N-cadherin, vimentin,  $\alpha$ -SMA, total and phosphorylated SMAD3 (p-SMAD 3) in TNBC cell lines. Our results demonstrated that miR-425 targets to TGF- $\beta$ 1, was a crucial mediator of EMT and development of TNBC through regulating TGF- $\beta$ 1/SMAD3 signaling pathway. Moreover, we did not find that it is involved in well-known molecules such as mTOR, Ki-67, *etc.*

Furthermore, the effects on Human TNBC xenografts and its mechanism were investigated using miR-425 agomir. Our results confirmed that the xenografts of control groups grew rapidly. But in the experiment group, which treated with agomir of miR-425, the xenografts grew slowly *in vivo*. Moreover, the size and weight of tumors in experiment group (treated with agomir of miR-425) was significantly less than that of control group. At the time of end, the average tumor volume of control group was much larger than that of experiment group, and the difference between the two groups had statistical significance. Therefore, the agomir of miR-425 could obviously inhibit tumor proliferation and its development process. Moreover, treatment with agomir of miR-425 could down-regulate the expression of TGF- $\beta$ 1 and SMAD3 with a statistically significant, while suppressed the expression of mesenchymal cell markers and

promoted epithelial markers E-cadherin and VE-cadherin expression in xenografts.

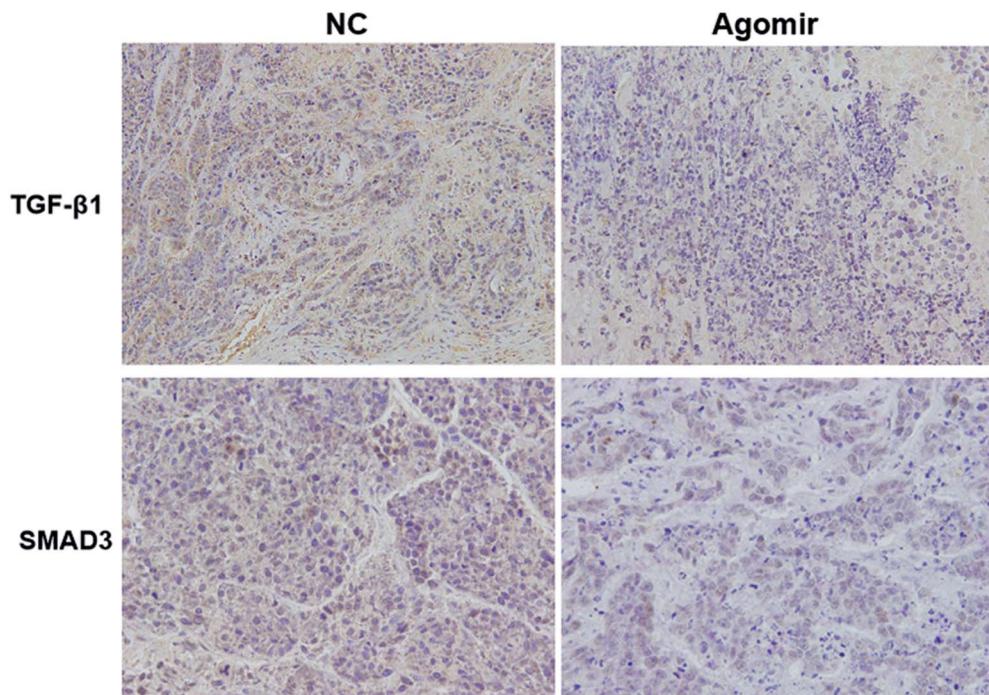
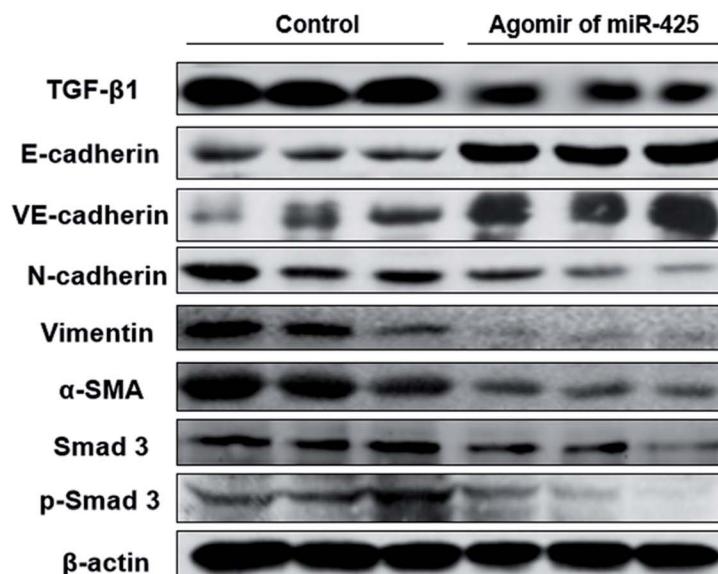
We thought that our manuscript was obviously composed only one part of the whole. The assays *in vivo* and *in vitro* were all separately used to identify our conclusions. In summary, we illustrated that miR-425 targets to TGF- $\beta$ 1. Moreover, miR-425 was a crucial suppressor of EMT and the development of TNBC through inhibiting TGF- $\beta$ 1/SMAD3 signaling pathway. It suggested that aim at TGF- $\beta$ 1/SMAD3 signaling pathway through enhancing miR-425 expression, was a feasible therapy strategy for TNBC.

## Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

## Ethics approval and consent to participate

This investigation and the utilization of human specimens in this work had been approved by the Ethics Committee of our hospitals (Beijing Obstetrics and Gynecology Hospital, Capital Medical University, Beijing, China) on the basis of the Declaration of Helsinki. We clearly confirmed that the informed consents were obtained from all patients. Moreover, the recorded and documented of participant consents were kept in our hospital.

**A****B**

**Fig. 10** The miR-425 agomir suppressed expression of mesenchymal cell markers in xenografts. The expression proteins in xenografts were determined with immunohistochemical staining (A) and western blotting methods (B), respectively. As we can see, the results revealed that when compare with control group (from left to right, control 1–3), treatment with agomir of miR-425 (from left to right, experiment 1–3) could down-regulate the expression of TGF- $\beta$ 1 and SMAD3 with a statistically significant, and suppressed mesenchymal markers expression of xenograft. The mesenchymal markers expression of N-cadherin, vimentin,  $\alpha$ -SMA, SMAD3, p-SMAD3 and endothelial markers E-cadherin, VE-cadherin in xenografts were determined.

All experiments related to animals were performed based on Laboratory Animal Center, Capital Medical University, Beijing, China, Ethical Committee Acts. The experimental mice were dealt with according to the standards supported by

the Animal Protection Committee of Capital Medical University. Moreover, the ethical approval was obtained for the animal experiments conducted in your study from Animal Protection Committee of Capital Medical University.



## Patient consent for publication

This investigation and the utilization of human specimens in this work had been approved by the Ethics Committee of our hospitals (Beijing Obstetrics and Gynecology Hospital, Capital Medical University, Beijing, China) on the basis of the Declaration of Helsinki. We clearly confirmed that the informed consents were obtained from all patients. Moreover, the recorded and documented of participant consents were kept in our hospital.

## Authors' contributions

LYP and ZY conducted the cell and animal experiments and collected the data and wrote the manuscript. LYP, YF, and CJL analyzed and interpreted the data from the experimental results. CJL and ZJQ were involved in the conception and design of the study, and revised the manuscript. All authors have read and approved the final version of the manuscript.

## Funding

This work was supported by grants from the National Natural Science Foundation of China (No. 30600524 and 81341067), and the National Natural Science Foundation of Guangdong Province, China (No. 2017A030313510), Introduction of Talent Fund of Guangdong Second Provincial General Hospital (No. YY2016-006), and Capital Clinical Featured Applied Research and Results Promotion projects (Z161100000516141). The study sponsors had no involvement in the work.

## Conflicts of interest

The authors declare no conflict of interest.

## References

- 1 J. Ferlay, I. Soerjomataram, R. Dikshit, S. Eser, C. Mathers, M. Rebelo, D. M. Parkin, D. Forman and F. Bray, Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012, *Int. J. Cancer*, 2015, **136**, 359–386.
- 2 Y. Xiao, Y. Li, H. Tao, B. Humphries, A. Li, Y. Jiang, C. Yang, R. Luo and Z. Wang, Integrin  $\alpha$ 5 down-regulation by miR-205 suppresses triple negative breast cancer stemness and metastasis by inhibiting the Src/Vav2/Rac1 pathway, *Cancer Lett.*, 2018, **433**, 199–209.
- 3 A. Prat and C. M. Perou, Deconstructing the molecular portraits of breast cancer, *Mol. Oncol.*, 2011, **5**, 5–23.
- 4 S. J. Schnitt, Classification and prognosis of invasive breast cancer: from morphology to molecular taxonomy, *Mod. Pathol.*, 2010, **23**, S60–S64.
- 5 S. B. Zeichner, H. Terawaki and K. Gogineni, A review of systemic treatment in metastatic triple-negative breast cancer, *Breast Cancer*, 2016, **10**, 25–36.
- 6 T. K. Lee, R. T. P. Poon, A. P. Yuen, M. T. Ling, W. K. Kwok, X. H. Wang, Y. C. Wong, X. Y. Guan, K. Man, K. L. Chau and S. T. Fan, Twist Overexpression Correlates with Hepatocellular Carcinoma Metastasis through Induction of Epithelial-Mesenchymal Transition, *Clin. Cancer Res.*, 2006, **12**, 5369.
- 7 G. Giannelli, C. Bergamini, E. Fransvea, C. Sgarra and S. Antonaci, Laminin-5 with Transforming Growth Factor- $\beta$ 1 Induces Epithelial to Mesenchymal Transition in Hepatocellular Carcinoma, *Gastroenterology*, 2005, **129**, 1375–1383.
- 8 Z. Y. Tang, S. L. Ye, Y. K. Liu, L. X. Qin, H. C. Sun, Q. H. Ye, L. Wang, J. Zhou, S. J. Qiu, Y. Li, X. N. Ji, H. Liu, J. L. Xia, Z. Q. Wu, J. Fan, Z. C. Ma, X. D. Zhou, Z. Y. Lin and K. D. Liu, A decade's studies on metastasis of hepatocellular carcinoma, *J. Cancer Res. Clin. Oncol.*, 2004, **130**, 187–196.
- 9 M. A. Huber, N. Kraut and H. Beug, Molecular requirements for epithelial-mesenchymal transition during tumor progression, *Curr. Opin. Cell Biol.*, 2005, **17**, 548–558.
- 10 A. R. Lourenço and P. J. Coffer, SOX4: Joining the Master Regulators of Epithelial-to-Mesenchymal Transition?, *Trends Cancer*, 2017, **3**, 571–582.
- 11 S. Yamada, B. C. Fuchs, T. Fujii, Y. Shimoyama, H. Sugimoto, S. Nomoto, S. Takeda, K. K. Tanabe, Y. Kodera and A. Nakao, Epithelial-to-mesenchymal transition predicts prognosis of pancreatic cancer, *Surgery*, 2013, **154**, 946–954.
- 12 M. A. Huber, N. Azoitei, B. Baumann, S. Grünert, A. Sommer, H. Pehamberger, N. Kraut, H. Beug and T. Wirth, NF- $\kappa$ B is essential for epithelial-mesenchymal transition and metastasis in a model of breast cancer progression, *J. Clin. Invest.*, 2004, **114**, 569–581.
- 13 B. De Craene and G. Berx, Regulatory networks defining EMT during cancer initiation and progression, *Nat. Rev. Cancer*, 2013, **13**, 97–110.
- 14 G. Finak, *et al.*, Stromal gene expression predicts clinical outcome in breast cancer, *Nat. Med.*, 2008, **14**, 518–527.
- 15 M. Sund and R. Kalluri, Tumor stroma derived biomarkers in cancer, *Cancer Metastasis Rev.*, 2009, **28**, 177–183.
- 16 P. Carmeliet and R. K. Jain, Angiogenesis in cancer and other diseases, *Nature*, 2000, **407**, 249–257.
- 17 D. Hanahan and R. A. Weinberg, Hallmarks of cancer: the next generation, *Cell*, 2011, **144**, 646–674.
- 18 L. E. Drake and K. F. Macleod, Tumour suppressor gene function in carcinoma-associated fibroblasts: from tumour cells *via* EMT and back again?, *J. Pathol.*, 2014, **232**, 283–288.
- 19 P. Mallini, T. Lennard, J. Kirby and A. Meeson, Epithelial-to-mesenchymal transition: What is the impact on breast cancer stem cells and drug resistance, *Cancer Treat. Rev.*, 2014, **40**, 341–348.
- 20 R. Kalluri and M. Zeisberg, Fibroblasts in cancer, *Nat. Rev. Cancer*, 2006, **6**, 392–401.
- 21 P. Mallini, T. Lennard, J. Kirby and A. Meeson, Epithelial-to-mesenchymal transition: What is the impact on breast cancer stem cells and drug resistance, *Cancer Treat. Rev.*, 2014, **40**, 341–348.
- 22 E. Dejana, Endothelial cell-cell junctions: happy together, *Nat. Rev. Mol. Cell Biol.*, 2004, **5**, 261–270.



23 J. M. Zarzynska, Two faces of TGF-beta1 in breast cancer, *Mediators Inflammation*, 2014, **2014**, 141747.

24 L. Y. Jia, M. K. Shanmugam, G. Sethi and A. Bishayee, Potential role of targeted therapies in the treatment of triple-negative breast cancer, *Anticancer Drugs*, 2016, **27**, 147–155.

25 O. Iriondo, Y. Liu, G. Lee, M. Elhodaky, C. Jimenez, L. Li, J. Lang, P. Wang and M. Yu, TAK1 mediates microenvironment-triggered autocrine signals and promotes triple-negative breast cancer lung metastasis, *Nat. Commun.*, 2018, **9**, 1994.

26 N. Erin, N. Ogan and A. Yerlikaya, Correction to: Secretomes reveal several novel proteins as well as TGF- $\beta$ 1 as the top upstream regulator of metastatic process in breast cancer, *Breast Cancer Res. Treat.*, 2018, **170**, 251–256.

27 N. E. Bhola, J. M. Balko, T. C. Dugger, M. G. Kuba, V. Sanchez, M. Sanders, J. Stanford, R. S. Cook and C. L. Arteaga, TGF-beta inhibition enhances chemotherapy action against triple-negative breast cancer, *J. Clin. Invest.*, 2013, **123**, 1348–1358.

28 B. Feng, Y. Cao, S. Chen, X. Chu, Y. Chu and S. Chakrabarti, miR-200b Mediates Endothelial-to-Mesenchymal Transition in Diabetic Cardiomyopathy, *Diabetes*, 2016, **65**, 768–779.

29 L. Zhang, D. Huang, Q. Wang, D. Shen, Y. Wang, B. Chen, J. Zhang and L. Gai, MiR-132 inhibits expression of SIRT1 and induces pro-inflammatory processes of vascular endothelial inflammation through blockade of the SREBP-1c metabolic pathway, *Cardiovasc. Drugs Ther.*, 2014, **28**, 303–311.

30 A. Kozomara and S. Griffiths-Jones, miRBase: integrating microRNA annotation and deep-sequencing data, *Nucleic Acids Res.*, 2011, **39**, D152–D157.

31 M. M. Martin, Cutadapt removes adaptor sequences from high-throughput sequencing reads, *EMBnet J.*, 2011, **17**, 10–12.

32 B. Langmead, C. Trapnell, M. Pop and S. L. Salzberg, Ultrafast and memory-efficient alignment of short DNA sequences to the human genome, *Genome Biol.*, 2009, **10**, R25.

33 W. Anna, S. Michal, K. Oskar, G. Wojciech, M. Monika, K. Monika, *et al.*, Next generation sequencing reveals microRNA isoforms in liver cirrhosis and hepatocellular carcinoma, *Int. J. Biochem. Cell Biol.*, 2014, **53**, 208–217.

34 W. Zhu, Y. Ma, X. Zhuang and X. Jin, MicroRNA-425 is downregulated in nasopharyngeal carcinoma and regulates tumor cell viability and invasion by targeting TNBC-derived growth factor, *Oncol. Lett.*, 2018, **15**, 6345–6351.

35 C. B. Nielsen, N. Shomron, R. Sandberg, E. Hornstein, J. Kitzman and C. B. Burge, Determinants of targeting by endogenous and exogenous microRNAs and siRNAs, *RNA*, 2007, **13**, 1894–1910.

36 C. Foroni, M. Broggini, D. Generali and G. Damia, Epithelial–mesenchymal transition and breast cancer: role, molecular mechanisms and clinical impact, *Cancer Treat. Rev.*, 2012, **38**, 689–697.

37 M. Tu, Z. Li, X. Liu, N. Lv, C. Xi, Z. Lu, J. Wei, G. Song, J. Chen, F. Guo, *et al.*, Vasohibin 2 promotes epithelial–mesenchymal transition in human breast cancer *via* activation of transforming growth factor  $\beta$  1 and hypoxia dependent repression of GATA-binding factor 3, *Cancer Lett.*, 2017, **388**, 187–197.

38 R. B. Runyan and P. Savagner, Epithelial–mesenchymal transition and plasticity in the developmental basis of cancer and fibrosis, *Dev. Dyn.*, 2018, **247**, 330–331.

39 D. L. Gibbons and C. J. Creighton, Pan-cancer survey of epithelial–mesenchymal transition markers across the Cancer Genome Atlas, *Dev. Dyn.*, 2017, **247**, 555–564.

40 H. Ungefroren, D. Witte and H. Lehnert, The role of small GTPases of the Rho/Rac family in TGF- $\beta$ -induced EMT and cell motility in cancer, *Dev. Dyn.*, 2017, **247**, 451–461.

41 H. Xia, L. L. Ooi and K. M. Hui, MicroRNA-216a/217-induced epithelial–mesenchymal transition targets PTEN and SMAD7 to promote drug resistance and recurrence of liver cancer, *Hepatology*, 2013, **58**, 629–641.

42 L. Zhai, C. Ma, W. Li, S. Yang and Z. Liu, miR-143 suppresses epithelial–mesenchymal transition and inhibits tumor growth of breast cancer through down-regulation of ERK5, *Mol. Carcinog.*, 2016, **55**, 1990–2000.

43 H. Škovierová, T. Okajčeková, J. Strnádel, E. Vidomanová and E. Halašová, Molecular regulation of epithelial–to–mesenchymal transition in tumorigenesis (review), *Int. J. Mol. Med.*, 2018, **41**, 1187–1200.

44 S. IXing, W. Yu, X. Zhang, Y. Luo, Z. Lei, D. Huang, J. Lin, Y. Huang, S. Huang, F. Nong, C. Zhou and G. Wei, Soviolanthin Extracted from *Dendrobium officinale* Reverses TGF- $\beta$ 1-Mediated Epithelial–Mesenchymal Transition in Hepatocellular Carcinoma Cells *via* Deactivating the TGF- $\beta$ /Smad and PI3K/Akt/mTOR Signaling Pathways, *Int. J. Mol. Sci.*, 2018, **19**(6).

45 G. H. Huang, M. Y. Dua, H. M. Zhua, N. Zhang, Z. W. Lua, L. X. Qiana, W. J. Zhang, X. H. Tians, H. Xia and L. Yina, MiRNA-34a reversed TGF- $\beta$ -induced epithelial–mesenchymal transition *via* suppression of SMAD4 in NPC cells, *Biomed. Pharmacother.*, 2018, **106**, 217–224.

46 J. Ma, J. Liu, Z. Wang, X. Gu, Y. Fan, W. Zhang, L. Xu, J. Zhang and D. Cai, NF- $\kappa$ B-dependent microRNA-425 upregulation promotes gastric cancer cell growth by targeting PTEN upon IL-1 $\beta$  induction, *Mol. Cancer*, 2014, **13**, 40.

