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ASK1-p38 cascaded signal mediates pulmonary microvascular endothelial barrier injury induced by the return of PHSMML in rats

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The return of post-hemorrhagic shock mesenteric lymph (PHSMML) induces pulmonary vascular endothelial barrier dysfunction, which results in acute lung injury. Activation of the apoptosis signal-regulated kinase 1 (ASK1) and p38 mitogen-activated protein kinase (p38 MAPK) pathway has been shown to trigger inflammatory responses. However, whether the ASK1-p38 MAPK pathway is involved in the PHSMML-induced pulmonary endothelial barrier dysfunction remains unclear. In the present study, permeability changes of pulmonary capillaries were found *in vivo*, and activation of the ASK1-p38 MAPK pathway was determined *in vitro*. PMVEC barrier dysfunction was determined by measuring TEER. Furthermore, junctional and cytoskeletal protein expressions were analyzed. The results showed that hemorrhagic shock led to a marked increase in the permeability of pulmonary capillaries *in vivo*, which was markedly alleviated by PHSMML drainage. In cultured pulmonary microvascular endothelial cells (PMVECs), PHSMML reduced the endothelial barrier function accompanied by upregulated p-ASK1 and p-p38 MAPK protein expression, impaired the cytoskeletal protein structure, and down-regulated junction protein expression. These adverse effects were eliminated by applying either Trx1 (ASK1 inhibitor) or SB203580 (p38 MAPK inhibitor). These results indicated that the ASK1-p38 MAPK pathway mediates PHSMML-induced pulmonary vascular endothelial barrier dysfunction during hemorrhagic shock.

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Introduction

Acute lung injury (ALI), challenged by lipopolysaccharide (LPS) through oxidative, inflammatory, and apoptotic effects, is a leading cause of death in severe hemorrhagic shock.^{1–3} Previous studies have shown that hemorrhagic shock-induced gut ischemia leads to gut barrier dysfunction, with the anti-inflammatory cytokines and bacterial toxins of gut flora directly carried into lymphatic vessels and the lungs to further cause ALI deterioration.^{4,5} Furthermore, intravenous infusion of post-hemorrhagic shock mesenteric lymph (PHSMML) directly increases the endothelial cell layer permeability of pulmonary capillaries, which results in vascular endothelial barrier dysfunction.⁶ These results suggest that the backflow of mesenteric lymph derived from ischemic gut is a primary cause of ALI induced by hemorrhagic shock.⁷

The barrier arrangement of pulmonary microvascular endothelial cells (PMVECs) is not identical to the capillary endothelial cells of other organs.^{8,9} The cytoskeleton that supports the PMVEC monolayer barrier mainly consists of three primary elements, namely, actin microfilaments, intermediate filaments, and microtubules.¹⁰ The endothelial barrier receives

signals from extracellular matrix (ECM) fibrous proteins linked to junction proteins between the endothelium.¹¹ Endothelial barrier integrity is stabilized by several structural proteins (inside and outside the membrane), including ZO-1, VE-cadherin, claudin-1, and F-actin.^{12–14} Therefore, altering the cytoskeleton elements or tight junction protein would lead to changes in pulmonary microvascular endothelial barrier function.

Many harmful factors produced by hemorrhagic shock can impair the cytoskeleton structure and reduce the barrier properties.¹⁵ The mitogen-activated protein kinase (MAPK) family plays a vital role in barrier dysfunction through the apoptosis pathway.¹⁶ There are three classical types of MAPK pathway activation cascade, namely, p38 MAPK, extracellular-signal-regulated kinase (ERK), and c-Jun N-terminal kinase (JNK).¹⁷ Apoptosis signal-regulated kinase 1 (ASK1) activation plays an important role in the apoptosis pathway, and can activate the p38 MAPK pathway, which induces local production of cytokines to aggravate the inflammatory cascade, and disorganize the cytoskeleton of distinctive cells.^{18,19} Although activation of the ASK1-p38 MAPK pathway is involved in the cytoskeleton-mediated endothelial barrier function, it remains unclear whether this pathway also mediates PHSMML-induced pulmonary microvascular endothelial barrier dysfunction following hemorrhagic shock. Therefore, the present study aimed to

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determine the impairing effect of PHSMML on the pulmonary microvascular endothelial barrier, and confirm the mediating role of the ASK1-p38 MAPK pathway in PHSMML-induced pulmonary endothelial barrier dysfunction.

Materials and methods

Pulmonary capillary permeability

A hemorrhagic shock model was established and PHSMML was drained as described previously.²⁰ The supernatant of PHSMML was centrifuged and stored at -80°C for subsequent cell experiments. A 10 mm^2 window was then opened in the right thoracic wall of mechanically ventilated Wistar rats.²¹ To measure the permeability of pulmonary capillaries *in vivo*, FITC-labeled albumin (50 mg kg^{-1}) was injected into the femoral vein.²² Using a digital camera system (dynamic *in vivo* imaging of microvasculature and perfusion by fluorescence microscopy, Gene & I, LTD), images were scanned and recorded, followed by analysis using Image-Pro Plus 6.0 software. Five fields were selected for evaluation in each rat. The A_o/A_i ratio (fluorescent intensity outside or inside of the alveolar capillary) was calculated as an indicator of FITC-albumin leakage (compared with the baseline). This study was performed in strict accordance with the NIH guidelines for the care and use of laboratory animals (NIH Publication no. 85-23 Rev. 1985) and was approved by the Institutional Animal Care and Use Committee of Hebei North University (Zhangjiakou, China).

Morphological observations

Rat PMVECs (PriCells, China) were cultured at 37°C in a humidified atmosphere of $5\%\text{ CO}_2$ – $95\%\text{ air}$ (v/v) in Dulbecco's modified Eagle's medium (DMEM, GIBCO, USA) supplemented with 10% (v/v) fetal bovine serum (FBS, GIBCO, USA), 1% antibiotic (penicillin, $10\,000\text{ U mL}^{-1}$; streptomycin, $10\text{ }\mu\text{g mL}^{-1}$; GIBCO), and Microvascular Growth Supplement (MVGS, GIBCO, USA). The PMVECs grew to contact-inhibited monolayers with a typical cobblestone morphology and were then passaged by trypsinization (0.25% trypsin/ 0.02% EDTA) and resuspended in fresh culture medium. The cells (80% confluence) were incubated in FBS-free DMEM medium for 24 h before treatment. Subsequent treatment with FBS-free low-glucose DMEM medium, PHSMML, lipopolysaccharide (LPS, $10\text{ }\mu\text{g mL}^{-1}$), PHSMML + TRX1 (ASK1 inhibitor, 10 ng mL^{-1}) or PHSMML + SB203580 (p38 MAPK inhibitor, $10\text{ }\mu\text{mol L}^{-1}$) was performed for 6 h . Images of PMVECs were obtained using a Leica inverted phase-contrast microscope and recorded at magnifications of $50\times$ to $200\times$, to observe the role of the ASK1-p38 MAPK pathway in PHSMML-induced PMVEC injury.

Monolayer permeability

To assess the role of the ASK1-p38 pathway in the PHSMML-induced increase in the PMVEC monolayer permeability, PMVECs were seeded at 1×10^5 cells per cm^2 on Costar Transwell inserts ($0.4\text{ }\mu\text{m}$ pores, surface area 0.3 cm^2) using DMEM medium with different treatments. Overnight incubation ensured temperature and pH equilibration, and prevented

transient electrical fluctuations, which might occur after media changes or repetitive measurements. Suspension cells ($100\text{ }\mu\text{L}$) were added into the apical compartment, and FBS-free low glucose DMEM medium ($600\text{ }\mu\text{L}$) was added into the basolateral compartment. When the cells were fully fused to form a dense monolayer, we replaced the medium with various factors, as listed above, for 6 h . Primarily, DMEM medium containing FITC-albumin ($100\text{ }\mu\text{L}$, 1 mg mL^{-1}) was added to exchange the medium of the apical compartment. The cells were then incubated for 45 min in the dark. The trans-epithelial electrical resistance (TEER) was measured using an epithelial voltohmmeter (WPI, USA). The media were collected from the apical ($2\text{ }\mu\text{L}$) and basolateral ($100\text{ }\mu\text{L}$) compartments of the 96 well plate, with that from the apical compartments diluted with $98\text{ }\mu\text{L}$ DMEM medium (50 -fold dilution), while that from the basolateral compartments was not diluted. The fluorescence intensity of each sample was measured using a Spectramax M3 fluorescence microplate reader (EX: 485 nm , EM: 525 nm). Apparent permeability coefficients (P_a) were estimated according to the following equation: $P_a = ([A]/t) \times (1/A) \times (v/[L])$, where $[A]$ is the fluorescence absorbance of the basolateral compartment, t is time (s), A is the surface area of the chamber filter (cm^2), v is the liquid volume of the basolateral compartment (mL), and $[L]$ is the fluorescence absorbance of the apical compartment. All permeability and adsorption values were means of at least triplicate measurements using separate filters.

Western blotting

Membrane and cytosolic proteins were isolated with a Membrane and Cytosol Extraction Kit (Beyotime, China). Protein concentrations were determined using a BCA Protein Assay Kit (Beyotime, China). Equal amounts of proteins were separated by 10% SDS gel electrophoresis under denaturing and nonreducing conditions, and then transferred to a PVDF membrane. The membrane was blocked with 5% non-fat milk in Tris-buffered saline and 1% Tween 20 (TBST) at room temperature for 1 h , and then incubated with primary antibody at 4°C overnight. After washing in TBST, the membrane was incubated with the secondary antibody. Membranes were exposed to enhanced chemiluminescent reagents (ECL Plus Western Blotting Detection System, Beyotime, China). Images were captured and analyzed using an ImageQuant LAS 4000 instrument.

Laser scanning confocal microscopy

PMVECs were seeded on sterile chamber slides at $5 \times 10^4/\text{mL}$ and cultured. After 6 h , cells were fixed for 30 min in 4% paraformaldehyde, permeabilized in 0.5% Triton X-100 for 30 min , and rinsed in PBS for 5 min . For F-actin staining, cells were incubated with FITC-conjugated phalloidin ($5\text{ }\mu\text{g mL}^{-1}$) for 20 min at room temperature in the dark, and then washed with PBS. Phalloidin binds to F-actin. Nuclei were stained with DAPI staining solution ($10\text{ }\mu\text{g mL}^{-1}$) for 5 min at room temperature and washed with PBS. The cells were visualized with a laser scanning confocal microscope (Nikon CIsi, Japan). Transverse sections were imaged from the dorsal surface down to the cell-



substratum interface at 2 μm intervals. F-actin protein expression was also detected with western blotting.

Statistical analysis

Data were calculated as means \pm standard deviation (SD). One-way analysis of variance (ANOVA) was used to assess significant differences between groups. $P < 0.05$ was considered significant. All analyses were performed using SPSS version 17.0 (SPSS, Chicago, USA).

Results

Permeability changes of rat pulmonary capillaries

In this study, we used the dynamics of FITC-labeled albumin leakage to assess the permeability of pulmonary capillaries, as shown in Fig. 1. The permeability was significantly increased in the shock group. In contrast, increased permeability was markedly alleviated in the shock with drainage group.

pASK1-p38 MAPK protein activation of PMVECs

To confirm that increased permeability was linked to the ASK1-p38 MAPK pathway, we examined the p-ASK1 and p-p38 MAPK protein expression of PMVECs treated with PHSM and/or inhibitors of ASK1 and p-38 MAPK. As shown in Fig. 2, PHSM increased the activation of p-ASK1 and p-p38 MAPK proteins, which were reduced by co-administration of either TRX1 or SB203580.

Morphological changes of PMVECs

PMVECs grew vigorously to create a cobblestone wall with well-defined margins in the control group (Fig. 3A and F). However, PMVECs treated with either PHSM or LPS exhibited increased

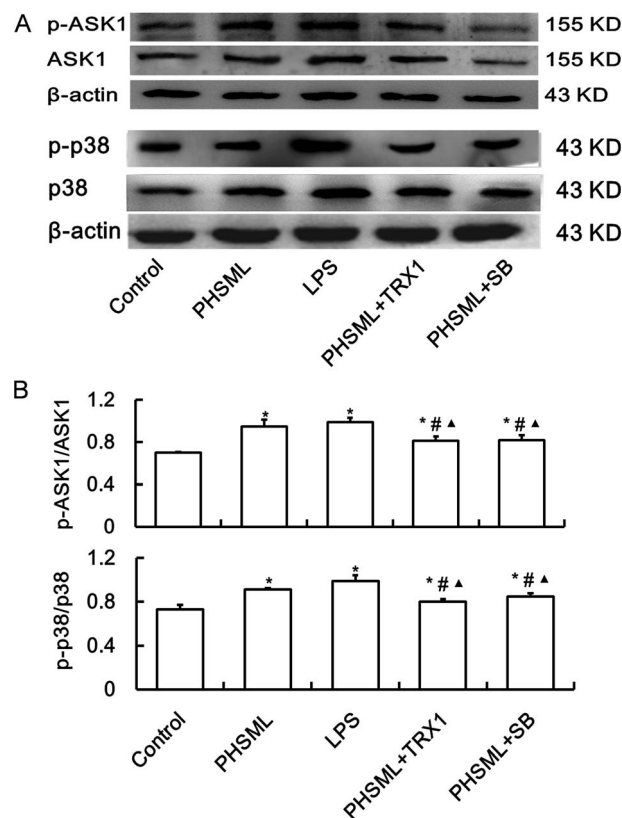


Fig. 2 Effects of TRX1 and SB203580 on the p-ASK1 and p-p38 MAPK expressions in PMVECs treated with PHSM. (A) Images of western blotting for ASK1 and p38 MAPK. (B) Relative density analysis of concentration. Results are presented as the mean \pm SD ($n = 3$). * $P < 0.05$ vs. control group, # $P < 0.05$ vs. PHSM group, $\Delta P < 0.05$ vs. LPS group.

cell gaps and undifferentiated cell numbers (Fig. 3B, G, C and H). These treatments also restricted cell growth, kept cells in suspension, and altered the cytoskeleton of individual cells. Co-administration of either TRX1 or SB203580 with PHSM reversed the PHSM-induced adverse effects (Fig. 3D, I, E and J).

Permeability changes of PMVECs

To examine the effects of PHSM and the ASK1-p38 MAPK pathway on PMVEC barrier dysfunction, the permeability of the PMVEC monolayer was determined by measuring TEER

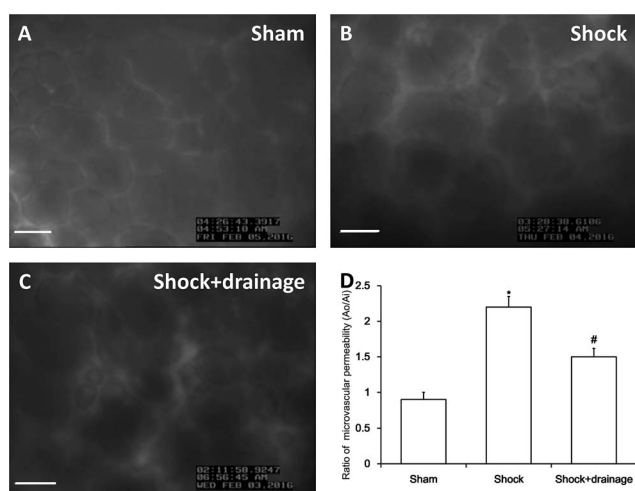


Fig. 1 Permeability changes of pulmonary capillaries following hemorrhagic shock in rats. (A–C) Images of FITC-albumin leakage from pulmonary capillaries under fluorescence microscopy among the three groups, bar = 50 μm . (D) Ratio of FITC-albumin leakage in different groups. Results are presented as means \pm SD ($n = 3$). * $P < 0.05$ vs. sham group, # $P < 0.05$ vs. shock group.

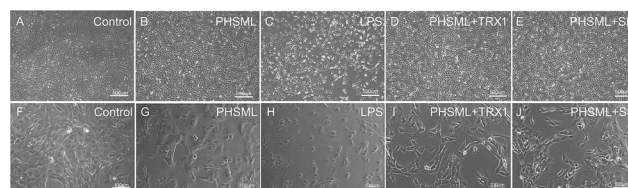


Fig. 3 Effects of TRX1 and SB203580 on the morphology changes of PMVECs treated with PHSM. (A–J) Images of control, PHSM, lipopolysaccharide (LPS), PHSM + TRX1 (ASK1 inhibitor) or PHSM + SB203580 (p38 MAPK inhibitor) incubated for 6 h (inverted microscope, $\times 50$ and $\times 200$).



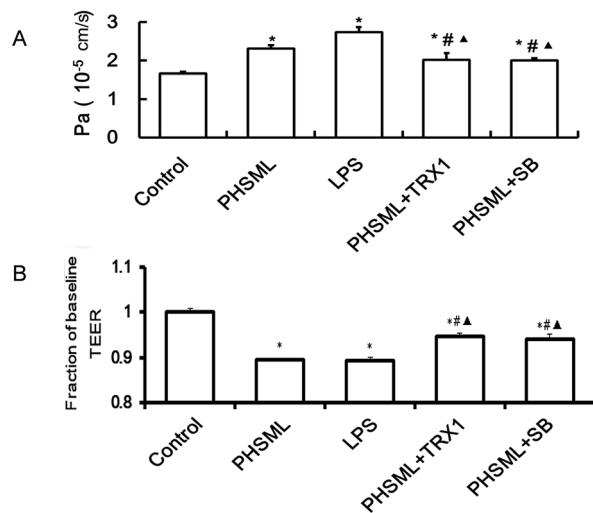


Fig. 4 Effects of TRX1 and SB203580 on the permeability of PMVECs treated with PHSM. (A) Permeability coefficient of the PMVEC barrier. (B) TEER of the PMVEC barrier. Results are presented as means \pm SD ($n = 3$). * $P < 0.05$ vs. control group, # $P < 0.05$ vs. PHSM group, ▲ $P < 0.05$ vs. LPS group.

(Fig. 4A) and the permeability coefficient (Fig. 4B). Both PHSM and LPS significantly decreased the TEER ($P < 0.05$) on the PMVEC monolayer, reflecting that the endothelial barrier was compromised. The concurrent application of either TRX1 or SB203580 significantly attenuated the TEER ($P < 0.05$) in response to either PHSM or LPS. Similarly, by measuring the permeability coefficient, the higher permeability of the PMVEC monolayer was evident during treatment with PHSM, which was inhibited by co-administration of either TRX1 or SB203580.

Junction protein expressions of PMVECs

As PHSM and ASK1-p38 MAPK pathways play a role in the permeability of the PMVEC monolayer, we further confirmed

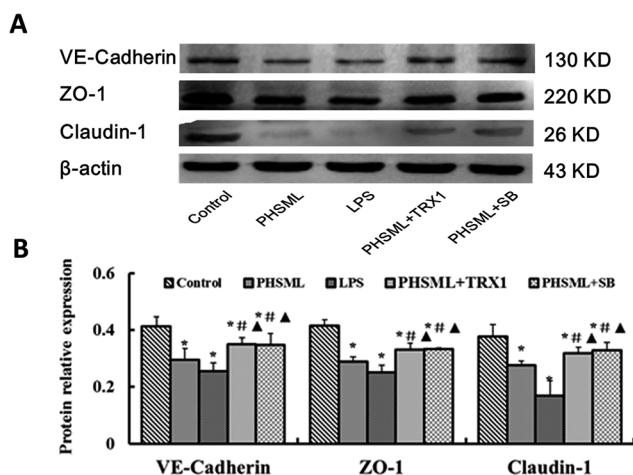


Fig. 5 Effects of TRX1 and SB203580 on junction protein expression in PMVECs treated with PHSM. (A) Images of western blotting for VE-cadherin, ZO-1, and claudin-1. (B) Relative density analysis of concentration. Results are presented as means \pm SD ($n = 3$). * $P < 0.05$ vs. control group, # $P < 0.05$ vs. PHSM group, ▲ $P < 0.05$ vs. LPS group.

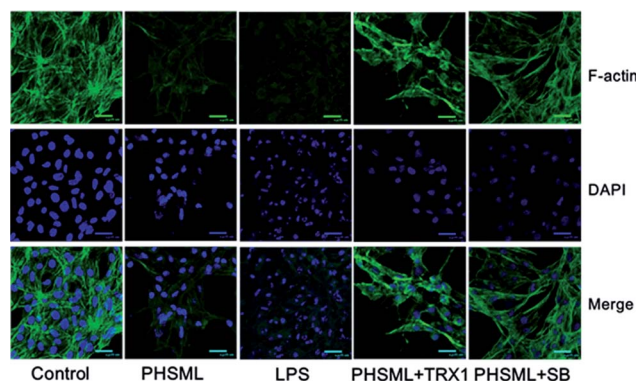


Fig. 6 Effects of TRX1 and SB203580 on F-actin expression in PMVECs treated with PHSM. F-actin was stained with phalloidin FITC (green); nuclei were stained with DAPI (blue); cells co-expressed F-actin and DAPI in the merged image.

the role of the ASK1-p38 MAPK pathway in barrier dysfunction. Changes in the junction protein expression were determined. We analyzed the expressions of proteins (VE-cadherin, ZO-1, and claudin-1) by western blotting (Fig. 5). Compared with the control group, PHSM or LPS dramatically reduced the expression of three junction proteins ($P < 0.05$). These reduced protein expressions were restored by treatment with either TRX1 or SB203580 ($P < 0.05$).

Cytoskeleton expression of PMVECs

Cytoskeleton changes were observed using a combination of F-actin staining with fluorescein-phalloidin and nuclear staining with DAPI. F-actin is situated in the cellular cortex in the cells. This localization is observable in cells, which present a strong accumulation of actin filaments underneath the plasma membrane. The central region of the growth cone contains a F-actin meshwork, because F-actin bundles are an essential part of motility. F-actin was stained with phalloidin and analyzed using confocal microscopy. Representative confocal images of phalloidin and DAPI co-stained in PMVECs are shown as Fig. 6. Both PHSM and LPS decreased phalloidin staining and destroyed the nuclei compared with the control group, whereas treatment of either TRX1 or SB203580 with PHSM restored the nuclei and increased F-actin expression.

Discussion

Pulmonary microvascular endothelial cells are irreversibly impaired after hypoxic-ischemic injury during hemorrhagic shock, which induces capillary leak syndrome or a large area of tissue edema (easily causing MOD).^{4,20,23} Activation of the ASK1-p38 MAPK pathway induced p38 MAPK phosphorylation and enhanced the inflammatory response and apoptosis, leading to ALI.^{24,25} In contrast, inhibition of shock mesenteric lymph reflux can reduce pulmonary vascular permeability and relieve ALI.⁷ We have previously demonstrated that the permeability of pulmonary capillaries was increased in shock rats (lung wet-to-dry weight ratio, Evan's blue dye technique).²⁰ In the present



study, we found that blocking the ASK1-p38 MAPK pathway abolished the impairing effects of PHSMML on PMVEC barrier. These results indicated that ASK1-p38 MAPK pathway activation mediated the impairing effects of PHSMML on the PMVEC barrier.

Mesenteric lymph return can trigger a systemic inflammatory response and ALI by pro-inflammatory mediators in hemorrhagic shock.²⁶ The MAPK pathway has been reported to play a major role in this process, especially the p38 MAPK pathway.²⁷ Suppressing the p38 MAPK pathway reduces lung dysfunction following hemorrhagic shock, but whether hemorrhagic shock directly upregulates the expression of p38 MAPK has not been reported previously.²⁸ Furthermore, p38 MAPK signaling has been proposed to be critical for lipopolysaccharide-induced apoptosis in PMVECs.²⁹ ASK1 is an important kinase for apoptosis, and its activation can activate the p38 MAPK pathway.^{18,19} Therefore, it is necessary to establish the relationship between the ASK1-p38 MAPK pathway and PHSMML in pulmonary endothelial barrier dysfunction. Our findings indicated that activation of the ASK1-p38 MAPK pathway mediated PHSMML-induced pulmonary microvascular endothelial barrier dysfunction following hemorrhagic shock. Interestingly, ASK1 expression was also inhibited by p38 inhibitor, although the specific mechanism should be explored in future studies.

To mimic the environment of PMVECs during hemorrhagic shock, we used PHSMML to stimulate PMVECs, and found that PHSMML induced significant morphological changes, which might be related to barrier dysfunction. Blocking of the ASK1-p38 MAPK pathway could reduce these morphological changes. Our results showed that the ASK1-p38 MAPK pathway plays an important role in damage to the PMVEC morphology induced by PHSMML.

PHSMML has been shown to activate the ASK1-p38 MAPK pathway through inflammatory cytokines, which are responsible for several intra-cellular responses.^{19,30} To examine the effect of the ASK1-p38 MAPK pathway on PMVEC barrier dysfunction produced by PHSMML, permeability changes were evaluated by TEER and permeability coefficient experiments. We found that PHSMML could decrease TEER and increase the PMVEC monolayer permeability. Both TRX1 and SB203580 reversed the PHSMML-induced changes in the TEER and PMVEC monolayer permeability. These results indicated that inhibiting ASK1-p38 MAPK significantly reduced PMVEC barrier damage and increased the permeability produced by PHSMML.

Our findings were in agreement with the p38 MAPK pathway being involved in disruption of the tight junction protein induced by LPS.³ PHSMML markedly reduced the expression of ZO-1, claudin-1, and VE-cadherin. Both TRX-1 and SB203580 restored PHSMML-reduced protein expression. The main cytoskeleton constituent was actin filament, which was mainly linked to the junction proteins.^{13,14} Confocal images also showed that the F-actin contents were reduced upon stimulation of the ASK1-p38 MAPK pathway. After inhibition of either SB203580 or TRX-1, the contents were restored to near control levels. Our results supported that F-actin filaments are responsible for junction protein integrity by maintaining the

barrier permeability.³¹ ASK1-p38 MAPK pathway activation led to actin filament derangement and disrupted the barrier properties by changing the junction protein.

Conclusions

We have demonstrated that the ASK1-p38 pathway is strongly correlated with the induction of PHSMML-induced pulmonary microvascular endothelial barrier dysfunction. PHSMML greatly altered the cytoskeletal and junction proteins (F-actin, VE-cadherin, claudin-1, and ZO-1). Inhibition of the ASK1-p38 pathway prevented damages of the PMVEC barrier induced by the PHSMML. These results indicate that the ASK1-p38 pathway mediates PHSMML-induced permeability changes of the PMVEC barrier.

Conflicts of interest

The authors declare no conflicts of interest.

Acknowledgements

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References

- 1 X. Wu, M. G. Schwacha, M. A. Dubick, A. P. Cap and D. N. Darlington, *Shock*, 2016, **46**, 108–114.
- 2 H. Yao, Y. Sun, S. Song, Y. Qi, X. Tao, L. Xu, L. Yin, X. Han, Y. Xu, H. Li, H. Sun and J. Peng, *Front. Pharmacol.*, 2017, **8**, 120.
- 3 S. Aggarwal, C. Dimitropoulou, Q. Lu, S. M. Black and S. Sharma, *Front. Physiol.*, 2012, **3**, 161.
- 4 E. A. Deitch, R. Forsythe, D. Anjaria, D. H. Livingston, Q. Lu, D. Z. Xu and H. Redl, *Shock*, 2004, **22**, 221–228.
- 5 G. Levy, J. E. Fishman, D. Xu, B. T. Chandler, E. Feketova, W. Dong, Y. Qin, V. Alli, L. Ulloa and E. A. Deitch, *Shock*, 2013, **39**, 39–44.
- 6 Y. M. Zhang, S. K. Zhang and N. Q. Cui, *World J. Gastroenterol.*, 2014, **20**, 4771–4777.
- 7 E. A. Deitch, C. Adams, Q. Lu and D. Z. Xu, *Surgery*, 2001, **129**, 39–47.
- 8 M. Shao, S. T. Tang, B. Liu and H. Q. Zhu, *Mol. Med. Rep.*, 2016, **13**, 529–535.
- 9 F. Huber, A. Boire, M. P. Lopez and G. H. Koenderink, *Curr. Opin. Cell Biol.*, 2015, **32**, 39–47.
- 10 S. F. Rodrigues and D. N. Granger, *Tissue Barriers*, 2015, **3**, e978720.
- 11 C. W. Wilson, L. H. Parker, C. J. Hall, T. Smyczek, J. Mak, A. Crow, G. Posthuma, A. De Maziere, M. Sagolla,



- C. Chalouni, P. Vitorino, M. Roose-Girma, S. Warming, J. Klumperman, P. S. Crosier and W. Ye, *Blood*, 2013, **122**, 3678–3690.
- 12 S. C. Nicklisch, J. E. Spahn, H. Zhou, C. M. Gruian and J. H. Waite, *Biochemistry*, 2016, **55**, 2022–2030.
- 13 C. Greene and M. Campbell, *Tissue Barriers*, 2016, **4**, e1138017.
- 14 O. L. Mooren, J. Li, J. Nawas and J. A. Cooper, *Mol. Biol. Cell*, 2014, **25**, 4115–4129.
- 15 S. M. Stamatovic, A. M. Johnson, R. F. Keep and A. V. Andjelkovic, *Tissue Barriers*, 2016, **4**, e1154641.
- 16 J. S. Arthur and S. C. Ley, *Nat. Rev. Immunol.*, 2013, **13**, 679–692.
- 17 C. Jiang, Y. Yuan, F. Hu, Q. Wang, K. Zhang, Y. Wang, J. Gu, X. Liu, J. Bian and Z. Liu, *Biol. Trace Elem. Res.*, 2014, **158**, 249–258.
- 18 W. Li, B. Huang, K. Liu, J. Hou and H. Meng, *J. Periodontol.*, 2015, **86**, 917–926.
- 19 A. Ray, N. Sehgal, S. Karunakaran, G. Rangarajan and V. Ravindranath, *Free Radical Biol. Med.*, 2015, **87**, 312–325.
- 20 G. X. Sun, Y. X. Guo, Y. P. Zhang, L. M. Zhang, Z. G. Zhao and C. Y. Niu, *J. Surg. Res.*, 2016, **203**, 47–55.
- 21 S. E. Gill, R. Taneja, M. Rohan, L. Wang and S. Mehta, *PLoS One*, 2014, **9**, e88501.
- 22 F. Lin, Y. Y. Liu, B. Xu, K. Sun, H. Y. Wang, Q. Li, C. S. Wang, J. Y. Fan, S. W. Zhang and J. Y. Han, *Shock*, 2013, **39**, 317–325.
- 23 A. Gulati, *Curr. Vasc. Pharmacol.*, 2016, **14**, 187–195.
- 24 R. K. Brobey, D. German, P. K. Sonsalla, P. Gurnani, J. Pastor, C. C. Hsieh, J. Papaconstantinou, P. P. Foster, M. Kuro-o and K. P. Rosenblatt, *PLoS One*, 2015, **10**, e0139914.
- 25 J. Wu, R. Wang, Z. Ye, X. Sun, Z. Chen, F. Xia, Q. Sun and L. Liu, *Biochem. Biophys. Res. Commun.*, 2015, **466**, 155–161.
- 26 S. Langness, T. W. Costantini, K. Morishita, B. P. Eliceiri and R. Coimbra, *PLoS One*, 2016, **11**, e0168322.
- 27 J. T. Hsu, Y. C. Hsieh, W. H. Kan, J. G. Chen, M. A. Choudhry, M. G. Schwacha, K. I. Bland and I. H. Chaudry, *Am. J. Physiol.: Heart Circ. Physiol.*, 2007, **292**, H2982–H2987.
- 28 H. Sato, K. Kasai, T. Tanaka, T. Kita and N. Tanaka, *Med. Sci. Monit.*, 2008, **14**, BR79–87.
- 29 Z. F. Liu, D. Zheng, G. C. Fan, T. Peng and L. Su, *Apoptosis*, 2016, **21**, 896–904.
- 30 M. Soga, A. Matsuzawa and H. Ichijo, *Int. J. Cell Biol.*, 2012, **2012**, 439587.
- 31 D. Coronado-Velazquez, A. Betanzos, J. Serrano-Luna and M. Shibayama, *J. Eukaryotic Microbiol.*, 2018, **65**, 804–819.

