


Cite this: *RSC Adv.*, 2019, 9, 3351

High-throughput metabolic profiling, combined with chemometrics and bioinformatic analysis reveals functional alterations in myocardial dysfunction†

Qun Liang,^a Han Liu,^b Xiuli Li,^a Panguo Hairong,^a Peiyang Sun,^a Yang Yang^a and Chunpeng Du^a

High-throughput metabolic profiling technology has been used for biomarker discovery and to reveal underlying metabolic mechanisms. Sepsis-induced myocardial dysfunction (SMD) is a common complication in sepsis patients, and severely affects their quality of life. However, the pathogenesis of SMD is currently unclear, and there has been inadequate basic research. In this study, metabolic profiling was explored by liquid chromatography/mass spectrometry (LC/MS) combined with chemometrics and bioinformatic analysis. The global metabolome data were analyzed using chemometrics analysis including principal component analysis and partial least squares discriminant analysis for significant metabolites. Variable importance for projection values obtained utilizing a pattern recognition method were used to identify potential biomarkers. The differential metabolites were putatively identified using the metabolome database and bioinformatics analysis was conducted via Ingenuity Pathway Analysis (IPA) to predict the likely functional alterations. In total, 21 differential metabolites were found in SMD and these were involved in phenylalanine, tyrosine and tryptophan biosynthesis, arachidonic acid metabolism, glycine, serine and threonine metabolism, and so on. The analysis revealed that the metabolites were strongly related to molecular transport, and small molecule biochemistry metabolic pathways. The present study indicates that high-throughput metabolic profiling, combined with chemometrics and a bioinformatic platform, can reveal the likely functional alterations in disease and could provide more precise and credible information in the basic research of disease pathogenesis.

Received 11th September 2018
Accepted 15th January 2019

DOI: 10.1039/c8ra07572g

rsc.li/rsc-advances

Introduction

Sepsis-induced myocardial dysfunction (SMD) is a leading cause of death in intensive care units (ICU) with high morbidity and mortality worldwide.^{1,2} Numerous studies have indicated that SMD exists in the high output stage of sepsis. Indeed, septic patients with SMD have a 70% chance of mortality compared to a 20% mortality rate for patients without myocardial dysfunction.^{3,4} Multiple mechanisms have been suggested to underlie myocardial dysfunction, including inflammatory responses, myocardial and mitochondrial disorders, and apoptosis.⁵ SMD contributes to a high risk of mortality and is linked to a detrimental prognosis of sepsis.⁶ The molecular mechanism of SMD has been studied with regard to cytokines, apoptosis, immune regulation, mitochondria, and so on.^{7–9} However, the precise

metabolic mechanisms and their role in the pathogenesis of SMD remain incompletely understood.

Clinical studies confirmed that myocardial dysfunction is the main clinical manifestation in sepsis.¹⁰ Therefore, it is critical that we clarify the mechanisms of SMD and explore effective treatments. Lipopolysaccharide (LPS)-induced myocardial injury is an animal model that is commonly used to assess sepsis-induced cardiac dysfunction.¹¹ Experimentally, the administration of LPS to laboratory animals, especially mice, has been widely used to study the mechanisms of SMD.¹² Metabolomics, the study of a biologic process involving all of the metabolites in a whole organism, has been integrated into multidisciplinary methods to profile changes in small molecules associated with disease.^{13–15} Metabolomics has been applied to identify metabolic biomarkers in several human diseases.^{16–18} Ultra-performance liquid chromatography (UPLC) coupled with mass spectrometry (MS) has higher resolution, high sensitivity and rapid separation, and has been widely used to investigate metabolite alterations in disease.^{19,20}

For the reasons given above, we hypothesized that metabolomics is a reliable method to assess LPS-induced SMD in

^aICU Center, First Affiliated Hospital, School of Pharmacy, Heilongjiang University of Chinese Medicine, Heping Road 24, Xiangfang District, Harbin 150040, China. E-mail: qunliangomics@yeah.net

^bSimon Fraser University (SFU), Burnaby, British Columbia, Canada

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c8ra07572g



a mouse model. Accordingly, metabolic profiling was explored by integrating a UPLC/MS platform with bioinformatics analysis and chemometrics analysis. Variable importance for projection values obtained using a chemometrics method were used to identify potential biomarkers. The differential metabolites were putatively identified and bioinformatics analysis was conducted *via* Ingenuity Pathway Analysis (IPA) to identify the predicted pathways and biological functions in a well-established mouse model of SMD.

Experimental methods

Reagents and materials

Acetonitrile, HPLC grade, was provided by Merck (Germany). Leucine enkephalin was supplied from Sigma-Aldrich (St. Louis, MO, USA). Water was supplied from the Milli-Q ultra-pure water system (Millipore, Billerica, USA). Formic acid, HPLC grade, was provided by Tedia (USA). LPS was supplied by Sigma (St. Louis, MO, USA).

Animals

Male C57BL/6J mice (8–10 weeks old, weighing 20–24 g) were obtained from the Animal Research Center of Heilongjiang University of Chinese Medicine. The animals were kept with a 12 h/12 h light/dark cycle at 21–23 °C and fed a standard chow diet *ad libitum* before experiments.

Ethical statement

The experimental procedures were approved by the Animal Care and Ethics Committee at Heilongjiang University of Chinese Medicine and all experiments were performed in accordance with the Declaration of Helsinki.

Lipopolysaccharide injection

LPS administration was used to induce sepsis as previously described.²¹ *E. coli* LPS was dissolved in sterile physiological saline solution (0.9% NaCl) at a concentration of 1 mg mL⁻¹. Forty mice were randomized into the following two groups: the control group ($n = 20$), the treatment group ($n = 20$). The treatment and control animals were injected with either 0.5 mL normal saline solution containing LPS (10 mg kg⁻¹ body weight, i.p.) or 0.5 mL normal saline only. This dose of LPS was based on previous observations of overt myocardial dysfunction.²² At the end of the experiments, blood collection was performed, and the hearts were removed for histological analyses.

Histopathological examination

At the end of the experiment, the whole heart was removed. Whole hearts were fixed in 10% formalin, embedded in paraffin, and sectioned onto slides. Serial sections of 4 µm were cut for Haematoxylin and Eosin staining (H&E) analysis.

Cytokine measurements

Blood was collected from the experimental and control mice to determine the serum levels of cytokines. Samples were allowed

to clot for 2 h at room temperature before centrifugation for 10 min at 2000 g. The serum was then removed and stored at -80 °C for subsequent enzyme-linked immunosorbent assay (ELISA) analysis. The levels of cytokines (tumor necrosis factor- α (TNF- α), interleukin-4 (IL-4)) were examined using a Quantikine Mouse ELISA kit (R&D Systems, MN).

Sample preparation

Sample preparation for the serum metabolomics analysis was conducted as follows: prior to analysis, 100 µL serum samples were thawed at 4 °C, followed by vigorous vortexing for 2 min, centrifugation for 10 min at a speed of 5000 rpm at 4 °C, and the supernatant was then filtered through a 0.22 µm membrane. The supernatant was placed into an auto sampler vial for UPLC/QTOF-MS/MS analysis in positive and negative electrospray ionization (ESI) mode. A mixed sample containing aliquots of all the serum samples was prepared as the quality control (QC) sample. The QC sample was analyzed once for every 10 test samples.

UPLC/QTOF-MS/MS analysis

We transferred a pretreated sample (2 µL) into a 2.1 µm × 100 µm × 1.7 µm C₈ column using a UPLC system (Waters Corp., Milford, USA). The column temperature was maintained at 45 °C. The gradient consisted of 0.1% formic acid in water as mobile phase A and 0.1% formic acid in acetonitrile as mobile phase B. The positive ion and negative ion mode involved 1–15% acetonitrile for 0–3.0 min, 15–50% for 3.0–5 min and 50–98% for 5–10 min; the concentration was held at 98% for 2 min. The injection volume was 3 µL and the flow rate was 0.4 mL min⁻¹. MS analysis was performed using a Waters Micro-mass Q-TOF/MS system (Waters Corp., Milford, USA). The parameters were set up as follows: desolvation temperature of 250 °C, source temperature of 100 °C, capillary voltage of 3000 V, cone voltage of 25 V, extraction cone voltage of 3 V, desolvation gas flow of 600 L h⁻¹, collision energy of 5 eV. MS data were collected from 50 to 1000 m/z in full scan mode.

Metabolite identification

MS/MS experiments were conducted to obtain fragmentation patterns of selected metabolites for determining their structures, and accurate molecular weights were also calculated. Accurate mass data and isotopic distributions for the precursor and product ions were compared to the spectral data. The mass tolerance between measured m/z values and exact mass was set to within 5 ppm. The metabolite product ion spectrum with its structure information was confirmed using MassFragment software. The relevant pathways of potential endogenous metabolites were assessed using the online website MetaboAnalyst 3.0.²³

Metabolic pathway analysis

Ingenuity Pathway Analysis (IPA, Ingenuity® Systems, <http://www.ingenuity.com>, Redwood City, CA, USA) enables bioinformaticians to identify the biological functions which are



most relevant to their work. The “core analysis” module in the IPA analysis system was used to analyze the differential metabolites and build networks of the differentially regulated metabolites. Canonical pathway analysis of those metabolites was also confirmed, with the criterion that a score of higher than 1 was used to obtain the most enriched network.

Statistical analysis

MS data were imported to MarkerLynx (Waters Corporation) for peak detection and alignment. We used the Pareto scaling method for data transformation before multivariate statistical analysis. Chemometrics analyses including principal component analysis (PCA) and orthogonal partial least squares discriminant analysis (OPLS-DA) were performed using EZinfo software (V2, Waters Corp., Milford, USA). VIP values (variable importance for projection) provided by OPLS-DA were used to evaluate the variable contribution and find potential biomarkers. All statistical tests were two-sided, with all statistical analyses performed using SPSS software (Version 17.0, SPSS Inc., Chicago, Illinois). Differences with $p < 0.05$ were considered statistically significant.

Results and discussion

Histological sections of the hearts treated with LPS were analyzed (Fig. S1†). In the control animals, the myocardiocytes

were in healthy shape and were well aligned. The sections demonstrated the occurrence of extravasation of red blood cells, and hyperemia of blood vessels. The treatment group showed more severe damage than the control group (Fig. S1A†). Fig. S1B† shows the levels of TNF- α and IL-4 in serum from mice subjected to LPS-induced sepsis. The treatment group showed significantly increased concentrations of TNF- α and IL-4 in the serum when compared with control mice. In our study, we applied UPLC/MS to collect metabolite information with respect to SMD. Based on the PCA, none of the samples were identified as outliers and excluded from further statistical analysis (Fig. 1 and S2†). The OPLS-DA approach was established to reveal the specific metabolic changes in the treatment and control groups and to improve their separation. The treatment and control mice could be separated into two distinct clusters, which indicated that the metabolic states were different between the treatment and control groups, and as displayed by the score plots. The loading ion plots of the OPLS-DA model built from treatment and control samples are presented in Fig. 2 and S3.†

In this study, an untargeted UPLC/MS technique was performed to investigate the metabolic signatures in serum samples from SMD mice. In total, 3601 variables were selected in the final data table for subsequent analyses. To screen the putative biomarkers for SMD using multivariate analysis, VIP values, which are based on weighted coefficients, were obtained from the OPLS-DA model. The key metabolites, which were

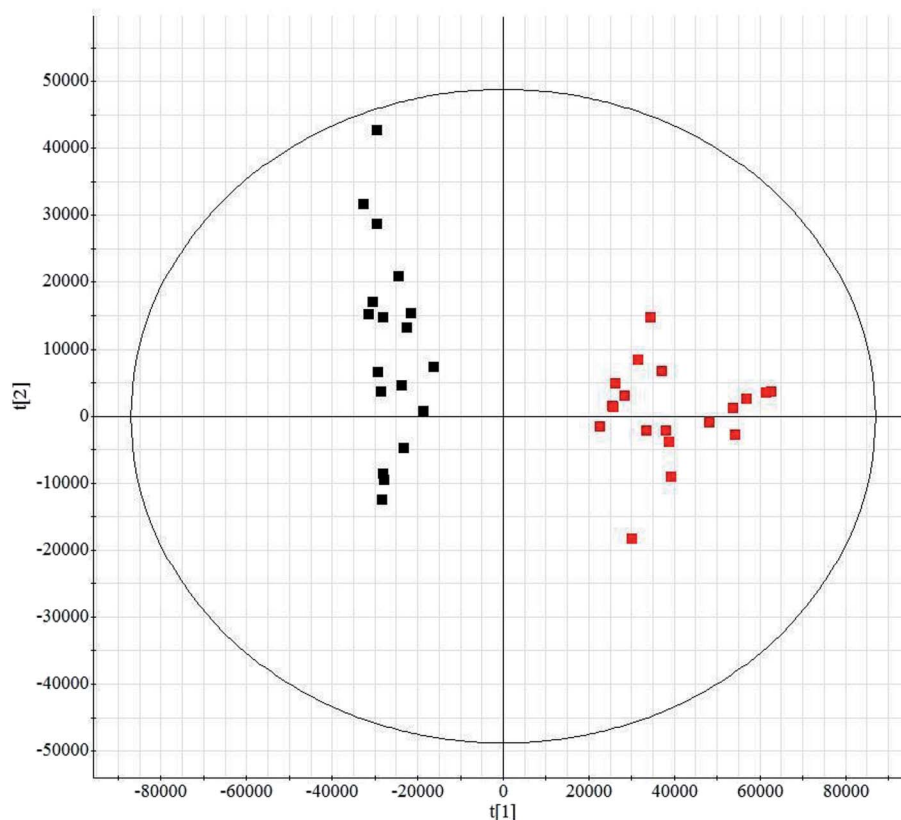


Fig. 1 Principal component analysis score plot of the UPLC/MS serum metabolite data obtained in positive ion mode by liquid chromatography/mass spectrometry. Note: black, control group; red, treatment group.



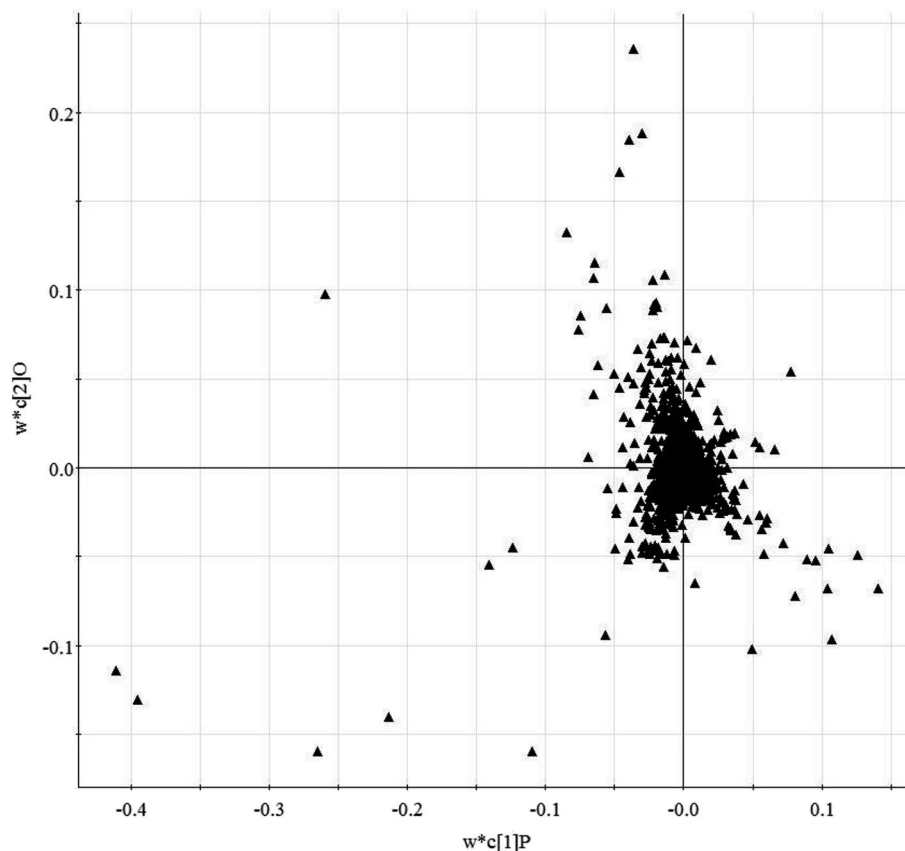


Fig. 2 Loading-plot of the serum metabolite data obtained in positive ion mode by liquid chromatography/mass spectrometry ($R^2X = 0.91$, $R^2Y = 0.77$, $Q^2Y = 0.82$).

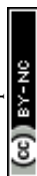
essential for distinguishing between the treatment and control cases, were selected from the VIP plot (Fig. 3 and S4†). A total of 21 potential metabolites such as pyroglutamic acid, 5-aminopentanoic acid, *p*-aminobenzoic acid, citric acid, *L*-tyrosine, pipercolic acid and *L*-methionine were selected, on the basis of the VIP ($VIP > 10$) and Student's *t* test ($p < 0.05$), and significant differences between treatment and control groups were also shown (Table S1†). This suggests that metabolic profiling might be helpful in determining the metabolic phenotype of SMD disease. The change in metabolite concentrations is shown in a heatmap (Fig. 4).

Myocardial dysfunction is a common clinical manifestation in sepsis.²⁴ SMD is being studied extensively. However, its pathogenesis is still unknown. Metabolomics is the non-targeted analysis of global changes in metabolites and a potential tool for discovering new biomarkers for disease.^{25–27} The identified biomarkers of SMD were imported into the MetPA website to analyze the relevant metabolic pathways. A total of 20 metabolic pathways were involved in disturbances relating to phenylalanine, tyrosine and tryptophan biosynthesis, arachidonic acid metabolism, glycine, serine and threonine metabolism, glyoxylate and dicarboxylate metabolism, phenylalanine metabolism, retinol metabolism, and tyrosine metabolism (Impact > 0.1 , Fig. S5, Table S2†). These results indicate that endogenous metabolites demonstrate a strong disturbance in

the whole metabolic profile which is closely connected with the SMD model.

In the present study, IPA was used to analyze the differentiated metabolites and associated network relationships, to examine the metabolic pathways that were most changed. The most significantly altered network was 'amino acid metabolism, molecular transport and small molecule biochemistry' (Fig. 5), with a score above 3. Based on IPA analysis, certain metabolites (1-18:2(9Z,12Z)lysophosphatidylcholine, 20-hydroxyeicosatetraenoic acid, arachidonic acid, citric acid, kynurenic acid, *L*-methionine, *L*-tyrosine, and phenylpyruvic acid) are involved in and play a central role in this network. These data strongly suggest that disturbance within this metabolism is characteristic for SMD.

SMD increases mortality in sepsis, and myocardial dysfunction is the leading cause of death in patients with sepsis, but the underlying pathophysiology of myocardial dysfunction is not completely understood, which limits effective therapy for this disorder. Experimentally, the administration of LPS to mice has been widely used to study the mechanisms of SMD. Despite more than three decades of clinical and basic research on SMD, there is no gold standard for evaluating cardiac function in sepsis. Recent research demonstrated that $TNF-\alpha$ and IL-4 served as indicators of damaged cardiac myocytes in SMD.²⁸ In our study, we observed that two indexes ($TNF-\alpha$ and IL-4)



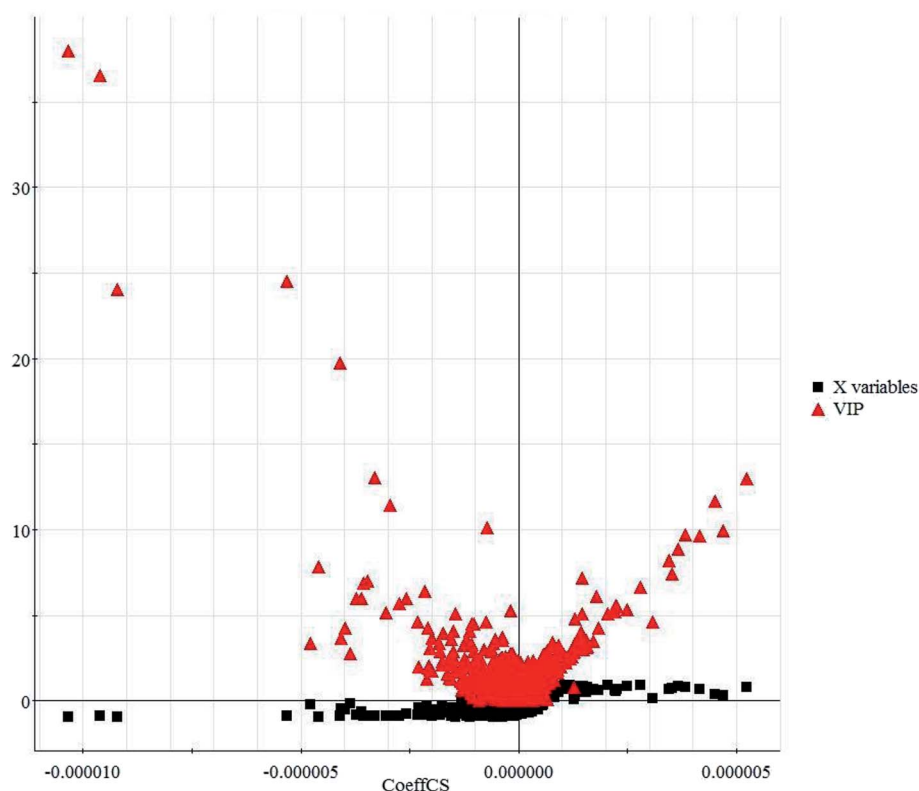


Fig. 3 VIP-plots based on the matched partial least squares discriminant analysis model for the interesting variables in the positive ion mode.

increased in mice after LPS administration when compared with controls. We provide evidence that increased expression of TNF- α and IL-4 in serum from mice may be involved in the pathogenesis of SMD. Taken together, histological and serum

results confirmed that myocardial injury occurred after LPS administration in the mouse model.

Previous research has identified amino acids measured using mass spectrometry in the plasma of septic patients.^{29–33}

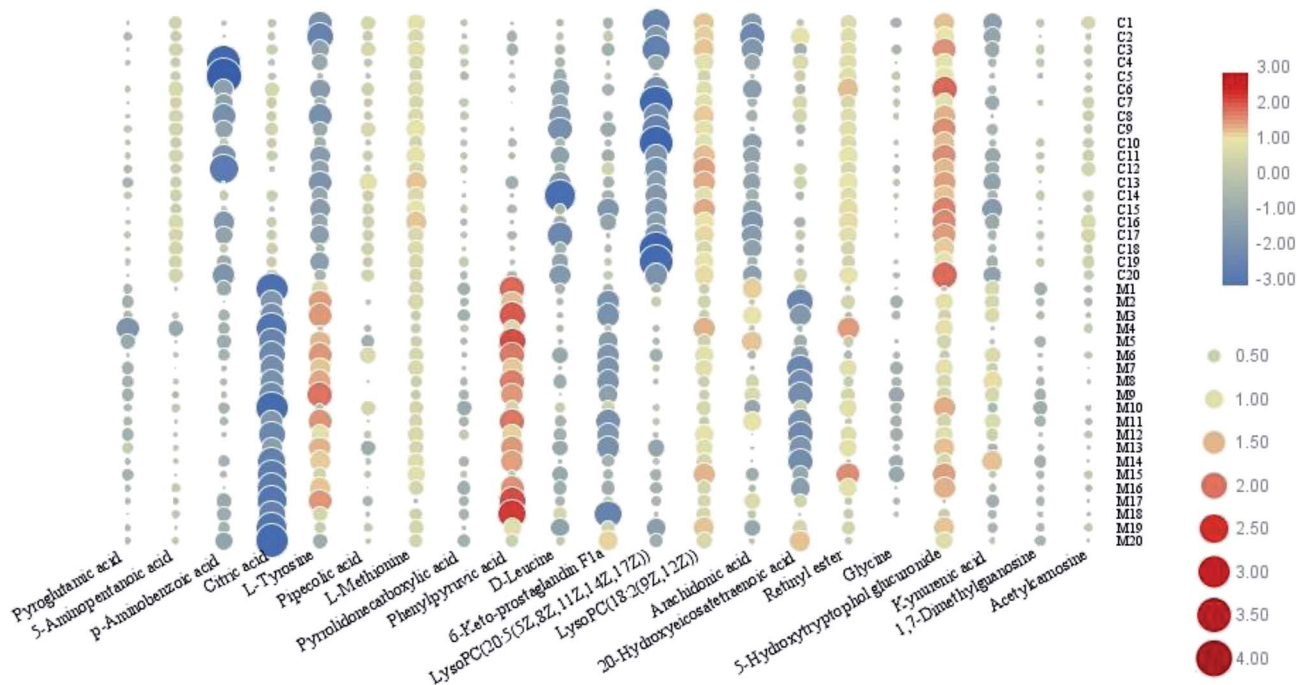


Fig. 4 The relative concentration changes in serum biomarkers in SMD mice.



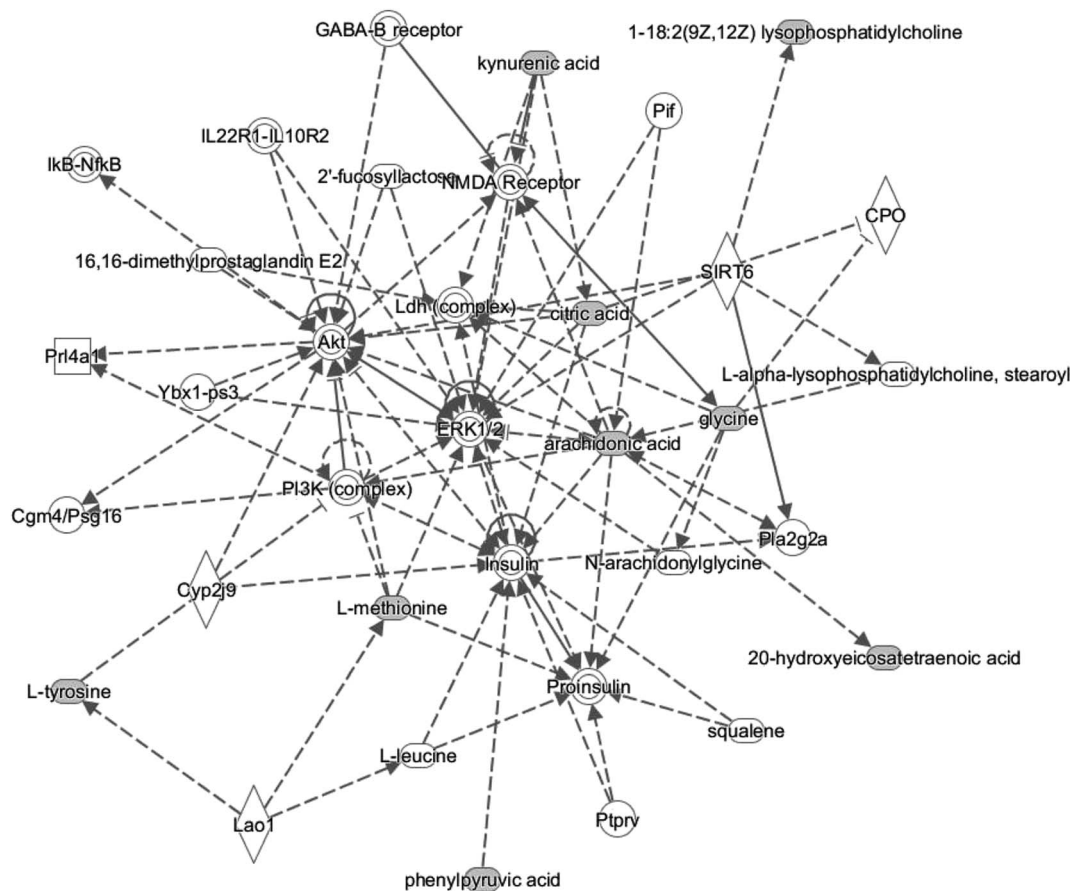


Fig. 5 Network analysis of the differential metabolites using the Ingenuity Pathway Analysis tool.

The differential metabolite expression may provide more reliable biomarkers for sepsis. A study explored the dynamic concentration profiles of amino acids and found that the metabolic spectrum of the amino acids changed dramatically in sepsis patients.³⁴ In this study, high-throughput metabolic phenotyping was used to characterize sensitive and economical peripheral biomarker(s) associated with SMD. The PCA score plot showed significant discrimination between the treatment and control cases. Serum metabolite profiling showed SMD to be associated with a profound abnormality in the metabolic phenotype. Metabolic pathway analysis indicated that endogenous metabolites such as pyroglutamic acid, 5-aminopentanoic acid, *p*-aminobenzoic acid, citric acid, *L*-tyrosine, pipecolic acid and *L*-methionine, which are closely connected with SMD, demonstrate a highly disturbed metabolism. Altogether, our results represent a good foundation for the full understanding of the pathogenesis of SMD and will provide beneficial effects for septic patients in the near future.

Conclusions

SMD increases mortality in sepsis, yet the underlying mechanism is unclear. Early recognition of intrinsic myocardial dysfunction is critical for the administration of the most appropriate therapy for septic patients. New approaches to

allow a deeper understanding of the underlying mechanism should help to improve the prognosis of patients with myocardial dysfunction in the near future. In this work, high-throughput UPLC/MS-based metabolic phenotyping was used to characterize serum biomarkers associated with the SMD metabolome. We found increased levels of pyroglutamic acid, 5-aminopentanoic acid, *p*-aminobenzoic acid, citric acid, *L*-tyrosine, pipecolic acid and *L*-methionine as compared to the control group. Further analysis of metabolic pathway variation indicated that many of the changed metabolites were involved in phenylalanine, tyrosine and tryptophan biosynthesis, arachidonic acid metabolism, glycine, serine and threonine metabolism, glyoxylate and dicarboxylate metabolism, phenylalanine metabolism, retinol metabolism, and tyrosine metabolism. The elucidation of these mechanisms should help identify new and specific therapeutic targets for septic patients. Our results demonstrate that integrating a metabolomics platform with chemometrics and bioinformatics is a promising approach for systematically dissecting the underlying metabolic mechanism of disease.

Conflicts of interests

The authors declare no competing interests.



Acknowledgements

This work was supported by grants from the Key Program of Natural Science Foundation of State (Grant No. 81774276, 81470196).

References

- 1 T. Suzuki, Y. Suzuki, J. Okuda, *et al.*, Sepsis-induced cardiac dysfunction and β -adrenergic blockade therapy for sepsis, *Journal of Intensive Care*, 2017, 5(1), 22.
- 2 R. Neviere, F. Delguste, A. Durand, *et al.*, Abnormal Mitochondrial cAMP/PKA Signaling Is Involved in Sepsis-Induced Mitochondrial and Myocardial Dysfunction, *Int. J. Mol. Sci.*, 2016, 17(12), 2075.
- 3 A. C. S. Freitas, M. J. Figueiredo, E. C. Campos, *et al.*, Activation of Both the Calpain and Ubiquitin-Proteasome Systems Contributes to Septic Cardiomyopathy through Dystrophin Loss/Disruption and mTOR Inhibition, *PLoS One*, 2016, 11(11), e0166839.
- 4 K. R. Walley, New Approaches to Modifying Inotropy in Sepsis-Induced Myocardial Dysfunction, *Crit. Care Med.*, 2017, 45(4), 754–756.
- 5 T. Janušonis and R. Paškevičiūtė, Clinical Case: Sepsis-Induced Cardiac Dysfunction, *Health Sciences*, 2016, 26(5), 26–28.
- 6 Y. Zhou, Y. Song, Z. Shaikh, H. Li, H. Zhang, Y. Caudle, S. Zheng, H. Yan, D. Hu, C. Stuart and D. Yin, MicroRNA-155 attenuates late sepsis-induced cardiac dysfunction through JNK and β -arrestin 2, *Oncotarget*, 2017, 8(29), 47317–47329.
- 7 Y. Okuhara, S. Yokoe, T. Iwasaku, *et al.*, Interleukin-18 gene deletion protects against sepsis-induced cardiac dysfunction by inhibiting PP2A activity, *Int. J. Cardiol.*, 2017, 243, 396–403.
- 8 Z. Zheng, H. Ma, X. Zhang, *et al.*, Enhanced Glycolytic Metabolism Contributes to Cardiac Dysfunction in Polymicrobial Sepsis, *J. Infect. Dis.*, 2017, 215(9), 1396–1406.
- 9 Q. Liang, H. Liu, H. Xing, Y. Jiang and A. Zhang, Urinary UPLC-MS metabolomics dissecting the underlying mechanisms of huaxian capsule protects against sepsis, *RSC Adv.*, 2016, 6, 40436–40441.
- 10 R. L. Wilson, V. Selvaraju, R. Lakshmanan, *et al.*, Thioredoxin-1 attenuates sepsis-induced cardiomyopathy after cecal ligation and puncture in mice, *J. Surg. Res.*, 2017, 220, 68–78.
- 11 E. Delile, R. Neviere, P. A. Thiébaud, *et al.*, Reduced insulin resistance contributes to the beneficial effect of protein tyrosine phosphatase-1B deletion in a mouse model of sepsis, *Shock*, 2017, 48(3), 355–363.
- 12 T. Fan, X. Feng, Y. Yang, *et al.*, Downregulation of PI3K- γ in a mouse model of sepsis-induced myocardial dysfunction, *Cytokines*, 2017, 96, 208–216.
- 13 Q. Liang, Y. Zhu, H. Liu, *et al.*, High-throughput lipidomics enables discovery of the mode of action of huaxian capsule impacting the metabolism of sepsis, *RSC Adv.*, 2017, 7(71), 44990–44996.
- 14 Y. Zhao, H. Lv, S. Qiu, *et al.*, Plasma metabolic profiling and novel metabolite biomarkers for diagnosing prostate cancer, *RSC Adv.*, 2017, 7(48), 30060–30069.
- 15 Q. Liang, H. Liu, L. Xie, *et al.*, High-throughput and multi-dimensional omics approach uncovers a huaxian capsule to ameliorate the dysregulated expression profiling of severe sepsis rats, *RSC Adv.*, 2017, 7(32), 19894–19903.
- 16 Q. Liang, H. Liu, L. Xie, X. Xue Li and A. H. Zhang, High-throughput metabolomics enables biomarker discovery in prostate cancer, *RSC Adv.*, 2017, 7, 2587–2593.
- 17 Q. Liang, H. Liu, L. Xie, *et al.*, High-throughput metabolomics enables biomarker discovery in prostate cancer, *RSC Adv.*, 2017, 7(5), 2587–2593.
- 18 X. Wang, J. Li and A. H. Zhang, Urine metabolic phenotypes analysis of extrahepatic cholangiocarcinoma disease using ultra-high performance liquid chromatography-mass spectrometry, *RSC Adv.*, 2016, 6(67), 63049–63057.
- 19 Q. Liang, H. Liu, Y. Jiang, *et al.*, Novel liquid chromatography-mass spectrometry for metabolite biomarkers of acute lung injury disease, *Anal. Methods*, 2016, 8, 6017.
- 20 Q. Liang, H. Liu, H. Xing, *et al.*, UPLC-QTOF/MS based metabolomics reveals metabolic alterations associated with severe sepsis, *RSC Adv.*, 2016, 6, 43293–43298.
- 21 H. Y. Chung, A. S. Kollmey, A. Schreppler, *et al.*, Adjustment of Dysregulated Ceramide Metabolism in a Murine Model of Sepsis-Induced Cardiac Dysfunction, *Int. J. Mol. Sci.*, 2017, 18(4), 839.
- 22 R. L. Wilson, V. Selvaraju, R. Lakshmanan, *et al.*, Thioredoxin-1 attenuates sepsis-induced cardiomyopathy after cecal ligation and puncture in mice, *J. Surg. Res.*, 2017, 220, 68–78.
- 23 J. Xia, I. V. Sinelnikov, B. Han and D. S. Wishart, MetaboAnalyst 3.0—making metabolomics more meaningful, *Nucleic Acids Res.*, 2015 Jul 1, 43(W1), W251–W257.
- 24 R. L. Wilson, V. Selvaraju, R. Lakshmanan, *et al.*, Thioredoxin-1 attenuates sepsis-induced cardiomyopathy after cecal ligation and puncture in mice, *J. Surg. Res.*, 2017, 220, 68–78.
- 25 Q. Liang, T. Zhang, Y. Jiang, H. Xing, C. Wang, B. Li and A. Zhang, Metabolomics of Alcoholic Liver Disease: A Clinical Discovery Study, *RSC Adv.*, 2015, 5, 80381–80387.
- 26 Q. Liang, H. Liu, H. Xing, Y. Jiang, T. Zhang and A. Zhang, High-resolution mass spectrometry for exploring metabolic signatures of sepsis-induced acute kidney injury, *RSC Adv.*, 2016, 6, 29863–29868.
- 27 Q. Liang, H. Liu, T. Zhang, Y. Jiang, H. Xing and A. Zhang, High-throughput metabolic profiling for discovering metabolic biomarkers of sepsis-induced acute lung injury, *RSC Adv.*, 2016, 6, 11008–11013.
- 28 O. Nduka and J. E. Parrillo, The pathophysiology of septic shock, *Crit. Care Nurs. Clin. North Am.*, 2011, 23(1), 41–66.
- 29 A. Tesfai, N. MacCallum, N. S. Kirkby, *et al.*, Metabolomic profiling of amines in sepsis predicts changes in NOS canonical pathways, *PLoS One*, 2017, 12(8), e0183025.



- 30 Y. Zhang, P. Liu, Y. Li and A. H. Zhang, Exploration of metabolite signatures using high-throughput mass spectrometry coupled with multivariate data analysis, *RSC Adv.*, 2017, 7(11), 6780–6787.
- 31 Y. F. Li, S. Qiu, L. J. Gao, *et al.*, Metabolomic estimation of the diagnosis of hepatocellular carcinoma based on ultrahigh performance liquid chromatography coupled with time-of-flight mass spectrometry, *RSC Adv.*, 2018, 8(17), 9375–9382.
- 32 Y. Zhao, H. Lv, S. Qiu, *et al.*, Plasma metabolic profiling and novel metabolite biomarkers for diagnosing prostate cancer, *RSC Adv.*, 2017, 7(48), 30060–30069.
- 33 Y. Zhao, H. Lv, S. Qiu, *et al.*, Plasma metabolic profiling and novel metabolite biomarkers for diagnosing prostate cancer, *RSC Adv.*, 2017, 7(48), 30060–30069.
- 34 L. Su, H. Li, A. Xie, *et al.*, Dynamic changes in amino acid concentration profiles in patients with sepsis, *PLoS One*, 2015, 10(4), e0121933.

