









Cite this: *Polym. Chem.*, 2019, **10**, 5626

Synthesis and supramolecular assembly of fluorinated biogenic amine recognition host polymers†

Ervin Kovács, ^a János Deme,^a Gábor Turczel,^a Tibor Nagy, ^a Vajk Farkas, ^a László Trif, ^a Sándor Kéki, ^b Péter Huszthy ^c and Robert Tuba ^{*a}

Copolymers containing hydroxyl (*i.e.* vinyl alcohol, VA) or fluorine functionalities are synthetic macromolecules having prominent biomedical applications. The concentration of hydroxyl groups along the polymer chain controls the polymer polarity. Moreover, the introduction of perfluorinated organic moieties *via* the OH functionalities may lead to macromolecules having potential magnetic resonance imaging (MRI) active properties. The ring-opening metathesis polymerization (ROMP) reaction using well-defined ruthenium-catalyzed systems is one of the most promising synthetic tools to fabricate such polymers. Co-polymerization of norbornene grafted pyridino-18-crown-6 ether (**7**) with fluorine-functionalized norbornenes (**10** and **11**) results in polymers bearing host molecular moieties. It has been demonstrated that the complexation of these host copolymers with biogenic amines including dopamine hydrochloride (**12**) and L-alanyl-L-lysine dipeptide hydrochloride (**13**) is straightforward. Based on the ¹H NMR investigation of the **7** and **12** complexation, an equilibrium constant of $\log K = 4.3 \pm 0.6$ could be calculated. The *in situ* ¹H NMR investigations have revealed that the complex formation of **13** with monomer **7** and perfluorinated copolymer **cp-7-10** takes place *via* both the lysine –NH₃⁺ and the alanine –NH₃⁺ moieties. However, in the case of homopolymer **poly-7**, the lysine–NH₃⁺ group coordination was observed exclusively. According to theoretical calculations, molecular switching of the crown ether structure of both the **7** monomer and its **cp-7-10** copolymer were observed from 90 degrees bent to planar structure upon –NH₃⁺ ion coordination.

Received 25th June 2019,
Accepted 26th September 2019
DOI: 10.1039/c9py00929a

rsc.li/polymers

Introduction

Supramolecular chemistry utilizes reversible non-covalent bonding for the assembly of molecular architectures.¹ Crown ethers (CEs) are well-known host molecules in supramolecular chemistry. Due to their excellent selectivity, crown ether-based molecular recognition motifs are widely used to synthesize polymers with unique properties.^{2,3} They can be involved either in the main chain or attached covalently to a polymer backbone as a side-chain.⁴ The synthesis of such host polymers can be carried out not only by classical polymerization

methods but also *via* ring opening metathesis polymerization (ROMP) reactions.^{5,6} Crown ethers can form stable complexes not only with metal cations but also with protonated amines.⁷ This possibility has prompted many studies of CEs with peptides and proteins.^{8,9} The utilization of CEs in medical applications is emerging.^{10,11} For example, amino-crown ethers have been utilized for the delivery of 5-fluorouracil (5-FU), an anticancer agent used for the treatment of metastatic carcinomas of pancreas and breast.¹² Moreover, several 18-crown-6 ethers have shown antitumor activity and reversal effect on multidrug resistance.¹³

We have recently reported the synthesis of vinyl alcohol (VA) copolymers having fine tunable polarities, which are considered as emerging nanocomposite materials for drug delivery applications.^{14a}

Fine tuning of the polarity of drug delivery polymers may enable preprogrammed and sustained drug release.^{14b} Moreover, it is envisioned that the fine tuning of the polarity of the CEs containing host polymers should make possible the transport of highly hydrophilic biogenic amines in a non-polar environment using a well-adjusted lipophilic polymer/nano-

^aInstitute of Materials and Environmental Chemistry, Research Centre for Natural Sciences, Hungarian Academy of Sciences, Magyar tudósok körútja 2, P.O. Box 286, H-1519 Budapest, Hungary. E-mail: tuba.robert@ttk.mta.hu

^bDepartment of Applied Chemistry, University of Debrecen, Egyetem tér 1, H-4032 Debrecen, Hungary

^cDepartment of Organic Chemistry and Technology, Budapest University of Technology and Economics, Szent Gellért tér 4, H-1111 Budapest, Hungary

†Electronic supplementary information (ESI) available. See DOI: 10.1039/c9py00929a

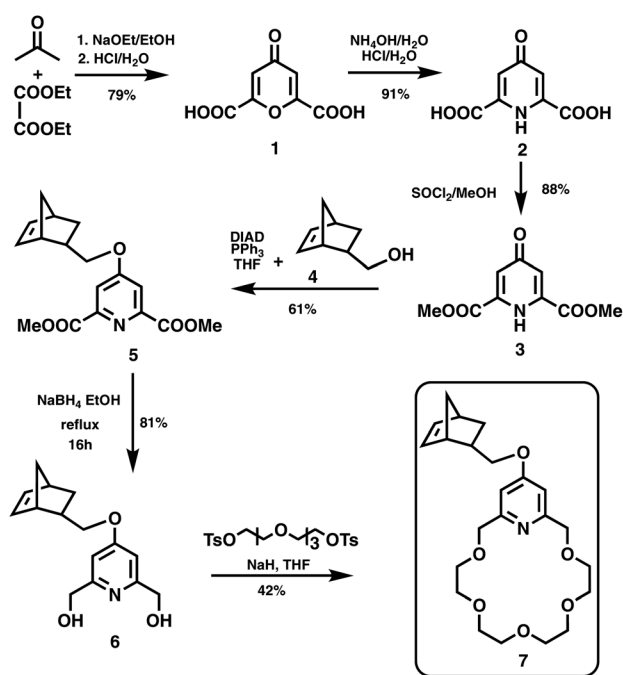


composite matrix. For example, it is tentatively supposed that tagging of dopamine *via* non-covalent reversible bonds to semi- or non-polar host polymers may enable its direct transport through the blood–brain barrier, leading to alternative treatment of Parkinson's disease.^{15,16} Furthermore, the introduction of perfluorinated organic moieties may open the way to magnetic resonance imaging (MRI) active properties of the potential drug carrier macromolecules.¹⁷ Here, we report the synthesis and supramolecular assembly of hydroxyl and fluorine functionalized biogenic amine recognition host polymers.

Results and discussion

Norbornene-functionalized pyridino-18-crown-6 ether (7) synthesis

As shown in Scheme 1, norbornene-functionalized pyridino-18-crown-6 ether 7 was synthesized *via* chelidonic acid (1) in 15% overall yield. 1 was prepared from diethyl oxalate and acetone in the presence of sodium ethoxide according to literature sources.¹⁸ Treatment of 1 with aqueous concentrated ammonium hydroxide resulted in chelidamic acid (2).¹⁹ Esterification was carried out with thionyl chloride and methanol leading to chelidamic acid dimethyl ester (3).²⁰ Following the Mitsunobu reaction of 3 with the *endo* norbornene derivative 4, the ether 5 could be obtained by column chromatography. The reduction of 5 to diol 6 followed by macrocyclization gave the crown ether 7 in moderate yield (42%). When the mixture of *endo/exo* isomers of 5 was used without separation, the product 7 contained *endo* and *exo* stereoisomers in 74% and 26% ratios, respectively.



Scheme 1 Synthesis of norbornene-functionalized pyridino-18-crown-6 ether (7).

Complexation of pyridino-18-crown-6 ether (7) with biogenic amines

The complexation of crown ether (7) with dopamine hydrochloride (12) (Fig. 1, 2 and Fig. S47†) and L-alanyl-L-lysine dipeptide hydrochloride (13) (Fig. 2 and Fig. S48†) has been investigated. Upon mixing 7 with 12, the C(py)-CH₂-O proton signals of 7 shifted upfield significantly from 4.64 to 4.57 ppm (Fig. 1). This is aligned with the literature data reported for the complexation of organic ammonium salts and similar pyridino-18-crown-6 ethers.^{21–24} The titration of 7 with 12 indicated a gradually increasing upfield shift of C(py)-CH₂-O signals of 7 (4.64 ppm) until reaching the stoichiometric ratio. However, as the stoichiometric ratio has been achieved there was no further significant shifts observed for the C(py)-CH₂-O signal in ¹H NMR spectra (Fig. 1). Based on the chemical shift changes, log *K* = 4.3 ± 0.6 could be calculated,²⁵ which is consistent with the reported data for similar complexes shown in Fig. S54.† Izatt *et al.* found a slightly lower log *K* value of 3.62 for the complexation of a bulkier isoalkyl-ammonium ion, (*R*)-1-phenylethylamine with a less flexible dimethylated pyridino-18-crown-6 ether molecule (see details of calculation in the ESI†).²⁵ The *K* values for complexation of crown ethers using ammonium salts are in general high, and the equilibrium is shifted toward the complex formation side.^{21–24} Reproduction of the complexation using dipeptide 13 resulted in a similar supramolecular complex formation (see also ESI Fig. S48†).

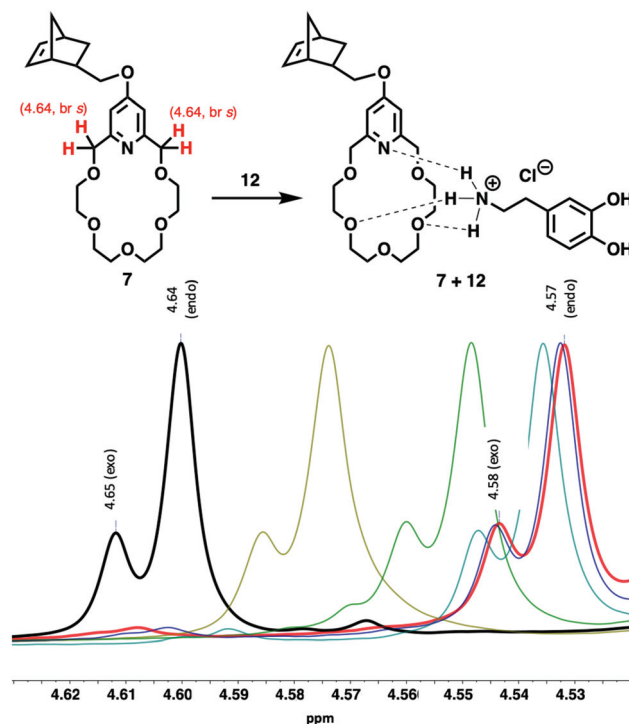


Fig. 1 Investigation of the complexation of 7 (*endo/exo* mixture) with 12 by a titration ¹H NMR method. Chemical shifts of pyridino-18-crown-6 ether methyldine protons (red) vs. 12 : 7 molar ratios. Black: 0; brown: 0.3; green: 0.6; light blue: 1.0; dark blue: 1.5; red: 2.0 (MeOD-CD₂Cl₂ 1 : 1 mixture, [7] = [12] = 0.01 mmol mL⁻¹).

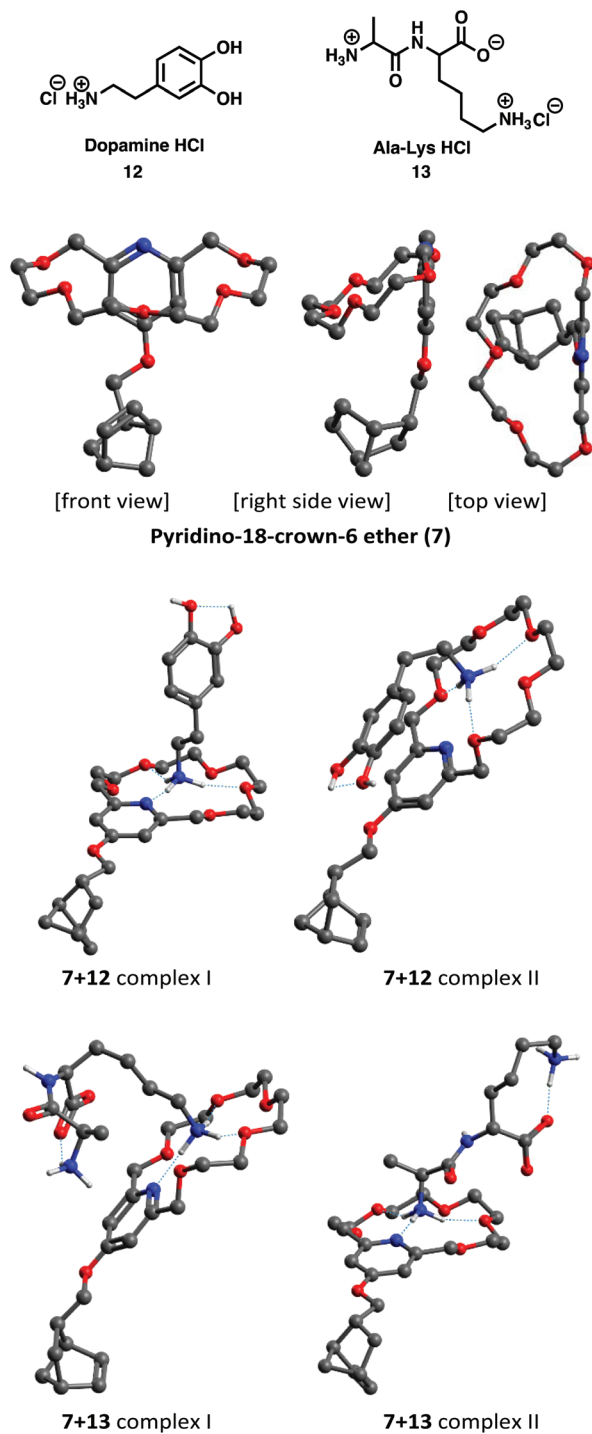


Fig. 2 Structural formulas of the dopamine HCl (**12**) and Ala-Lys HCl (**13**) (top two 2D structures). DFT optimized minimum energy conformer of pyridino-18-crown-6 ether (**7**) in methanol, shown in three perpendicular directions (top). 2–2 DFT optimized low energy conformers of complexes **7+12** (middle two) and **7+13** (bottom two) in an implicit methanol solvent (without the chloride ion). See text for the description of various complexes. In the ball and stick representation, carbon, nitrogen, oxygen and heteroatom-bound hydrogen atoms are drawn as grey, blue, red and white balls, respectively.

Electronic structure calculations using Gaussian 09 program package²⁶ were carried out to determine the lowest-energy conformers of **7** and its complexes formed with dopamine-HCl (**12**) and L-alanyl-L-lysine dipeptide-HCl (**13**) at the M06-2X/cc-pVDZ level of density functional theory (DFT)^{27,28} using the SMD implicit solvent model.²⁹ Coordinates of optimized geometries are listed in the ESI.† Molecular dynamics (MD) studies with explicit solvent molecules and accurately parameterized solvent-solute interaction are needed to account for such a complex environment.³⁰ Implicit solvent models do not allow the realistic treatment of solvent mixtures, especially in the case of different solvent types (*e.g.*, protic *vs.* aprotic, highly polar *vs.* slightly polar/apolar).

The recent MD study of Benay and Wipff investigated solvation and complexation of alkali cations (picrate salts) by 18-crown-6 in a 90 : 10 chloroform/methanol mixture and found that the ions and the highly polar moieties are surrounded mainly by the polar and protic methanol molecules.³⁰ Accordingly, they found that the calculated free energy change of complexation in the solvent mixture was very much like in pure methanol as intense interaction energies of polar and ionic groups dominated the energy change of the process. While specific MD studies are beyond the scope of the present work, these findings allow a realistic treatment of the complexation in methanol/dichloromethane and methanol/chloroform 50/50 solvent mixtures even with an implicit solvent model if pure methanol solvent is assumed.

Consequently, all electronic structure calculations presented here were carried out using methanol as an implicit solvent. A systematic (though probably non-exhaustive) DFT exploration of the high-dimensional conformational space of the crown ether ring identified several low-energy conformers for **7**.

The one with the lowest energy is shown in three perpendicular directions in Fig. 2 (top three 3D structures). The minimum energy structure of the pyridino-18-crown-6 ether ring is largely different from the regular chair-type S_6 -symmetric structure expected for 18-crown-6 ethers. Driven by secondary interactions, the crown shrinks and folds back over the pyridine ring. Complexation of **7** with dopamine and L-alanyl-L-lysine dipeptide hydrochloride (**12** and **13**) was investigated in methanol as an implicit solvent without explicit consideration of the chloride ion and by assuming a zwitterionic form for the dipeptide (Fig. 2).

During complexation, the crown ether folds out, and the cavity of the macrocycle expands and takes on a much more regular shape that can host the primary ammonium cations. Complexation of **12** proceeds by forming three H-bonds (distributed at every $\sim 120^\circ$) with the crown ether by two different ways: either involving the nitrogen atom of the pyridine ring (**7+12** complex I) or without it (**7+12** complex II) (Fig. 2 middle two 3D structures). The former is energetically favored when no other interactions of the ligand with the crown ether are considered. However, when the nitrogen atom is not involved in the complexation, the conformational motion of the dopamine allows the aromatic ring of the dopamine to fold back over the pyridine ring (Fig. 2 middle right) and this structure



is stabilized by π - π interactions.²⁵ The protonated dipeptide cation can form a complex with the crown ether *via* either its lysine side chain (7+13 complex I) or its N-terminal (7+13 complex II) (Fig. 2 bottom two 3D structures). The formation of the intramolecular H-bond on the one hand significantly lowers the energy of the latter structure, and on the other hand, it hinders its conformational motion. The crown ether (7) and all complexes of 7+12 and 7+13 turned out to be very flexible as they have several low-energy very low-frequency vibrational modes (~ 20 cm⁻¹). Thus, to assess their relative stability through their standard free energy differences, full conformational sampling using MD simulations³⁰ would be required. This, however, is beyond the scope of this study. Nevertheless, one can conclude that in proteins, which are the main targets of the proposed application, complexation through the lysine side chain will be dominant statistically as the number of lysine residues in them is usually much higher than one.

Polymer synthesis

Cycloolefin-functionalized crown ethers are expected to participate in polymerization and co-polymerization reactions such as ruthenium-catalyzed ring-opening metathesis polymerization (ROMP).^{31,32} Indeed, it was found that the norbornene-functionalized pyridino-18-crown-6 ether can readily be polymerized using commercially available ruthenium metathesis catalysts **G2** (Grubbs 2nd generation catalyst) and **G3** (Grubbs 3rd generation catalyst). Although **G2** provides polynorbornene having a moderate polydispersity ($D > 1.5$) – especially at relatively high catalyst loading – it has a higher functional group tolerance compared to the **G3** metathesis catalysts.³³ The ROMP reactions of norbornenes, in general, are relatively straightforward reactions, and they can be carried out at a low catalyst loading in any common organic solvent giving insoluble high molecular weight polymers.³⁴

In these tests, however, a relatively high (2 mol%) catalyst loading was applied to obtain relatively short polymers (M_w (mass-average molar mass) < 20 kDa) with reasonable THF solubility for GPC analysis. The ROMP of **7** was carried out at room temperature in dichloromethane using the **G2** catalyst giving the homopolymer **poly-7** in 99% isolated yield (Table 1). The formed polymer was sparingly soluble in CDCl₃, CD₂Cl₂

and CD₂Cl₂/MeOD mixture (1:1); however, it was poorly soluble in THF, which did not allow its GPC analysis. When the polymerization was carried out using the **G3** catalyst, the product showed increased THF solubility (M_w : 1.53×10^3 Da, D (dispersity): 1.17). This observation can be explained by the significantly faster initiation and living polymerization character of the **G3** catalyst leading to polymers with lower M_w values and narrow dispersity (D).³³ MALDI-TOF MS measurements showed molecular weights corresponding to the oligomers containing 7 units (m/z : 419, Fig. S15†). Preliminary co-polymerizations of **7** have been carried out with norbornene (**8**) at 1:1 and 1:5 molar ratios using the **G3** catalyst.³³ The M_w of the copolymers were 4.49×10^3 Da (D : 1.68) at 7:8 = 1:1 and 7.19×10^3 Da, (D : 1.87) at 7:8 = 1:5 molar ratio, respectively. Co-polymerization carried out at the 7:8 = 1:1 molar ratio with the **G2** catalyst resulted in a non-THF soluble polymer (98% yield) (Scheme 2). However, the co-polymerization of **7** and **8** at the 1:5 molar monomer ratio led to a THF soluble polymer in 97% yield with a comparable molecular weight (M_w : 5.97×10^3 Da) and higher polydispersity (D : 1.80). The MALDI-TOF MS measurements indicate that the oligomers are comprised of both **7** (m/z : 419) and **8** (m/z : 94) monomer units (Fig. S23†), as expected. Considering the observed molecular weights for **cp-7-8** polymers (Table 1), it can be concluded that these data are consistent with the M_w value of polynorbornene synthesized under the similar condition and a slightly lower catalyst loading (0.6 mol%).³⁵

Following the preliminary co-polymerization tests with **8**, the incorporation of hydroxyl and perfluoro-*tert*-butyl groups into the host polymer chain was investigated. Perfluorinated moieties containing monomer **10** was synthesized by the reaction of the tosyl ester derivative of **4** (**Tos-4**)³⁶ with the sodium salt of nonafluoro-*tert*-butyl alcohol³⁷ in reasonable yield (60%). Meanwhile, the bis-perfluorinated moieties containing monomer **11** has been synthesized by the Mitsunobu reaction³⁸ *via* the reaction of **9** and nonafluoro-*tert*-butyl alcohol in moderate yield (27%). The co-polymerization of crown ether **7** with **4** and **9** and perfluoro-*tert*-butyl-functionalized norbornenes **10** and **11** gave copolymers having randomly distributed dyads.

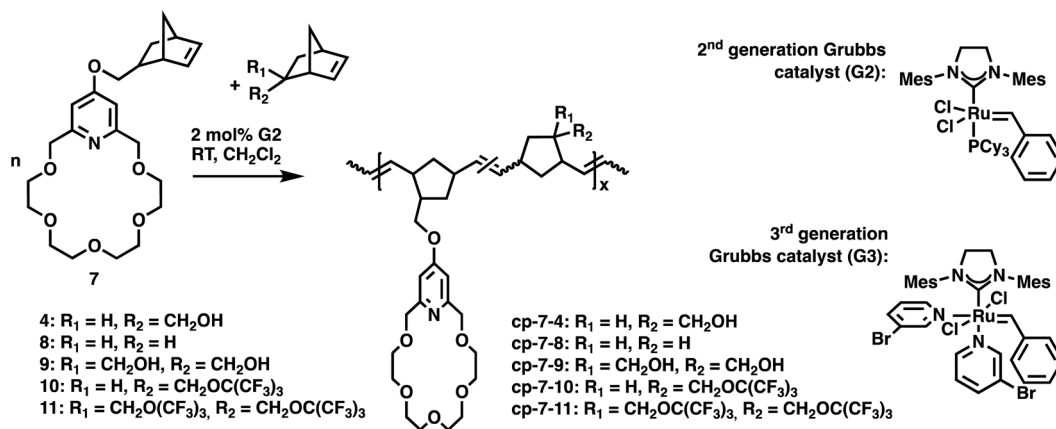
The co-polymerization of **7** with **4** in the stoichiometric ratio resulted in the **cp-7-4** copolymer in quantitative yield. The polymer does not render any THF solubility. However, it is

Table 1 MW, polydispersity (D) and glass transition (T_g) and decomposition (T_d) temperature of the synthesized host polymers

Entry	Polymer	Yield (%)	Cp-M-M' M/M'	Catalysts	THF solubility	M_w (Da) $\times 10^3$	D	T_g (°C)	T_d (°C)
1	poly-7	99	NA	G2	Poor	—	—	−36	347
2	poly-7	93	NA	G3	Sparing	1.53	1.17	—	—
3	cp-7-8	98	1/1	G2	Poor	—	—	14.9	329
4	cp-7-8	95	1/1	G3	Moderate	4.49	1.68	—	—
5	cp-7-8	91	1/5	G3	Moderate	7.19	1.87	—	—
6	cp-7-8	97	1/5	G2	Moderate	5.97	1.80	19.7	248
7	cp-7-10	92	1/1	G2	Poor	—	—	−16	295
8	cp-7-10	98	1/5	G2	Sparing	9.85	1.43	^a	307
9	cp-7-11	98	1/5	G2	Moderate	5.59	2.16	^a	403

^a Crystalline structure, no T_g data were obtained. Cp-M-M': copolymer of monomer 1 (M) and monomer 2 (M'). M/M': theoretical monomer 1 and monomer 2 ratio in the copolymer.





Scheme 2 Co-polymerization of crown ether-functionalized norbornenes.

sparingly soluble in the methylene chloride-methanol mixture. The co-polymerization of **7** with **9** led to the formation of entirely insoluble copolymer **cp-7-9**. These observations are consistent with literature data reporting that **poly-4** and **poly-9** have minimal solubility in common organic solvents.³⁹ However, the introduction of perfluoro-*tert*-butyl units has significantly improved the copolymer solubility even in tetrahydrofuran. The co-polymerization of **7** has been carried out with **10** at 1 : 1 and 1 : 5 molar ratios using the G2 catalyst (**cp-7-10** (1 : 1): 92% and **cp-7-10** (1 : 5): 98% yield) in dichloromethane solution. **cp-7-10** (1 : 1) was only sparingly soluble in THF; however, **cp-7-10** (1 : 5) rendered reasonable THF solubility. The GPC tests of **cp-7-10** (1 : 5) revealed a reasonable $M_w = 9.85 \times 10^3$ Da (D : 1.43) value (Table 1). As it was expected, the MALDI-TOF MS investigations have clearly indicated that the copolymers are comprised of both **7** (m/z : 419) and **10** (m/z : 342) monomer units as well (Fig. 3). Polymer **cp-7-11** containing the bis-perfluoro-*tert*-butyl moieties in 1 : 5 = 7 : 11 ratio has been synthesized in high yield (98%). The polymer rendered a similar molecular weight and dispersity (5.59×10^3 Da, D : 2.16) to copolymers **cp-7-8** and **cp-7-10** synthesized under similar reaction conditions (Table 1). Based on the DSC measurements (Table 1), it can be concluded that the inclusion of side groups increases the T_d values (as seen at **poly-7**: 347 °C and **cp-7-11** (1 : 5): 403 °C vs. **cp-7-8** (1 : 1): 329 °C; **cp-7-8** (1 : 5): 248 °C). The co-polymerization of **7** with **8** at the 1 : 1 monomer ratio significantly increased the T_g value (**poly-7** = -36 °C; **poly-7-8** (1 : 1) = 14.9 °C). However, increasing the 7 : 8 monomer ratio up to 1 : 5 resulted in only a slight increase of the T_g value (**poly-7-8** (1 : 5) = 19.7 °C).

Complexation of pyridino-18-crown-6 ether (7)-functionalized polymers with biogenic amines

The complexation of **poly-7** with **12** (Fig. 4 and Fig. S49†) and **13** (Fig. S50†) have been investigated by ¹H NMR spectroscopy in the MeOD-CD₂Cl₂ 1 : 1 solvent mixture at 30 °C. As expected, upon complexation the **poly-7** broad aromatic proton signals shifted upfield from 6.83 to 6.79 ppm; meanwhile, the aromatic protons of **12** have also shown significant shifts from

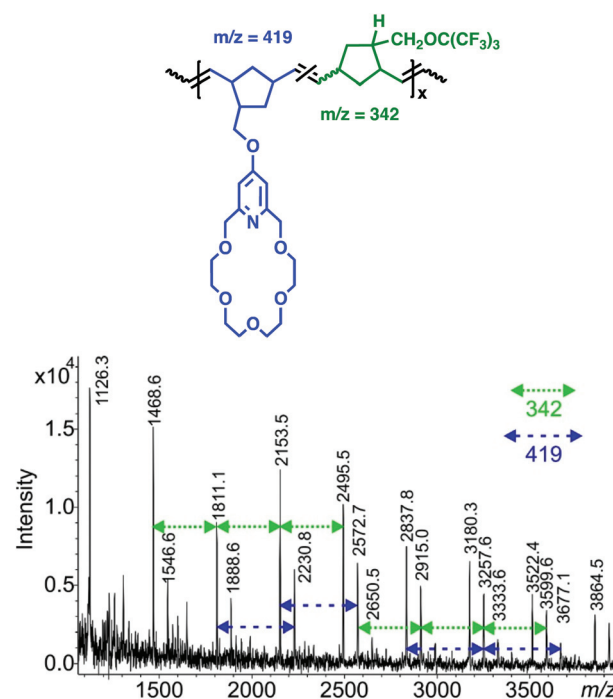


Fig. 3 MALDI TOF MS spectrum of copolymer **cp-7-10**, **7** (m/z : 419), **10** (m/z : 342) (7/10 = 1/5) (linear mode, DHB/NaTFA).

6.80 (d), 6.76 (d) and 6.62 (dd) ppm to 6.76, 6.71 and 6.55 ppm, respectively (Fig. 4).

These observations are in accordance with the literature data reported for the supramolecular complex formation of pyridino-18-crown-6 ethers with protonated primary amines.²¹ Similar chemical shift could be clearly observed in the aliphatic proton region as well. The aliphatic CH₂ proton signals of **12** shifted from 3.13 (t) and 2.86 (t) ppm to 3.09 and 2.80 ppm, respectively (Fig. 4). As was expected, the proton signals of **12** significantly broadened upon complexation with the crown ether-functionalized polymer.

The complexation of **poly-7** with **13** also revealed significant changes of the chemical shifts of the reacting molecules. The



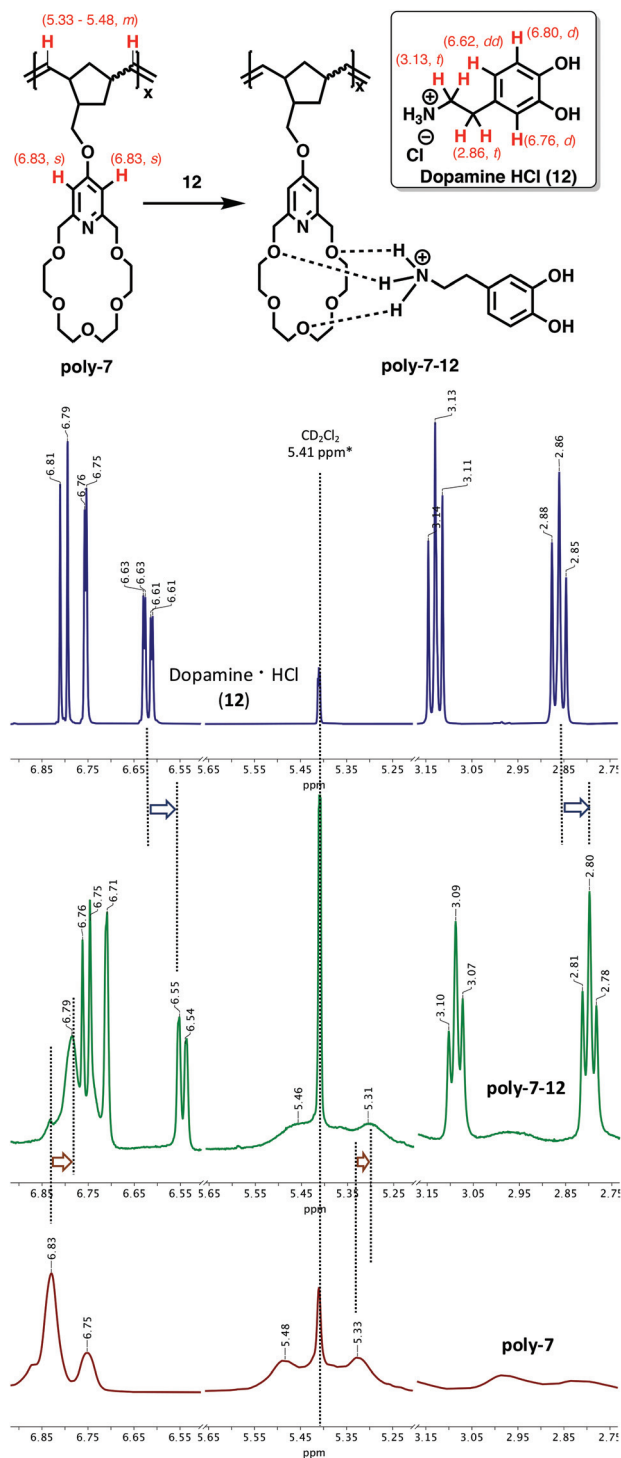


Fig. 4 Complexation of **poly-7** (bottom) with Dopamine-HCl, **12** (top), and **poly-7-12** complex (middle) (MeOD-CD₂Cl₂ 1 : 1 mixture, 30 °C, [poly-7] = [12] = 0.01 mmol mL⁻¹).

aromatic proton signals of **poly-7** shifted upfield from 6.83 ppm to 6.79 ppm (Fig. S50†). Unlike the complexation of **7** with **12**, it can be seen that upon complexation of **poly-7** with **13** the most characteristic peaks CH (t) (4.33 ppm) and CH₂ (t) (2.95 ppm) of **13** have revealed significant chemical shifts to

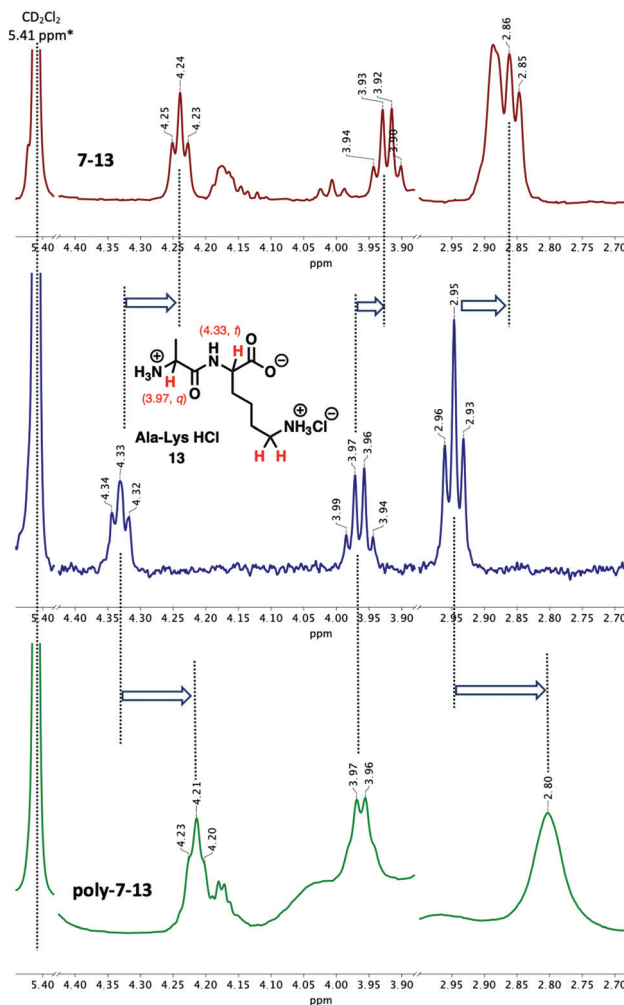


Fig. 5 Complexation of **13** (middle) with pyridino-18-crown-6 ether, **7-13** (top) and polymer, **poly-7-13** (bottom) (MeOD-CD₂Cl₂ 1 : 1 mixture, 30 °C, [7] = [13] = 0.01 mmol mL⁻¹).

4.21 ppm and 2.80 ppm; meanwhile, the CH (q) peak at 3.97 ppm did not show any significant changes (Fig. 5, bottom). This observation indicates that the complexation between **poly-7** and **13** most probably occurs exclusively *via* the lysine -NH₃⁺ group rather than the alanine -NH₃⁺ moiety, unlike the complexation between **7** and **13** where all the three peaks revealed significant upshift indicating that it takes place by both -NH₃⁺ moieties (Fig. 5, top).

Following the investigation of the complexation of homopolymer **poly-7** with **12**, the complexation of perfluoro *tert*-butyl group-functionalized copolymer **cp-7-10** has been investigated (Fig. 6).

As expected, upon addition of **12** to a MeOD-CD₂Cl₂ solution of **cp-7-10** (1 : 1), the aromatic proton signals of **12** (6.81 (d), 6.75 (d) and 6.62 (dd) ppm) significantly shifted to 6.79 (d), 6.73 (d) and 6.60 (dd) ppm, respectively. Meanwhile, broadening of the proton peaks of **12** was observed.

The aliphatic proton signals of **12** have also shifted upfield from 3.13 (t) and 2.86 (t) ppm to 3.12 (t) and 2.84 (t) ppm, respectively (Fig. 6 and Fig. S51†). Meanwhile, the broad aro-



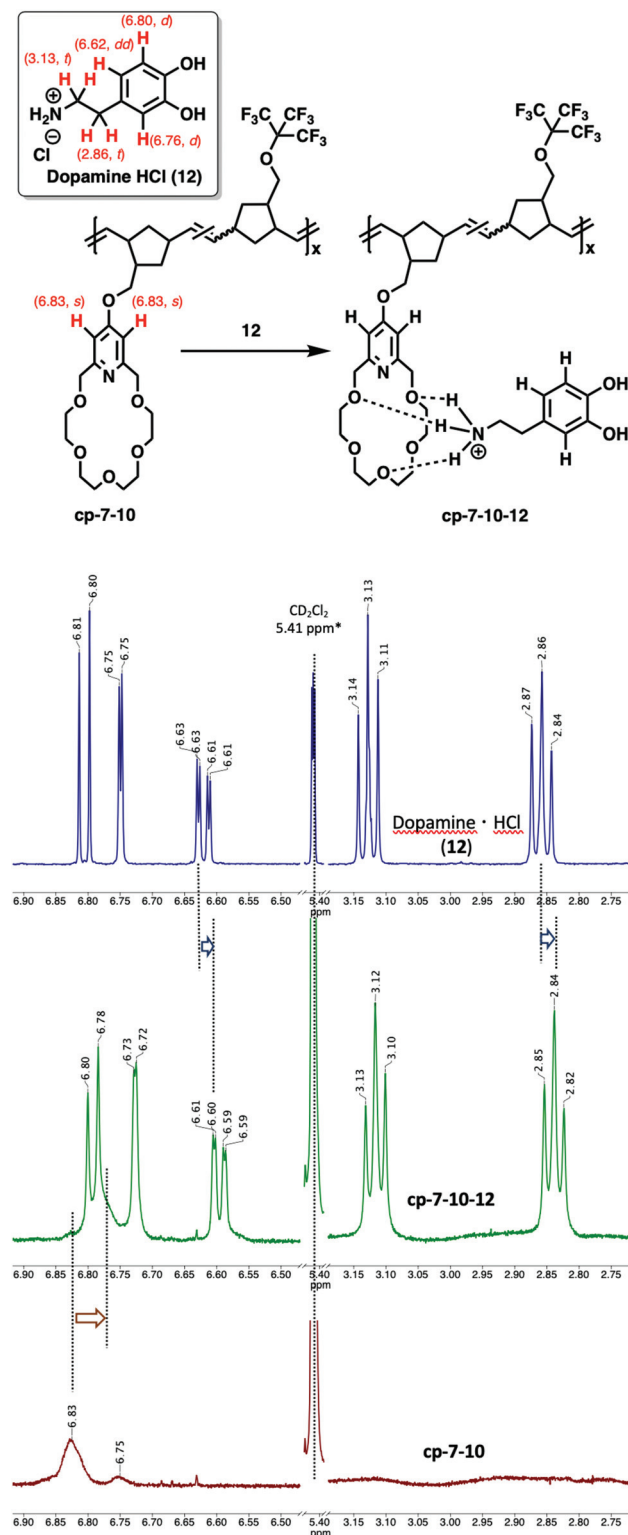


Fig. 6 Complexation of poly-7-10 (1:1, bottom) with Dopamine-HCl, 12 (top), and poly-7-10-12 complex (middle) (MeOD-CD₂Cl₂ 1:1 mixture, 30 °C, [cp-7-10] = [12] = 0.01 mmol mL⁻¹).

matic CH proton peaks of **cp-7-10** were also shifted upfield from 6.83 ppm to 6.77 ppm. The observed simultaneous upfield shifts and peak broadenings of **12** clearly indicate an

interaction between the polymer (**cp-7-10**) and Dopamine-HCl (**12**). The complexation of **cp-7-10** with Ala-Lys-HCl (**13**) led to similar shifts of the pyridino-18-crown-6 ether aromatic protons (from 6.83 (br) to 6.78 (br) ppm) and lysine protons (from 4.33 (t) to 4.26 (br) ppm and from 2.95 (t) to 2.91 (br) ppm) (Fig. S52†). Just as in the case of complexation of **cp-7-10** with **12**, the proton signal broadening of **13** were unambiguous, indicating the specific intermolecular interaction between **cp-7-10** and Ala-Lys-HCl (**13**). It should be noted that based on the ¹H NMR spectra, there was significant upshift for the quartet signal of the Ala unit from 3.97 ppm to 3.91 ppm (CH-CH₃), indicating that most probably the complexation occurs not only *via* the protonated lysine side chain but also on the protonated N-terminal alanine dyad (Fig. S52†).

The formed copolymers are not only non-periodic, but also highly flexible; thus, they do not exhibit a well-defined geometry. The all-*trans* H-(poly-7-10)₄-H alternating cooligomer can be considered as the most abundant possible dyad⁴⁰ of the formed host polymers. A local optimum structure of its complex formed with **12** was determined in an implicit methanol solvent using the PM6 semiempirical theory and is shown as an illustration in Fig. 7 to help the reader to visualize them.⁴¹

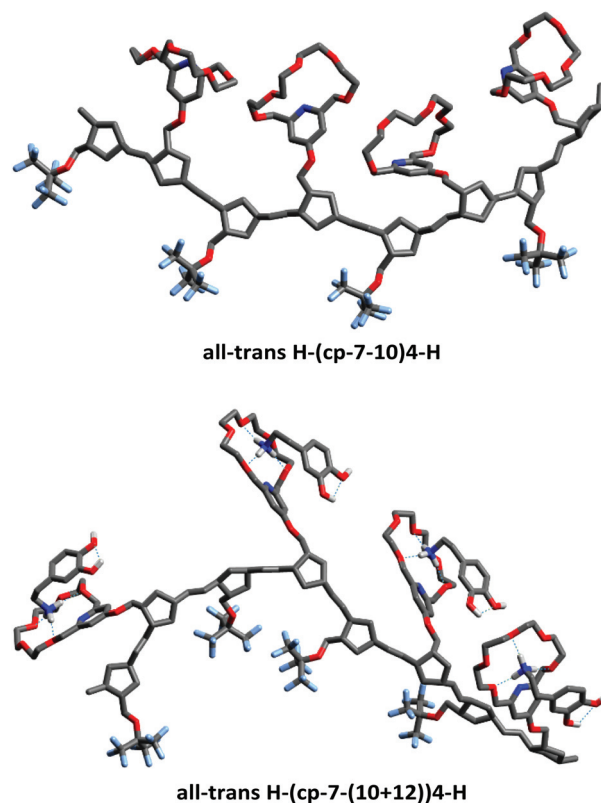


Fig. 7 A local optimum structure of the all-*trans* H-(cp-7-10)₄-H cooligomer and its complex with **12** (cp-7-(10+12))₄-H determined at the PM6 level of theory in an implicit methanol solvent. Molecular switching to planar structure of pyridino-18-crown-6 ether moieties upon complexation with **12**. In the ball and stick representation carbon, nitrogen, oxygen and heteroatom-bound hydrogen atoms are drawn as grey, blue, red and white balls, respectively.



Coordinates of optimized geometries are listed in the ESI.† If no complexation takes place, the strong attraction between the tilted and folded crown ether units aligns them and bends the copolymer chain toward the side groups. In the complex state, the side groups are positively charged due to the presence of the primary ammonium ions. The strong electrostatic repulsion is only partially ameliorated by the methanol solvent (and the scattered, solvated chloride ions). Thus, the side groups tilt away from each other and the polymer chain becomes distorted.

Conclusions

In summary, a wide range of copolymers having host (pyridino-18-crown-6 ether), OH and perfluoro *tert*-butyl functionalities have been synthesized *via* ruthenium-catalyzed ring opening metathesis polymerization.

The complexation of **poly-7** and its perfluoro-*tert*-butyl group containing copolymers (**poly-7-10**) with biogenic amines including dopamine (**12**) and L-alanyl-L-lysine dipeptide (**13**) has led to the formation of the supramolecular complexes indicated by ¹H NMR spectroscopy. Upon complexation, significant upfield shift of the proton signals (0.02–0.06 ppm) of both reactants and peak broadening of the biogenic amines were observed. The investigations have revealed that the complex formation of **13** with monomer **7** and copolymer **cp-7-10** may take place *via* either the lysine or alanine –NH₃⁺ moieties. However, in the case of **poly-7**, the lysine amine group coordination was observed exclusively.

The experimental data have been supported by theoretical calculations, also indicating that the complexation of a biogenic amine to the presented host polymer is straightforward.

It is envisioned that these polymers may have potential to serve as “biogenic amine carriers” opening new alternatives for drug delivery applications.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was funded by grants provided by the National Competitiveness and Excellence Program, Hungary (NVKP-16-1-2016-0007), and by the BIONANO-GINOP-2.3.2-15-2016-00017 project. The authors are grateful to Materia, Inc., for providing G2. Tibor Nagy and Péter Huszthy are grateful to the Hungarian Academy of Sciences and National Research, Development and Innovation Office (NKFIH) for financial support under Grant No. PD 120776 and K128473, for János Bolyai Research Fellowship BO/00279/16/7 and for the Hungarian NIIF HPC infrastructure. Ervin Kovács is grateful to the National Research, Development and Innovation Office (NKFIH) for Postdoctoral Excellence Award (PD 128612). Attila

Domján is acknowledged for solid NMR analysis. The authors thank Prof. John A. Gladysz (TAMU) for the revision and the constructive comments.

Notes and references

- 1 L. Brunsveld, B. J. B. Folmer, E. W. Meijer and R. P. Sijbesma, *Chem. Rev.*, 2012, **101**, 4071–4097.
- 2 C. R. South, M. N. Higley, K. C. F. Leung, D. Lanari, A. Nelson, R. H. Grubbs, J. F. Stoddart and M. Weck, *Chem. – Eur. J.*, 2006, **12**, 3789–3797.
- 3 C. R. South, K. C. F. Leung, D. Lanari, J. F. Stoddart and M. Weck, *Macromolecules*, 2006, **39**, 3738–3744.
- 4 U. Tunca and Y. Yagci, *Prog. Polym. Sci.*, 1994, **19**, 233–286.
- 5 R. Ji, C. G. Chao, Y. C. Huang, Y. K. Lan, C. L. Lu and T. Y. Luh, *Macromolecules*, 2010, **43**, 8813–8820.
- 6 E. Eder, P. Preishuber-Pflügl and F. Stelzer, *J. Mol. Catal. A: Chem.*, 2000, **160**, 63–69.
- 7 V. Rüdiger, H. J. Schneider, V. P. Solov'ev, V. P. Kazachenko and O. A. Raevsky, *Eur. J. Org. Chem.*, 1999, 1847–1856.
- 8 S. W. Lee, H. N. Lee, H. S. Kim and J. L. Beauchamp, *J. Am. Chem. Soc.*, 1998, **120**, 5800–5805.
- 9 R. R. Julian and J. L. Beauchamp, *J. Am. Soc. Mass Spectrom.*, 2002, **13**, 493–498.
- 10 X. Liu, S. Zhu, S. Wu, P. Wang and G. Han, *Colloids Surf., A*, 2013, **417**, 140–145.
- 11 I. A. Darwish and I. F. Uchegbu, *Int. J. Pharm.*, 1997, **159**, 207–213.
- 12 R. Muzzalupo, F. P. Nicoletta, S. Trombino, R. Cassano, F. Iemma and N. Picci, *Colloids Surf., B*, 2007, **58**, 197–202.
- 13 I. Guberović, M. Marjanović, M. Mioč, K. Ester, I. Martin-kleiner, T. Š. Ramljak, K. Mlinarić-majerski and M. Kralj, *Sci. Rep.*, 2018, **8**, 14467.
- 14 (a) T. Feczko, G. Merza, G. Babos, B. Varga, E. Gyetvai, L. Trif, E. Kovács and R. Tuba, *Int. J. Pharm.*, 2019, **562**, 333–341; (b) S. Miyazaki, S. Takeuchi, W. Hou, N. Hashiguchi, C. Yokouchi, M. Takada, M. Hosokawa, Y. Koga and H. Kobayashi, *Chem. Pharm. Bull.*, 1985, **33**, 2490–2498.
- 15 C. Saraiva, C. Praça, R. Ferreira, T. Santos, L. Ferreira and L. Bernardino, *J. Controlled Release*, 2016, **235**, 34–47.
- 16 A. M. Grabrucker, B. Ruozzi, D. Belletti, F. Pederzoli, F. Forni, M. A. Vandelli and G. Tosi, *Tissue Barriers*, 2016, **4**, e1153568.
- 17 S. Decato, T. Bemis, E. Madsen and S. Mecozzi, *Polym. Chem.*, 2014, **5**, 6461–6471.
- 18 E. R. Riegel and F. Zwiilmeyer, *Org. Synth.*, 1937, **2**, 126–128.
- 19 E. R. Riegel and M. C. Reinhard, *J. Am. Chem. Soc.*, 1926, **48**, 1334–1345.
- 20 J. S. Bradshaw, P. Huszthy, T. Wang, C. Zhu, A. Y. Nazarenko and R. M. Izatt, *Supramol. Chem.*, 1993, **1**, 267–275.
- 21 T.-M. Wang, J. S. Bradshaw, J. C. Curtis, P. Huszthy and R. M. Izatt, *J. Inclusion Phenom. Mol. Recognit. Chem.*, 1993, **16**, 113–122.



- 22 T.-M. Wang, J. S. Bradshaw and R. M. Izatt, *J. Heterocycl. Chem.*, 1994, **31**, 1097–1114.
- 23 C.-Y. Zhu, J. S. Bradshaw, J. L. Oscarson and R. M. Izatt, *J. Inclusion Phenom. Mol. Recognit. Chem.*, 1992, **12**, 275–289.
- 24 (a) J. S. Bradshaw, P. Huszthy, C. W. McDaniel, C.-Y. Zhu, N. K. Dalley, R. M. Izatt and S. Lifson, *J. Org. Chem.*, 1990, **55**, 3129–3137; (b) S. V. Brignell and D. K. Smith, *New J. Chem.*, 2007, **31**, 1243–1249.
- 25 R. M. Izatt, T. Wang, J. K. Hathaway, X. X. Zhang, J. C. Curtis, J. S. Bradshaw, C. Y. Zhu and P. Huszthy, *J. Inclusion Phenom. Mol. Recognit. Chem.*, 1994, **17**, 157–175.
- 26 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, Ö. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, Gaussian Inc., Wallingford, CT, 2009.
- 27 Y. Zhao and D. G. Truhlar, *Theor. Chem. Acc.*, 2008, **120**, 215–241.
- 28 T. H. Dunning, *J. Chem. Phys.*, 1989, **90**, 1007–1023.
- 29 A. V. Marenich, C. J. Cramer and D. G. Truhlar, *J. Phys. Chem. B*, 2009, **113**, 6378–6396.
- 30 G. Benay and G. Wipff, *New J. Chem.*, 2016, **40**, 4662–4671.
- 31 C. W. Bielawski and R. H. Grubbs, in *Controlled and Living Polymerizations*, 2009, pp. 297–342.
- 32 C. W. Bielawski and R. H. Grubbs, *Prog. Polym. Sci.*, 2007, **32**, 1–29.
- 33 T. Choi and R. H. Grubbs, *Angew. Chem., Int. Ed.*, 2003, **42**, 1743–1746.
- 34 J. Suriboot, Y. Hu, T. J. Malinski, H. S. Bazzi and D. E. Bergbreiter, *ACS Omega*, 2016, **1**, 714–721.
- 35 R. Tuba, R. Corrêa Da Costa, H. S. Bazzi and J. A. Gladysz, *ACS Catal.*, 2012, **2**, 155–162.
- 36 V. De Matteis, F. L. Van Delft, J. Tiebes and F. P. Rutjes, *Eur. J. Org. Chem.*, 2006, 1166–1176.
- 37 D. Szabó, J. Mohl, A. M. Bálint, A. Bodor and J. Rábai, *J. Fluor. Chem.*, 2006, **127**, 1496–1504.
- 38 K. C. Kumara Swamy, N. N. Bhuvan Kumar, E. Balaraman and K. V. P. Pavan Kumar, *Chem. Rev.*, 2009, **109**, 2551–2651.
- 39 U. Frenzel, T. Weskamp, F. J. Kohl, W. C. Schattenmann, O. Nuyken and W. A. Herrmann, *J. Organomet. Chem.*, 1999, **586**, 263–265.
- 40 R. Tuba, M. Al-Hashimi, H. Bazzi and R. Grubbs, *Macromolecules*, 2014, **47**, 8190–8195.
- 41 J. J. P. Stewart, *J. Mol. Model.*, 2007, **13**, 1173–1213.

