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Reactive block copolymers for patterned surface immobilization with sub-30 nm spacing†

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Phase-segregating block copolymers are powerful platforms for nanofabrication, particularly when employed as lithographic mask precursors. Surface-reactive polymeric films with distinct sub-30 nm domains are also proposed as covalent docking platforms for scalable, high-resolution molecular patterned immobilization. Here, the well-known self-assembling polystyrene-*block*-polyisoprene system is the starting point to produce a small library of derivatives with distinct reactive pendant groups (halide, azide, pentafluorophenylalkyl) by nitroxide-mediated radical polymerization. We find that controlling film thickness is crucial to obtain a perpendicular lamellar morphology and that the presence of the functional groups has a limited impact on self-assembly, yet may influence characteristic domain dimensions. Differential scanning calorimetry (DSC), small-angle X-ray scattering (SAXS), and atomic force microscopy (AFM) are utilized in concert to assess the phase behavior of the polymers and the surface features of the nanostructures. As a proof-of-concept for the surface reactivity, click chemistry-driven immobilization of a model water-soluble polymer is evidenced by X-ray photoelectron spectroscopy (XPS) and preservation of the underlying morphology is investigated by AFM.

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Introduction

Surface chemical patterning with spatial resolution exceeding 50 nm is a challenge which has so far only been met by a few methods, *e.g.*, dip-pen nanolithography,¹ e-beam lithography,² scanning near-field photolithography.^{3,4} Despite being sophisticated, these techniques bear drawbacks such as high costs of required apparatus and long processing times in case of large-area patterning, due to the inherent nature of top-down approaches. Only nanocontact printing has already been able to produce sub-50 nm features in a high throughput fashion on large areas, yet adjustment of the feature intervals at such a low scale remains complicated.⁵ Therefore, alternative methods for patterned covalent immobilization on surfaces at

such reduced length scales are highly sought after. In this respect, block copolymer-based technologies have gained significant attention in the last decades. Indeed, the ability of block copolymers (BCPs) to phase-separate and self-assemble into nanostructured motifs frequently led experts to regard them as having great potential in micro- and nanotechnology.^{6–9}

The most studied type of BCPs is that of linear AB diblock copolymers, which can exhibit various equilibrium structures, with lamellae and cylinders being those relevant for patterning. The relative volume fractions of the two blocks ($f_A + f_B = 1$) and the segregation strength (a function of the Flory–Huggins interaction parameter χ_{AB} and of the overall degree of polymerization N) govern the microphase separation in bulk.^{10–13} In thin films, however, there can be additional factors such as the air/polymer and polymer/substrate interactions. The thickness of the films can also alter the microdomain orientation and consequently the morphology displayed at the surface of the film,¹⁴ which is the crucial feature for soft chemical patterning of molecules. To date, nanostructured block copolymer thin films have been employed almost exclusively for so-called “block copolymer lithography”, *i.e.*, as masks/templates for inorganic nanostructures in the areas of electronics, catalysis, or optics, among other fields.^{15–17}

Yet, the stunning phase-segregating property of BCPs can be further exploited in order to align (bio)molecules in close proximity with each other in nanodimensions (10–50 nm). While the race for the smallest pitch size is fierce,^{18–27} with

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future storage and microelectronic devices in mind, many applications may not require such an extreme resolution. For instance, BCP nanostructures with a 20–50 nm period could be employed to design advanced catalytic systems and biosensors based on enzyme cascades, where confinement and proximity between active single components is an important aspect.²⁸ The literature offers a few examples of patterned molecular assemblies directed by BCPs through simple preferential adsorption on a specific domain.^{29–36} Functional groups introduced in the block copolymer structure logically enables a higher level of control. If expressed at the surface of the nanostructured thin films, they should behave as selective anchoring points for molecules *via* specific chemical reactions. Only a handful of examples for this approach can be found so far.^{33,37,38} For instance, Lynn and co-workers reported the patterning of a fluorescent amine on a BCP nanostructured surface of which one block was entirely made of a custom, functional amine-reactive monomer unit.³⁸ Our idea is that it would be a more versatile method to employ a well-studied phase-segregating BCP system and introduce a functional comonomer during the synthesis in an amount that is (i) sufficient for further functionalization, (ii) yet low enough to preserve the phase behavior of the original, non-functional system. This way, every time one desires to employ a different functionality, tedious investigations of the phase behavior would potentially not be necessary. Although some studies showed that small changes in structures can alter the morphologies and nanodomain periodicities,^{39,40} a few reports suggested that incorporation of a small percentage (<10%) of functionalities into or at the end of the polymer backbones does not influence the phase separation behavior to a considerable extent.^{37,41–44} During the course of our study, Wiesner, Estroff, and co-workers have used such an elegant and versatile approach with a well-known phase-separating polystyrene-*b*-polyethylene glycol scaffold (PS-*b*-PEG), where a significant amount of allyl glycidyl ether was introduced in the PEG block to provide allyl anchors for further thiol-ene-based functionalization.⁴⁵ Nevertheless, only one type of functionality was reported and, in fact, the polymerization method which was used, *i.e.*, anionic polymerization, may limit the range of accessible functionalities.

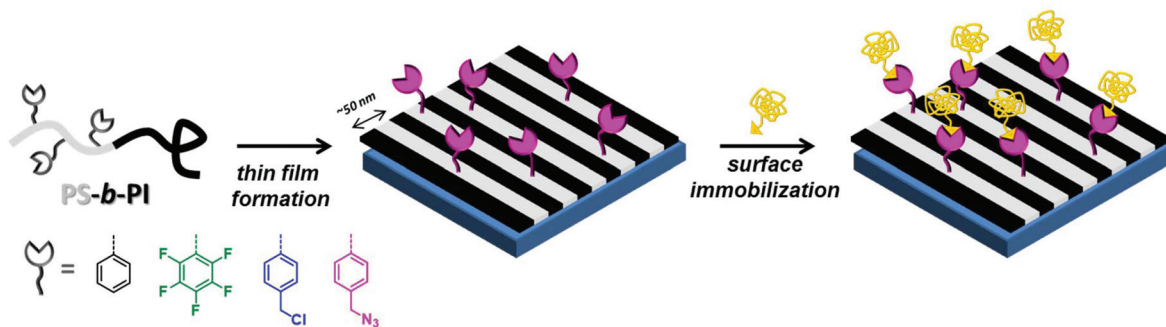
We reported very recently a library of mono- or bifunctional PMMA-*b*-PS BCPs and their self-assembly behavior in thin films, where minor changes in spacing were generally observed.⁴⁶ In parallel, we have been working on lamellae-forming polystyrene-*block*-polyisoprene (PS-*b*-PI) – a very well-known phase-segregating system^{47–54} – which has been modified in the PS block with a variety of functional comonomers. Various styrenic derivatives are commercially available or readily synthesized, while polyisoprene possesses inherent reactivity through the remaining double bonds present after synthesis. Our small set of functional PS-*b*-PI derivatives offers potential reactivity in ene reactions (in the PI block), halide nucleophilic substitution, azide-alkyne 1,3-dipolar cycloaddition, and *para*-fluoro-thiol reaction. Here, together with the synthesis and characterization of the block copolymers, as well as the investigation of their self-assembly behavior, we present the azide-alkyne variant as a preliminary and exemplary functionalization (Scheme 1).

Experimental section

Materials

Styrene (99%, Acros), 2,3,4,5,6-pentafluorostyrene (PFS; 98%, ABCR), and *N,N*-dimethylacrylamide (DMAAm; 99%, Acros) were passed through a short basic alumina column prior to use. Isoprene (99%, ABCR) was distilled at 40 °C prior to use. 2,2'-Azobis(isobutyronitrile) (AIBN; 98%, Sigma Aldrich) was recrystallized from methanol. 4-Vinylbenzyl chloride (97%, Sigma-Aldrich), copper(II) sulfate pentahydrate (CuSO₄·5H₂O; ≥ 99.5%, Roth), tetrabutylammonium fluoride solution (TBAF; 1.0 M in THF, Sigma Aldrich), butylated hydroxytoluene (BHT; 99%, Alfa Aesar), sodium azide (NaN₃, ≥99%, Roth), and magnesium sulfate (MgSO₄, ≥99%, Roth) were used as received. All following solvents were used as received: methanol and toluene were purchased from Fisher; isopropanol and pyridine were purchased from Roth; 1,4-dioxane, dichloromethane (DCM), and *N,N*-dimethylformamide (DMF) were purchased from VWR with high purity.

N-tert-Butyl-*N*-[1-diethylphosphono-(2,2-dimethylpropyl)] nitroxide (SG1),⁵⁵ 2-methyl-2-[*N*-tert-butyl-*N*-(1-diethoxyphosphoryl)-



Scheme 1 Schematic depiction of the concept of molecular patterning with reactive block copolymer films. Note that for clarity sake only a few surface-expressed anchors were depicted.



2,2-dimethylpropyl)aminoxy]propionic acid (MAMA-SG1),⁵⁶ tris(3-hydroxypropyltriazolylmethyl)amine (THPTA),⁵⁷ and 3-(trimethylsilyl)prop-2-yn-1-yl 2-(((dodecylthio)carbonothioyl)thio)propanoate (TMS-DOPAT)⁵⁸ were synthesized according to the previously reported procedures.

Si wafers (p-type, boron-doped, <100>, $\rho = 1\text{--}30 \text{ } \Omega \text{ cm}$) were purchased from Si-Mat (Kaufering, Germany). Ace pressure tubes (Ref. 8648-75) equipped with a PTFE plug (Ref. 5845-47) were purchased from Sigma Aldrich. A benchtop dual wavelength CAMAG lamp was used for photocrosslinking at 254 nm (8 W).

Characterization

Size-exclusion chromatography (SEC). For PS and PS-*b*-PI, SEC measurements were performed on a TOSOH Eco-SEC HLC-8320 GPC system, which comprised an autosampler, a SDV 5 μm bead size guard column ($50 \times 8 \text{ mm}$, PSS) followed by three SDV 5 μm columns ($300 \times 7.5 \text{ mm}$, subsequently 100, 1000, and 105 \AA pore size, PSS), and a differential refractive index (DRI) detector with THF as the eluent at 30 $^{\circ}\text{C}$ with a flow rate of 1 mL min^{-1} . The SEC system was calibrated by using linear polystyrene standards ranging from 266 to $2.52 \times 10^6 \text{ g mol}^{-1}$, with linear PMMA standards ranging from 800 to $1.82 \times 10^6 \text{ g mol}^{-1}$. Calculation of the molar masses proceeded by using the Mark-Houwink-Sakurada (MHS) parameters for PS in THF at 30 $^{\circ}\text{C}$, that is, $K = 14.1 \times 10^{-5} \text{ dL g}^{-1}$, $\alpha = 0.70$.⁵⁹

For PDMAAm, SEC measurements were performed on a Polymer Laboratories (Varian) PL-GPC 50 Plus Integrated System, comprising an autosampler, a PLgel 5 μm beads size guard column ($50 \times 7.5 \text{ mm}$) followed by three PLgel 5 μm Mixed-C columns ($300 \times 7.5 \text{ mm}$) and a differential refractive index detector using DMAc containing 0.3 wt% LiBr as the eluent at 50 $^{\circ}\text{C}$ with a flow rate of 1 mL min^{-1} . The SEC system was calibrated using linear polystyrene standards ranging from 160 to $6 \times 10^6 \text{ g mol}^{-1}$ and linear poly(methyl methacrylate) standards ranging from 700 to $2 \times 10^6 \text{ g mol}^{-1}$. Calculation of the molar mass proceeded by using the Mark-Houwink-Sakurada (MHS) parameters for PMMA in THF at 30 $^{\circ}\text{C}$, that is, $K = 12.8 \times 10^{-5} \text{ dL g}^{-1}$, $\alpha = 0.69$.⁶⁰

NMR spectroscopy. Measurements were performed on a Bruker AM 500 spectrometer at 500 MHz. The analytes were dissolved in CDCl_3 and the residual solvent signals were employed for shift correction. For the ^{19}F -CPD NMR spectrum, at least one of the following corrections was made by using MestReNova 9.0 software: baseline correction with the method Bernstein polynomial fit (with polynomial order of 14), manual correction, or multipoint baseline correction.

Differential scanning calorimetry (DSC). Thermographs were obtained using a Q200 differential scanning calorimeter from TA Instruments. Each sample was heated first to +150 $^{\circ}\text{C}$ at a rate of 10 $^{\circ}\text{C min}^{-1}$ to erase the thermal history of the sample, then cooled down to -90 $^{\circ}\text{C}$. For determination of the glass transition temperatures, samples were then heated from -90 to +150 $^{\circ}\text{C}$ at a rate of 10 $^{\circ}\text{C min}^{-1}$.

Small-angle X-ray scattering (SAXS). The block copolymer morphology and its orientation were determined using a S3-Micro instrument from Hecus X-ray Systems GmbH (Graz,

Austria) combined with a 2D CCD detector from Photonic Science. The distance between sample and detector was 280.1 mm. The pixel size of the detector was 29 μm in each dimension. With this combination, a q range from 0.08 to 5 nm^{-1} was accessible. The polymers were annealed for 24 h at 150 $^{\circ}\text{C}$ under vacuum in O-rings (2 mm diameter) serving as sample supports. Measurements were directly performed in these O-rings with the X-ray beam position adjusted to the middle of the sample. Both the background signal of the CCD detector and the scattering of the system itself were subtracted after radial averaging of the data. No correction for the final dimension of the beam has been performed, therefore the scattering peaks are broadened compared to a measurement using a SAXS beamline at a synchrotron facility. The beam dimensions were 0.3 mm \times 0.5 mm (FWHM), with a 100 μm entrance slit in scattering direction and a 400 μm side limit vertical to the scattering direction.

Ellipsometry. Film thicknesses were determined with a spectroscopic ellipsometer (J. A. Woollam) with a wavelength range of 400–800 nm at a 75 $^{\circ}$ angle of incidence, on at least three different points of each film. A three-layer model consisting of a Si layer, a 3 nm SiO_2 layer, and a Cauchy layer was used to simulate experimental data.

Atomic force microscopy (AFM). Micrographs were acquired with a MultiMode 2 Bruker instrument (MMAFM-2) using tapping mode. The probes used during AFM analysis were n-type silicon probes HQ:NSC14/Al BS (160 kHz, 5 N m^{-1}) and HQ:NSC35-No Al (150–300 kHz, 5.4–16 N m^{-1}) and were purchased from MikroMasch. AFM images were acquired using the Nanoscope software. Characteristic domain spacing values L_0 were obtained by performing Fast Fourier transforms of the AFM images using the Gwyddion software, measuring at least 10 profiles across the obtained circle patterns and averaging the radius values, corresponding to L_0 .

X-ray photoelectron spectroscopy (XPS). XPS investigations were performed on a K-Alpha+ spectrometer (ThermoFisher Scientific, East Grinstead, UK) using a microfocused, monochromated Al K α X-ray source (400 μm spot size). The kinetic energy of the electrons was measured by a 180 $^{\circ}$ hemispherical energy analyzer operated in the constant analyzer energy mode (CAE) at 50 eV pass energy for elemental spectra. Data acquisition and processing using the Thermo Avantage software is described elsewhere.⁶¹ The spectra were fitted with one or more Voigt profiles (BE uncertainty: $\pm 0.2 \text{ eV}$). The analyzer transmission function, Scofield sensitivity factors and effective attenuation lengths (EALs) for photoelectrons were applied for quantification.⁶² EALs were calculated using the standard TPP-2 M formalism. All spectra were referenced to the C 1s peak (C-C, C-H) at 285.0 eV binding energy controlled by means of the well-known photoelectron peaks of metallic Cu, Ag, and Au, respectively. The signals presented in Fig. 5 were normalized according to the signal amplitude of the baseline.

Synthesis

Polystyrene macroinitiator MI1. In a 25 mL round bottom flask, styrene (10.36 g, 99.4 mmol) was mixed with MAMA-SG1



(98.1 mg, 0.26 mmol) and the flask was sealed with a rubber septum. Subsequently, the content was purged with N_2 for 30 min and placed in a preheated oil bath at 120 °C. After 2 h, the flask was cooled to ambient temperature. The resulting polymer was precipitated twice in cold methanol and **MI1** was recovered as a white powder (Scheme S1† and Fig. 1 left).

Polystyrene-*b*-polyisoprene (PS-*b*-PI) BCP1. **MI1** (100.6 mg) was dissolved in pyridine (1280 μ L) in a pressure tube. The tube was sealed with a rubber septum and the content was purged with N_2 for 30 min. A large amount of isoprene was placed in a flask which was then sealed with a rubber septum and purged with nitrogen while being in an ice bath. The isoprene flask and the pressure tube were taken into a glovebox and isoprene (1280 μ L, 870.4 mg, 12.78 mmol) was added into the pressure tube before the latter was quickly sealed with a fitted screw cap. Outside of the glovebox, the pressure tube was placed in a preheated oil bath at 115 °C. After 13 h 40 min, the reaction was stopped by cooling the flask to ambient temperature. Pyridine was removed under vacuum. The residue was dissolved in a minimal amount of THF and precipitated in a methanol/isopropanol mixture (3 : 1 v/v) including a small amount of BHT as stabilizer. **BCP1** was recovered as a white powder (Fig. 1 left and S5,† and Scheme S1†).

Pentafluorophenyl-functionalized PS macroinitiator MI2. Styrene (4.63 g, 44.5 mmol), PFS (449 mg, 2.3 mmol), and MAMA-SG1 (46.1 mg, 0.12 mmol) were mixed in a 10 mL round bottom flask. The flask was placed in a preheated oil bath at 120 °C after being purged with N_2 for 30 min. The reaction mixture was cooled to ambient temperature after 135 min. The same purification steps were applied as for **MI1** and **MI2** was recovered as white powder (Fig. 1 middle and Scheme S2†).

Pentafluorophenyl-functionalized PS-*b*-PI BCP2. **MI2** (200.6 mg) was dissolved in pyridine (2560 μ L) in a pressure tube. The same procedure was applied as for the synthesis of **BCP1** with isoprene (2560 μ L) for a total reaction time of 13 h 55 min. The same purification steps were applied and yielded

BCP2 as a white powder (Fig. 1 middle and S5, and Scheme S2†).

Chloromethylbenzyl-functionalized PS macroinitiator MI3. Styrene (3.67 g, 35.1 mmol), 4-vinylbenzyl chloride (282 mg, 1.84 mmol), and MAMA-SG1 (35.5 mg, 0.09 mmol) were mixed in a 10 mL round bottom flask. The flask was placed in a preheated oil bath at 120 °C after being purged with N_2 for 30 min. The reaction mixture was cooled to ambient temperature after 135 min. The same purification steps were applied as for **MI1** and **MI3** was recovered as a white powder (Fig. 1 right and Scheme S3†).

Chloromethylbenzyl-functionalized PS-*b*-PI BCP3. **MI3** (200.4 mg) was dissolved in 1,4-dioxane (2560 μ L) in a pressure tube. The same procedure was applied as for the synthesis of **BCP1** with addition of isoprene (2560 μ L) and the reaction was run for a total time of 15 h. The same purification steps were applied to yield **BCP3** as a white powder (Fig. 1 right and S5, and Scheme S3†).

Azidomethylbenzyl-functionalized PS-*b*-PI BCP4. **BCP3** (100.8 mg) was dissolved in THF (3 mL). DMF (50 mL) was added. NaN_3 (10 mg, 0.15 mmol) was dissolved in this solution. The content was stirred at RT for 4 days. Solvents were removed in vacuum. Subsequently, the product was dissolved in DCM (20 mL) and extracted with water (3 \times 80 mL). The organic layer was dried over $MgSO_4$, filtered, and concentrated in vacuum. **BCP4** was precipitated in a methanol/isopropanol mixture (3 : 1 v/v) including a small amount of BHT as stabilizer and obtained as a white powder (Fig. 1 right, S2,† and S5†).

Alkyne-functionalized poly(*N,N*-dimethylacrylamide) \equiv -PDMAAm. DMAAm (461.1 mg, 4.65 mmol), TMS \equiv -DOPAT (43 mg, 0.093 mmol), and AIBN (6.1 mg, 0.037 mmol) were dissolved in 1,4-dioxane (1.857 mL) in a 5 mL round bottom flask. The flask was sealed with a rubber septum. The content was purged with N_2 for 30 min and then heated to 70 °C for 15 min, time after which the flask was brought to ambient temperature. The solvent was removed and the residue dissolved in THF and precipitated twice in cold pentane, yielding TMS \equiv -PDMAAm as a sticky yellow solid. SEC (DMAC): $M_n = 4090$ g mol $^{-1}$; $D = 1.09$ (Scheme S4, Fig. S10 and S11†).

TMS \equiv -PDMAAm (155 mg) was dissolved in THF (865 μ L). TBAF (80 μ L, 80 μ mol) was added dropwise. The content was stirred for 1 h at RT. \equiv -PDMAAm was precipitated in cold pentane and recovered as a sticky yellow polymer (Scheme S4, Fig. S10 and S11†). SEC (DMAC): $M_n = 4180$ g mol $^{-1}$; $D = 1.08$.

Surface experiments

Thin film preparation. Freshly cut 1 cm 2 Si wafers were placed in acidic piranha solution ($H_2SO_4:H_2O_2$ 3 : 1 v/v) (Caution! Explosive when in contact with organic solvents!) at room temperature for 45 min for substrate activation. The wafers were then thoroughly rinsed with distilled water and ethanol, consecutively, and finally dried in a stream of N_2 . Afterwards, a 0.2 wt% solution of PS-*b*-PI in toluene was spin-coated on the previously activated Si substrates at 10 000 rpm for 5 min resulting in a film thickness of ca. 7 nm.

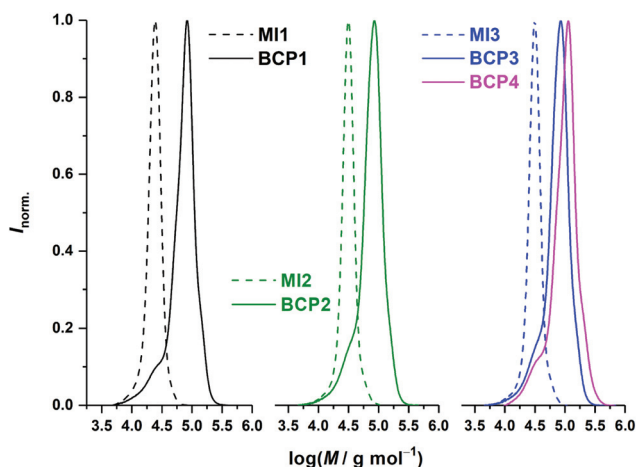


Fig. 1 SEC traces of (left) **MI1** and **BCP1**, (middle) **MI2** and **BCP2**, and (right) **MI3**, **BCP3**, and **BCP4**.



Functionalization of BCP4 films by copper-catalyzed azide-alkyne cycloaddition. Two aqueous solutions containing the CuAAC catalytic components were prepared: a 0.4 M CuSO₄ and 0.4 M THPTA on the one hand and a 1.4 M sodium ascorbate on the other hand. Two BCP4-coated Si wafers (film thickness *ca.* 7 nm) were placed in 5 mL vials separately. The first vial was used for the reaction whereas the second one was used as a control experiment. For the reaction sample, water (550 μL) was added, followed by the addition of ≡-PDMAAm solution (200 μL, 1 mg mL⁻¹ in water). The CuSO₄/THPTA solution (30 μL) was added. Finally, the sodium ascorbate solution (70 μL) was added. The total volume was 850 μL. The vial was closed and entire immersion of the wafer with the coated side facing upwards was ensured. For the control sample, only water (650 μL) and the ≡-PDMAAm solution (200 μL, 1 mg mL⁻¹) were added, omitting the entire catalytic system. Both samples were placed on a benchtop shaker at a speed of 150 rpm for 1 hour at room temperature. The samples were then taken out and rinsed thoroughly with water for 10–20 seconds. Afterwards, they were placed in conical tubes filled with water (14 mL) and placed back onto the shaker at 150 rpm for 15 min. Water was renewed once and the samples incubated for another 15 minutes, before being eventually dried in a stream of air.

UV-induced crosslinking of BCP4 thin films. A BCP4-coated sample (*ca.* 7 nm) was placed in a quartz flask connected to a vacuum pump and was crosslinked by UV light irradiation at $\lambda = 254$ nm for 15 min using a classic thin-layer chromatography UV lamp (CAMAG UV Lamp 4, 8 W), placed at 1 cm from the film. Afterwards, the film was subjected to successive washings with multiple solvents, *e.g.*, acetone, toluene, DCM, and was finally incubated in DMSO for 30 min.

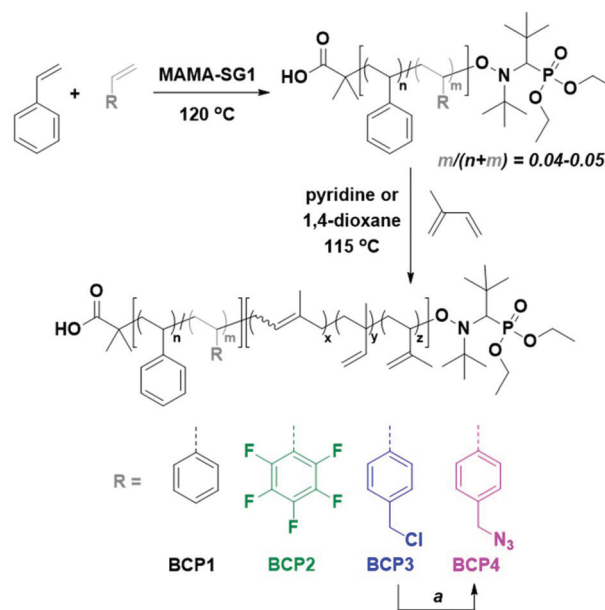
Results and discussion

Synthesis of functional PS-*b*-PI derivatives

In contrast to most studies on PS-*b*-PI employing anionic polymerization, we synthesized our BCPs using nitroxide-mediated polymerization (NMP), because radical polymerization is known to be tolerant to a wide range of functional moieties.⁶³ The nitroxide SG1 has recently been shown to enable a good control over isoprene NMP.^{64,65} To the best of our knowledge, we present the synthesis of such functional BCPs by SG1-mediated NMP for the first time. In fact, we have used the present library of functional PS-*b*-PI BCPs in a recent article dealing with nanoprecipitation.⁶⁶ Yet the polymerization data had yet to be thoroughly presented. As our intention was to obtain stripe patterns which originate from lamellae-based systems, the synthesis of BCPs with block of near-identical volume fraction was necessary. In addition, we targeted a rather small domain size, having in mind to evidence the potential of the approach to compete with other nanopatterning techniques (*vide supra*). An inspection of the literature revealed that a molar mass of about 40 000 g mol⁻¹ would yield domain sizes in the sub-50 nm range.³⁰ When one con-

siders the synthesis of a diblock copolymer, two routes are possible with respect to the order in which the blocks will be created. In the present case, either a polyisoprene or a polystyrene macroinitiator could be synthesized first. The former route is certainly the most convenient to establish a library of PS-*b*-PI BCPs with various comonomers in the PS block, since a single PI-SG1 macroinitiator could serve as a starting point. However, PI possessing a very low glass transition temperature (T_g) and consequently behaving like a viscous oil is not practical to handle. We therefore directed our efforts towards the synthesis of a set of functional PS-SG1 macroinitiators, which were then chain-extended with isoprene. The general synthetic strategy for the set of functional PS-*b*-PIs is given in Scheme 2.

We first established the synthetic procedure for plain PS-*b*-PI starting with the alkoxyamine MAMA-SG1 as the initiator. The synthesis of PS-SG1 is straightforward and already extensively reported. To maximize the livingness of the macroinitiator, conversion was limited to 50%. PS-SG1 **MI1** with a number-average molar mass of approx. 22 kg mol⁻¹ and a dispersity value of 1.1 was obtained (Fig. 1, left, dashed line), and after purification, was subsequently extended with isoprene to yield **BCP1**. For this second step, conditions similar to those reported for the synthesis of PI-SG1 by Nicolas and co-workers were employed, *i.e.*, benzyl-type SG1 alkoxyamine initiator (low molar mass analogue of PS-SG1), 115 °C, and pyridine as an accelerating solvent.^{64,65} A low molar mass tailing was observed, which seems to indicate the presence of a fraction of non-reacted PS dead macroinitiator or potentially extended with a short segment of isoprene (Fig. 1, left, full line). This distribution pattern was not observed for the low molar mass initiator, which is certainly due to the fact that it would lead to species not detectable in SEC. It must however



Scheme 2 Chemical structures of the PS-*b*-PI derivatives employed in the current study and corresponding synthetic route. (a) NaN₃, DMF, RT.



be noted that the presence of homopolymer should not prevent phase separation and may only alter domain size.⁶⁷ We subsequently followed the same protocol for functional BCP derivatives by introducing 5 mol% of a functional styrene derivative in the synthesis of PS macroinitiators, namely, pentafluorostyrene (PFS) and vinylbenzyl chloride (VBC) (Scheme 2). Each macroinitiator displayed a dispersity value of 1.1 (Fig. 1, middle and right, dashed lines; Table S1†). Using a functional analogue of the main comonomer theoretically favors straightforward statistical incorporation of functionality. Indeed, VBC exhibited an ideal behavior: a feed of 5 mol% resulted in 4.8 mol% incorporation in **MI3**. However, from previous studies,^{68–70} it was known that PFS reacts faster than styrene in copolymerization. Therefore, to limit its incorporation below 5 mol%, the initial comonomer mixture contained 3.2 mol% of PFS. While styrene conversion reached 55%, that of PFS was 67%, leading to 4 mol% functionality in the corresponding macroinitiator **MI2**, subsequently used to synthesize **BCP2** (Fig. S1†).

As mentioned above, pyridine is used as a solvent for the synthesis of the polyisoprene block. It was however not suitable for the synthesis of **BCP3**, due to the presence of labile chlorine atoms in **MI3**. Indeed, pyridine is a basic solvent and a good medium for dehalogenation.⁷¹ Therefore, **BCP3** was obtained by polymerization in 1,4-dioxane, which was also reported as an acceptable medium for the NMP of isoprene.⁶⁵ To obtain **BCP4**, direct incorporation of 4-(azidomethyl)styrene in a macroinitiator was not possible due to its instability at the employed polymerization temperatures. Instead, **BCP4** was obtained by azidation of **BCP3**, according to a procedure reported in the literature (Scheme 2).⁷² Quantitative conversion was confirmed by ¹H NMR spectroscopy (Fig. S2†). Table 1 compiles all characterization data obtained for the block copolymers. It must be noted that during chain extensions, for a conversion in isoprene superior to 10% significant shoulder formation in the high molar mass region was observed. This imposed the use of large monomer-to-macroinitiator ratios and low conversion. Even reasonable ratios which would lead to the desired chain lengths at conversions of 30% or 20% started to show this phenomenon at ca. 15% (Fig. S3†), which could suggest early termination reactions. Despite further optimization attempts, a slight shoulder at high molar masses persists in our samples at conversions of approx. 10%. Notwithstanding, all BCPs exhibit reasonably narrow distributions ($D = 1.32–1.36$). Using PS and PI density values (1.05

and 0.913, respectively), final volume fractions were calculated by ¹H NMR: f_{PI} was found to be in the 0.50–0.54 range, which is ideal to produce lamellar phases. It is to note that for the calculation of volume fractions the functional styrene derivatives were treated as styrene units, *i.e.*, as if they had no impact on the density of a plain PS. Although differences cannot be excluded, these minor fluctuations would still surely allow the BCP compositions to remain within the range corresponding to a lamellar regime.

Differential scanning calorimetry measurements

Thermal characterization can be exploited to obtain information on the phase behavior of polymeric materials which possess several structural motifs. For an AB-type block copolymer, provided phase separation occurs, differential scanning calorimetry (DSC) measurements should reveal two glass transition temperatures (T_g) which roughly match those of the corresponding homopolymers A and B.⁷³ PS-*b*-PI consists in one *soft* segment (PI) and one *hard* segment (PS). PS is glassy at room temperature, due to a T_g typically comprised between 95 and 100 °C.^{74–79} The T_g of polyisoprene varies with the fractions of isomeric repeating units ((3,4), (1,2), (1,4)), as well as of *cis* and *trans* diastereoisomeric forms of the (1,4) unit.⁸⁰ These distributions are themselves highly depending on the polymerization technique employed for the synthesis. For instance, both being obtained by Ziegler–Natta polymerization with specific catalysts, *cis*-1,4-polyisoprene possesses a T_g of ca. –70 °C while that of *trans*-1,4-polyisoprene is about 10 °C higher.⁷⁴ Here, the PI block is obtained by SG1-mediated NMP, which typically leads to a monomer unit distribution dominated by the (1,4) motif (>80 mol%) along 10–15 mol% of (3,4) units, with (1,2) motifs forming the minor fraction (Fig. S4†).⁶⁴ In any case, the T_g of polyisoprene is well below room temperature.

Fig. 2 compiles the DSC thermographs obtained for the four PS-*b*-PI derivatives. All measurements revealed two clear glass transitions ($T_{g,1} < T_{g,2}$). This fact alone validates the ability of all BCPs to undergo phase separation. $T_{g,1}$ values lie in the –64(–61) °C range and correspond to the PI block. In spite of the various incorporated comonomers, the glass transition temperatures of PS-based blocks ($T_{g,2}$) span a narrower range (93.5–95 °C). In the present case, the thermal behavior of PS-*b*-PI is thus not significantly altered by the presence of small amounts of functional units.

Table 1 Characteristics of the block polymers reported in the present study

Name	$M_{n,SEC}^a$ (g mol ⁻¹)	D^a	$M_{n,NMR}^b$ (g mol ⁻¹)	$f_{PS}:f_{PI}$	T_g^c (°C)	$L_{0,SAXS}^d$ (nm)	$L_{0,AFM}^e$ (nm)
BCP1	57.7	1.36	44.5	0.46 : 0.54	–61.8/95.0	48.3	48.7 ± 1.8
BCP2	61.9	1.32	47.3	0.50 : 0.50	–63.4/95.0	48.3	45.3 ± 1.0
BCP3	61.8	1.36	49.4	0.47 : 0.53	–62.8/94.0	58.7	56.2 ± 1.1
BCP4	63.5	1.33	49.4	0.47 : 0.53	–63.6/93.6	51.5	59.1 ± 2.5

^a Values obtained by SEC. ^b Values obtained by NMR. ^c Values obtained by DSC. ^d Values obtained by SAXS. ^e Values obtained by AFM. Values reported for $L_{0,AFM}$ are mean ± SD.



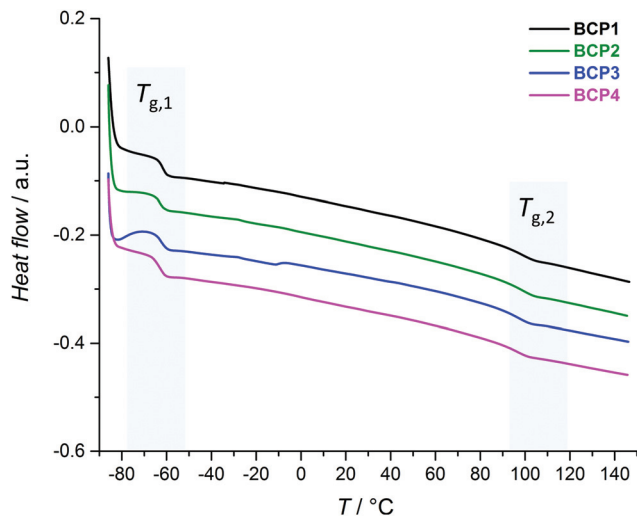


Fig. 2 DSC thermographs of PS-*b*-PI derivatives BCP1–4.

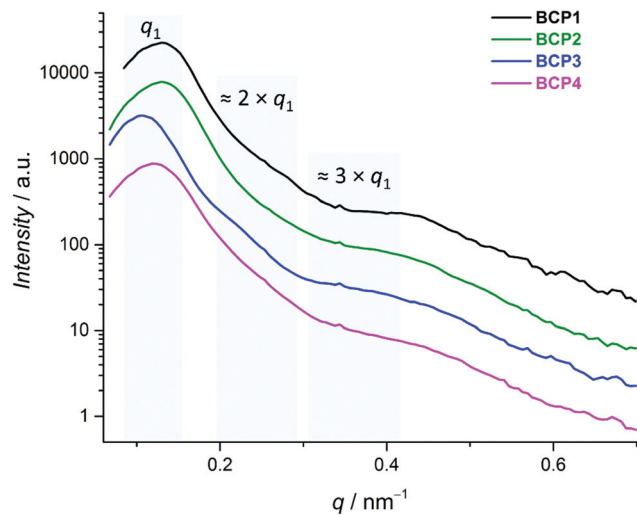


Fig. 3 SAXS profiles of PS-*b*-PI derivatives BCP1–4. For clearer representation, multiplication factors were applied.

Small angle X-ray scattering analysis

While DSC can evidence the occurrence of phase separation in a block copolymer system, it is unable to provide any structural or dimensional information. In consequence, we turned to small-angle X-ray scattering (SAXS) to characterize the bulk structure of the four PS-*b*-PI. SAXS can give access to the dimensions of nanoscale structural heterogeneities, that is in the case of BCPs, the mean values of the interdomain spacing, noted $L_{0,\text{SAXS}}$. From the scattering pattern, the nature of the ordered structures (*e.g.*, cylinders, lamellae) can also be obtained.⁸¹ While grazing-incidence SAXS (GI-SAXS) measurements would allow evaluation of the system in conditions similar to those envisioned for applications (*i.e.*, thin films), particularly giving information on the surface of the samples, access to related instruments is scarce. Therefore, classic SAXS measurement were carried out in the present study, reasoning that parallel observation by atomic force microscopy (*vide infra*) would provide sufficient morphological information.

The obtained SAXS profiles are presented in Fig. 3. All four samples display a comparable behavior: a main peak at a similar position ($q_1 = 0.11\text{--}0.13\text{ nm}^{-1}$) and a shoulder at approx. three times q_1 . At two times q_1 , only **BCP3** exhibits a faint inflexion.⁸² The absence of peaks at positions different than integer multiples of q_1 is absolutely characteristic of a lamellar structure. The fact that the second-order peak q_2 is mostly suppressed suggests that the two phases have nearly identical dimensions. This phenomenon is only witnessed in the occurrence of a lamellar structure. Indeed, one can find other examples in the literature in which the even-ordered peaks on SAXS profiles are less pronounced than the odd-ordered ones in the presence of equal volumes of two separate blocks.^{83–85} In combination with the molecular design based on volume ratio adequate for this phase separation mode,⁸⁶ one can confidently conclude to that all BCPs exhibit this type of bulk nanostructuration.

The first Bragg peak gives the periodicity of the system according to $L_{0,\text{SAXS}} = 2\pi/q_1$.⁸¹ In the strong segregation limit

($\chi N > 100$), this value is related to $N^{2/3}$, $\chi^{1/6}$, and an average segment length reflecting the stiffness of the chain.^{87,88} Bearing in mind these dependencies, a variation in χ of 50% would result in a change of periodicity of only 7%. Due to the low degree of functionalization in this study, the influence of a variation of χ can certainly be neglected (only as long as χN is great enough to remain in the segregation range). Therefore, only the chain length and the average segment length remain as parameters of interest. One possible measure for both is the radius of gyration, which displays a dependency on both the number of segments and the stiffness, *e.g.*, the average segment length. The variation of the molar mass in the current BCP series is at most *ca.* 5%, while the difference between the smallest and the largest $L_{0,\text{SAXS}}$ values is of about 21–22%. This clearly shows that the small molar mass fluctuations are not responsible for the observed variations in periodicity and points rather at the influence of the introduced functional groups, mainly by alteration of the stiffness and hence the average segment length. Bulkier side groups occupy more space and more polar groups will bring additional interactions between the polymeric chains. The first contribution should increase the stiffness of the chain and therefore the radius of gyration, while the influence of the latter is difficult to predict. Taking into account the relatively similar dimensional characteristics of hydrogen and fluorine and of their respective bonds with carbon atoms, the pentafluorophenyl group is supposed to possess a similar bulkiness compared to a simple phenyl ring. In that case, the difference in polarity does not seem strong enough to induce a change in nanostructure periodicity, as both **BCP1** and **BCP2** exhibit the same q_1 value (Fig. 2B), hence an identical $L_{0,\text{SAXS}}$ (48.3 nm; Table 1).

However, **BCP3** and **BCP4** possess a significantly higher interdomain spacing ($L_{0,\text{SAXS}} = 58.7$ and 51.5 nm, respectively). This could be explained by an increased bulkiness as a pendant group of atoms is added at the *para* position of the phenyl ring. The notably pronounced difference between the



two is however rather unexpected, even though chlorides may be more polar than azides of the same structure⁸⁹ and could possibly induce a change. Conclusions should therefore be drawn with cautiousness. Nevertheless, all copolymers gratifyingly yield bulk structures containing sub-50 nm heterogeneities which can be attributed to a lamellar morphology and are thus potential candidates for chemical patterning.

Formation of surface-nanostructured films

As DSC measurements indicated the ability of **BCP1–4** to phase separate and a bulk lamellar structure for all samples was evidenced by SAXS, the fabrication of reactive nanostructured thin films was confidently tackled. Films were obtained by spin-coating on activated Si wafers (1 cm²). After film formation, an annealing step is often carried out to induce an effective pattern formation or at least to enhance the ordering. For instance, for PS-*b*-PI systems some studies report thermal annealing.^{90–93} Notwithstanding, Keddie and co-workers employed PS-*b*-PI thin films without annealing, as coated.³⁰ A similar strategy was employed for **BCP1–4**: the films were investigated by atomic force microscopy (AFM) directly after spin-coating (Fig. 4). As a starting point, a solution of the non-functional PS-*b*-PI **BCP1** in toluene was investigated. Being the softer component and therefore dissipating more energy during AFM analysis, PI domains appear darker.⁹⁴ Based on the aforementioned NMR calculations and SAXS study, lamellae are expected, ideally with a perpendicular orientation in the view of patterning applications. However, it is well known that the domain orientation depends on interfacial interactions of the polymer segments with the substrate

on the one hand and the atmosphere on the other hand. In a similar setup, the report of Keddie and co-workers pointed out that PI preferentially wets silicon substrates (*i.e.*, the native oxide layer covering any non-etched Si wafer), which led to PI circular dot patterns at the surface of the film (polymer–air interface) for PS-*b*-PI films above a given thickness.³⁰ In the current study, the same phenomenon was witnessed for thicknesses of 33 and 20 nm (as measured by ellipsometry; see Fig. S7A, B and D†). Increasing spinning rate and time as well as reducing polymer concentration during the spin-coating step typically lead to a decrease of film thickness. Here, for a 0.2 wt% polymer solution spun at 10 000 rpm for five minutes, an ultra-thin film exhibiting a thickness of *ca.* 7 nm was obtained. This value is significantly lower than $L_{0,SAXS}$, which has the consequence of forcing the phases to accommodate a perpendicular orientation, leading to the observation of so-called stripe-like patterns in atomic force micrographs (Fig. 4A), typical for perpendicular lamellae without long-range order.⁹⁵

In most lithographical studies based on BCP thin films, thicknesses close or equal to one domain spacing are employed.⁹⁶ Although more intricate structures can form for BCP film thicknesses below L_0 , lamellar nanodomains can indeed orient perpendicularly to the substrate in such cases.⁷ This morphological behavior was previously described in theoretical and experimental studies,^{97,98} particularly for PS-*b*-PI.³⁰ Consequently, an identical coating strategy was implemented for the three functional PS-*b*-PIs **BCP2–4**. All 4 BCPs have sensibly the same volume fractions (Table 1 and Fig. S5†) and behave similarly in bulk, as suggested by the SAXS data. Comparable patterns are therefore expected. However, the presence of functional groups could lead to alterations due to modification of interfacial interactions. Working at ultra-low thicknesses may have prevented any issue of the sort because very similar morphologies were indeed found for all PS-*b*-PIs (Fig. 4).

In order to carry out statistical measurements, for each BCP, multiple films were produced. For each of them, several areas were analyzed. Similar morphologies were consistently observed. However, note that the “nonequilibrium in-frozen” nature of the films³⁰ precludes one to claim perfect reproducibility. For advanced applications, it may be worthwhile to investigate the opportunity of solvent annealing to provide improved domain ordering. Nevertheless, here, the periodicity of the surface line patterns, *i.e.*, $L_{0,AFM}$, was determined by applying fast Fourier transform and determining the radius of the characteristic circular pattern (see insets in Fig. 4). The obtained values are comprised between 45 and 59 nm and are rather congruent with SAXS results, with only minor deviations (1–12% range). Note also that height-based AFM analysis showed a variation in the range of 2 nm – that is, below the film thickness – corresponding to a full surface coverage, as previously observed in such PS-*b*-PI thin films.³⁰

Surface functionalization by copper-catalyzed azide–alkyne cycloaddition

Following the aforementioned structural investigations, and particularly the successful formation of binary surface

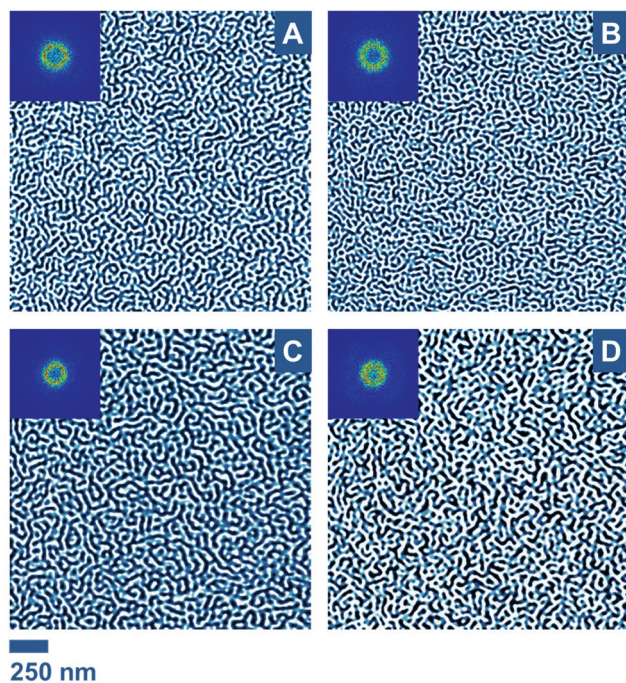


Fig. 4 AFM phase images of ultra-thin films of (A) **BCP1**, (B) **BCP2**, (C) **BCP3**, and (D) **BCP4**, all obtained in identical spin-coating conditions (0.2 wt% in toluene, 10 000 rpm, 5 min). Insets are fast Fourier transform (FFT) images.



patterns, surface functionalization could be tackled by using the ultra-thin films. Since PS and PI are both relatively hydrophobic polymers,⁹⁹ PS-*b*-PI was assumed to be stable in water. Therefore, the current films were considered suitable candidates as platforms for reactions in aqueous media.

As previously mentioned, our small series of functional BCPs encompasses reactivity in nucleophilic substitution ($-\text{CH}_2\text{Cl}$), copper-catalyzed azide-alkyne cycloaddition (CuAAC) ($-\text{CH}_2\text{N}_3$), *para*-fluoro thiol substitution (pentafluorophenyl moieties), and ene reactions (PI double bonds). Here, we focus on CuAAC as a proof of concept. Further studies concerning the other ligations are underway. CuAAC has been in the last decade one of the most popular reactions in materials chemistry, and particularly for surface functionalization.^{100–105} To assess the availability of the azide groups at the surface of the films made of **BCP4**, an alkyne-functionalized water-soluble poly(*N,N*-dimethylacrylamide)-based polymer ($\equiv\text{-PDMAAm}$) was synthesized (Fig. 5A and Scheme S4†). As each of its repeating units carries an amide moiety, PDMAAm is convenient for investigating the surface reaction by X-ray photoelectron spectroscopy (XPS).¹⁰⁶ In addition, since $\equiv\text{-PDMAAm}$ was obtained by RAFT polymerization and is of rather low mass, XPS should enable the detection of sulfur-related peaks arising from the trithiocarbonate end chain. For unambiguous

characterization, a control experiment was conducted in the absence of the copper catalyst. XPS results indicate a clear increase in nitrogen content upon reaction from 0.4 at% in the initial **BCP4** film sample to 3.8 at% in the so-called CuAAC sample (Fig. 5B, left). Particularly, a clear signal at 400.2 eV is observed and can be attributed to the amide group of $\equiv\text{-PDMAAm}$.¹⁰⁷ The control sample also shows a higher nitrogen content than the initial sample, yet to a significantly lower extent (from 0.4 to 1 at%), suggesting a potential limited physisorption of the polymer. The 167–160 eV region of the XPS spectrum is more convincing (Fig. 4B, right). Indeed, while the control sample did not show any characteristic signal, the CuAAC sample possessed a readily detectable sulfur content and exhibited a clear peak related to the S 2p orbital. This comparison alone evidences the success of the reaction and discards the occurrence of physisorption.

While proving that a reaction occurs at the surface is an important step, it is equally important to verify the preservation of the surface nanostructure. If the film is stable, the morphology of the film should not evolve. Considering the low amount of functional groups present in the polymer film – hence at the surface – and the relatively low molar mass of the grafted polymer chains, no change of topology is expected either. The AFM phase image of the CuAAC sample (Fig. 6B)

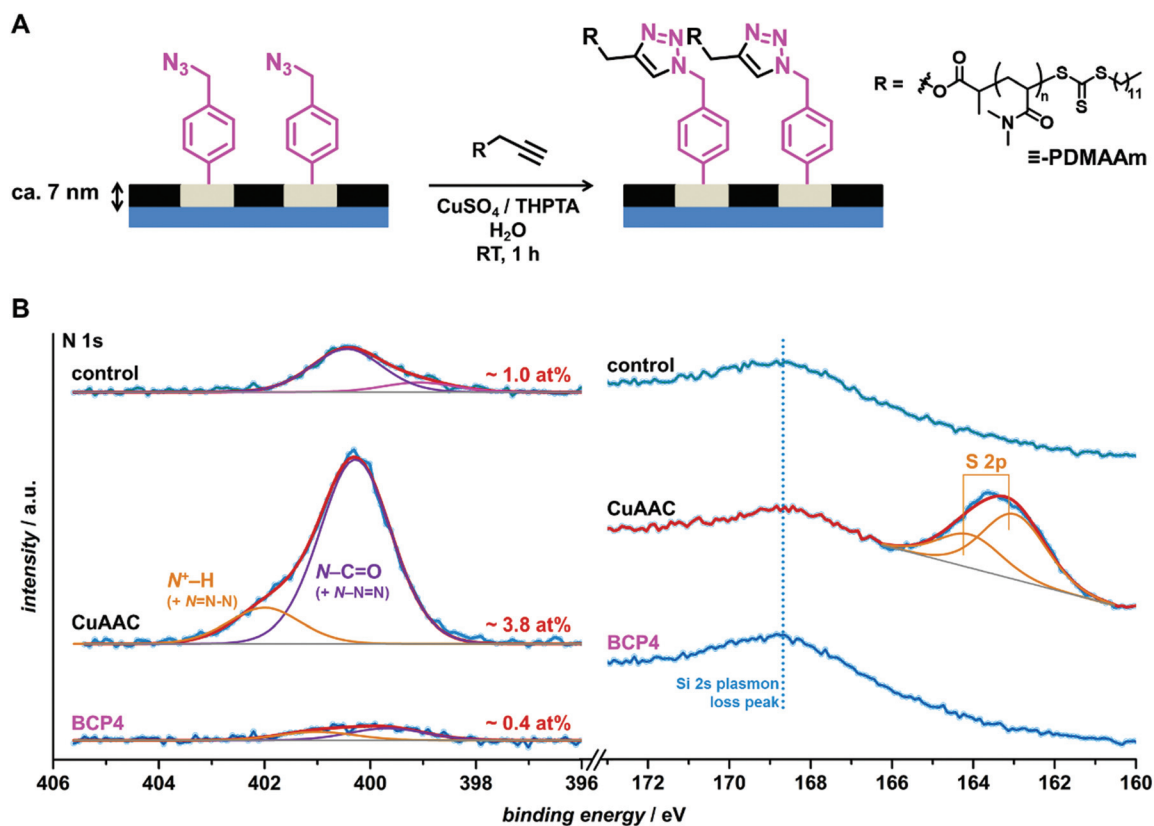


Fig. 5 (A) Schematic representation of the grafting of $\equiv\text{-PDMAAm}$ on an ultra-thin (ca. 7 nm) **BCP4** film via copper-catalyzed azide-alkyne coupling. (B) N 1s (left) and S 2p (right) selected regions of XPS spectra of an as-coated **BCP4** thin film (bottom) and after incubation with $\equiv\text{-PDMAAm}$ in the presence (middle, CuAAC) or in the absence (top, control) of the copper-based catalytic system. Note that the amide signals may overlap with triazole-based peaks.⁵³



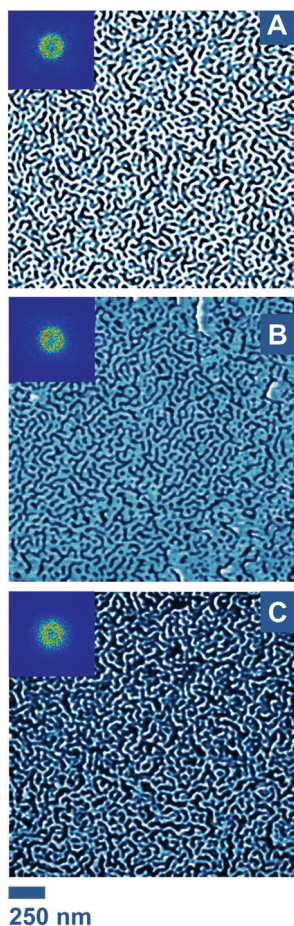


Fig. 6 AFM phase images of an ultra-thin (ca. 7 nm) BCP4 film (A) before reaction and after incubation (B) with \equiv -PDMAAm in the presence (CuAAC sample) or (C) in the absence of the copper-based catalytic system (control sample). Insets are fast Fourier transform (FFT) images.

displayed features very similar to those of the original BCP4 film (Fig. 6A). The morphology of the control sample was also overall preserved, as expected (Fig. 6C), yet the AFM data exhibited slightly different contrast. Besides AFM phase images, height images were also analyzed. In this case, further slight changes were observed between the original BCP4 film and the CuAAC sample (Fig. S8A and B[†]). We assign these alterations to a general slightly poor stability of the film upon incubation and through numerous washings in/with water because the control sample presented very similar alterations (Fig. S8C[†]). For future developments, a stabilization protocol would therefore be required. For instance, the presence of azide moieties allow for photocrosslinking (see Fig. S9[†]) and should be further exploited to obtain reactive films stable not only in aqueous conditions but also in organic media.

Conclusions

In the current contribution, we introduce a new system for molecular surface ordering at the nanoscale by employing

lamellae-forming block copolymer thin films as pre-organized docking sites. A series of functional PS-*b*-PI block copolymers possessing alkene, pentafluorophenyl, chloromethyl, or azide groups were synthesized by nitroxide-mediated polymerization and employed to fabricate surface-reactive nanostructured films with sub-30 nm features. Thermal characterization demonstrated the phase segregating behavior of the block copolymers, while SAXS measurements evidenced the lamellar nature of the two phases corresponding to PS and PI domains, respectively. Observation of the surface morphology by AFM revealed the near-identical features of all block copolymer films. It is shown that no particular process optimization is needed to obtain controlled morphology, despite the presence of additional functionalities. The key element is to keep the amount of the latter to a minimum. In order to illustrate the concept, an exemplary reaction proved the presence of azide groups at the surface by grafting of a model alkyne-functionalized polymer. Nevertheless, the films were shown to exhibit limited stability, even in aqueous medium, therefore requiring a stabilization protocol such as photocrosslinking. Further work is underway to optimize this crosslinking protocol in order to leave free reactive groups for surface reaction. Additional chemical routes for functionalization, *i.e.*, ene reactions on PI and nucleophilic substitutions on PS, are also being evaluated. We believe that employing reactive block copolymer films is a very promising alternative for high-throughput and large-area chemical patterning, because most applications do not require more than two components to be immobilized. Furthermore, progress is continuously witnessed in the area of block copolymer thin film technology. Notably, the combination with directed self-assembly methods may be beneficial and will also be explored in the future.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- B. Basnar and I. Willner, *Small*, 2009, **5**, 28–44.
- C. M. Kolodziej and H. D. Maynard, *Chem. Mater.*, 2012, **24**, 774–780.
- S. A. Alang Ahmad, L. S. Wong, E. ul-Haq, J. K. Hobbs, G. J. Leggett and J. Micklefield, *J. Am. Chem. Soc.*, 2011, **133**, 2749–2759.
- M. Montague, R. E. Ducker, K. S. L. Chong, R. J. Manning, F. J. M. Rutten, M. C. Davies and G. J. Leggett, *Langmuir*, 2007, **23**, 7328–7337.
- H.-W. Li, B. V. O. Muir, G. Fichet and W. T. S. Huck, *Langmuir*, 2003, **19**, 1963–1965.
- C. Park, J. Yoon and E. L. Thomas, *Polymer*, 2003, **44**, 6725–6760.
- H.-C. Kim, S.-M. Park and W. D. Hinsberg, *Chem. Rev.*, 2010, **110**, 146–177.
- C. T. Black, R. Ruiz, G. Breyta, J. Y. Cheng, M. E. Colburn, K. W. Guarini, H. C. Kim and Y. Zhang, *IBM J. Res. Dev.*, 2007, **51**, 605–633.
- M. Luo and T. H. Epps, *Macromolecules*, 2013, **46**, 7567–7579.
- F. S. Bates and G. H. Fredrickson, *Phys. Today*, 1999, **52**, 32–38.
- T. P. Lodge, *Macromol. Chem. Phys.*, 2003, **204**, 265–273.
- A.-V. Ruzette and L. Leibler, *Nat. Mater.*, 2005, **4**, 19–31.
- J. Bang, U. Jeong, D. Y. Ryu, T. P. Russell and C. J. Hawker, *Adv. Mater.*, 2009, **21**, 4769–4792.
- X. Jin, Y. Pang and S. Ji, *Chin. J. Polym. Sci.*, 2016, **34**, 659–678.
- R. A. Segalman, *Mater. Sci. Eng., R*, 2005, **48**, 191–226.
- A. P. Lane, M. J. Maher, C. G. Willson and C. J. Ellison, *ACS Macro Lett.*, 2016, **5**, 460–465.
- C. M. Bates, M. J. Maher, D. W. Janes, C. J. Ellison and C. G. Willson, *Macromolecules*, 2014, **47**, 2–12.
- C. Sinturel, F. S. Bates and M. A. Hillmyer, *ACS Macro Lett.*, 2015, **4**, 1044–1050.
- J. D. Cushen, C. M. Bates, E. L. Rausch, L. M. Dean, S. X. Zhou, C. G. Willson and C. J. Ellison, *Macromolecules*, 2012, **45**, 8722–8728.
- S. Park, D. H. Lee, J. Xu, B. Kim, S. W. Hong, U. Jeong, T. Xu and T. P. Russell, *Science*, 2009, **323**, 1030–1033.
- D. Borah, M. T. Shaw, J. D. Holmes and M. A. Morris, *ACS Appl. Mater. Interfaces*, 2013, **5**, 2004–2012.
- J. D. Cushen, I. Otsuka, C. M. Bates, S. Halila, S. Fort, C. Rochas, J. A. Easley, E. L. Rausch, A. Thio, R. Borsali, C. G. Willson and C. J. Ellison, *ACS Nano*, 2012, **6**, 3424–3433.
- H. S. Suh, D. H. Kim, P. Moni, S. Xiong, L. E. Ocola, N. J. Zaluzec, K. K. Gleason and P. F. Nealey, *Nat. Nanotechnol.*, 2017, **12**, 575.
- J. Kwak, A. K. Mishra, J. Lee, K. S. Lee, C. Choi, S. Maiti, M. Kim and J. K. Kim, *Macromolecules*, 2017, **50**, 6813–6818.
- J. G. Kennemur, L. Yao, F. S. Bates and M. A. Hillmyer, *Macromolecules*, 2014, **47**, 1411–1418.
- Y. Sakai-Otsuka, S. Zaioncz, I. Otsuka, S. Halila, P. Rannou and R. Borsali, *Macromolecules*, 2017, **50**, 3365–3376.
- G. Jeong, D. M. Yu, J. K. D. Mapas, Z. Sun, J. Rzaev and T. P. Russell, *Macromolecules*, 2017, **50**, 7148–7154.
- S. Schoffelen and J. C. M. van Hest, *Soft Matter*, 2012, **8**, 1736–1746.
- N. Kumar and J. Hahn, *Langmuir*, 2005, **21**, 6652–6655.
- D. Liu, T. Wang and J. L. Keddie, *Langmuir*, 2009, **25**, 4526–4534.
- S. Watanabe, R. Fujiwara, M. Hada, Y. Okazaki and T. Iyoda, *Angew. Chem., Int. Ed.*, 2007, **46**, 1120–1123.
- J. Chai and J. M. Buriak, *ACS Nano*, 2008, **2**, 489–501.
- I. C. Reinhout, G. Delaittre, H.-C. Kim, R. J. M. Nolte and J. J. L. M. Cornelissen, *J. Mater. Chem. B*, 2013, **1**, 3026–3030.
- W. A. Lopes and H. M. Jaeger, *Nature*, 2001, **414**, 735–738.
- H. Schönherr, C. L. Feng, N. Tomczak and G. J. Vancso, *Macromol. Symp.*, 2005, **230**, 149–157.
- C. Minelli, I. Geissbuehler, C. Hinderling, H. Heinzelmann, H. Vogel, R. Pugin and M. Liley, *J. Nanosci. Nanotechnol.*, 2006, **6**, 1611–1619.
- H. Tran, K. Ronaldson, N. A. Bailey, N. A. Lynd, K. L. Killips, G. Vunjak-Novakovic and L. M. Campos, *ACS Nano*, 2014, **8**, 11846–11853.
- F. W. Speetjens, M. C. D. Carter, M. Kim, P. Gopalan, M. K. Mahanthappa and D. M. Lynn, *ACS Macro Lett.*, 2014, **3**, 1178–1182.
- J. G. Kennemur, M. A. Hillmyer and F. S. Bates, *Macromolecules*, 2012, **45**, 7228–7236.
- E. W. Cochran and F. S. Bates, *Macromolecules*, 2002, **35**, 7368–7374.
- J. Stadermann, M. Riedel, H. Komber, F. Simon and B. Voit, *J. Polym. Sci., Part A: Polym. Chem.*, 2012, **50**, 1351–1361.
- J. Stadermann, M. Riedel and B. Voit, *Macromol. Chem. Phys.*, 2013, **214**, 263–271.
- S. Chae, J.-H. Kim, P. Theato, R. Zentel and B.-H. Sohn, *Macromol. Chem. Phys.*, 2014, **215**, 654–661.
- T. Hosoda, S. P. Gido, J. W. Mays, T. Huang, C. R. Park and T. Yamada, *J. Polym. Eng.*, 2013, **33**, 49–59.
- K. W. Oleske, K. P. Barteau, M. Z. Turker, P. A. Beaucage, L. A. Estroff and U. Wiesner, *Macromolecules*, 2017, **50**, 542–549.
- H. Turgut, D. Varadharajan, N. Dingenouts and G. Delaittre, *Macromol. Rapid Commun.*, 2018, **39**, 1800231.
- S. Park, D. Y. Ryu, J. K. Kim, M. Ree and T. Chang, *Polymer*, 2008, **49**, 2170–2175.
- Y. Noda, T. Kumada, T. Hashimoto and S. Koizumi, *J. Appl. Crystallogr.*, 2011, **44**, 503–513.



- 49 S.-Y. Chen, Y. Huang and R. C.-C. Tsiang, *J. Polym. Sci., Part A: Polym. Chem.*, 2008, **46**, 1964–1973.
- 50 T. Higuchi, M. Shimomura and H. Yabu, *Macromolecules*, 2013, **46**, 4064–4068.
- 51 H. Yabu, S. Sato, T. Higuchi, H. Jinnai and M. Shimomura, *J. Mater. Chem.*, 2012, **22**, 7672–7675.
- 52 D. Liu, C. A. Che Abdullah, R. P. Sear and J. L. Keddie, *Soft Matter*, 2010, **6**, 5408–5416.
- 53 X. Li, J. Guo, Y. Liu and H. Liang, *J. Chem. Phys.*, 2009, **130**, 74908.
- 54 S. H. Kim, K.-S. Kim, K. Char, S. Il Yoo and B.-H. Sohn, *Nanoscale*, 2016, **8**, 10823–10831.
- 55 L. Hlalele and B. Klumperman, *Macromolecules*, 2011, **44**, 6683–6690.
- 56 S. Harrisson, P. Couvreur and J. Nicolas, *Polym. Chem.*, 2011, **2**, 1859–1865.
- 57 V. Hong, S. I. Presolski, C. Ma and M. G. Finn, *Angew. Chem., Int. Ed.*, 2009, **48**, 9879–9883.
- 58 C. J. Durr, S. G. J. Emmerling, P. Lederhose, A. Kaiser, S. Brandau, M. Klimpel and C. Barner-Kowollik, *Polym. Chem.*, 2012, **3**, 1048–1060.
- 59 C. Strazielle, H. Benoit and O. Vogl, *Eur. Polym. J.*, 1978, **14**, 331–334.
- 60 S. Beuermann, D. A. Paquet, J. H. McMinn and R. A. Hutchinson, *Macromolecules*, 1996, **29**, 4206–4215.
- 61 K. L. Parry, A. G. Shard, R. D. Short, R. G. White, J. D. Whittle and A. Wright, *Surf. Interface Anal.*, 2006, **38**, 1497–1504.
- 62 J. H. Scofield, *J. Electron Spectrosc. Relat. Phenom.*, 1976, **8**, 129–137.
- 63 J. Nicolas, Y. Guillauneuf, C. Lefay, D. Bertin, D. Gigmes and B. Charleux, *Prog. Polym. Sci.*, 2013, **38**, 63–235.
- 64 S. Harrisson, P. Couvreur and J. Nicolas, *Macromolecules*, 2011, **44**, 9230–9238.
- 65 S. Harrisson, P. Couvreur and J. Nicolas, *Macromol. Rapid Commun.*, 2012, **33**, 805–810.
- 66 D. Varadharajan, H. Turgut, J. Lahann, H. Yabu and G. Delaittre, *Adv. Funct. Mater.*, 2018, **28**, 1800846.
- 67 U. Jeong, D. Y. Ryu, D. H. Kho, J. K. Kim, J. T. Goldbach, D. H. Kim and T. P. Russell, *Adv. Mater.*, 2004, **16**, 533–536.
- 68 E. L. Malins, S. Amabilino, G. Yilmaz, F. H. Isikgor, B. M. Gridley and C. R. Becer, *Eur. Polym. J.*, 2015, **62**, 347–351.
- 69 H. Turgut and G. Delaittre, *Chem. – Eur. J.*, 2016, **22**, 1511–1521.
- 70 D. Varadharajan and G. Delaittre, *Polym. Chem.*, 2016, **7**, 7488–7499.
- 71 R. Breslow, M. Brandl, J. Hunger, N. Turro, K. Cassidy, K. Krogh-Jespersen and J. D. Westbrook, *J. Am. Chem. Soc.*, 1987, **109**, 7204–7206.
- 72 J.-P. O'Shea, V. Solovyeva, X. Guo, J. Zhao, N. Hadjichristidis and V. O. Rodionov, *Polym. Chem.*, 2014, **5**, 698–701.
- 73 W. Jakubowski, A. Juhari, A. Best, K. Koynov, T. Pakula and K. Matyjaszewski, *Polymer*, 2008, **49**, 1567–1578.
- 74 J. Brandrup, E. H. Immergut and E. A. Grulke, *Polymer Handbook*, John Wiley & Sons, Chichester, 4th edn, 1999.
- 75 M. Dionisio, L. Ricci, G. Pecchini, D. Masseroni, G. Ruggeri, L. Cristofolini, E. Rampazzo and E. Dalcanale, *Macromolecules*, 2014, **47**, 632–638.
- 76 C. B. Roth and J. R. Dutcher, *Eur. Phys. J. E*, 2003, **12**, 103–107.
- 77 G. B. Kharas, E. Hanawa, A. B. Agpasa, T. O. Fedczyna, B. M. Flaherty, J. L. Fernandes, M. Liosatos, M. E. Lukowski, K. G. Skinner and R. N. Theiler, *Polym. Bull.*, 2013, **70**, 707–714.
- 78 V. Thirtha, R. Lehman and T. Nosker, *Polym. Eng. Sci.*, 2005, **45**, 1187–1193.
- 79 J. Liu, Q. Deng and Y. C. Jean, *Macromolecules*, 1993, **26**, 7149–7155.
- 80 C. Kow, M. Morton, L. J. Fetters and N. Hadjichristidis, *Rubber Chem. Technol.*, 1982, **55**, 245–252.
- 81 I. Hamley, *The Physics of Block Copolymers*, Oxford University Press, 1998.
- 82 We observed a pronounced peak broadening caused by the finite beam dimensions combined with the short distance between sample and detector, inherent to laboratory SAXS equipment. Therefore, it is normally difficult to identify the exact morphology because the peak broadening may hide peaks at non-integer multiples of q indicating other structures than lamellae. In this case, we are of the opinion that it is possible to conclude that we have a lamellar structure in spite of these problems. The presence of a third-order scattering peaks at three times q indicates a rather high degree of ordering, this could be shown by folding of theoretical scattering curves with our experimental beam profile. With a higher polydispersity in layer distance, the peak broadening would suppress all higher order peaks. This does not imply a long-range order, but a rather low deviation in layer distance and a grain size of at least 10 layers. Due to this achieved degree of ordering, we are able to see that the second-order peak at two times q is nearly suppressed. Even with the peak broadening in mind, one can conclude that the second order is clearly less pronounced than the third order.
- 83 T. Hashimoto, K. Nagatoshi, A. Todo, H. Hasegawa and H. Kawai, *Macromolecules*, 1974, **7**, 364–373.
- 84 H. Hasegawa, T. Hashimoto, H. Kawai, T. P. Lodge, E. J. Amis, C. J. Glinka and C. C. Han, *Macromolecules*, 1985, **18**, 67–78.
- 85 Y. Matsushita, K. Mori, R. Saguchi, Y. Nakao, I. Noda and M. Nagasawa, *Macromolecules*, 1990, **23**, 4313–4316.
- 86 F. S. Bates, *Science*, 1991, **251**, 898–905.
- 87 M. W. Matsen and F. S. Bates, *J. Polym. Sci., Part B: Polym. Phys.*, 1997, **35**, 945–952.
- 88 N. A. Lynd and M. A. Hillmyer, *Macromolecules*, 2005, **38**, 8803–8810.
- 89 No precise data could be found in the literature for the respective polarity of halides and azides. Yet, experimental observation from colleagues reveals that the transformation of alkyl bromides into alkyl azides essentially do not



lead to any major change of retention factor in thin-layer chromatography. Chlorine being more electronegative than bromine, one could infer that alkyl chloride are – at least slightly – more polar than the corresponding alkyl azides.

- 90 C. Harrison, M. Park, P. M. Chaikin, R. A. Register, D. H. Adamson and N. Yao, *Polymer*, 1998, **39**, 2733–2744.
- 91 C. Harrison, M. Park, P. M. Chaikin, R. A. Register and D. H. Adamson, *J. Vac. Sci. Technol., B: Microelectron. Nanometer Struct.–Process., Meas., Phenom.*, 1998, **16**, 544–552.
- 92 M. Park, C. Harrison, P. M. Chaikin, R. A. Register and D. H. Adamson, *Science*, 1997, **276**, 1401–1404.
- 93 T. A. Mykhaylyk, O. O. Mykhaylyk, S. Collins and I. W. Hamley, *Macromolecules*, 2004, **37**, 3369–3377.
- 94 C. Lei, K. Ouzineb, O. Dupont and J. L. Keddie, *J. Colloid Interface Sci.*, 2007, **307**, 56–63.
- 95 M. J. Fasolka, D. J. Harris, A. M. Mayes, M. Yoon and S. G. J. Mochrie, *Phys. Rev. Lett.*, 1997, **79**, 3018–3021.
- 96 H.-C. Kim and W. D. Hinsberg, *J. Vac. Sci. Technol., A*, 2008, **26**, 1369–1382.
- 97 M. W. Matsen, *J. Chem. Phys.*, 1997, **106**, 7781–7791.
- 98 M. J. Fasolka, P. Banerjee, A. M. Mayes, G. Pickett and A. C. Balazs, *Macromolecules*, 2000, **33**, 5702–5712.
- 99 K. Motoyoshi, A. Tajima, T. Higuchi, H. Yabu and M. Shimomura, *Soft Matter*, 2010, **6**, 1253–1257.
- 100 A. Lucena-Serrano, C. Lucena-Serrano, R. Contreras-Cáceres, A. Díaz, M. Valpuesta, C. Cai and J. M. López-Romero, *Appl. Surf. Sci.*, 2016, **360**(Part), 419–428.
- 101 N. K. Devaraj and J. P. Collman, *QSAR Comb. Sci.*, 2007, **26**, 1253–1260.
- 102 R. Zirbs, F. Kienberger, P. Hinterdorfer and W. H. Binder, *Langmuir*, 2005, **21**, 8414–8421.
- 103 L. Li, X.-X. Deng, Z.-L. Li, F.-S. Du and Z.-C. Li, *Macromolecules*, 2014, **47**, 4660–4667.
- 104 L. Gonzalez-Macia, S. Griveau, F. d'Orlyé, A. Varenne, C. Sella, L. Thouin and F. Bedioui, *Electrochem. Commun.*, 2016, **70**, 78–81.
- 105 G. A. Hudalla and W. L. Murphy, *Langmuir*, 2009, **25**, 5737–5746.
- 106 N. Zydziak, C. Hubner, M. Bruns, A. P. Vogt and C. Barner-Kowollik, *Polym. Chem.*, 2013, **4**, 1525–1537.
- 107 A. Belouqui, S. Baur, V. Trouillet, A. Welle, J. Madsen, M. Bastmeyer and G. Delaitte, *Small*, 2016, **12**, 1716–1722.

