



Cite this: *Polym. Chem.*, 2019, **10**, 1186

Received 5th October 2018,
Accepted 23rd January 2019

DOI: 10.1039/c8py01437j

rsc.li/polymers

Microscale synthesis of multiblock copolymers using ultrafast RAFT polymerisation†

Joji Tanaka, *^a Pratik Gurnani, ^a Alexander B. Cook, ^a Satu Häkkinen, ^a Junliang Zhang, ^a Jie Yang,^a Andrew Kerr,^a David M. Haddleton, ^a Sébastien Perrier ^{a,b} and Paul Wilson ^{a,b}

We demonstrate that ultrafast RAFT in the presence of air can be scaled down to 2 μL with good control using microvolume insert vials as the polymerisation vessel. By careful cooling and mixing of the sequential monomers, well-defined pentablock copolymers were generated with a final volume of 10 μL .

High-throughput screening is becoming increasingly prevalent industrially and academically in identifying pharmaceutical targets and in optimising synthetic routes.¹ Consequently, scaling down reactions is of paramount importance for exploring enormous numbers of possible permutations of parameters involved in a chemical synthesis.² Polymerisation in standard chemistry laboratory reaction vessels becomes increasingly difficult at smaller scales. The transfer of advanced polymer synthesis techniques to smaller scales will allow for the high-throughput screening of polymer compositions for biomaterial discovery, and has been previously exploited in step-growth polymerisations.^{3,4} However, small scale screening using controlled polymerisation methods was only recently achieved by Boyer *et al.* in the investigation of the influence of polymer architecture on material properties.^{5,6}

Polymerisations are typically carried out with reaction volumes between 50 mL and 0.5 mL.^{7–15} These ranges are practical for the conventional reaction vessels and deoxygenation processes necessary for typical Reversible Deactivation Radical Polymerisation (RDRP). Note that the latter condition limits scales of the reactions, because using nitrogen sparging or freeze–pump–thaw cycles to deoxygenate the reaction media is not practical at ultralow volumes, due to the inherent loss of volatile monomers and solvents. Hence, oxygen tolerant RDRP protocols are necessary to allow polymerisations to be carried

out at the microscale. To this end, Boyer *et al.* have performed ultralow volume reactions (20 μL) in 96 well plates, using photo-catalysed redox Reversible Addition–Fragmentation Chain Transfer (RAFT) polymerisation without deoxygenation in the presence of air.¹⁶ This enabled screening of different homopolymers, diblock copolymers, star architectures and nanoparticle formulations.

RDRP protocols without deoxygenation have become an emerging topic;^{17,18} however, many of these protocols require external stimuli,¹⁹ additives¹⁶ or oxygen scavenging enzymes,^{5,20,21} resulting in the deviation from the simplicity of RDRP protocols. To address this, Gody *et al.* demonstrated standard RAFT polymerisation using only conventional ingredients without deoxygenation, in vessels open to air.²² This elegant and simple approach takes advantage of the fast propagation of acrylamidic monomers in water, a solvent known to increase the rate of radical polymerisation, which is further accelerated at elevated temperatures. This ultrafast RAFT polymerisation was generally demonstrated with acrylamide-based monomers with 2,2'-azobis[2-(2-imidazolin-2-yl)propane]dihydrochloride (VA-044) as the initiator (10 h $t_{1/2}$ = 44 °C) and at 100 °C. This allowed the synthesis of multiblock copolymers (MBCPs) through iterative chain extensions where full monomer conversion was achieved within 3 minutes per block, before the initiator was fully decomposed (approximately 80%).²²

MBCPs are macromolecules with defined control over the block sequence that can be synthesised from just simple chemical ingredients without complex biological machineries, and are amenable for industrial scales.^{23–25} The synthesis of MBCPs has progressed more recently with RDRP, using copper-mediated polymerisation^{26–34} and RAFT polymerisation.^{35–41} In spite of the inevitable small number of radical termination events, 21 iterative block extensions have been reliably demonstrated with RDRP.⁴² Furthermore, these routes are popular as they allow the incorporation of monomers of various functional groups^{43–48} and do not require immaculately dry reagents and environments, necessary for

^aDepartment of Chemistry, University of Warwick, Gibbet Hill Road, CV4 7AL Coventry, UK. E-mail: j.tanaka@warwick.ac.uk

^bMonash Institute of Pharmaceutical Sciences, Monash University (Parkville Campus), 399 Royal Parade, Parkville, Victoria 3152, Australia

†Electronic supplementary information (ESI) available. See DOI: 10.1039/c8py01437j



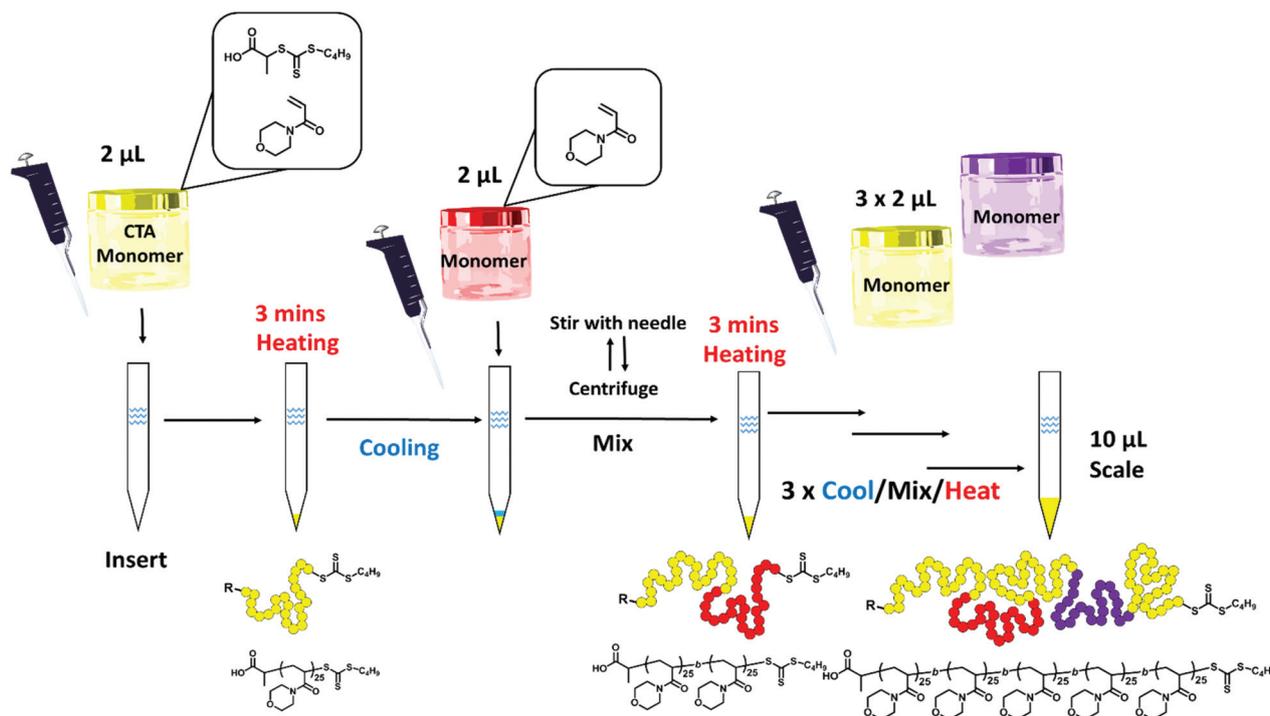
ionic living polymerisation systems.⁴⁹ Recently, work on sulfur-free RAFT polymerisation has offered the potential for MBCP synthesis amenable for industrial scales.^{42,50,51} However, the possible benefits of scaling down MBCP synthesis are often overlooked in academic settings. Industries often rely on the inexpensive small scale combinatorial reactions for optimisation before larger scale synthesis. Microscale MBCP synthesis could have applicability in high-throughput microarrays,² thus allowing the rapid investigation of permutations of monomers and block lengths of MBCPs.

We postulate that the aforementioned ultrafast RAFT protocol could be applicable in the microliter scale due to the rapid consumption of the monomer without deoxygenation; thus demonstrating the synthesis of MBCPs at a microscale suitable for potential applications such as microarray patterning and combinatorial chemistry with only conventional ingredients used for RAFT polymerisation.

To counteract the inherent problem of the increased air/water interface when scaling down the protocol proposed by Gody *et al.*, we used narrow micro-volume glass inserts (4.6 mm diameter, 200 μL capacity) with conical bottoms (cone volume = approximately 20 μL) that are typically fitted into standard 2 mL vials for low volume HPLC/GPC analysis.⁵² A master mix of the RAFT agent, monomer, solvent and initiator was made as an “all-in-one” stock solution and added into the insert using a standard micropipette (Scheme 1). This mix was

made following the published protocol,²² using VA-044 as the initiator (3×10^{-3} M, $[\text{CTA}]/[\text{I}]_0 = 40$), *N*-acryloylmorpholine (NAM) as a suitable acrylamidic monomer ($[\text{M}]_0 = 3$ M) in an aqueous mixture and 2-(((butylthio)-carbonothioyl)thio)propanoic acid (PABTC) as the RAFT agent. The inserts were then heated in an oil bath at 100 $^{\circ}\text{C}$ for 3 minutes. In contrast to the previous study where the increase in reaction temperature was gradual, taking 80 seconds to reach 96 $^{\circ}\text{C}$,²² we assume the temperature of the reaction to reach equilibrium almost immediately. Conveniently, as the polymerisations were carried out in SEC insert vials, the reaction mixture could be directly diluted with the SEC eluent within the insert, and injected directly for SEC analysis (Fig. S2†). A duplicate reaction was carried out to dilute with NMR solvent (DMSO-d_6) to measure the monomer conversion by NMR.

Preliminary experiments were designed to investigate the absolute limit of scale for the polymerisation. Initially this was investigated with a targeted degree of polymerisation (DP) of 25 using 20 vol% dioxane in water to aid the solubility of the CTA. The experimental number-average molar mass ($M_{n,\text{SEC}}$) and dispersity (D) were measured using SEC analysis to investigate the control maintained at lower volumes. At 10 μL ($M_{n,\text{SEC}} = 2200$ g mol⁻¹; $D = 1.23$), 5 μL ($M_{n,\text{SEC}} = 2600$ g mol⁻¹; $D = 1.23$), and 2 μL ($M_{n,\text{SEC}} = 2600$ g mol⁻¹; $D = 1.29$), we were able to reproducibly obtain PNAM₂₅ as observed by SEC analysis (Fig. S1†) with only a slight increase in dispersity,



Scheme 1 General scheme: “master mix” (with a monomer, CTA, initiator and solvent) was added into the microvolume insert using a regular air displacement micropipette. After 3 min of heating at 100 $^{\circ}\text{C}$ in an oil bath, polymerisation was complete and the insert was cooled with liquid nitrogen. For sequential chain extension, a separate monomer master mix was directly added and mixed by stirring with a needle and centrifugation, before reheating for further 3 minutes for block extension. This cycle was repeated to yield a pentablock copolymer. All the polymerisations were carried out without deoxygenation and in the presence of open air.



nevertheless they maintained good control comparable to the polymerisation carried out in a 5.4 mL test tube (termed macroscale in this paper) ($M_{n,SEC} = 2700 \text{ g mol}^{-1}$; $D = 1.19$). At 1 μL scale polymers were obtained ($M_{n,SEC} = 2600 \text{ g mol}^{-1}$); however, the D increased to 1.42. The weight loss due to the evaporation of the reaction mixture was also noted (Table S1†), which seemingly increased at lower scales and may have been a contributing factor to the loss of control at the 1 μL scale. Hence we concluded that 2 μL is the lowest scale to maintain good control. Our next objective was to apply this protocol to longer polymer chain lengths of PNAM $_n$ (Fig. 1). Increasing the chain length fourfold (DP = 100) required a slight modification of the master mix (10 vol% dioxane in water; $[I]_0 = 1 \times 10^{-3} \text{ M}$, $[CTA]/[I]_0 = 30$). Pleasingly, polymerisation yielded PNAM $_{100}$ at the 2 μL scale ($M_{n,SEC} = 9200 \text{ g mol}^{-1}$; $D = 1.36$), and in contrast to the macroscale its molecular weight distribution was relatively broader as revealed by SEC analysis ($M_{n,SEC} = 8900 \text{ g mol}^{-1}$; $D = 1.19$). Increasing the length further (DP = 200), increased the dispersity at the 2 μL scale (PNAM $_{200}$, $M_{n,SEC} = 26800 \text{ g mol}^{-1}$; $D = 1.43$), compared to the macroscale equivalent ($M_{n,SEC} = 15000 \text{ g mol}^{-1}$; $D = 1.23$). SEC analysis in all cases revealed a slightly higher dispersity due to the appearance of low molecular tailing. Also the ^1H NMR spectra revealed that more residual monomer was present (approximately 2–3% more). As the targeting DP of 25 of NAM yielded a relatively narrow dispersity at the 2 μL scale, we therefore decided to keep this a constant block length for our MBCPs. In order to generate MBCPs through iterative chain extension with the current protocol, it was important to consider the limitation of mixing sequential monomers in the polymerisation mixture, as stirring during polymerisation is unfeasible at the 2 μL scale. To maximise the mixing of each monomer aliquot the polymerisation reaction mixture was cooled prior to the addition of a new monomer and stirred before heating again at 100 $^\circ\text{C}$ for a successful sequential chain extension. This circumvented the need for continual stirring during the addition of sequential monomers. Thus by adopting this

necessary measure of cooling and mixing before reheating (Scheme 1), we were able to successfully demonstrate successive chain extensions within the insert vials to synthesize a homopolymer in five successive chain extensions, P(NAM $_{25}$) $_5$, at 5 μL per block (final $M_{n,SEC} = 10600 \text{ g mol}^{-1}$; $D = 1.25$) and 2 μL per block (final $M_{n,SEC} = 10000 \text{ g mol}^{-1}$; $D = 1.35$) (Fig. 2). Centrifugation was necessary to collect the new monomer solution at the bottom of the insert before stirring (see the ESI† for full details). It is important to note that the monomer concentration of the chain extension stock solution was kept constant at 3 M, such that the same DP per chain extension could be targeted by sequentially adding the same volume as the original block. It is noteworthy that all the chain extension stock solutions had contained the same initiator concentration of $2.2 \times 10^{-3} \text{ M}$. This was designed to give a constant overall macroCTA/initiator ratio of 40 per block, whilst assuming that 20% of the initiator is still remaining from the previous block. This gave a good balance of quantitative monomer consumption (>96%, Table 1) at each block whilst keeping the theoretical livingness of each block high (>98%), thus minimising the number of dead chains being formed (see the ESI for calculation and S6–S10† for detailed experimental conditions).

The monomer consumption was followed by ^1H NMR spectroscopy and the succession of the sequential chain extension was confirmed by GPC analysis of polymerisation at each block. To analyse each block extension, the same number of replicate reactions as the number of iterative blocks was prepared, whereby the representative vessel at each stage was used as a whole for each analysis (see the ESI† for the detailed procedure). In all cases, a linear increase in $M_{n,SEC}$ was observed with the increasing number of iterative block extensions (Fig. 2), suggesting excellent control in polymerisation at the microscale. In comparison the molecular weight distributions of P(NAM $_{25}$) $_5$ were only slightly broader at the microscale compared to the macroscale with our protocol (final $M_{n,SEC} = 12600 \text{ g mol}^{-1}$; $D = 1.23$). At the macroscale, bimodal distri-

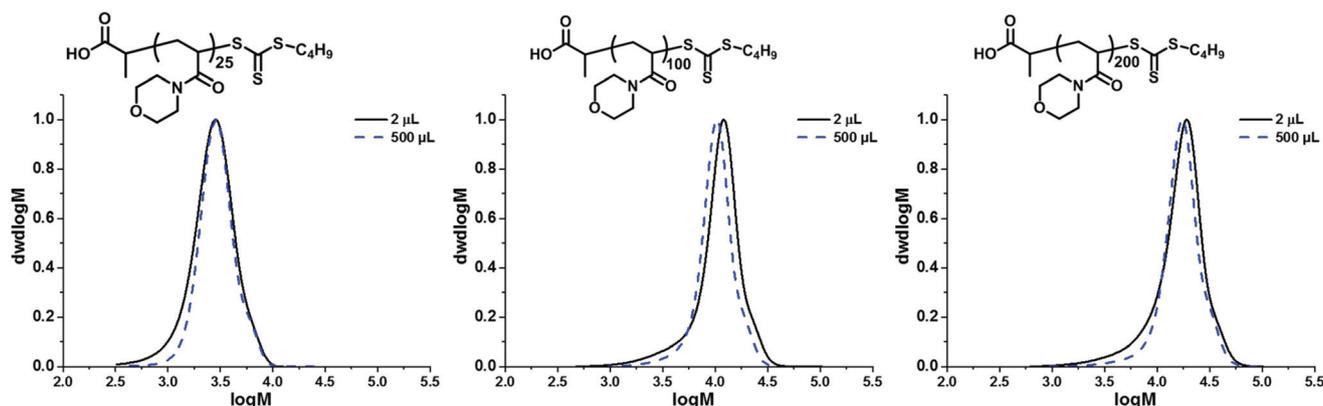


Fig. 1 SEC analysis (dRI, THF) of PNAM $_n$ ($n = 25, 100$ and 200) prepared at the microscale (2 μL) in microvolume inserts and at the normal scale (500 μL) in conventional test tubes (5.4 mL). All the polymerisations were carried out in 3 minutes open to air without stirring and deoxygenation. PMMA was used as the standard for SEC analysis.



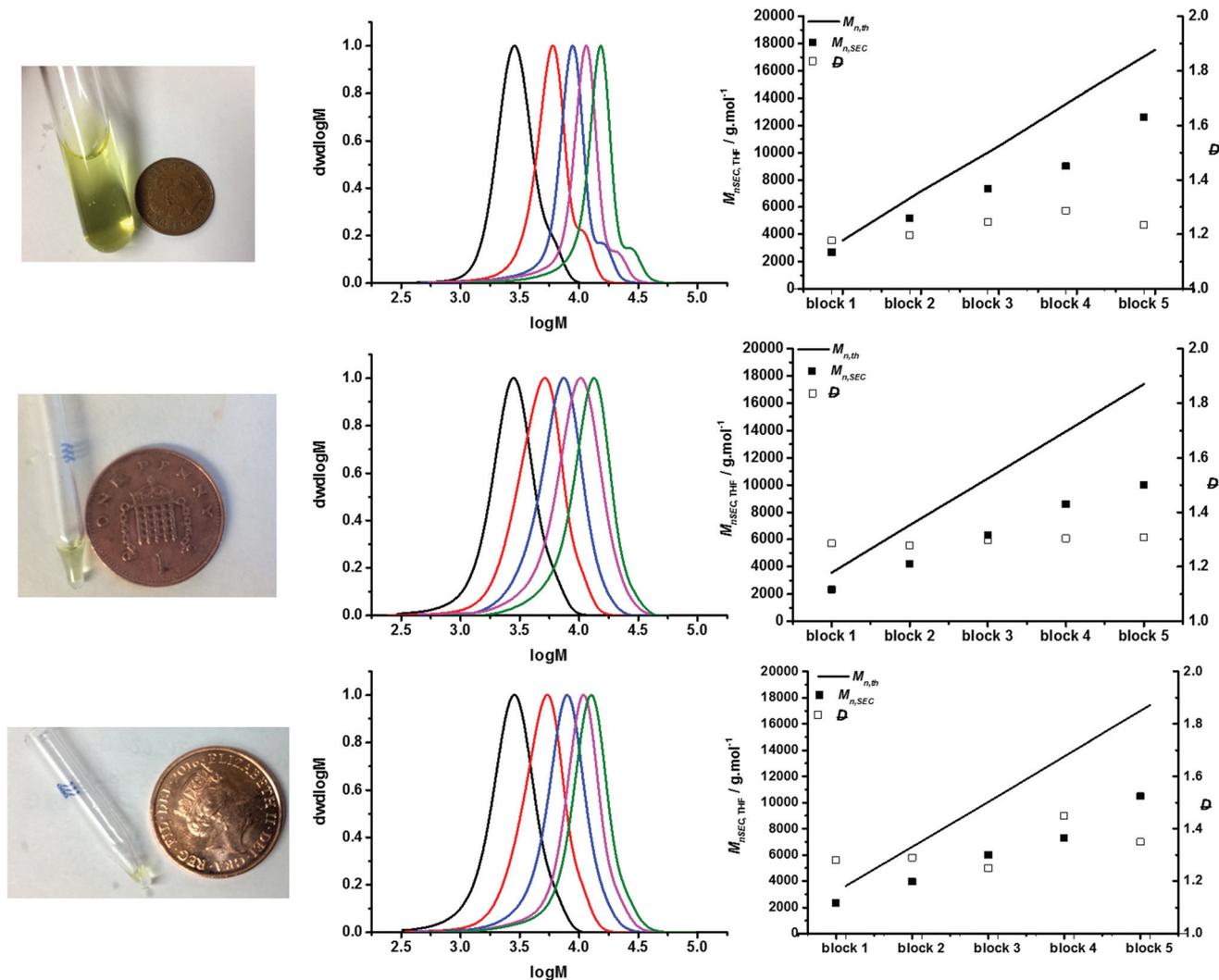


Fig. 2 Multichain extension to generate $P(\text{NAM}_{25})_5$ with ultrafast RAFT polymerisation at different scales: Top row = 2.5 ml, 0.5 ml per block; middle row = 25 μL , 5 μL per block; bottom row = 10 μL , 2 μL per block. Left column: Photograph after the reaction next to a British penny coin (20.3 mm in diameter) as a reference to the size of the scale. Middle column: SEC chromatograms for successive chain extensions. Right column: Evolution of number-average molar masses and dispersity values with the succession of chain extensions during the preparation of $P(\text{NAM}_{25})_5$. The black line represents the theoretical molar mass calculated using eqn (2) (see the ESI †). The filled squares represent the experimental molar masses and empty squares represent the dispersity values, both determined by SEC in THF with PMMA standards.

butions were observed, due to backbiting induced β -scission and subsequent branching, with successive chain extensions.⁵³ Although this is typically a characteristic of more labile methine backbone hydrogens of acrylic monomer families, this was observed here with an acrylamide monomer as a result of high temperature. This was indeed the case in previous work manifested as a high molecular skew.²² We suspect that at the microscale (25 μL and 10 μL), this feature is still present despite molecular weight distributions appearing to be unimodal, due to the broadening of the molecular weight distribution. We attribute this as a result of the increased interface between the air and solution phase when scaling down, leading to an increased viscosity and uneven reaction mixture within the reaction vessel from evaporation as well as increased oxygen related termination events being more preva-

lent. It is noteworthy that the relative weight loss for each chain extension was considerably greater at the microscale compared to the conventional scale (Table S2 †).

To further demonstrate the robustness of this method, MBCPs were synthesized with blocks of different monomers: *N,N*-dimethylacrylamide (DMA) and *N*-hydroxyethylacrylamide (HEAm). Pentablocks of $\text{PNAM}_{2.5}\text{-}b\text{-PDMA}_{2.5}\text{-}b\text{-PNAM}_{2.5}\text{-}b\text{-PHEAm}_{2.5}\text{-}b\text{-PNAM}_{2.5}$ were prepared in the inserts at the micro-liter scale (2 μL per block) following our protocol, with a well-defined molecular weight distribution ($M_{n,\text{SEC}} = 14\,000\text{ g mol}^{-1}$; $D = 1.35$) which was comparable to the macroscale synthesis ($M_{n,\text{SEC}} = 15\,100\text{ g mol}^{-1}$; $D = 1.32$). Note that switching the SEC eluent to DMF was necessary to aid the solubility of the HEAm incorporated pentablock. The resulting $M_{n,\text{SEC}}$ showed better agreement with the theoretical number-average



Table 1 The range of investigated reaction volumes and the corresponding monomer conversion, theoretical and experimental number-average molar masses and the dispersities of the synthesised homopolymers and multiblock copolymers

Polymer	Scale ^a (μL)	Conv. ^b %	$M_{n,th}$ ^c (g mol ⁻¹)	$M_{n,SEC}$ ^d (g mol ⁻¹)	D^d
PNAM ₂₅	500	>97	3600	2700	1.18
	10	>98	3600	2200	1.23
	5	>98	3600	2600	1.23
	2	>98	3600	2600	1.29
	1	>96	3600	2600	1.42
PNAM ₁₀₀	500	>99	14 000	8900	1.19
	2	>97	13 600	9200	1.36
PNAM ₂₀₀	500	>98	26 800	15 000	1.23
	2	>98	26 800	13 000	1.43
PNAM ₂₅ - <i>b</i> -PNAM ₂₅ - <i>b</i> -PNAM ₂₅ - <i>b</i> -PNAM ₂₅ - <i>b</i> -PNAM ₂₅	2500	>99.9	17 200	12 600	1.23
	25	>99	17 200	10 600	1.25
	10	>99	17 200	10 000	1.35
PNAM ₂₅ - <i>b</i> -PDMA ₂₅ - <i>b</i> -PNAM ₂₅ - <i>b</i> -PHEAm ₂₅ - <i>b</i> -PNAM ₂₅	2500	>96	15 400	15100 ^e	1.32 ^e
	10	>98	15 500	14000 ^e	1.35 ^e

^a All polymerisations were carried out in insert vials unless the scale is above or equal to 500 μL in which case they were carried out in a test tube (5.4 mL). ^b Determined by ¹H-NMR (DMSO-*d*₆) and calculated using eqn (1) in the ESI. ^c As calculated from eqn (2) in the ESI. ^d Determined by SEC in THF with PMMA standards, unless stated otherwise. ^e Determined by SEC in DMF with PMMA standards.

molar mass ($M_{n,th}$) owing to a better comparison with the PMMA calibrant in DMF (Fig. S4 and S5†).

Conclusions

To conclude, we have demonstrated the downscaling of the robust, oxygen-tolerant ultrafast RAFT polymerisation method, suitable for acrylamidic monomers in water in microvolume insert vials. Increased evaporation of the reaction mixture was observed with an increasing surface-to-volume ratio of the reaction mixture. This set a practical lower limit; however, we found that good control was still maintained at the 2 μL scale. The resulting polymer was successfully chain extended by sequential monomer additions. The reported method enables the synthesis of pentablock copolymers using only a final volume of 10 μL of the reaction mixture, thus demonstrating the high potential of ultrafast RAFT as a high-throughput screening method for MBCPs. Further studies are being carried out to investigate the robustness of the protocol in the synthesis of complex architectures and using different solvent systems, as well as its applicability to biological sciences.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

The authors gratefully acknowledge financial support from the Engineering and Physical Sciences Research Council (EPSRC) under grant EP/F500378/1 through the Molecular Organisation and Assembly in Cells Doctoral Training Centre (MOAC-DTC). S. H. acknowledges Lubrizol for financial support. A. B. C. acknowledges Syngenta for funding. The authors also

wish to acknowledge the facilities and personnel (S. P., P. W.) enabled by the Monash–Warwick Alliance. S. P. acknowledges a Royal Society Wolfson Merit Award (WM130055). P. W. thanks the Leverhulme Trust for the award of an Early Career Fellowship (ECF/2015-075). The authors would like to thank Dr Daniel Lester and the Polymer Characterisation Research Technology Platform for GPC facilities.

Notes and references

- R. Macarron, M. N. Banks, D. Bojanic, D. J. Burns, D. A. Cirovic, T. Garyantes, D. V. S. Green, R. P. Hertzberg, W. P. Janzen, J. W. Paslay, U. Schopfer and G. S. Sittampalam, *Nat. Rev. Drug Discovery*, 2011, **10**, 188.
- N. J. Gesmundo, B. Sauvagnat, P. J. Curran, M. P. Richards, C. L. Andrews, P. J. Dandliker and T. Cernak, *Nature*, 2018, **557**, 228–232.
- D. G. Anderson, S. Levenberg and R. Langer, *Nat. Biotechnol.*, 2004, **22**, 863.
- J. J. Green, R. Langer and D. G. Anderson, *Acc. Chem. Res.*, 2008, **41**, 749–759.
- R. Chapman, A. J. Gormley, M. H. Stenzel and M. M. Stevens, *Angew. Chem., Int. Ed.*, 2016, **55**, 4500–4503.
- G. Ng, J. Yeow, R. Chapman, N. Isahak, E. Wolvetang, J. J. Cooper-White and C. Boyer, *Macromolecules*, 2018, **51**, 7600–7607.
- G. Moriceau, G. Gody, M. Hartlieb, J. Winn, H. Kim, A. Mastrangelo, T. Smith and S. Perrier, *Polym. Chem.*, 2017, **8**, 4152–4161.
- J. Tanaka, S. Tani, R. Peltier, E. H. Pilkington, A. Kerr, T. P. Davis and P. Wilson, *Polym. Chem.*, 2018, **9**, 1551–1556.
- J. Tanaka, A. S. Gleinich, Q. Zhang, R. Whitfield, K. Kempe, D. M. Haddleton, T. P. Davis, S. Perrier, D. A. Mitchell and P. Wilson, *Biomacromolecules*, 2017, **18**, 1624–1633.



- 10 J. Zhang, J. Tanaka, P. Gurnani, P. Wilson, M. Hartlieb and S. Perrier, *Polym. Chem.*, 2017, **8**, 4079–4087.
- 11 B. Couturaud, P. G. Georgiou, S. Varlas, J. R. Jones, M. C. Arno, J. C. Foster and R. K. O'Reilly, *Macromol. Rapid Commun.*, 2018, 1800460.
- 12 A. M. Lunn and S. Perrier, *Macromol. Rapid Commun.*, 2018, **39**, 1800122.
- 13 A. B. Cook, R. Peltier, M. Hartlieb, R. Whitfield, G. Moriceau, J. A. Burns, D. M. Haddleton and S. Perrier, *Polym. Chem.*, 2018, **9**, 4025–4035.
- 14 P. Gurnani, C. Sanchez-Cano, K. Abraham, H. Xandri-Monje, A. B. Cook, M. Hartlieb, F. Lévi, R. Dallmann and S. Perrier, *Macromol. Biosci.*, 2018, 1800213.
- 15 P. Gurnani, A. M. Lunn and S. Perrier, *Polymer*, 2016, **106**, 229–237.
- 16 J. Yeow, R. Chapman, J. Xu and C. Boyer, *Polym. Chem.*, 2017, **8**, 5012–5022.
- 17 J. Yeow, R. Chapman, A. J. Gormley and C. Boyer, *Chem. Soc. Rev.*, 2018, **47**, 4357–4387.
- 18 E. Liarou, R. Whitfield, A. Anastasaki, N. G. Engelis, G. R. Jones, K. Velonia and D. M. Haddleton, *Angew. Chem., Int. Ed.*, 2018, **57**, 8998–9002.
- 19 J. Tan, D. Liu, Y. Bai, C. Huang, X. Li, J. He, Q. Xu and L. Zhang, *Macromolecules*, 2017, **50**, 5798–5806.
- 20 R. Chapman, A. J. Gormley, K.-L. Herpoldt and M. M. Stevens, *Macromolecules*, 2014, **47**, 8541–8547.
- 21 L. Zhifen, L. Yue and A. Zesheng, *Angew. Chem., Int. Ed.*, 2017, **129**, 14040–14044.
- 22 G. Gody, R. Barbey, M. Danial and S. Perrier, *Polym. Chem.*, 2015, **6**, 1502–1511.
- 23 M. Zamfir and J.-F. Lutz, *Nat. Commun.*, 2012, **3**, 1138.
- 24 M. Ouchi, N. Badi, J.-F. Lutz and M. Sawamoto, *Nat. Chem.*, 2011, **3**, 917.
- 25 J.-F. Lutz, *Polym. Chem.*, 2010, **1**, 55–62.
- 26 A. Anastasaki, B. Oschmann, J. Willenbacher, A. Melker, M. H. C. van Son, N. P. Truong, M. W. Schulze, E. H. Discekici, A. J. McGrath, T. P. Davis, C. M. Bates and C. J. Hawker, *Angew. Chem., Int. Ed.*, 2017, **56**, 14483–14487.
- 27 A. Anastasaki, V. Nikolaou and D. M. Haddleton, *Polym. Chem.*, 2016, **7**, 1002–1026.
- 28 F. Alsubaie, A. Anastasaki, V. Nikolaou, A. Simula, G. Nurumbetov, P. Wilson, K. Kempe and D. M. Haddleton, *Macromolecules*, 2015, **48**, 5517–5525.
- 29 F. Alsubaie, A. Anastasaki, V. Nikolaou, A. Simula, G. Nurumbetov, P. Wilson, K. Kempe and D. M. Haddleton, *Macromolecules*, 2015, **48**, 6421–6432.
- 30 C. Waldron, Q. Zhang, Z. Li, V. Nikolaou, G. Nurumbetov, J. Godfrey, R. McHale, G. Yilmaz, R. K. Randev, M. Girault, K. McEwan, D. M. Haddleton, M. Driesbeke, A. J. Haddleton, P. Wilson, A. Simula, J. Collins, D. J. Lloyd, J. A. Burns, C. Summers, C. Houben, A. Anastasaki, M. Li, C. R. Becer, J. K. Kiviahio and N. Risangud, *Polym. Chem.*, 2014, **5**, 57.
- 31 Q. Zhang, A. Anastasaki, G.-Z. Li, A. J. Haddleton, P. Wilson and D. M. Haddleton, *Polym. Chem.*, 2014, **5**, 3876–3883.
- 32 A. Anastasaki, V. Nikolaou, Q. Zhang, J. Burns, S. R. Samanta, C. Waldron, A. J. Haddleton, R. McHale, D. Fox and V. Percec, *J. Am. Chem. Soc.*, 2014, **136**, 1141–1149.
- 33 Q. Zhang, P. Wilson, Z. Li, R. McHale, J. Godfrey, A. Anastasaki, C. Waldron and D. M. Haddleton, *J. Am. Chem. Soc.*, 2013, **135**, 7355–7363.
- 34 A. H. Soeriyadi, C. Boyer, F. Nyström, P. B. Zetterlund and M. R. Whittaker, *J. Am. Chem. Soc.*, 2011, **133**, 11128–11131.
- 35 G. Gody, T. Maschmeyer, P. B. Zetterlund and S. Perrier, *Macromolecules*, 2014, **47**, 3451–3460.
- 36 G. Gody, T. Maschmeyer, P. B. Zetterlund and S. Perrier, *Nat. Commun.*, 2013, **4**, 2505.
- 37 A. Kerr, M. Hartlieb, J. Sanchis, T. Smith and S. Perrier, *Chem. Commun.*, 2017, **53**, 11901–11904.
- 38 J. Zhang, R. Deubler, M. Hartlieb, L. Martin, J. Tanaka, E. Patyukova, P. D. Topham, F. H. Schacher and S. Perrier, *Macromolecules*, 2017, **50**, 7380–7387.
- 39 A. Kuroki, P. Sangwan, Y. Qu, R. Peltier, C. Sanchez-Cano, J. Moat, C. G. Dowson, E. G. L. Williams, K. E. S. Locock, M. Hartlieb and S. Perrier, *ACS Appl. Mater. Interfaces*, 2017, **9**, 40117–40126.
- 40 J. Zhang, G. Gody, M. Hartlieb, S. Catrouillet, J. Moffat and S. Perrier, *Macromolecules*, 2016, **49**, 8933–8942.
- 41 C. Bray, R. Peltier, H. Kim, A. Mastrangelo and S. Perrier, *Polym. Chem.*, 2017, **8**, 5513–5524.
- 42 N. G. Engelis, A. Anastasaki, G. Nurumbetov, N. P. Truong, V. Nikolaou, A. Shegiwal, M. R. Whittaker, T. P. Davis and D. M. Haddleton, *Nat. Chem.*, 2016, **9**, 171.
- 43 T. R. Barlow, J. C. Brendel and S. Perrier, *Macromolecules*, 2016, **49**, 6203–6212.
- 44 C. Footman, P. A. de Jongh, J. Tanaka, R. Peltier, K. Kempe, T. P. Davis and P. Wilson, *Chem. Commun.*, 2017, **53**, 8447–8450.
- 45 V. Nikolaou, A. Simula, M. Driesbeke, N. Risangud, A. Anastasaki, K. Kempe, P. Wilson and D. M. Haddleton, *Polym. Chem.*, 2016, **7**, 2452–2456.
- 46 J. Collins, J. Tanaka, P. Wilson, K. Kempe, T. P. Davis, M. P. McIntosh, M. R. Whittaker and D. M. Haddleton, *Bioconjugate Chem.*, 2015, **26**, 633–638.
- 47 Q. Zhang, Z. Li, P. Wilson and D. M. Haddleton, *Chem. Commun.*, 2013, **49**, 6608–6610.
- 48 C. Boyer and T. P. Davis, *Chem. Commun.*, 2009, 6029–6031.
- 49 D. R. Carroll, A. P. Constantinou, N. Stingelin and T. K. Georgiou, *Polym. Chem.*, 2018, **9**, 3450–3454.
- 50 N. G. Engelis, A. Anastasaki, R. Whitfield, G. R. Jones, E. Liarou, V. Nikolaou, G. Nurumbetov and D. M. Haddleton, *Macromolecules*, 2018, **51**, 336–342.
- 51 A. Lotierzo, R. M. Schofield and S. A. F. Bon, *ACS Macro Lett.*, 2017, **6**, 1438–1443.
- 52 E. Liarou, A. Anastasaki, R. Whitfield, C. E. Iacono, G. Patias, N. G. Engelis, A. Marathianos, G. R. Jones and D. M. Haddleton, *Polym. Chem.*, 2019, DOI: 10.1039/C8PY01720D.
- 53 A. Postma, T. P. Davis, G. Li, G. Moad and M. S. O'Shea, *Macromolecules*, 2006, **39**, 5307–5318.

