Diphenylaminostyryl-substituted quinolizinium derivatives as fluorescent light-up probes for duplex and quadruplex DNA†

Avijit Kumar Das, Heiko Ihmels* and Sarah Kölsch

(E)-2-[(Diphenylamino)styryl]quinolizinium (3a) and 2,2′-[(phenylimino)-bis[(E)-1′,1″-styryl]]-bis[quinolizinium] (3b) were synthesized, and their interactions with duplex DNA and quadruplex DNA were investigated with a particular focus on their ability to operate as DNA-sensitive fluorescent probes. Due to the significantly different size and steric demand of these quinolizinium derivatives they exhibit different binding modes. Thus, 3a intercalates into duplex DNA and binds through π stacking to quadruplex DNA, whereas 3b favours groove binding to both DNA forms. The emission intensity of these compounds is very low in aqueous solution, but it increases drastically upon association with duplex DNA by a factor of 11 (3a) and >100 (3b) and with quadruplex DNA by a factor of >100 (3a) and 10 (3b), with emission bands between 600 and 750 nm.

Introduction

The detection of nucleic acids is an important tool in clinical, forensic, and biological studies and applications.1 And among the different chemical or biological tools that may be used for DNA analysis,2 emission spectroscopy is one of the most versatile and efficient methods.3 As a result, numerous fluorescent probes have been reported that allow the staining as well as qualitative and quantitative detection of DNA in cell-free medium and in cells.4 Specifically, these molecular probes that indicate the nucleic acid selectively with an increase in the emission intensity upon association with the biomacromolecule (light-up probes) are very useful fluorimetric markers.5 Along with the regular duplex DNA there is also an increasing interest in the detection of non-canonical DNA forms, with G-quadruplex DNA (G4-DNA) being the most prominent one. As this DNA form is proposed to have essential biological relevance and function,6 probes have been developed recently that enable its selective fluorimetric detection.7

In this context, it has been demonstrated that styryl dyes with a cationic hetarene unit exhibit favourable properties for DNA sensing:10–12 (i) the cationic charge of the dyes increases the affinity of the probe to DNA;8 (ii) in donor–acceptor substituted styryl dyes ICT (intramolecular charge transfer) states are possible, and these properties may change upon binding of the dye to DNA, thus enabling sensitive detection; and (iii) the large Stokes shifts in donor–acceptor substituted styryl dyes reduce the interference between excitation and detection light.8,9 As a result, styryl dyes are widely used as probes in fluorescence-based bioanalytical applications, specifically DNA staining.10–12

In our studies on the development of DNA-sensitive fluorescent probes, we have demonstrated that anelated quinolizinium derivatives are versatile DNA-binding ligands that may be used for fluorimetric detection.13 In addition, we have shown that especially donor-substituted quinolizinium derivatives have favourable photophysical properties and may therefore be used as versatile building blocks in fluorescent light-up probes.14 Based on these observations we proposed that styryl-substituted quinolizinium derivatives may also be promising DNA-sensitive light-up probes with useful absorption and emission properties. So far, only a few styrylquinolizinium derivatives are known, and some of these have been already applied for fluorimetric DNA detection.15 Among the derivatives investigated so far, the aminostyryl-substituted quinolizinium derivatives appeared to be the most promising candidates as they exhibit the typical advantages of donor–acceptor dyes. Therefore, we decided to vary this motif and employ a diphenylaminostyryl substituent as a donor unit, because the triarylamine unit is well known for its strong electron-donating properties,16 and it was already shown to be a useful functionality in fluorescent chemosensors.17 Herein, we present the synthesis of the resulting diphenylamino-substituted styryl-quinolizinium derivatives 3a and 3b along with investigations of their interactions with duplex and quadruplex DNA with a
particular focus on their potential application as DNA-sensitive fluorescent probes.

Results

Synthesis

The styrylquinolizinium derivatives 3a and 3b were synthesized by a Knoevenagel-condensation of 2-methylquinolizinium (1) with the mono- and diformyl-substituted triphenylamine derivatives 2a and 2b in 70% and 67% yield, respectively (Scheme 1). The chemical structures of the ligands were confirmed by 1D and 2D $^1$H NMR and $^{13}$C NMR spectroscopy, mass spectrometry and elemental analysis (cf. ESI†).

Absorption and emission properties

The absorption and emission properties of compounds 3a and 3b were investigated in different solvents, namely CH$_2$Cl$_2$, CHCl$_3$, CH$_3$CN, MeOH, EtOH, DMSO and aqueous buffer at pH 7. The absorption spectra of 3a and 3b display a long-wavelength absorption maximum ranging from 450 nm (3a) and 473 nm (3b) in aqueous buffer to 500 nm (3a) and 512 nm (3b) in CH$_2$Cl$_2$ (Table 1, Fig. S1†). In most solvents, the emission intensity of derivatives 3a and 3b is very weak. Thus, in DMSO, MeOH, CH$_3$CN and buffer solution only a weak, broad emission band was observed between 600 and 750 nm, whereas in the protic polar solvent EtOH the emission intensity at 695 nm is slightly higher.

In contrast, both 3a and 3b show more intense emission in the less polar, chlorinated solvents CHCl$_3$ (3a: 638 nm, 3b: 620 nm) and CH$_2$Cl$_2$ (3a: 695 nm, 3b: 690 nm). In general, the emission quantum yields in CHCl$_3$ ($\Phi_{fl} = 0.34$ and 0.27) are higher as compared to those in CH$_2$Cl$_2$ ($\Phi_{fl} = 0.21$ and 0.12) (cf. ESI, Fig. S1, B1 and B2†).

To assess whether the low emission quantum yields of compounds 3a and 3b are caused by conformational changes in the excited state, the fluorescence spectra were recorded in media of high viscosity, namely aqueous solutions with increasing glycerol content. 22 Whereas in aqueous solution compounds 3a and 3b are basically non-fluorescent ($\Phi_{fl} \approx 0.002$), an increasing content of glycerol (wt% glycerol: 0, 50, 100%) resulted in a large increase in the fluorescence quantum yield ($\Phi_{fl} = 0.30$ for 3a and $\Phi_{fl} = 0.14$ for 3b), along with a red shift of the absorption maxima from $\lambda_{abs} = 450$ nm to 472 nm (3a) and $\lambda_{abs} = 473$ nm to 493 nm (3b) (Fig. S2†).

DNA binding properties

Spectrophotometric and spectrofluorimetric titrations. The interactions of derivatives 3a and 3b with calf thymus (ct) DNA and the quadruplex forming oligonucleotide d[AGGGTAA]$_3$GGG (22AG) were monitored by photometric and fluorimetric titrations in BPE buffer solution (ct DNA) or in K-phosphate buffer (22AG) at 20 °C and pH 7 (Fig. 1 and 2). In both cases, the absorption spectrum of the quinolinium derivatives 3a

<p>| Table 1 | Absorption and emission properties of 3a and 3b in different solvents |</p>
<table>
<thead>
<tr>
<th>Solvent</th>
<th>$\lambda_{abs}^{a}$/nm</th>
<th>$\epsilon^{b}$/M$^{-1}$cm$^{-1}$</th>
<th>$\lambda_{fl}^{c}$/nm</th>
<th>$\lambda_{abs}^{d} - \lambda_{fl}^{d}$/cm$^{-1}$</th>
<th>$\Phi_{fl}^{e}$</th>
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<tr>
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<tr>
<td>3b</td>
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<td></td>
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<td></td>
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</tr>
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<td>4.79</td>
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$^a$Long-wavelength absorption maximum; $^b$ $\epsilon$ M$^{-1}$cm$^{-1}$; $^c$ Fluorescence emission maximum; $^d$ Difference between absorption maximum and emission maximum at lowest energy. $^e$ Fluorescence quantum yield relative to rhodamine B in EtOH ($\Phi_{fl} = 0.7$) (ref. 21). $^f$ In 10 mM BPE buffer solution at pH 7.0 and 20 °C.

![Fig. 1](image-url) Photometric titration of 3a (1) and 3b (2) with ct DNA (A, $c = 1.88$ mM in base pairs) in BPE buffer (10 mM, pH 7.0; solutions of ligands with 10% v/v DMSO, $c_{ligand} = 20$ µM) and with 22AG (B, $c = 200$ µM) in K-phosphate buffer at pH 7.0 at 20 °C ($c_{ligand} = 20$ µM). The arrows indicate the changes of absorption upon addition of DNA. Inset: Changes of the ligand absorption with increasing DNA concentration.
DNA with binding constants of $\Phi$ increased significantly in the presence of DNA (Fig. 2). Thus, $\Phi_{30}$ (ct DNA, continuous increase of the weak emission band by a factor of $8.8 \times 10^4 \text{ M}^{-1}$) and $\Phi_{30}$ (ct DNA) were achieved. Further addition of ct DNA led to an increase in the absorbance with a red shift of the absorption maximum to 470 nm ($\Delta A = 20 \text{ nm}$; 3a) and 513 nm ($\Delta A = 40 \text{ nm}$, 3b) (Fig. 1A). On titration of 22AG to compounds 3a and 3b the absorbance at 450 nm (3a) and 473 nm (3b) decreased until LDRs of >20 (3a) and >18.6 (3b) were achieved. At a higher concentration of 22AG (LDR <0.4 and <0.7, resp.) a red shifted absorption peak was formed at 480 nm ($\Delta A = 30 \text{ nm}$) (3a) and 505 nm ($\Delta A = 32 \text{ nm}$) (3b) (Fig. 1B). The data from spectrophotometric titrations were used to determine the binding constant, $K_b$, obtained from the fitting of the experimental binding isotherms to the theoretical model (cf. ESI, Fig. S3†). Thus, ligands 3a and 3b bind to ct DNA with binding constants of $K_b = 4.3 \times 10^4 \text{ M}^{-1}$ and $K_b = 8.8 \times 10^4 \text{ M}^{-1}$.

The emission intensities of both compounds 3a and 3b increased significantly in the presence of DNA (Fig. 2). Thus, the addition of ct DNA and 22AG to derivative 3a resulted in a continuous increase of the weak emission band by a factor of 30 (ct DNA, $\Phi_{30} = 0.16$) and 176 (22AG, $\Phi_{30} = 0.26$) and small blue shifts ($\Delta \lambda \approx 20 \text{ nm}$) of the emission maximum (Fig. 2, A1 and B1).

Similarly, the weak emission of compound 3b increased by factors of 128 and 10 on addition of ct DNA ($\Phi_{30} = 0.40$) and 22AG ($\Phi_{30} = 0.13$), respectively, and the emission maxima were slightly blue shifted to 660 nm and 655 nm on addition of ct DNA and 22AG (Fig. 2, A2 and B2). From the fluorescence titration experiments, the limit of detection (LOD) of 3a and 3b was estimated to be 0.02 $\mu$M and 0.79 $\mu$M for ct DNA and 0.05 $\mu$M and 0.01 $\mu$M for 22AG, respectively (cf. ESI, Table S1†). To check whether the fluorimetric analysis is disturbed by the formation of singlet oxygen and subsequent DNA damage, the irradiation of a 3b–DNA complex (LDR = 0.3) under aerobic conditions was monitored by CD spectroscopy (cf. ESI, Fig. S5†). Under these conditions, no significant changes in the CD signals corresponding to the DNA absorption (≈300 nm) were observed, which indicates negligible changes in the DNA structure after irradiation. In another experiment, we examined the influence of oxygen on the excited singlet state of 3b by recording emission spectra at different oxygen concentrations. Namely, the spectra were measured under anaerobic conditions, in air or in oxygen-saturated solution under otherwise identical conditions (cf. ESI, Fig. S6†). Notably, the fluorescence intensity of ligand 3b and of the 3b–ct DNA complex is essentially independent of the oxygen concentration, which indicates that oxygen does not significantly quench the excited singlet state.

**CD and LD-spectroscopic analysis**

Complementary analysis with circular dichroism (CD) and flow linear dichroism (LD) spectroscopy revealed that in the presence of ct DNA, ligands 3a and 3b display induced circular dichroism (ICD) and LD bands in the absorption range of the ligands (Fig. 3). A solution of ligand 3a with ct DNA exhibited negative ICD bands at 247 nm, 310 nm, 420 nm and 500 nm, all of which increased in intensity from LDR = 0.3 to 1.0. At higher LDR values, however, the negative long-wavelength bands at 420 nm and 500 nm developed into a very intense bisignate band with zero transition at the absorption maximum. Simultaneously, the initially intense positive CD signal of the DNA at 278 nm decreased drastically (Fig. 3, A1). At the same time, the addition of ct DNA to the solution of ligand 3a (LDR = 0–1.0) led to the formation of a gradually increasing negative LD signal in the absorption region of the ligand (350–550 nm). Notably, the intensity of the negative LD signal at 258 nm decreased with increasing LDR values (Fig. 3, B1).

The complexation of compound 3b with ct DNA resulted in the formation of positive ICD signals at 347 nm and 545 nm, along with a negative signal at 460 nm. Up to an LDR value of 0.5, the intensities of the two very strong positive CD bands at 545 nm and 460 nm increased, but at a larger LDR of 1.0 these CD bands decreased (Fig. 3, A2). The LD spectra of complexes of 3b and ct DNA revealed a very strong positive band at 536 nm that increased from LDR = 0.3 to 0.5 and subsequently disappeared at larger LDR = 1 in favour of a weak bisignate band with maxima at 540 nm and 440 nm. Concurrently, very
weak positive and negative LD signals were observed at 340 nm (+), 375 nm (−) and 440 nm (−) that strongly broadened with increasing LDR values (Fig. 3, B2).

On addition of derivatives 3a and 3b to G4-DNA 22AG the characteristic CD pattern of the hybrid-type structure of the quadruplex, i.e. with positive bands at 290 nm and 270 nm along with a negative band at 235 nm, did not change essentially (Fig. 3, C1 and C2). In the complex of 3a and 22AG, only a very weak bisignate ICD band was observed with maxima at 345 nm (+) and 410 nm (−) (Fig. 3, C1). In contrast, more pronounced CD signals developed in the absorption range of the derivative 3b upon addition of G4-DNA 22AG. Specifically, with increasing LDR from 0 to 2.0, a positive band at 350 nm and two very broad negative bands with maxima at 440 nm and 540 nm were observed (Fig. 3, C2). CD-spectroscopic analysis was also performed to determine the influence of the association of 3a and 3b on the quadruplex melting temperature, \(T_m\) (cf. ESI, Fig. S7†). Thus, the determination of the DNA melting temperature by monitoring the temperature-dependent CD intensity revealed an increase in the melting temperature, \(\Delta T_m\), in the presence of the ligands (3a: \(\Delta T_m = 3 \, ^\circ C\), 3b: \(\Delta T_m = 9 \, ^\circ C\), at LDR = 2).

**Discussion**

The absorption and emission properties of 3a and 3b in different solvents resemble the ones that have been reported previously for styryl substituted quinolizinium derivatives.13d Although the absorption and emission shifts of these compounds are slightly different in various polar aprotic and polar protic solvents (Table 1), there was no clear relationship identified between the absorption and emission energy and a particular solvent property. This observation indicates that several solvent properties, such as polarity and hydrogen bonding, contribute to the overall solvent effect to different extents. Notably, in CH\(_2\)Cl\(_2\) and CHCl\(_3\), derivatives 3a and 3b exhibit significant red shifts of the absorption bands (Fig. S1†) which are frequently observed for cationic dyes due to the high polarizability of chloroalkane solvents.26 In addition, it should be considered that these compounds are poorly soluble in these solvents, which leads to dye aggregation. Thus, the red-shifted absorption in these solvents may indicate the aggregate formation.27 The emission intensity of 3a and 3b is very low in polar solvents (<0.04), most likely due to the radiationless deactivation of the excited state by conformational changes, such as rotation about the vinyl-arene bond, as commonly observed for styryl dyes.28 This assumption was clearly supported by the observation that the emission intensity increases with increasing viscosity of the solution, namely in glycerol–water mixtures (cf. ESI, Fig. S2†), as the rotational relaxation of the excited molecule becomes slower than the fluorescence at higher viscosity. Presumably, the slightly larger emission quantum yields of 3a and 3b in CHCl\(_3\) and CH\(_2\)Cl\(_2\) (\(\Phi_\text{f} = 0.1\) to 0.3) are the result of their low solubility in these solvents and are thus caused by an aggregation-induced enhancement of the emission intensity.29

The binding studies of 3a and 3b with ct DNA and 22AG by photometric, fluorimetric and polarimetric titrations clearly confirmed the association of these ligands with DNA. Firstly, the photometric titrations show a development of absorption bands that is characteristic of DNA-binding ligands, i.e. a hypochromic effect along with a red shift of the absorption band with progressing titration of DNA.30 Nevertheless, the lack of isosbestic points and the appearance of two distinctly different sections of the titration (Fig. 1) point to different binding modes that depend on the LDR. Thus, at the beginning of the titration, i.e. at large LDR values, the ligands form aggregates along the DNA backbone because there are not enough binding sites available. At smaller LDR values, i.e. with ample number of binding sites, the ligands may associate with the DNA by intercalation or groove binding. Notably, the sterically demanding diphenylamino-substituent severely hinders the access of a ligand close to an already occupied DNA binding site, which explains the large amount of DNA required to reach saturation. Unfortunately, this heterogeneous binding of both ligands with duplex and quadruplex DNA, that even changes in the course of the titrations, seriously hampers the determination of meaningful binding constants from the binding isotherms, as the ones calculated only reflect an apparent binding constant, i.e. an average constant from all...
contributing binding modes, thus showing different overall affinities of the ligands toward a particular DNA form. Hence, the apparent binding constants of ligands 3a and 3b with ct DNA show a higher overall affinity of 3b to this DNA form. The $K_b$ values ($3a: K_b = 4.3 \times 10^4$ M$^{-1}$, $3b: K_b = 8.8 \times 10^4$ M$^{-1}$) differ only by a factor of ca. 2 and are both comparable to the ones obtained for classical intercalators.

Additional information about the binding mode was obtained by CD- and LD-spectroscopic studies. Thus, the strong bisignate ICD band in the long-wavelength absorption range of compound 3a at high LDR clearly confirms the aggregation of the dye at the DNA backbone (Fig. 3, A1). At lower LDR, a less intense negative ICD band appears, which indicates a different binding mode. But it should be noted that the latter band is still overlapped by the ICD signal of the aggregate, resulting in a combination of spectra that cannot be dissected. Similarly, the CD bands in the absorption range of DNA and the ligand (<320 nm) also severely overlap. As a result, this combination of overlapping bands, whose contribution also changes with LDR, does not allow a conclusive binding mode analysis at low LDR. At the same time, the negative LD signal in the absorption range of 3a (Fig. 3, B1) clearly indicates an intercalating binding mode of this ligand with ct DNA.

In the case of ligand 3b, the very intense bisignate CD signal between 400 nm and 600 nm is maintained at all employed LDR ratios, and in each case the bands correspond to the two absorption maxima observed during photometric titration (Fig. 3 A2 and B2). This observation shows that this CD spectrum is also a combination of separate bands. Notably, the CD bands of the ligand are very strong, as seen directly from the comparison with the DNA bands, and this is usually observed for groove binders. The LD-spectroscopic analysis supports this interpretation, as the positive LD band at the long-wavelength absorption of the ligand unambiguously indicates groove binding. The weak positive LD bands at 375 nm and 440 nm seem to contradict the latter interpretation, but it should be noted that the size and shape of the ligand do not allow for a complete accommodation and fit of the whole molecule within the groove, so that at least partially one of the quinolinium units point outside the groove or even intercalate. In this arrangement, this quinolinium chromophore is decoupled from the conjugated π system and essentially oriented in the same direction as the DNA bases, thus resulting in a positive LD signal. Such bisignate LD spectra have also been observed with bichromophoric bis-ammonaphthalimide-substituted Tröger bases and interpreted similarly. It is worth noting, however, that the intensity of CD and LD signals decreased significantly at higher LDR, presumably because at this higher ligand loading there are not sufficient binding sites available in the grooves, thus reducing the relative number of intercalated or groove-bound ligands.

The two ligands 3a and 3b do not carry special substituents that may be involved in specific interactions with the DNA, such as hydrogen bonding. Even the amino group is not available as it is sterically protected and the free lone pair of the amino group is highly delocalized. Therefore, the association of these ligands should be mainly governed by attractive dispersion interactions, such as π stacking and van der Waals interactions, with the binding site and by the thermodynamically favorable counter-ion release from the DNA upon association of the cationic ligand. Therefore, the different binding modes of ligands 3a and 3b are mainly based on their different size and shape. Both compounds are structurally flexible containing triphenylamine as a rather hydrophobic unit and the quinolinium as the DNA-binding fragment. In the case of 3a, the bulky diaryl unit may point outside the binding site or even accommodate partly in the groove when the quinolinium intercalates into DNA. Obviously, this binding mode is not favoured with 3b as only one of the two quinolinium units can intercalate and the majority of the molecule would still protrude from the binding site so that it is still exposed to the solvent, thus reducing the role of the hydrophobic effect as a driving force for intercalation. As a result, ligand 3b binds to the grooves because in this binding mode this sterically demanding molecule fits better to the binding site as a whole, which may be driven by entropic factors.

The association of 3a and 3b with G4-DNA 22AG was confirmed with CD spectroscopy. In the presence of 3a and 3b, the CD bands of the quadruplex 22AG change only marginally (Fig. 3, C1 and C3), which indicates the preservation of the quadruplex structure upon complex formation. The complex formation is also clearly indicated by the increased melting temperature, $ΔT_m$, of the quadruplex in the presence of ligands 3a and 3b, which indicates significant thermodynamic stabilization of the quadruplex toward unfolding due to ligand association. At the same time, the disappearance of the weak shoulder around 250 nm usually denotes the disappearance of one minor form from the equilibrium of different quadruplex structures because of the stabilization of a basket-type or chair-type quadruplex structure by the ligand. In the case of ligand 3a, only a very weak ICD signal in the absorption range of the ligand developed, which is often observed for ligands that bind to the quadruplex by terminal π stacking. In contrast, the association of ligand 3b with 22AG leads to the formation of a much more pronounced ICD signal, which is proposed to be a characteristic feature of groove-binding G4-DNA ligands.

In the absence of DNA, the styrylquinolinium derivatives 3a and 3b are weakly fluorescent with very low quantum yields in aqueous buffer solution (Table 1) due to radiationless deactivation by conformational relaxation of the excited state (see Discussion above). Upon association with duplex and quadruplex DNA, however, the emission intensity of these ligands increases significantly (Fig. 2). As observed for several DNA-sensitive fluorescent light-up probes, this effect is most likely the result of the restricted conformational freedom of movement of the ligand within the binding site, which is comparable to the light-up effect observed in glycerol. Similar fluorescence light-up effects on DNA binding have been observed with annelated quinolinium derivatives and diphenylamino-substituted derivatives of quinolinium, pyridinium and imidazolium.
Most interesting is the observation that the fluorescence light-up effect of 3b on binding with ct DNA is 4 times stronger than that of 3a (Fig. 4A). Considering that the light-up effect is mainly caused by the restricted conformational flexibility of the ligand in the sterically constrained binding site, and not necessarily with the binding strength, this observation indicates that the groove-bound ligand 3b is accommodated more tightly in its binding site than the intercalated molecule 3a. Conversely, when bound to G4-DNA, the light-up factor of 3a is 17 times larger than that of 3b (Fig. 4B), so that in this case the conformational flexibility of 3a is more suppressed in the DNA binding site.

Conclusion

In summary, it was demonstrated that the diphenylaminotyryl-substituted quinolizinium derivatives 3a and 3b bind to duplex and quadruplex DNA. Due to the significantly different size and steric demand of these ligands they exhibit different binding modes. Thus, 3a intercalates into ct DNA and binds through \( \pi \) stacking to G4-DNA 22AG, whereas 3b favours groove binding to both DNA forms. As the very low emission intensity of these compounds increases strongly upon association with DNA and the emission wavelength is close to the NIR range, which is favourable for biological applications, these compounds are promising platforms for the development of DNA-sensitive fluorescent probes. In this regard, the different effects of the actual DNA form and binding mode on the extent of the light-up effect may be used for selective DNA detection.

Conflicts of interest

There are no conflicts to declare.

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Notes and references


