



Cite this: *Org. Biomol. Chem.*, 2019, **17**, 8816

Received 27th August 2019,
Accepted 17th September 2019

DOI: 10.1039/c9ob01881f

rsc.li/obc

Vinylboronic acid-caged prodrug activation using click-to-release tetrazine ligation†

Lianne P. W. M. Lelieveldt, Selma Eising, Abel Wijen and Kimberly M. Bonger *

Bioorthogonal reactions can be performed selectively in the presence of any biological functional group and are widely used to achieve site-selective chemical modifications of biomolecules. The click-to-release reaction is a bioorthogonal bond-cleavage variant that has gained much interest over the last few years. The bioorthogonal reaction between tetrazines and *trans*-cyclooctenes or vinyl ethers, for example, initiates the release of a small molecule immediately after the cycloaddition with tetrazines. Recently, our group reported that vinylboronic acids (VBAs) give exceptionally high reaction rates in the bioorthogonal inverse electron-demand Diels–Alder reaction with tetrazines that are substituted with boron-coordinating ligands. In the present study, we show that VBAs can be used in a click-to-release variant and demonstrate its bioorthogonality with a VBA-protected doxorubicin prodrug. We show that the cytotoxicity of doxorubicin is silenced by the attachment of the VBA, and activity can be largely restored upon the reaction with a tetrazine, inducing cell death.

recently reported a new bioorthogonal tetrazine ligation utilizing vinylboronic acids (VBAs).^{4–7} We showed that by introducing a boronic acid moiety on an alkene, reaction rates of an inverse electron-demand Diels–Alder (iEDDA) reaction with pyridyl-containing tetrazines improved by several orders of magnitude compared to the non-modified linear alkene.⁴ The hydrophilic properties and the small size of VBAs compared to other bioorthogonal reactants make them attractive for use in biomolecular labeling experiments. VBAs were shown to be biocompatible, non-toxic, and highly stable in aqueous media and cell lysates. In addition, while the number of bioorthogonal reactions used *in situ* and *in vivo* is limited due to the need for toxic reagents (*e.g.* copper, in copper(i)-catalyzed alkyne–azide cycloaddition⁸) or possible side reactions of the reactants (*e.g.* strained alkenes or alkynes can react with cysteines⁹), VBAs appeared to be useful in cellular experiments.⁷ Reactions with vicinal diols, present in carbohydrates for example, were not observed when performing reactions in living cells.

Introduction

Chemical site-selective protein modifications have become increasingly popular for modification and control over protein functions *in vitro* and in living systems. Several reactions have been developed to decorate biomolecules with a desired functionality such as fluorophores, affinity probes or reactive tags. All these reactions have some properties in common as they must be selective over other functional groups in biomolecules, have fast reaction rates and proceed in aqueous media around physiological pH.¹ Reactions such as the strain-promoted alkyne–azide cycloaddition (SPAAC)² or the inverse electron-demand Diels–Alder reaction with tetrazines³ have been used extensively for modifying purposes. Our group has

The reactivity of VBAs depends on the tetrazine and its substituents. Since the boronic acid coordinates to the pyridyl ring of dipyrindyl-*s*-tetrazine, reaction rates were enhanced in comparison with tetrazines lacking such a potential Lewis base.⁶ This unique feature allows one to perform orthogonal bioorthogonal reactions with two different tetrazines, one with and one without substituents allowing for coordination, and using a VBA and a strained alkene.

The interest in bioorthogonal bond-cleavage reactions for activation of protected biologically relevant molecules *in vivo* has increased significantly (Fig. 1A). A substantial number of so-called ‘click-to-release’ reactions have been developed in the last few years.^{10–12} These reactions enable new applications *in vitro* and *in vivo*, such as the activation of prodrugs or fluorogenic compounds. Most reactions are based on the iEDDA reaction between a tetrazine and a (strained) alkene.¹¹

In 2013, the group of Robillard reported the first bioorthogonal click-to-release reaction based on the reaction of tetrazines with *trans*-cyclooctenes (TCOs) that possess a carbamate substituent at the allylic position (Fig. 1A).¹³ After the iEDDA

Department of Biomolecular Chemistry and Synthetic Organic Chemistry, Radboud University Nijmegen, The Netherlands. E-mail: k.bonger@science.ru.nl

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c9ob01881f



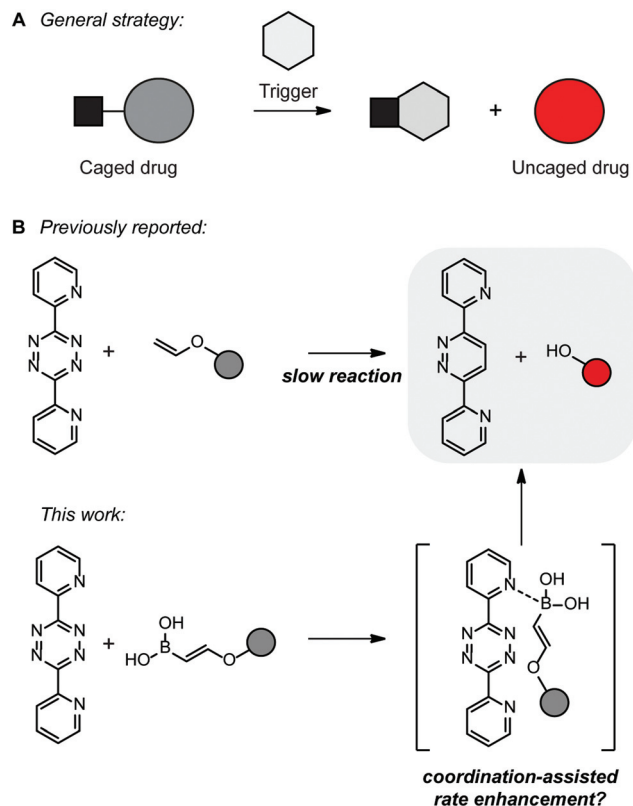


Fig. 1 (A) General scheme of a click-to-release reaction to uncage a prodrug. (B) Overview of the inverse electron-demand Diels–Alder cycloaddition with a tetrazine and a vinyl ether or vinylboronic acid to release an alcohol functionality.

reaction of the tetrazine and TCO, the 4,5-dihydropyridazine eliminates the leaving group to liberate both CO_2 and the amine. Unfortunately, part of the dihydropyridazine tautomerizes to the 1,4-dihydropyridazine, which does not lead to a release reaction and therefore this click-to-release process did not result in complete elimination. Over the last few years, the reaction has been optimized^{14,15} and the scope was further expanded¹⁶ and applied in biological systems for activation of prodrugs,^{17–23} RNA²⁴ or caged proteins.^{14,25,26} Recently, it was noted that this click-to-release reaction is environmentally sensitive and improvements on both the TCO and tetrazine resulted in ultrafast click-to-release reaction rates and the complete elimination of the leaving group.²⁷

In 2016 and 2017, the groups of Devaraj and Bernardes reported both the bioorthogonal click-to-release reactions of tetrazines and vinyl ethers (Fig. 1A).^{28,29} These alkenes bear their leaving group directly on the alkene and therefore this results in immediate full release of the alcohol after the iEDDA reaction. The vinyl ethers appeared to be stable in aqueous solutions for at least 8 hours and were suitable for the release of a prodrug in living cells.³⁰ The vinyl ethers have the advantage of being small in size, but have disappointingly low rate constants when reacting with tetrazines.

Since VBAs have proven to be useful in bioorthogonal conjugation, we envisioned that by introducing a boronic acid group on the vinyl ethers, we could increase the rate of these click-to-release reactions. In addition, similar to vinyl ethers, we expect that VBA ethers fully release the alcohol, or the amine when linked *via* a carbamate, after the iEDDA reaction. The proposed iEDDA click-to-release reaction mechanism of vinylboronic acids with tetrazines is shown in Fig. 1B. Upon cycloaddition and retro-Diels–Alder reaction, N_2 , the boric acid and the uncaged drug are released. We here explore the applicability of the VBA click-to-release strategy and develop a synthetic route to VBA protected prodrugs, which can be released using a tetrazine in the presence of cells.

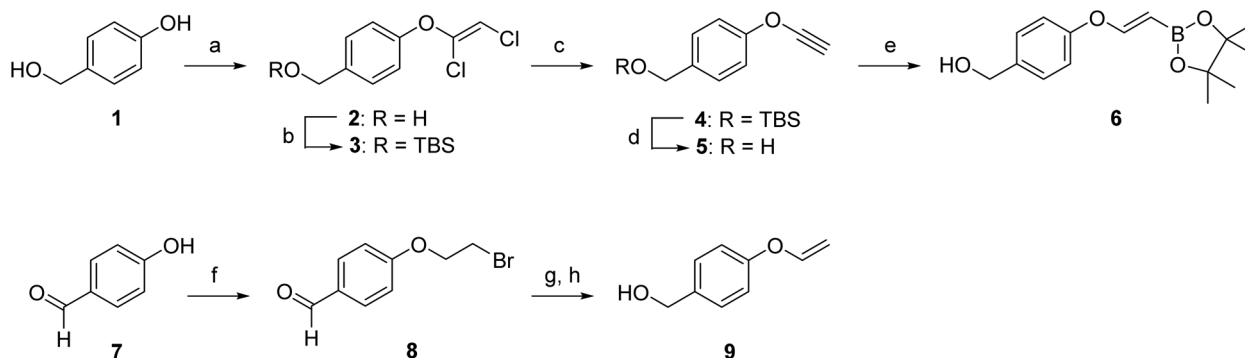
Results and discussion

We first aimed to compare the reactivity of the VBA ether with the unsubstituted vinyl ethers and synthesized VBA **6** and its primary alkene derivative **9** (Scheme 1). The synthesis of VBA **6** started with the etherification of 4-hydroxybenzyl alcohol **1** with trichloroethylene and subsequent silyl protection of the primary alcohol. Next, dichlorovinyl ether **3** was treated with *n*-BuLi to eliminate hydrogen chloride and to provide alkynyl ether **4**. Deprotection of the primary alcohol and hydroboration of alkyne **5** with pinacolborane yielded VBA pinacol ester **6**. In our kinetic experiments, we used the pinacol ester as a precursor for the free boronic acid as it hydrolyzes quickly in aqueous media (see Fig. SI-1†). The synthesis of vinyl ether **9** was performed following a literature procedure.³¹ Here, 4-hydroxybenzaldehyde (**7**) was first reacted with 1,2-dibromoethane to yield compound **8** which was subsequently eliminated using *t*-BuOK and the aldehyde was reduced using NaBH_4 to yield the corresponding benzyl alcohol **9**.

Having alkenes **6** and **9** in hand, we examined the second-order rate constants with 3,6-dipyridyl-*s*-tetrazine **10** in 75% MeOH in PBS at a controlled temperature of 20 °C (Fig. 2A). As we used the pinacol ester as a precursor, we incubated ester **6** for 2 h in the solvent mixture to achieve full hydrolysis to the free boronic acid before measuring its k_2 value. Second-order rate constants were obtained by following the decrease of the tetrazine UV absorbance in the visible region. The pseudo first-order constant k_{obs} was first determined by plotting the decay of tetrazine absorbance against time (minutes) for five different amounts of excess alkene (see also the ESI†). Next, the k_{obs} was plotted against the five different alkene concentrations and a linear regression fit gave the linear function, of which the slope corresponded to the k_2 (Fig. SI-2†). The second-order rate constant for VBA **6** was $3.3 \times 10^{-3} \text{ M}^{-1} \text{ s}^{-1}$ and 4-fold higher than the rate constant of vinyl ether **9**, clearly indicating a positive effect of the boronic acid on the reactivity of the alkene.

The observed rate constant of VBA ether **6** was lower than expected as we previously observed rate enhancements of up to 100-fold compared to the unsubstituted alkene derivatives.⁴ We hypothesize that the moderate rate enhancement is due to





Scheme 1 Synthesis of VBA ether **6** and vinyl ether **9**. (a) Trichloroethylene, K_2CO_3 , DMF, 70 °C, 16 h, 87%. (b) TBSCl, imidazole, DMF, 2 h, quant. (c) *n*-BuLi, Et₂O, -78 to -40 °C, 87%. (d) TBAF, THF, 0 °C, 91%. (e) Pinacolborane, RuHClCO(PPh₃)₃, toluene, 50 °C, 16 h, 76%. (f) Dibromoethane, K_2CO_3 , MeCN, reflux, 20 h, 79%. (g) *t*-BuOK, DMSO, r.t., 10 min. (h) NaBH₄, MeOH, r.t., 1.5 h, 18%.

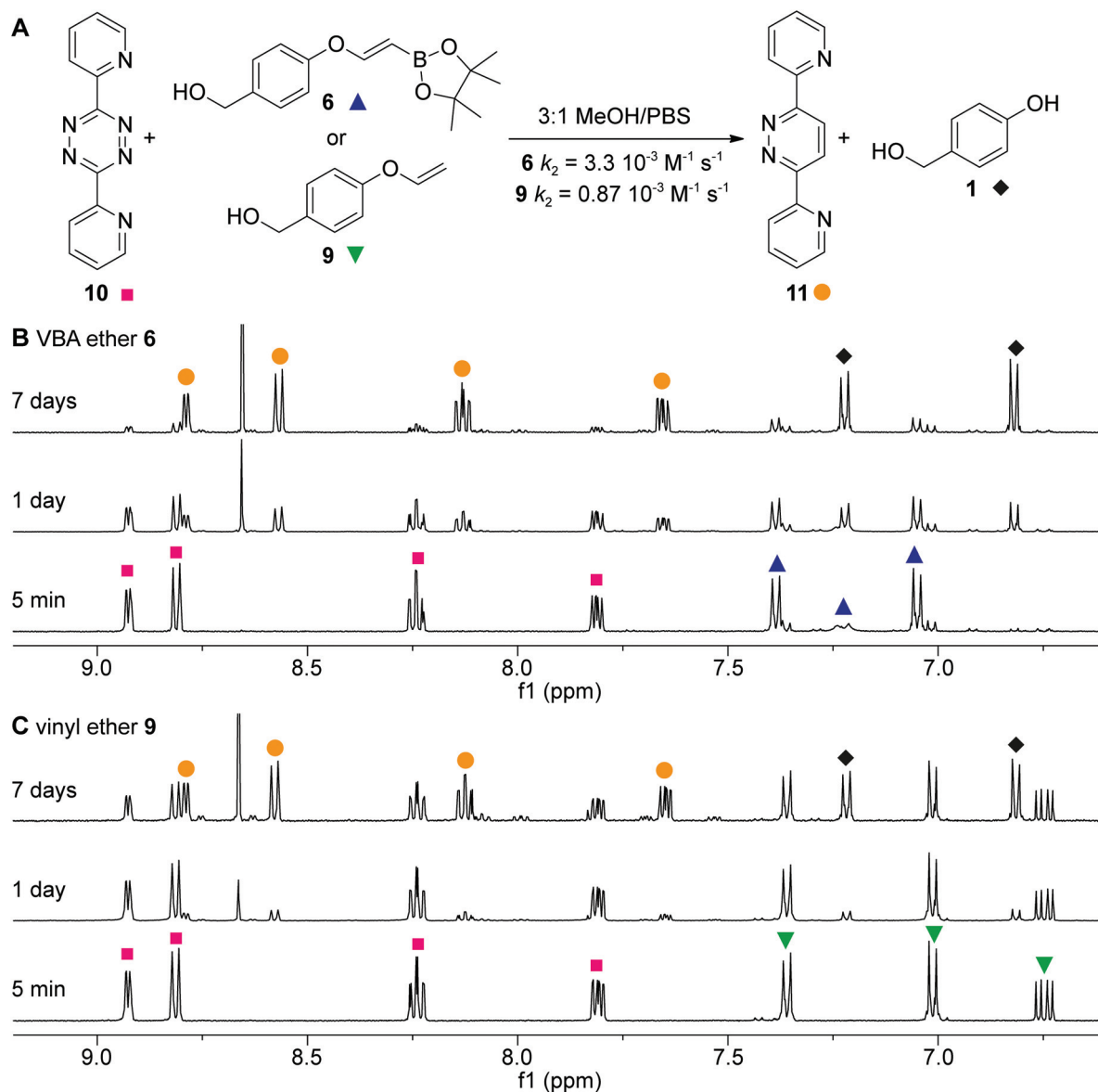


Fig. 2 Click-to-release reactions and reaction rates. (A) Scheme of the reaction of 3,6-dipyridyl-*s*-tetrazine Tz with VBA ether **6** and vinyl ether **9**. k_2 values were measured in 3 : 1 MeOH/PBS; (B) $^1\text{H-NMR}$ study of the reaction between tetrazine **10** (■) and VBA ether **6** (▲) yielding pyridazine **11** (●) and 4-hydroxybenzyl alcohol **1** (◆) in 3 : 1 CD₃OD/deuterated PBS; (C) similar to (B) with the reaction of Tz with vinyl ether **9** (▼).



the ether on the vinylic position. Since this alcohol is slightly electron-donating, we assume that the alkene becomes more electron-rich, thereby lowering the acidity of the boronic acid, resulting in less favorable coordination to the pyridyl substituents of the tetrazine.³²

Next, we looked in more detail into the click-to-release reaction of 3,6-dipyridyl-*s*-tetrazine **10** and alkenes **6** and **9** by ¹H NMR spectroscopy in 75% CD₃OD in deuterated PBS over time (Fig. 2B and C). A near to complete conversion of VBA ether **6** was observed after 7 days, while vinyl ether **9** gave around 50% conversion. Both reactions resulted in the clean formation of pyridazine **11** and 4-hydroxybenzyl alcohol **1**. No intermediates were observed showing that the initial cycloaddition step is rate limiting for both VBA ether **6** and vinyl ether **9**.

To further examine the click-to-release reaction with a VBA ether in a more complex system, we synthesized a VBA-caged cytotoxin doxorubicin and explored the effect on HeLa cells upon addition of a water soluble tetrazine **12** (Fig. 3A). For this, we aimed to couple doxorubicin to VBA alcohol **6**, which acts as a self-cleavable linker by 1,6 elimination upon release (Fig. 3B).³³ We commenced the synthesis from VBA alcohol **6** and activated the alcohol using bis(pentafluorophenyl)carbonate and Et₃N as a base, yielding successfully VBA-PFP carbonate **13**. Doxorubicin was subsequently added and reacted over-

night to obtain VBA-caged doxorubicin **14** (VBA-Dox) in reasonable yields.

First, we tested the stability of the VBA-Dox **14** at 100 μM in PBS at 37 °C before performing the click-to-release reactions with cells. As shown in Fig. SI-3,† hydrolysis of the pinacol ester was observed, as expected, while minor decomposition (around 10%) of VBA-Dox **14** was observed after 48 hours. Decomposition likely results from protodeboronation of the VBA cage.³⁴ Importantly, no free doxorubicin was observed after incubating VBA-Dox **14** in PBS for up to 7 days. Encouraged by these results we next evaluated the cycloaddition with a water soluble tetrazine **12** and followed the subsequent release over time in a buffer system with LCMS. Deprotection of **14** at 100 μM in PBS at 37 °C, using 1 equivalent of tetrazine **12** resulted in decaging and release of doxorubicin (Fig. SI-4†). No intermediate products were observed, indicating that the cleavage of the linker occurred instantaneously after the cycloaddition.

Next, we tested the VBA release and toxicity of doxorubicin on living HeLa cells (Fig. 3A). We first incubated HeLa cells with free doxorubicin and observed a dose dependent toxic effect, as expected (Fig. SI-5†). In addition, we incubated VBA-caged doxorubicin in the absence of a tetrazine. No toxicity was observed with 100 nM VBA-doxorubicin **14** while 1 μM

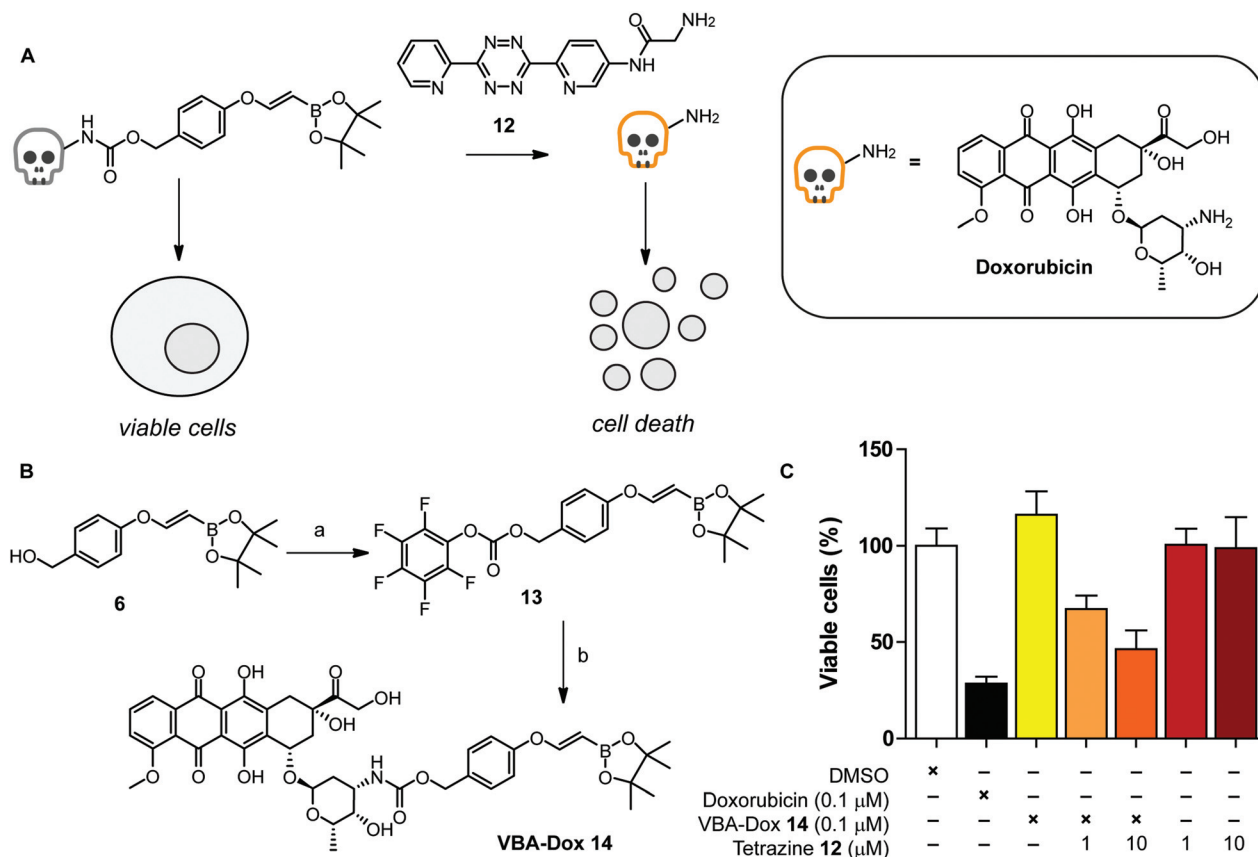


Fig. 3 Click-to-release reaction of VBA-Dox **14** to yield the free doxorubicin. (A) General overview of the caging strategy. (B) Synthetic route of VBA-Dox. (a) Bis(pentafluorophenyl)carbonate, Et₃N, r.t., 1 h, 84%. (b) Doxorubicin HCl, Et₃N, DMF, r.t., 24 h, 40%. (C) Evaluation of cytotoxicity of VBA-Dox **14** incubated with tetrazine **12** (1 or 10 μM, respectively, 10 equiv. or 100 equiv.) for 72 h with HeLa cells.



VBA-Dox did show some toxicity after 3 days of incubation. As we observed more than 70% cell death with free doxorubicin at the lower concentration of 100 nM we used these conditions to test the efficiency of the VBA click-to-release strategy with tetrazine **12**.

We incubated VBA-doxorubicin **14** with 10 and 100 equivalents of tetrazine **12** with HeLa cells for 3 days. To our delight, we observed a dose-dependent toxicity of tetrazine-activated doxorubicin and an almost similar toxicity level was obtained to that when incubated with free doxorubin (Fig. 3C). Using higher amounts of tetrazine **12** also showed signs of toxicity of the tetrazine itself, so we omitted this concentration in further cell studies (Fig. SI-5†).

Conclusion

In this work we have shown that VBAs can be used in bioorthogonal click-to-release tetrazine ligations. The release was instantaneous after the tetrazine cycloaddition as no reaction intermediates were observed by NMR experiments. We assume that the slight electron-donating nature of the ether at the vinylic position of the boronic acid reduces the reaction rate as compared to VBAs lacking the ether functionality. The boronic acid moiety increases the reactivity of vinyl ethers in the click-to-release tetrazine ligation several folds when tetrazines bearing boron-coordinating ligands are used.

The use of VBA-caged prodrugs may be of interest as we have recently shown that this moiety can orthogonally react with tetrazines bearing a coordinating phenol substituent. These tetrazines are too electron rich to react with vinyl ethers.⁵ It would further be interesting to explore different substituents on the VBA cage, such as carbamates, to study the electronic effects in the click-to-release reaction with different coordinating tetrazines. These studies are currently ongoing in our laboratory.

Conflicts of interest

No conflict declared.

Acknowledgements

This research was funded by the NWO gravity program 'Institute for Chemical Immunology' NWO-024.002.009 and the Institute for Molecules and Materials at Radboud University. We thank Selina Thijssen for her pioneering work on this project and Professor Floris Rutjes for fruitful discussions.

References

- 1 E. M. Sletten and C. R. Bertozzi, *Angew. Chem., Int. Ed.*, 2009, **48**, 6974–6998.
- 2 J. Dommerholt, F. P. J. T. Rutjes and F. L. van Delft, *Top. Curr. Chem.*, 2016, **374**, 1–20.
- 3 B. L. Oliveira, Z. Guo and G. J. L. Bernardes, *Chem. Soc. Rev.*, 2017, **46**, 4895–4950.
- 4 S. Eising, F. Lelivelt and K. M. Bongers, *Angew. Chem., Int. Ed.*, 2016, **55**, 12243–12247.
- 5 S. Eising, A. H. J. Engwerda, X. Riedijk, F. M. Bickelhaupt and K. M. Bongers, *Bioconjugate Chem.*, 2018, **29**, 3054–3059.
- 6 S. Eising, B. T. Xin, F. Kleinpenning, J. Heming, B. I. Florea, H. S. Overkleeft and K. M. Bongers, *ChemBioChem*, 2018, **19**, 1648–1165.
- 7 S. Eising, N. G. A. van der Linden, F. Kleinpenning and K. M. Bongers, *Bioconjugate Chem.*, 2018, **29**, 982–986.
- 8 E. Haldón, M. C. Nicasio and P. J. Pérez, *Org. Biomol. Chem.*, 2015, **13**, 9528–9550.
- 9 R. Van Geel, G. J. M. Pruijn, F. L. Van Delft and W. C. Boelens, *Bioconjugate Chem.*, 2012, **23**, 392–398.
- 10 J. Li and P. R. Chen, *Nat. Chem. Biol.*, 2016, **12**, 129–137.
- 11 X. Ji, Z. Pan, B. Yu, L. Kimberly, D. La Cruz, Y. Zheng, B. Ke and B. Wang, *Chem. Soc. Rev.*, 2019, **48**, 1077–1094.
- 12 S. Davies, B. J. Stenton, G. J. L. Bernardes, S. Davies and B. Stenton, *Chimia*, 2018, **72**, 771–776.
- 13 R. M. Versteegen, R. Rossin, W. Ten Hoeve, H. M. Janssen and M. S. Robillard, *Angew. Chem., Int. Ed.*, 2013, **52**, 14112–14116.
- 14 X. Fan, Y. Ge, F. Lin, Y. Yang, G. Zhang, W. S. C. Ngai, Z. Lin, S. Zheng, J. Wang, J. Zhao, J. Li and P. R. Chen, *Angew. Chem., Int. Ed.*, 2016, **55**, 14046–14050.
- 15 J. C. T. Carlson, H. Mikula and R. Weissleder, *J. Am. Chem. Soc.*, 2018, **140**, 3603–3612.
- 16 R. M. Versteegen, W. Hoeve, R. Rossin, M. A. R. De Geus, H. M. Janssen and M. S. Robillard, *Angew. Chem., Int. Ed.*, 2018, **57**, 10494–10499.
- 17 J. M. M. Oneto, I. Khan, L. Seebald and M. Royzen, *ACS Cent. Sci.*, 2016, **2**, 476–482.
- 18 R. Rossin, S. M. J. Van Duijnhoven, W. Ten Hoeve, H. M. Janssen, L. H. J. Kleijn, F. J. M. Hoeben, R. M. Versteegen and M. S. Robillard, *Bioconjugate Chem.*, 2016, **27**, 1697–1706.
- 19 R. Rossin, R. M. Versteegen, J. Wu, A. Khasanov, H. J. Wessels, E. J. Steenbergen, W. Hoeve, H. M. Janssen, A. H. A. M. Van Onzen, P. J. Hudson and M. S. Robillard, *Nat. Commun.*, 2018, **9**, 1484–1494.
- 20 S. Davies, L. Qiao, B. L. Oliveira and C. D. Navo, *ChemBioChem*, 2019, **20**, 1541–1546.
- 21 I. Khan, P. F. Agris, M. V. Yigit and M. Royzen, *Chem. Commun.*, 2016, **52**, 6147–6177.
- 22 S. Davies, B. L. Oliveira and G. J. L. Bernardes, *Org. Biomol. Chem.*, 2019, **17**, 5725–5730.
- 23 Q. Yao, F. Lin, X. Fan, Y. Wang, Y. Liu, Z. Li, P. R. Chen and Y. Gao, *Nat. Commun.*, 2018, **9**, 5032.
- 24 E. Agustin, P. N. Asare Okai, I. Khan, M. R. Miller, R. Wang, J. Sheng and M. Royzen, *Chem. Commun.*, 2016, **52**, 1405–1408.



- 25 J. Li, S. Jia and P. R. Chen, *Nat. Chem. Biol.*, 2014, **10**, 1003–1005.
- 26 G. Zhang, J. Li, R. Xie, X. Fan, Y. Liu, S. Zheng, Y. Ge and P. R. Chen, *ACS Cent. Sci.*, 2016, **2**, 325–331.
- 27 J. C. T. Carlson, H. Mikula and R. Weissleder, *J. Am. Chem. Soc.*, 2018, **140**, 3603–3612.
- 28 H. Wu, S. C. Alexander, S. Jin and N. K. Devaraj, *J. Am. Chem. Soc.*, 2016, **138**, 11429–11432.
- 29 E. Jiménez-Moreno, Z. Guo, B. L. Oliveira, I. S. Albuquerque, A. Kitowski, A. Guerreiro, O. Boutureira, T. Rodrigues, G. Jiménez-Osés and G. J. L. Bernardes, *Angew. Chem., Int. Ed.*, 2017, **56**, 243–247.
- 30 K. Neumann, S. Jain, A. Gambardella, S. E. Walker, E. Valero, A. Lilienkamp and M. Bradley, *ChemBioChem*, 2017, **18**, 91–95.
- 31 J. Jiang, W. Liu, J. Cheng, L. Yang, H. Jiang, D. Bai and W. Liu, *Chem. Commun.*, 2012, **48**, 8371–8373.
- 32 D. G. Hall, *Boronic acids, Preparation and Applications in Organic Synthesis, Medicine and Materials*, WILEY-VCH, Weinheim, 2011.
- 33 C. A. Blencowe, A. T. Russell, F. Greco, W. Hayes and D. W. Thornthwaite, *Polym. Chem.*, 2011, **2**, 773–790.
- 34 P. A. Cox, A. G. Leach, A. D. Campbell, G. C. Lloyd-jones, P. A. Cox, A. G. Leach, A. D. Campbell and G. C. Lloyd-jones, *J. Am. Chem. Soc.*, 2016, **138**, 9145–9157.

