



Cite this: *Green Chem.*, 2019, **21**, 2868

Progress in understanding the four dominant intra-particle phenomena of lignocellulose pyrolysis: chemical reactions, heat transfer, mass transfer, and phase change

M. Brennan Pecha, ^{*a} Jorge Ivan Montoya Arbelaez,^b Manuel Garcia-Perez, ^c Farid Chejne ^b and Peter N. Ciesielski ^a

Four principal intra-particle phenomena occur in a highly concerted manner during the pyrolysis of lignocellulosic materials: heat transfer, mass transfer, chemical reactions, and phase changes. Achieving a holistic understanding of these processes has been challenged by their intricate coupling, high temperatures, and rapid rates at which they occur. Heat and mass transfer have been well studied at the single-particle level but their coupling with chemical reactions and phase change within single particle models remains problematic. Equally challenging is the multiscale coupling of reactor- and single particle-scale models. Too little attention has been given to phase change. Similarly, the presence of oligomeric compounds (constituting up to 20% of the oil) has not been fully accounted for in chemical reaction schemes and physical models developed for pyrolysis. Recent studies have shown that a multiscale approach is key to predictive modelling across a variety of reactor systems. Historical and recent developments are outlined in this pyrolysis review paper regarding these four intra-particle phenomena, as well as modelling efforts to capture their effect on product yields and composition. It is critical for the design of future biomass pyrolysis systems to appropriately account for all four intra-particle phenomena and their inter-connectivities in order to predict, achieve, and maintain optimal operation.

Received 15th February 2019,
Accepted 17th April 2019

DOI: 10.1039/c9gc00585d

rscl.li/greenchem

^aBiosciences Center, National Renewable Energy Laboratory, Golden, CO, USA.

E-mail: brennan.pecha@nrel.gov; Tel: +1-303-275-4817

^bFacultad de Minas, Universidad Nacional de Colombia, Medellín, Colombia

^cBiological Systems Engineering, Washington State University, Pullman, Washington, USA

Introduction

The total worldwide energy consumption in recent years is estimated to be approximately 515 EJ y⁻¹, 80% of which is cur-



M. Brennan Pecha

Dr M. Brennan Pecha is a researcher at the National Renewable Energy Laboratory. During his PhD in Chemical Engineering at Washington State University he studied fundamental heat transfer, mass transfer, chemistry, and phase change phenomena in biomass pyrolysis. His current work at NREL focuses on simulation of chemical reactions in biomass pyrolysis and catalytic systems.



Jorge Ivan Montoya Arbelaez

Dr Jorge Montoya studied Chemical Engineering at Universidad Nacional de Colombia-Medellín. He finished his MSc in chemical engineering and PhD in engineering-Energetic systems at Universidad Nacional de Colombia-Medellín. In 2015 he was at WSU for one year, working in a fundamental aspects of biomass pyrolysis. Since 2017 Jorge Montoya has been working at CELSIA-Colombia developing projects at industrial scale of cogeneration with biomass and biogas.



rently supplied by fossil fuels like coal and petroleum.^{1,2} Global energy consumption has increased exponentially over the last century due to population growth and increased per capita demands, particularly from emerging countries like Brazil, Russia, India, and China (BRIC).^{3,4} Renewable energy sources such as biomass, solar radiation, wind, water and geothermal are all potential alternatives that can be used to offset fossil energy consumption. Of these sustainable sources, biomass is the only direct source of carbon for renewable fuels and chemicals production that can be inserted into existing markets. The global biomass supply potential is estimated from 97–147 EJ in 2030.⁵ While it is feasible to use all of this for power, heating, transportation,⁵ further technical advances and investments need to be made first. Recent progress has been made in developing advanced bioproducts and biomaterials, which greatly enhance profit margins for biofuels technologies.⁶ Fast pyrolysis processes offer a promising opportunity for the rapid thermal processing of agro-industrial wastes into bio-oil. The development of this technology is relevant and appropriate with benefits over current biofuel production methods. In particular, it need not compete with food security, unlike first generation transesterification and fermentation processes commonly used to produce ethanol and biodiesel, and has the potential to be incorporated into existing petroleum refining infrastructure.^{7–12}

Pyrolysis processes require heating of biomass at temperatures between 400–600 °C in an inert or anoxic atmosphere to produce bio-oil, biochar, and non-condensable gases with high calorific value. Heat transfer, chemical reactions, phase change, and mass transport occur in concert, as illustrated in Fig. 1. Volatiles are released as both condensable compounds (bio-oil or pyrolysis oil) and non-condensable gases rich in CO, CH₄, and H₂. The remaining solid char (biochar or biocarbon) from this process is high in carbon content with a low oxygen/carbon ratio. Bio-oil is considered to be largely compatible

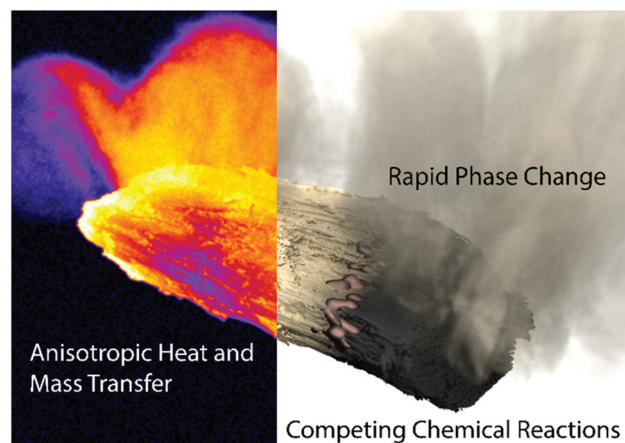


Fig. 1 Depiction of the four primary phenomena that occur during biomass pyrolysis: heat and mass transfer occur as anisotropic processes due to the aspherical geometry and directional internal porosity of biomass particles (left); these are coupled to thermally-driven chemical reactions which promote transition into molten and vapor phases (right).

when blended with vacuum gas oil in FCC units of oil refineries¹² while the solid char can be combusted to provide heat to the pyrolysis reactor and other integrated processes. Low-ash char is also attractive for metallurgical applications as a reducing agent, as a gas/water filtration medium, soil additive, *etc.*^{13–15} Without government regulation on carbon emissions, bringing char to market is currently more profitable than bio-oil,¹⁶ as argued early on by Antal *et al.*^{17,18} Non-condensable pyrolysis gas are typically combusted and recirculated into the reactor to provide heat and act as fluidization medium, or can be used in downstream processes such as catalytic methane reforming or Fisher Tropsch.^{19,20}



Manuel Garcia-Perez

Dr Manuel Garcia-Perez is a professor for the Biological Systems Engineering department at Washington State University. He has been working for the last 20 years on projects related to thermochemical conversion of lignocellulosic materials for producing bio-fuels and chemicals. Dr Garcia-Perez has studied thermochemical reactions of cellulose, hemicelluloses and lignin as well as the characterization and uses of crude bio-oils and chars.

He is currently working on the development of more selective pyrolysis reactors and on new concepts to refine pyrolysis oils. Dr Garcia-Perez is also very active on the development and characterization of engineering carbonaceous materials.



Farid Chejne

Dr Farid Chejne-Janna is an associate professor in the Department of Processes and Engineering at the Universidad Nacional de Colombia-Medellin. He is also a member of the Colombian Academy of Science. His research focuses are modeling and process simulations, analysis of energy systems, advanced thermodynamics, and energy resource evaluations.



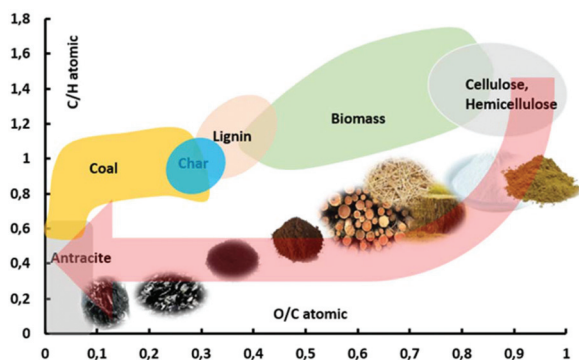


Fig. 2 Van Krevelen diagram comparing biomass and pyrolysis products to coal.

Depending on operating conditions, the pyrolysis process has historically been classified typically as slow pyrolysis, intermediate pyrolysis and fast pyrolysis. Slow pyrolysis, with particles larger than 1 cm, is used primarily for char production whereby 30–35% w/w of the carbon is converted to char with the remainder being bio-oil and non-condensable gases.²¹ The char can resemble a high-rank coal as it converts, as shown in Fig. 2 in a van Krevelen diagram.²² Fast pyrolysis, with high heating rates and small particle sizes typically 1–5 mm, is employed when the primary desired product is bio-oil which is produced at yields ranging from 50–80% yield in conjunction with minor yields of biochar (10–20% w/w) and non-condensable gas (10–20% w/w).^{11,23} In each case, yields are strongly influenced by the type of biomass, operating temperature, gas residence time in the reactor, heating rate, particle size and mineral content. High heating rates and small particle sizes are key for achieving high selectivity towards the production of bio-oil. A variety of pyrolysis technologies have been developed for producing bio-oils *via* fast pyrolysis, some of which have been put into commercial operation by entities including Ensyn, BTG, Dynamotive, GTI, KIT, VTT, and PyroTec.^{24,25}

Despite substantial research in the field of biomass pyrolysis, very few companies have succeeded in commercializing



Peter N. Ciesielski

Dr Peter Ciesielski is a Senior Scientist at the National Renewable Energy Laboratory. He has an interdisciplinary background and training in Chemical and Biological Engineering and Materials Science. Dr Ciesielski's research encompasses many aspects of bioenergy and biomaterials science and aims to improve ways by which renewable feedstocks can be used for the sustainable production of fuels, chemicals, and materials.

the technologies that produce bio-oil as the primary product, although many examples exist of companies who employ “slow” pyrolysis to produce char briquettes such as Kingsford and Weber. It has been argued that one reason for failure to scale up is a lack of understanding of the fundamental phenomena of biomass pyrolysis.²⁶ The purpose of this review is to describe the four principal intra-particle phenomena of biomass pyrolysis: chemical reactions, heat transfer, mass transfer, and phase change. The authors believe that too little attention has been given to phase change, particularly with regards to the liquid intermediate stage between native biopolymer and vaporized organic compounds. This work will have a special emphasis on recent developments in understanding this phenomenon.

Chemical reactions

Chemical composition of lignocellulosic materials

Biomass can be defined as an organic material composed primarily of a heterogeneous mixture of polymers and a small fraction of inert materials which comes from previously living plants. The organic fraction consists of polymers containing three major macromolecules: cellulose, hemicellulose, and lignin as shown in Fig. 3. In addition to these primary polymers, some biomasses contain a small quantity of lipids, pectin, and extractives, which do not exceed 10% w/w.^{11,27}

Cellulose makes up about 40 to 60% of the total biomass weight, hemicellulose 15–25%, and lignin 15–25%.^{28–30} Cellulose is a linear, mostly crystalline, polymer composed of glucose structural units linked by β -D type glycosidic bonds. These monomeric units link together to form chains of 10 000 or more structural units. Cellulose polymers form fibers inside biomass, and are responsible for the fibrous nature of cell walls. Hemicelluloses are polysaccharides with an amorphous, branched polymeric structure and a substantially lower degree of polymerization than cellulose, approximately 100, primarily of the following types: xylans, homoxylylans, glucuronoxylans, arabinoxylans, glucuronoarabinoxylans, mannans, and others.³¹ It is primarily composed of sugars with 5 and 6 carbons per unit like xylose, mannose, arabinose, galactose and glucose. Angiosperm species have mostly xylan and gymnosperm species have mostly glucomannan based hemicellulose.³² Lignin is a complex, branched, non-crystalline macromolecule, composed of three primary aromatic monomeric constituents: sinapyl, coniferyl, and coumaryl alcohols linked by β -ether linkages and other variety of C–C and C–O–C bonds exist between monomer units of lignin as β -O-4, α -O-4, β -5, 5–5, 4-O-5, β -1, and β - β bonds.^{29,33} Both lignin and hemicellulose are responsible for binding cellulose fibers together, maintaining the structure of the cell walls.^{33,34}

Less-discussed are the “extractives” and mineral matter in biomass. Extractives include chemical families such as turpentine, rosin, tall oil fatty acids, tanning materials, camphor, volatile oils, gums, and rubber.³⁵ Huge industries have historically been built around these high-value chemicals. Extractives



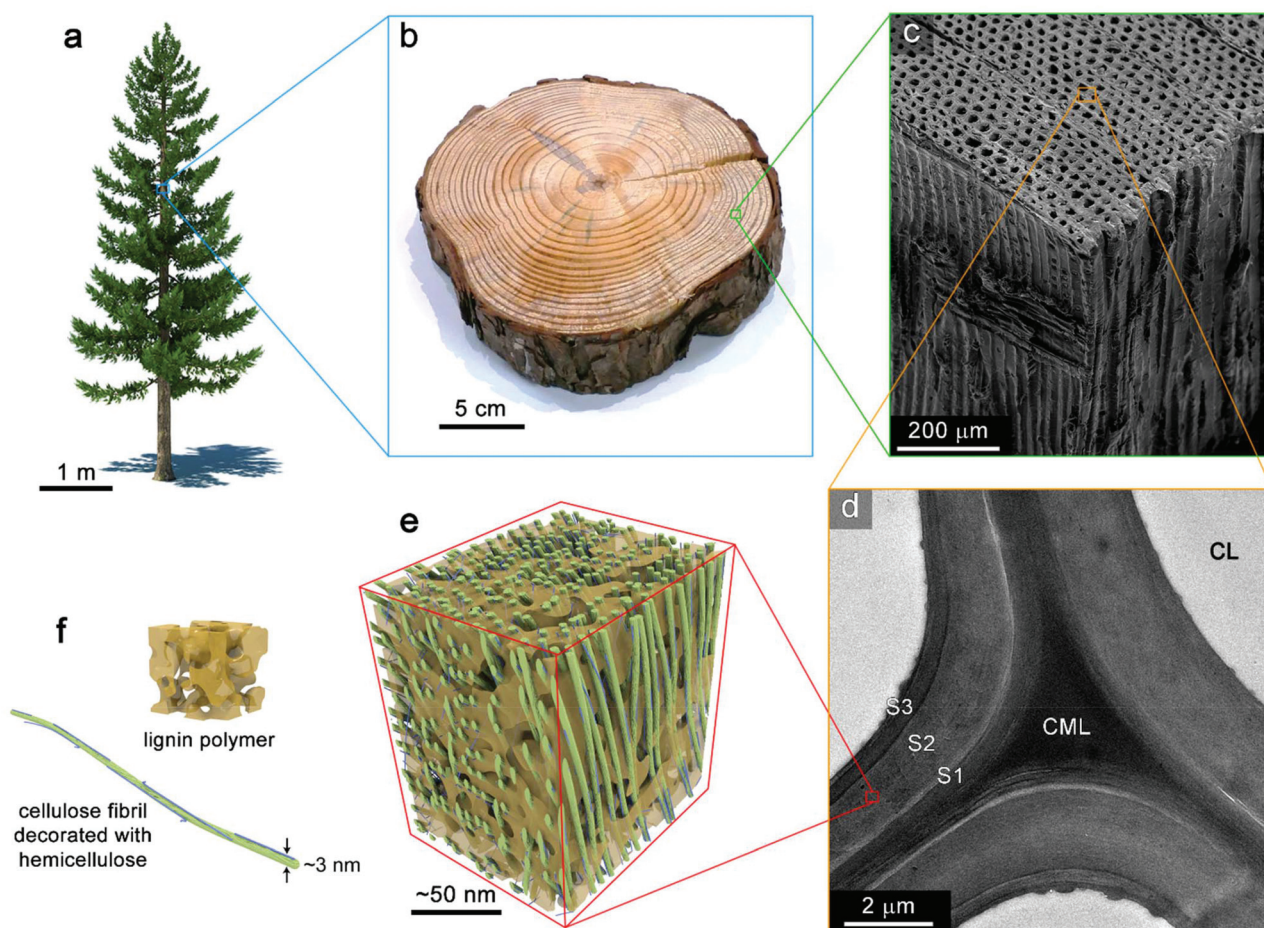


Fig. 3 Hierarchical structure of wood. (a) Coniferous tree. (b) Photograph of trunk section from a pine tree. (c) SEM image of the tissue structure soft-wood (yellow pine). (d) TEM image showing the ultrastructure wood cell wall. CL, cell lumen; CML, compound middle lamella; S1, S2, and S3 denote layers of the secondary cell wall. (e) Schematic of the nanoscale architecture of lignocellulose. (f) Idealized depiction of an amorphous lignin polymer and a cellulose elementary fibril decorated with hemicellulose (microfibril). Reprinted (adapted) with permission from Zhu *et al.*³⁹ Copyright (2016) American Chemical Society.

are more abundant in oily biomass species like olive pomace and nut shells than common wood species like pine and poplar, which typically less than 2 wt%. Metal matter content depends on the type of biomass, but comprises 2–25% of the total solid weight and commonly consist of minerals such as Si, Na, K, Ca, Mg, Mn, Co, Zn, and Cu. These minerals are present as like oxides and salts such as chlorides, carbonates, phosphates and sulphates.^{36–38} Mineral matter plays an important role in thermal decomposition of biomass, acting as catalyst for critical reactions, such as dehydration and catalytic cracking of the volatiles, though Si is not considered a catalytic species.

The complex, hierarchical structure of biomass visualized in Fig. 3 originates from tissue structure of the once-living plant. This structure is highly specific to the plant species of origin and can vary dramatically. At the nanoscale, the lignocellulosic material comprising plant cell walls is an impressive, highly-ordered assembly of biopolymeric components. Bundles of several 18–24 chains of cellulose form elementary cellulose fibrils which are often further aggregated into macro-

fibrils. These are decorated and crosslinked by hemicelluloses. Lignin acts as matrixing material which provides additional structural support, defence from fungal and microbial attack, moisture control. This nanocomposite material forms the cell walls of plant tissue. Assemblies of cells make up the microstructure of plants in highly directional pores that transport water and nutrients throughout the organism during its lifetime. The anisotropic transport of heat and mass throughout biomass during pyrolysis results from this microstructure which is retained within biomass particles after comminution processes.

Biopolymer breakdown at pyrolysis temperatures: studies on isolated constituents

The compounds present in pyrolysis oil are derived from the biopolymers present in the biomass. Thus, the composition of bio-oil is highly dependent on the proportions and characteristics of cellulose, lignin, hemicellulose, and extractives as well as interactions between these moieties inside biomass solid matrix.^{40–43} This variety of biopolymers causes thermal



decomposition to progress in several stages. Initially, hemicellulose breaks down at temperatures between 200–300 °C. This is followed by the degradation of cellulose, which decomposes between 300 and 450 °C. Lignin decomposition occurs across the full temperature range of 250–500 °C, but is the last polymer to fully degrade. Lignin is the major contributor to char production at the end of pyrolysis.^{43–46}

Cellulose has been by far the most studied component of biomass. The position and orientation of glucan linkages do not have a significant effect on levoglucosan yields, which is the chief monomeric compound obtained from the devolatilization of cellulose.⁴⁷ Other fundamental work demonstrated that cellulose that has a high degrees of polymerization (DP) produced more levoglucosan and less char than cellulose with lower DP.⁴⁸ The regions under maximum stress, and therefore the most likely to fracture during pyrolysis, are precisely those that are located at the interface of the amorphous and crystalline structures.⁴⁹ It has also been illustrated that the maximum temperature required to achieve cellulose decomposition is higher for crystalline cellulose structures than for amorphous cellulose due to the extra energy required to disrupt the crystal lattice structure.^{50,51} As cellulose degrades, the chains continue to break down in a continuous fashion, passing through oligomeric compounds (two or more glucan units) all the way to monomeric levoglucosan.⁵² Cellulose pyrolysis has been recently reviewed in detail for further reading.⁵³

Due to the complex structure of lignin within biomass, the final structure of isolated lignins shows significant differences in molecular weight and aromatic structures distribution based on the separation methods employed and biomass used.^{54,55} Product yields have been found to vary substantially depending on the type of lignin used for pyrolysis.^{56,57} Saka *et al.*⁵⁷ reported that lignins from softwoods tend to pyrolyze at lower temperatures and produce more volatiles and less char than lignins derived from hardwoods. This observation is attributed to the molecular structure of hardwood lignins, which are more complex, and have a high molecular weight than softwood lignins, providing greater resistance to thermal decomposition. Unfortunately, there are very few studies regarding the effect of the type and nature of lignins on the product distribution during pyrolysis. Further review on lignin pyrolysis studies can be found elsewhere.⁵⁸

Pyrolysis of isolated hemicelluloses has been studied even less than lignin. This is largely because of the difficulty associated with extracting well-preserved hemicelluloses. Nevertheless, some studies have been performed on isolated polysaccharides similar to hemicelluloses, with primary products being acetic acid, furfural, formic acid, furfural, 5-hydroxymethyl furfural, and neutral sugars, as reviewed in detail by Zhou *et al.*³¹ Furthermore, studies using xylan, one of many types of hemicellulose, are often unreliable because commercial xylan is up to 10 wt% potassium (a catalyst for gas production).⁵⁹ At temperatures between 150 and 240 °C, the methoxy and acetyl bonds of hemicellulose dehydrate.⁶⁰ Between 240 and 350 °C, the glycosidic bonds break, similar to cellulose.^{60–62} Char formation reactions in xylan dominate above 300 °C.^{61,62}

Further reading on studies to understand the chemistry of biomass pyrolysis can be found by Wang *et al.*⁶³ and Collard *et al.*⁵³

Catalytic effect of metal content native to biomass

Biomass contains traces of mineral material (K, Na, Ca, Zn, Cu, P, Mg, Mn), usually in the form of oxides or salts such as chlorides, sulphates, carbonates and phosphates.^{36,38,64–66} The proportions of these minerals depends on the type of biomass, as well as collection, drying, and grinding conditions. The content of inorganic material affects product yields and is an important parameter for the study of side reactions; generally, as the mineral material content increases, bio-oil yields decrease and gas and char production increase.^{11,25,30,37,44,67–69} Changes in product composition are largely due to the catalytic effect of the mineral matter on several reaction pathways including the dehydration and decarboxylation reactions.⁷⁰ Alkalis strongly interact with functional groups such as –COOH and –OH, forming alkali-oxygen clusters that promote cracking reactions.⁶⁶ The common metals with the greatest impact on volatile yields were, in descending impact: K⁺, Na⁺, Ca²⁺, Mg²⁺. Low concentrations, *e.g.* 0.5% w/w, increase char yields by up to 10% and increase the yield of low molecular weight species such as acetol, glycol aldehyde, and formic acid while the anhydrosugar (*e.g.* levoglucosan) yield decreases.^{65,70} Although some studies differ on impacts of various metals, consensus is that potassium is the biggest offender.^{36,38,64,71–73}

Additional effects on reaction pathways have also been observed; for example, sodium restricts cellulose and hemicellulose transglycosylation reactions but facilitates demethoxylation, demethylation, and dehydration of lignin.⁷⁴ The presence of silicon has also been found to decrease the production of volatiles.⁷² In contrast to the effects of alkali and alkaline metals, adding anions (chloride, nitrates, sulfates) has been found to increase levoglucosan production and decrease char yields.⁷³ In recent years, numerous works have been published that examine the effect of mineral matter on final product distribution.^{36,38,65,75–77} Despite these efforts, very few theoretical models for the quantitative prediction of the effects of inorganic material on the product distribution of fast pyrolysis processes are available.^{78,79}

The presence of mineral matter has also been found to affect morphological changes during biomass pyrolysis. Iwasaki *et al.*⁸⁰ showed that a possible cause of particles agglomeration inside fluidized bed reactors is the formation of an intermediate liquid phase during devolatilization, that covers all particles inside the fluidized bed and promotes agglomeration. The formation of this liquid phase is favored for biomasses that have been pretreated with acids. Visualization of inert material particles from fluidized beds revealed a coating which acts to promote particle agglomeration. The formation of this film is most evident for biomasses that were previously pre-treated with organic acids.⁸¹ Oudenhoven *et al.*⁸¹ have shown that part of the solid residue left by biomasses treated with acid has lost its original structure, forming new structures similar to a melted polymer. Cavities were observed that presumably formed when bubbles rupture



inside the liquid phase. Recent work has showed the corollary result that materials that still have mineral content tend to maintain their original tissue structure after pyrolysis.⁸²

Effect of reaction temperature on chemical reactions

The devolatilization process is sequential and depends on the temperature reached by the particle. The first stage occurs between 20–120 °C and corresponds to water evaporation from the biomass. The second stage between 120–300 °C shows no appreciable biomass weight loss, though small quantities of some gases, such as CO, CO₂, and steam are released from dehydration reactions and decarboxylation of R–CO–R groups belonging to hemicellulose and lignin.^{29,83,84} Between 250 and 370 °C,^{50,85–88} cellulose undergoes endothermic reactions and becomes plasticized, often called “active cellulose”, wherein the amorphous parts of the cellulose degrade and expose reducing ends of cellulose chains.⁵¹ Above 300 °C, an intermediate liquid phase known by many researchers as “molten cellulose”, or “intermediary liquid compound” due to formation of anhydrous oligomers that have low vapor pressure at these temperatures.^{87,89,90} Between 300 and 400 °C, about 80% weight loss can occur without mass or heat transfer limitations.⁸⁷ In this stage, random fragmentation of glycosidic bonds in cellulose, hemicellulose and their oligomers generate volatile compounds with high oxygen content, leaving a carbonaceous residue known as char or biochar.^{23,91,92} At temperatures above 400 °C, CO and CO₂ are released by depolymerization reactions from the lignin-rich aromatic carbonaceous matrix.

Fig. 4 provides one example of a large particle in which temperature was measured *in situ*. Though spherical particles are not representative of industrially-relevant biomass feed-

stocks,⁹³ the thermal pathway is still characteristic of wood. Here it can be seen that endothermic phenomena occur around 400–500 °C and exothermic phenomena dominate between 600 and 800 °C. The first event around 630 K (357 °C) is associated with the pyrolysis of cellulose and the evaporation of the products of this reaction. The exothermic phenomena which occur at higher temperatures are due to the release of heat by polycondensation reactions.^{94,95} “Thermal runaway” is notable in larger particles, where conductive heat transfer is slow compared to exothermic secondary reactions of slowly diffusing products.⁹⁶

The effect of temperature on bio-oil production is well understood and has been explored in the literature. It has been shown that there is a parabolic behavior for bio-oil production with temperature wherein the maximum bio-oil yield is obtained with reactor temperatures between 400 and 550 °C, depending on the type and size of biomass and heat transfer mechanism.^{10,36,80,97–100} The maximum yields of lignin-derived oligomers has been reported between 450 and 500 °C;¹⁰¹ this reflects the balance between primary reactions, which release oligomeric products from biomass, and secondary reactions which degrade these products into monomers and eventually small organics and gas above 500 °C.¹⁰²

As temperature increases, there is less char. However, at higher temperatures above 500 °C, volatile cracking reactions reduce the yield of bio-oil and increase the production of non-condensable gases.^{103–106} Due to this phenomenon, many studies show that maximum production of bio-oil in systems with good heat transfer and small particles is achieved 450 °C, with yields of up to 80% for cellulose but more commonly 60% for wood.^{107–110}

The *composition* of the resultant oil also changes with temperature.¹¹² At low temperatures (<300 °C), most volatiles orig-

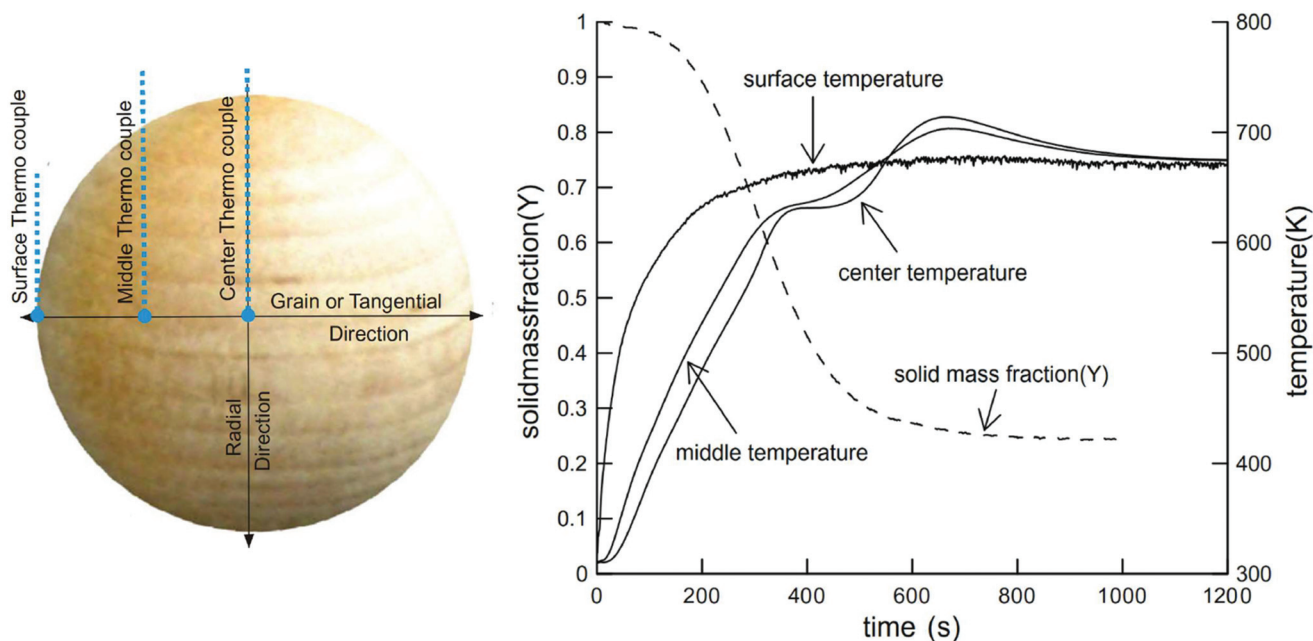


Fig. 4 Solid mass fraction and temperature of wood sphere pyrolysis at 688 K. Reprinted from Park *et al.*,¹¹¹ Copyright (2010), with permission from Elsevier.



inate from hemicellulose degradation and the scission of weak cellulose linkages, with the main products levoglucosan, levoglucosone, hydroxymethyl furfural, acetic acid, acetone, guaiacyl acetone, glyoxal, methanol and formic acid. Fu *et al.*¹¹³ demonstrated the formation of short chain vapors and gases, including: formic acid, methanol, ethane, ethylene, formaldehyde, acetone, hydrogen cyanide, carbon monoxide and carbon dioxide, during pyrolysis of cotton stalk, corn and rice, at a temperature range of 300–400 °C. At temperatures above 400 °C, intact rings such as benzene, carboxylic acids, phenols, p-cresol, furans, furfural, and sugars are the most abundant species.³⁶ As the temperature increases further, the yield of these volatile compounds decreases and the yield of more thermally stable polyhydroxyl aromatic compounds (PHA) increases. Between 580–800 °C, new aromatic compounds such as benzene, naphthalene, cresol, and toluene can be formed.^{37,83,114} Chaiwat *et al.*¹¹⁵ proposed a set of competitive reactions in cellulose depolymerization and dehydration, as shown in Fig. 5. At low temperatures and low heating rates, strong intra-chain hydrogen bonds of cellulose functional groups, increasing the probability of dehydration reactions. At high heating rates, intra-chain hydrogen bonds weaken and decrease the possibility of collisions that facilitate dehydration reactions.¹¹⁶

Lignin oligomers are lignin derived compounds with two or more aromatic rings, with molecular weight greater than 180 g mol⁻¹. Garcia-Perez *et al.*¹⁰¹ found that the lignin oligomer yields increase with temperature until 500 °C. At temperatures higher than 500 °C, decomposition reactions for aerosols, which contain these oligomers, become more intense, generating secondary lower molecular weight volatiles. Westerhof *et al.*¹⁰⁰ reported an increase in lignin oligomer yields between 380–480 °C for pine pyrolysis, attributing this behavior to increased biomass conversion. Above 480 °C, oligomer yields decreased presumably due to secondary vapor cracking reactions.¹¹⁷ It is also thought that the particle size reduces the yield of large compounds like lignin oligomers due to mass transfer issues. In one recent publication, the yield of pyrolytic lignin increased as particle size

decreased; the authors hypothesized that this is due to collision of aerosol droplets with the cell walls after they are ejected,¹¹⁸ as will be discussed in the next subsection.

Despite the wide assortment of studies conducted to evaluate the effect of temperature on the yield of char, bio-oil and non-condensable gases, there is ambiguity in the definition of the optimum temperature of pyrolysis. It is not typically possible to directly measure the temperature of very small biomass particles, and so it is often assumed to be equal to the reactor temperature^{84,85} or in some cases is estimated by mathematical modelling,^{86,119} with some exceptions when using large particles, as illustrated in Fig. 4.¹¹¹ For slow pyrolysis conditions utilizing small particle sizes (less than 1 mm), the assumption that the biomass particle temperature is equal to the reactor temperature is valid due to longer heating times, which allows thermal equilibrium between the biomass particles and surroundings to be achieved.^{89,104} However, these assumptions are often far from reality for fast pyrolysis conditions, resulting in mismatches (“thermal lag”) of temperatures of 100 °C or more. This thermal lag causes errors in estimating the actual particle temperature, which can result in over-estimates of kinetic parameters such as the activation energy and the coefficient of the reaction rate.^{104,105}

There are many reports in the literature that point out the importance of heating rate effects on bio-oil and char yields. Onay *et al.*¹²⁰ studied the fast pyrolysis of safflower seeds at heating rates of 100, 300 and 800 °C min⁻¹. Results showed that above 300 °C min⁻¹, maximum yields of bio-oil (55%) and char (17%) were achieved. Thangalazhy-Gopakumar *et al.*¹²¹ pyrolyzed pine and grass fodder at reactor heating rates of 50, 100, 500, 1000, 2000 °C s⁻¹ and did not find an impact of heating rate due to slow heat transfer into the biomass. This highlights the importance of accurately measuring temperature in basic studies.¹²²

As with temperature, the heating rate plays an important role on the quality of bio-oil and char structure obtained. Bio-oils can be obtained with lower moisture content at higher heating rates, primarily due to the inhibition of secondary volatile dehydration and cracking reactions.¹²³ Higher heating rates also reduce the formation of the water-soluble fractions (formic acid, methanol, acetic acid) and aromatic fractions (rich in phenol and its derivatives). In contrast, CO and CO₂ yield increase with higher heating rates.⁶⁹ Ketones, levoglucosan, phenol and toluene yields also increase with heating rate for the fast pyrolysis of pine wood and grasses when heating rates exceed 50 °C s⁻¹.

Chemical reaction schemes to describe the effect of temperature and composition

Due to the complex physicochemical nature of biomass pyrolysis, there are a wide array of reaction mechanisms and products associated with thermal decomposition. The search for these reaction mechanisms is a developing science; new results are frequently published, with sophisticated experimental equipment, new analytical techniques, and computational tools to elucidate the evolution of species and thereby kinetics of devolatilization processes. The reaction mechanisms and rate expressions are the heart of the analysis of

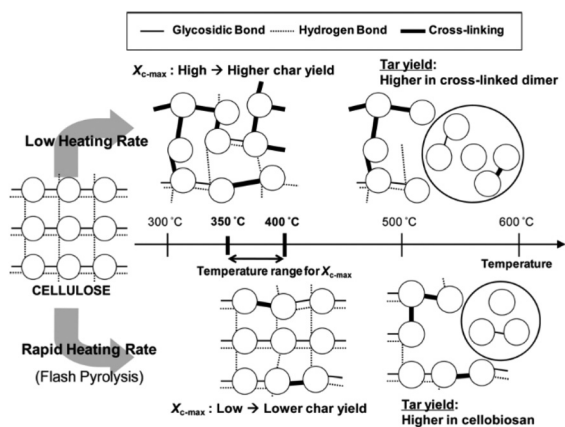


Fig. 5 Effect of the heating rate on the mechanism of thermal decomposition of the cellulose. Reprinted (adapted) with permission; Chaiwat *et al.*¹¹⁵ Copyright (2009) American Chemical Society.



pyrolysis processes.^{23,122} Most models are lumped schemes with global products obtained from biomass (total volatiles, total light gases, char) and simultaneous multi-stage and/or competitive reactions. These models can be classified under the following categories: global models in a single step, global models in multiple steps, semi-global models, and models focused on the formation of volatile species.

Global lumped models in a single step. In single step, global lumped models, it is assumed that all the products are directly released from the biomass in a single reaction stage:^{124,125}



The expressions for the reaction rate are represented as a function of temperature and biomass conversion (see eqn (2) and (3)) according to the mathematical expressions:

$$\frac{d\alpha}{dt} = k(T)f(\alpha) \quad (2)$$

$$\alpha = \frac{m(t) - m_0}{m_\infty - m_0} \quad (3)$$

And where da/dt represents the rate of decomposition of biomass, t represents the reaction time, α represents the biomass conversion, T represents the temperature of the sample, $f(\alpha)$ represents the expression for the conversion function, $m(t)$ represents the weight of biomass at a given time t . For non-isothermal systems such as thermogravimetric processes, the mathematical expression of the reaction rate may

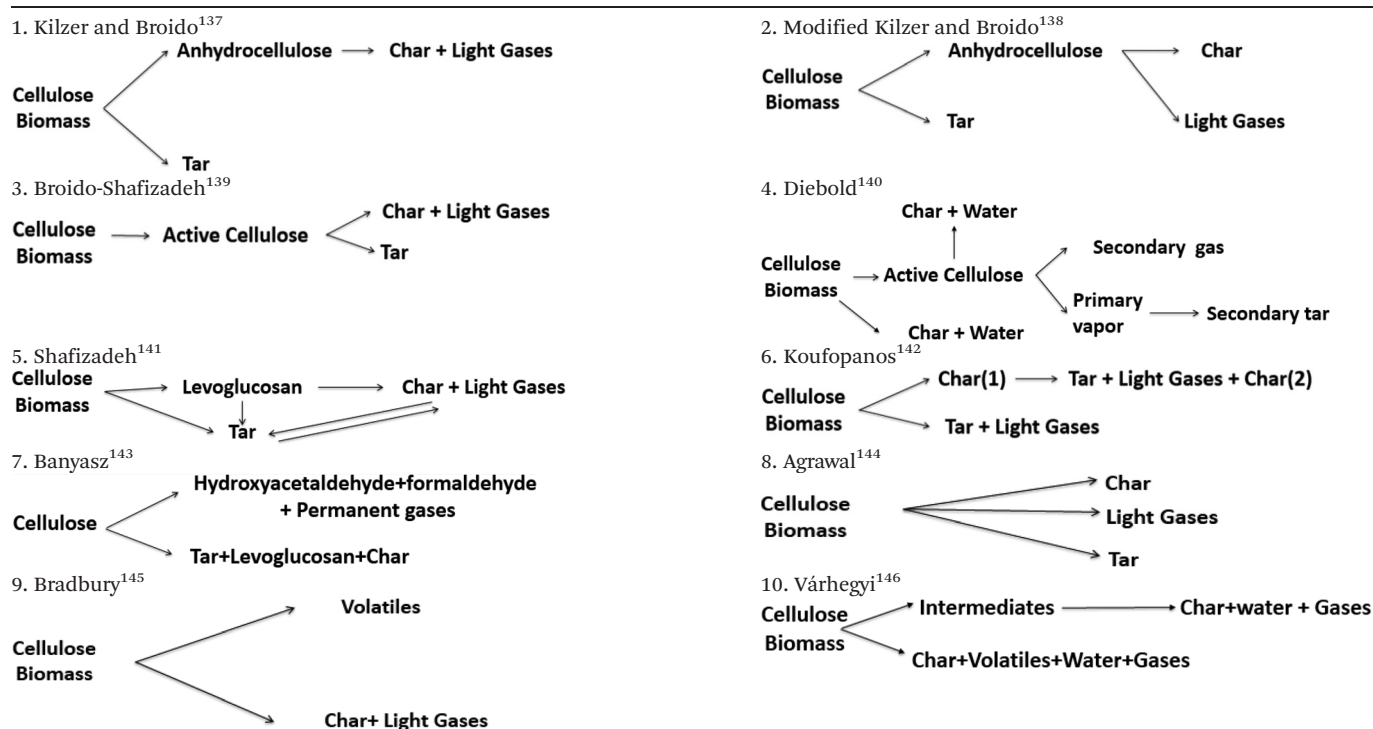
be expressed as a function of temperature such that the reaction rate can be expressed as:

$$\frac{d\alpha}{dt} = \frac{d\alpha}{dT} \frac{dT}{dt} = \frac{1}{\beta} k(T)f(\alpha) \quad (4)$$

Grønli,¹²⁵ Orfão,¹²⁴ Várhegyi,^{126,127} Antal,¹²⁸ and others have extensively used this reaction scheme, which can accurately represent the thermogravimetric curves without byzantine data-fitting algorithms. However, because of the simplicity of the physical model used, this type of model does not allow prediction of the individual species generated in devolatilization. The reaction rate is monitored by following the change in the weight of the solid residue with time, which is usually done using a thermobalance; this limits most knowledge to slow-heating conditions which are not comprehensive enough to represent fast pyrolysis processes.¹²⁹

Multi stage-global lumped models. Many researchers have proposed a variety of more complex reaction schemes with multiple stages that monitor the yield of all the macro-products involved in the overall reaction (condensable volatile tar, light gases, char, and biomass).⁷⁶ Table 1 summarizes schemes commonly used to describe the decomposition of biomass in multiple stages. Some models contain an "active" intermediate that approximates depolymerization and the presence of a liquid intermediate phase.^{9,85,87,110,122,130–132} Schemes 1, 2, 4, 5, 6, 7, 9 and 10 have two competing reactions in the first stage. One of these reactions is associated with dehydration of the cellulose to form anhydro-cellulose and water which can then react again to produce char and non-condensa-

Table 1 Multistage mechanisms for biomass pyrolysis



ble gases according to the kinetic schemes (1, 2, 4, 5, and 10). The second reaction involves the depolymerization of cellulose to produce anhydrous tar that is rich in levoglucosan. Though useful, with these models it is not possible to explain the changes in oil composition. However science is established that dehydration reactions are favored at low heating rates and temperatures below 300 °C, increasing the production of char and light gases, whereas at higher temperatures (300–500 °C) and high heating rates depolymerization reactions are promoted that increase the yield of condensable volatiles.^{23,129,133}

Models 3, 4, 5, and 10 have the novel inclusion of an intermediate plastic/liquid phase.⁸⁷ The presence of this phase explains the “Lag” in biomass mass loss at temperatures below 320 °C when evaluating the experimental results obtained by thermobalance.^{134,135} These schemes have been widely used by many researchers studying biomass slow and fast pyrolysis.^{131,133,136} Alexander⁶⁷ compared five different reaction schemes for slow and fast pyrolysis of Loblolly pine and found that the kinetic parameters obtained using low and high heating rates are distinct and not interchangeable; *i.e.* kinetic parameters obtained at low heating rates (5–20 K min⁻¹) do not explain behavior at high rates heating (400 °C s⁻¹). These results corroborate those presented by Milosavljevic.¹²⁹

Di Blasi¹³⁵ presented the results of a comparative study of most models presented in Table 1 (models 1, 3, 4, 6, 8) for cellulose and biomass slow pyrolysis at heating rates of between 20 and 50 K min⁻¹ at a temperature range between 550 and 1000 K. The primary findings of this study were:

- For cellulose, all schemes have the same trend: volatile species and char increase and non-condensable gases decrease with higher heating rate and temperature.
- Models 1 and 3 predict similar volatile yields, while model 4 under predicts volatile yields.
- For biomass, from the qualitative point of view, there are large differences in data trends, especially in the yield of non-condensable gases.
- Models 6 and 8 predict that char yield remained constant as the heating rate increases from 40 to 160 K min⁻¹. The authors suggest that at high temperatures, the reaction is thermally controlled, and no kinetic control exists. The primary reactions responsible for the formation of condensable volatile compounds are strongly endothermic, with part of the energy supplied to the reaction used to evaporate the tar condensate (liquid phase within the biomass), cooling the particle as the vapors escape.

All kinetic models obtained for both cellulose and biomass, were developed using data collected under different particle sizes and temperature ranges and different process equipment, including fluidized bed, fixed-bed thermobalance, and pyroprobe reactors. This means that there is subjectivity in each model since, in some, kinetic control could exist and not others, or the experimental conditions are tightly controlled by some teams, minimizing the risk of uncontrolled secondary effects.

In summary, the study of the reaction rate expressions within these models is based on monitoring global yields, such as the total weight of the solid residue char (char + unprocessed biomass + liquid phase), total volatiles (obtained

by condensing the total gas), and light, non-condensable gases (gas chromatography). These models do not predict the evolution of species released during devolatilization, and many of them contain high degrees of freedom for the estimate the kinetic parameters (more variables than experimental data), which limits their predictive utility for systems and feedstocks other than those from which they were obtained. This topic has been reviewed elsewhere in the literature.^{68,147}

Semi-global lumped models. Semi-global models describe the reaction rate of biomass as a linear combination of reaction rates for each of its components (cellulose, hemicellulose and lignin), see eqn (5). For each constituent, models have been adopted that are analogous to the reactions outlined in Table 1.

$$r_{\text{biomass}} = x_{\text{cellulose}}r_{\text{cellulose}} + x_{\text{hemicellulose}}r_{\text{hemicellulose}} + x_{\text{lignin}}r_{\text{lignin}} \quad (5)$$

where r_i and x_i are the reaction rate and the weight fraction of species i (cellulose, hemicellulose, lignin). This type of model has generated substantial controversy. While many authors claim that there is no interaction between biomass components and therefore valid to model the overall biomass pyrolysis behavior,^{142,148–150} others argue that it is virtually impossible to achieve the same structure and pattern of actual biomass interactions with simple mixtures of components.^{40,43,151,152}

Yang *et al.*¹⁴⁸ found that synthetic biomass pyrolysis can be explained as the sum of the decomposition of each component separately and that this behavior is similar that of real biomass. Rao *et al.*¹⁵⁰ proposed a kinetic model of order n to assess the behavior of cellulose, hazel, wood, rice husks and olive husk under slow pyrolysis conditions (20 K min⁻¹), using predefined kinetics parameters for cellulose, hemicellulose and lignin, finding good comparison between the model and experiments. Oyedun *et al.*¹⁵³ adopted a two-stage model (model 7 Table 1) to explain the behavior of each biomass component during bamboo pyrolysis. The results showed that this model represented the experimental data well. The activation energy obtained varied from 33 to 144 kJ mol⁻¹ for the formation of volatiles and 60–91 kJ mol⁻¹ for the formation of char for each component.

Hosoya *et al.*⁴⁰ concluded that cellulose–hemicellulose interactions are negligible but that interactions between cellulose and lignin enhanced the formation of guaiacol, 4-methyl-guaiacol, 4-vinyl-guaiacol, and light gases such as CO and CO₂ while decreasing the formation of char. Couhert *et al.*⁴³ obtained similar results, concluding that it is not possible to represent the behavior of a biomass as the sum of components. The authors suggest that the type of contact between constituents is very important in raw biomass, since there are components that are inter-linked by covalent and non-covalent intermolecular forces that hold them together. The complex arrangement of biopolymers and their interactions established during biosynthesis are not adequately represented by a simple mixing components, but may reflect general degradation patterns during thermal devolatilization.

Williams⁶⁷ concluded that a two-component two-stage model (model 7, Table 1) is not a suitable representation for



loblolly pine pyrolysis. As mentioned in the previous section, there is uncertainty regarding the use of additive laws to represent the behavior of whole biomass from its components because of arguments regarding the arrangement and potential interaction of each component in raw biomass. In most of these models, only global components such as total volatiles, char, and light gases are used to describe the evolution of species or of the condensed intermediate phase. Experiments used to develop such models are usually restricted to the use of a thermobalance to measure the experimental conditions. The next subsection describes kinetic schemes developed with chemical speciation.

Kinetic schemes with chemical speciation. Some models allow for targeted species present in the volatile and non-condensable gases to be determined as a function of the operating parameters (temperature, heating rate). Examples of this type of model are limited in the literature⁴⁴ and many that do exist are simply extensions of the models described in Table 1. Only a few studies have been published on the evolution of species from cellulose and glucose derivatives from the perspective of micro kinetic mechanism.¹⁵⁵ Ranzi *et al.*^{44,156} proposed a multicomponent kinetic model that takes into account changes in the primary gas in the condensable and non-condensable gas fractions, such as CO, CO₂, H₂, CH₄, H₂O, HMF, levoglucosan, phenol, ethanol and methanol. The model was validated using results from various biomass pyrolysis tests conducted in a fluidized bed reactor. Despite being very descriptive, it does not accurately predict the behavior of levoglucosan, HMF, ethanol and other species, especially at elevated temperatures. In addition, Ranzi *et al.* do not report the experimental conditions under which the model was developed, and claim that it is only a first approximation, with more work needed to improve reproducibility.¹⁵⁷ Anca-Couce extended the Ranzi scheme to account for high charring biomass species like those with catalytic ash content and a trapped gas mechanism to represent delayed release of these species.^{79,158} The authors advise using semi-empirical “X factors” in the mechanism to account for the impact of ash, such that high charring materials have higher X values than low charring materials as shown in Table 2. A similar approach was used by Ranzi *et al.*, 2017.⁷⁸

Liao *et al.*¹⁵⁹ created a modified Broido-Shafizadeh model that considers the evolution of levoglucosan, acetol, 5-HMF, HAA, CO, CO₂, H₂, H₂O, and furfural. Although the model includes the formation of an intermediate phase (activated cellulose), this phase is not directly monitored but is fit implicitly from experimental data. Other similar schemes may be found in the literature.^{160,161} Vinu *et al.*¹⁵⁵ developed a microkinetic model for pyrolysis of glucose-based carbohydrates. The scheme is rigorous, involving 41 elementary reactions, including depolymerization, cracking, drying, aldol condensation, and tautomerization reactions among others. This model describes the step by step the evolution of the primary volatile condensable and non-condensable gases, as well as the evolution of char and intermediate anhydrosugars. Yanez *et al.*¹⁶² recently published a detailed lignin pyrolysis microkinetic model based on advanced lignin modelling that has 1615 species and 4313 reactions.

Some commercial packages have also been developed to explain the evolution of species during biomass devolatilization. However these models remain “black boxes” which do not disclose the underlying theory and formulation of their implementation: Prime Kinetics Model (Reaction Network), Bio-Flashing,¹⁶³ Bio-FG-VDC (Functional Group-Depolymerization-Volatilization-Crosslinking).

Distribution activation energy model (DAEM). Another kind of kinetic model that has been originally developed for coal is the distribution of activation energies model (DAEM).^{164–171} These schemes are often multi-stage and multicomponent which involve the formation of a species from a very large number of independent parallel reactions that reflect variations in bond cleavage and reactant structures. These models have been used to explain data obtained under slow pyrolysis conditions using a thermobalance,^{164,169,170,172–176} but so far few applications have been reported for fast pyrolysis of biomass.

Usually in the study of the effect of heating rate on the overall yield of volatiles, char and non-condensable gases, there are very few models that extend to the study of individual chemical species of both condensable and non-condensable gases.¹⁷⁷ To achieve this level of detail in the analysis of product species, integral kinetic models, such as the one given in eqn (6) are employed. The main advantage of this model is that it covers a wide range of experimental conditions, including temperatures and heating rates, and can represent slow and fast heating kinetics in one kinetics scheme. The activation energy distribution can be represented by continuous functions as normal, lognormal and Weibull distributions.

$$\frac{V^* - V}{V^*} = \frac{1}{\sigma\sqrt{2\pi}} \int_{-\infty}^{\infty} \exp\left[-A \int_0^t \exp\left(\frac{-E}{RT(t)}\right) dt\right] \left[\frac{(E - E_0)^2}{2\sigma^2}\right] dE \quad (6)$$

Here V^* and V represent the maximum amount of volatile or individual species in the gas or vapor phase at a given time t , A is the rate constant, E is the activation energy, σ is the variance and E_0 is the mean of the statistical distribution of activation energies. The kinetic parameters can be obtained by regression methods using numerical methods to solve the exponential integral^{170,174,178} or with differential methods as proposed by Miura, *et al.*^{179,180} This type of model is becoming widely recognized as the most accurate available to correlate the experimental data from biomass pyrolysis. These models avoid underestimating the activation energy compared to single stage reaction schemes, but the most striking feature of this scheme is that a mathematical model is derived that is applicable at various heating rates, unlike either single or multi stage that only apply to a single heating rate.¹⁸¹ The difficulty with this technique is that the right side integral has no analytic solution, so approximations or numerical methods must be used to obtain a solution.

Secondary reactions

Secondary reactions convert primary products into char and gas. These reactions can occur in the gas phase of the reactor,



Table 2 Kinetic scheme proposed by Anca-Couce *et al.*⁷⁹ based on the Ranzi scheme⁴⁴ to predict condensable and non-condensable gas yields from cellulose, hemicellulose, and lignin pyrolysis in native lignocellulose. Extractives kinetics from Debiagi *et al.*¹⁵⁴

Reaction	$A [s^{-1}]$	$E [kJ mol^{-1}]$	$\Delta h [kJ g^{-1}]$
Cellulose scheme			
1 CELL \rightarrow CELLA		188.37	0.0
2 CELLA $\rightarrow (1 - x_{CELL}) \times (0.45 \text{ HAA} + 0.2 \text{ GLYOX} + 0.3 \text{ C}_3\text{H}_6\text{O} + 0.25 \text{ HMFU} + 0.05 \text{ H}_2 + 0.31 \text{ CO} + 0.41 \text{ CO}_2 + 0.4 \text{ CH}_2\text{O} + 0.15 \text{ CH}_3\text{OH} + 0.1 \text{ CH}_3\text{CHO} + 0.83 \text{ H}_2\text{O} + 0.02 \text{ HCOOH} + 0.05 \text{ G}\{\text{H}_2\} + 0.2 \text{ G}\{\text{CH}_4\} + 0.61 \text{ Char}) + x_{CELL} \times (5.5 \text{ Char} + 4 \text{ H}_2\text{O} + 0.5 \text{ CO}_2 + \text{H}_2)$	4×10^{13}	80.0	$0.56 (1 - x_{CELL}) - 1.47x_{CELL}$
3 CELLA $\rightarrow (1 - x_{CELL}) \times (\text{LVG/same products as R}_2) + x_{CELL} \times (5.5 \text{ Char} + 4 \text{ H}_2\text{O} + 0.5 \text{ CO}_2 + \text{H}_2)$	2×10^6	41.86	$0.53 (1 - x_{CELL}) - 1.47x_{CELL}$
Hemicellulose scheme			
4 HCE $\rightarrow 0.4 \text{ AA (HW)}/0.1 \text{ AA (SW)} + 0.58 \text{ HCEA1} + 0.42 \text{ HCEA2}$	1×10^{10}	129.77	0.0
5 HCEA1 $\rightarrow (1 - x_{HCE}) \times (0.5 \text{ CO} + 0.5 \text{ CO}_2 + 0.325 \text{ CH}_4 + 0.8 \text{ CH}_2\text{O} + 0.1 \text{ CH}_3\text{OH} + 0.25 \text{ C}_2\text{H}_4 + 0.125 \text{ ETOH} + 0.025 \text{ H}_2\text{O} + 0.025 \text{ HCOOH} + 0.275 \text{ G}\{\text{CO}_2\} + 0.4 \text{ G}\{\text{COH}_2\} + 0.125 \text{ G}\{\text{H}_2\} + 0.45 \{\text{CH}_3\text{OH}\} + 0.875 \text{ Char}) + x_{HCE} \times (4.5 \text{ Char} + 3 \text{ H}_2\text{O} + 0.5 \text{ CO}_2 + \text{H}_2)$	1.2×10^9	125.58	$0.25 (1 - x_{HCE}) - 1.42x_{HCE}$
6 HCEA1 $\rightarrow (1 - x_{HCE}) \times (0.1 \text{ CO} + 0.8 \text{ CO}_2 + 0.3 \text{ CH}_2\text{O} + 0.25 \text{ H}_2\text{O} + 0.05 \text{ HCOOH} + 0.15 \text{ G}\{\text{CO}_2\} + 0.15 \text{ G}\{\text{CO}\} + 1.2 \text{ G}\{\text{COH}_2\} + 0.2 \text{ G}\{\text{H}_2\} + 0.625 \text{ G}\{\text{CH}_4\} + 0.375 \text{ G}\{\text{C}_2\text{H}_4\} + 0.875 \text{ Char}) + x_{HCE} \times (4.5 \text{ Char} + 3 \text{ H}_2\text{O} + 0.5 \text{ CO}_2 + \text{H}_2)$	$0.15 \times T$	33.5	$-0.64 (1 - x_{HCE}) - 1.42x_{HCE}$
7 HCEA1 $\rightarrow (1 - x_{HCE}) \times (\text{XYL/same as R}_6) + x_{HCE} \times (4.5 \text{ Char} + 3 \text{ H}_2\text{O} + 0.5 \text{ CO}_2 + \text{H}_2)$	$3 \times T$	46.05	$0.77 (1 - x_{HCE}) - 1.42x_{HCE}$
8 HCEA2 $\rightarrow (1 - x_{HCE}) \times (0.2 \text{ HAA} + 0.175 \text{ CO} + 0.275 \text{ CO}_2 + 0.5 \text{ CH}_2\text{O} + 0.1 \text{ ETOH} + 0.2 \text{ H}_2\text{O} + 0.025 \text{ HCOOH} + 0.4 \text{ G}\{\text{CO}_2\} + 0.925 \text{ G}\{\text{COH}_2\} + 0.25 \text{ G}\{\text{CH}_4\} + 0.3 \text{ G}\{\text{CH}_3\text{OH}\} + 0.275 \text{ G}\{\text{C}_2\text{H}_4\} + \text{Char}) + x_{HCE} \times (4.5 \text{ Char} + 3 \text{ H}_2\text{O} + 0.5 \text{ CO}_2 + \text{H}_2)$	0.5×10^{10}	138.14	$-0.14 (1 - x_{HCE}) - 1.42x_{HCE}$
Lignin scheme			
9 LIG-C $\rightarrow 0.35 \text{ LIG-CC} + 0.1 \text{ pCOUMARYL} + 0.08 \text{ PHENOL} + 0.32 \text{ CO} + 0.3 \text{ CH}_2\text{O} + \text{H}_2\text{O} + 0.7 \text{ G}\{\text{COH}_2\} + 0.495 \text{ G}\{\text{CH}_4\} + 0.41 \text{ G}\{\text{C}_2\text{H}_4\} + 5.735 \text{ Char}$	1.33×10^{15}	203.02	-0.47
10 LIG-H $\rightarrow \text{LIG-OH} + 0.25 \text{ HAA} + 0.5 \text{ C}_3\text{H}_6\text{O} + 0.5 \text{ G}\{\text{C}_2\text{H}_4\}$	0.67×10^{13}	156.97	0.10
11 LIG-O $\rightarrow \text{LIG-OH} + \text{CO}_2$	0.33×10^9	106.74	-0.21
12 LIG-CC $\rightarrow (1 - x_{LIG}) \times (0.35 \text{ HAA} + 0.3 \text{ pCOUMARYL} + 0.2 \text{ PHENOL} + 0.4 \text{ CO} + 0.65 \text{ CH}_4 + 0.6 \text{ C}_2\text{H}_4 + 0.7 \text{ H}_2\text{O} + 0.4 \text{ G}\{\text{CO}\} + \text{G}\{\text{COH}_2\} + 6.75 \text{ Char}) + x_{LIG} \times (15 \text{ Char} + 4 \text{ H}_2\text{O} + 3 \text{ H}_2)$	3×10^7	131.86	$-0.09 (1 - x_{LIG}) - 1.30x_{LIG}$
13 LIG-OH $\rightarrow \text{LIG} + 0.55 \text{ CO} + 0.05 \text{ CO}_2 + 0.1 \text{ CH}_4 + 0.6 \text{ CH}_3\text{OH} + 0.9 \text{ H}_2\text{O} + 0.05 \text{ HCOOH} + 0.6 \text{ G}\{\text{CO}\} + 0.85 \text{ G}\{\text{COH}_2\} + 0.1 \text{ G}\{\text{H}_2\} + 0.35 \{\text{CH}_4\} + 0.3 \text{ G}\{\text{CH}_3\text{OH}\} + 0.2 \text{ G}\{\text{C}_2\text{H}_4\} + 4.15 \text{ Char}$	1×10^8	125.58	-0.17
14 LIG $\rightarrow (1 - x_{LIG}) \times \text{FE2MACR} + x_{LIG} \times (10.5 \text{ Char} + 3 \text{ H}_2\text{O} + 0.5 \text{ CO}_2 + 3 \text{ H}_2)$	$4 \times T$	50.2	$0.95 (1 - x_{LIG}) - 1.52x_{LIG}$
15 LIG $\rightarrow (1 - x_{LIG}) \times (0.2 \text{ C}_3\text{H}_6\text{O} + \text{CO} + 0.2 \text{ CH}_4 + 0.2 \text{ CH}_2\text{O} + 0.4 \text{ CH}_3\text{OH} + 0.2 \text{ CH}_3\text{CHO} + 0.95 \text{ H}_2\text{O} + 0.05 \text{ HCOOH} + 0.45 \text{ G}\{\text{CO}\} + 0.5 \text{ G}\{\text{COH}_2\} + 0.4 \{\text{CH}_4\} + 0.65 \text{ G}\{\text{C}_2\text{H}_4\} + 5.5 \text{ Char}) + x_{LIG} \times (10.5 \text{ Char} + 3 \text{ H}_2\text{O} + 0.5 \text{ CO}_2 + 3 \text{ H}_2)$	0.4×10^9	125.58	$-0.35 (1 - x_{LIG}) - 1.52x_{LIG}$
16 LIG $\rightarrow (1 - x_{LIG}) \times (0.4 \text{ CO} + 0.2 \text{ CH}_4 + 0.4 \text{ CH}_2\text{O} + 0.6 \text{ H}_2\text{O} + 0.2 \text{ G}\{\text{CO}\} + 2 \text{ G}\{\text{COH}_2\} + 0.4 \{\text{CH}_4\} + 0.4 \text{ G}\{\text{CH}_3\text{OH}\} + 0.5 \text{ G}\{\text{C}_2\text{H}_4\} + 6 \text{ Char}) + x_{LIG} \times (10.5 \text{ Char} + 3 \text{ H}_2\text{O} + 0.5 \text{ CO}_2 + 3 \text{ H}_2)$	$0.083 \times T$	33.5	$-0.50 (1 - x_{LIG}) - 1.52x_{LIG}$
Trapped gas scheme			
17 $\text{G}\{\text{CO}_2\} \rightarrow \text{CO}_2$	1×10^5	100.46	0.0
18 $\text{G}\{\text{CO}\} \rightarrow (1 - x_{G\{\}}) \times \text{CO} + x_{G\{\}} \times (0.5 \text{ Char} + 0.5 \text{ CO}_2)$	3×10^{13}	209.3	$-3.08x_{G\{\}}$
19 $\text{G}\{\text{COH}_2\} \rightarrow 0.75\text{G}_2\{\text{COH}_2\} + 0.25 (\text{H}_2 + 0.5 \text{ CO} + 0.25 \text{ CO}_2 + 0.25 \text{ Char})$	1×10^6	100.46	0.31
20 $\text{G}\{\text{H}_2\} \rightarrow \text{H}_2$	1×10^{12}	313.96	0.0
21 $\text{G}\{\text{CH}_4\} \rightarrow \text{CH}_4$	2×10^{13}	300.0	0.01
22 $\text{G}\{\text{CH}_3\text{OH}\} \rightarrow (1 - x_{G\{\}}) \times \text{CH}_3\text{OH} + x_{G\{\}} \times (\text{Char} + \text{H}_2\text{O} + \text{H}_2)$	1.2×10^{13}	209.3	$-1.27x_{G\{\}}$
23 $\text{G}\{\text{C}_2\text{H}_4\} \rightarrow 0.3 \text{ C}_2\text{H}_4 + 0.7 (\text{CH}_4 + \text{Char})$	1×10^6	100.46	0.46
24 $\text{G}_2\{\text{COH}_2\} \rightarrow 0.2 \text{ G}_3\{\text{COH}_2\} + 0.8 (\text{CO} + \text{H}_2)$	1.5×10^9	209.3	0.0
Extractives scheme			
25 TANN \rightarrow FENOL + ITANN	50	46.02	
26 ITANN $\rightarrow 5\text{CHAR} + 3\text{CO} + \text{GCOH}_2 + 2\text{H}_2\text{O}$	6100	25.52	
27 TGL \rightarrow acrolein + 3FFA	7×10^{12}	191.2	
Reaction parameters			
X_CELL	Low	High	
X_HCE	charring	charring	
X_LIG	0.025	0.1	
X_G{\}	0.05	0.2	
X_LIG	0.1	0.4	
X_G{\}	0.1	0.4	
Sugar formation in R ₃ and R ₈	Yes	No	

or inside the particle if transport out of the particle is slow. Several types of intraparticle secondary reactions have been described, including reactions of condensable volatiles at the gas/liquid interface,⁵⁷ reactions of condensable volatile in the

liquid phase^{182–184} (polymerization, crosslinking, dehydration) and reactions of gases and condensable volatiles with the mineral matter in char.^{185,186} The Ranzi scheme discussed above has secondary reactions in the scheme.^{78,79,158}



The most studied reactions are thermal decomposition, or cracking, reactions that occur in the gas phase and solid–gas phase reactions including gasification, combustion, and oxidation.^{187–190} Cracking reactions are important at high temperatures ($T > 500$ °C) and are more prominent when long residence time in the hot zones exist.^{188,191,192} Numerous studies related to solid–gas secondary reactions have also been reported.^{189,193–196} These reactions have been studied at an experimental level, and have been measured by increases in char production and decreases in volatiles caused by an increase in the time of solid–gas contact.

Several studies conducted at both the experimental and theoretical level, have reported the reactions responsible for the thermal decomposition of volatiles. Hayashi *et al.*¹⁸⁸ reports a rigorous theoretical and experimental study of volatile cracking reactions released from biomass fast pyrolysis. The model predicted with good accuracy the behavior of light gases (CO, CO₂, H₂, CH₄, C₂H₄), but provided less accurate predictions for furanic compounds as well as benzene, toluene, acetic acid, aldehyde, methanol. Shin *et al.*¹⁹⁷ proposed a 3 stage kinetic model for the decomposition of volatiles produced from cellulose, levoglucosan and 5-HMF pyrolysis. They found that in 0.6 seconds at 750 °C, 80% of the volatiles generated in during pyrolysis decompose. Further experimental studies at fast pyrolysis conditions were carried out by Morf *et al.*,¹⁹² Graham *et al.*,¹⁹⁸ and Zhang *et al.*¹⁹⁹ Each evaluated char, total volatiles, condensable species and light gases yields as a function of temperature and gas residence time in two stage reactors. In these studies, it was found that the maximum decomposition rate was achieved at a temperature range of between 600–700 °C. The poly-hydroxy-aromatic (PHA) formation is another interesting secondary reaction that results primarily from the volatile aromatic derivatives of lignin. Investigations have shown that these reactions are favored at very high temperatures (>800 °C) and long residence times (>10 s),^{199,200} which are typical operational conditions for gasification and combustion processes.

Another set of solid–gas and gas phase secondary reactions are related to char gasification with steam or CO₂. These reactions include char oxidation, the Boudouard reaction, reforming reactions, water gas-shift reactions and methanation, which have been widely explored, in biomass and coal gasification and combustion processes.^{72,201,202}

The liquid–gas and liquid–liquid phase reactions that occur during biomass pyrolysis are largely unexplored with no theoretical models to report. This is primarily due to uncertainty regarding the components present in the liquid phase and the difficulties associated with monitoring these species during pyrolysis. Understanding of the liquid phase reaction is one of the main challenges to be addressed in order to better elucidate of the reactivity of biomass during the pyrolysis process.¹²² Despite this lack of knowledge, it is known that reactions in the liquid phase are the primary contributors to the production of secondary char, light oxygenated compounds and permanent gases.^{132,182}

Paulsen²⁰³ looked at the reactivity of mixtures of levoglucosan/fructose to simulate the environment and interactions

inside the intermediate liquid phase formed during cellulose fast pyrolysis. Cyclization and elimination reactions were found to be responsible for the production of light oxygenated compounds and char. Both reactions are more intense in the pyrolysis of powders than for thin films. Furthermore, the author suggested that catalysts, such as palladium, promote decarbonylation reactions in the liquid phase, reducing aldehydic furans and improving the bio-oil quality (less oxygen content in bio-oil). Bai *et al.*²⁰⁴ suggested a competitive mechanism between levoglucosan evaporation and polymerization during cellulose pyrolysis in TGA. The intensity of both reactions depends on the quantity of sample, the carrier gas flow rate, and the heating rate. They show that low sample quantities and high carrier gas flow promote evaporation due to lower mass transfer resistance. Similar mechanisms were proposed by Hosoya *et al.*²⁰⁵ for levoglucosan pyrolysis in an ampoule reactor. Other important reactions in the liquid phase are related to the decomposition of lignin oligomers to produce monomers, char and permanent gases. However, there are few studies focused on the pyrolysis of lignin oligomers, and none relating experimental results using kinetic parameters. It has been suggested that these compounds are generated by thermal ejection caused by intense bubbling of the liquid intermediates.^{117,206,207} These aerosols can be trapped inside the solid matrix after ejection and the collision probability will increase with particle size.^{106,207,208}

Heat transfer

Effect of particle size and pore structure on heat transfer

Biomass from any source is a poor conductor of heat, with a thermal conductivity on the order of 0.1 W m⁻¹ K⁻¹ which promotes formation significant temperature gradients within particles when subjected to rapid heating. Temperature gradients are strongly influenced by particle size and shape, and are typically anisotropic due to the direction microstructure of biomass and the aspherical shape typical of milled particles, as illustrated in Fig. 6.^{209–211} The primary purpose of milling biomass before pyrolysis is to enhance heat transfer. In an industrial or pilot scale process, the cost of this milling may be a significant operating expense, so it is critical to find the “sweet spot” that maximizes yield and minimizes reactor cost. Bridgwater *et al.*⁶⁹ presented specifications for particle sizes to be used with each of the major technologies available. For example: particle sizes less than 6 mm are well suited for fluidized bed reactor and circulating bed designs, and up to 10 cm are suitable for ablative rotary disc processes.

The shape and hierarchical pore structure also impact the heat and mass transfer properties of wood. Ciesielski *et al.*⁹³ have shown, for example, that spherical representations of realistic biomass feedstocks fail to accurately capture the thermal gradients in a particle, though some effort has also been made to employ develop low-order mathematical approximations in spherical models to make them more accurate.²¹³ Pecha *et al.* have further illustrated that the pore structure between even pine



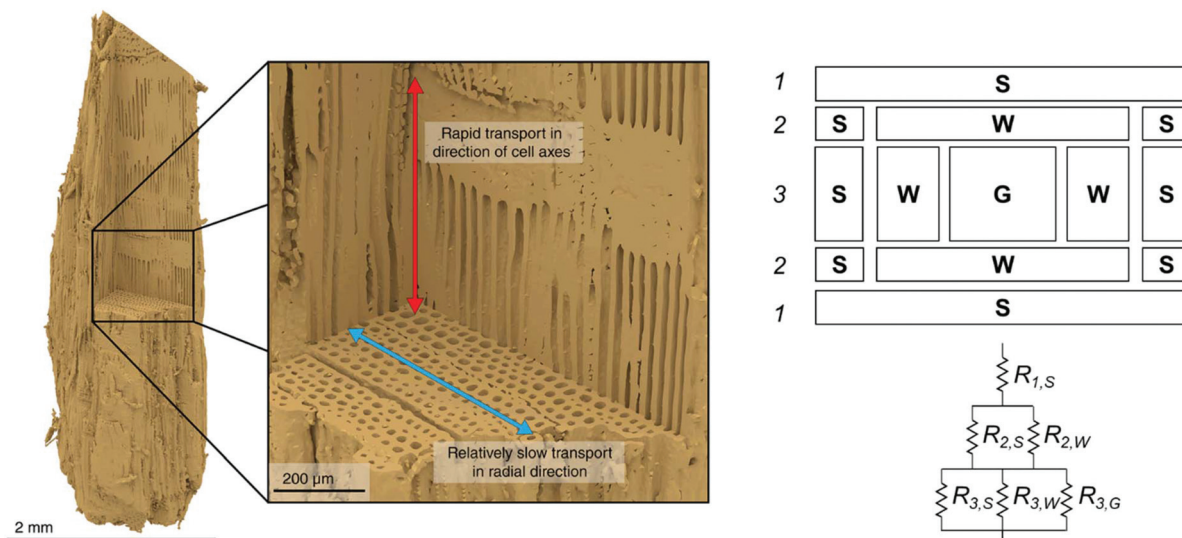


Fig. 6 (Left) A visual reconstruction of the softwood fiber structure of pine based on X-ray tomography data²⁶ illustrates that heat and mass both move more readily in the direction of cell axes than in radial direction and (right) heat flux resistance pathways for wood, depending on the direction of the flux.²¹² Left image reprinted (adapted) with permission from Ciesielski et al.²⁶ Copyright (2018) Wiley.

and poplar wood makes for differences in the external heat transfer coefficients.²¹⁴ Thunman and Leckner²¹² developed a model for thermal conductivity in wood based on the pore structure for perpendicular and parallel directions relative to the fibers in the wood, as well as density and moisture content. Conductivity parallel to the fiber was found to be approximately twice (e.g. $0.4 \text{ W m}^{-1} \text{ K}^{-1}$) the conductivity in perpendicular direction ($0.2 \text{ W m}^{-1} \text{ K}^{-1}$). Some unique models for approximating the resistance of cell walls have been developed (see Fig. 6).²¹²

Several dimensionless numbers have been developed for examining heat transfer in pyrolysis of biomass particles. The Biot number compares external and internal heat transfer rates:

$$\text{Bi} = \frac{hR}{\lambda} \quad (7)$$

Pyrolysis number 1 compares internal heat transfer rate and reaction rate,

$$\text{Py}_1 = \frac{\lambda}{(k\rho c_p R^2)}, \quad (8)$$

and pyrolysis number 2 compares external heat transfer with reaction rate,

$$\text{Py}_2 = \frac{h}{k\rho c_p R}, \quad (9)$$

where R is radius, λ is thermal conductivity, ρ is bulk particle density, k is apparent reaction rate constant, and h is external heat transfer coefficient. In the region in which heat conduction is limiting the conversion rate, mass transfer of vapors leaving the particle will also be a concern (larger than 1 mm for most reactors).

Pyle and Zaror authored a seminal paper describing various regimes in which one phenomenon controls the effective rate

Table 3 Regions of validity for models single particle models²¹⁵

Model	Approximate range of validity		
	Bi	Py ₁	Py ₂
I: Non-controlled conditions	All	All	All
II: External heat transfer controlled	<1	>1	>1
III: Kinetics controlled	<1	>10	>10
IV: Internal heat transfer controlled	>50	<10 ⁻³	<<1

of pyrolysis, as shown in Table 3.²¹⁵ If the particle is large enough that Biot number is greater than 10 and the Py_1 is less than 1, the heating is limited by internal thermal conduction. Hence, heating rate of the reactor in some respects is less important than particle size.^{216–218} Dauenhauer's group extended this analysis into a pyrolysis transport map as shown in Fig. 7.^{48,122}

In micro-reactors at pyrolysis temperatures, conductive and convective heat transfer account for 90% of the heat flux, while radiation accounts for less than 10% of the total heat transfer.²¹⁹ However, in larger wall-heated reactors it is well-known that radiative heat transfer can account for a significant portion of the total heat transfer and can be modeled using the Stefan–Boltzmann law.²²⁰ For more accurate use of the dimensionless numbers described above total surface heat transfer may be accounted for in a lumped heat transfer coefficient comprised of the sum of the convective and radiative heat transfer coefficients.²²¹

Although heating rate is a critical factor to define the conditions of fast pyrolysis, it cannot typically be measured experimentally; it is instead estimated with theoretical models or by using engineering approaches to minimize the thermal thickness, like using a thin film.²²² Often, these parameters are subjective and depend strongly on the pyrolysis system, measurement equipment, model assumptions, and experimental con-



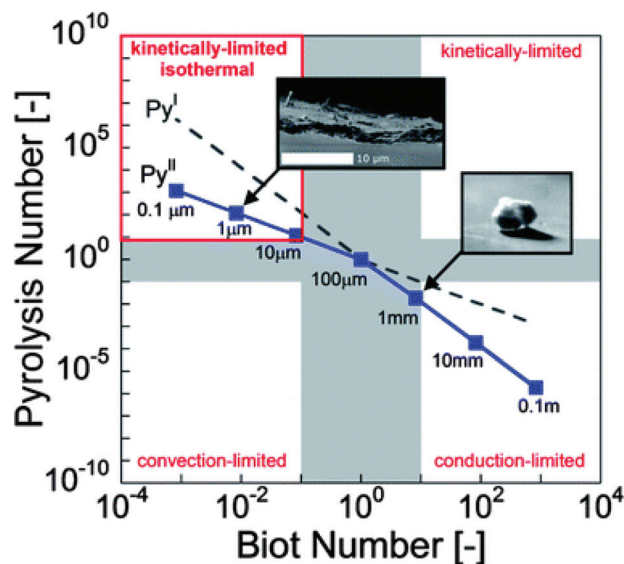


Fig. 7 Pyrolysis transport map. Relative rates of biomass reaction and heat transfer by conduction or convection at 500 °C are compared in terms of Pyrolysis number and Biot number; four pyrolysis regimes are identified (clockwise from top left): isothermal and reaction limited, reaction-limited, conduction-limited, and convection-limited.¹²² Republished with permission of the Royal Society of Chemistry, Mettler *et al.*,¹²² permission conveyed through Copyright Clearance Center, Inc.

ditions (see Table 4). This makes it difficult to compare experimental results obtained for global yields and kinetic parameters published in scientific papers. There is ambiguity in determining the correct heating rate from values reported in publications. In most cases, heating rate is reported as being the programmed rate for heating elements (electrical resistance, radiation lamp, hot plate), rather than a particle heating rate.

Particle heating is commonly assumed to follow a linear heating, mathematically represented as: $T = \beta t + T_0$ with β , t and T_0 as the heating rate, time and initial temperature of the particle, respectively.^{85,216,223} This is a classic simplification used for “lumped” heat transfer wherein spatial temperature gradients are neglected. This approximation is typically considered applicable for systems exhibiting a Biot number ≤ 0.1 and can be valid under certain experimental conditions such as those in thermogravimetric analysis or pyrolysis subjected to kinetic control conditions using thin films and small particle sizes. If this condition is met, and there are not convective heat transfer limitations, the reaction rate constants may be considered “intrinsic”, meaning transport phenomena are lumped with chemical reaction rates.

Fig. 4 illustrated that for large particles (>1 mm), temperature gradients exist and heating rates at each point are different, modifying reactivity patterns at each point within the biomass particle (the reaction rate depends on the heating

Table 4 Common reactors used to study biomass pyrolysis

Reactor	T (°C)	HR (°C s ⁻¹)	Time (s)	P (atm)	Sample	Remarks
Drop tube	1500	10^4 – 10^5	4	Vac – 69	Powder	High heating rates. It is not possible to measure particle temperatures. Moderate gas residence time. Small sample size
Wire mesh	1500	1000	1–3600	Vac – 69	Powder	Uncertainty in particle temperature measurement. Short residence times of volatiles. Low uncertainty in the mass balances
TGA	1200	1	h	Vac – 2	<2 mm	Low heating rates and temperatures. High volatile residence time. Good estimation of sample temperature. Online monitoring sample weight
Radiation	2000	10^4 – 10^6	ms	1	Powder	High heating rates. Difficult temperature measurement
Shock tubes	700–2200	10^6	ms	Vac	Fine	The temperature history of the system is well established. In shock tube reaction, times longer than 3.0 ms cannot be achieved. Also, sampling of the reaction zone for gas chromatographic analysis is time consuming
Py-MS	700	20 000	10–3600	Vac – 1	Fine	Direct measurement of volatile and light gases. Indirect measurement of the sample temperature
Fluid bed	700–1200	1000–10 000	1–3600	1–100	1–5 mm	High uncertainties in the closure of mass balances. Volatile quantifying uncertain. It is not possible to accurately determine particle temperature and residence time
Plate reactor	300–700	10^2 – 10^6	s	1–2	μ m–film	Accurate temperature measurement of the particle. Uniformity on solid and temperature distribution. Visualization of solid sample
Ablative	600	—	1	1	cm–m	Volatile cracking reactions. Solids temperature measurement is not possible. Difficult volatile monitoring
Auger	600	—	min	1	Chips	Bio-oil contamination of sand. Difficult estimation of solids temperature. Higher volatile residence times
Curie point	300–900	—	s	1	μ m–mm	The temperature ceases to rise when the Curie-point of the metal has been reached; that is the exact reproducible temperature at which the ferromagnetic material loses its magnetism. Temperature control is limited
Microwave	500–2000 W	—	min	1	Powder	Suffers from uneven sample heating. Selective heating of polar groups

HR: heating rate, Vac: vacuum, P: pressure.



rate and reaction temperature). This consideration has been ignored in many studies,^{50,67,86} where normally the same reaction rate expression is used to describe the reactivity of the whole particle. This can cause problems when predicting the final product distribution, because each point inside the particle has a different effective reaction rate due to changes in the heating rates and temperatures and rate data is this not intrinsic kinetic data. Care must be taken to review the experimental protocols used to extract rate data which not collated with heat transfer or mass transfer through a large biomass particle or dust pile (>100 μm thick).¹²² For fast pyrolysis studies, there are many techniques and devices, which can ensure conditions of high heating rates, such as: flash radiation lamps, wire mesh reactors, discharge tubes, and free fall reactors. Table 4 summarizes the main characteristics of these devices. For intrinsic kinetic studies, the most commonly used devices are free fall reactors, Pyrolysis-Mass Spectrometry (Py-MS) reactors, and wire mesh reactors, all of which minimize heat transfer limitations and secondary vapor phase reactions.^{223–227} For further reading on how to design a system for extracting intrinsic pyrolysis kinetics, please see Fig. 7 and a recent publication by the Dauenhauer research group.²²²

Mass transfer

Influence of pore structure on intraparticle mass transport

The rapid evolution of pyrolysis vapors can generate significant pressure gradients within particles, particularly in larger particles, because the volatiles cannot be instantly evacuated. This causes some internal structures to crack, increasing the quantity of both macropores and micropores. When the biomass passes through a molten (or metaplastic) phase, the internal structures can warp and block pores at high heating rates.^{112,113}

Shen *et al.*¹⁰⁶ found that bio-oil yields obtained from mallee (Australian Eucalyptus) in a fluidized bed reactor decreases as the particle diameter increases from 0.3 to 1.5 mm, and thereafter bio-oil yield is insensitive to particle size increases. Koçkar *et al.*²²⁸ reported that the maximum bio-oil yield is achieved with a 0.42 mm particle size. Koçkar *et al.*²²⁸ and Nurul *et al.*²²⁹ have reported ideal particle sizes of between 0.85 to 200 μm for fixed-bed pyrolysis of rapeseed, with bio-oil yields of up to 60 wt%. Laboratory scale studies of primary biomass pyrolysis reactions show that rapid removal of products the solid matrix is critical to achieve high oil yields. This removal is facilitated by the use of small particle sizes (<100 μm) and vacuum conditions.^{227,230}

Increasing vapor residence time favors cracking reactions that reduce the yield of condensable vapors.^{11,28,92} Therefore, secondary reactions between char and volatiles are more prominent in larger particles, which increases the yield of char and gases by consuming pyrolysis oil.⁸⁴ Zhou *et al.*¹¹⁷ showed the effect of particle size on the formation of char and lignin oligomers during beech wood pyrolysis. The authors suggest two distinct pyrolysis regimes to explain the oligomer yields obtained. In the first regime, for particles with diameters between

0.3–3 mm, the lignin oligomer yield decreased when particle size was increased. For particles >3 mm, no changes were observed in oligomer yield when particle size changed. The authors hypothesized that lignin oligomers come from aerosol ejection from the intermediate liquid phase which form char while diffusing through large particles. In contrast, char yield increased with increasing particle size. A reasonable explanation for this behavior is that below 1 mm, all the pore ends are open. This allows for vapors to leave without being trapped.^{100,231} Similar results have been reported elsewhere.^{11,24,37,68,69,84,131}

The anisotropic pore structure⁹³ of most lignocellulose species (like wood) leads to differences in gas permeabilities in the axial and radial directions of up to 4 orders of magnitude.²³² Permeability scales with pore size to the second power, so this is to be expected.²¹⁰ Adding complication to modelling, macroporosity changes dynamically as biomass is converted to char.²³³ Diffusion constants used in Fick's laws for porous media transport through macropores can be estimated as $D_{\text{eff}} = \epsilon D / \tau$, where ϵ is porosity, D is gas diffusivity for a species, and τ is tortuosity (often estimated as $\tau = \epsilon^{-1/3}$). Diffusion coefficients should be represented as a tensor, since porosity is anisotropic. Anisotropic permeability and diffusion have been clearly illustrated experimentally in laser-induced fluorescence experiments shown in Fig. 8.²³⁴

Influence of reactor residence times on pyrolysis

A short vapor residence time increases the bio-oil yield and minimize secondary vapor phase reactions.^{37,69,99,100,235} However high feedstock residence times are needed to ensure complete devolatilization.^{37,100,192} These times are directly related to the

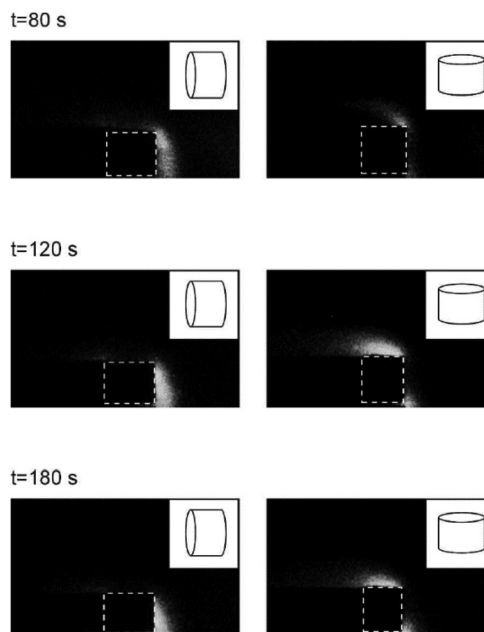


Fig. 8 Laser-induced fluorescence images of pyrolysis volatiles leaving the axial ends of wood cylinders due to anisotropic permeation and diffusion.²³⁴ © 2003, Optical Society of America, reprinted with permissions.



operating conditions, such as carrier gas flow for fluidized beds, and the technology used in the pyrolysis process. For instance, the residence time of solids in fixed bed reactors is higher (minutes to hours) while in fluid bed reactors the residence time for both solids and volatiles is very short (a few seconds), depending on the height of the reactor and the gas flow. Scott *et al.*²³⁶ have observed in pyrolysis of sorghum bagasse at 550 °C that as the vapor residence time increases, the bio-oil yield drastically reduces from 75% to 57% and char and gas yields increase. Further, heavy tars are formed with more vapor residence time. Some authors suggest taking advantage of these reactions to increase monomer yield by cracking oligomers.²¹

The volatile residence time in a fluidized bed reactor is manipulated by changing the carrier gas flow,^{99,237,238} however it can also be altered by gas phase pressure. At high pressures the volatile residence time within particles is high, promoting secondary cracking reactions.^{190,209,239} In wire mesh and hot plate reactors this condition is commonly achieved by operating the reactor under vacuum (<200 mbar).^{230,240,241} Under these conditions, evaporation of high molecular weight oligomers from cellulose decomposition is possible. Aerosol ejection from the liquid phase is also more intense under these conditions.^{242,243}

Phase change

Liquid intermediate formation

A recent study conducted by Zhou *et al.*¹⁸⁴ showed that for both slow pyrolysis (2 K s⁻¹) and fast pyrolysis (1000 K s⁻¹), lignin decomposes to form an intermediate liquid phase, which swells, evaporates and shrinks to form a droplet. Other studies at high heating rates showed that an intense foaming of oligomeric compounds in the liquid phase occurs, which may be ejected from the surface as aerosolized droplets.²⁴⁴ At low heating rates, the intermediate liquid provides an ionic medium in which cross-linking reactions can occur, promoting char formation and decreasing high molecular weight oligomers yield.¹³² Thus, it could be concluded that minimizing the time that compounds stay in that phase will enhance the yield of oligomeric compounds through unique physical mechanisms. It has been shown that during boiling of a liquid at high heating rates, an intense bubbling is produced due to violent expansion of gases in over very short periods of time which facilitates fluid collapse by micro-explosions that release small liquid droplets as aerosols into the gas stream.^{243,245,246}

The existence of an intermediate liquid phase during biomass pyrolysis has sparked much debate for several decades. Goring *et al.*⁸⁸ observed that for cellulose, lignin, and hemicellulose a softening occurred when samples were immersed in a hot oil bath. The authors attributed this behavior to the formation of an intermediate liquid phase due to thermal degradation rather than a phase change associated with the glass transition point. The softening point was estimated as the temperature at which the solid structure collapsed (227–253 °C for cellulose), higher than the glass transition point of the cellulose (145–175 °C).

In 1974, Nordin *et al.*²⁴⁷ carried out an interesting experiment in which visual evidence of a condensed phase during flash pyrolysis was presented when exposing various types of leaf paper to a heating laser in less than 0.1 ms. Once samples reach the pyrolysis temperature, volatiles were collected, rapidly cooled and analyzed. Based on these results the authors argued that the structure of cellulose was clearly broken and the crystallinity index had decreased by 35%. These results were attributed to a physical phenomenon, namely fusion of cellulose, instead of thermal degradation. They concluded that cellulose and biomass pass through a condensed phase, although it could not be determined if this behavior was due to a chemical thermal degradation process or a condensation of monomers. Over the course of this decade, the first theoretical models to explain the phase change of cellulose and biomass during pyrolysis were developed. These schemes include an intermediate liquid phase referred to as activated cellulose in some studies.

Kilzer and Broido¹³⁸ rigorously analyzed the data reported in the literature related to cellulose pyrolysis obtained using a thermobalance. They proposed a two-step competitive model (model 1, Table 1). In this model they describe the transformation of cellulose into an unstable intermediate compound, which in most cases is associated with monomeric levoglucosan. Between 1971 and 1975, Broido and Weinstein,¹³⁴ Shafizadeh,¹⁴¹ and Bradbury¹⁴⁵ proposed similar reaction schemes. Patai *et al.*²⁴⁸ and Golova²⁴⁹ studied changes in degree of polymerization of cellulose at a temperature range of 170–230 °C, where the mass loss only reaches approximately 5%. These researchers associated the reduction of the degree of polymerization with random breaking of cellulose glycosidic bonds, giving rise to anhydrous glucose derived sugars (levoglucosan, cellobiosan, cellotriosan).

In 1980, during the Specialists' Workshop on Biomass Fast Pyrolysis held at Copper Mountain (USA), Diebold²⁵⁰ carried out in front of all attendees an experiment in which he demonstrated a hot nichrome wire passing through a piece of biomass as if it were butter, without a trace of char. A detailed analysis of the biomass edges after cutting showed virgin biomass structures covered by a beige layer, like a varnish. The explanation for this observation was the formation of a molten phase, which is liquid at pyrolysis temperatures and solid at room temperature. After Diebold's demonstration, in 1985 Lédé *et al.*²⁵¹ presented the results of a biomass pyrolysis experiment in a spinning disk reactor. In this experiment large cylindrical wood pieces were pressed at high pressure (0.1–3.5 MPa) against a hot disk (500–900 °C) that rotates at constant velocity, thereby degrading biomass. During the reaction on the heated disc, a liquid film formed in the wake of the biomass, like a lubricant, direct evidence of the formation of an intermediate liquid phase that faded away with time. This evidence corroborated the results presented by Diebold.²⁵⁰

In 1989, Pouwels *et al.*²⁵² carried out a number of interesting experiments for the study of the fast pyrolysis *in situ* and *ex situ* of cellulose, using a mass spectrometer Desorption Chemical Ionization Mass Spectrometry (DCI-MS), Desorption



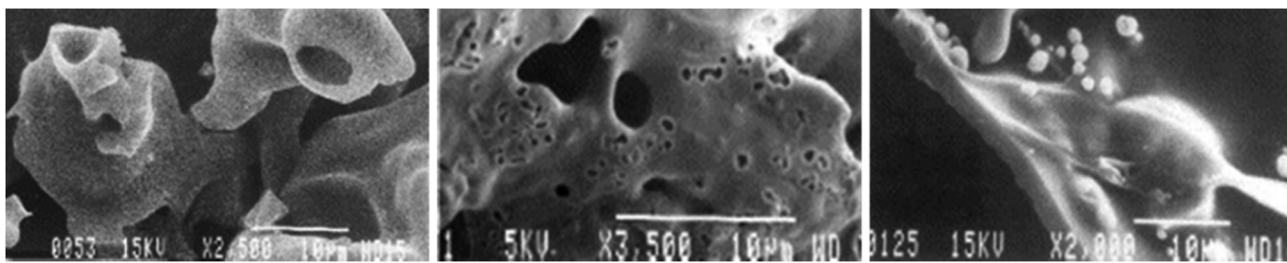


Fig. 9 SEM micrographs illustrate structural changes of (left) avicel cellulose, (center) tobacco, and (right) alkali lignin during pyrolysis, which clearly show a melt phase has occurred. Reprinted from McGrath *et al.*,²⁵⁶ Copyright (2003), with permission from Elsevier.

Ionization Mass Spectrometry (DC-MS) and curie-point-DCI-MS. The authors identified anhydrous sugars containing up to 6 degrees of polymerization (cellohexoses) at high concentrations, thereby indicating that the first step of the devolatilization process is depolymerization of lower molecular weight compounds with high melting points (>250 °C). Boiling points between 339–550 °C hint that levoglucosan and cellobiosan make up the intermediate liquid phase.

In 1998 Boutin *et al.*²⁵³ studied cellulose fast pyrolysis in a reactor heated by radiation and confirmed the formation of an off-yellow film, on the cellulose surface. This film was soluble in water and polar solvents, indicating that this was a new compound and not molten cellulose, which has a hydrophobic behavior. Piskorz *et al.*²⁵⁴ concluded that it is very difficult to reach the glass transition point without undergoing any thermal degradation. Piskorz *et al.*²⁵⁵ later presented experimental evidence showing that it is possible to depolymerize cellulose to produce oligosaccharides with varying degrees of polymerization. They showed by scanning electron microscopy (SEM) analysis that the cellulose structure was deformed like a swollen polymer. Qualitative analysis by HPLC of the methanol soluble solid char showed a variety of anhydrosugars with degrees of polymerization of 1–7, and a heavier fraction which could not be identified. These results again corroborate that cellulose passes through an intermediate liquid-state product during depolymerization.

In 2003 McGrath *et al.*²⁵⁶ studied the char obtained from pyrolysis of tobacco leaves and its biopolymer constituents. Using scanning electron microscopy (SEM), they demonstrated that snuff char passes through a melt phase between 450 and 550 °C (see Fig. 9). The authors did not assess the chemical composition of melt phase. The structural behavior of the cellulose and hemicellulose presented in these images is similar to that found for the tobacco leaves, as shown by Dufour.²⁵⁷

In 2008, Liu *et al.*²⁵⁸ carried out experiments in an oven heated by radiation (under fast pyrolysis conditions) to estimate the evolution of the condensed phase formed during cellulose pyrolysis. The authors confirmed results that had been obtained in previous studies.^{131,253,259} They demonstrated that up to 68% (w/w) of the initial cellulose formed a water soluble compound of yellow color, consisting primarily of oligomers of glucose. They proved that under lower severity heating conditions lower yields of the condensed phase were obtained, with maximum yields dropping to 57% (w/w).

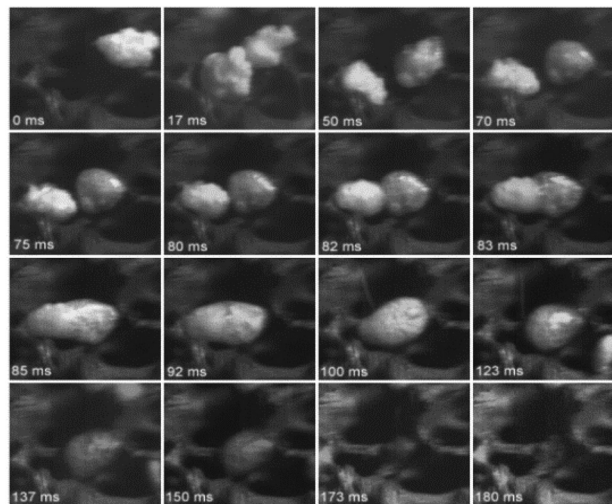


Fig. 10 Photographic record of the evolution of the cellulose during fast pyrolysis. Republished with permission of the Royal Society of Chemistry, Dauenhauer *et al.*,¹¹⁰ permission conveyed through Copyright Clearance Center, Inc.

An article presented by Mamleev *et al.*¹³² in 2009 investigated a new kinetic model called a “two-phase model”. This model incorporates the formation and growth of liquid cavities within the cellulose that contain high boiling point compounds within the liquid phase (cellobiosan, cellotriosan). The condensed phase is an ionic medium and promotes the decomposition of the reduced cellulose chains (after depolymerization) by two competing reactions: β -elimination and transglycolization. The first promotes char formation, while the second promotes the production of anhydrous sugars and volatiles. They suggest that the quick removal of liquid compounds by evaporation at high heating rates minimizes the effects of dehydration and crosslinking which are more prevalent at lower heating rates.

Dauenhauer *et al.*¹¹⁰ presented convincing visual evidence (Fig. 10) on the formation of the condensed phase. For the first time, a photographic record was made using a high-speed camera to monitor the evolution of cellulose during fast pyrolysis. For this study, 300 μm particles were pyrolyzed using a hot plate reactor at 700 °C. The melt phase begins to form at 50 ms (see Fig. 10), and it is completely formed after 120 ms. Inside the melted phase, intense bubbling and evaporation



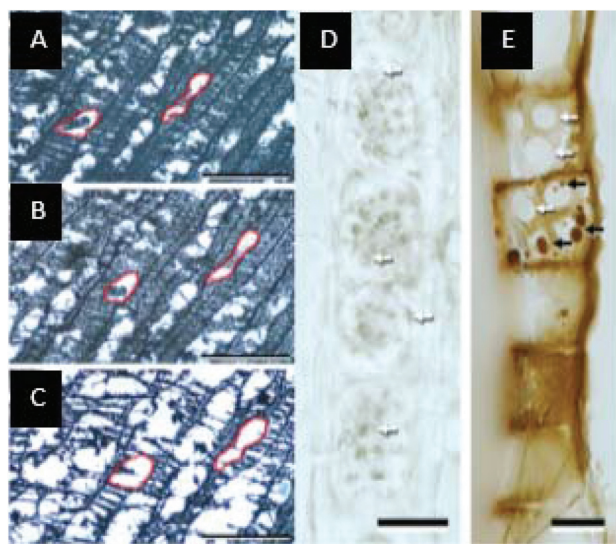


Fig. 11 Images of the evolution of the biomass during the slow pyrolysis process. (A) 260 °C, (B) 299 °C, and (C) 501 °C. Red outlines of xylem cells track individual areas through each frame. (D) Longitudinally sectioned control poplar showing cell wall faces of ray parenchyma, with white arrows pointing to examples of cell wall pores (plasmodesma). (E) Longitudinally sectioned hot-stage poplar shows cell wall faces of ray parenchyma with collected pyrolysis products (black arrows). Reprinted (adapted) with permission from Haas *et al.*²⁶³ Copyright (2009) American Chemical Society.

promote aerosol ejection and shrinking of the liquid phase. This phase is completely consumed after 150 ms, without a trace of char formation.

Inside the intermediate liquid phase, gas bubbles are formed by chemical reactions and evaporation. The intensity of bubbling depends of the liquid temperature, heating rate, pressure and nature of the liquid phase.^{260–262} The bubbles bursting promotes aerosol ejection by a reactive boiling ejection mechanism.²⁴⁴ These aerosols can be trapped inside the cell walls. Evidence of this was provided by Haas *et al.*²⁶³ through *in situ* visualization inside the cell walls which showed the presences of oligomeric liquids (see Fig. 11). In this study biomass pyrolysis was monitored in real time *via* light microscopy and TEM. Observations showed swelling and tissue collapse in poplar wood fiber and the appearance of droplets of liquid encapsulated in the biomass pores, due to impingement of aerosols within the cell walls (see Fig. 11). The existence of the liquid phase within the cell walls has critical implications for attempts to define mass and energy transfer during pyrolysis.

In 2011, Teixeira *et al.*²⁴⁴ used a high-speed camera to monitor the pyrolysis of cellulose and sucrose. The authors visualized the mechanism of aerosol formation by bubbles bursting in the intermediate liquid phase and devised a new mechanism called *reactive boiling ejection* to explain the results. Visualization of the ejection of droplets from the condensed phase of the sucrose during pyrolysis is presented in Fig. 12, along with cartoon representations to more clearly illustrate the process. The ratio between bubble size before

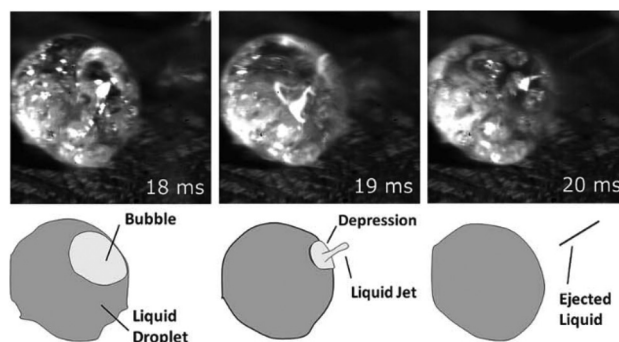


Fig. 12 Ejection mechanism of liquid droplets from the intermediate liquid phase originating from the fast pyrolysis of sucrose (top) with cartoon representations of the observed behavior (bottom). Republished with permission of the Royal Society of Chemistry, Teixeira *et al.*,²⁴⁴ permission conveyed through Copyright Clearance Center, Inc.

bursting and aerosol size ejected is close to 0.1. The ejection velocity of the liquid droplets was 0.1 m s^{-1} and had an average diameter of $1 \mu\text{m}$. The composition of the aerosols was primarily cellobiosan. With this information a mathematical model was developed to predict the mechanism for the formation of aerosols when a bubble bursts.

In 2012, Carlsson *et al.*,²⁶⁴ examined *in situ* pyrolysis of hardwood with a particle size of $300 \mu\text{m}$. They captured the behavior and evolution of the condensed as the sample was heated phase (see Fig. 13). They found that during biomass

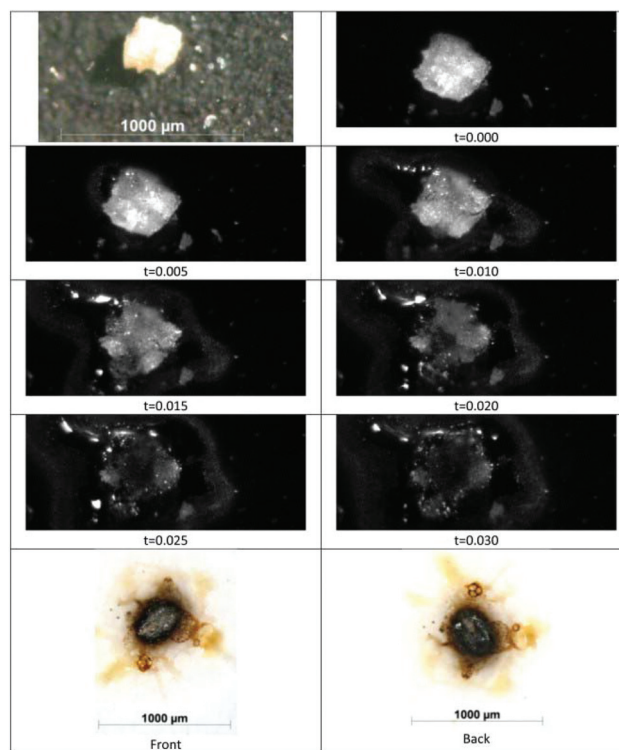


Fig. 13 Evolution of the intermediate liquid phase during the fast pyrolysis of hardwood. Reprinted from Carlsson *et al.*,²⁶⁴ Copyright (2013) with permission from Elsevier.



pyrolysis, the tissues swelled, and formed a brown film that covered the biomass surface. This film was water-soluble and had different levels of transparency as the pyrolysis process progressed. Zhu *et al.*²⁶⁵ visualized soybean waste pyrolysis in a quartz capillary at 400 °C. This technique allows *in situ* monitoring of biomass pyrolysis during heating. There was a change in color as the reaction progressed, resulting in the formation of a liquid with an oily appearance at 240 °C. This liquid became darker as the temperature increased due to decomposition and polymerization reactions.

In 2013, Dufour *et al.*¹⁵² pyrolyzed biomass (miscanthus) and its biopolymer components in a parallel hot plate rheometer specifically adapted for *in situ* tracking of the viscoelastic behavior of the intermediate liquid phase formed during pyrolysis of biomass and its components. Experiments showed that melted lignin has an elastic behavior at 200 °C, hemicellulose at 270 °C, cellulose at 300 °C, and biomass at 280 °C. The elastic behavior of the biomass melt phase could not be estimated as a combination of each of its components. A valuable observation was that viscous behavior was identified after the onset of elastic behavior, *i.e.* structural changes occur first which correspond to the glass transition and then bond breaking occurs to form a viscous fluid phase. The researchers also found that cellulose imposes its viscoelastic behavior on biomass during thermal degradation. This viscoelastic behavior was found to be variable; after 400 °C all materials increased their elastic and viscous moduli. This indicates that a rigid phase formed at higher temperatures which, in the case of pyrolysis, corresponds to the char formation. These results were validated and compared with ¹H-NMR.

Wang *et al.*⁵¹ and Liu *et al.*⁵⁰ found that amorphous cellulose produces a higher content of the intermediate liquid phase, and that this formation begins at lower temperatures compared to crystalline cellulose. Amorphous cellulose produces a variety of anhydrous sugars with high degrees of polymerization, while the crystalline cellulose is rich in monomers and dimers of glucose. One explanation given by the authors was that there is a protective effect of the crystal lattices that gives greater rigidity to the structure and prevents the breaking of glucose rings. In 2016, Westerhof *et al.*²⁶⁶ showed that depolymerization of cellulose produces sugar oligomers in the liquid phase that are composed of anhydrosugars with a degree of polymerization >2. At high heating rates and under vacuum (2 mbar_{abs}) these compounds escape by evaporation and aerosol ejection without producing char or permanent gases, indicating that for cellulose pyrolysis these fractions are produced by secondary reactions. The yield of levoglucosan increases when operating at 1 bar, because the boiling point of heavy sugars increases which reduces the evaporation rate of these compounds and allows depolymerization reactions to progress until forming levoglucosan, which evaporates.

Zhou *et al.*¹⁸⁴ showed the importance of heating rate on the intermediate liquid phase formation during lignin pyrolysis. Using a fast speed camera the formation of intermediate liquid phase was observed in 3 stages: complete lignin melting, foaming and swelling of liquid phase by bubbles, and

shrinking by evaporation and bubble collapse. Pecha *et al.*^{52,82,206} published a series of manuscripts in 2017 detailing experiments which studied the effect of vacuum on pyrolysis of cellulose, lignin, and xylan. These works illustrated, through high speed photography and pyrolysis product analysis, that the liquid intermediate is made up of partially pyrolyzed oligomers which will either break down into monomers or turn into char at atmospheric pressure. Further studies with Montoya *et al.*^{207,267,268} demonstrated protocols to analyze and model ejection of aerosols from a thin film of pyrolyzing pseudo-biomass components.

In summary, it has been demonstrated that the intermediate liquid phase during biomass pyrolysis does exist, especially under high heating rate conditions; however, this phase has not been adequately monitored experimentally. Particle-level mathematical models for both the bubble formation and thermal aerosol ejection mechanisms have not yet been completely explored. Other mechanisms for ejection of aerosols have not been studied, like viscous movement of liquid films along the inner walls of biomass pores.

Vaporization

As oligomeric compounds depolymerize into dimers and monomers with high vapor pressures and boiling points at pyrolysis temperatures, they leave the liquid intermediate as a vapor. For example, the boiling points of pyrolytic cellulose sugars levoglucosan, cellobiosan, and cellotriosan at 1 atm are 385 °C,²⁶⁹ 580 °C,²⁶⁹ and 790 °C,²⁷⁰ respectively. Oja and Suuberg have studied the vapor pressures of polycyclic aromatic hydrocarbons similar to lignin dimers and oligomers^{271,272} as well as other sugar monomers,²⁷³ as illustrated in Fig. 14.

Recent studies have elucidated the effect of vaporization on the nature of products from pyrolysis of cellulose, lignin, and wood. In cellulose, for example, lower pressures allow for larger sugar compounds from cellulose pyrolysis to evaporate (like cellobiosan and cellotriosan) and avoid conversion into polyaromatic rings and eventually char *via* dehydration and polycondensation reactions, as illustrated in Fig. 15 with high-

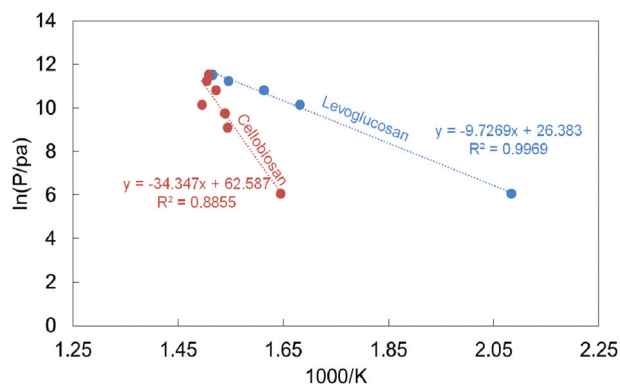


Fig. 14 Vapor pressures of levoglucosan and cellobiosan with temperature illustrate how much more rapidly levoglucosan evaporates than heavier sugars. Reprinted from Pecha,⁵⁸ Copyright (2017) with permission.



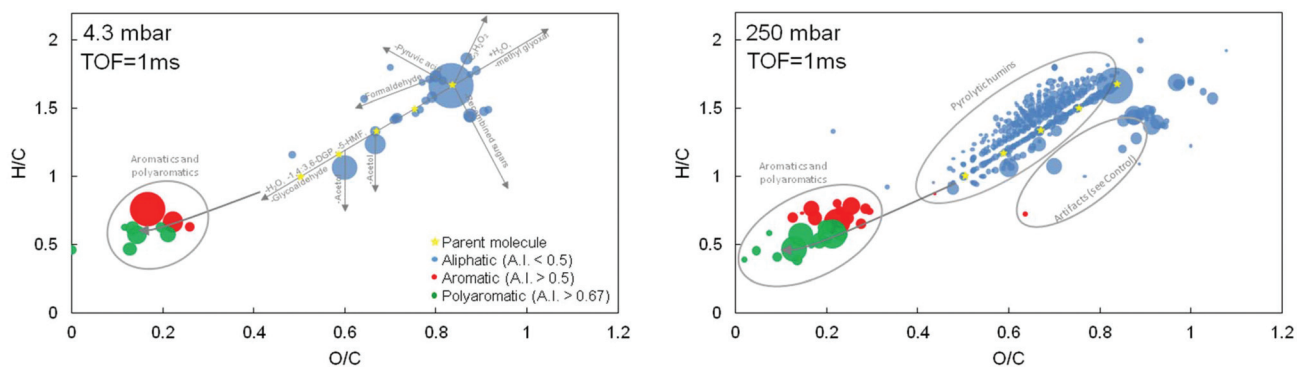


Fig. 15 Van Krevelen plots for cellulose at two pressures (4.3 and 250 mbar_{abs}) based on high resolution mass spectrometry illustrate the pathway for conversion of sugars into aromatics and char in the liquid intermediate if they cannot evaporate. Note that stars represent sugars like cellobiosan.⁵² Reprinted from Pecha,⁵⁸ Copyright (2017) with permission.

resolution mass spectrometry of high molecular weight products.⁵² Similar trends were observed for lignin and poplar, where the char yield increases as pressure increases due to evaporation of compounds at lower pressures.²⁰⁶ Some work has also been done to characterize evaporation of pyrolysis oil droplets, which may be used to model heat of vaporization in simulations.^{241,274} Lumped reaction schemes include vaporization rate and enthalpy in the step for conversion of biopolymer to “oil”, but because of this they may not be extendible to significantly different pressures.

Char formation

Char is made up of a solid polycyclic aromatic structure and forms through both intra- and intermolecular rearrangement reactions.^{275,276} Some key steps include formation of aromatic benzene the combining of these rings into polycyclic clusters that generates the rigid hierarchical structure.⁵³ Furthermore, char formation reactions are accompanied by release of water and light gas.^{77,270,277} Char formation reactions in lignin proceed through aldol condensation²⁷⁸ and radical induced cross-linking.²⁷⁹ Carbonization of cellulose is thought to proceed through oxygen-mediated reactions like aldol condensation at temperatures as low as 200 °C²⁷⁶ and free radical cross-linking and condensation reactions. Aromatic rings from cellulose include furans, pyrans, and 5–6 membered anoxic aromatic rings.²⁷⁶ C–C cross-linking into polyaromatic structures occurs as low as 400 °C; aromatic condensation increases until 500 °C. There is considerable increase in ring size as temperature increases to 700 °C.²⁸⁰ Evidence that trapped heavy oligomeric products form char can be seen in Fig. 16, where cellobiosan forms more char than levoglucosan due to its lower vapor pressure.⁵² Similar results have also been reported for lignin and wood.²⁰⁶

It is well known that char largely retains the tissue structure of its native lignocellulosic feedstock.^{280,281} However, if pyrolysis proceeds through a melt phase it will undergo alterations to its pore structure at some scales. This implies that biopolymers can cross-link before lysing, or that some other phenomenon is in play. Recent research actually indicates that

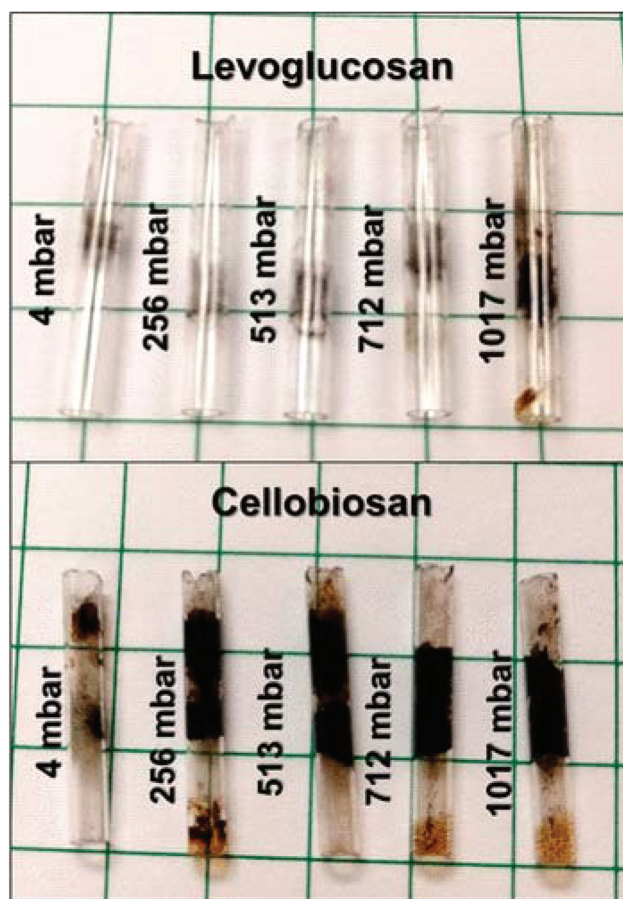


Fig. 16 Char yields from pyrolysis of levoglucosan and cellobiosan at various low pressures up to 1 atm.⁵⁸ Reprinted from Pecha,⁵⁸ Copyright (2017) with permission.

metal (“ash”) content plays a role in preserving the structural porous backbone.⁸² It is possible that the ash catalyzes char formation reactions and since the ash is solid and less mobile than liquids, it acts as a template on which polycyclic rings form.



Incorporating the main phenomena into predictive models for biomass pyrolysis

Since 1940, there have been significant developments in the modelling of biomass particles under pyrolysis conditions. The complexity and coupling of the phenomena involved (transport of mass, energy, momentum, reactivity), has led to continued research and development of these models to achieve better representations of real processes. In general, modelling can be as rigorous as desired, however; this does not imply that better representations of real phenomena will result. Modelling can also be associated with different scales of study, for example, molecular, tissue, particle, or reactor scale models can be designed.²³¹ The focus of this review are the particle-level models; however, some models that have developed for biomass fast pyrolysis at the reactor scale can be consulted.^{86,282–284}

Most articles published to date have considered one-dimensional non-steady state models, in one or two phases (solid–gas) with thermal equilibrium between phases (solid/gas), and global reaction schemes as described in Table 1. Some researchers suggest that, for small particles (<1 mm), the global kinetics scheme is sufficient to predict the product distribution.²⁴ However for large particles, both physical and chemical changes occur simultaneously, and it is essential to consider the coupling of both phenomena to accurately obtain the global pyrolysis rate. A summary of models reported in the literature is presented in Table 5.

Two dimensional models attempt to explicitly describe the anisotropy (different properties depending on the direction and position), which real-life biomass exhibits. These properties include thermal conductivity, permeability, density, porosity, and chemical composition. Typically, two-dimen-

Table 5 Particle scale models that account for heat and mass transport

Citation	Characteristics	Areas for improvement
Non dimensional models		
Diebold ¹⁴⁰ 1994	Explains the pyrolysis of cellulose. Predicts behavior of cellulose under thermos-gravimetric (slow pyrolysis) operating conditions such as temperature, heating rate and pressure	Does not include the mass transfer restrictions, energy, water evaporation, or gas flow. Particle sizes used in the experiment were not reported. Cannot be used to predict product composition
Moghtaderi <i>et al.</i> ²⁹⁰ 1997	Evaluates the temperature and mass loss of the biomass as a function of time. Modeled only the solid phase, considering two areas of interest: virgin biomass and char	Adopted a kinetic scheme with a single step. Mass loss of the solid to generate volatile phenomena is modeled as a sublimation of virgin biomass and it only takes into account the char-virgin biomass interface. Cannot be used to predict product composition
One-dimensional models		
Bamford <i>et al.</i> ²⁹¹ 1946	Predicts temperature and weight loss profiles. Predicts volatiles evolution rate	Cannot be used to predict product yields or composition. No sensitivity analyses were carried out. No convection term. Assumed constant physical properties
Kanury & Blackshear ²⁹² 1970	Shows that convection term is needed in heat balance equation. Suggests that burning rate depends on particle size	Can only be used to investigate the importance of convective heat transfer
Kung, ²⁹³ 1972	Predicts temperature and density profiles. Predicts volatiles release rate. Suggests that both thermal conductivity and specific heat of char, reaction heat, and particle size are important parameters in wood pyrolysis	Cannot be used to predict product yields or composition
Kanury ²⁹⁴ 1972	Predicts volatiles and gas yields. Simple model gives good agreement with experimental work	Supposes lignin pyrolysis as dominant step
Maa and Bailie ²⁹⁵ 1973	Predicts under what conditions heat transfer or chemical reaction are rate controlling. Predicts reaction rate and Temperature profiles. Predicts reaction times. Derived simple expressions to estimate time reaction by using either effective thermal conductivity (large particles) or effective activation energy (small particle)	Cannot be used to predict product yields or composition. Does not predict volatiles evolution rate
Fan <i>et al.</i> ²⁹⁶ 1977	Introduces a new parameter, Lewis no., the ratio of thermal diffusivity to mass diffusivity. Predicts concentration and temperature profiles. Suggests that with a higher Lewis no., greater solid conversion is reached and there will be smaller temperature gradients within the particle. Suggests that heat of reaction affects the pyrolysis rate	Cannot be used to predict product yields or composition. Does not predict volatile evolution rate
Kansa <i>et al.</i> ²⁹⁷ 1977	Predicts mass loss, pressure, and temperature profiles. Suggests that reaction heat, heat transfer by conductivity, specific heat of char and the permeability are important parameters in wood pyrolysis	Cannot be used to predict product yields or composition. Does not predict volatiles release rate
Pyle and Zaror ²⁹⁸ 1982	Introduces two pyrolysis numbers, Py I (ratio: reaction time to heat penetration time) and PyII (Biot/Py I). Evaluates the importance of external and internal heat transfer. Predicts conversion and temperature profiles. Predicts conversion time. Carried out sensitivity study to check the particle size effect on conversion	Cannot be used to predict product yields. Does not predict volatiles release rate. No convection term



Table 5 (Contd.)

Citation	Characteristics	Areas for improvement
Becker <i>et al.</i> ²⁹⁹ 1984	Predicts mass loss profiles. Predicts surface and internal temperature profiles. Predicts char yields	Does not predict volatiles yields. Does not predict volatiles release rate. Assumes physical properties constant. For large particles, kinetics and heat transfer are not coupled. Cannot be used to predict product yields or composition. Does not predict volatile release rate
Antal ³⁰⁰ 1985	Predicts temperature profiles. Predicts weight loss. Calculates reaction times. Derives simple expressions to calculate heat up time and devolatilization time in pyrolysis	Predicts product yields and composition. Predicts volatiles release rate. Predicts temperature profile. Predicts moisture effects. Carried out sensitivity studies
Chan <i>et al.</i> ³⁰¹ 1985	Predicts optimal conditions for product yields (parametric study). Predicts temperature profiles and optimum heating rates in hollow fibrous structure of wood. Primary and secondary reactions are included	Neglects mass transfer resistance, which may play a role in pyrolysis
Hastaoglu & Berruti ³⁰² 1989	Predicts high temperature drying profiles at $T > 150$ °C. Applied to wet particles up to the free-water continuity point	Specific to flash pyrolysis. Cannot be used to predict product yields or composition
Alves ³⁰³ 1989	Predicts under what conditions heat transfer or chemical reaction is rate controlling. Introduces a thermal Thiele Modulus (ratio of heat penetration time to reaction time). Predicts temperature profile. Predicts reaction time	Neglects water vapor diffusion, bound water, diffusion, and internal pressure gradients. Dimensions in longitudinal and transverse directions must be similar. Cannot be used to predict product yields or composition. Does not predict volatiles evolution rate. Thermal properties are constant
Lédé <i>et al.</i> ³⁰⁴ 1994	Transient, one-dimensional for the fast pyrolysis of wood. Proposes a 3-stage kinetic model to predict the overall composition of char, gas and volatile, and a two-stage model to explain the secondary reactions of the volatiles out of the particle. The model considers thermal equilibrium between solid and gas, ideal gas behavior, change of properties with biomass conversion and convective, diffusive transport of gases, heat transfer by conduction, convection and radiation, and particle shrinkage. Predicts the temperature profiles, velocity, and overall concentration of species inside the particle as a function of position and time	Does not include condensed liquid intermediate phase, volatile species evolution, or the effect of the heating rate on the reaction rate expression
Di Blasi ²⁸⁵ 1996	Transient model for the pyrolysis of large solid fuels (wood, coal, <i>etc.</i>). The model simultaneously evaluates the devolatilization process and drying. Kinetic model is a single stage scheme	Does not evaluate convective, diffusive processes or water vapor within the particle; <i>i.e.</i> instantaneous evaporation is supposed on the front of reaction at constant temperature 100 °C. Ignores secondary reactions, evolution of species, liquid intermediate phase
Richard ³⁰⁵ 1996	Transient, for a flat particle, thin and dry biomass, which is subjected to an external source of heat in an inert atmosphere. Is considered changing the porosity and volume of the solid matrix. The pressure gradients are estimated using Darcy's law. Predicts the temperature development, and condensable volatile fraction of non-condensable volatiles escape velocity, depending on particle size, position (spatial gradients) and reaction time	Does not take into account the effect of the intermediate liquid; disregards the effect of the heating rate and evolution of species, including secondary reactions of the volatiles. Cannot be used to predict composition of products
Di Blasi ²¹¹ 2002	Unsteady state to predict the behavior of biomass pyrolysis. Includes heat transfer by conduction and convection and mass transfer by diffusion and convection. For different particle sizes and shapes, the model predicts overall yields of char, volatiles, and permanent gases. The author found that the pyrolysis of small particles (<1 mm) and spherical produces more volatile and the reaction time is shorter compared to flat and cylindrical shapes. For large particles (1 cm) the reaction time increases and the char yield is independent of the external heating rate, although it depends on the geometry of the particle	Does not predict volatiles species evolution rate. Applied for slow heating rates
Van de Weerdhof ²⁸ 2010	The model, which accounts for the combined effect of various parameters such as particle shrinkage and drying, predicts temperature profiles, drying rate, char, and volatiles yields	Local thermal equilibrium (temperature gradient) was not considered between vapor and solid phase. Re-condensation of volatiles in cooler regions of the particle is not taken into account, due to higher permeability of char as compared to solid biomass. The volatile/gas phase follows ideal gas behavior. The kinetic and potential energies of the particle were neglected in the energy balance. The reaction heats for gases and secondary tar formation were considered equal. Particle shape did not change during the degradation process, <i>i.e.</i> cracking and fragmentation not considered
Sharma ³⁰⁶ 2014		



Table 5 (Contd.)

Citation	Characteristics	Areas for improvement
Two dimensional models		
Di Blasi ¹³⁵ 1998	Energy transfer by conduction, convection and radiation; mass transfer by convection; diffusion of momentum using Darcy's law. This model estimates physicochemical changes in two dimensions (anisotropy). The results were compared with those of a one-dimensional model with non-steady state. The comparison between the two models shows that the reactions are faster and produce more volatile for configurations in two dimensions. An interesting feature of this work is the evaluation of the evaporation of moisture within the model and studying different schemes of shrinkage of the particle as uniform shrinkage, shrinkage layers (cell shrinkage), and shrinkage by cylinders. Showed that the shrinkage model affects the calculated heating rate, which, depending on the geometry, may have a positive or negative effect	This model proposes multi steps overall reaction scheme, but does not include the formation of intermediate liquid phase, the formation of side reactions, or the evolution of species
Bellais ³⁰⁷ 2007	Predict the pyrolysis of biomass cylindrical large particles (order of cm). Includes changing properties of the conversion and the direction (anisotropy), and evaluates the mechanisms of heat and mass transfer by diffusion and convection as well as pressure profiles <i>via</i> Darcy's law. This model adopts multistage overall reaction schemes, regards secondary reactions, and evaluates the effect of the condensed phase on chemical reactions	Does not consider gaseous species evolution such (CO, CO ₂ , H ₂). The formation of intermediate liquid phase is not considered. Limited to slow pyrolysis conditions
Kersten group ^{235,308} 2005 Zeng <i>et al.</i> ³⁰⁹ 2007	Single particle models (1D and 2D) compared to experiments 2-d cylindrical model for concentrated solar radiation pyrolysis; simple reaction scheme with "intermediate solid" stage	Lumped kinetics, no coupling to secondary vapor phase reactions Isotropic intraparticle transport; only 4 product species in reaction scheme
Okekunle <i>et al.</i> ²³⁹ 2012	Effect of particle size and aspect ratio on tar decomposition. Predict temperature profiles and provides the char, volatiles and gas yields. Compares the temperature profiles and char yields considering isotropic and anisotropic structures	Does not predict the rate of water drying. Doesn't include mass transfer by diffusion. Particle shrinking is not included
Paulsen ²⁰³ 2014	Predict temperature profiles, drying rate, and particle shrinkage. The carbohydrates profiles are predicted inside the particle. Classify the solid particle as: unreacted carbohydrate, virgin biomass and char	Isotropic particle. Mass transfer by diffusion is neglected. Ideal gas behavior. Secondary reactions not taken in account
Anca-Couce <i>et al.</i> ¹⁵⁷ 2017	2-d cylindrical single particle model incorporating highly speciated RAC reaction scheme and secondary reactions in homogenous phase of reactor. Predicts trends for compound release for common monomers better than any other available reaction scheme	Some notable errors in yield predictions; reaction scheme does not account for oligomeric compounds
Pecha <i>et al.</i> ³¹⁰ 2018	2-d cylindrical model with Di Blasi scheme. Distribution of particle size and morphology. Microscopy-informed internal properties. Coupling with reactor scale model for residence times of various sized particles and vapor residence time. Close prediction to experiment without any fitted parameters	Simplistic Di Blasi reaction scheme gives no semblance of bio-oil quality
Three dimensional models		
Leonardi ³¹¹ 2007	Spherical particles. Transient model for pyrolysis of wet biomass. This model is stringent because it includes water evaporation spatial changes in temperature, concentration, velocity profiles, and changes in physicochemical properties (anisotropy)	The reaction scheme is a global model adopted in one step without side reactions in the gas phase, gas-solid reactions, or the development of species of condensable and non-condensable volatiles. Nor is the formation of a liquid phase and its respective resistance to the processes of energy and mass transport included
Mahmoudi ³¹² 2014	Numerical study based on the Lattice-Boltzmann Method (LBM) is proposed to solve one, two, and three-dimensional heat and mass transfer for isothermal carbonization of thick wood particles. The model is used to study the effect of reactor temperature and thermal boundary conditions on the evolution of the local temperature and the mass distributions of the wood particle during carbonization. Takes into account diffusive mechanisms for volatiles through this phase	Does not include secondary reactions. The physical properties do not depend on the conversion degree of the biomass. Does not consider evolution of species or the effect of the heating rate on the reaction rate



Table 5 (Contd.)

Citation	Characteristics	Areas for improvement
Gentile <i>et al.</i> ³¹³ 2017	Highly speciated Ranzi reaction scheme with 3D spherical and cylindrical model; moving mesh particle shrinking in OpenFOAM; decent matchup with experiments; transport anisotropy	Shrinking does not match up well with experiments, predicts high velocity from sides of particle; reaction scheme does not include liquid intermediate or oligomeric products
Models with explicit intermediate liquid phase Oh <i>et al.</i> ²⁸⁷ 1989	Model applied for coal pyrolysis with high swelling (softening coals). Includes heat and mass transfer through the condensed phase (meta plastic) <i>via</i> a bubble growth model uniformly distributed (population balance). The behavior is in transient state. The model predicts the production of tar, permanent gases, char, and evolution of the condensed phase as a function of time and temperature	Applied for coal. No spatial gradients of temperature or concentration of species. The overall kinetic scheme is a single stage and does not evaluate the evolution of individual species
Sezen ³¹⁴ 1989	Presents a model for studying transient dimensional fast pyrolysis of spherical coal (coal softening). <i>This model is the most rigorous</i> we have found which includes the effects of heat and mass transfer in the condensed phase. The model considers diffusive and convective transport of volatiles, heat transfer by conduction and convection, and uses a multi-stage kinetic mechanism	Does not study the evolution of individual species and their respective secondary reactions. Applies only for coal
Di Blasi ²⁸⁵ 1996	Model for the pyrolysis of cellulose cylindrical particles in an ablative reactor (spinning-disk) as described by Ledé <i>et al.</i> ²⁵¹ The model developed is transient, and one-dimensional, using a Broido-Shafizadeh kinetic scheme. It does not regard side reactions or effects of diffusion and convection on volatiles. This proposed modeled considers the formation and evaporation of the condensed liquid intermediate	Does not consider diffusive mechanisms of volatiles through liquid phase. Does not consider the evolution of species or effect of the heating rate on the reaction rate
Ghazaryan ³¹⁵ 2000	Aerosol formation through bubble nucleation followed by condensation, evaporation, and collision within porous material	This model only considers aerosol filtration but does not consider the formation of gases through pyrolysis and ignores secondary reactions that produce char (changing the structure). The structural arrangement is very simple and may not reflect the real structure of lignocellulosic biomass
Boutin ¹³¹ 2002	Model for the fast pyrolysis of cellulose pellets subjected to radiative heating. The model considers the formation and consumption of the intermediate liquid phase. This is a one-dimensional model, split in 3 areas (virgin biomass, liquid phase and char). Each zone formation is estimated by mass and energy balances, and the interface boundary conditions give continuity to the variables. The model only predicts the temperature profiles and the evolution of global species over time	Does not consider secondary reactions or the loss of moisture from the biomass, pressure gradients, or convection and mass transfer through each reaction zone
Bounaceur ²⁵⁷ 2011	Intra-particle model for the pyrolysis of biomass fast pyrolysis under non-steady state. Includes the formation and consumption of the intermediate liquid phase. Evaporation of the liquid phase is modeled with a Clausius–Clapeyron expression and Darcy's law for estimating the volatile release. The model predicts the overall yields of species depending on the heating rate, and reaction time, and estimates the time and temperature where maximum conversion gives the intermediate liquid phase	The model does not consider spatial change (zero dimensional), so the mechanisms of conduction heat transfer, mass transfer by diffusion and convection, are not considered
Teixeira ²⁴⁴ 2011	2D-CFD modelling to explain boiling ejection in aerosol generation. Navier–Stokes equations are solved using finite volume. Models interface transport and capillary effects with the Eulerian volume of fluids (VOF) method. The current approach is capable of handling topological changes in the interface, such as the formation of droplets and bubbles. The simulation specifically considers ejection from the curved interface of a droplet, rather than an infinite pool with a flat surface. The model predicts the aerosol ejection velocity, aerosol size and distribution, and physical properties of molten phase	Only for cellulose pyrolysis. The populations to describe the coalescence, nucleation, breakage of bubbles formed inside the molten phase are not clear
Montoya <i>et al.</i> ²⁶⁷ 2016	A slab of particle on a hot plate was modelled with a statistical model for bubbling frequency and growth in the liquid intermediate. Collision of aerosols along pore walls was considered such that large particles release less aerosols. Particle shrinking was considered. Yields of oligomeric compounds were close to those observed experimentally	Difficult to validate bubbling model because it is not possible to visualize this phenomenon inside particles



sional models are represented by cylinders. Three-dimensional models further expand the anisotropy in a third dimension. Typically, three-dimensional models do not improve the accuracy of results enough to justify the extra amount of computational time required. Although many single particle models have been developed, only a few have looked at the explicit species-specific microporosity of cell walls.^{93,214} See Table 5 for examples.

The majority of models at the particle level developed until now do not consider the intermediate liquid phase evolution. They also do not include other phenomena like gas bubble formation inside the liquid that explain the mechanism for aerosol formation.^{183,244} There are models proposed by Dufour *et al.*,²⁵⁷ Boutin *et al.*,¹³¹ Di Blasi,²⁸⁵ Lédé *et al.*,²⁵¹ and Brown *et al.*²⁸⁶ which take into account the physiochemical properties of the liquid intermediate phase. However, these models still do not consider bubble formation and aerosol formation. Dufour incorporated evaporation of the liquid intermediate phase into the model. A model describing the levoglucosan was published in which the competition between evaporation and poly-condensation reactions was described.²⁸⁶ Recently, Montoya *et al.*²⁶⁸ developed a single particle model which incorporates thermal ejection of aerosols, though more work needs to be done to advance this topic.

Today, models for cellulose and biomass do not exist which include the liquid phenomena of gas bubbling and aerosol ejection, which are thought to be the primary sources for large molecules that are always observed in the bio-oil. However, these phenomena have been included for models with other raw materials like coal and polymers;^{261,287} these models describe bubble formation using a discrete phase population balance and couple this information with the global energy balance and mass transfer inside the particle. The important conclusion of these papers is that modelling the presence of the liquid intermediate dramatically changed the results from a model that ignores the liquid intermediate. In pyrolysis, this will impact the secondary reactions inside the particles. One important phenomenon unaccounted for in these models ignored is the formation of aerosols from bubble explosion at the surface of the liquid phase. See Table 5 for more detailed information about these models. Further discussion on coupling atomic scale models, single particle models, and reactor scale models can be seen in Ciesielski *et al.*²⁶ and a focus on reactor scale modelling in Xiong *et al.*^{288,289}

Conclusions

From this review, the following conclusions and recommendations can be drawn:

- Bio-oil production by pyrolysis is still an immature technology and it is not currently commercially viable. Bio-oil production technologies still have to overcome many technical and economic barriers to compete with traditional fossil fuels and diversify its spectrum of value-added products.
- Fast pyrolysis is a complex process, with many interdependent phenomena including chemical reactions. Heating rate effects mass and energy transport inside particles. The

real temperature in the particle is not identical to surface temperature. The particle structure changes through the pyrolysis process. The evolution of the intermediate phase (liquid phase) impacts product yields.

- New experimental methodologies to measure different parameters such as temperature, gas flow, and chemical composition have recently added to the scientific understanding of biomass pyrolysis, particularly with regards to the liquid intermediate phase and the oligomeric fraction of pyrolysis oil. However there is still limited experimental information regarding this phase, its impact on the evolution of species condensable and non-condensable volatile, and its incorporation into physical models.
- Recent studies have shown that aerosols rich levoglucosan dimers and trimers and lignin oligomers are formed directly from the ejection of fluid from the intermediate liquid phase. This phenomenon explains the presence of heavy compounds in bio-oil recovered from the fast pyrolysis of biomass, as well as the agglomeration and crusting of biomass particles and inert material in the reactor. Experiments at the laboratory level and rigorous models are still needed to understand many of the fundamentals associated with bubble formation and aerosol ejection.
- The integration of reaction and transport phenomena in a single particle model is not a trivial undertaking, with the validation the model an even more challenging task. Future pyrolysis models for particles must be able to predict volatilization as a function of time for different biomass feedstocks (necessary to design the reactor and optimize operating conditions) and predict the distribution of pyrolysis products. These models are likely to be based on existing transport models for particles, but must include more detailed estimation methods for critical transport parameters (*e.g.*, thermal conductivity, heat capacity, liquid and gas viscosity *etc.*), condensed phase thermodynamics, and the reaction kinetics. These models should also be non-dimensionalized to ease application into reactor-scale models.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was authored in part by Alliance for Sustainable Energy, LLC, the manager and operator of the National Renewable Energy Laboratory for the U.S. Department of Energy (DOE) under Contract No. DE-AC36-08GO28308. Funding provided by the U.S. Department of Energy Office of Energy Efficiency and Renewable Energy Bioenergy Technologies Office. Jorge Montoya thanks Colciencias for the financial support during the development of the PhD with the call 526, and during the postdoctoral stay with the Call 784. Farid Chejne would like to thank 'Strategy of transformation of the Colombian energy sector in the horizon 2030' funded by the call 788 of Colciencias Scientific Ecosystem. Contract number FP44842-210-2018.



References

- IEA, *Key World Energy Statistics 2010*, OECD Publishing, 2010.
- G. Akhmat, K. Zaman, T. Shukui and F. Sajjad, *Renewable Sustainable Energy Rev.*, 2014, **36**, 123–134.
- International Energy Agency and I. E. Agency, *Key World Energy*, OECD Publishing, 2011.
- E. C. Marín, H. S. Mahecha and S. P. Carrasco, *Dyna*, 2009, **76**, 101–110.
- S. Nakada, D. Saygin and D. Gielen, *Global bioenergy supply and demand projections: a working paper for REmap 2030*, 2014.
- R. E. Davis, N. J. Grundl, L. Tao, M. J. Bidy, E. C. Tan, G. T. Beckham, D. Humbird, D. Thompson and M. S. Roni, *Process Design and Economics for the Conversion of Lignocellulosic Biomass to Hydrocarbon Fuels and Coproducts: 2018 Biochemical Design Case Update; Biochemical Deconstruction and Conversion of Biomass to Fuels and Products via Integrated Biorefinery Path*, National Renewable Energy Lab. (NREL), Golden, CO, USA, 2018.
- A. Dutta, R. M. Swanson, A. Aden, J. A. Satrio, M. M. Wright, A. Platon, D. E. Dugaard, R. C. Brown, D. D. Hsu, R. P. Anex, F. K. Kazi, G. Kothandaraman and J. Fortman, *Fuel*, 2010, **89**, S29–S35.
- E. Henrich, F. Schultmann, F. Trippe, R. Stahl and M. Fröhling, *Waste Biomass Valorization*, 2010, **1**, 415–430.
- A. Dufour, C. Rogaume, Y. Rogaume, P. Girods and A. Zoulalian, *J. Anal. Appl. Pyrolysis*, 2009, **85**, 171–183.
- M. C. Tirado, M. J. Cohen, N. Aberman, J. Meerman and B. Thompson, *Food Res. Int.*, 2010, **43**, 1729–1744.
- P. de Wild, H. Reith and E. Heeres, *Biofuels*, 2011, **2**, 185–208.
- H. L. Chum, *Biomass Pyrolysis to Hydrocarbon Fuels in the Petroleum Refining Context: Cooperative Research and Development Final Report, CRADA Number CRD-12-500*, National Renewable Energy Lab. (NREL), Golden, CO, United States, 2018.
- J. Lehmann and S. Joseph, *Biochar for Environmental Management: Science and Technology*, Earthscan, Washington DC, 2009.
- M. Garcia-Perez, J. Garcia-Nunez, T. Lewis, C. Kruger and S. Kantor, *Methods for Producing Biochar and Advanced Biofuels in Washington State. Part 3: Literature Review of Technologies for Product Collection and Refining*, Department of Biological Systems Engineering and the Center for Sustaining Agriculture and Natural Resources, Washington State University, Pullman, WA, 2011.
- J. Hyväluoma, S. Kulju, M. Hannula, H. Wikberg, A. Källi and K. Rasa, *Environ. Sci. Pollut. Res.*, 2017, 1–11.
- J. A. Garcia-Nunez, D. T. Rodriguez, C. A. Fontanilla, N. E. Ramirez, E. E. Silva Lora, C. S. Frear, C. Stockle, J. Amonette and M. Garcia-Perez, *Biomass Bioenergy*, 2016, **95**, 310–329.
- L. Wang, Ø. Skreiberg, S. Van Wesenbeeck, M. Grønli and M. J. Antal, *Energy Fuels*, 2016, **30**, 7994–8008.
- M. J. Antal, E. Croiset, X. Dai, C. DeAlmeida, W. S.-L. Mok, N. Norberg, J.-R. Richard and M. Al Majthoub, *Energy Fuels*, 1996, **10**, 652–658.
- M.-K. Bahng, C. Mukarakate, D. J. Robichaud and M. R. Nimlos, *Anal. Chim. Acta*, 2009, **651**, 117–138.
- W. R. Kang and K. B. Lee, *Korean J. Chem. Eng.*, 2013, **30**, 1386–1394.
- A. Sharma, V. Pareek and D. Zhang, *Renewable Sustainable Energy Rev.*, 2015, **50**, 1081–1096.
- D. W. van Krevelen, *Fuel*, 1950, **29**, 269–284.
- A. V. Bridgwater, *Fast Pyrolysis of Biomass: A Handbook*, PyNe, 3rd edn, 2008.
- D. Meier, B. van de Beld, A. V. Bridgwater, D. C. Elliott, A. Oasmaa and F. Preto, *Renewable Sustainable Energy Rev.*, 2013, **20**, 619–641.
- A. C. Cuevas, C. Reinoso and J. Lamas, in *Biomass for Energy, Environment, Agriculture, and Industry: Proceedings of the 8th European Biomass Conference, Vienna, Austria, 3rd–5th October 1994*, ed. P. Chartier, A. A. C. M. Beenackers and G. Grassi, Pergamon Press, 1995, pp. 1506–1512.
- P. N. Ciesielski, M. B. Pecha, V. S. Bharadwaj, C. Mukarakate, G. J. Leong, B. Kappes, M. F. Crowley, S. Kim, T. D. Foust and M. R. Nimlos, *Wiley Interdiscip. Rev.: Energy Environ.*, 2018, **7**, e297.
- G. Xiu-juan, W. Shu-Rong, W. Kai-Ge, L. Qian and L. Zhong-Yang, *J. Fuel Chem. Technol.*, 2010, **38**, 42–46.
- M. W. Van de Weerdhof, PhD Thesis, Eindhoven University of Technology, 2010.
- M. T. Klein and P. S. Virk, *Energy Fuels*, 2008, **22**, 2175–2182.
- D. M. Alonso, S. G. Wettstein and J. A. Dumesic, *Chem. Soc. Rev.*, 2012, **41**, 8075–8098.
- X. Zhou, W. Li, R. Mabon and L. J. Broadbelt, *Energy Technol.*, 2016, **5**, 52–79.
- P. Eronen, M. Österberg, S. Heikkinen, M. Tenkanen and J. Laine, *Carbohydr. Polym.*, 2011, **86**, 1281–1290.
- M. W. Smith, G. L. Helms, J.-S. McEwen and M. Garcia-Perez, *Carbon*, 2017, **116**, 210–222.
- L. Li, P. Pérré, X. Frank and K. Mazeau, *Carbohydr. Polym.*, 2015, **127**, 438–450.
- I. S. Goldstein, in *Organic chemicals from biomass*, CRC Press, 2018, pp. 9–18.
- P. R. Patwardhan, J. A. Satrio, R. C. Brown and B. H. Shanks, *Bioresour. Technol.*, 2010, **101**, 4646–4655.
- J. Akhtar and N. Saidina Amin, *Renewable Sustainable Energy Rev.*, 2012, **16**, 5101–5109.
- H. Yang, R. Yan, H. Chen, C. Zheng, D. H. Lee and D. T. Liang, *Combust. Flame*, 2006, **146**, 605–611.
- H. Zhu, W. Luo, P. N. Ciesielski, Z. Fang, J. Y. Zhu, G. Henriksson, M. E. Himmel and L. Hu, *Chem. Rev.*, 2016, **116**, 9305–9374.
- T. Hosoya, S. Saka and H. Kawamoto, *J. Anal. Appl. Pyrolysis*, 2007, **80**, 118–125.
- T. J. Hilbers, Z. Wang, B. Pecha, R. J. M. Westerhof, S. R. A. Kersten, M. R. Pelaez-Samaniego and M. Garcia-Perez, *J. Anal. Appl. Pyrolysis*, 2015, **114**, 197–207.



- 42 A. Dufour, M. Castro-Diaz, N. Brosse, M. Bouroukba and C. Snape, *ChemSusChem*, 2012, **5**, 1258–1265.
- 43 S. Salvador, C. Couhert and J.-M. Commandre, *Fuel*, 2009, **88**, 408–417.
- 44 E. Ranzi, A. Cuoci, T. Faravelli, A. Frassoldati, G. Migliavacca, S. Pierucci and S. Sommariva, *Energy Fuels*, 2008, **22**, 4292–4300.
- 45 W. F. Degroot, M. D. Rahman, W.-P. Pan and G. N. Richards, *J. Anal. Appl. Pyrolysis*, 1988, **13**, 221–231.
- 46 C. Branca and C. Di Blasi, *Ind. Eng. Chem. Res.*, 2006, **45**, 5891–5899.
- 47 G. R. Ponder, T. T. Stevenson and G. N. Richards, *J. Anal. Appl. Pyrolysis*, 1992, **22**, 217–229.
- 48 M. S. Mettler, A. D. Paulsen, D. G. Vlachos and P. J. Dauenhauer, *Green Chem.*, 2012, **14**, 1284.
- 49 A. Broido, A. C. Javier-Son, A. C. Ouano and E. M. Barrall, *J. Appl. Polym. Sci.*, 1973, **17**, 3627–3635.
- 50 D. Liu, Y. Yu and H. Wu, *Energy Fuels*, 2013, **27**, 1371–1380.
- 51 Z. Wang, A. G. McDonald, R. J. M. Westerhof, S. R. A. Kersten, C. M. Cuba-Torres, S. Ha, B. Pecha and M. Garcia-Perez, *J. Anal. Appl. Pyrolysis*, 2013, **100**, 56–66.
- 52 M. B. Pecha, J. I. Montoya, F. Chejne and M. Garcia-Perez, *Ind. Eng. Chem. Res.*, 2017, **56**, 4288–4301.
- 53 F. X. Collard and J. Blin, *Renewable Sustainable Energy Rev.*, 2014, **38**, 594–608.
- 54 J. R. Obst and T. K. Kirk, *Methods Enzymol.*, 1988, **161**, 3–12.
- 55 L. Yang, D. Wang, D. Zhou and Y. Zhang, *Int. J. Biol. Macromol.*, 2016, **85**, 417–424.
- 56 V. B. F. Custodis, C. Bährle, F. Vogel and J. A. van Bokhoven, *J. Anal. Appl. Pyrolysis*, 2015, **115**, 214–223.
- 57 M. Asmadi, H. Kawamoto and S. Saka, *J. Anal. Appl. Pyrolysis*, 2011, **92**, 417–425.
- 58 M. B. Pecha, PhD Thesis, Washington State University, 2017.
- 59 P. R. Patwardhan, R. C. Brown and B. H. Shanks, *ChemSusChem*, 2011, **4**, 636–643.
- 60 D. K. Shen, S. Gu and A. V. Bridgwater, *J. Anal. Appl. Pyrolysis*, 2010, **87**, 199–206.
- 61 T. Hosoya, H. Kawamoto and S. Saka, *J. Anal. Appl. Pyrolysis*, 2007, **78**, 328–336.
- 62 M. Widyawati, T. L. Church, N. H. Florin and A. T. Harris, *Int. J. Hydrogen Energy*, 2011, **36**, 4800–4813.
- 63 S. Wang, *Prog. Energy Combust. Sci.*, 2017, **62**, 33–86.
- 64 M. Sohrabi, M. R. Hajaligol, M. Nik-Azar and B. Dabir, *Fuel Process. Technol.*, 1997, **51**, 7–17.
- 65 K. Raveendran, A. Ganesh and K. C. Khilar, *Fuel*, 1995, **74**, 1812–1822.
- 66 Y. Yürüm and N. Öztaş, *Fuel*, 2000, **79**, 1221–1227.
- 67 A. W. Williams, PhD Thesis, Georgia Institute of Technology, 2011.
- 68 G. Alexander, K. Wolframg, R. Sonia, A. Gómez, W. Klose and S. Rincón, *Pirólisis de Biomasa: Cuesco de palma de aceite*, Kassel University Press GmbH, Bogotá, 2008.
- 69 A. V. Bridgwater and G. V. C. Peacocke, *Fast pyrolysis processes for biomass*, 2000, vol. 4.
- 70 P. R. Patwardhan, PhD Thesis, Iowa State University, 2010.
- 71 K. Shimomura, H. Watanabe and K. Okazaki, *Proc. Combust. Inst.*, 2012, **34**, 2339–2345.
- 72 G. N. Richards and G. Zheng, *J. Anal. Appl. Pyrolysis*, 1991, **21**, 133–146.
- 73 Z. A. Mayer, A. Apfelbacher and A. Hornung, *J. Anal. Appl. Pyrolysis*, 2012, **96**, 196–202.
- 74 G. Gellerstedt and M. Kleen, *J. Anal. Appl. Pyrolysis*, 1995, **35**, 15–41.
- 75 F. Sulaiman and N. Abdullah, *Energy*, 2011, **36**, 2352–2359.
- 76 J. E. White, W. J. Catallo and B. L. Legendre, *J. Anal. Appl. Pyrolysis*, 2011, **91**, 1–33.
- 77 J. Valette, J. Blin, F.-X. Collard and A. Bensakhria, *J. Anal. Appl. Pyrolysis*, 2012, **95**, 213–226.
- 78 E. Ranzi, P. E. A. Debiagi and A. Frassoldati, *ACS Sustainable Chem. Eng.*, 2017, **5**, 1–7.
- 79 A. Anca-Couce and R. Scharler, *Fuel*, 2017, **206**, 572–579.
- 80 T. Iwasaki, S. Suzuki and T. Kojima, *Energy Environ. Res.*, 2014, **4**, 64–72.
- 81 S. R. G. Oudenhoven, C. Lievens, R. J. M. Westerhof and S. R. A. Kersten, *Biomass Bioenergy*, 2016, **89**, 78–90.
- 82 J. Montoya, B. Pecha, F. C. Janna and M. Garcia-Perez, *J. Anal. Appl. Pyrolysis*, 2017, **123**, 307–318.
- 83 R. M. Baldwin, B. S. Donohoe, M. R. Nimlos, K. A. Magrini-Bair, J. E. Hensley, P. Pepiot and S. D. Phillips, *Appl. Catal., B*, 2012, **115–116**, 320–329.
- 84 P. Basu, *Biomass Gasification and Pyrolysis*, © 2010 Elsevier Inc., 2010, vol. 45.
- 85 J. Lédé, *Energies*, 2010, **3**, 886–898.
- 86 C. Di Blasi, *Chem. Eng. Sci.*, 2000, **55**, 5999–6013.
- 87 J. Lédé, *J. Anal. Appl. Pyrolysis*, 2012, **94**, 17–32.
- 88 D. A. I. Goring, *Pulp Pap. Mag. Can.*, 1963, **64**, T517–T527.
- 89 Y. Lin, J. Cho, G. A. Tompsett, P. R. Westmoreland and G. W. Huber, *J. Phys. Chem. C*, 2009, **113**, 20097–20107.
- 90 J. Lédé, J. P. Diebold, G. V. C. Peacocke and J. Piskorz, in *Developments in Thermochemical Biomass Conversion*, ed. Springer Science + Business Media, CPL Press, Netherlands, 1997, pp. 27–42.
- 91 P. Pineda-Gomez, C. M. Bedoya-Hincapié and A. Rosales-Rivera, *Dyna*, 2011, **76**, 207–214.
- 92 A. Melgar, D. Borge and J. F. Pérez, *Dyna*, 2008, **75**, 123–131.
- 93 P. N. Ciesielski, M. F. Crowley, M. R. Nimlos, A. W. Sanders, G. M. Wiggins, D. Robichaud, B. S. Donohoe and T. D. Foust, *Energy Fuels*, 2015, **29**, 242–254.
- 94 I. Milosavljevic, V. Oja and E. M. Suuberg, *Ind. Eng. Chem. Res.*, 1996, **35**, 653–662.
- 95 C. Di Blasi, C. Branca, F. E. Sarnataro and A. Gallo, *Energy Fuels*, 2014, **28**, 2684–2696.
- 96 C. Di Blasi, C. Branca, A. Galgano and B. Gallo, *Energy Fuels*, 2015, **29**, 2514–2526.
- 97 J. Montoya, B. Pecha, D. Roman, F. C. Janna and M. Garcia-Perez, *J. Anal. Appl. Pyrolysis*, 2016, **123**, 347–363.
- 98 R. K. Singh and B. Ruj, *Fuel*, 2016, **174**, 164–171.
- 99 J. I. Montoya, C. Valdés, F. Chejne, C. A. Gómez, A. Blanco, G. Marrugo, J. Osorio, E. Castillo, J. Aristóbulo and J. Acero, *J. Anal. Appl. Pyrolysis*, 2015, **112**, 379–387.



- 100 R. J. Westerhof, PhD Thesis, University of Twente, 2011.
- 101 M. Garcia-Perez, S. Wang, J. Shen, M. Rhodes, W. J. Lee and C. Z. Li, *Energy Fuels*, 2008, **22**, 2022–2032.
- 102 S. Zhou, M. Garcia-Perez, B. Pecha, A. G. McDonald, S. R. A. Kersten and R. J. M. Westerhof, *Energy Fuels*, 2013, **27**, 1428–1438.
- 103 P. Basu, *Biomass gasification and pyrolysis practical design and theory*, Academic Press, Burlington, MA, 2010.
- 104 R. Narayan and M. J. J. Antal, *Ind. Eng. Chem. Res.*, 1996, **35**, 1711–1721.
- 105 G. Pokol, G. Várhegyi and D. Dollimore, *CRC Crit. Rev. Anal. Chem.*, 2008, **19**, 65–93.
- 106 J. Shen, X.-S. Wang, M. Garcia-Perez, D. Mourant, M. J. Rhodes and C.-Z. Li, *Fuel*, 2009, **88**, 1810–1817.
- 107 A. V. Bridgwater, *Fast pyrolysis of biomass: a handbook*, CPL Press, Newbury, 1999.
- 108 A. S. Kalgo, PhD Thesis, Aston University, 2011.
- 109 M. Hajaligol, T. Fisher, B. Waymack and D. Kellogg, *J. Anal. Appl. Pyrolysis*, 2002, **62**, 331–349.
- 110 P. J. Dauenhauer, J. L. Colby, C. M. Balonek, W. J. Suszynski and L. D. Schmidt, *Green Chem.*, 2009, **11**, 1555–1561.
- 111 W. C. Park, A. Atreya and H. R. Baum, *Combust. Flame*, 2009, **157**, 481–494.
- 112 D. S. Scott, J. Piskorz, M. A. Bergougnou, R. P. Overend and R. Graham, *Ind. Eng. Chem. Res.*, 1988, **27**, 8–15.
- 113 P. Fu, S. Hu, J. Xiang, P. Li, D. Huang, L. Jiang, A. Zhang and J. Zhang, *J. Anal. Appl. Pyrolysis*, 2010, **88**, 117–123.
- 114 M. Garcia-Perez, *The Formation of Polyaromatic Hydrocarbons and Dioxins During Pyrolysis: A Review of the Literature with Descriptions of Biomass Composition*, Fast Pyrolysis Technologies and Thermochemical Reactions, Pullman, WA, 2008.
- 115 W. Chaiwat, I. Hasegawa, T. Tani, K. Sunagawa and K. Mae, *Energy Fuels*, 2009, **23**, 5765–5772.
- 116 V. Agarwal, P. J. Dauenhauer, G. W. Huber and S. M. Auerbach, *J. Am. Chem. Soc.*, 2012, **134**, 14958–14972.
- 117 S. Zhou, M. Garcia-Perez, B. Pecha, A. G. McDonald and R. J. M. Westerhof, *Fuel*, 2014, **125**, 15–19.
- 118 R. J. M. Westerhof, D. W. F. Brilman, M. Garcia-Perez, Z. Wang, S. R. G. Oudenhoven, W. P. M. Van Swaaij and S. R. A. Kersten, *Energy Fuels*, 2011, **25**, 1817–1829.
- 119 J. Diebold and J. Scahill, in *Fundamentals of Thermochemical Biomass Conversion*, ed. R. P. Overend, T. A. Milne and L. K. Mudge, Springer Netherlands, 1985, pp. 539–555.
- 120 O. Onay, *Fuel Process. Technol.*, 2007, **88**, 523–531.
- 121 S. Thangalazhy-Gopakumar, S. Adhikari, R. B. Gupta and S. D. Fernando, *Energy Fuels*, 2011, **25**, 1191–1199.
- 122 M. S. Mettler, D. G. Vlachos and P. J. Dauenhauer, *Energy Environ. Sci.*, 2012, **5**, 7797.
- 123 N. Ozbay, A. E. Putun and E. Putun, *Int. J. Energy Res.*, 2006, **30**, 501–510.
- 124 J. J. M. Orfão, F. J. A. Antunes and J. L. Figueiredo, *Fuel*, 1999, **78**, 349–358.
- 125 M. G. Grønli, PhD Thesis, Norwegian University of Science and Technology, 1996.
- 126 G. Várhegyi and T. Székely, *Thermochim. Acta*, 1982, **57**, 13–28.
- 127 G. Várhegyi, *Thermochim. Acta*, 1987, **110**, 95–99.
- 128 M. J. Antal, *Ind. Eng. Chem. Res.*, 1995, **34**, 703–717.
- 129 I. Milosavljevic and E. Suuberg, *Ind. Eng. Chem. Res.*, 1995, **34**, 1081–1091.
- 130 D. Radlein, A. Grinshpun, J. Piskorz and D. S. Scott, *J. Anal. Appl. Pyrolysis*, 1987, **12**, 39–49.
- 131 O. Boutin, M. Ferrer and J. Lédé, *Chem. Eng. Sci.*, 2002, **57**, 15–25.
- 132 M. Le Bras, J. Yvon, S. Bourbigot and V. Mamleev, *J. Anal. Appl. Pyrolysis*, 2009, **84**, 1–17.
- 133 O. Boutin, J. Lede, G. Olalde and A. Ferriere, *J. Phys. IV Fr.*, 1999, **9**, 367–372.
- 134 A. Broido and M. W. Weinstein, in *Proceedings of the 3rd International conference on thermal Analysis*, Birkhauser Verlag, 1972, pp. 285–296.
- 135 C. Di Blasi, *J. Anal. Appl. Pyrolysis*, 1998, **47**, 43–64.
- 136 J. Lédé, J. P. Diebold, G. V. C. Peacocke and J. Piskorz, in *Developments in Thermochemical Biomass Conversion*, Springer Netherlands, 1997, pp. 27–42.
- 137 F. J. Kilzer and A. Broido, *Pyrodynamic*, 1965, **2**, 151–163.
- 138 F. J. Kilzer and A. Broido, *Pyrodynamic*, 1965, **2**, 151–163.
- 139 G. Várhegyi, E. Jakab and M. J. Antal Jr., *Energy Fuels*, 1994, **8**, 1345–1352.
- 140 J. P. Diebold, *Biomass Bioenergy*, 1994, **7**, 75–85.
- 141 F. Shafizadeh, M. L. Wolfson and R. S. Tipson, *Adv. Carbohydr. Chem.*, 1968, **23**, 419–474.
- 142 C. A. Koufopoulos, G. Maschio and A. Lucchesi, *Can. J. Chem. Eng.*, 1989, **67**, 75–84.
- 143 J. L. Banyasz, S. Li, J. L. Lyons-Hart and K. H. Shafer, *J. Anal. Appl. Pyrolysis*, 2001, **57**, 223–248.
- 144 R. W. Agrawal, *Can. J. Chem. Eng.*, 1988, **66**, 403–412.
- 145 A. G. W. Bradbury, Y. Sakai and F. Shafizadeh, *J. Appl. Polym. Sci.*, 1979, **23**, 3271–3280.
- 146 G. Várhegyi, M. J. Antal, E. Jakab and P. Szabó, *J. Anal. Appl. Pyrolysis*, 1997, **42**, 73–87.
- 147 C. Șerbănescu, *Chem. Pap.*, 2014, **67**, 847–860.
- 148 H. Yang, R. Yan, H. Chen, C. Zheng, D. H. Lee and D. T. Liang, *Energy Fuels*, 2006, **20**, 388–393.
- 149 R. Font, A. Marcilla, E. Verdu and J. Devesa, *Ind. Eng. Chem. Res.*, 1990, **29**, 1846–1855.
- 150 T. R. Rao and A. Sharma, *Energy*, 1998, **23**, 973–978.
- 151 X. Guo, Z. Luo, K. Wang and S. Wang, *J. Anal. Appl. Pyrolysis*, 2011, **91**, 183–189.
- 152 A. Dufour, M. Castro-Díaz, P. Marchal, N. Brosse, R. Olcese, M. Bouroukba and C. Snape, *Energy Fuels*, 2012, **26**, 6432–6441.
- 153 A. O. Oyedun, T. Gebreegziabher and C. W. Hui, *Fuel Process. Technol.*, 2012, **106**, 595–604.
- 154 P. E. A. Debiagi, C. Pecchi, G. Gentile, A. Frassoldati, A. Cuoci, T. Faravelli and E. Ranzi, *Energy Fuels*, 2015, **29**, 6544–6555.



- 155 R. Vinu and L. J. Broadbelt, *Energy Environ. Sci.*, 2012, **5**, 9808.
- 156 M. Calonaci, R. Grana, E. B. Hemings, G. Bozzano, M. Dente and E. Ranzi, *Energy Fuels*, 2010, **24**, 5727–5734.
- 157 A. Anca-Couce, P. Sommersacher and R. Scharler, *J. Anal. Appl. Pyrolysis*, 2017, **127**, 411–425.
- 158 A. Anca-Couce, R. Mehrabian, R. Scharler and I. Obernberger, *Energy Convers. Manage.*, 2014, **87**, 687–696.
- 159 Y. Liao, S. Wang and X. Ma, *Prepr. Pap. - Am. Chem. Soc., Div. Fuel Chem.*, 2004, **49**, 407–412.
- 160 R. H. Venderbosch and W. Prins, *Biofuels, Bioprod. Biorefin.*, 2010, **4**, 178–208.
- 161 D. S. Scott, D. Radlein, J. Piskorz and L. Paterson, *J. Anal. Appl. Pyrolysis*, 2001, **57**, 169–176.
- 162 A. J. Yanez, P. Natarajan, W. Li, R. Mabon and L. J. Broadbelt, *Energy Fuels*, 2018, **32**, 1822–1830.
- 163 S. Niksa, *Proc. Combust. Inst.*, 2000, **28**, 2727–2733.
- 164 C. P. Please, M. J. McGuinness and D. L. S. McElwain, *Combust. Flame*, 2003, **133**, 107–117.
- 165 Q. Xiong, J. Zhang, F. Xu, G. Wiggins and C. Stuart Daw, *J. Anal. Appl. Pyrolysis*, 2016, **117**, 176–181.
- 166 D. K. Shen, S. Gu, B. Jin and M. X. Fang, *Bioresour. Technol.*, 2011, **102**, 2047–2052.
- 167 Q.-V. Bach, K.-Q. Tran and Ø. Skreiberg, *Appl. Energy*, 2017, **185**, 1059–1066.
- 168 L. Fiori, M. Valbusa, D. Lorenzi and L. Fambri, *Bioresour. Technol.*, 2012, **103**, 389–397.
- 169 J. Cai and R. Liu, *Bioresour. Technol.*, 2008, **99**, 2795–2799.
- 170 A. A. Jain, A. Mehra and V. V. Ranade, *Fuel*, 2016, **165**, 490–498.
- 171 S. Wang, H. Lin, B. Ru, W. Sun, Y. Wang and Z. Luo, *J. Anal. Appl. Pyrolysis*, 2014, **108**, 78–85.
- 172 M. V. Navarro, R. Murillo, A. M. Mastral, N. Puy and J. Bartroli, *AIChE J.*, 2009, **55**, 2700–2715.
- 173 J. Cai, S. Yang and T. Li, *Bioresour. Technol.*, 2011, **102**, 3642–3644.
- 174 A. Meng, H. Zhou, L. Qin, Y. Zhang and Q. Li, *J. Anal. Appl. Pyrolysis*, 2013, **104**, 28–37.
- 175 A. N. Hayhurst, J. F. Davidson, S. A. Scott and J. S. Dennis, *Chem. Eng. Sci.*, 2006, **61**, 2339–2348.
- 176 C. Ulloa, A. L. Gordon and X. Garcia, *J. Anal. Appl. Pyrolysis*, 2004, **71**, 465–483.
- 177 Y. Chen, R. Bassilakis, W. W. Smith, R. M. Carangelo and M. A. Wójtcowicz, *J. Anal. Appl. Pyrolysis*, 2003, **66**, 235–261.
- 178 T. Sonobe and N. Worasuwanarak, *Fuel*, 2008, **87**, 414–421.
- 179 J. Cai, T. Li and R. Liu, *Bioresour. Technol.*, 2011, **102**, 3894–3899.
- 180 K. Miura and T. Maki, *Energy Fuels*, 1998, **12**, 864–869.
- 181 S. Paea, Master's Thesis, Victoria University of Wellington, 2008.
- 182 M. S. Mettler, S. H. Mushrif, A. D. Paulsen, A. D. Javadekar, D. G. Vlachos and P. J. Dauenhauer, *Energy Environ. Sci.*, 2012, **5**, 5414–5424.
- 183 Z. Wang, B. Pecha, R. J. M. Westerhof, S. R. A. Kersten, C.-Z. Li, A. G. McDonald and M. Garcia-Perez, *Ind. Eng. Chem. Res.*, 2014, **53**, 2940–2955.
- 184 S. Zhou, B. Pecha, M. van Kuppevelt, A. G. McDonald and M. Garcia-Perez, *Biomass Bioenergy*, 2014, **66**, 398–409.
- 185 A. Dieguez-Alonso, A. Anca-Couce, N. Zobel and F. Behrendt, *Fuel*, 2015, **153**, 102–109.
- 186 P. R. Patwardhan, D. L. Dalluge, B. H. Shanks and R. C. Brown, *Bioresour. Technol.*, 2011, **102**, 5265–5269.
- 187 B. J. Vreugdenhil, R. W. R. Zwart and R. W. R. Zwart, *Tar formation in pyrolysis and gasification*, Petten, Netherlands, 2009.
- 188 J. Hayashi, T. Shoji, S. Kudo and K. Norinaga, *Fuel*, 2011, **103**, 141–150.
- 189 M. L. Boroson, J. B. Howard, J. P. Longwell and W. A. Peters, *Energy Fuels*, 1989, **3**, 735–740.
- 190 M. L. Boroson, J. B. Howard, J. P. Longwell and W. A. Peters, *AIChE J.*, 1989, **35**, 120–128.
- 191 C. F. Palma, *Appl. Energy*, 2013, **111**, 129–141.
- 192 P. Hasler, T. Nussbaumer and P. Morf, *Fuel*, 2002, **81**, 843–853.
- 193 P. Ahuja, S. Kumar and P. C. Singh, *Chem. Eng. Technol.*, 1996, **19**, 272–282.
- 194 E. Hoekstra, R. J. M. Westerhof, W. Brilman, W. P. M. Van Swaaij, S. R. A. Kersten, K. J. A. Hogendoorn and M. Windt, *AIChE J.*, 2012, **58**, 2830–2842.
- 195 H. N. Stiles and R. Kandiyoti, *Fuel*, 1989, **68**, 275–282.
- 196 J. Hayashi, S. Amamoto, K. Kusakabe and S. Morooka, *Energy Fuels*, 1996, **9**, 290–294.
- 197 E. J. Shin, M. R. Nimlos and R. J. Evans, *Fuel*, 2001, **80**, 1697–1709.
- 198 R. G. Graham, M. A. Bergougnou and B. A. Freil, *Biomass Bioenergy*, 1994, **7**, 33–47.
- 199 Y. Oki, M. Ashizawa, S. Kajitani and Y. Zhang, *Fuel*, 2010, **89**, 302–309.
- 200 H. Böhm and H. Jander, *Phys. Chem. Chem. Phys.*, 1999, **1**, 3775–3781.
- 201 J. Moon, J. Lee, U. Lee and J. Hwang, *Bioresour. Technol.*, 2013, **133**, 429–436.
- 202 S. Ma, Y. Tan and Y. Han, *J. Nat. Gas Chem.*, 2011, **20**, 440–435.
- 203 A. D. Paulsen, PhD Thesis, University of Massachusetts, 2014.
- 204 X. Bai, P. Johnston and R. C. Brown, *J. Anal. Appl. Pyrolysis*, 2013, **99**, 130–136.
- 205 T. Hosoya, H. Kawamoto and S. Saka, *J. Anal. Appl. Pyrolysis*, 2008, **83**, 64–70.
- 206 M. B. Pecha, E. Terrell, J. I. Montoya, F. Stankovikj, L. J. Broadbelt, F. Chejne and M. Garcia-Perez, *Ind. Eng. Chem. Res.*, 2017, **56**, 9079–9089.
- 207 J. Montoya, B. Pecha, F. C. Janna and M. Garcia-Perez, *J. Anal. Appl. Pyrolysis*, 2017, **125**, 69–82.
- 208 R. J. M. Westerhof, H. S. Nygård, W. P. M. van Swaaij, S. R. A. Kersten and D. W. F. Brilman, *Energy Fuels*, 2012, **26**, 2274–2280.



- 209 A. Bharadwaj, L. L. Baxter and A. L. Robinson, *Energy Fuels*, 2004, **18**, 1021–1031.
- 210 A. Bliet, W. M. Van Poelje, W. P. M. Van Swaaij and F. P. H. Van Beckum, *AIChE J.*, 1985, **31**, 1666–1681.
- 211 C. Di Blasi, *AIChE J.*, 2002, **48**, 2386–2397.
- 212 H. Thunman and B. Leckner, *Biomass Bioenergy*, 2002, **23**, 47–54.
- 213 G. M. Wiggins, P. N. Ciesielski and C. S. Daw, *Energy Fuels*, 2016, **30**, 4960–4969.
- 214 M. B. Pecha, M. Garcia-Perez, T. D. Foust and P. N. Ciesielski, *ACS Sustainable Chem. Eng.*, 2017, **5**, 1046–1053.
- 215 D. L. Pyle and C. A. Zaror, *Chem. Eng. Sci.*, 1984, **39**, 147–158.
- 216 J. Lédé and O. Authier, *J. Anal. Appl. Pyrolysis*, 2015, **113**, 1–14.
- 217 A. D. Paulsen, M. S. Mettler and P. J. Dauenhauer, *Energy Fuels*, 2013, **27**, 2126–2134.
- 218 M. B. Pecha, J. I. Montoya, C. Ivory, F. Chejne and M. Garcia-Perez, *Ind. Eng. Chem. Res.*, 2017, **56**, 5185–5200.
- 219 J. Proano-Aviles, J. K. Lindstrom, P. A. Johnston and R. C. Brown, *Energy Technol.*, 2017, **5**, 189–195.
- 220 G. Flamant, J. D. Lu and B. Variot, *J. Heat Transfer*, 1994, **116**, 652–659.
- 221 M. Sakin, F. Kaymak-Ertekin and C. Ilcali, *J. Food Eng.*, 2009, **94**, 344–349.
- 222 S. Maduskar, G. G. Facas, C. Papageorgiou, C. L. Williams and P. J. Dauenhauer, *ACS Sustainable Chem. Eng.*, 2018, **6**, 1387–1399.
- 223 P. R. Solomon, M. A. Semo and E. M. Suuberg, *Prog. Energy Combust. Sci.*, 1992, **18**, 133–220.
- 224 X. Gong, Y. Yu, X. Gao, Y. Qiao, M. Xu and H. Wu, *Energy Fuels*, 2014, **28**, 5204–5211.
- 225 A. V. Sepman and L. P. H. de Goey, *Biomass Bioenergy*, 2011, **35**, 2903–2909.
- 226 S. S. Liaw, V. Haber Perez, S. Zhou, O. Rodriguez-Justo and M. Garcia-Perez, *J. Anal. Appl. Pyrolysis*, 2014, **109**, 140–151.
- 227 K. J. A. Hogendoorn, E. Hoekstra, S. R. A. Kersten and W. P. M. van Swaaij, *Chem. Eng. J.*, 2012, **191**, 45–58.
- 228 Ö. M. Koçkar, Ö. Onay and S. H. Beis, *Renewable Energy*, 2002, **26**, 21–32.
- 229 M. N. Islam, M. R. A. Beg and M. R. Islam, *Renewable Energy*, 2005, **30**, 413–420.
- 230 E. Hoekstra, W. P. M. Van Swaaij, S. R. A. Kersten and K. J. A. Hogendoorn, *Chem. Eng. J.*, 2012, **187**, 172–184.
- 231 S. Kersten and M. Garcia-Perez, *Curr. Opin. Biotechnol.*, 2013, **24**, 414–420.
- 232 G. L. Comstock, *Wood Fiber*, 1970, **1**, 283–289.
- 233 D. W. Rutherford, R. L. Wershaw and L. G. Cox, *Changes in Composition and Porosity Occurring During the Thermal Degradation of Wood and Wood Components*, US Department of the Interior, US Geological Survey, 2004.
- 234 C. Brackmann, M. Aldén, P.-E. Bengtsson, K. O. Davidsson and J. B. Pettersson, *Appl. Spectrosc.*, 2003, **57**, 216–222.
- 235 S. R. A. Kersten, X. Wang, W. Prins and W. P. M. van Swaaij, *Ind. Eng. Chem. Res.*, 2005, **44**, 8773–8785.
- 236 D. S. Scott, P. Majerski, J. Piskorz and D. Radlein, *J. Anal. Appl. Pyrolysis*, 1999, **51**, 23–37.
- 237 W. Shurong, F. Mengxiang, Y. Chunjiang, L. Zhongyang, C. Kefa, S. Wang, M. Fang, C. Yu, Z. Luo and K. Cen, *China Particuol.*, 2005, **3**, 136–140.
- 238 A. V. Bridgwater, *Biomass Bioenergy*, 2012, **38**, 68–94.
- 239 P. O. Okeunle, H. Watanabe, T. Pattanotai and K. Okazaki, *J. Therm. Sci. Technol.*, 2012, **7**, 1–15.
- 240 M. R. Hajaligol, PhD Thesis, Massachusetts Institute of Technology, 1980.
- 241 M. Garcia-Perez, P. Lappas, P. Hughes, L. Dell, A. Chaala, D. Kretschmer and C. Roy, *IFRF Combust. J.*, 2006, 1–27.
- 242 J. Kizito, R. Balasubramaniam, H. Nahra, J. Agui and D. Troung, in *47th AIAA Aerospace Sciences Meeting including The New Horizons Forum and Aerospace Exposition*, Aerospace Research Central, Orlando, Florida, 2009, pp. 1–11.
- 243 M. M. Avulapati, L. C. Ganippa, J. Xia and A. Megaritis, *Fuel*, 2016, **166**, 59–66.
- 244 A. R. Teixeira, K. G. Mooney, J. S. Kruger, C. L. Williams, W. J. Suszynski, L. D. Schmidt, D. P. Schmidt and P. J. Dauenhauer, *Energy Environ. Sci.*, 2011, **4**, 4306.
- 245 E. Mura, P. Massoli, C. Josset, K. Loubar and J. Bellettre, *Exp. Therm. Fluid Sci.*, 2012, **43**, 63–70.
- 246 S. Das, D. S. Kumar and S. Bhaumik, *Appl. Therm. Eng.*, 2016, **96**, 555–567.
- 247 S. Nordin, J. Nyren and E. Back, *Text. Res. J.*, 1974, **44**, 915–917.
- 248 Y. Halpern and S. Patai, *Isr. J. Chem.*, 1969, **7**, 673–683.
- 249 O. P. Golova, *Russ. Chem. Rev.*, 1975, **44**, 687–697.
- 250 J. Diebold and J. Scahill, Ablative pyrolysis of macroparticles of biomass, in *Proceedings of Specialists' Workshop on fast Pyrolysis of Biomass, Copper Mountain*, Springer Netherlands, Copper Mountain, 1985.
- 251 J. Lédé, J. Panagopoulos, H. Z. Li and J. Villermaux, *Fuel*, 1985, **64**, 1514–1520.
- 252 A. D. Pouwels, G. B. Eijkel, P. W. Arisz and J. J. Boon, *J. Anal. Appl. Pyrolysis*, 1989, **15**, 71–84.
- 253 O. Boutin, M. Ferrer and J. Lédé, *J. Anal. Appl. Pyrolysis*, 1998, **47**, 13–31.
- 254 J. Piskorz, D. S. Scott and D. Radlein, in *Frontiers of pyrolysis: biomass conversion and polymer recycling*, Breckenridge-USA, 1995.
- 255 J. Piskorz, A. Vladars-Usas, D. Radlein, P. Majerski and D. Scott, *J. Anal. Appl. Pyrolysis*, 2000, **56**, 145–166.
- 256 T. McGrath, V. Baliga, D. Miser, M. Hajaligol and R. Sharma, *J. Anal. Appl. Pyrolysis*, 2003, **66**, 191–215.
- 257 R. Bounaceur, B. Quartassi, A. Dufour and A. Zoulalian, *Chem. Eng. Res. Des.*, 2011, **89**, 2136–2146.
- 258 Q. Liu, S. Wang, K. Wang, X. Guo, Z. Luo and K. Cen, *Acta Phys.-Chim. Sin.*, 2008, **24**, 1957–1963.
- 259 H. Z. Li, J. Villermaux, H. Martin and J. Lédé, *J. Anal. Appl. Pyrolysis*, 1987, **10**, 291–308.



- 260 M. K. Koch, A. Voßnacke, J. Starflinger, W. Schütz and H. Unger, *J. Aerosol Sci.*, 2000, **31**, 1015–1028.
- 261 K. Butler, *A numerical model for combustion of bubbling theroplastic materials in microgravity*, National Institute of Standards and Technology, Boulder, CO, USA, 2002.
- 262 M. Shojaeian and A. Koşar, *Exp. Therm. Fluid Sci.*, 2015, **63**, 45–73.
- 263 T. J. Haas, M. R. Nimlos and B. S. Donohoe, *Energy Fuels*, 2009, **23**, 3810–3817.
- 264 K. Voorhees, D. M. Blazsó, D. C. Schwarzingler, P. Carlsson, H. Lycksam, P. Gren, R. Gebart, H. Wiinikka and K. Iisa, *J. Anal. Appl. Pyrolysis*, 2013, **103**, 278–286.
- 265 G. Zhu, X. Zhu, Z. Xiao, R. Zhou and F. Yi, *Waste Manage.*, 2012, **32**, 2287–2293.
- 266 R. J. M. Westerhof, S. R. G. Oudenshoven, P. S. Marathe, M. Englen, M. Garcia-Pérez, Z. Wang and S. R. A. Kersten, *React. Chem. Eng.*, 2016, **1**, 555–566.
- 267 J. Montoya, B. Pecha, F. C. Janna and M. Garcia-Perez, *J. Anal. Appl. Pyrolysis*, 2016, **122**, 106–121.
- 268 J. Montoya, B. Pecha, F. C. Janna and M. Garcia-Perez, *J. Anal. Appl. Pyrolysis*, 2017, **124**, 204–218.
- 269 T. Shoji, H. Kawamoto and S. Saka, *J. Anal. Appl. Pyrolysis*, 2014, **109**, 185–195.
- 270 V. Mamleev, S. Bourbigot, M. Le Bras and J. Yvon, *J. Anal. Appl. Pyrolysis*, 2009, **84**, 1–17.
- 271 V. Oja and E. M. Suuberg, *J. Chem. Eng. Data*, 1998, **43**, 486–492.
- 272 E. Suuberg and V. Oja, *Vapor pressures and heats of vaporization of primary coal tars*, Federal Energy Technology Center, Morgantown, WV (US), Federal Energy Technology Center, Pittsburgh, PA (US), 1997, vol. 12.
- 273 V. Oja and E. M. Suuberg, *J. Chem. Eng. Data*, 1999, **44**, 26–29.
- 274 W. L. H. Hallett and N. A. Clark, *Fuel*, 2006, **85**, 532–544.
- 275 T. E. McGrath, W. G. Chan and M. R. Hajaligol, *J. Anal. Appl. Pyrolysis*, 2003, **66**, 51–70.
- 276 I. Pastorova, R. E. Botto, P. W. Arisz and J. J. Boon, *Carbohydr. Res.*, 1994, **262**, 27–47.
- 277 J. Scheirs, G. Camino and W. Tumiatti, *Eur. Polym. J.*, 2001, **37**, 933–942.
- 278 H. Kawamoto, S. Horigoshi and S. Saka, *J. Wood Sci.*, 2007, **53**, 268–271.
- 279 M. Brebu and C. Vasile, *Cellul. Chem. Technol.*, 2010, **44**, 353.
- 280 M. W. Smith, B. Pecha, G. Helms, L. Scudiero and M. Garcia-Perez, *Biomass Bioenergy*, 2017, **104**, 17–35.
- 281 H. Watanabe, *Energy Fuels*, 2018, **32**, 4248–4254.
- 282 J. Ratte, F. Marias, J. Vaxelaire and P. Bernada, *J. Hazard. Mater.*, 2009, **170**, 1023–1040.
- 283 N. Bech, P. A. Jensen, K. Dam-Johansen and M. B. Larsen, *Biomass Bioenergy*, 2009, **33**, 999–1011.
- 284 P. K. Agarwal, *Chem. Eng. Sci.*, 1991, **46**, 1115–1127.
- 285 C. Di Blasi, *Chem. Eng. Sci.*, 1996, **51**, 2211–2220.
- 286 X. Bai and R. C. Brown, *J. Anal. Appl. Pyrolysis*, 2014, **105**, 363–368.
- 287 M. S. Oh, W. A. Peters and J. B. Howard, *AIChE J.*, 1989, **35**, 775–792.
- 288 Q. Xiong, Y. Yang, F. Xu, Y. Pan, J. Zhang, K. Hong, G. Lorenzini and S. Wang, *ACS Sustainable Chem. Eng.*, 2017, **5**, 2783–2798.
- 289 Q. Xiong, F. Xu, Y. Pan, Y. Yang, Z. Gao, S. Shu, K. Hong, F. Bertrand and J. Chaouki, *Chem. Eng. Process. - Process Intensif.*, 2018, **127**, 206–212.
- 290 B. Moghtaderi, V. N. D. Fletcher and J. H. Kent, *Fire Mater.*, 1997, **21**, 7–16.
- 291 C. H. Bamford, J. Crank, D. H. Malan and A. H. Wilson, *Math. Proc. Cambridge Philos. Soc.*, 1946, **42**, 166.
- 292 A. M. Kanury and P. L. Blackshear, *Combust. Sci. Technol.*, 1970, **2**, 5–9.
- 293 H.-C. Kung, *Combust. Flame*, 1972, **18**, 185–195.
- 294 A. M. Kanury, *Combust. Flame*, 1972, **18**, 75–83.
- 295 P. S. Maa and R. C. Bailie, *Combust. Sci. Technol.*, 1973, **7**, 257–269.
- 296 L. T. Fan, L.-S. Fan, K. Miyanami, T. Y. Chen and W. P. Walawender, *Can. J. Chem. Eng.*, 1977, **55**, 47–53.
- 297 E. J. Kansa, H. E. Perlee and R. F. Chaiken, *Combust. Flame*, 1977, **29**, 311–324.
- 298 C. A. Zaror and D. L. Pyle, *Proc. - Indian Acad. Sci., Sect. C*, 1982, **5**, 269–285.
- 299 H. A. Becker, A. M. Phillips and J. Keller, *Combust. Flame*, 1984, **58**, 163–189.
- 300 M. J. J. Antal, *Fuel*, 1985, **64**, 1483–1486.
- 301 W. C. R. Chan, M. Kelbon and B. B. Krieger, *Fuel*, 1985, **64**, 1505–1513.
- 302 M. A. Hastaoglu and F. Berruti, *Fuel*, 1989, **68**, 1408–1415.
- 303 S. S. Alves and J. L. Figueiredo, *Chem. Eng. Sci.*, 1989, **44**, 2861–2869.
- 304 J. Lédé, *Biomass Bioenergy*, 1994, **7**, 49–60.
- 305 J.-R. Richard and J. Saastamoinen, *Combust. Flame*, 1996, **106**, 288–300.
- 306 A. Sharma, V. Pareek, S. Wang, Z. Zhang, H. Yang and D. Zhang, *Comput. Chem. Eng.*, 2014, **60**, 231–241.
- 307 M. Bellais, PhD Thesis, Royal Institute of Technology, Stockholm, 2007.
- 308 X. Wang, S. R. A. Kersten, W. Prins and W. P. M. van Swaaij, *Ind. Eng. Chem. Res.*, 2005, **44**, 8786–8795.
- 309 Y. Zeng and C. F. F. Lee, *Proc. Combust. Inst.*, 2007, **31 II**, 2185–2193.
- 310 M. B. Pecha, E. Ramirez, G. M. Wiggins, D. Carpenter, B. Kappes, S. Daw and P. N. Ciesielski, *Energy Fuels*, 2018, **32**, 10683–10694.
- 311 E. Leonardi, G. de Vahl Davis, R. K. K. Yuen and G. H. Yeoh, *Int. J. Heat Mass Transfer*, 2007, **50**, 4371–4386.
- 312 A. Mahmoudi, I. Mejri, M. A. Abbassi and A. Omri, *Int. J. Math. Comput. Phys. Quantum Eng.*, 2014, **8**, 668–677.
- 313 G. Gentile, P. E. A. Debiagi, A. Cuoci, A. Frassoldati, E. Ranzi and T. Faravelli, *Chem. Eng. J.*, 2017, **321**, 458–473.
- 314 Y. Sezen, *Int. J. Heat Mass Transfer*, 1989, **32**, 1992–1987.
- 315 L. Ghazaryan, PhD Thesis, University of Twente, 2000.

