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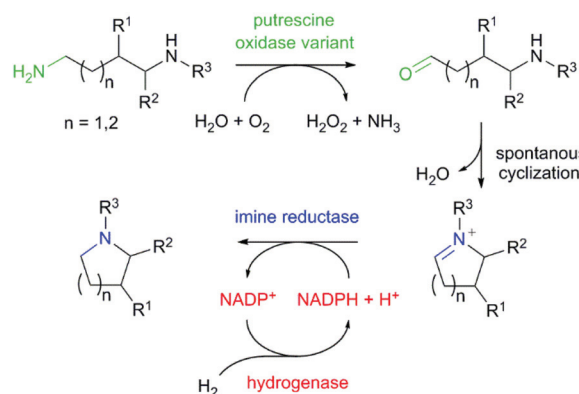
Synthesis of *N*-heterocycles from diamines via H₂-driven NADPH recycling in the presence of O₂[†]

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Herein, we report an enzymatic cascade involving an oxidase, an imine reductase and a hydrogenase for the H₂-driven synthesis of *N*-heterocycles. Variants of putrescine oxidase from *Rhodococcus erythropolis* with improved activity were identified. Substituted pyrrolidines and piperidines were obtained with up to 97% product formation in a one-pot reaction directly from the corresponding diamine substrates. The formation of up to 93% ee gave insights into the specificity and selectivity of the putrescine oxidase.

Saturated *N*-heterocycles are widespread in biologically active molecules. In particular, pyrrolidines and piperidines are increasingly attractive scaffolds found in agrochemicals, pharmaceuticals and natural alkaloid products.¹ Among the different strategies reported for the preparation of saturated nitrogen heterocycles,² the biocatalytic synthesis *via* cascades has attracted particular interest. Biocatalytic cascades coupling several enzymatic transformations in a one-pot process allow the formation of complex molecular architectures under ambient conditions and without the need of protecting groups and purification of intermediates.^{3,4} To date, current enzymatic cascade strategies for the synthesis of pyrrolidine and piperidine heterocycles involve ω -transaminases, monoamine oxidases, carboxylic acid reductases and imine reductases including also artificial imine reductases.^{5–7} Although these cascades have been mainly used for the transformation of keto acids, diketones and keto aldehydes to the corresponding heterocyclic products, only one example has been described for the preparation of *N*-heterocycles from the corresponding diamines. The group of Turner has described the transamin-

ation of terminal aliphatic diamines and polyamines using putrescine transaminase and pyruvate as amine acceptor to provide different *N*-heterocycles in good yields.^{8,9} Highest activity was observed towards the natural substrate putrescine (1,4-diaminobutane). However, highly decreased activity using a methyl-substituted diamine has been obtained. The low activity of putrescine transaminase towards this compound might result from the rather narrow substrate access channel to the active site pocket. In the transformation of substituted diamines, there is still a requirement for an efficient oxidation. We wondered if we could access pyrrolidine and piperidine *N*-heterocycles in a biocatalytic cascade from diamines employing a diamine oxidase (putrescine oxidase) for the first oxidation step followed by the reduction of the *in-situ* formed cyclic imine using an imine reductase (IRED) (Scheme 1).



Scheme 1 Synthesis of pyrrolidine and piperidine heterocyclic scaffolds from diamines by an enzymatic cascade. In the first step one amino group is oxidized with molecular oxygen to a carbonyl group using putrescine oxidase variant (green). Subsequently a spontaneous cyclisation forms the cyclic imine, which is reduced by the imine reductase (blue) to obtain a saturated *N*-heterocycle. The consumed NADPH cofactor is regenerated *via* H₂-driven enzymatic cofactor regeneration without by-product formation, using a NADP⁺-reducing hydrogenase (red). Diamine substrates with methyl substituents either on site R¹, R² or R³ were selected.

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The asymmetric reduction of prochiral imines with NADPH-dependent IREDs has enjoyed particular success for the generation of *N*-heterocyclic compounds including pyrrolidines, piperidines, indolines and tetrahydroisoquinolines.^{10–17} Given the high cost, stoichiometric use, and instability of NADPH, regeneration of this cofactor is essential for practical application. Therefore, we have based our approach on emerging “green” chemistry. This involves no utilization of toxic compounds and solvents and 100% atom efficiency by applying the H₂ hydrogenase cofactor regeneration system.¹⁸

For the initial oxidation of diamines and polyamines, we selected the putrescine oxidase (PuO) from *Rhodococcus erythropolis*.¹⁹ PuO is a flavin-dependent amine oxidase that catalyses the oxidation of its natural substrate putrescine (**1**) with molecular oxygen (O₂) as electron acceptor. This makes oxidases inexpensive and straightforward in usage compared to other redox enzymes. Unfortunately, PuO possesses a narrow substrate specificity accepting only small aliphatic diamines as substrates such as putrescine (**1**) and cadaverine (**2**). In an attempt to increase the substrate spectrum, we initially engineered PuO through directed evolution using error prone PCR (epPCR). We exchanged an average number of four nucleotides that corresponds to one or two amino acid substitutions. For the screening of variants with improved activity, a solid phase assay was used.²⁰ We examined the altered oxidation activity by screening libraries against 1,5-diamino-2-methylpentane (**3**) and 1,5-diaminohexane (**4**).

Interestingly, the best variant PuO^{E203G} for the oxidation of substituted substrate **4** possesses a single amino acid substitution of glutamic acid at position 203 to glycine. Analysis of the crystal structure revealed that this residue is located at the access channel to the active site pocket of PuO. We assume that the introduction of a smaller amino acid residue at this position expands the access channel to the active site and facilitates the passage of substituted or elongated diamine substrates (Fig. 1). In this light, we tested a panel of nine diamines (**1**–**9**) and two polyamines (**21**–**22**) with purified PuO^{native} and PuO^{E203G} (Fig. 1). Both wildtype and variant were equipped with a *Strep-tag* II and purified by using affinity chromatography (Fig. S1†). We could demonstrate that the activity of both enzymes was decreased when one amino group harboured a methyl substituent (**5**) and was abolished when the diamine was highly substituted (**9**), indicating that the less hindered ω-amino group is oxidized by PuO.

For the variant, increased activities were observed with five different non-natural substrates including 1,5-diaminopentane (**2**), 1,5-diaminohexane (**4**) and *N*-methyl putrescine (**5**). We speculate that the configuration of the active site in PuO^{E203G} enables almost exclusively the oxidation of the less-hindered amino group. Photometric analysis of the cofactor content revealed 0.94 ± 0.2 and 1.2 ± 0.14 FAD per PuO^{native} and PuO^{E203G}, respectively, which is in contrast to previous studies with only 0.5 FAD per enzyme.^{19,21} The previous low FAD loading is related to competing binding of the inhibitor

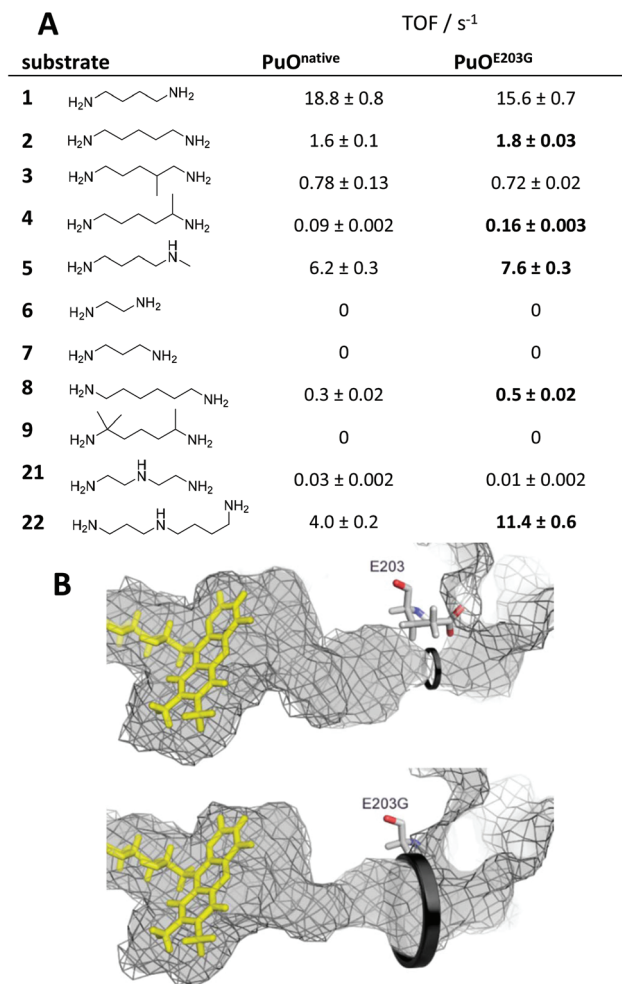


Fig. 1 Engineering of putrescine oxidase and substrate scope. (A) Turnover frequencies of PuO^{native} and PuO^{E203G} are shown for nine different diamines (**1**–**9**) and two polyamine substrates (**21**–**22**). Improved activities of the variant are highlighted. (B) Comparison of substrate channels in PuO^{native} (top, PDB code: 2YG4) and PuO^{E203G} (bottom, homology model via SWISS-MODEL-server).²³ Inner surfaces of channels are shown in grey visualized with PyMOL. The structures were calculated with Yasara,²⁴ running energy minimizations in water and subsequent MD refinements (0.5 ns, force field: YAMBER3). The steric bottleneck of the channel next to glutamic acid at position E203 is opened by introducing glycine at this position (indicated with black circles).

adenosine diphosphate to the active site.²² The nearly total FAD occupancy of purified PuO derivatives can be explained by optimized PuO production conditions at low temperature (18 °C) in TB complex media. The UV/Vis spectra showed typical characteristics of flavoenzymes (Fig. S2†).

We next addressed the enzyme kinetics of PuO^{E203G} compared to PuO^{native} using four model diamine substrates **1**–**4** (Table 1, Fig. S3A–D†) and evaluated the Michaelis constant for the co-substrate O₂ (Fig. S3E†). In order to shed light on the optimal biotransformation conditions, we first measured the affinity of PuO to O₂ using a peroxidase-coupled assay similar to Hellemond and co-workers.¹⁹



Table 1 Kinetic data of PuO^{native} and variant PuO^{E203G} for substrates 1,4-diaminobutane (1), 1,5-diaminopentane (2), 1,5-diamino-2-methylpentane (3) and 1,5-diaminohexane (4). All kinetic parameters were determined at 30 °C, pH 8 and 100% oxygen

Substrate	PuO ^{native}			PuO ^{E203G}		
	K_M (μM)	k_{cat} (s^{-1})	k_{cat}/K_M ($\text{s}^{-1} \text{mM}^{-1}$)	K_M (μM)	k_{cat} (s^{-1})	k_{cat}/K_M ($\text{s}^{-1} \text{mM}^{-1}$)
1	54	18	331	213	34	160
2	69	1.1	16	315	3.4	11
3	30	0.5	15	177	1.5	8
4	11	0.1	0.9	201	0.1	0.5

The O₂ concentration was precisely determined by an optical sensor before measuring the kinetics with **1**. The steady state kinetic for **1** as a function of O₂ (Fig. S4†) indicated an apparent ping-pong reaction mechanism, where no ternary complex of enzyme–substrate–oxygen is formed similar to *Desa et al.*²¹ The K_m value of 30 μM of native PuO for **1** at ambient O₂ concentration was similar to the result obtained by Kopacz and co-workers.²⁵ In contrast to ambient O₂ concentration, a higher K_M (51.8 μM) for **1** at 1.24 mM O₂ (100% O₂) was observed (Fig. S4†) in line with a ping-pong mechanism. Stopped-flow spectroscopic experiments indicated a bifurcated mechanism at ambient O₂ concentrations due to comparable rate constants of product release from the reduced enzyme and reoxidation of the reduced enzyme–product complex.²⁵ Thus, the results support the assumption that at lower or higher O₂ concentrations, the contribution of the ping-pong and ordered sequential mechanism can change.²⁵ K_M values for O₂ using PuO^{native} and PuO^{E203G} were 246 μM and 296 μM , respectively. It is worth mentioning that affinities for O₂ are at levels similar to those observed for other flavoenzymes and the atmospheric O₂ concentration of 260 μM . Due to the high K_M for O₂, all further kinetic determinations were performed under 100% O₂ saturation (1.24 mM). Under these conditions the glucose-6-phosphate dehydrogenase/glucose-6-phosphate system for cofactor regeneration was used since no H₂ is present for SH-mediated regeneration.

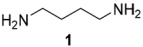

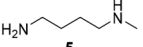

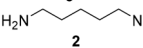

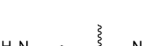

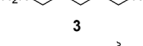

Furthermore, up to 3-fold improved k_{cat} values were observed for variant PuO^{E203G} compared to PuO^{native} (Table 1). However, due to high K_M values the catalytic efficiency was reduced. Interestingly, catalytic efficiencies were 16-fold reduced for 1,5-diaminohexane (**4**) compared to 1,5-diamino-2-methylpentane (**3**), which indicates that the position of the methyl group in the molecule influences the affinity and the acceptance of the substrate.

To assess the potential for the application of variant PuO^{E203G} in the preparation of different pyrrolidine and piperidine heterocycles, we combined PuO^{E203G} with an *R*-selective imine reductase from *Streptosporangium roseum* (*R*-IRED_{Sr}).¹⁶ The IRED-catalysed reduction of imines requires a sufficient supply of NADPH as a reducing equivalent. For the regeneration of NADPH, the enzyme glucose-6-phosphate dehydrogenase is frequently used converting glucose-6-phosphate by NADP⁺ reduction to 6-phosphoglucono- δ -lactone. A highly attractive alternative for NADPH regeneration constitutes the

O₂-tolerant NAD⁺-reducing hydrogenase (SH) from *Ralstonia eutropha* H16, which utilises only molecular hydrogen as the reductant. In contrast to commonly used cofactor regeneration systems, the H₂-based NADH recycling is 100% atom-efficient and relies on a cheap, carbon-free reducing agent.¹⁸ Unlike other hydrogenases, the *R. eutropha* enzyme is O₂-tolerant with its activity remaining unchanged even at ambient O₂ concentrations.²⁶ The new SH-based NADH regenerating enzyme was able to be successfully applied in many multi-cascade reactions *in vitro* as well as in whole cell systems.^{27–30} Recently, the cofactor specificity of SH was altered from NADH to NADPH by rational design (manuscript submitted). The engineered SH^{E341A/S342R} variant retained its O₂ tolerance and permits the use of SH in numerous of reactions where NADPH is required as cofactor. For the regeneration of NADPH both the glucose-6-phosphate dehydrogenase (1 U mL^{−1})/glucose-6-phosphate system (20 mM) and the NADP⁺-dependent variant SH^{E341A/S342R} (1 U mL^{−1}) with H₂ as reducing agent were examined. In this context it is worth noting that primary amines are described to react with aldehydes including also the open-chain form of glucose-6-phosphate to generate imine derivatives.³¹ Indeed, we found that glucose-6-phosphate reacted with **1** and **2** in solution overnight (Fig. S09–S13, Schemes S1 and S2†). Similar observations are reported for different sugars that might react with primary and secondary amines. Even though, those reactions are reversible in aqueous solution and no drastic negative effects were observed in the overall enzyme activity (Table S4†), these side reactions will hamper the isolation and purification of desired products. The glucose/glucose dehydrogenase cofactor regeneration system was reported to show promiscuous enzyme activity towards the reduction of imine compounds,³² which would complicate IRED activity analysis and was therefore not applied. The presented alternative H₂-driven cofactor regeneration circumvents these problems, since the only needed substrate is H₂. In this setup, biotransformations were performed combining PuO^{E203G}, *R*-IRED-*Sr* and SH^{E341A/S342R} in a one-pot process (Table 2). Furthermore, we employed catalase for the decomposition of H₂O₂, which is generated by PuO. Due to the low solubility of H₂ and O₂ in aqueous solutions, biotransformations were performed in small explosive secured and enclosed vessels with 1 mL reaction mixture and excess of headspace (7 mL) containing a 1 : 1 ratio of H₂ and O₂ gas mixtures. This ensured sufficient supply as well as O₂ and H₂ concentrations



Table 2 Product formations and selectivities in the transformation of diamines (1–5) to pyrrolidine and piperidine derivatives

Substrate	Product	Product formation ^a (%)	Optimized conditions for increased product formation ^b (%)	Optimized conditions for increased selectivity ^c (%)
		71 ± 7	n.d.	—
		30 ± 4	34 ± 2	—
		97 ± 2	n.d.	—
		56 ± 6 (<i>ee</i> = 25% <i>R</i>)	99 ± 1 (<i>racemic</i>)	22 ± 3 (<i>ee</i> = 93% <i>R</i>)
		24 ± 12 (<i>ee</i> = 40% <i>S</i>)	86 ± 3 (<i>ee</i> = 40% <i>S</i>)	16 ± 4 (<i>ee</i> = 57% <i>S</i>)

^a Enzyme cascades were performed in Tris-HCl buffer (50 mM, pH 7.5) with 10 mM diamine and 2 mM NADP⁺ in 8 mL glass-vials (horizontal shaking, 180 rpm) for 4 hours at 25 °C. Concentrations of PuO^{E203G} and *R*-IRED_{Sr} were adapted to substrate related activities (PuO: 0.05–1.5 mg mL⁻¹; IRED: 1–2 mg mL⁻¹) and combined with SH^{E341A,S342R} and catalase. Samples were aerated with O₂ and H₂ in the ratio 1:1. ^b Increased product formations were obtained under modified reaction conditions with 5 mM diamine after 2 hours (Table S2, Fig. S5 and S6). ^c Increased enantiomeric excesses were obtained when reactions were stopped after 30 minutes under modified reaction conditions with 5 mM diamine (Table S2). n.d. not determined.

for maximum amine oxidase and hydrogenase activity. With this setup, the transformations of five model diamine substrates to the corresponding *N*-heterocyclic products were employed with up to 97% of product formed (Table 2). Thereby, reaction conditions emerged as crucial for the overall performance of the cascade as well as for the enantiomeric excess (*ee*) of generated products. For substrates which are just poorly accepted by PuO^{E203G}, for example in the transformation of diamine 4, the product formation can be improved either by higher O₂-concentrations (Table S2†) or by increasing the ratio of catalysts PuO^{E203G}/*R*-IRED_{Sr}.

Pyrrolidine (11), piperidine (12) and methylpiperidines (13, 14) were formed in high to excellent product formations (up to 97%) starting from the corresponding diamines (1–4). In addition, chiral methylpiperidines 13 and 14 were obtained after 30 min reaction time with 93% *ee* (*R*) and 57% *ee* (*S*), respectively. It is worth mentioning that selectivities are not induced by *R*-IRED_{Sr}, because the oxidation of diamines mainly causes the formation of the non-prochiral imine intermediate species (Fig. S7 and S8†). Instead, PuO^{E203G} is able to distinguish between both enantiomers from the racemic diamine substrate with one enantiomer being faster converted than the other. In the course of the reaction, however, the concentration of the preferred substrate enantiomer decreases and thus affects the *ee* due to the conversion of the slower-reacting remaining substrate enantiomer (Table S2†). The resulting kinetic resolution allowed for example the asymmetric formation of 13 with up to 93% *ee*. In contrast to 13, the *ee* was just slightly improved for 14 after 30 min reaction time (Table S2†). This reveals that PuO^{E203G} is more selective in the kinetic resolution of 3 compared to 4 and that the enzymatic cascade can be tuned towards productivity or selectivity but not in both directions. Moreover, the secondary amine substrate *N*-methyl 1,4-butanedi-amine (5) was successfully

converted (30% product formation) to the corresponding tertiary amine product *N*-methylputrescine (15). Such products are not accessible *via* other reported enzyme cascades starting from dicarbonyls.

Conclusions

In conclusion, we have developed an enzymatic cascade reaction, allowing access to pyrrolidine and piperidine heterocycles from substituted diamines. The importance of substituted nitrogen heterocycles, in particular pyrrolidine and piperidine types as subunits of bioactive molecules or pharmaceuticals stimulates the development of new synthetic methods. Directed evolution of the diamine oxidase resulted in variants with improved oxidation activity towards non-natural substrates. The presented cascade enabled the production of various *N*-heterocycles and gave insights into the selectivity and specificity of the putrescine oxidase. By optimized conditions for increased selectivity, the enantiomeric excess was enhanced up to 93% *ee* for 13 and demonstrated the kinetic resolution behaviour of the applied oxidase. Importantly, the H₂-driven NADPH regeneration by soluble hydrogenase was well sufficient and surpassed the commonly used glucose-6-phosphate dehydrogenase system with no by-product formation and 100% atom efficiency. Further benefits of the described enzymatic cascade include the prevention of organic solvents and toxic compounds demonstrating the “greenness” of our approach. To the best of our knowledge, this work constitutes an important advance in the field by opening a one-pot and selective access to pyrrolidine and piperidine nitrogen heterocycles from diamines, thus definitely increasing the attractiveness of the H₂-driven regeneration platform.



Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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References

- 1 C. Lamberth and J. Dinges, in *Bioactive Heterocyclic Compound Classes*, John Wiley & Sons, Ltd, 2012, pp. 1–20.
- 2 C. V. T. Vo and J. W. Bode, *J. Org. Chem.*, 2014, **79**, 2809–2815.
- 3 J. H. Schrittwieser, J. Sattler, V. Resch, F. G. Mutti and W. Kroutil, *Curr. Opin. Chem. Biol.*, 2011, **15**, 249–256.
- 4 E. Ricca, B. Brucher and J. H. Schrittwieser, *Adv. Synth. Catal.*, 2011, **353**, 2239–2262.
- 5 S. P. France, L. J. Hepworth, N. J. Turner and S. L. Flitsch, *ACS Catal.*, 2017, **7**, 710–724.
- 6 J. H. Schrittwieser, S. Velikogne, M. Hall and W. Kroutil, *Chem. Rev.*, 2018, **118**, 270–348.
- 7 M. Genz, V. Köhler, M. Krauss, D. Singer, R. Hoffmann, T. R. Ward and N. Sträter, *ChemCatChem*, 2014, **6**, 736–740.
- 8 I. Slabu, J. L. Galman, N. J. Weise, R. C. Lloyd and N. J. Turner, *ChemCatChem*, 2016, **8**, 1038–1042.
- 9 J. L. Galman, I. Slabu, N. J. Weise, C. Iglesias, F. Parmeggiani, R. C. Lloyd and N. J. Turner, *Green Chem.*, 2017, **19**, 361–366.
- 10 G. A. Aleku, S. P. France, H. Man, J. Mangas-Sanchez, S. L. Montgomery, M. Sharma, F. Leipold, S. Hussain, G. Grogan and N. J. Turner, *Nat. Chem.*, 2017, **9**, 961.
- 11 M. Gand, H. Müller, R. Wardenga and M. Höhne, *J. Mol. Catal. B: Enzym.*, 2014, **110**, 126–132.
- 12 S. Hussain, F. Leipold, H. Man, E. Wells, S. P. France, K. R. Mulholland, G. Grogan and N. J. Turner, *ChemCatChem*, 2015, **7**, 579–583.
- 13 F. Leipold, S. Hussain, D. Ghislieri and N. J. Turner, *ChemCatChem*, 2013, **5**, 3505–3508.
- 14 H. Man, E. Wells, S. Hussain, F. Leipold, S. Hart, J. P. Turkenburg, N. J. Turner and G. Grogan, *ChemBioChem*, 2015, **16**, 1052–1059.
- 15 D. Wetzl, M. Berrera, N. Sandon, D. Fishlock, M. Ebeling, M. Müller, S. Hanlon, B. Wirz and H. Iding, *ChemBioChem*, 2015, **16**, 1749–1756.
- 16 P. N. Scheller and B. M. Nestl, *Appl. Microbiol. Biotechnol.*, 2016, **100**, 10509–10520.
- 17 P. N. Scheller, S. Fademrecht, S. Hofelzer, J. Pleiss, F. Leipold, N. J. Turner, B. M. Nestl and B. Hauer, *ChemBioChem*, 2014, **15**, 2201–2204.
- 18 L. Lauterbach, O. Lenz and K. A. Vincent, *FEBS J.*, 2011, **280**, 3058–3068.
- 19 E. W. van Hellemond, M. van Dijk, D. P. H. M. Heuts, D. B. Janssen and M. W. Fraaije, *Appl. Microbiol. Biotechnol.*, 2008, **78**, 455–463.
- 20 M. Alexeeva, R. Carr and N. J. Turner, *Org. Biomol. Chem.*, 2003, **1**, 4133–4137.
- 21 R. J. DeSa, *J. Biol. Chem.*, 1972, **247**, 5527–5534.
- 22 E. W. van Hellemond, H. Mazon, A. J. Heck, R. H. H. van den Heuvel, D. P. H. M. Heuts, D. B. Janssen and M. W. Fraaije, *J. Biol. Chem.*, 2008, **283**, 28259–28264.
- 23 A. Waterhouse, M. Bertoni, S. Bienert, G. Studer, G. Tauriello, R. Gumienny, F. T. Heer, T. A. P. de Beer, C. Rempfer, L. Bordoli, R. Lepore and T. Schwede, *Nucleic Acids Res.*, 2018, **46**, W296–W303.
- 24 E. Krieger, K. Joo, J. Lee, J. Lee, S. Raman, J. Thompson, M. Tyka, D. Baker and K. Karplus, *Proteins: Struct., Funct., Bioinf.*, 2009, **77**, 114–122.
- 25 M. M. Kopacz, D. P. H. M. Heuts and M. W. Fraaije, *FEBS J.*, 2014, **281**, 4384–4393.
- 26 L. Lauterbach and O. Lenz, *J. Am. Chem. Soc.*, 2013, **135**, 17897–17905.
- 27 A. K. Holzer, K. Hiebler, F. G. Mutti, R. C. Simon, L. Lauterbach, O. Lenz and W. Kroutil, *Org. Lett.*, 2015, **17**, 2431–2433.
- 28 J. Ratzka, L. Lauterbach, O. Lenz and M. B. Ansorge-Schumacher, *Biocatal. Biotransform.*, 2011, **29**, 246–252.
- 29 M. Andersson, H. Holmberg and P. Adlercreutz, *Biotechnol. Bioeng.*, 2000, **57**, 79–86.
- 30 T. H. Lonsdale, L. Lauterbach, S. Honda Malca, B. M. Nestl, B. Hauer and O. Lenz, *Chem. Commun.*, 2015, **51**, 16173–16175.
- 31 M. A. Sprung, *Chem. Rev.*, 1940, **26**, 297–338.
- 32 S. Roth, A. Präg, C. Wechsler, M. Marolt, S. Ferlaino, S. Lüdeke, N. Sandon, D. Wetzl, H. Iding, B. Wirz and M. Müller, *ChemBioChem*, 2017, **18**, 1703–1706.

